

RIII Supporting Information Figures

1 **Title:** Unraveling the Role of Lipid Droplets and Perilipin 2 in Bovine Luteal Cells¹

2
3 **Running Title:** Perilipin 2 in Bovine Luteal Cells

4 **Key Words:** Lipid Droplets, Perilipin 2, Steroidogenesis, Progesterone, Corpus Luteum, Prostaglandin F2 α

5 **Authors:** Michele R. Plewes^{2,3,4,#}, Heather A. Talbott², Micah B. Schott⁴, Jennifer R. Wood⁵, Andrea S. Cupp⁵,
6 and John S. Davis^{2,3,4}

7 ²Olson Center for Women's Health, Department of Obstetrics and Gynecology, University of Nebraska Medical
8 Center, 983255 Nebraska Medical Center, Omaha, NE 68198-3255, USA

9 ³Veterans Affairs Nebraska Western Iowa Health Care System, 4101 Woolworth Ave, Omaha, NE 68105, USA

10 ⁴Department of Biochemistry and Molecular Biology, University of Nebraska Medical Center, 985870 Nebraska
11 Medical Center, Omaha, NE 68198-5870, USA

12 ⁵Department of Animal Sciences, University of Nebraska–Lincoln, Lincoln, NE 68583-0908, USA

13 ¹This project was supported by Agriculture and Food Research Initiative Competitive Grant no. 2018-67012-
14 29531 (MRP) and 2017-67015-26450 (JSD) from the USDA National Institute of Food and Agriculture; IK2
15 BX004911-01A1 (MRP) and I01 BX004272 (JSD) from the U.S Department of Veterans Affairs; NIH grants R01
16 HD087402 and R01HD092263; and The Olson Center for Women's Health. JSD is the recipient of a VA Senior
17 Research Career Scientist Award IK6 BX005797.
18

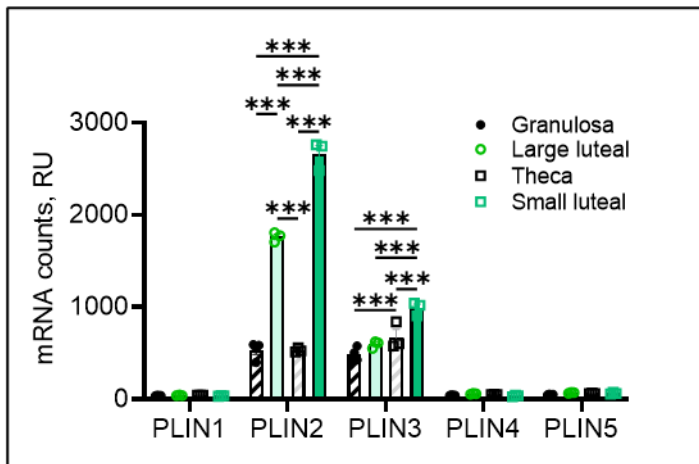
19 **#Correspondence:** Michele R. Plewes, Olson Center for Women's Health, University of Nebraska Medical
20 Center, 983255 Nebraska Medical Center, Omaha, NE 68198-3255, USA.

21 E-mail: michele.plewes@unmc.edu; ORCID: 0000-0002-6086-0104; Phone: 402-559-8627

22 **Disclosure Statement:** The authors have nothing to disclose.
23
24
25
26
27
28
29
30
31
32
33
34
35
36
37
38
39
40
41
42
43
44
45
46
47

