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The Journal of Allergy and Clinical Immunology
**IL-6 serum levels predict severity and response to Tocilizumab in COVID-19: an
 observational study**
 --Manuscript Draft--

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Abstract:	<p>Background</p> <p>COVID-19 patients can develop a cytokine release syndrome that eventually leads to acute respiratory distress syndrome (ARDS) requiring invasive mechanical ventilation (IMV). Since interleukin-6 (IL-6) is a relevant cytokine in ARDS, the blockade of its receptor with Tocilizumab (TCZ) could reduce mortality and/or morbidity in severe COVID-19.</p> <p>Objective</p> <p>To determine whether baseline IL-6 serum levels can predict the need for IMV and the response to TCZ.</p> <p>Methods</p> <p>Retrospective observational study performed in hospitalized patients diagnosed of COVID-19. Clinical information and laboratory findings, including IL-6 levels, were collected approximately 3 and 9 days after admission to be matched with pre- and post-administration of TCZ. Multivariable logistic and linear regressions, and survival analysis were performed depending on outcomes: need for IMV, evolution of arterial oxygen tension/fraction of inspired oxygen ratio (PaO_2 / FiO_2) or mortality.</p> <p>Results</p> <p>One hundred and forty-six patients were studied, predominantly male (66%); median age was 63 years. Forty-four patients (30%) required IMV, and 58 patients (40%) received treatment with TCZ. IL-6 levels >30 pg/ml was the best predictor for IMV (OR: 7.1; $p < 0.001$). Early administration of TCZ was associated with improvement of oxygenation (PaO_2 / FiO_2) in patients with high IL-6 ($p = 0.048$). Patients with high IL-6 not treated with TCZ showed high mortality (HR: 4.6; $p = 0.003$), as well as those with low IL-6 treated with TCZ (HR: 3.6; $p = 0.016$). No relevant serious adverse events were observed in TCZ-treated patients.</p> <p>Conclusion</p> <p>Baseline IL-6 >30 pg/ml predicts IMV requirement in patients with COVID-19 and contributes to establish an adequate indication for TCZ administration.</p>

Dr. Cezmi A. Akdis, MD
Dr. Zuhair K. Ballas
Co-Editors-in-Chief
Journal of Allergy & Clinical Immunology

Manuscript # JACI-D-20-01110_R3

Madrid, September 7th, 2020

Dear Dr Akdis and Dr Ballas,

We completely agree with your comment regarding research on COVID-19 being a continuously evolving scenario. We consider that unpublished data provided in press releases from Roche (COVACTA study, NCT04320615) and Sanofi (ex-US study, NCT04327388) are relevant enough to be discussed within the manuscript. In light of these results, we understand that this is the right time to highlight the possible role that measuring IL-6 serum levels can play in selecting patients that can benefit from IL-6 blockade as derived from our results.

We believe our results are of importance for guiding clinicians and scientists to better select patients that can benefit from IL-6 blockade. Therefore, we resubmit our final revised manuscript entitled "IL-6 serum levels predict severity and response to Tocilizumab in COVID-19: an observational study" (#JACI-D-20-01110R3) by Dr Galvan-Roman et al., according to the editors' recommendations.

Editor's comments:

#1 "The revision addressed the previous concerns adequately."

Response: Thank you for recognizing our efforts fulfilling all of the reviewers' demands.

#2 "However, since then there have been new developments in the COVID area that need to be addressed. In particular, the COVACTA Phase III trial, a controlled trial, showed that anti-IL-6 failed to show statistically significant difference. The authors will need to address this issue. The editors believe that this manuscript offers additional data not in COVACTA; that is the correlation of the level of serum IL-6 with potential response. The editors that suggest that this finding be expanded further."

Response: We fully agree with this statement. The fact that the phase III clinical trials with tocilizumab and sarilumab for patients with severe COVID-19 did not reach statistical significance does not invalidate our results at all. In fact, we think that this lack of significance could be due to a non-optimal selection of patients, just as our work indicates. As we have commented in the discussion of our work, and highlighted in this new version, only patients with high levels of interleukin 6 respond to treatment with tocilizumab, not being effective for those with low baseline levels, despite presenting data on systemic inflammation. In accordance we have modified the following sections of the manuscript:

- a) Abstract Conclusion in line 143: "... to establish an adequate indication for TCZ administration"
- b) Capsule summary in line 177: "... and should be used to guide..."
- c) Discussion in lines 450-61: "...mortality. These observations pose the question whether it is possible that severe patients with low IL-6 included in phase 3 trials with Tocilizumab and Sarilumab account for the failure to meet their primary endpoints (29, 30). Therefore, our results support the measurement of baseline IL-6 levels in hospitalized COVID-19 patients, since in those severe or critical patients with low IL-6 levels, other cytokines such as IL-1 or tumor necrosis factor (TNF) could be driving the exacerbated inflammatory response in lungs (25). Probably, this specific group could benefit from receiving other anti-inflammatory agents such as IL-1 or TNF blockers. In this regard, the need for biomarkers of response to TCZ has been recently highlighted (31) and

neither COVACTA study (Tocilizumab trial; NCT04320615) nor ex-US Sarilumab clinical trial (NCT04327388) listed increased baseline IL-6 serum levels within the inclusion criteria. Nevertheless, both clinical trials reported a decrease in duration of hospital stay in the active arm, that was statistically significant in COVACTA (29, 30)."

- d) Discussion in lines 470-1: "use of TCZ was safe and it did not increase the number of serious bacterial infections. These findings are consistent with the results of both the COVACTA and Sarilumab trials (29, 30)."
- e) Discussion in line 491: can instead of may
- f) References in lines 639-51: As a consequence of the inclusion of the press releases of Roche and Sanofi as references 29 and 30, the previous references 29 and 30 are now 31 and 32 respectively.

#3 In particular, supplementary figure 4 should probably be in the body of the manuscript rather than in an online repository.

Response: We think that the editors have probably misinterpreted figure numbering, since there is no supplementary figure 4. In fact, we think that the Figure 4 displaying survival depending on IL-6 levels and need for TCZ is the most important figure. On the initial submission, it was panel E in figure 3. However, following the indications of reviewer 1 in the first review of the work, we gave more importance to this figure being the unique panel in figure 4. Since version R1, the importance of measuring IL-6 in order to prescribe TCZ is clearly shown in this one panel figure.

Other possibility is that editors refer to Supplementary Table 4. However, the most interesting results of this table, the effect of TCZ on PaFiO₂ depending on IL-6 levels, are displayed in a more visual way at panel C of Figure 3.

All in all, we believe that the manuscript has been significantly improved as a result of the changes we have incorporated, and hope it is now suitable for publication in the Journal of Allergy and Clinical Immunology. We are convinced that our article provides very important information for physicians to guide an early and proper prescription of IL-6R antibodies in COVID-19 patients.

Thank you for your kind consideration.

Sincerely,

Jose María Galvan-Roman

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1 **TITLE**

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4
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68

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76 interpretation of data; in the writing of the report; and in the decision to submit the article
77 for publication.

78

79 **CONFLICTS OF INTEREST**

80 IG-A reports grants from Instituto de Salud Carlos III, during the course of the study; personal
81 fees from Lilly and Sanofi; personal fees and non-financial support from BMS; personal fees
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106 Pharma, outside the submitted work.

107 CF-D reports personal fees from BMS, non-financial support from Novartis, outside the
108 submitted work.

109 DR-S reports personal fees from MSD, outside the submitted work.

110 CM-C reports competitive grants from ISCIII during the conduct of the study.

111 The remaining authors report no competing interests with the submitted work.

112

113 **RUNNING TITLE**

114 IL-6 levels predict response to TCZ in COVID-19

115

116

117 **ABSTRACT**

118 **Background**

119 COVID-19 patients can develop a cytokine release syndrome that eventually leads to acute
120 respiratory distress syndrome (ARDS) requiring invasive mechanical ventilation (IMV). Since
121 interleukin-6 (IL-6) is a relevant cytokine in ARDS, the blockade of its receptor with
122 Tocilizumab (TCZ) could reduce mortality and/or morbidity in severe COVID-19.

123 **Objective**

124 To determine whether baseline IL-6 serum levels can predict the need for IMV and the
125 response to TCZ.

126 **Methods**

127 Retrospective observational study performed in hospitalized patients diagnosed of COVID-19.
128 Clinical information and laboratory findings, including IL-6 levels, were collected
129 approximately 3 and 9 days after admission to be matched with pre- and post-administration
130 of TCZ. Multivariable logistic and linear regressions, and survival analysis were performed
131 depending on outcomes: need for IMV, evolution of arterial oxygen tension/fraction of
132 inspired oxygen ratio ($\text{PaO}_2/\text{FiO}_2$) or mortality.

133 **Results**

134 One hundred and forty-six patients were studied, predominantly male (66%); median age was
135 63 years. Forty-four patients (30%) required IMV, and 58 patients (40%) received treatment
136 with TCZ. IL-6 levels >30 pg/ml was the best predictor for IMV (OR:7.1; $p<0.001$). Early
137 administration of TCZ was associated with improvement of oxygenation ($\text{PaO}_2/\text{FiO}_2$) in
138 patients with high IL-6 ($p=0.048$). Patients with high IL-6 not treated with TCZ showed high
139 mortality (HR: 4.6; $p=0.003$), as well as those with low IL-6 treated with TCZ (HR: 3.6; $p=0.016$).
140 No relevant serious adverse events were observed in TCZ-treated patients.

141 **Conclusion**

142 Baseline IL-6>30 pg/ml predicts IMV requirement in patients with COVID-19 and contributes
143 to establish an adequate indication for TCZ administration.

144

145 **CLINICAL IMPLICATIONS**

146 Elevated levels of circulating IL-6 predict the need for IMV in severe COVID-19 patients and
147 may contribute to establish the indication for timely administration of TCZ, possibly reducing
148 ICU demand.

149

150 **KEYWORDS**

151 COVID-19, Interleukin-6, Tocilizumab, Invasive Mechanical Ventilation

152

153 **ABBREVIATIONS**

154 AEMPS: Spanish Agency for Drugs and Health Devices

155 ARDS: acute respiratory distress syndrome

156 AUC: Area under curve

157 CAR: chimeric antigen receptor

158 COPD: chronic obstructive pulmonary disease

159 COVID-19: coronavirus disease 2019

160 CRP: C-reactive protein

161 CRS: cytokine release syndrome

162 IL: interleukin

163 IMV: invasive mechanical ventilation

164 IQR: interquartile range

- 165 LR+: positive likelihood ratio; LR- negative likelihood ratio
- 166 PaO₂: arterial oxygen tension
- 167 PaO₂/FiO₂: arterial oxygen tension/fraction of inspired oxygen ratio
- 168 PCT: procalcitonin
- 169 ROC: receiver operating characteristic
- 170 SatO₂: mean oxygen saturation
- 171 SD: standard deviation
- 172 STROBE: Strengthening the Reporting of Observational Studies in Epidemiology
- 173 TCZ: Tocilizumab
- 174 TNF: tumor necrosis factor

175

176 **CAPSULE SUMMARY**

177 Baseline IL-6 serum levels >30 pg/ml identify severe COVID-19 patients and ~~can~~ should be used

178 to guide the intervention with IL-6R inhibitors, aiming to improve their use in an uncertain and

179 evolving therapeutic scenario.

180 **INTRODUCTION**

181 The recent exponential increase in cases of severe acute respiratory syndrome caused by
182 coronavirus 2 (SARS-CoV-2) has led the World Health Organization to declare a pandemic. The
183 disease, known as coronavirus disease 2019 (COVID-19), has pushed national health systems
184 to the brink of collapse, prompting national governments to impose complete population
185 lockdowns in an attempt to slow down the dynamics of infection (1, 2)

186 The spectrum of COVID-19 clinical manifestations varies widely, from mild to severe cases of
187 atypical pneumonia, some of them developing acute respiratory distress syndrome (ARDS),
188 which often requires invasive mechanical ventilation (IMV) and is the leading cause of death.

189 It is suggested that the severity of the respiratory disease caused by SARS-CoV-2 is largely due
190 to an exacerbated immune response against the virus (3, 4). This response has been observed
191 in previous respiratory virus outbreaks and can be also seen in patients treated with chimeric
192 antigen receptor (CAR) T cell therapy (5-7). Pro-inflammatory cytokines such as interleukin (IL)
193 1 and IL-6 are crucial mediators of this process (3, 6). In this regard, recent studies have
194 indicated the usefulness of IL-6 serum levels, lymphocyte count, fibrinogen or D-dimer to
195 evaluate the development of ARDS and its mortality (8-11). However, further evidence is
196 required to support these observations (12).

197 Tocilizumab (TCZ), an anti-IL-6 receptor (IL-6R) antibody, is the only drug currently licensed for
198 the treatment of the cytokine release syndrome (CRS) associated with CAR-T cell therapy (13).

199 Due to the virulence of the current outbreak, its use has been advised in severe cases of
200 COVID-19 (14-16). The rationale is to curb the deleterious effects of inflammation, thereby
201 limiting lung damage. Early promising results have prompted ongoing randomized clinical
202 trials (RCT) (17). We aimed to explore the ability of IL-6 serum levels at baseline to predict the
203 need for IMV and the response to TCZ in severe COVID-19 patients.

204 **METHODS**

205 ***Study design and population***

206 This is a retrospective observational study including 146 consecutive patients with confirmed
207 detection of SARS-CoV-2 RNA, baseline IL-6 serum level measurement and admitted to the
208 Hospital Universitario La Princesa (HUP) with severe to critical COVID-19 (18), from February
209 24th to March 23rd 2020 (Flow chart in Suppl. Fig. 1).

