

Washington University School of Medicine

Digital Commons@Becker

2020-Current year OA Pubs

Open Access Publications

7-2-2024

Baseline levels and longitudinal changes in plasma A β 42/40 among Black and white individuals

Chengjie Xiong

Washington University School of Medicine in St. Louis

Jingqin Luo

Washington University School of Medicine in St. Louis

Rachel L Henson

Washington University School of Medicine in St. Louis

Tammie L S Benzinger

Washington University School of Medicine in St. Louis

Quoc Bui

Washington University School of Medicine in St. Louis

See next page for additional authors

Follow this and additional works at: https://digitalcommons.wustl.edu/oa_4



Part of the [Medicine and Health Sciences Commons](#)

Please let us know how this document benefits you.

Recommended Citation

Xiong, Chengjie; Luo, Jingqin; Henson, Rachel L; Benzinger, Tammie L S; Bui, Quoc; Agboola, Folasade; Grant, Elizabeth; Gremminger, Emily N; Moulder, Krista L; Babulal, Ganesh; Cruchaga, Carlos; Holtzman, David M; Bateman, Randall J; Morris, John C; Schindler, Suzanne E; and et al., "Baseline levels and longitudinal changes in plasma A β 42/40 among Black and white individuals." *Nature Communications*. 15, 1. 5539 (2024).

https://digitalcommons.wustl.edu/oa_4/4053

This Open Access Publication is brought to you for free and open access by the Open Access Publications at Digital Commons@Becker. It has been accepted for inclusion in 2020-Current year OA Pubs by an authorized administrator of Digital Commons@Becker. For more information, please contact vanam@wustl.edu.

Authors

Chengjie Xiong, Jingqin Luo, Rachel L Henson, Tammie L S Benzinger, Quoc Bui, Folasade Agboola, Elizabeth Grant, Emily N Gremminger, Krista L Moulder, Ganesh Babulal, Carlos Cruchaga, David M Holtzman, Randall J Bateman, John C Morris, Suzanne E Schindler, and et al.

Baseline levels and longitudinal changes in plasma A β 42/40 among Black and white individuals

Received: 20 December 2023

Accepted: 21 June 2024

Published online: 02 July 2024

 Check for updates

Chengjie Xiong^{1,2,10}, Jingqin Luo^{3,4,10}, David A. Wolk⁵, Leslie M. Shaw^{5,6}, Erik D. Roberson⁷, Charles F. Murchison⁷, Rachel L. Henson², Tammie L. S. Benzinger^{2,8}, Quoc Bui¹, Folasade Agboola¹, Elizabeth Grant¹, Emily N. Gremminger¹, Krista L. Moulder², David S. Geldmacher⁷, Olivio J. Clay^{7,9}, Ganesh Babulal², Carlos Cruchaga², David M. Holtzman², Randall J. Bateman², John C. Morris² & Suzanne E. Schindler²✉

Blood-based biomarkers of Alzheimer disease (AD) may facilitate testing of historically under-represented groups. The Study of Race to Understand Alzheimer Biomarkers (SORTOUT-AB) is a multi-center longitudinal study to compare AD biomarkers in participants who identify their race as either Black or white. Plasma samples from 324 Black and 1,547 white participants underwent analysis with C₂N Diagnostics' PrecivityAD test for A β 42 and A β 40. Compared to white individuals, Black individuals had higher average plasma A β 42/40 levels at baseline, consistent with a lower average level of amyloid pathology. Interestingly, this difference resulted from lower average levels of plasma A β 40 in Black participants. Despite the differences, Black and white individuals had similar longitudinal rates of change in A β 42/40, consistent with a similar rate of amyloid accumulation. Our results agree with multiple recent studies demonstrating a lower prevalence of amyloid pathology in Black individuals, and additionally suggest that amyloid accumulates consistently across both groups.

Biomarkers of Alzheimer's disease (AD), including fluid and imaging biomarkers of amyloid and tau pathology, have enabled a better understanding of AD pathophysiology, facilitated clinical trials that have led to the development of amyloid-lowering treatments, and increased the accuracy of clinical dementia diagnosis¹. While cerebrospinal fluid (CSF)- and positron emission tomography (PET)-based biomarkers accurately detect AD brain pathology, the scale of testing with these modalities is limited by their requirements for specialized

personnel and equipment, perceived risks, and high costs¹⁻⁴. Additionally, individuals from minoritized groups may be less likely to present to memory clinics that perform biomarker testing with CSF and PET⁵. In contrast, blood tests are considered highly accessible, acceptable, and scalable, making blood-based biomarkers ideal tools for research, clinical trials, and clinical practice^{6,7}. Blood-based biomarkers may enable testing of individuals who would not be comfortable with CSF or PET testing and may allow for testing in a community-

¹Division of Biostatistics, Washington University, St. Louis, MO, USA. ²Department of Neurology, Washington University School of Medicine, St. Louis, MO, USA. ³Division of Public Health Sciences, Department of Surgery, Washington University School of Medicine, St. Louis, MO, USA. ⁴Siteman Cancer Center Biostatistics and Qualitative Research Shared Resource, Washington University School of Medicine, St. Louis, MO, USA. ⁵Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA, USA. ⁶Department of Pathology and Laboratory Medicine, University of Pennsylvania, Philadelphia, PA, USA. ⁷Alzheimer's Disease Center, Department of Neurology, University of Alabama at Birmingham, Birmingham, AL, USA. ⁸Mallinckrodt Institute of Radiology, Washington University, St. Louis, MO, USA. ⁹Department of Psychology, University of Alabama at Birmingham, Birmingham, AL, USA. ¹⁰These authors contributed equally: Chengjie Xiong, Jingqin Luo. ✉e-mail: schindler.s.e@wustl.edu

based setting rather than a major medical center⁸. Therefore, blood-based biomarkers of AD may increase biomarker testing of individuals from minoritized groups that have historically been underrepresented in AD research studies and clinical trials^{8–11}.

Multiple epidemiological studies have reported a higher prevalence of dementia in self-identified Black or African American and Hispanic individuals as compared to non-Hispanic white individuals^{12–14}. Despite the higher reported prevalence of dementia, several research studies have reported a lower rate of AD biomarker abnormalities in Black and Hispanic individuals compared to non-Hispanic white individuals^{15–21}, although other studies have found the opposite result or no differences between these groups^{22,23}. The seeming disconnect between the reported prevalence of dementia and the rate of AD biomarker abnormalities has raised concerns that the major etiologies of dementia may vary across racial and ethnic groups and/or that biomarkers may not reflect AD pathology consistently across groups^{19,20,24–26}. Adding to these concerns, concentrations of some plasma biomarkers, including amyloid- β 42 and 40 (A β 42 and A β 40, respectively) and tau phosphorylated at positions 181 and 217 (p-tau181 and p-tau217, respectively) can be affected by medical conditions (e.g., chronic kidney disease and obesity) that are more prevalent in some racial and ethnic groups^{16,27–29}. However, some evidence indicates that plasma biomarker ratios such as A β 42/40 and the ratio of phosphorylated to non-phosphorylated tau (p-tau ratio) may normalize for non-AD-related individual differences and provide more consistent performance in classifying amyloid status across groups^{16,28–30}.

We have previously reported that plasma A β 42/40, as measured by a high-precision mass spectrometry-based assay, has more consistent performance in classifying amyloid status across racial groups as compared to concentrations of phosphorylated tau¹⁶. This finding suggests that plasma A β 42/40, as measured by high-precision assays, may enable consistent classification of amyloid status in diverse groups. Our study and the few other studies that have compared plasma biomarkers in different racial groups only reported cross-sectional data^{16,23,31–34}. Therefore, it is unknown whether the longitudinal rates of change in plasma biomarkers vary by race. The rate of change is particularly important in clinical trials, as it represents the placebo trajectory that is intended to be modified by treatments.