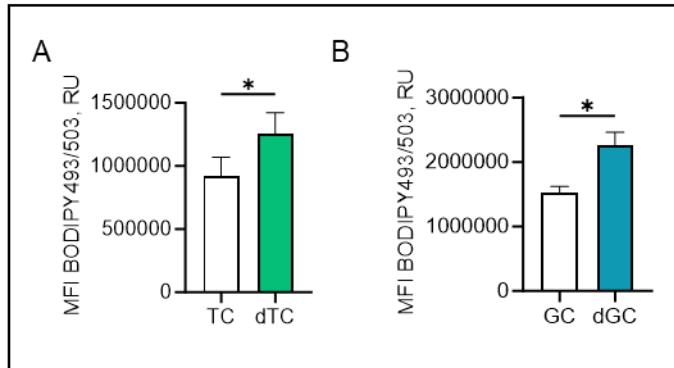
48
49
50
51
52
53
54
55
56
57
58
59
60
61
62
63
64
65
66
67
68
69
70
71
72
73
74
75
76
77
78
79
80
81
82
83
84
85
86
87
88
89
90
91
92
93
94
95
96
97
98
99
100
101
102
103
104
105
106

Supporting Information Figure 1: PLIN2 is highly enriched in bovine luteal cells



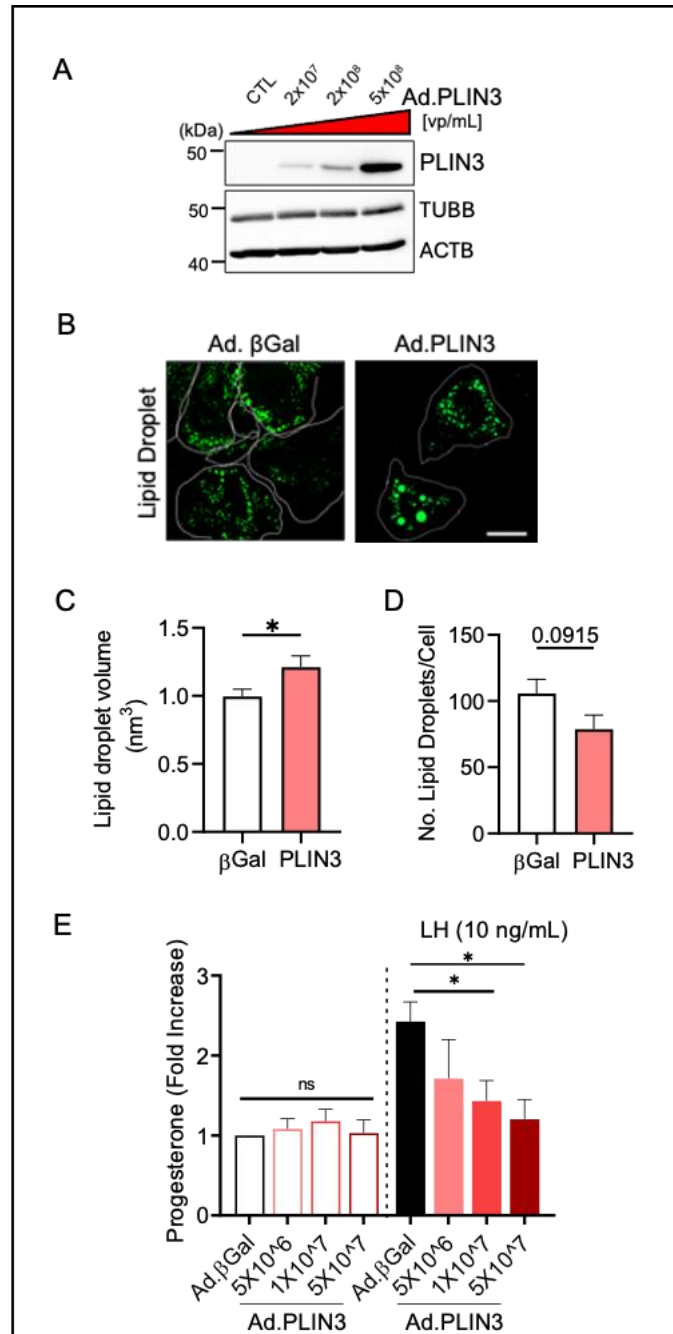
107
108
109
110
111
112
113
114
115
116
117
118
119
120
121
122
123
124
125
126
127
128
129
130
131
132
133
134
135
136
137
138
139
140
141
142
143
144
145
146
147
148
149
150
151
152
153
154
155
156
157
158
159
160
161
162
163
164
165

Supporting Information Figure 2: Lipid droplet accumulation increases as follicular cells undergoing the follicular to luteal transition.