210
211 This time frame was decided based on the fact that after March 23rd, clinicians involved in the
212 treatment of these patients were aware of the possibility of measuring IL-6 serum levels to
213 decide treatment initiation with TCZ (RoActemra, Roche). Furthermore, around that date
214 Spanish facilities capable of providing IMV were overrun (Suppl. Fig. 2), thus the decision for
215 IMV application was based on availability rather than the patients' needs. Notwithstanding,
216 during the study period, patients were prompted to IMV when they presented a marked
217 worsening of oxygenation (arterial oxygen tension/fraction of inspired oxygen ratio
218 $[\text{PaO}_2/\text{FiO}_2] < 200$), provided that their baseline conditions did not contraindicate it.

219 220 ***Data collection***

221 Clinical, laboratory and therapeutic data were collected from electronic clinical records and
222 included in an anonymized database. Baseline evaluation was performed around the third day
223 of admission (median=3 days, interquartile range [IQR] 2 to 5). The second evaluation was
224 obtained around the ninth day of admission (median=9 days, IQR 7-12). No significant
225 differences at both evaluation time points were observed between patients treated and not
226 treated with TCZ.

227 In addition, twenty-three serum samples from healthy donors obtained before the pandemic
228 onset were used to determine the variability of IL-6 serum levels in baseline conditions.

229

230 ***SARS-CoV-2 RNA detection***

231 Samples from nasopharyngeal and throat exudates were obtained with specific swabs as
232 previously described (19). Then, we performed real-time RT-PCR assay targeting the E gene of
233 SARS-CoV-2 as the first-line screening tool, with Real Time ready RNA Virus Master (Roche),
234 followed by confirmatory testing with the assay TaqPath™ COVID-19 CE-IVD Kit RT-PCR
235 (Applied Biosystems™), including three assays that target SARS-CoV-2 genes (Orf1ab, S gene,
236 N gene) and one positive control assay that targets the human RNase P RPPH1 gene (20). All
237 the procedures were performed on Applied Biosystems™ Quant Studio-5 Real-Time PCR
238 System.

239

240 ***IL-6 serum level measurement***

241 Surplus sera from laboratory routine determinations were used to assess IL-6 levels, which
242 were retrospectively quantified in duplicate with the Human IL-6 Quantikine high sensitivity
243 enzyme-immune assay from R&D Systems Europe Ltd. (Abingdon, UK). The intra-assay and
244 inter-assay variability were 3% and 5%, respectively.

245

246 ***Tocilizumab treatment***

247 The rationale to treat severe COVID-19 patients with TCZ was based on its previously
248 approved indication for treating the CRS associated with CAR-T cell therapy, as well as
249 unpublished experience in COVID-19 patients from China and Italy. Treatment required

250 approval by the HUP COVID-19 Committee, following the recommendations of the Spanish
251 Agency for Drugs and Health Devices (AEMPS) (21):

- 252 • Interstitial pneumonia with severe respiratory failure (score = 2);
- 253 • Rapid respiratory worsening requiring non-invasive or invasive ventilation (score \geq 3
254 on the COVID respiratory severity scale);
- 255 • Presence of extrapulmonary organ failure (shock or score \geq 3 on the SOFA scale);
- 256 • Criteria for severe systemic inflammatory response. In adults: elevated levels of IL-6
257 ($>$ 40 pg / ml); alternatively, increased levels of D-dimer ($>$ 1500 ng / ml) or
258 progressively increasing D-dimer.
- 259 • Patients who, according to their baseline clinical condition, would be IMV subsidiary.

260
261 Therefore, the decision to treat with TCZ was based on the AEMPS criteria, excluding IL-6 $>$ 40
262 pg/ml, since no information about IL-6 results was available to physicians during the study
263 period. After a preliminary analysis of data by the end of March, IL-6 measurement was
264 included in the baseline assessment of COVID-19 patients.

265 The administration schedule of TCZ at the time of the study was a first intravenous infusion of
266 8 mg/kg (maximum 800 mg) followed by a second one after 12 hours.

267
268 **Variables**

269 To analyze whether IL-6 levels can predict disease severity, two main outcomes were
270 considered: need for IMV and all-cause mortality.

271 To determine the effect of TCZ we analyzed the evolution of PaO₂/FiO₂ between both
272 evaluation times. In 160 out of 267 evaluations arterial oxygen tension (PaO₂) was

273 unavailable. So, in order to avoid missing data in these relevant outcomes, this parameter was
274 estimated from mean oxygen saturation (SatO₂) as proposed elsewhere (22).

275 IL-6 serum levels showed a heterogeneous distribution in patients and in healthy donors
276 (Suppl. Fig. 3A). To improve its representation in figures, this variable was normalized through
277 natural logarithmic transformation (Suppl. Fig. 3B). The procedure to determine the cut-off for
278 high baseline IL-6 is described below.

279

280 **Statistical analysis**

281 Statistical analyses were performed using Stata 14 for Windows (Stata Corp LP, College
282 Station, TX, USA). Quantitative variables following a non-normal distribution were
283 represented as median and IQR and the Mann Whitney test was used to assess significant
284 differences. Variables with a normal distribution were described by mean±standard deviation
285 (SD) and differences between groups were assessed with Student's t-test. Qualitative
286 variables were described as counts and proportions and χ^2 or Fisher's exact test was used for
287 comparisons. Correlation between quantitative variables was analyzed using the Pearson
288 correlation test. To estimate the 95% confidence interval of correlation coefficients we used
289 the *ci2* command of Stata.

290 To determine whether IL-6 serum levels were able to discriminate between: i) COVID-19
291 patients vs. healthy donors; ii) patients requiring IMV vs. those that did not; or iii) patients
292 treated with TCZ vs. not treated, receiver operating characteristic (ROC) analysis was
293 performed using the *roctab* command. Each cut-off point was selected based on the best
294 trade-off values between sensitivity, specificity and the percentage of patients correctly
295 classified. Positive and negative likelihood ratios and ROC curves were also obtained.

296 To determine the variables associated with the need for IMV, we performed a multivariable
297 logistic regression analysis that was first modeled by adding all the variables with a p value
298 lower than 0.15 in the bivariable analysis, namely total lymphocyte count, D-dimer, LDH,
299 PaO₂/FiO₂, COPD, obesity, hypertension, C-reactive protein, and IL-6 (high vs low). The final
300 model was reached with backward stepwise removal of variables with p-value higher than
301 0.15, and using Wald tests to demonstrate that each model was better than its previous
302 iteration.

303 Next, we performed a multivariable analysis using generalized linear models nested by patient
304 and visit (*xtgee* command) in which the dependent variable was PaO₂/FiO₂. This approach
305 allowed us to identify which variables influenced the evolution of PaO₂/FiO₂. The first model
306 included all variables with a p value <0.15 in the bivariable analysis, namely hypertension,
307 baseline radiological pattern, LDH, total lymphocyte count, baseline C-reactive protein, IMV.

308 After that, through backward stepwise approach, we obtained the best model as described
309 above. Then, to assess the role of IL-6 as predictor of TCZ effect on PaO₂/FiO₂, the composite
310 variable IL-6/TCZ (low IL-6/no TCZ, low IL-6/Early TCZ, low IL-6/Late TCZ, high IL-6/no TCZ, high
311 IL-6/Early TCZ and high IL-6/Late TCZ) was forced in the model.

312 Survival time was analyzed by Kaplan-Meier method with the *sts* command of Stata. Date of
313 admission was considered the date of entry and for exit date we considered the exitus date.
314 For those patients without the event, the last revision of the database (electronic chart or
315 telephone call) on May 21st was used to censor their follow-up. Differences in time to death
316 by different variables were analyzed by log-rank test.

317

318 ***Ethics***

319 This study was approved by the local Research Ethics Committee (register number 4070) and

320 it was carried out following the ethical principles established in the Declaration of Helsinki. All
321 included patients (or their representatives) were informed about the study and gave an oral
322 informed consent as proposed by AEMPS due to COVID-19 emergency.

323 This article was written following the Strengthening the Reporting of Observational Studies in
324 Epidemiology (STROBE) guidelines taking into consideration the difficulties to obtain all
325 needed information in the setting of the COVID-19 pandemic.

326

327

328 **RESULTS**

329 ***Demographic and clinical characteristics of the study population***

330 One hundred and forty-six patients were included; their main demographic and clinical
331 characteristics are shown in Table 1. Median age was 63 (IQR [54-71]; range, 30 to 86), 97
332 (66%) were men and 100 (69%) presented comorbidities. The most frequent were
333 hypertension, 55 (38%); obesity, 23 (16%); diabetes mellitus, 26 (18%); and chronic
334 obstructive pulmonary disease (COPD) 9 (6%); 19 (13%) patients had a history of malignancy.
335 Median duration of symptoms before admission was 6 days (IQR 4-7); 36 (25%) arrived at the
336 emergency room presenting fever ($\geq 38^{\circ}\text{C}$), with a SatO_2 of $91\% \pm 5\%$. Most individuals (121
337 [83%]) were admitted to the internal Medicine or Pneumology wards; however, 16 (11%)
338 patients admitted directly to the Intensive Care Unit because of IMV requirement, and 9 (6%)
339 to the Hematology ward because of pre-existing conditions. Additional details of patient
340 baseline features can be found in supplementary table 1.

341

342 ***IL-6 serum levels and disease severity***

343 IL-6 serum levels above 10 pg/ml discriminated COVID-19 patients from healthy donors with
344 low accuracy (Area under ROC curve [AUC] 0.695; sensitivity [Se] 84%, specificity [Sp] 46%,
345 positive likelihood ratio [LR+] 1.5, LR- 0.4; Suppl. Fig. 3C), probably due to their intrinsic
346 heterogeneity (Suppl. Fig. 3A and B).

347 No significant correlation was found between PaO_2 and IL-6 serum levels at baseline ($r = -0.09$
348 [95% CI: -0.270 to 0.085]; $p = 0.299$; Figure 1A), likely due to a higher oxygen supply in the most
349 severe cases; in fact, serum IL-6 levels showed a significant negative correlation with

350 PaO₂/FiO₂ (Figure 1B; r= -0.38 [95% CI: -0.526 to -0.218]; p<0.001), meaning that higher levels
351 of IL-6 at baseline were associated with lower PaO₂/FiO₂.

352 In this regard, forty-four patients (30%) required IMV at some point during their
353 hospitalization. As expected, these patients showed significantly worse PaO₂/FiO₂ levels than
354 those not requiring IMV (p<0.001; Table 2). In addition, they showed increased leukocytes,
355 total lymphocytes, IL-6, C-reactive protein (CRP), and procalcitonin (PCT) showing a higher
356 inflammatory status than those not requiring IMV (p≤0.001 except p=0.003 for CRP and
357 p=0.029 for lymphocyte count; Table 2).

358 Furthermore, a baseline IL-6 above 30 pg/ml (henceforth high IL-6) discriminated patients
359 requiring IMV with 68% Se and 73% Sp. AUC was 0.725 (Figure 1C; LR+ 2.5, LR- 0.4). A logistic
360 regression model, adjusted for COPD and baseline white blood cell count, also showed that
361 high baseline IL-6 was a predictive biomarker for IMV (OR:7.1; 95% CI: 3.0 to 16.6; Supp Table
362 2).

363

364 ***Response to Tocilizumab***

365 Fifty-eight patients (40%) received treatment with TCZ. No significant differences between
366 groups were observed in most sociodemographic and therapeutic variables, except for
367 patients not treated with TCZ, which were more often obese and COPD (p=0.023 and p=0.071
368 respectively, Supplementary Table 3). Importantly, patients in the TCZ-treated group
369 presented several baseline findings indicating that they suffered more severe COVID-19
370 disease, such as lower PaO₂/FiO₂ (p<0.001; Table 3), higher levels of serum Lactate
371 Dehydrogenase (p<0.001), CRP (p=0.005), IL-6 levels at baseline (p=0.007), and total
372 lymphocyte count (p=0.001). Other elevated markers in this group included Aspartate amino-
373 transferase, ferritin, and procalcitonin (p<0.05 for all comparisons. Table 3).

374

375 Even before physicians were aware of the potential value of IL-6 serum levels as a predictor of
376 severe disease, those patients with high IL-6 were more frequently treated with TCZ (Figure
377 1D; AUC 0.634; 30 pg/ml as cut-off showed Se 57%, Sp 69%, LR+ 1.9, LR- 0.7), although with
378 less accuracy than for IMV. The median time from the beginning of symptoms to TCZ
379 treatment was 11 days (IQR: 8-12.5). Therefore, we considered early TCZ when the treatment
380 was applied before 11 days of disease duration and late TCZ after this cut-off.

381 As a consequence of IL-6R blockade with TCZ, a significant trend toward higher IL-6 serum
382 levels after administration of the drug was observed (Figure 2A; $p=0.005$). However, IL-6R
383 blockade induced a fast and significant down-regulation of CRP (Figure 2B; $p<0.001$) and PCT
384 (Figure 2C; $p=0.026$) and a non-significant decrease of ferritin (Figure 2D) and LDH (Figure 2E).
385 Conversely, it was associated with a significant increase of D-Dimer levels (Figure 2F; $p<0.001$).