In this study, we assembled a large cohort to examine plasma A β measures (A β 42, A β 40, and A β 42/40) for potential differences in baseline levels and rates of change in self-identified Black and white individuals. Participants from three AD Research Centers (Washington University, University of Pennsylvania, and University of Alabama at Birmingham) were included. Plasma samples were analyzed with the C₂N Diagnostics mass spectrometry-based assays that are currently being used in clinical trials and clinical practice^{35,36}. Linear models were used to estimate the baseline levels and rates of change for plasma biomarker measures in both groups. Analyses also examined whether factors such as sex, *APOE* ϵ 4 carrier status, cognitive status, or medical conditions (hypertension and diabetes) modified potential racial differences.

Results

Participant characteristics

The study cohort included a total of 324 Black participants and 1547 white participants with plasma A β measures from at least one sample (Table 1). The time interval between clinical assessment and plasma collection was 0.16 ± 0.18 years (mean \pm standard deviation). Black and white participants had similar ages at baseline (70.2 ± 8.6 versus 70.5 ± 9.5 years, respectively, $p = 0.26$), and there was no difference in the proportion carrying an *APOE* ϵ 4 allele (45.1% versus 42.6%, $p = 0.35$). Most participants completed at least 12 years of education, with Black participants completing slightly fewer years of education on average compared to white participants (15.3 ± 2.9 years versus 15.8 ± 2.8 years, $p = 0.002$). Black participants were more likely to be cognitively unimpaired than white participants at baseline (72.2%

versus 66.4%, $p = 0.041$) and were more likely than white participants to be female (72.2% versus 53.5%, $p < 0.0001$). Black participants were more likely to be overweight or obese than white participants (69.4% versus 52.0%, $p < 0.0001$) and were more likely than white participants to have a history of hypertension (65.4% versus 42.3%, $p < 0.0001$) or diabetes (17.3% versus 6.0%, $p < 0.0001$). A lower percentage of Black participants were fasting at the time of the baseline plasma collection compared to white participants (36.1% versus 65.5%, $p < 0.0001$). This is because fasting samples were collected at the time of lumbar puncture for many participants, and Black individuals were less likely to choose to undergo lumbar puncture.

Racial differences in baseline biomarker levels

Baseline levels of plasma A β were compared in Black and white participants. The largest comparison available was for unadjusted plasma A β levels because some individuals were missing data on covariates. Unadjusted levels of plasma A β 42/40 were higher in 324 Black individuals compared to 1547 white individuals in both cognitively unimpaired (0.107 ± 0.0119 versus 0.102 ± 0.0106 , $p = 0.0367$) and cognitively impaired groups (0.0995 ± 0.0121 versus 0.0964 ± 0.0105 , $p < 0.0001$), consistent with Black individuals having less brain amyloid (Fig. 1). Next, baseline levels of plasma A β were examined after adjustment for covariates (age, sex, *APOE* ϵ 4 carrier status, years of education, cognitive status, fasting status, BMI, and status for hypertension, diabetes and stroke) (Table 2). Consistent with the unadjusted plasma A β 42/40 results, the covariate-adjusted mean plasma A β 42/40 values were higher (less abnormal) in 214 Black individuals than in 1113 white individuals (0.1201 ± 0.0030 versus 0.1155 ± 0.0030 , $p < 0.0001$). Interestingly, the higher plasma A β 42/40 values in Black individuals compared to white individuals resulted from lower plasma A β 40 levels (177.8 ± 22.7 versus 199.5 ± 22.4 pg/mL, $p = 0.0002$); plasma A β 42 levels were not different in Black and white individuals (26.25 ± 2.40 pg/mL versus 26.41 ± 2.36 pg/mL, $p = 0.80$).

Further analyses examined whether racial differences in the adjusted mean levels of plasma A β were modified by amyloid status, age, sex, *APOE* ϵ 4 carrier status, years of education, cognitive status, BMI, hypertension, or diabetes. Amyloid status by CSF or amyloid PET did not significantly affect racial differences in plasma A β 42/40, A β 42, or A β 40 (Supplementary Table 1). Racial differences in plasma A β 42/40 were larger in cognitively unimpaired compared to cognitively impaired individuals ($p = 0.017$) (Supplementary Table 2), but racial differences in plasma A β 42 or A β 40 were not significantly affected by cognitive status (Supplementary Tables 3, 4). Racial differences in plasma A β 42 or A β 40 were affected by age group ($p = 0.031$ for A β 42, Supplementary Table 3; $p = 0.027$ for A β 40, Supplementary Table 4). Notably, there were no other significant interactions between race and any of the other covariates in models of plasma A β levels, suggesting that Black participants had higher mean levels of plasma A β 42/40 than white participants regardless of amyloid status, sex, *APOE* ϵ 4 carrier status, years of education, BMI, hypertension or diabetes.

Using the same covariate-adjusted models, levels of the amyloid burden by PET and CSF A β 42/40 were compared for Black and white groups (Table 2). Notably, the power to discern differences was lower because many fewer individuals underwent amyloid PET or CSF collection. The covariate-adjusted mean amyloid PET Centiloid values were slightly lower in 89 Black individuals compared to 626 white individuals, although this did not reach significance (21.6 ± 13.6 Centiloids versus 28.2 ± 13.4 Centiloids, $p = 0.072$). The covariate-adjusted mean CSF A β 42/40 values were higher, consistent with lower amyloid burden, in 80 Black individuals compared to 806 white individuals (0.1122 ± 0.0068 versus 0.1069 ± 0.0068 , $p = 0.014$). CSF A β 42 levels were slightly (8.2%) lower in Black individuals compared to white individuals (840 ± 117 versus 915 ± 116 pg/mL, $p = 0.041$), but CSF A β 40 levels were much (23.8%) lower in Black individuals than white individuals (5552 ± 1277 versus 7284 ± 1272 pg/mL, $p < 0.0001$). CSF t-tau and