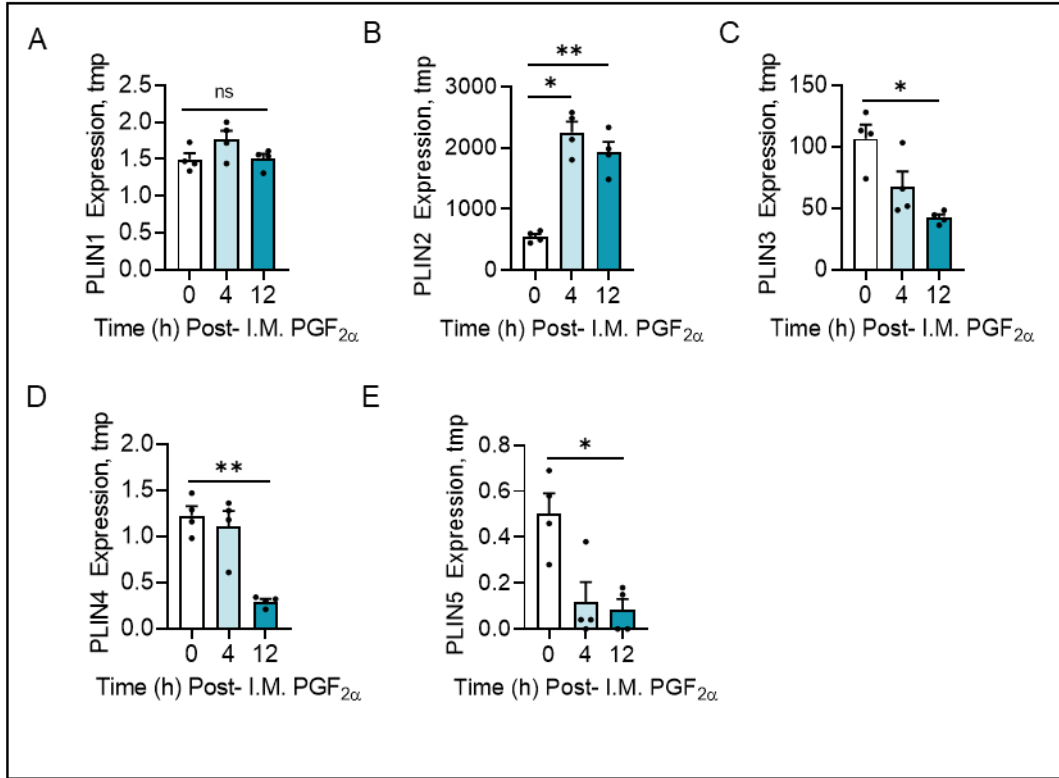


166
167
168
169
170
171
172
173
174
175
176
177
178
179
180
181
182
183
184
185
186
187
188
189
190
191
192
193
194
195
196
197
198
199
200
201
202
203
204
205
206
207
208
209
210
211
212
213
214
215
216
217
218
219
220
221
222
223

Revised Supporting Information Figure 3:
Overexpression of Perilipin 3 (PLIN3) increases
lipid droplet volume and attenuates
progesterone production in bovine small luteal
cells.



Supporting Information Figure 4: Effects of PGF₂alpha on Perilipin mRNA expression *in vivo*.



Supporting information Figure Captions

283

284 **Supporting Information Figure 1: Microarray expression of Bovine Perilipins (1-5).** We mined bovine gene
285 expression arrays from the NCBI GEO repository (GSE83524) to analyze the expression of Perilipins (1-5) in
286 freshly isolated bovine granulosa and theca cells from large follicles and purified preparations of bovine small
287 and large luteal cells from mature corpora lutea. Microarray analysis of the Perilipin (1-5) family. Bovine
288 granulosa cells (n = 4; open black hatch bars; circle symbol), theca cells (n = 3; open grey hatch bars; square
289 symbol), large luteal cells (n = 3; closed light green bars; circle symbol), and small luteal cells (n = 3; closed
290 green bars; square symbol) Significant differences were identified as changes greater than 1.5-fold and $P < 0.05$
291 supported by unpaired t-tests.