386 After an average 6 days of TCZ treatment, despite this improvement of inflammatory
387 parameters, $\text{PaO}_2/\text{FiO}_2$ did not show a significant improvement in the whole population (Data
388 not shown). Only patients with high IL-6 that underwent early TCZ treatment showed a
389 significant $\text{PaO}_2/\text{FiO}_2$ increase (Figure 3A mid boxes; $p=0.048$). Patients with low IL-6 did not
390 improve their $\text{PaO}_2/\text{FiO}_2$ after treatment with TCZ (Figure 3B). Those patients not treated with
391 TCZ showed a heterogeneous behavior in their $\text{PaO}_2/\text{FiO}_2$ with a trend to improve in patients
392 with low IL-6 (Figure 3A and B two left boxes of each panel).

393 Since relevant differences were observed between patients treated and not treated with TCZ,
394 we used a multivariable analysis to determine which variables influenced the evolution of
395 $\text{PaO}_2/\text{FiO}_2$ at the short-term. Baseline $\text{PaO}_2/\text{FiO}_2$ and radiological pattern, HTA, LDH and CRP
396 levels and total lymphocyte blood count significantly explained variation in $\text{PaO}_2/\text{FiO}_2$ (Suppl
397 Table 4). Adjusted by these confounders, the best $\text{PaO}_2/\text{FiO}_2$ evolution was achieved in

398 patients with low IL-6 not requiring TCZ treatment (Figure 3C first dot on the left; reference
399 group in analysis showed in Suppl Table 4), likely because they were the less severe patients.
400 Patients with low IL-6 that due to their bad evolution were prescribed with TCZ showed a
401 significant worsening of PaO₂/FiO₂ (Figure 3C, 2nd and 3rd dots; Suppl Table 4). A similar
402 evolution was observed in patients with high IL-6 and late TCZ treatment, whereas those with
403 high IL-6 not treated or treated early with TCZ showed no significant differences compared
404 with the reference group (Figure 3C; Suppl Table 4).

405 406 ***Mortality***

407 Next, we were interested in the long-term evolution of patients depending on baseline IL-6
408 and treatment with TCZ. After a median follow-up of 61 days (IQR: 58-64) we observed 30
409 deaths in our sample (21%). The survival curves according to the baseline level of IL-6 and TCZ
410 treatment are shown in Figure 4. Six patients out of 59 (10%) died in the reference group (low
411 IL-6/no TCZ), 9 out of 28 (32%) in the low IL-6/TCZ treated group (HR 3.6 [CI 95%: 1.3 – 10.0],
412 p=0.016), 10 out of 28 (36%) in the high IL-6/no TCZ group (HR 4.6 [CI 95%: 1.7 – 12.7],
413 p=0.003) and 5 out of 31 (16%) in the high IL-6/TCZ treated group (HR 1.6 [CI 95%: 0.5 – 5.4],
414 p=0.411).

415 416 ***Safety of Tocilizumab treatment***

417 Regarding safety, no relevant cytopenia, hypertransaminasemia, bowel perforation or
418 secondary bacterial infections were observed during or after treatment with TCZ for the time
419 of the study. Ten (7%) patients had positive blood cultures; most of them in the non-TCZ
420 treated group (7 vs 3; p=0.03).

421
422

423 **DISCUSSION**

424 To our knowledge, this is the first study showing that high baseline IL-6 levels predict both the
425 need for IMV and the response to TCZ in severe patients hospitalized with COVID-19. Our
426 results confirm the hypothesis that respiratory failure in the advanced phase of severe COVID-
427 19 is mainly due to an exacerbated inflammatory response. These findings are in accordance
428 with cytokine storms described in previous experiences with H5N1 influenzae virus (5) and
429 previous coronaviruses SARS-CoV and MERS (7), as well as CAR T cell therapy (23, 24).
430 Together, these data suggest a key role for inflammation of the small distal airways in the
431 severity of this condition (25). Hence, approximately one-third of patients with ARDS display
432 elevated levels of inflammatory mediators (IL-6, IL-8 and soluble TNF receptor 1, among
433 others), increasing the prevalence of shock and mortality (26). Accordingly, our data show
434 that IL-6 levels higher than 30 pg/ml predict the need for IMV, and correlated with other
435 severity data. A similar threshold has been described to discriminate between mild and severe
436 COVID-19 in Chinese patients (27).

437 The association of high IL-6 with a more severe disease in our population supports the use of
438 TCZ to treat COVID-19 patients. TCZ is a humanized antibody that blocks both soluble and
439 membrane-bound forms of IL-6 receptor. Thus, TCZ prevents ligand binding, which likely
440 explains why IL-6 serum levels significantly increased after treatment (28). However, the TCZ-
441 mediated blockade of IL-6R signaling led to the observed relevant decrease in circulating
442 inflammatory mediators. Severe high IL-6 COVID-19 patients treated with TCZ showed an early
443 respiratory improvement, represented by a moderate but significantly increased PaO₂/FiO₂
444 when TCZ was prescribed before 11th day of symptoms and lower overall mortality,
445 independently of other treatments or clinical factors. Furthermore, patients with severe
446 COVID-19 and high IL-6 levels that were not treated with TCZ displayed a higher mortality.

447 On the other hand and of outstanding interest, patients with low IL-6 that were treated with
448 TCZ due to a severe or critical COVID-19 did not improve and showed significantly higher
449 mortality. These observations pose the question whether it is possible~~It is likely~~ that severe
450 patients with low IL-6 included in phase 3 trials with Tocilizumab and Sarilumab account for
451 the failure to meet their primary endpoints (29, 30). Therefore, our results support the
452 measurement of baseline IL-6 levels in hospitalized COVID-19 patients, since ~~that~~ in these
453 severe or critical patients with low IL-6 levels, other cytokines ~~{such as~~ IL-1 or tumor necrosis
454 factor ~~{(TNF)}~~ could be ~~responsible for~~ driving the exacerbated inflammatory response in lungs
455 (25), ~~and p~~ Probably, this specific group could benefit ~~they should be treated with~~ from
456 receiving other anti-inflammatory agents such as IL-1 or TNF blockers. In this regard, the need
457 for biomarkers of response to TCZ has been recently highlighted (31) and neither COVACTA
458 study (Tocilizumab trial; NCT04320615) nor ex-US Sarilumab clinical trial (NCT04327388) listed
459 increased baseline IL-6 serum levels within the inclusion criteria. Nevertheless, both clinical
460 trials reported a decrease in duration of hospital stay in the active arm, that was statistically
461 significant in COVACTA (29, 30).

462 In agreement with our data, other case series with few patients have shown that tocilizumab
463 can improve the outcomes of COVID-19 patients with ARDS (6, 8, 31, 32); in some of these
464 reports described the use of, lower doses of TCZ were used. In this regard, the protocol in our
465 hospital, after these preliminary data, has evolved towards lower doses of TCZ (a single dose
466 of TCZ 400 mg if <80 kg and 600 mg if >80 kg) administered earlier with similar efficacy
467 (unpublished observation).

468 Additionally, during our study the use of TCZ was safe and it did not increase the number of
469 serious bacterial infections. These findings are consistent with the results of both the
470 COVACTA and Sarilumab trials (29, 30). The only unexpected observation was the rise in D-

471 dimer levels at the second evaluation. Possible explanations are either: i) IL-6 does not play a
472 role in the regulation of D-dimer production; or ii) more likely, D-dimer production has a
473 slower kinetics than CRP or other acute phase reactants, since in patients not treated with TCZ
474 a similar rise in D-dimer levels was observed (data not shown).

475 Apart from the novelty and the immediate clinical utility of these findings, a drawback of our
476 study is its retrospective and observational nature involving mainly very severe cases in the
477 group of treatment with TCZ. A stricter selection of a control group through a propensity score
478 strategy was unfeasible, since once the physicians were aware of IL-6 measurement they
479 focused their efforts on treating with TCZ those patients with the highest IL-6 levels.
480 Therefore, prospective studies should be carried out to confirm our observations.

481 In addition, there is some controversy about the reliability of PaO₂/FiO₂ as an outcome for
482 improvement of lung damage, especially in patients with IMV. However, our population was a
483 mix of patients with IMV and non-IMV, so despite the many factors that could interfere with
484 PaO₂/FiO₂, we considered it the most objective outcome for patients' assessment.

485 Finally, our findings are of relevance for clinical decision making in the ongoing COVID-19
486 pandemic. Increased levels of circulating IL-6 predict IMV requirement in patients with severe
487 disease, and ~~may~~ can contribute to establish the indication for timely TCZ administration.

488 Furthermore, the improvement of respiratory parameters achieved upon treatment with TCZ
489 may reduce IMV demand in these patients. On the other hand, in our population there was a
490 small group of patients with severe COVID-19 and low IL-6 serum levels, which probably
491 should have been treated with blockade of IL1 or TNF- α since their evolution with TCZ was
492 inadequate.

493

494

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648 single-arm multicentre study on off-label use of tocilizumab in patients with severe COVID-19.
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651 experience. *J Med Virol*. 2020;92(7):814-818. doi: 10.1002/jmv.25801
652
- 653

654 Table 1. Baseline clinical characteristics and laboratory findings of the study population

	Study Population (n=146)
Age	63 (54-71)
Male sex	97 (66)
Comorbidities	100(69)
Duration of symptoms at admission (days)	6 (4-7)
Baseline PaO ₂ /FiO ₂	215 (112-310)
Treatment during hospitalization	
Hydroxychloroquine	137 (96)
Lopinavir/Ritonavir	119 (83)
Azithromycin	82 (57)
Interferon-β	7 (5)
Glucocorticoids	85 (59)
Methylprednisolone bolus	61 (42)
Laboratory Findings	
White Blood Count (10 ³ /mm ³)	7.64 (5.25–10.68)
Lymphocyte Count (10 ³ /mm ³)	0.83 (0.60–11.7)
Creatinine. mg/dl	0.86 (0.70-1.10)
LDH (U/L)	341 (256-461)
CK (U/L)	72 (48-155)
Serum IL-6 (pg/ml)	21.36 (7.53-54.21)
Ferritin (ng/ml)	1598 (830-2305)
CRP (mg/dL)	11.55 (5.16-22.53)
PCT (ng/ml)	0.15 (0.10-0.35)
D-dimer (mg/ml)	0.75 (0.48–1.48)

655 All categorical variables are expressed as number (%) and quantitative variables as
656 median (p25-p75). PaO₂/FiO₂: arterial oxygen tension - fraction of inspired oxygen
657 ratio; LDH: Lactate Dehydrogenase; IL-6: Interleukin-6; CRP: C-reactive protein; PCT:
658 Procalcitonin.
659

660 Table 2. Baseline clinical characteristics of groups requiring vs not requiring Invasive
 661 Mechanical Ventilation.

	Invasive Mechanical Ventilation		p value
	Required (n=44)	Not-required (n=102)	
Age	63.5 (56.5-72)	62 (54-71)	0.517
Male sex	32 (73)	65 (64)	0.291
Comorbidities	30 (68)	70 (69)	0.893
Duration of symptoms at admission (days)	5 (5-7)	7 (4-8)	0.265
Baseline PaO ₂ /FiO ₂	125.5 (75-207)	247 (172-348)	<0.001
Treatment during hospitalization			
Hydroxychloroquine	38 (86)	99 (100)	<0.001
Lopinavir/Ritonavir	38 (86)	81 (82)	0.502
Azithromycin	24 (55)	58 (59)	0.652
Interferon-β	3 (7)	4 (4)	0.676
Glucocorticoids	27 (61)	58 (59)	0.755
Methylprednisolone bolus	21 (48)	40 (40)	0.414
Laboratory Findings			
White Blood Count (10 ³ /mm ³)	9.39 (6.59-13.31)	6.93 (5.13-8.78)	<0.001
Lymphocyte Count (10 ³ /mm ³)	0.74 (0.58-1.08)	0.87 (0.62-1.26)	0.029
Creatinine (mg/dl)	0.99 (0.71-1.20)	0.85 (0.72-1.1)	0.398
LDH (U/L)	413 (315-496)	302 (224-443)	0.001
CK (U/L)	67 (39.50-167.50)	94 (59-140)	0.617
Serum IL-6 (pg/ml)	49.20 (17.28-103.57)	16.08 (6.09-42.03)	<0.001
Ferritin (ng/ml)	1665 (602-2765)	1573 (1012-2300)	0.832
CRP (mg/dL)	17.09 (7.69-28.98)	10.13 (4.83-18.48)	0.003
PCT (ng/ml)	0.29 (0.14-0.46)	0.13 (0.08-0.26)	0.001
D-dimer (mg/ml)	0.92 (0.56-2.31)	0.71 (0.48-1.19)	0.058

662 All categorical variables are expressed as number (%) and quantitative variables as
 663 median (p25-p75). PaO₂/FiO₂: arterial oxygen tension - fraction of inspired oxygen ratio;
 664 LDH: Lactate Dehydrogenase; IL-6: Interleukin-6; CRP: C-reactive protein; PCT:
 665 Procalcitonin.
 666

667 **Table 3.** Baseline clinical characteristics of groups treated vs not treated with Tocilizumab.