Table 1 | Characteristics of cohort at baseline

Characteristics	Overall			Cognitively unimpaired			Cognitively impaired		
	Black N = 324 ^a	White N = 1547	<i>p</i>	Black N = 234	White N = 1027	<i>p</i>	Black N = 89	White N = 520	<i>p</i>
Sample characteristics									
Site (<i>n</i> for WU/UPenn/UAB)	249/63/12	1412/115/20	<0.0001	181/47/6	934/83/10	<0.0001	67/16/6	478/32/10	<0.0001
Number of plasma samples per individual (<i>n</i> with 1/2/3/≥ 4 samples)	166/74/ 53/31	788/340/ 207/212	0.15	98/58/ 50/28	392/258/ 181/196	0.056	68/16/3/2	396/82/ 26/16	0.84
Years between the 1st and last plasma sample (mean, SD) ^b	5.11 (3.52)	6.93 (4.17)	<0.0001	5.22 (3.54)	7.17 (4.22)	<0.0001	4.01 (2.94)	5.70 (3.67)	0.018
Fasting status (<i>n</i> for fasting/non-fasting, % fasting)	117/ 207 (36.1%)	1015/ 532 (65.6%)	<0.0001	91/ 143 (38.9%)	734/ 293 (71.5%)	<0.0001	25/ 64 (28.1%)	281/ 239 (54.0%)	<0.0001
Demographics									
Baseline age (mean, SD)	70.2 (8.6)	70.5 (9.5)	0.26	69.0 (7.78)	68.3 (9.67)	0.42	73.8 (9.41)	74.8 (7.55)	0.40
Sex (<i>n</i> , % female)	234 (72.2%)	827 (53.5%)	<0.0001	172 (73.5%)	595 (57.9%)	<0.0001	61 (68.5%)	232 (44.6%)	<0.0001
Years of education (mean, SD)	15.3 (2.92)	15.8 (2.81)	0.002	15.4 (2.84)	16.3 (2.55)	0.0001	14.8 (3.07)	14.9 (3.06)	0.71
APOE ε4 status (<i>n</i> for carrier/non-carrier/missing, % carrier)	146/172/ 6 (45.1%)	659/878/ 10 (42.6%)	0.35	96/136/ 2 (41.0%)	359/667/ 1 (35.0%)	0.08	50/35/ 4 (56.2%)	300/211/ 9 (57.7%)	1.00
Medical conditions									
BMI status (<i>n</i> for overweight or obese/underweight or normal/missing, % overweight or obese)	225/52/ 47 (69.4%)	805/426/ 316 (52.0%)	<0.0001	179/30/ 25 (76.5%)	569/282/ 176 (55.4%)	<0.0001	46/22/ 21 (51.7%)	236/144/ 140 (45.4%)	0.46
Hypertension status (<i>n</i> for positive/negative/missing, % positive)	212/108/ 4 (65.4%)	655/878/ 14 (42.3%)	<0.0001	154/79/ 1 (65.8%)	402/618/ 7 (39.1%)	<0.0001	58/29/ 2 (65.2%)	253/260/ 7 (48.7%)	0.004
Diabetes status (<i>n</i> for positive/negative/missing, % positive)	56/223/ 45 (17.3%)	93/1150/ 304 (6.0%)	<0.0001	44/167/ 23 (18.8%)	53/805/ 169 (5.2%)	<0.0001	12/56/ 21 (13.5%)	40/345/ 135 (7.7%)	0.13
Stroke status (<i>n</i> for positive/negative/missing, % positive)	6/258/ 60 (1.9%)	25/1401/ 121 (1.6%)	0.74	3/188/ 43 (1.3%)	14/945/ 68 (1.4%)	1.00	3/70/ 16 (3.4%)	11/456/ 53 (2.1%)	0.63

The significance of differences between the Black and white groups are shown. *p* values were unadjusted from two-sided Wilcoxon rank sum test and Chi-square test for quantitative and categorical characteristics, respectively.

^aData on baseline cognitive status is missing for one of *N* = 324 Black participants.

^bA sub-cohort had plasma Aβ measures from at least two samples.

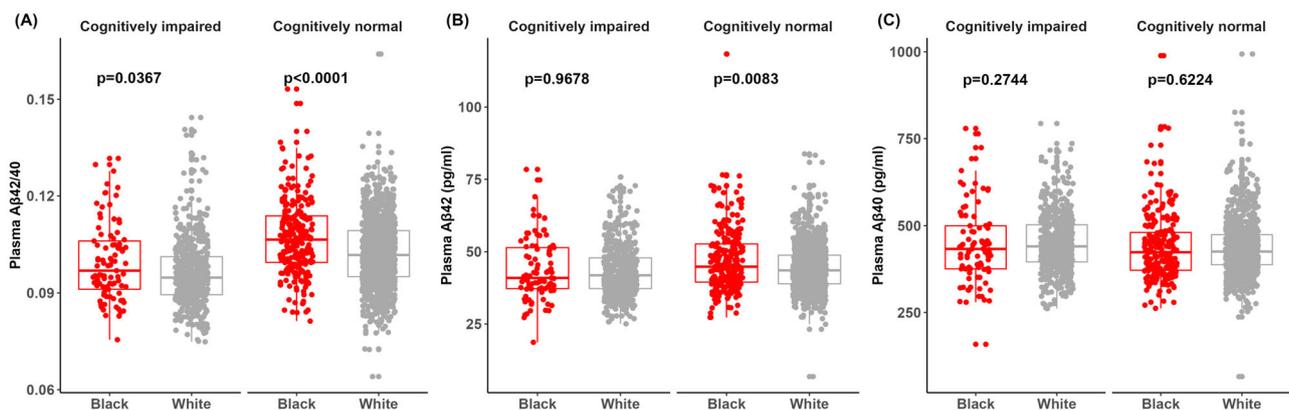


Fig. 1 | Plasma Aβ values stratified by race and cognitive status. Values for plasma Aβ42/40 (A), Aβ42 (B), and Aβ40 (C) are shown for Black (*N* = 323) and white (*N* = 1547) participants stratified by cognitive status. The box indicates 50% of the data from the 25% quantile to the 75% quantile, with the horizontal lines representing the median. The individual data points were jittered horizontally for

better visualization. The whiskers extend to 1.5 times of the 25% and 75% quantile. Overlaid are individual data points with gray for white participants and red for Black participants. The significance of unadjusted racial differences was evaluated by two-sided Wilcoxon rank sum tests with unadjusted *p* values reported. Source data are provided as a Source Data file.

p-tau181 levels were less abnormal in Black individuals compared to white individuals (*t*-tau: 114.2 ± 83.1 versus 198.1 ± 82.9 pg/mL, *p* = 0.001; *p*-tau181: 17.42 ± 12.31 versus 28.46 ± 12.27 pg/mL, *p* = 0.004).

Racial differences in the relationships of plasma biomarkers with CSF biomarkers, amyloid PET, and cognitive composite scores

Spearman correlations of plasma Aβ42/40, Aβ42 and Aβ40 with CSF biomarkers, amyloid PET, and cognitive composite scores were

examined within each group and compared across Black and white groups (Fig. 2). For both Black and white groups, the strongest correlation was between plasma Aβ42/40 and CSF Aβ42/40, but Black individuals had a weaker correlation ($\rho = 0.51$ versus 0.64, difference -0.13 , raw *p* < 0.05). This racial difference persisted after adjustment for covariates ($\rho = 0.38$ versus 0.49, difference -0.11 , raw *p* < 0.05) (Supplementary Fig. 1), but was no longer significant after FDR-based multiplicity adjustment. In fact, no significant differences in the partial correlations of plasma Aβ biomarkers with CSF biomarkers and

Table 2 | Adjusted mean baseline biomarker values for groups of Black and white individuals

Biomarker	Group	N	Mean adjusted value	Standard error	p
Plasma Aβ42/Aβ40	Black	214	0.1201	0.0030	
	White	1113	0.1155	0.0030	
	Difference (Black–white)		0.0046	0.0008	<0.0001
Plasma Aβ42 (pg/mL)	Black	214	26.25	2.40	
	White	1113	26.41	2.36	
	Difference (Black–white)		−0.16	0.61	0.80
Plasma Aβ40 (pg/mL)	Black	214	177.8	22.7	
	White	1113	199.5	22.4	
	Difference (Black–white)		−21.6	5.7	0.0002
Amyloid PET Centiloid	Black	89	21.6	13.6	
	White	626	28.2	13.4	
	Difference (Black–white)		−6.6	3.7	0.072
CSF Aβ42/40	Black	80	0.1122	0.0068	
	White	806	0.1069	0.0068	
	Difference (Black–white)		0.0053	0.0021	0.014
CSF Aβ42 (pg/mL)	Black	80	840	117	
	White	806	915	116	
	Difference (Black–white)		−75	37	0.041
CSF Aβ40 (pg/mL)	Black	80	5552	1277	
	White	806	7284	1272	
	Difference (Black–white)		−1732	400	<0.0001
CSF t-tau (pg/mL)	Black	80	114.2	83.1	
	White	806	198.1	82.9	
	Difference (Black–white)		−84.0	26.1	0.001
CSF p-tau181 (pg/mL)	Black	80	17.42	12.31	
	White	806	28.46	12.27	
	Difference (Black–white)		−11.04	3.86	0.004

The linear regression models included the main effects of race and the covariates of age, sex, APOE ε4 carrier status, years of education, cognitive status (unimpaired, CDR 0; or impaired, CDR > 0), fasting status if plasma biomarker (fasting or non-fasting), BMI, and status for hypertension, diabetes, and stroke (positive or negative). The two-sided t-test raw p-values testing for the significance of differences between the Black and white groups are shown.

amyloid PET and cognitive measures were found across racial groups after the multiplicity adjustment.