292

293 **Supporting Information Figure 2: Lipid droplet accumulation increases as follicular cells undergoing the**
294 **follicular to luteal transition.** Bovine granulosa (GC) and theca cells (TC) were cultured for up to four days in
295 medium contain 1% fetal calf serum with or without insulin/transferrin/selenium, the adenylyl cyclase activator
296 forskolin (10 μM) and phorbol myristate acetate (PMA, 20 nM). (A) Quantitative analyses of the mean
297 fluorescence intensity of BODIPY493/503 in TC or differentiated TC (dTC). (B) Quantitative analyses of the mean
298 fluorescence intensity of BODIPY493/503 in GC or differentiated GC (dGC). Statistics were performed by t-tests
299 to evaluate paired responses. Bars represent means \pm sem. Significant difference between treatments, $*P <$
300 0.05.

301

302 **Supporting Information Figure 3: Overexpression and knockdown of lipid droplet-associated proteins,**
303 **Perilipin 3 (PLIN3), in bovine luteal cells.** Replication-deficient adenoviruses (Ad) containing Beta-galactose
304 (Ad. β Gal; control) or PLIN3 (Ad.PLIN3) were utilized to overexpress PLIN3 in bovine small luteal cells. (A)
305 Representative western blot of dose-dependent overexpression of Ad.PLIN3 in small luteal cells. Small luteal
306 cells were infected with Ad. β Gal or Ad.PLIN3 as described above. After 48 h, luteal cells were equilibrated for
307 two hours and stimulated with luteinizing hormone (LH; 10 ng/mL) for 4 h. Small luteal cells were infected with
308 Ad. β Gal or Ad.PLIN3 and lipid droplets were labeled (Lipi-blue 1 μM) and visualized by confocal microscopy. (B)
309 Representative micrographs of lipid droplets obtained from small luteal cells infected with Ad. β Gal or Ad.PLIN3.
310 (C) Quantification of lipid droplet volume (nm³) in small luteal cells infected with Ad. β Gal or Ad.PLIN3. (D)
311 Quantification of lipid droplet number and in small luteal cells infected with Ad. β Gal or Ad.PLIN3. Statistics were
312 performed by t-tests to evaluate paired responses. Data are means \pm standard error, n=3. (E) Medium
313 progesterone obtained from small luteal cells infected with Ad. β Gal or increasing concentrations Ad.PLIN3
314 following stimulation with LH. Statistics were performed by two-way ANOVA was used to evaluate repeated
315 measures with Tukey's multiple comparison tests. Bars represent means \pm sem, n = 3. Significant difference
316 between treatments, $*P < 0.05$. Micron bar represents 20 μm . Beta Tubulin (TUBB; loading control); Beta Actin
317 (ACTB; loading control).

318

319 **Supporting Information Figure 4: Effects of in vivo administration of Prostaglandin (PG) F2 α on Perilipins**
320 **(PLIN1-5) mRNA expression in bovine corpora lutea.** Mid-luteal phase cows were injected with saline

321 (Control) or PGF2 α , (25 mg, i.m.) and ovariectomized after 4- and 12-h to collect corpora lutea. RNA sequencing
322 of whole luteal tissue was performed. (A) mRNA levels of PLIN1, (B) PLIN2, (C) PLIN3, (D) PLIN4, and (E) PLIN5
323 in the bovine corpus luteum at midcycle and 4- and 12 h post- PGF2 α injection. Data are presented as mean
324 number of transcripts per million (TPM) \pm SEM. n = 4; *p < 0.05, **p < 0.01, compared to 0 h by DESeq2 analysis,
325 Benjamini Hochberg correction. P-values shown are adjusted p-values for multiple comparisons.

326
327