	Tocilizumab		p value
	Treated (n=58)	Not treated (n=88)	
Age	61 (54-70)	64 (54-72)	0.288
Male sex	40 (69)	57 (65)	0.600
Comorbidities	35 (61)	64 (73)	0.124
Duration of symptoms at admission (days)	6 (5-7)	7 (4-8)	0.612
Baseline PaO ₂ /FiO ₂	137 (88-232)	248 (183-348)	<0.001
Treatment during hospitalization			
Hydroxychloroquine	53 (93)	84 (98)	0.171
Lopinavir/Ritonavir	51 (89)	68 (79)	0.103
Azithromycin	33 (58)	49 (57)	0.913
Interferon-β	2 (4)	5 (6)	0.532
Glucocorticoids	38 (67)	47 (55)	0.152
Methylprednisolone bolus	31 (54)	30 (35)	0.018
Laboratory Findings			
White Blood Count (10 ³ /mm ³)	7.99 (5.17-11.85)	7.52 (5.4–10.36)	0.527
Lymphocyte Count (10 ³ /mm ³)	0.74 (0.52–0.997)	0.93 (0.66–1.47)	0.001
Creatinine (mg/dl)	0.83 (0.70-1.05)	0.90 (0.72-1.14)	0.177
LDH (U/L)	425 (302-510)	293.5 (221-388)	<0.001
CK (U/L)	69 (38-270)	75.5 (49-125)	0.785
Serum IL-6 (pg/ml)	41.85 (12.37-71.95)	16.25 (6.27-44.95)	0.007
Ferritin (ng/ml)	1888 (1152-2844)	1461 (471-1861)	0.038
CRP (mg/dL)	13.73 (8.75-27.08)	9.09 (4.78-19.31)	0.005
PCT (ng/ml)	0.25 (0.13-0.36)	0.14 (0.1-0.3)	0.045
D-dimer (mg/ml)	0.75 (0.48–1.48)	0.71 (0.53–1.22)	0.491

668 All categorical variables are expressed as number (%) and quantitative variables as median (p25-p75).

669 PaO₂/FiO₂: arterial oxygen tension - fraction of inspired oxygen ratio; LDH: Lactate Dehydrogenase; IL-

670 6: Interleukin-6; CRP: C-reactive protein; PCT: Procalcitonin.

671

672 FIGURE LEGENDS

673 **Figure 1.** IL-6 serum levels predict disease severity and tocilizumab (TCZ) use. (A) Correlation
674 between log-transformed IL-6 serum levels and arterial oxygen tension (PaO₂). (B) Correlation
675 between log-transformed IL-6 serum levels and arterial oxygen tension - fraction of inspired
676 oxygen ratio (PaO₂/FiO₂). Data in panels A and B are shown as dot-plot and their fitted linear
677 prediction with 95% confidence interval (transparent grey shadow) estimated using the
678 twoway command of Stata with the lfitci option. (C) ROC curve showing the ability of log-
679 transformed IL-6 serum levels to discriminate between patients requiring vs not-requiring
680 invasive mechanical ventilation (IMV). (D) ROC curve for the ability of log-transformed IL-6
681 serum levels to discriminate between TCZ treated and non-treated patients. The best cut-off
682 for discrimination of patients requiring IMV (panel C) or TCZ treatment (panel D) was 30
683 pg/ml.

684 **Figure 2.** Response of laboratory parameters to Tocilizumab (TCZ) treatment. (A) Differences
685 in log-transformed IL-6 serum levels, (B) C-Reactive Protein and (C) Procalcitonin, (D) Ferritin,
686 (E) Lactate Dehydrogenase (LDH), and (F) D-Dimer. Data are presented as the interquartile
687 range (p75 upper edge, p25 lower edge, p50 midline), p95 (line above the box), and p5 (line
688 below the box) of levels for each parameter before (grey boxes) and after (white boxes)
689 treatment with TCZ.

690 **Figure 3.** Change of PaO₂/FiO₂ in COVID-19 patients treated early (before 11 day of symptoms
691 onset) or late with Tocilizumab and not-treated. (A) Patients with high baseline IL-6 (cut-off
692 30 pg/ml). (B) Subjects with low baseline IL-6 serum levels. Data in A and B are shown as the
693 interquartile range (p75 upper edge, p25 lower edge, p50 midline), p95 (line above the box),
694 and p5 (line below the box) before (grey boxes) and after (white boxes) treatment with TCZ. In
695 non-treated patients PRE and POST mean first and second evaluation, respectively. Statistical

696 significance was determined with the Mann-Whitney test. (C) The graph represents the
697 predicted mean (dots) with 95% confidence intervals (bars) of PaO₂/FiO₂ according to baseline
698 IL-6 levels and early or late TCZ treatment. Data were obtained with the command
699 marginsplot of Stata, after adjustment by baseline PaO₂/FiO₂ and radiological pattern,
700 Hypertension, Lactic dehidrogenase and C-reactive protein levels, lymphocyte blood count
701 and need for IMV, according to the multivariable analysis displayed in Supplementary Table 2
702 (see Methods for further information).

703 **Figure 4.** Survival curves of COVID-19 patients grouped according to baseline IL-6 levels and
704 TCZ treatment. Statistical significance was established with log-rank test.

705

706

707

1 **TITLE**

2 IL-6 serum levels predict severity and response to Tocilizumab in COVID-19: an observational
3 study

4
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112

113 **RUNNING TITLE**

114 IL-6 levels predict response to TCZ in COVID-19

115

116

117 **ABSTRACT**

118 **Background**

119 COVID-19 patients can develop a cytokine release syndrome that eventually leads to acute
120 respiratory distress syndrome (ARDS) requiring invasive mechanical ventilation (IMV). Since
121 interleukin-6 (IL-6) is a relevant cytokine in ARDS, the blockade of its receptor with
122 Tocilizumab (TCZ) could reduce mortality and/or morbidity in severe COVID-19.

123 **Objective**

124 To determine whether baseline IL-6 serum levels can predict the need for IMV and the
125 response to TCZ.

126 **Methods**

127 Retrospective observational study performed in hospitalized patients diagnosed of COVID-19.
128 Clinical information and laboratory findings, including IL-6 levels, were collected
129 approximately 3 and 9 days after admission to be matched with pre- and post-administration
130 of TCZ. Multivariable logistic and linear regressions, and survival analysis were performed
131 depending on outcomes: need for IMV, evolution of arterial oxygen tension/fraction of
132 inspired oxygen ratio ($\text{PaO}_2/\text{FiO}_2$) or mortality.

133 **Results**

134 One hundred and forty-six patients were studied, predominantly male (66%); median age was
135 63 years. Forty-four patients (30%) required IMV, and 58 patients (40%) received treatment
136 with TCZ. IL-6 levels >30 pg/ml was the best predictor for IMV (OR:7.1; $p<0.001$). Early
137 administration of TCZ was associated with improvement of oxygenation ($\text{PaO}_2/\text{FiO}_2$) in
138 patients with high IL-6 ($p=0.048$). Patients with high IL-6 not treated with TCZ showed high
139 mortality (HR: 4.6; $p=0.003$), as well as those with low IL-6 treated with TCZ (HR: 3.6; $p=0.016$).
140 No relevant serious adverse events were observed in TCZ-treated patients.

141 **Conclusion**

142 Baseline IL-6>30 pg/ml predicts IMV requirement in patients with COVID-19 and contributes
143 to establish an adequate indication for TCZ administration.

144

145 **CLINICAL IMPLICATIONS**

146 Elevated levels of circulating IL-6 predict the need for IMV in severe COVID-19 patients and
147 may contribute to establish the indication for timely administration of TCZ, possibly reducing
148 ICU demand.

149

150 **KEYWORDS**

151 COVID-19, Interleukin-6, Tocilizumab, Invasive Mechanical Ventilation

152

153 **ABBREVIATIONS**

154 AEMPS: Spanish Agency for Drugs and Health Devices

155 ARDS: acute respiratory distress syndrome

156 AUC: Area under curve

157 CAR: chimeric antigen receptor

158 COPD: chronic obstructive pulmonary disease

159 COVID-19: coronavirus disease 2019

160 CRP: C-reactive protein

161 CRS: cytokine release syndrome

162 IL: interleukin

163 IMV: invasive mechanical ventilation

164 IQR: interquartile range

- 165 LR+: positive likelihood ratio; LR- negative likelihood ratio
- 166 PaO₂: arterial oxygen tension
- 167 PaO₂/FiO₂: arterial oxygen tension/fraction of inspired oxygen ratio
- 168 PCT: procalcitonin
- 169 ROC: receiver operating characteristic
- 170 SatO₂: mean oxygen saturation
- 171 SD: standard deviation
- 172 STROBE: Strengthening the Reporting of Observational Studies in Epidemiology
- 173 TCZ: Tocilizumab
- 174 TNF: tumor necrosis factor

175

176 **CAPSULE SUMMARY**

177 Baseline IL-6 serum levels >30 pg/ml identify severe COVID-19 patients and should be used to

178 guide the intervention with IL-6R inhibitors, aiming to improve their use in an uncertain and

179 evolving therapeutic scenario.

180 **INTRODUCTION**

181 The recent exponential increase in cases of severe acute respiratory syndrome caused by
182 coronavirus 2 (SARS-CoV-2) has led the World Health Organization to declare a pandemic. The
183 disease, known as coronavirus disease 2019 (COVID-19), has pushed national health systems
184 to the brink of collapse, prompting national governments to impose complete population
185 lockdowns in an attempt to slow down the dynamics of infection (1, 2)

186 The spectrum of COVID-19 clinical manifestations varies widely, from mild to severe cases of
187 atypical pneumonia, some of them developing acute respiratory distress syndrome (ARDS),
188 which often requires invasive mechanical ventilation (IMV) and is the leading cause of death.

189 It is suggested that the severity of the respiratory disease caused by SARS-CoV-2 is largely due
190 to an exacerbated immune response against the virus (3, 4). This response has been observed
191 in previous respiratory virus outbreaks and can be also seen in patients treated with chimeric
192 antigen receptor (CAR) T cell therapy (5-7). Pro-inflammatory cytokines such as interleukin (IL)
193 1 and IL-6 are crucial mediators of this process (3, 6). In this regard, recent studies have
194 indicated the usefulness of IL-6 serum levels, lymphocyte count, fibrinogen or D-dimer to
195 evaluate the development of ARDS and its mortality (8-11). However, further evidence is
196 required to support these observations (12).

197 Tocilizumab (TCZ), an anti-IL-6 receptor (IL-6R) antibody, is the only drug currently licensed for
198 the treatment of the cytokine release syndrome (CRS) associated with CAR-T cell therapy (13).

199 Due to the virulence of the current outbreak, its use has been advised in severe cases of
200 COVID-19 (14-16). The rationale is to curb the deleterious effects of inflammation, thereby
201 limiting lung damage. Early promising results have prompted ongoing randomized clinical
202 trials (RCT) (17). We aimed to explore the ability of IL-6 serum levels at baseline to predict the
203 need for IMV and the response to TCZ in severe COVID-19 patients.

204 **METHODS**

205 ***Study design and population***

206 This is a retrospective observational study including 146 consecutive patients with confirmed
207 detection of SARS-CoV-2 RNA, baseline IL-6 serum level measurement and admitted to the
208 Hospital Universitario La Princesa (HUP) with severe to critical COVID-19 (18), from February
209 24th to March 23rd 2020 (Flow chart in Suppl. Fig. 1).

210

211 This time frame was decided based on the fact that after March 23rd, clinicians involved in the
212 treatment of these patients were aware of the possibility of measuring IL-6 serum levels to
213 decide treatment initiation with TCZ (RoActemra, Roche). Furthermore, around that date
214 Spanish facilities capable of providing IMV were overrun (Suppl. Fig. 2), thus the decision for
215 IMV application was based on availability rather than the patients' needs. Notwithstanding,
216 during the study period, patients were prompted to IMV when they presented a marked
217 worsening of oxygenation (arterial oxygen tension/fraction of inspired oxygen ratio
218 $[PaO_2/FiO_2]<200$), provided that their baseline conditions did not contraindicate it.

219

220 ***Data collection***

221 Clinical, laboratory and therapeutic data were collected from electronic clinical records and
222 included in an anonymized database. Baseline evaluation was performed around the third day
223 of admission (median=3 days, interquartile range [IQR] 2 to 5). The second evaluation was
224 obtained around the ninth day of admission (median=9 days, IQR 7-12). No significant
225 differences at both evaluation time points were observed between patients treated and not
226 treated with TCZ.

227 In addition, twenty-three serum samples from healthy donors obtained before the pandemic
228 onset were used to determine the variability of IL-6 serum levels in baseline conditions.

229

230 ***SARS-CoV-2 RNA detection***

231 Samples from nasopharyngeal and throat exudates were obtained with specific swabs as
232 previously described (19). Then, we performed real-time RT-PCR assay targeting the E gene of
233 SARS-CoV-2 as the first-line screening tool, with Real Time ready RNA Virus Master (Roche),
234 followed by confirmatory testing with the assay TaqPath™ COVID-19 CE-IVD Kit RT-PCR
235 (Applied Biosystems™), including three assays that target SARS-CoV-2 genes (Orf1ab, S gene,
236 N gene) and one positive control assay that targets the human RNase P RPPH1 gene (20). All
237 the procedures were performed on Applied Biosystems™ Quant Studio-5 Real-Time PCR
238 System.

239

240 ***IL-6 serum level measurement***

241 Surplus sera from laboratory routine determinations were used to assess IL-6 levels, which
242 were retrospectively quantified in duplicate with the Human IL-6 Quantikine high sensitivity
243 enzyme-immune assay from R&D Systems Europe Ltd. (Abingdon, UK). The intra-assay and
244 inter-assay variability were 3% and 5%, respectively.

245

246 ***Tocilizumab treatment***

247 The rationale to treat severe COVID-19 patients with TCZ was based on its previously
248 approved indication for treating the CRS associated with CAR-T cell therapy, as well as
249 unpublished experience in COVID-19 patients from China and Italy. Treatment required

250 approval by the HUP COVID-19 Committee, following the recommendations of the Spanish
251 Agency for Drugs and Health Devices (AEMPS) (21):

- 252 • Interstitial pneumonia with severe respiratory failure (score = 2);
- 253 • Rapid respiratory worsening requiring non-invasive or invasive ventilation (score \geq 3
254 on the COVID respiratory severity scale);
- 255 • Presence of extrapulmonary organ failure (shock or score \geq 3 on the SOFA scale);
- 256 • Criteria for severe systemic inflammatory response. In adults: elevated levels of IL-6
257 ($>$ 40 pg / ml); alternatively, increased levels of D-dimer ($>$ 1500 ng / ml) or
258 progressively increasing D-dimer.
- 259 • Patients who, according to their baseline clinical condition, would be IMV subsidiary.