Racial differences in the longitudinal changes of plasma Aβ biomarkers

A sub-cohort of 158 Black participants and 759 white participants had plasma Aβ measures from at least two samples with a mean interval between the first and last plasma sample of 5.11 ± 3.52 years for Black individuals and 6.93 ± 4.17 years for white individuals. Longitudinal trajectories of plasma Aβ42, Aβ40, and Aβ42/40 appeared relatively linear (Fig. 3), justifying the use of linear models (Supplementary Fig. 2). Covariate-adjusted linear mixed-effects models were used to estimate the rate of change of plasma Aβ in 112 Black and 566 white individuals (Table 3). Plasma Aβ42/40 decreased longitudinally in Black and white individuals at a rate that did not vary by race ($p = 0.38$).

However, plasma Aβ42 increased longitudinally at a faster rate in Black compared to white individuals (0.5719 ± 0.1142 versus 0.2642 ± 0.0495 pg/mL/year, $p = 0.013$). Plasma Aβ40 also increased at a faster rate in Black compared to white individuals (7.357 ± 1.047 versus 5.464 ± 0.454 pg/mL/year, $p = 0.093$), although this difference did not reach statistical significance.

Further analyses examined whether racial differences in the rate of change of plasma Aβ were modified by baseline amyloid status, age, sex, APOE ε4 carrier status, years of education, cognitive status, BMI, hypertension, or diabetes. Amyloid status by CSF or amyloid PET did not affect racial differences in the rate of change of plasma Aβ42/40, Aβ42, or Aβ40 (Supplementary Table 5). Further, no other covariates significantly affected racial differences in the rate of change of plasma Aβ42/40 (Supplementary Table 6). However, for plasma Aβ42 and Aβ40, there was a significant interaction between the racial group and baseline age such that younger but not older Black participants had a faster increase in Aβ42 ($p = 0.0059$, Supplementary Table 7) and Aβ40 ($p = 0.018$, Supplementary Table 8) than the white counterparts. Overall, plasma Aβ42/40 changed relatively consistently across racial groups despite multiple potentially confounding factors.

Discussion

This study examined potential differences in baseline levels and rates of longitudinal change in plasma Aβ measures (Aβ42, Aβ40, and Aβ42/40) in self-identified Black and white participants, in one of the largest cohorts studied so far. Black participants had higher average plasma Aβ42/40 levels at baseline than white participants, consistent with Black participants having a lower average level of amyloid pathology. Despite the baseline differences, the Black and white individuals had similar longitudinal rates of change in plasma Aβ42/40, consistent with a similar rate of amyloid accumulation in both groups.

The finding of the lower average level of amyloid pathology in Black individuals compared to white individuals aligns with three recent CSF and imaging biomarker studies. One imaging study of 144 Black and 3689 white cognitively normal individuals reported that the Black participants had a lower rate of amyloid positivity and lower average amyloid burden¹⁵. A second imaging study with 635 Black and 15,322 white cognitively impaired individuals reported that Black participants were less likely to be amyloid PET positive¹⁷. In a cohort overlapping with the current study cohort that included 266 Black and 1977 white participants with CSF biomarkers, Black participants had less abnormality of multiple CSF biomarkers, including Aβ42/40, total tau, p-tau181, and neurofilament light²¹. Other studies have found the opposite result or no differences between these groups^{22,23}. It is possible that differences in study findings could be related to variable methods of recruitment³⁷. Still, growing evidence suggests that Black individuals have a lower average level of amyloid pathology compared to white individuals, at least within the population of individuals in AD research studies and clinical trials.

In our current study, Black participants had higher mean levels of plasma Aβ42/40 than white participants regardless of amyloid status, sex, APOE ε4 carrier status, years of education, BMI, hypertension, or diabetes status. However, the possibility remains that variables not currently in our dataset, including additional medical conditions (e.g., chronic kidney disease) and/or social and structural determinants of health, might explain these racial differences³⁸. Until these factors are identified and included in datasets for analyses, the significant effect of race in models of plasma Aβ42/40, despite adjustment for key covariates, implies that statistical analyses of plasma AD biomarkers are more valid when race is included as a variable. Moreover, the converging findings across plasma biomarkers, CSF biomarkers, and amyloid PET suggest that Black individuals may truly have a lower average level of amyloid pathology compared to white individuals for reasons that we do not yet fully understand.



Fig. 2 | Correlations of plasma Aβ biomarkers with CSF biomarkers, amyloid PET, and cognitive composites. Spearman correlations between plasma Aβ biomarkers and CSF biomarkers, amyloid PET, or cognitive composite are shown for Black (A) and white (B) individuals. Racial differences in correlations (Black–white) are shown in (C). Only significant correlations or differences (raw unadjusted

$p < 0.05$) from the two-sided correlation test are shown. Colored blocks visualize correlation direction and magnitude, with warm to cold colors representing most negative correlations to most positive correlations and darker colors and larger block sizes corresponding to greater absolute magnitudes. Source data are provided as a Source Data file.

Table 3 | Estimated annual rates of longitudinal change in plasma Aβ

Biomarker	Black N = 112			White N = 566			Difference (Black–white)		
	Slope	SE	p	Slope	SE	p	Slope	SE	p
Plasma Aβ42/40	-0.00047	0.0001	0.002	-0.00060	0.00006	<0.0001	0.00013	0.00015	0.38
Plasma Aβ42	0.5719	0.1142	<0.0001	0.2642	0.0495	<0.0001	0.3076	0.1226	0.013
Plasma Aβ40	7.357	1.047	<0.0001	5.464	0.454	<0.0001	1.893	1.125	0.093

The linear mixed-effects models included the main effects of race and time, and race by time interaction, as well as the covariates of age, sex, APOE ε4 carrier status, years of education, cognitive status (unimpaired, CDR 0; or impaired, CDR > 0), fasting status (fasting or non-fasting), BMI, and status for hypertension and diabetes (positive or negative). Whether the slope is significantly different from zero for Black and white participants, and the significance of racial differences from two-sided t-test are shown.

Lower average levels of amyloid pathology in Black individuals may have a major impact on efforts to make AD clinical trials more representative. A recent study of four AD clinical trials that required amyloid positivity for inclusion found much lower eligibility of Black individuals¹⁸. There have been suggestions to decrease the level of amyloid biomarker abnormality required for Black individuals to increase enrollment in this under-represented group. However, the use of race-specific cut-offs could have unintended consequences and even lead to systematic racial discrimination³⁹. For example, if Black individuals included in clinical trials had less amyloid pathology, they may be less responsive to amyloid-lowering treatments, potentially leading to the erroneous conclusion that the treatments are not effective in Black individuals. These concerns provide a strong rationale for not using race-specific cut-offs, even if the intent is to increase the representation of Black individuals in AD clinical trials.