260
261 Therefore, the decision to treat with TCZ was based on the AEMPS criteria, excluding IL-6 $>$ 40
262 pg/ml, since no information about IL-6 results was available to physicians during the study
263 period. After a preliminary analysis of data by the end of March, IL-6 measurement was
264 included in the baseline assessment of COVID-19 patients.

265 The administration schedule of TCZ at the time of the study was a first intravenous infusion of
266 8 mg/kg (maximum 800 mg) followed by a second one after 12 hours.

267
268 **Variables**

269 To analyze whether IL-6 levels can predict disease severity, two main outcomes were
270 considered: need for IMV and all-cause mortality.

271 To determine the effect of TCZ we analyzed the evolution of PaO₂/FiO₂ between both
272 evaluation times. In 160 out of 267 evaluations arterial oxygen tension (PaO₂) was

273 unavailable. So, in order to avoid missing data in these relevant outcomes, this parameter was
274 estimated from mean oxygen saturation (SatO₂) as proposed elsewhere (22).

275 IL-6 serum levels showed a heterogeneous distribution in patients and in healthy donors
276 (Suppl. Fig. 3A). To improve its representation in figures, this variable was normalized through
277 natural logarithmic transformation (Suppl. Fig. 3B). The procedure to determine the cut-off for
278 high baseline IL-6 is described below.

279

280 **Statistical analysis**

281 Statistical analyses were performed using Stata 14 for Windows (Stata Corp LP, College
282 Station, TX, USA). Quantitative variables following a non-normal distribution were
283 represented as median and IQR and the Mann Whitney test was used to assess significant
284 differences. Variables with a normal distribution were described by mean±standard deviation
285 (SD) and differences between groups were assessed with Student's t-test. Qualitative
286 variables were described as counts and proportions and χ^2 or Fisher's exact test was used for
287 comparisons. Correlation between quantitative variables was analyzed using the Pearson
288 correlation test. To estimate the 95% confidence interval of correlation coefficients we used
289 the *ci2* command of Stata.

290 To determine whether IL-6 serum levels were able to discriminate between: i) COVID-19
291 patients vs. healthy donors; ii) patients requiring IMV vs. those that did not; or iii) patients
292 treated with TCZ vs. not treated, receiver operating characteristic (ROC) analysis was
293 performed using the *roctab* command. Each cut-off point was selected based on the best
294 trade-off values between sensitivity, specificity and the percentage of patients correctly
295 classified. Positive and negative likelihood ratios and ROC curves were also obtained.

296 To determine the variables associated with the need for IMV, we performed a multivariable
297 logistic regression analysis that was first modeled by adding all the variables with a p value
298 lower than 0.15 in the bivariable analysis, namely total lymphocyte count, D-dimer, LDH,
299 PaO₂/FiO₂, COPD, obesity, hypertension, C-reactive protein, and IL-6 (high vs low). The final
300 model was reached with backward stepwise removal of variables with p-value higher than
301 0.15, and using Wald tests to demonstrate that each model was better than its previous
302 iteration.

303 Next, we performed a multivariable analysis using generalized linear models nested by patient
304 and visit (*xtgee* command) in which the dependent variable was PaO₂/FiO₂. This approach
305 allowed us to identify which variables influenced the evolution of PaO₂/FiO₂. The first model
306 included all variables with a p value <0.15 in the bivariable analysis, namely hypertension,
307 baseline radiological pattern, LDH, total lymphocyte count, baseline C-reactive protein, IMV.
308 After that, through backward stepwise approach, we obtained the best model as described
309 above. Then, to assess the role of IL-6 as predictor of TCZ effect on PaO₂/FiO₂, the composite
310 variable IL-6/TCZ (low IL-6/no TCZ, low IL-6/Early TCZ, low IL-6/Late TCZ, high IL-6/no TCZ, high
311 IL-6/Early TCZ and high IL-6/Late TCZ) was forced in the model.

312 Survival time was analyzed by Kaplan-Meier method with the *sts* command of Stata. Date of
313 admission was considered the date of entry and for exit date we considered the exitus date.
314 For those patients without the event, the last revision of the database (electronic chart or
315 telephone call) on May 21st was used to censor their follow-up. Differences in time to death
316 by different variables were analyzed by log-rank test.

317

318 ***Ethics***

319 This study was approved by the local Research Ethics Committee (register number 4070) and

320 it was carried out following the ethical principles established in the Declaration of Helsinki. All
321 included patients (or their representatives) were informed about the study and gave an oral
322 informed consent as proposed by AEMPS due to COVID-19 emergency.

323 This article was written following the Strengthening the Reporting of Observational Studies in
324 Epidemiology (STROBE) guidelines taking into consideration the difficulties to obtain all
325 needed information in the setting of the COVID-19 pandemic.

326

327

328 **RESULTS**

329 ***Demographic and clinical characteristics of the study population***

330 One hundred and forty-six patients were included; their main demographic and clinical
331 characteristics are shown in Table 1. Median age was 63 (IQR [54-71]; range, 30 to 86), 97
332 (66%) were men and 100 (69%) presented comorbidities. The most frequent were
333 hypertension, 55 (38%); obesity, 23 (16%); diabetes mellitus, 26 (18%); and chronic
334 obstructive pulmonary disease (COPD) 9 (6%); 19 (13%) patients had a history of malignancy.
335 Median duration of symptoms before admission was 6 days (IQR 4-7); 36 (25%) arrived at the
336 emergency room presenting fever ($\geq 38^{\circ}\text{C}$), with a SatO_2 of $91\% \pm 5\%$. Most individuals (121
337 [83%]) were admitted to the internal Medicine or Pneumology wards; however, 16 (11%)
338 patients admitted directly to the Intensive Care Unit because of IMV requirement, and 9 (6%)
339 to the Hematology ward because of pre-existing conditions. Additional details of patient
340 baseline features can be found in supplementary table 1.

341

342 ***IL-6 serum levels and disease severity***

343 IL-6 serum levels above 10 pg/ml discriminated COVID-19 patients from healthy donors with
344 low accuracy (Area under ROC curve [AUC] 0.695; sensitivity [Se] 84%, specificity [Sp] 46%,
345 positive likelihood ratio [LR+] 1.5, LR- 0.4; Suppl. Fig. 3C), probably due to their intrinsic
346 heterogeneity (Suppl. Fig. 3A and B).

347 No significant correlation was found between PaO_2 and IL-6 serum levels at baseline ($r = -0.09$
348 [95% CI: -0.270 to 0.085]; $p = 0.299$; Figure 1A), likely due to a higher oxygen supply in the most
349 severe cases; in fact, serum IL-6 levels showed a significant negative correlation with

350 PaO₂/FiO₂ (Figure 1B; r= -0.38 [95% CI: -0.526 to -0.218]; p<0.001), meaning that higher levels
351 of IL-6 at baseline were associated with lower PaO₂/FiO₂.

352 In this regard, forty-four patients (30%) required IMV at some point during their
353 hospitalization. As expected, these patients showed significantly worse PaO₂/FiO₂ levels than
354 those not requiring IMV (p<0.001; Table 2). In addition, they showed increased leukocytes,
355 total lymphocytes, IL-6, C-reactive protein (CRP), and procalcitonin (PCT) showing a higher
356 inflammatory status than those not requiring IMV (p≤0.001 except p=0.003 for CRP and
357 p=0.029 for lymphocyte count; Table 2).

358 Furthermore, a baseline IL-6 above 30 pg/ml (henceforth high IL-6) discriminated patients
359 requiring IMV with 68% Se and 73% Sp. AUC was 0.725 (Figure 1C; LR+ 2.5, LR- 0.4). A logistic
360 regression model, adjusted for COPD and baseline white blood cell count, also showed that
361 high baseline IL-6 was a predictive biomarker for IMV (OR:7.1; 95% CI: 3.0 to 16.6; Supp Table
362 2).

363

364 ***Response to Tocilizumab***

365 Fifty-eight patients (40%) received treatment with TCZ. No significant differences between
366 groups were observed in most sociodemographic and therapeutic variables, except for
367 patients not treated with TCZ, which were more often obese and COPD (p=0.023 and p=0.071
368 respectively, Supplementary Table 3). Importantly, patients in the TCZ-treated group
369 presented several baseline findings indicating that they suffered more severe COVID-19
370 disease, such as lower PaO₂/FiO₂ (p<0.001; Table 3), higher levels of serum Lactate
371 Dehydrogenase (p<0.001), CRP (p=0.005), IL-6 levels at baseline (p=0.007), and total
372 lymphocyte count (p=0.001). Other elevated markers in this group included Aspartate amino-
373 transferase, ferritin, and procalcitonin (p<0.05 for all comparisons. Table 3).

374

375 Even before physicians were aware of the potential value of IL-6 serum levels as a predictor of
376 severe disease, those patients with high IL-6 were more frequently treated with TCZ (Figure
377 1D; AUC 0.634; 30 pg/ml as cut-off showed Se 57%, Sp 69%, LR+ 1.9, LR- 0.7), although with
378 less accuracy than for IMV. The median time from the beginning of symptoms to TCZ
379 treatment was 11 days (IQR: 8-12.5). Therefore, we considered early TCZ when the treatment
380 was applied before 11 days of disease duration and late TCZ after this cut-off.

381 As a consequence of IL-6R blockade with TCZ, a significant trend toward higher IL-6 serum
382 levels after administration of the drug was observed (Figure 2A; $p=0.005$). However, IL-6R
383 blockade induced a fast and significant down-regulation of CRP (Figure 2B; $p<0.001$) and PCT
384 (Figure 2C; $p=0.026$) and a non-significant decrease of ferritin (Figure 2D) and LDH (Figure 2E).
385 Conversely, it was associated with a significant increase of D-Dimer levels (Figure 2F; $p<0.001$).

386 After an average 6 days of TCZ treatment, despite this improvement of inflammatory
387 parameters, $\text{PaO}_2/\text{FiO}_2$ did not show a significant improvement in the whole population (Data
388 not shown). Only patients with high IL-6 that underwent early TCZ treatment showed a
389 significant $\text{PaO}_2/\text{FiO}_2$ increase (Figure 3A mid boxes; $p=0.048$). Patients with low IL-6 did not
390 improve their $\text{PaO}_2/\text{FiO}_2$ after treatment with TCZ (Figure 3B). Those patients not treated with
391 TCZ showed a heterogeneous behavior in their $\text{PaO}_2/\text{FiO}_2$ with a trend to improve in patients
392 with low IL-6 (Figure 3A and B two left boxes of each panel).

393 Since relevant differences were observed between patients treated and not treated with TCZ,
394 we used a multivariable analysis to determine which variables influenced the evolution of
395 $\text{PaO}_2/\text{FiO}_2$ at the short-term. Baseline $\text{PaO}_2/\text{FiO}_2$ and radiological pattern, HTA, LDH and CRP
396 levels and total lymphocyte blood count significantly explained variation in $\text{PaO}_2/\text{FiO}_2$ (Suppl
397 Table 4). Adjusted by these confounders, the best $\text{PaO}_2/\text{FiO}_2$ evolution was achieved in

398 patients with low IL-6 not requiring TCZ treatment (Figure 3C first dot on the left; reference
399 group in analysis showed in Suppl Table 4), likely because they were the less severe patients.
400 Patients with low IL-6 that due to their bad evolution were prescribed with TCZ showed a
401 significant worsening of PaO₂/FiO₂ (Figure 3C, 2nd and 3rd dots; Suppl Table 4). A similar
402 evolution was observed in patients with high IL-6 and late TCZ treatment, whereas those with
403 high IL-6 not treated or treated early with TCZ showed no significant differences compared
404 with the reference group (Figure 3C; Suppl Table 4).

405 406 ***Mortality***

407 Next, we were interested in the long-term evolution of patients depending on baseline IL-6
408 and treatment with TCZ. After a median follow-up of 61 days (IQR: 58-64) we observed 30
409 deaths in our sample (21%). The survival curves according to the baseline level of IL-6 and TCZ
410 treatment are shown in Figure 4. Six patients out of 59 (10%) died in the reference group (low
411 IL-6/no TCZ), 9 out of 28 (32%) in the low IL-6/TCZ treated group (HR 3.6 [CI 95%: 1.3 – 10.0],
412 p=0.016), 10 out of 28 (36%) in the high IL-6/no TCZ group (HR 4.6 [CI 95%: 1.7 – 12.7],
413 p=0.003) and 5 out of 31 (16%) in the high IL-6/TCZ treated group (HR 1.6 [CI 95%: 0.5 – 5.4],
414 p=0.411).

415 416 ***Safety of Tocilizumab treatment***

417 Regarding safety, no relevant cytopenia, hypertransaminasemia, bowel perforation or
418 secondary bacterial infections were observed during or after treatment with TCZ for the time
419 of the study. Ten (7%) patients had positive blood cultures; most of them in the non-TCZ
420 treated group (7 vs 3; p=0.03).

421
422

423 **DISCUSSION**

424 To our knowledge, this is the first study showing that high baseline IL-6 levels predict both the
425 need for IMV and the response to TCZ in severe patients hospitalized with COVID-19. Our
426 results confirm the hypothesis that respiratory failure in the advanced phase of severe COVID-
427 19 is mainly due to an exacerbated inflammatory response. These findings are in accordance
428 with cytokine storms described in previous experiences with H5N1 influenzae virus (5) and
429 previous coronaviruses SARS-CoV and MERS (7), as well as CAR T cell therapy (23, 24).
430 Together, these data suggest a key role for inflammation of the small distal airways in the
431 severity of this condition (25). Hence, approximately one-third of patients with ARDS display
432 elevated levels of inflammatory mediators (IL-6, IL-8 and soluble TNF receptor 1, among
433 others), increasing the prevalence of shock and mortality (26). Accordingly, our data show
434 that IL-6 levels higher than 30 pg/ml predict the need for IMV, and correlated with other
435 severity data. A similar threshold has been described to discriminate between mild and severe
436 COVID-19 in Chinese patients (27).

437 The association of high IL-6 with a more severe disease in our population supports the use of
438 TCZ to treat COVID-19 patients. TCZ is a humanized antibody that blocks both soluble and
439 membrane-bound forms of IL-6 receptor. Thus, TCZ prevents ligand binding, which likely
440 explains why IL-6 serum levels significantly increased after treatment (28). However, the TCZ-
441 mediated blockade of IL-6R signaling led to the observed relevant decrease in circulating
442 inflammatory mediators. Severe high IL-6 COVID-19 patients treated with TCZ showed an early
443 respiratory improvement, represented by a moderate but significantly increased PaO₂/FiO₂
444 when TCZ was prescribed before 11th day of symptoms and lower overall mortality,
445 independently of other treatments or clinical factors. Furthermore, patients with severe
446 COVID-19 and high IL-6 levels that were not treated with TCZ displayed a higher mortality.

447 On the other hand and of outstanding interest, patients with low IL-6 that were treated with
448 TCZ due to a severe or critical COVID-19 did not improve and showed significantly higher
449 mortality. These observations pose the question whether it is possible that severe patients
450 with low IL-6 included in phase 3 trials with Tocilizumab and Sarilumab account for the failure
451 to meet their primary endpoints (29, 30). Therefore, our results support the measurement of
452 baseline IL-6 levels in hospitalized COVID-19 patients, since in those severe or critical patients
453 with low IL-6 levels, other cytokines such as IL-1 or tumor necrosis factor (TNF) could be
454 driving the exacerbated inflammatory response in lungs (25). Probably, this specific group
455 could benefit from receiving other anti-inflammatory agents such as IL-1 or TNF blockers. In
456 this regard, the need for biomarkers of response to TCZ has been recently highlighted (31) and
457 neither COVACTA study (Tocilizumab trial; NCT04320615) nor ex-US Sarilumab clinical trial
458 (NCT04327388) listed increased baseline IL-6 serum levels within the inclusion criteria.
459 Nevertheless, both clinical trials reported a decrease in duration of hospital stay in the active
460 arm, that was statistically significant in COVACTA (29, 30).

461 In agreement with our data, other case series with few patients have shown that tocilizumab
462 can improve the outcomes of COVID-19 patients with ARDS (6, 8, 31, 32); in some of these
463 reports described the use of, lower doses of TCZ were used. In this regard, the protocol in our
464 hospital, after these preliminary data, has evolved towards lower doses of TCZ (a single dose
465 of TCZ 400 mg if <80 kg and 600 mg if >80 kg) administered earlier with similar efficacy
466 (unpublished observation).

467 Additionally, during our study the use of TCZ was safe and it did not increase the number of
468 serious bacterial infections. These findings are consistent with the results of both the
469 COVACTA and Sarilumab trials (29, 30). The only unexpected observation was the rise in D-
470 dimer levels at the second evaluation. Possible explanations are either: i) IL-6 does not play a

471 role in the regulation of D-dimer production; or ii) more likely, D-dimer production has a
472 slower kinetics than CRP or other acute phase reactants, since in patients not treated with TCZ
473 a similar rise in D-dimer levels was observed (data not shown).

474 Apart from the novelty and the immediate clinical utility of these findings, a drawback of our
475 study is its retrospective and observational nature involving mainly very severe cases in the
476 group of treatment with TCZ. A stricter selection of a control group through a propensity score
477 strategy was unfeasible, since once the physicians were aware of IL-6 measurement they
478 focused their efforts on treating with TCZ those patients with the highest IL-6 levels.
479 Therefore, prospective studies should be carried out to confirm our observations.

480 In addition, there is some controversy about the reliability of $\text{PaO}_2/\text{FiO}_2$ as an outcome for
481 improvement of lung damage, especially in patients with IMV. However, our population was a
482 mix of patients with IMV and non-IMV, so despite the many factors that could interfere with
483 $\text{PaO}_2/\text{FiO}_2$, we considered it the most objective outcome for patients' assessment.

484 Finally, our findings are of relevance for clinical decision making in the ongoing COVID-19
485 pandemic. Increased levels of circulating IL-6 predict IMV requirement in patients with severe
486 disease, and can contribute to establish the indication for timely TCZ administration.
487 Furthermore, the improvement of respiratory parameters achieved upon treatment with TCZ
488 may reduce IMV demand in these patients. On the other hand, in our population there was a
489 small group of patients with severe COVID-19 and low IL-6 serum levels, which probably
490 should have been treated with blockade of IL1 or TNF- α since their evolution with TCZ was
491 inadequate.

492

493

494

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654 Table 1. Baseline clinical characteristics and laboratory findings of the study population

	Study Population (n=146)
Age	63 (54-71)
Male sex	97 (66)
Comorbidities	100(69)
Duration of symptoms at admission (days)	6 (4-7)
Baseline PaO ₂ /FiO ₂	215 (112-310)
Treatment during hospitalization	
Hydroxychloroquine	137 (96)
Lopinavir/Ritonavir	119 (83)
Azithromycin	82 (57)
Interferon-β	7 (5)
Glucocorticoids	85 (59)
Methylprednisolone bolus	61 (42)
Laboratory Findings	
White Blood Count (10 ³ /mm ³)	7.64 (5.25–10.68)
Lymphocyte Count (10 ³ /mm ³)	0.83 (0.60–11.7)
Creatinine. mg/dl	0.86 (0.70-1.10)
LDH (U/L)	341 (256-461)
CK (U/L)	72 (48-155)
Serum IL-6 (pg/ml)	21.36 (7.53-54.21)
Ferritin (ng/ml)	1598 (830-2305)
CRP (mg/dL)	11.55 (5.16-22.53)
PCT (ng/ml)	0.15 (0.10-0.35)
D-dimer (mg/ml)	0.75 (0.48–1.48)

655 All categorical variables are expressed as number (%) and quantitative variables as
656 median (p25-p75). PaO₂/FiO₂: arterial oxygen tension - fraction of inspired oxygen
657 ratio; LDH: Lactate Dehydrogenase; IL-6: Interleukin-6; CRP: C-reactive protein; PCT:
658 Procalcitonin.

659

660 Table 2. Baseline clinical characteristics of groups requiring vs not requiring Invasive
 661 Mechanical Ventilation.

	Invasive Mechanical Ventilation		p value
	Required (n=44)	Not-required (n=102)	
Age	63.5 (56.5-72)	62 (54-71)	0.517
Male sex	32 (73)	65 (64)	0.291
Comorbidities	30 (68)	70 (69)	0.893
Duration of symptoms at admission (days)	5 (5-7)	7 (4-8)	0.265
Baseline PaO ₂ /FiO ₂	125.5 (75-207)	247 (172-348)	<0.001
Treatment during hospitalization			
Hydroxychloroquine	38 (86)	99 (100)	<0.001
Lopinavir/Ritonavir	38 (86)	81 (82)	0.502
Azithromycin	24 (55)	58 (59)	0.652
Interferon-β	3 (7)	4 (4)	0.676
Glucocorticoids	27 (61)	58 (59)	0.755
Methylprednisolone bolus	21 (48)	40 (40)	0.414
Laboratory Findings			
White Blood Count (10 ³ /mm ³)	9.39 (6.59-13.31)	6.93 (5.13-8.78)	<0.001
Lymphocyte Count (10 ³ /mm ³)	0.74 (0.58-1.08)	0.87 (0.62-1.26)	0.029
Creatinine (mg/dl)	0.99 (0.71-1.20)	0.85 (0.72-1.1)	0.398
LDH (U/L)	413 (315-496)	302 (224-443)	0.001
CK (U/L)	67 (39.50-167.50)	94 (59-140)	0.617
Serum IL-6 (pg/ml)	49.20 (17.28-103.57)	16.08 (6.09-42.03)	<0.001
Ferritin (ng/ml)	1665 (602-2765)	1573 (1012-2300)	0.832
CRP (mg/dL)	17.09 (7.69-28.98)	10.13 (4.83-18.48)	0.003
PCT (ng/ml)	0.29 (0.14-0.46)	0.13 (0.08-0.26)	0.001
D-dimer (mg/ml)	0.92 (0.56-2.31)	0.71 (0.48-1.19)	0.058

662 All categorical variables are expressed as number (%) and quantitative variables as
 663 median (p25-p75). PaO₂/FiO₂: arterial oxygen tension - fraction of inspired oxygen ratio;
 664 LDH: Lactate Dehydrogenase; IL-6: Interleukin-6; CRP: C-reactive protein; PCT:
 665 Procalcitonin.
 666

667 **Table 3.** Baseline clinical characteristics of groups treated vs not treated with Tocilizumab.

	Tocilizumab		p value
	Treated (n=58)	Not treated (n=88)	
Age	61 (54-70)	64 (54-72)	0.288
Male sex	40 (69)	57 (65)	0.600
Comorbidities	35 (61)	64 (73)	0.124
Duration of symptoms at admission (days)	6 (5-7)	7 (4-8)	0.612
Baseline PaO ₂ /FiO ₂	137 (88-232)	248 (183-348)	<0.001
Treatment during hospitalization			
Hydroxychloroquine	53 (93)	84 (98)	0.171
Lopinavir/Ritonavir	51 (89)	68 (79)	0.103
Azithromycin	33 (58)	49 (57)	0.913
Interferon-β	2 (4)	5 (6)	0.532
Glucocorticoids	38 (67)	47 (55)	0.152
Methylprednisolone bolus	31 (54)	30 (35)	0.018
Laboratory Findings			
White Blood Count (10 ³ /mm ³)	7.99 (5.17-11.85)	7.52 (5.4–10.36)	0.527
Lymphocyte Count (10 ³ /mm ³)	0.74 (0.52–0.997)	0.93 (0.66–1.47)	0.001
Creatinine (mg/dl)	0.83 (0.70-1.05)	0.90 (0.72-1.14)	0.177
LDH (U/L)	425 (302-510)	293.5 (221-388)	<0.001
CK (U/L)	69 (38-270)	75.5 (49-125)	0.785
Serum IL-6 (pg/ml)	41.85 (12.37-71.95)	16.25 (6.27-44.95)	0.007
Ferritin (ng/ml)	1888 (1152-2844)	1461 (471-1861)	0.038
CRP (mg/dL)	13.73 (8.75-27.08)	9.09 (4.78-19.31)	0.005
PCT (ng/ml)	0.25 (0.13-0.36)	0.14 (0.1-0.3)	0.045
D-dimer (mg/ml)	0.75 (0.48–1.48)	0.71 (0.53–1.22)	0.491

668 All categorical variables are expressed as number (%) and quantitative variables as median (p25-p75).

669 PaO₂/FiO₂: arterial oxygen tension - fraction of inspired oxygen ratio; LDH: Lactate Dehydrogenase; IL-

670 6: Interleukin-6; CRP: C-reactive protein; PCT: Procalcitonin.

671

672 FIGURE LEGENDS

673 **Figure 1.** IL-6 serum levels predict disease severity and tocilizumab (TCZ) use. (A) Correlation
674 between log-transformed IL-6 serum levels and arterial oxygen tension (PaO₂). (B) Correlation
675 between log-transformed IL-6 serum levels and arterial oxygen tension - fraction of inspired
676 oxygen ratio (PaO₂/FiO₂). Data in panels A and B are shown as dot-plot and their fitted linear
677 prediction with 95% confidence interval (transparent grey shadow) estimated using the
678 twoway command of Stata with the lfitci option. (C) ROC curve showing the ability of log-
679 transformed IL-6 serum levels to discriminate between patients requiring vs not-requiring
680 invasive mechanical ventilation (IMV). (D) ROC curve for the ability of log-transformed IL-6
681 serum levels to discriminate between TCZ treated and non-treated patients. The best cut-off
682 for discrimination of patients requiring IMV (panel C) or TCZ treatment (panel D) was 30
683 pg/ml.