Interestingly, the higher average plasma Aβ42/40 levels in Black participants resulted from lower levels of plasma Aβ40 but similar levels of plasma Aβ42 compared to white participants. Aligned with this finding, CSF Aβ40 levels were much lower in Black participants compared to white participants. Importantly, it is the ratio of Aβ42 to Aβ40 that reflects sequestration of Aβ42 into amyloid plaques and is most strongly associated with amyloid plaque burden^{40,41}. Recent work has found that individuals have significant differences in overall levels

of CSF proteins that are driven by non-AD related factors and that CSF Aβ40 is a useful measure of an individual’s overall CSF protein level^{42,43}. The reasons for differences in overall levels of CSF proteins are not yet well understood but may be related to physiological factors in CSF production and clearance that are associated with age, sex, ventricular volumes, and circadian rhythms^{43,44}. Additional factors may influence the proportion of central nervous system-derived proteins that reach the plasma⁴⁵. Normalizing fluid biomarkers of AD (e.g., CSF p-tau181) to Aβ40 may decrease non-AD-related variance and improve associations with AD pathology^{42,43}. Our findings suggest that Black individuals have lower levels of CSF and plasma Aβ40. It is possible that normalization to Aβ40 may reduce racial differences in biomarkers of AD pathology^{16,28}.

An important finding of this study was that the longitudinal rate of change in plasma Aβ42/40 did not vary significantly between groups of Black and white individuals, despite racial differences in baseline plasma Aβ42/40. While this finding must be replicated in other cohorts, this suggests that while a lower average level of amyloid pathology may result in lower enrollment of Black participants in studies and trials that use biomarkers of amyloid pathology as inclusion criteria, once participants are enrolled and randomized, changes in plasma Aβ42/40 may be consistent across racial groups. Furthermore, changes in plasma Aβ42/40 were not differentially affected by

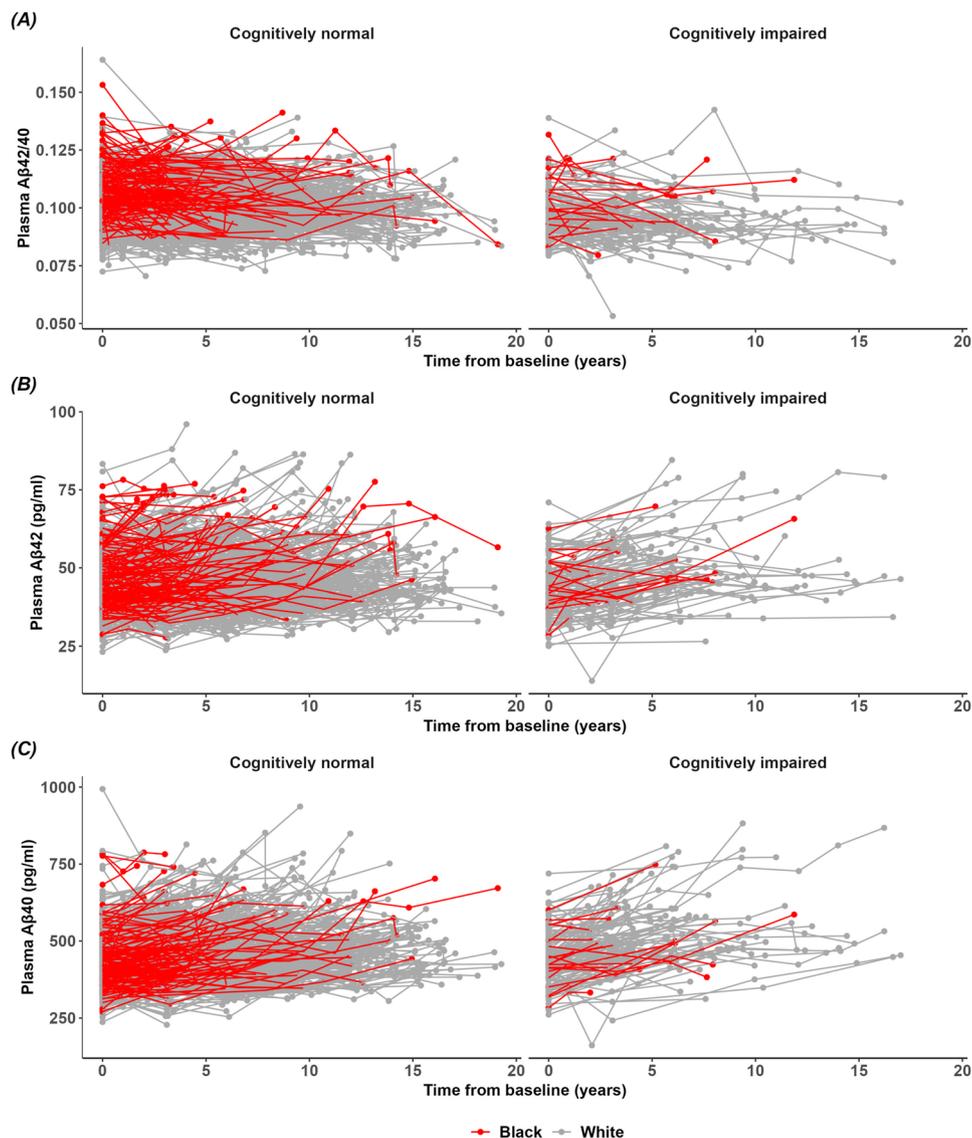


Fig. 3 | Spaghetti plots of plasma biomarkers stratified by cognitive status. Plasma A β 42/40 (panel A), A β 42 (panel B), and A β 40 (panel C) were plotted against time since baseline between cognitively normal (CDR 0, left panel) and impaired

(CDR > 0, right panel) Black ($N = 158$) and white ($N = 759$) participants. Each line represents an independent individual connecting longitudinal data points, gray for white participants and red for Black participants.

race for most covariates except cognitive status. The consistency in the rate of change may allow plasma A β 42/40 to be used in cohorts with Black and white participants to establish the effect of treatments on biomarkers. Specifically, the placebo arm in future clinical trials may estimate the same rate of change in plasma A β 42/40 for both Black and white groups.

This study has multiple major strengths. The cohort included a relatively large number of Black participants from three sites that had similar inclusion and exclusion criteria and used a uniform clinical assessment protocol⁴⁶. Plasma samples were collected according to standard protocols that did not vary by racial group, and samples were assayed together at a single lab. The plasma A β assay, PrecivityADTM, was previously shown to accurately and consistently classify amyloid status in an overlapping cohort with Black and white participants¹⁶. This test is currently being used in clinical trials as well as in clinical care³⁶, making our results of interest to researchers, clinical trialists, and clinicians. Limitations of our study include that AD research cohorts are not representative of the general population and do not represent a random sample from the community. There was very limited data on structural and social

determinants of health, including socioeconomic status, especially life course experience, and discrimination. Some individuals were missing data on covariates, reducing the number of individuals in covariate-adjusted analyses and, hence, statistical power. Further, data on more recently developed plasma p-tau217 assays were not currently available for most samples in our cohort; an examination of racial differences in plasma p-tau217 is planned once sufficient data is available. Finally, it is possible that an even larger cohort with longer longitudinal follow-up might reveal racial differences in the rate of change of plasma A β 42/40.

In summary, we found that Black research participants have higher average plasma A β 42/40 at baseline, consistent with less amyloid pathology, compared to white participants. Interestingly, despite these racial differences at baseline, the rate of change of plasma A β 42/40 was consistent in both Black and white groups. Further, plasma A β 42/40 had relatively consistent associations with CSF A β 42/40, amyloid PET, and cognitive measures across both groups. These results suggest that plasma A β 42/40 may be useful in providing a biomarker outcome for research and clinical trials that is consistent across racial groups.

Methods

Ethical approval

All participants provided written informed consent at recruitment from their parent studies. The Washington University Human Research Protection Office approved the current study with additional approvals from the Institutional Review Boards of the other sites.

Participants

The study cohort included individuals with plasma A β measures and clinical/cognitive data who participated in the Study of Race to Understand Alzheimer Biomarkers (SORTOUT-AB; NIH/NIA R01 AG067505), which aims to understand potential racial differences in harmonized biomarker data collected by multiple research studies of memory and aging in middle-aged and older individuals²¹. Participants in the current study represented three of the SORTOUT-AB sites: the Washington University (WU) Knight Alzheimer's Disease Research Center (ADRC), the University of Pennsylvania (UPenn) ADRC, and the University of Alabama at Birmingham (UAB) ADRC. Details of recruitment for these studies have been described previously^{20,21,47}. Participants with conditions that could prevent participation or affect long-term participation (e.g., metastatic cancer) were excluded. Participants underwent clinical and/or cognitive assessments within 2 years of their baseline plasma assessments. A sub-cohort of the participants also had CSF or imaging assessments within 2 years of their baseline plasma sample collection.

Clinical and cognitive assessments

Clinical and cognitive assessments from WU, UPenn, and UAB followed protocols consistent with the National Alzheimer's Coordinating Center Uniform Data Set (UDS)⁴⁶. Demographic information, body mass index (BMI), and medical history were collected. Race and sex were self-identified by participants. Cognitive impairment was scored with the Clinical Dementia Rating[®]™ (CDR[®]™)⁴⁸. Individuals with a CDR of 0 were categorized as cognitively unimpaired. Individuals with a CDR of 0.5 or greater were categorized as cognitively impaired, and the probable etiology was formulated by clinicians based on clinical features in accordance with standard criteria⁴⁹. The cognitive battery of the UDS included tasks of episodic memory, working memory, semantic knowledge, executive function and attention, and visuospatial ability, and were harmonized across UDS versions⁵⁰. Global cognitive and episodic memory composite scores were calculated as previously described²¹.

Apolipoprotein E genotyping

Apolipoprotein E (*APOE*) genotyping was performed as previously described⁵¹. Participants were classified as *APOE* ϵ 4 carriers (one or two ϵ 4 alleles) and non-carriers.

Blood collection and analysis

At WU, blood was collected from non-fasting participants at the time of clinical assessment or fasting participants at the time of lumbar puncture (LP)⁵². Blood was collected from non-fasting participants at UPenn and fasting participants at UAB at the time of clinical assessment. Blood was collected in EDTA-containing tubes and centrifuged to separate plasma from blood cells. Plasma was aliquoted into polypropylene tubes and stored at -80°C until analysis.

All plasma samples were analyzed at C₂N Diagnostics with the PrecivityAD assay, which has previously been described^{53,54}. Briefly, A β 40 and A β 42 were simultaneously immunoprecipitated from plasma via a monoclonal anti-A β mid-domain antibody⁵³. Proteins were digested into peptides using LysN endoprotease. Liquid chromatography–mass spectrometry was performed on a Thermo Scientific Orbitrap Lumos Tribrid mass spectrometer interfaced with a nano-Acquity chromatography system (LC–MS/MS)⁵³.

CSF collection and analysis

At WU, CSF samples (20–30 mL) were collected at 8 AM after overnight fasting by gravity drip, briefly centrifuged at low speed, and aliquoted into polypropylene tubes prior to freezing at -80°C . CSF samples from participants enrolled at the UPenn and UAB ADRCs were collected in accordance with protocols for the Alzheimer's Disease Neuroimaging Initiative (ADNI)⁵⁵.

An automated immunoassay (LUMIPULSE G1200, Fujirebio, Malverne, PA) was used to measure CSF concentrations of A β 40, A β 42, total tau (t-tau), and tau phosphorylated at position 181 (p-tau181)^{56,57}. A bridging subset of the CSF samples ($n = 114$) from the UPenn ADRC was selected to represent a wide range of values for all analytes and was run at the same time and with the same reagents as the WU samples to evaluate and adjust for systematic differences between the WU and UPenn sites. A model fitted on the values of the bridging samples was used to harmonize the CSF biomarker values between WU and UPenn⁵⁸. Positive CSF biomarker status was defined as a CSF A β 42/40 value < 0.0673 ⁵⁷.

Imaging processing and analysis

Structural brain MRI and amyloid PET protocols were consistent with those used by the ADNI²¹. A standardized uptake value ratio (SUVR) with correction for partial volume effects was calculated for the FreeSurfer regions of interest (ROIs) for PiB, Florbetapir, or Florbetaben⁵⁹. The cerebellum was used as the reference region. A summary measure of amyloid burden was calculated using the averaged SUVR values in the lateral orbitofrontal, medial orbitofrontal, precuneus, rostral middle frontal, superior frontal, superior temporal, and middle temporal regions. To harmonize SUVR values across different tracers (PiB, Florbetapir, or Florbetaben), values from the summary measure were converted into Centiloid units⁶⁰. Positive amyloid PET status was defined as a Centiloid value > 20 ³⁶.

Statistical analyses

The baseline characteristics of participants were summarized with the mean and SD for continuous variables or count and percentage for categorical variables. General linear models were implemented to evaluate for cross-sectional racial differences in levels of fluid or imaging biomarkers. These models included the main effects of race and the covariates of age, sex, *APOE* ϵ 4 carrier status, years of education, cognitive status (unimpaired, CDR 0; or impaired, CDR > 0), fasting status if plasma biomarker (fasting or non-fasting), BMI, and status for hypertension, diabetes, and stroke (positive or negative). Additional analyses included interactions of each variable with the racial group to examine whether each of the covariates modified the racial differences; stroke status was not included in these interaction models due to a small number of individuals with stroke.

Fluid biomarker measures were correlated with the established AD biomarkers by Spearman correlations, and the correlations between groups were compared by a two-sided standard normal test after Fisher's *Z*-transformation⁶¹. Because of the large number of comparisons in the correlations, we adjusted statistical significance for a false discovery rate (FDR)⁶² of 5%.

General linear mixed models with random slopes and intercepts were implemented to evaluate for racial differences in the longitudinal rates of change of fluid biomarker measures⁶³. All models included race and a race-by-time interaction (at baseline time = 0), and the same covariates as in the cross-sectional models. The annual rates of change between groups were compared by a two-sided approximate Student *t*-test; the degree of freedom was estimated by the Satterthwaite method. All models were implemented in Rstudio (version 2023.9.1.494 running R version 4.2.1) via the R package *lmerTest* (version 3.1-3). An assessment found no clear non-linear longitudinal patterns, likely because of the small number of plasma samples for most individuals.

Reporting summary

Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

Data availability

Anonymized data that support the findings of this study are available from the corresponding author and co-first authors upon request from qualified investigators. Briefly, a formal application should be submitted to provide the rationale and objectives of the research. Data will be provided after an application is reviewed and approved. The study participants provided their data with these conditions for use of the data. Source data are provided with this paper.

Code availability

Publicly available software, Rstudio, was used for all analyses as described in the “Methods” section and the accompanying Reporting Summary.