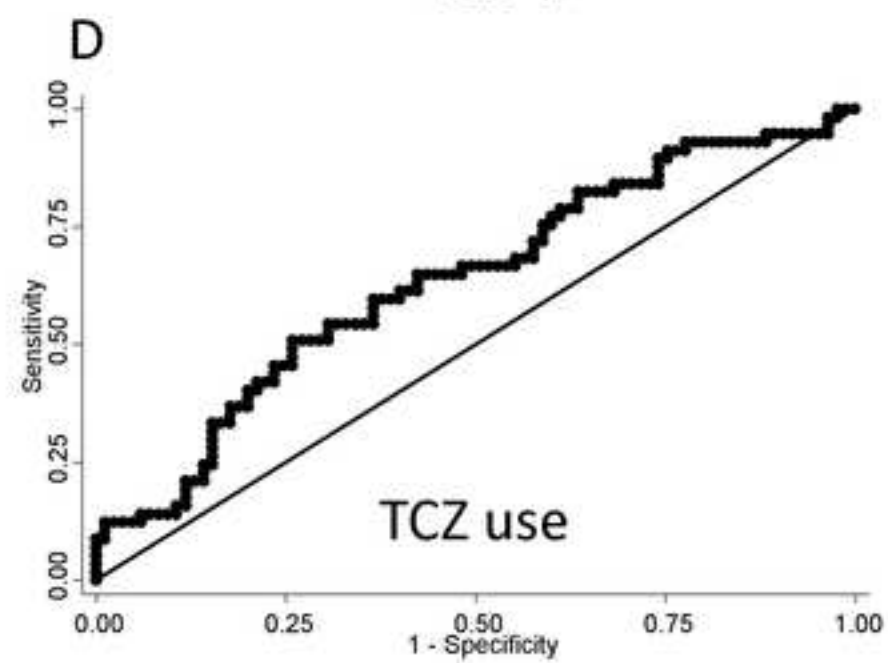
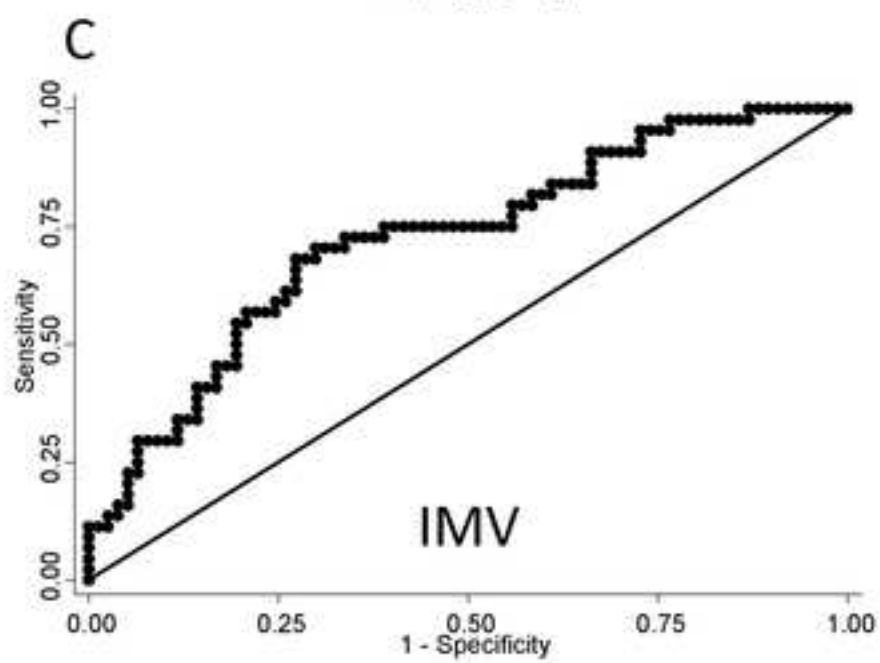
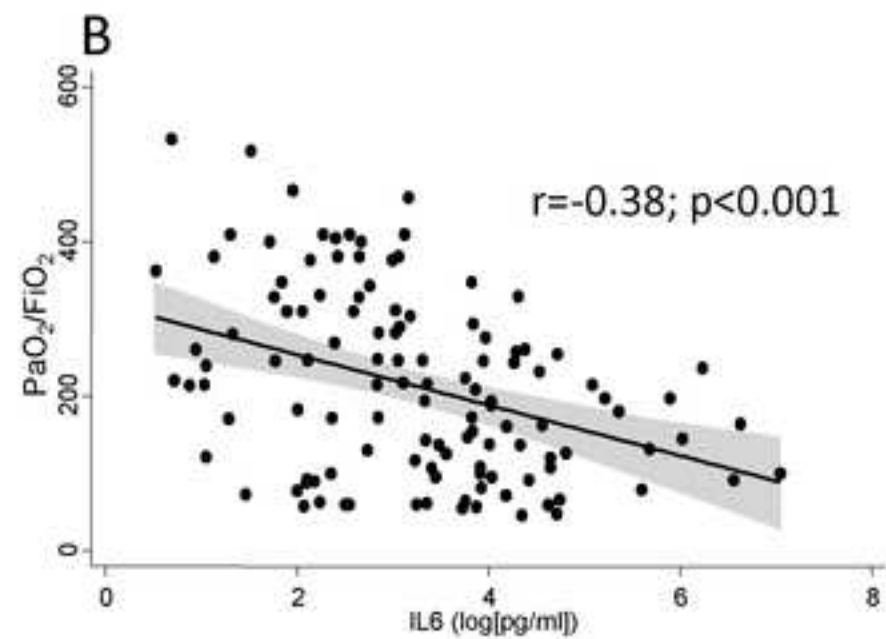
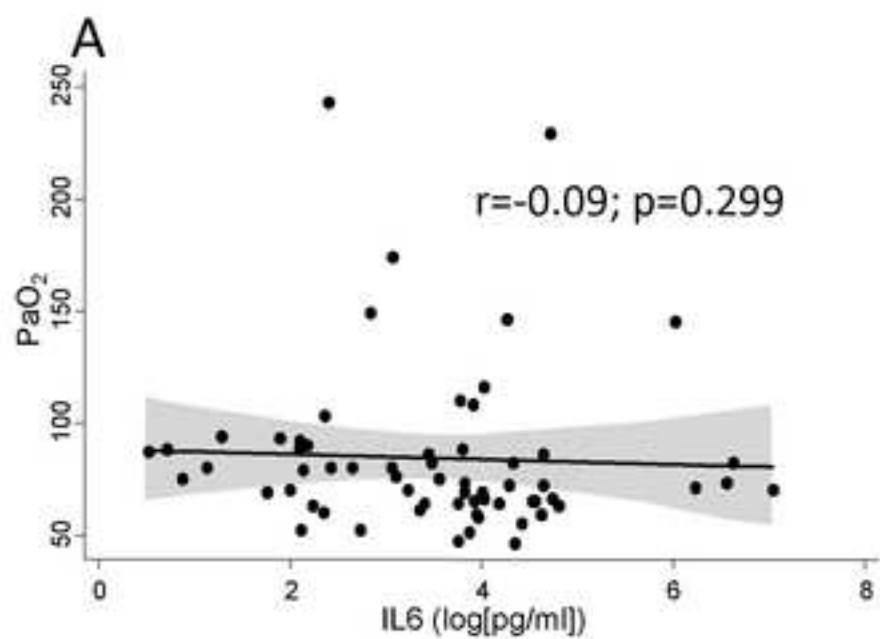
684 **Figure 2.** Response of laboratory parameters to Tocilizumab (TCZ) treatment. (A) Differences
685 in log-transformed IL-6 serum levels, (B) C-Reactive Protein and (C) Procalcitonin, (D) Ferritin,
686 (E) Lactate Dehydrogenase (LDH), and (F) D-Dimer. Data are presented as the interquartile
687 range (p75 upper edge, p25 lower edge, p50 midline), p95 (line above the box), and p5 (line
688 below the box) of levels for each parameter before (grey boxes) and after (white boxes)
689 treatment with TCZ.

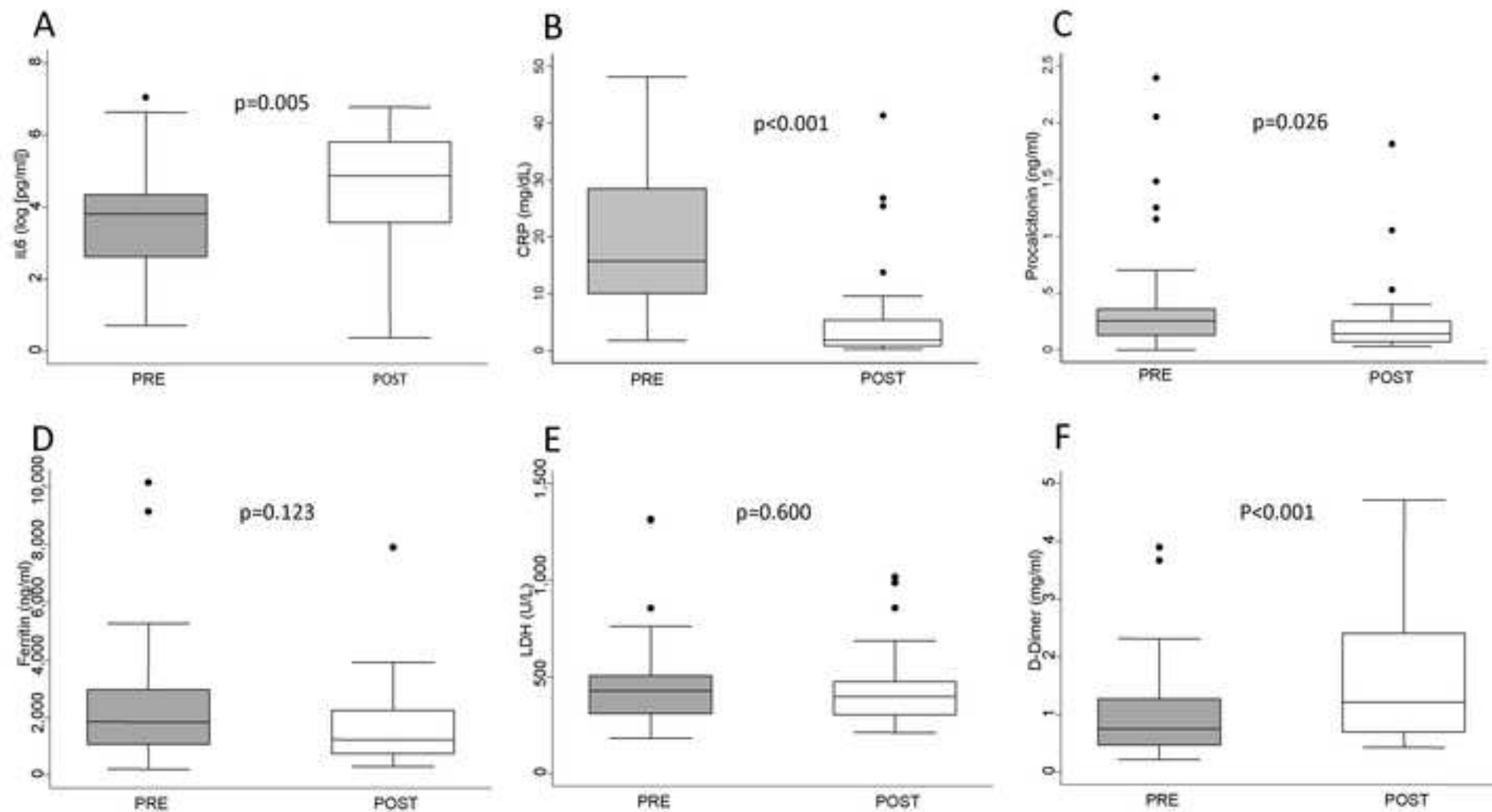
690 **Figure 3.** Change of PaO₂/FiO₂ in COVID-19 patients treated early (before 11 day of symptoms
691 onset) or late with Tocilizumab and not-treated. (A) Patients with high baseline IL-6 (cut-off
692 30 pg/ml). (B) Subjects with low baseline IL-6 serum levels. Data in A and B are shown as the
693 interquartile range (p75 upper edge, p25 lower edge, p50 midline), p95 (line above the box),
694 and p5 (line below the box) before (grey boxes) and after (white boxes) treatment with TCZ. In
695 non-treated patients PRE and POST mean first and second evaluation, respectively. Statistical

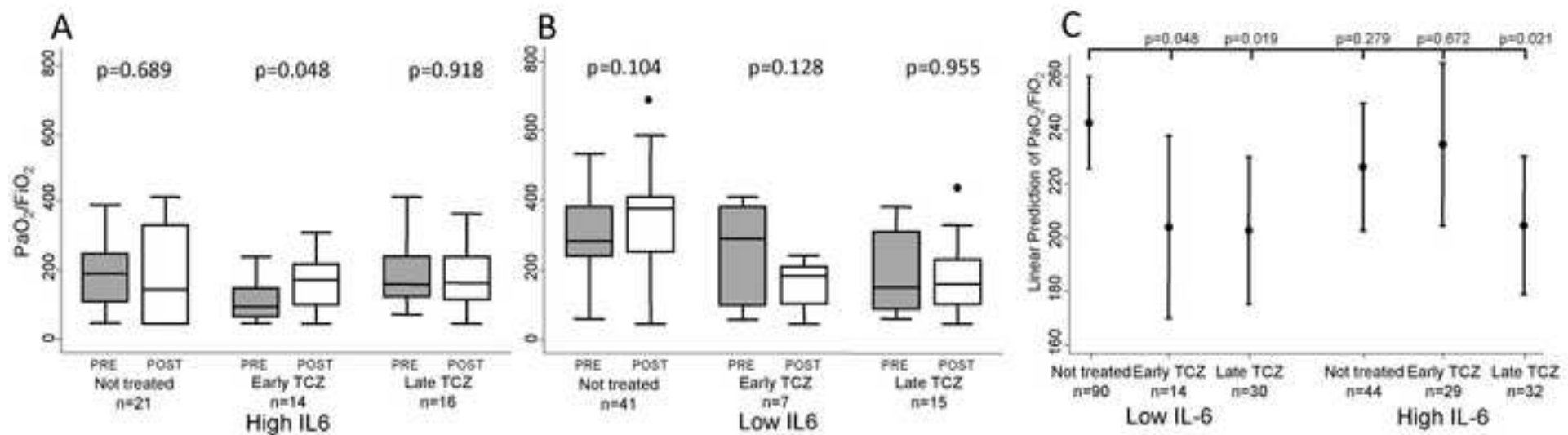
696 significance was determined with the Mann-Whitney test. (C) The graph represents the
697 predicted mean (dots) with 95% confidence intervals (bars) of PaO₂/FiO₂ according to baseline
698 IL-6 levels and early or late TCZ treatment. Data were obtained with the command
699 marginsplot of Stata, after adjustment by baseline PaO₂/FiO₂ and radiological pattern,
700 Hypertension, Lactic dehydrogenase and C-reactive protein levels, lymphocyte blood count
701 and need for IMV, according to the multivariable analysis displayed in Supplementary Table 2
702 (see Methods for further information).