References

- Schindler, S. E. & Atri, A. The role of cerebrospinal fluid and other biomarker modalities in the Alzheimer’s disease diagnostic revolution. *Nat. Aging* **3**, 460–462 (2023).
- Johnson, K. A. et al. Appropriate use criteria for amyloid PET: a report of the Amyloid Imaging Task Force, the Society of Nuclear Medicine and Molecular Imaging, and the Alzheimer’s Association. *Alzheimer’s Dement.* **9**, e1-16 (2013).
- Shaw, L. M. et al. Appropriate use criteria for lumbar puncture and cerebrospinal fluid testing in the diagnosis of Alzheimer’s disease. *Alzheimer’s Dement.* **14**, 1505–1521 (2018).
- Bonomi, S., Gupta, M. R. & Schindler, S. E. Inadequate reimbursement for lumbar puncture is a potential barrier to accessing new Alzheimer’s disease treatments. *Alzheimer’s Dement.* **19**, 5849–5851 (2023).
- Lewis, A. et al. Association between socioeconomic factors, race, and use of a specialty memory clinic. *Neurology* **101**, e1424–e1433 (2023).
- Hampel, H. et al. Blood-based biomarkers for Alzheimer’s disease: current state and future use in a transformed global healthcare landscape. *Neuron* **111**, 2781–2799 (2023).
- Hansson, O., Blennow, K., Zetterberg, H. & Dage, J. Blood biomarkers for Alzheimer’s disease in clinical practice and trials. *Nat. Aging* **3**, 506–519 (2023).
- Karikari, T. K. Blood tests for Alzheimer’s disease: increasing efforts to expand and diversify research participation is critical for widespread validation and acceptance. *J. Alzheimer’s Disease* **90**, 967–974 (2022).
- Babulal, G. M. et al. Perspectives on ethnic and racial disparities in Alzheimer’s disease and related dementias: update and areas of immediate need. *Alzheimer’s Dement.* **15**, 292–312 (2019).
- Adkins-Jackson, P. B. et al. The structural and social determinants of Alzheimer’s disease related dementias. *Alzheimer’s Dement.* **19**, 3171–3185 (2023).
- Howell, J. C. et al. Research lumbar punctures among African Americans and Caucasians: perception predicts experience. *Front. Aging Neurosci.* **8**, 296 (2016).
- Kornblith, E. et al. Association of race and ethnicity with incidence of dementia among older adults. *JAMA* **327**, 1488–1495 (2022).
- Power, M. C. et al. Trends in relative incidence and prevalence of dementia across non-Hispanic Black and White individuals in the United States, 2000–2016. *JAMA Neurol.* **78**, 275–284 (2021).
- Manly, J. J. et al. Estimating the prevalence of dementia and mild cognitive impairment in the US: the 2016 Health and Retirement Study Harmonized Cognitive Assessment Protocol Project. *JAMA Neurol.* **79**, 1242–1249 (2022).
- Deters, K. D. et al. Amyloid PET imaging in self-identified non-Hispanic Black participants of the anti-amyloid in asymptomatic Alzheimer’s Disease (A4) study. *Neurology* **96**, e1491–e1500 (2021).
- Schindler, S. E. et al. Effect of race on prediction of brain amyloidosis by plasma A β ₄₂/A β ₄₀, phosphorylated tau, and neurofilament light. *Neurology* **99**, e245–e257 (2022).
- Wilkins, C. H. et al. Racial and ethnic differences in amyloid PET positivity in individuals with mild cognitive impairment or dementia: a secondary analysis of the Imaging Dementia-Evidence for Amyloid Scanning (IDEAS) Cohort Study. *JAMA Neurol.* **79**, 1139–1147 (2022).
- Grill, J. D. et al. Eligibility rates among racially and ethnically diverse US participants in Phase 2 and Phase 3 placebo-controlled, double-blind, randomized trials of lecanemab and elenbecestat in early Alzheimer disease. *Ann. Neurol.* **95**, 288–298 (2024).
- Garrett, S. L. et al. Racial disparity in cerebrospinal fluid amyloid and tau biomarkers and associated cutoffs for mild cognitive impairment. *JAMA Netw. Open* **2**, e1917363 (2019).
- Morris, J. C. et al. Assessment of racial disparities in biomarkers for Alzheimer disease. *JAMA Neurol.* **76**, 264–273 (2019).
- Bonomi, S. et al. Relationships of cognitive measures with cerebrospinal fluid but not imaging biomarkers of Alzheimer disease vary between Black and White individuals. *Ann. Neurol.* **95**, 495–506 (2023).
- Gottesman, R. F. et al. The ARIC-PET amyloid imaging study: brain amyloid differences by age, race, sex, and APOE. *Neurology* **87**, 473–480 (2016).
- Windon, C. et al. Comparison of plasma and CSF biomarkers across ethnorracial groups in the ADNI. *Alzheimer’s Dement.* **14**, e12315 (2022).
- Xiong, C., Luo, J., Coble, D., Agboola, F., Kukull, W. & Morris, J. C. Complex interactions underlie racial disparity in the risk of developing Alzheimer’s disease dementia. *Alzheimer’s Dement.* **16**, 589–597 (2020).
- Xiong, C., et al. Racial differences in longitudinal Alzheimer’s disease biomarkers among cognitively normal adults. *Alzheimer’s Dement.* **18**, 2570–2581 (2022).
- Barnes, L. L. Alzheimer disease in African American individuals: increased incidence or not enough data? *Nat. Rev. Neurol.* **18**, 56–62 (2022).
- Mielke, M. M. et al. Performance of plasma phosphorylated tau 181 and 217 in the community. *Nat. Med.* **28**, 1398–1405 (2022).
- Schindler, S. E. & Karikari, T. K. Comorbidities confound Alzheimer’s blood tests. *Nat. Med.* **28**, 1349–1351 (2022).
- Syrjanen, J. A. et al. Associations of amyloid and neurodegeneration plasma biomarkers with comorbidities. *Alzheimer’s Dement.* **18**, 1128–1140 (2022).
- Janelidze, S., Barthelemy, N. R., He, Y., Bateman, R. J. & Hansson, O. Mitigating the associations of kidney dysfunction with blood biomarkers of Alzheimer disease by using phosphorylated tau to total tau ratios. *JAMA Neurol.* **80**, 516–522 (2023).
- Brickman, A. M. et al. Plasma p-tau181, p-tau217, and other blood-based Alzheimer’s disease biomarkers in a multi-ethnic, community study. *Alzheimer’s Dement.* **17**, 1353–1364 (2021).
- Ramanan, V. K. et al. Association of plasma biomarkers of Alzheimer disease with cognition and medical comorbidities in a biracial cohort. *Neurology* **101**, e1402–e1411 (2023).
- Hajjar, I. et al. Association of plasma and cerebrospinal fluid Alzheimer disease biomarkers with race and the role of genetic ancestry, vascular comorbidities, and neighborhood factors. *JAMA Netw. Open* **5**, e2235068 (2022).
- Mohs, R. C. et al. The Bio-Hermes Study: biomarker database developed to investigate blood-based and digital biomarkers in

- community-based, diverse populations clinically screened for Alzheimer's disease. *Alzheimer's Dement.* **20**, 2752–2765 (2024).
35. Monane, M. et al. A blood biomarker test for brain amyloid impacts the clinical evaluation of cognitive impairment. *Ann. Clin. Transl. Neurol.* **10**, 1738–1748 (2023).
 36. Rissman, R. A. et al. Plasma A β 42/A β 40 and phospho-tau217 concentration ratios increase the accuracy of amyloid PET classification in preclinical Alzheimer's disease. *Alzheimer's Dement.* **20**, 1214–1224 (2024).
 37. Gleason, C. E. et al. Association between enrollment factors and incident cognitive impairment in Blacks and Whites: data from the Alzheimer's Disease Center. *Alzheimer's Dement.* **15**, 1533–1545 (2019).
 38. Wilkins, C. H., Schindler, S. E. & Morris, J. C. Addressing health disparities among minority populations: why clinical trial recruitment is not enough. *JAMA Neurol.* **77**, 1063–1064 (2020).
 39. Powe, N. R. Black kidney function matters: use or misuse of race? *JAMA* **324**, 737–738 (2020).
 40. Wisch, J. K. et al. Predicting continuous amyloid PET values with CSF and plasma Abeta42/Abeta40. *Alzheimer's Dement.* **15**, e12405 (2023).
 41. Patterson, B. W. et al. Age and amyloid effects on human central nervous system amyloid-beta kinetics. *Ann. Neurol.* **78**, 439–453 (2015).
 42. Guo, T. et al. Normalization of CSF pTau measurement by Abeta(40) improves its performance as a biomarker of Alzheimer's disease. *Alzheimers Res. Ther.* **12**, 97 (2020).
 43. Karlsson, L. et al. Cerebrospinal fluid reference proteins increase accuracy and interpretability of biomarkers for brain diseases. *Nat. Commun.* **15**, 3676 (2024).
 44. Lucey, B. P. et al. Associations between beta-amyloid kinetics and the beta-amyloid diurnal pattern in the central nervous system. *JAMA Neurol.* **74**, 207–215 (2017).
 45. Roberts, K. F. et al. Amyloid-beta efflux from the central nervous system into the plasma. *Ann. Neurol.* **76**, 837–844 (2014).
 46. Morris, J. C. et al. The Uniform Data Set (UDS): clinical and cognitive variables and descriptive data from Alzheimer Disease Centers. *Alzheimer Dis. Assoc. Disord.* **20**, 210–216 (2006).
 47. Wolk, D. A., Mancuso, L., Klot, D., Arnold, S. E. & Dickerson, B. C. Familiarity-based memory as an early cognitive marker of pre-clinical and prodromal AD. *Neuropsychologia* **51**, 1094–1102 (2013).
 48. Morris, J. C. The Clinical Dementia Rating (CDR): current version and scoring rules. *Neurology* **43**, 2412–2414 (1993).
 49. McKhann, G. M. et al. The diagnosis of dementia due to Alzheimer's disease: recommendations from the National Institute on Aging-Alzheimer's Association workgroups on diagnostic guidelines for Alzheimer's disease. *Alzheimer's Dement.* **7**, 263–269 (2011).
 50. Monsell, S. E. et al. Results from the NACC uniform data set neuropsychological battery crosswalk study. *Alzheimer Dis. Assoc. Disord.* **30**, 134–139 (2016).
 51. Pastor, P. et al. Apolipoprotein Epsilon4 modifies Alzheimer's disease onset in an E280A PS1 kindred. *Ann. Neurol.* **54**, 163–169 (2003).
 52. Schindler, S. E. et al. High-precision plasma beta-amyloid 42/40 predicts current and future brain amyloidosis. *Neurology* **93**, e1647–e1659 (2019).
 53. Hu, Y. et al. Assessment of a plasma amyloid probability score to estimate amyloid positron emission tomography findings among adults with cognitive impairment. *JAMA Netw. Open* **5**, e228392 (2022).
 54. West, T. et al. A blood-based diagnostic test incorporating plasma Abeta42/40 ratio, ApoE proteotype, and age accurately identifies brain amyloid status: findings from a multi cohort validity analysis. *Mol. Neurodegener.* **16**, 30 (2021).
 55. Shaw, L. M. et al. Cerebrospinal fluid biomarker signature in Alzheimer's disease neuroimaging initiative subjects. *Ann. Neurol.* **65**, 403–413 (2009).
 56. Alcolea, D. et al. Agreement of amyloid PET and CSF biomarkers for Alzheimer's disease on Lumipulse. *Ann. Clin. Transl. Neurol.* **6**, 1815–1824 (2019).
 57. Barthélemy, N. R. et al. CSF tau phosphorylation occupancies at T217 and T205 represent improved biomarkers of amyloid and tau pathology in Alzheimer's disease. *Nat. Aging* **3**, 391–401 (2023).
 58. Henson, R. L. et al. A methodology for normalizing fluid biomarker concentrations across reagent lots. *Alzheimer's Dement.* **18**, e066912 (2022).
 59. Rousset, O. G., Ma, Y. & Evans, A. C. Correction for partial volume effects in PET: principle and validation. *J. Nucl. Med.* **39**, 904–911 (1998).
 60. Su, Y. et al. Utilizing the Centiloid scale in cross-sectional and longitudinal PiB PET studies. *Neuroimage Clin.* **19**, 406–416 (2018).
 61. Graybill, F. A. *Theory and Application of the Linear Model* (Duxbury, North Scituate, MA, 1976).
 62. Benjamini, Y. & Hochberg, Y. Controlling the false discovery rate—a practical and powerful approach to multiple testing. *J. R. Stat. Soc. B* **57**, 289–300 (1995).
 63. Laird, N. M. & Ware, J. H. Random-effects models for longitudinal data. *Biometrics* **38**, 963–974 (1982).

Acknowledgements

The authors thank the research volunteers who participated in the studies from which these data were obtained and their families. The authors express gratitude to Dr. Anne M. Fagan for establishing the large WU biorepository used in this study. We acknowledge the WU, UPenn, and UAB ADRC biospecimen cores. The authors thank C₂N Diagnostics for processing the plasma samples and conducting the QC of the plasma biomarker data. This work was supported in part by funding from the National Institutes of Health grants R01 AG067505 (PI: Xiong), R01 AG070941 (PI: Schindler), P30 AG066444 (PI: Holtzman), P01 AG026276 (PI: Morris), P01 AG003991 (PI: Morris), P30 AG072979 (PI: David Wolk), P20 AG068024 (PI: Erik Roberson), and R44 AG059489 (C₂N Diagnostics). Additional funding was provided by BrightFocus (CA2016636), the Gerald and Henrietta Rauenhurst Foundation, the Cure Alzheimer's Fund (PI: Moulder), and the Alzheimer's Drug Discovery Foundation (GC-201711-2013978).

Author contributions

C.X., J.L., D.A.W., L.M.S., E.D.R., T.L.S.B., R.J.B., C.C., J.C.M., and S.E.S. contributed to the conception and design of the study. C.X., D.A.W., L.M.S., E.D.R., C.F.M., R.L.H., T.L.S.B., Q.B., F.A., E.G., E.N.G., K.L.M., D.S.G., O.J.C., C.C., D.M.H., R.J.B., J.C.M., and S.E.S. contributed to the acquisition and analysis of data. C.X., J.L., C.H.M., R.L.H., Q.B., F.A., E.N.G., K.L.M., D.S.G., O.J.C., G.B., C.C., D.M.H., R.J.B., J.C.M., and S.E.S. contributed to drafting the text or preparing figures.

Competing interests

D.A.W. has served as a paid consultant for Eli Lilly, GE Healthcare and Qynapse, and serves on a DSMB for Functional Neuromodulation. E.D.R. serves on a data monitoring committee for Eli Lilly. T.L.S.B. participates as a site investigator in clinical trials sponsored by Avid Radiopharmaceuticals, Eli Lilly and Company, Biogen, Eisai, Janssen, and Roche. D.S.G. participates as a site investigator in clinical trials sponsored by Biogen and Janssen. He serves as a consultant to Eisai, Lilly, and Roche. DMH and RJB co-founded and have equity in C₂N Diagnostics. D.M.H. serves on the scientific advisory board of C2N Diagnostics, Genentech, Denali, Cajal Neurosciences, and Asteroid. S.E.S. has served on advisory boards for

Eisai. Washington University has a financial interest in C₂N Diagnostics and may financially benefit if the company is successful in marketing its product(s) that is/are related to this research. The other authors declare no competing interests.

Additional information

Supplementary information The online version contains supplementary material available at <https://doi.org/10.1038/s41467-024-49859-w>.

Correspondence and requests for materials should be addressed to Suzanne E. Schindler.

Peer review information *Nature Communications* thanks Thomas Wisniewski and the other, anonymous, reviewers for their contribution to the peer review of this work. A peer review file is available.

Reprints and permissions information is available at <http://www.nature.com/reprints>

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2024