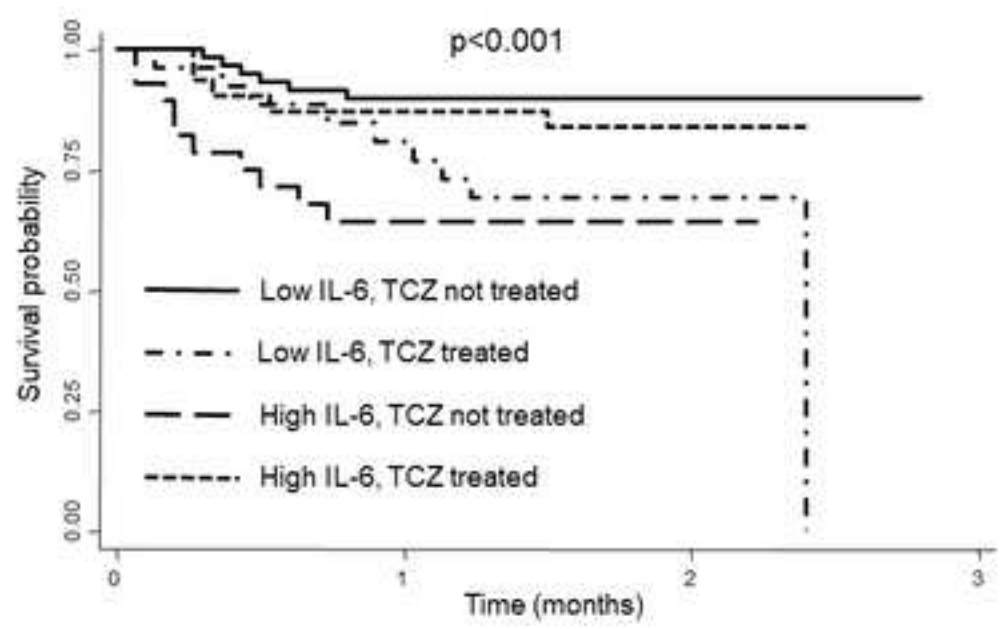
703 **Figure 4.** Survival curves of COVID-19 patients grouped according to baseline IL-6 levels and
704 TCZ treatment. Statistical significance was established with log-rank test.

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SUPPLEMENTARY DATA

Supplementary Table1: Baseline clinical characteristics of the study population and groups requiring vs not requiring Invasive Mechanical Ventilation.

	Study Population (n=146)	Invasive Mechanical Ventilation		p value
		Required (n=44)	Not-required (n=102)	
Age	63 (54-71)	63.5 (56.5-72)	62 (54-71)	0.517
Male sex	97 (66)	32 (73)	65 (64)	0.291
Comorbidities (1, 0.68%)	100(69)	30 (68)	70 (69)	0.893
Hypertension (1, 0.68%)	55 (38)	20 (45)	35 (35)	0.218
Obesity (1, 0.68%)	23 (16)	11 (25)	12 (12)	0.047
Diabetes Mellitus (1, 0.68%)	26 (18)	8 (18)	18 (18)	0.883
COPD (1, 0.68%)	9 (6)	5 (11)	4 (4)	0.089
Immune-mediated Disease† (1, 0.68%)	8 (6)	3 (7)	5 (5)	0.702
History of Malignancy‡ (1, 0.68%)	19 (13)	5 (11)	14 (14)	0.792
Others§ (1, 0.68%)	76 (52)	25 (57)	51 (52)	0.558
Duration of symptoms at admission (days) (1, 0.68%)	6 (4-7)	5 (5-7)	7 (4-8)	0.265
Fever at admission ($\geq 38^{\circ}\text{C}$) (15, 10%)	36 (27)	14 (32)	21 (25)	0.389
Baseline PaO ₂ /FiO ₂ (5, 3%)	215 (112-310)	125.5 (75-207)	247 (172-348)	<0.001
Treatment during hospitalization				
Hydroxychloroquine (2, 1.35%)	137 (96)	38 (86)	99 (100)	<0.001
Lopinavir/Ritonavir (2, 1.35%)	119 (83)	38 (86)	81 (82)	0.502
Azithromycin (2, 1.35%)	82 (57)	24 (55)	58 (59)	0.652
Interferon- β (2, 1.35%)	7 (5)	3 (7)	4 (4)	0.676
Glucocorticoids (2, 1.35%)	85 (59)	27 (61)	58 (59)	0.755
Methylprednisolone bolus (1, 0.68%)	61 (42)	21 (48)	40 (40)	0.414
Laboratory Findings				
White Blood Count ($10^3/\text{mm}^3$) (9, 6%)	7.64 (5.25–10.68)	9.39 (6.59-13.31)	6.93 (5.13-8.78)	<0.001
Lymphocyte Count ($10^3/\text{mm}^3$) (10, 6.76%)	0.83 (0.60–11.7)	0.74 (0.58-1.08)	0.87 (0.62-1.26)	0.029
Creatinine. mg/dl (7, 4.73%)	0.86 (0.70-1.10)	0.99 (0.71-1.20)	0.85 (0.72-1.1)	0.398
Bilirubin. mg/dl (9, 6.08%)	0.55 (0.41-0.87)	0.62 (0.44-1.15)	0.53 (0.39-0.78)	0.057
AST. U/L (9, 6.08%)	41 (28-64)	43 (30-60)	39 (27-70)	0.526
ALT. U/L (9, 6.08%)	37 (24-68)	30 (22-63)	39 (24-71)	0.287
GGT. U/L (26, 17.57%)	73 (36-159)	81.5 (41-152)	67 (36-159)	0.623
LDH. U/L (14, 9.46%)	341 (256-461)	413 (315-496)	302 (224-443)	0.001
CK. U/L (102, 69%)	72 (48-155)	67 (39.50-167.50)	94 (59-140)	0.617
Serum IL-6. pg/ml (7, 4.73%)	21.36 (7.53-54.21)	49.20 (17.28-103.57)	16.08 (6.09-42.03)	<0.001
Ferritin. ng/ml (96, 64.86%)	1598 (830-2305)	1665 (602-2765)	1573 (1012-2300)	0.832
CRP. mg/dL (23, 15.54%)	11.55 (5.16-22.53)	17.09 (7.69-28.98)	10.13 (4.83-18.48)	0.003
PCT. ng/ml (52, 35.14%)	0.15 (0.10-0.35)	0.29 (0.14-0.46)	0.13 (0.08-0.26)	0.001
D-dimer. mg/ml (25, 16.89%)	0.75 (0.48–1.48)	0.92 (0.56-2.31)	0.71 (0.48-1.19)	0.058
Radiologic findings (1, 0.68%)				0.209
Clear	11 (8)	4 (9)	8 (7)	
Diffuse ground glass opacities	50 (35)	13 (30)	37 (37)	
Unilateral alveolar pattern	20 (14)	3 (7)	17 (17)	
Bilateral alveolar pattern	63 (43)	24 (55)	37 (37)	

All categorical variables are expressed as number (%) and quantitative variables as median (p25-p75). Missing data in each clinical characteristic are expressed as: (number, %). Variables not disclosing it do not present any missing values. COPD: Chronic obstructive pulmonary disease; PaO₂/FiO₂: arterial oxygen tension - fraction of inspired oxygen ratio; AST: Aspartate amino-transferase; ALT: Alanine amino- transferase; GGT: Gamma-glutamyl transferase; LDH: Lactate Dehydrogenase; CK: Creatine-kinase; IL-6: Interleukin-6; CRP: C-reactive protein; PCT: Procalcitonin.

†Includes Rheumatoid Arthritis, Systemic Lupus Erythematosus, Ulcerative Colitis, etc

‡Includes solid organ and hematologic malignancies

§Includes obstructive sleep apnea syndrome, asthma, hypothyroidism, Ischemic cardiomyopathy, etc

Supplementary Table 2. Logistic regression model for invasive mechanical ventilation.

	OR	p	95%CI
COPD	5.407	0.030	1.172 to 24.938
White Blood Count (10^3)	1.050	0.116	0.998 to 1.115
High IL-6 baseline serum levels	7.087	<0.001	3.022 to 16.617

OR: Odds Ratio; 95%CI: 95% Confidence Interval ;COPD: Chronic Obstructive Pulmonary Disease; IL-6: interleukin 6; High IL-6 was considered if >30 pg/ml.

Supplementary Table 3: Baseline clinical characteristics of groups treated vs not treated with Tocilizumab.

	Tocilizumab (TCZ)		p value
	Treated (n=58)	Not-treated (n=88)	
Age	61 (54-70)	64 (54-72)	0.288
Male sex	40 (69)	57 (65)	0.600
Comorbidities (1, 0.68%)	35 (61)	64 (73)	0.124
Hypertension (1, 0.68%)	17 (30)	37 (42)	0.124
Obesity (1, 0.68%)	14 (25)	9 (10)	0.023
Diabetes Mellitus (1, 0.68%)	9 (16)	16 (18)	0.687
COPD (1, 0.68%)	1 (2)	8 (9)	0.071
Immune-mediated Disease† (1, 0.68%)	5 (9)	3 (3)	0.181
History of Malignancy# (1, 0.68%)	7 (12)	12 (14)	0.763
Others‡ (1, 0.68%)	30 (52)	46 (52)	0.892
Duration of symptoms at admission (days) (1, 0.68%)	6 (5-7)	7 (4-8)	0.612
Fever at admission ($\geq 38^{\circ}\text{C}$) (15, 10%)	17 (31)	19 (25)	0.483
Baseline PaO ₂ /FiO ₂ (5, 3%)	137 (88-232)	248 (183-348)	<0.001
Treatment during hospitalization			
Hydroxychloroquine (2, 1.35%)	53 (93)	84 (98)	0.171
Lopinavir/Ritonavir (2, 1.35%)	51 (89)	68 (79)	0.103
Azithromycin (2, 1.35%)	33 (58)	49 (57)	0.913
Interferon- β (2, 1.35%)	2 (4)	5 (6)	0.532
Glucocorticoids (2, 1.35%)	38 (67)	47 (55)	0.152
Methylprednisolone bolus (1, 0.68%)	31 (54)	30 (35)	0.018
Laboratory Findings			
White Blood Count ($10^3/\text{mm}^3$) (9, 6%)	7.99 (5.17-11.85)	7.52 (5.4-10.36)	0.527
Lymphocyte Count ($10^3/\text{mm}^3$) (10, 6.76%)	0.74 (0.52-0.997)	0.93 (0.66-1.47)	0.001
Creatinine. mg/dl (7, 4.73%)	0.83 (0.70-1.05)	0.90 (0.72-1.14)	0.177
Bilirubin. mg/dl (9, 6.08%)	0.62 (0.46-1.04)	0.52 (0.38-0.78)	0.070
AST. U/L (9, 6.08%)	47 (29-77.50)	34.5 (28-53)	0.047
ALT. U/L (9, 6.08%)	37.5 (25-77)	36 (22.5-64.5)	0.588
GGT. U/L (26, 17.57%)	73 (41-186)	71 (35-145)	0.374
LDH. U/L (14, 9.46%)	425 (302-510)	293.50 (221.50-388.50)	<0.001
CK. U/L (102, 69%)	69 (38-270)	75.5 (49-125)	0.785
Serum IL-6. pg/ml (7, 4.73%)	41.85 (12.37-71.95)	16.25 (6.27-44.95)	0.007
Ferritin. ng/ml (96, 64.86%)	1888 (1152-2844)	1461 (471-1861)	0.038
CRP. mg/dL (23, 15.54%)	13.73 (8.75-27.08)	9.09 (4.78-19.31)	0.005
PCT. ng/ml (52, 35.14%)	0.25 (0.13-0.36)	0.14 (0.1-0.3)	0.045
D-dimer. mg/ml (25, 16.89%)	0.75 (0.48-1.48)	0.71 (0.53-1.22)	0.491
Radiologic findings (1, 0.68%)			0.818
Clear	4 (7)	8 (8)	
Diffuse ground glass opacities	18 (32)	32 (37)	
Unilateral alveolar pattern	10 (18)	10 (11)	
Bilateral alveolar pattern	25 (44)	37 (43)	

All categorical variables are expressed as number (%) and quantitative variables as median (p25-p75). Missing data are expressed as number (%). Variables not disclosing it do not present any missing values.

COPD: Chronic obstructive pulmonary disease; PaO₂/FiO₂: arterial oxygen tension - fraction of inspired oxygen ratio; AST: Aspartate amino-transferase; ALT: Alanine amino- transferase; GGT: Gamma-glutamyl transferase; LDH: Lactate Dehydrogenase; CK: Creatine-kinase; IL-6: Interleukin-6; CRP: C-reactive protein; PCT: Procalcitonin.

†Includes Rheumatoid Arthritis, Systemic Lupus Erythematosus, Ulcerative Colitis, etc

‡Includes solid organ and hematologic malignancies

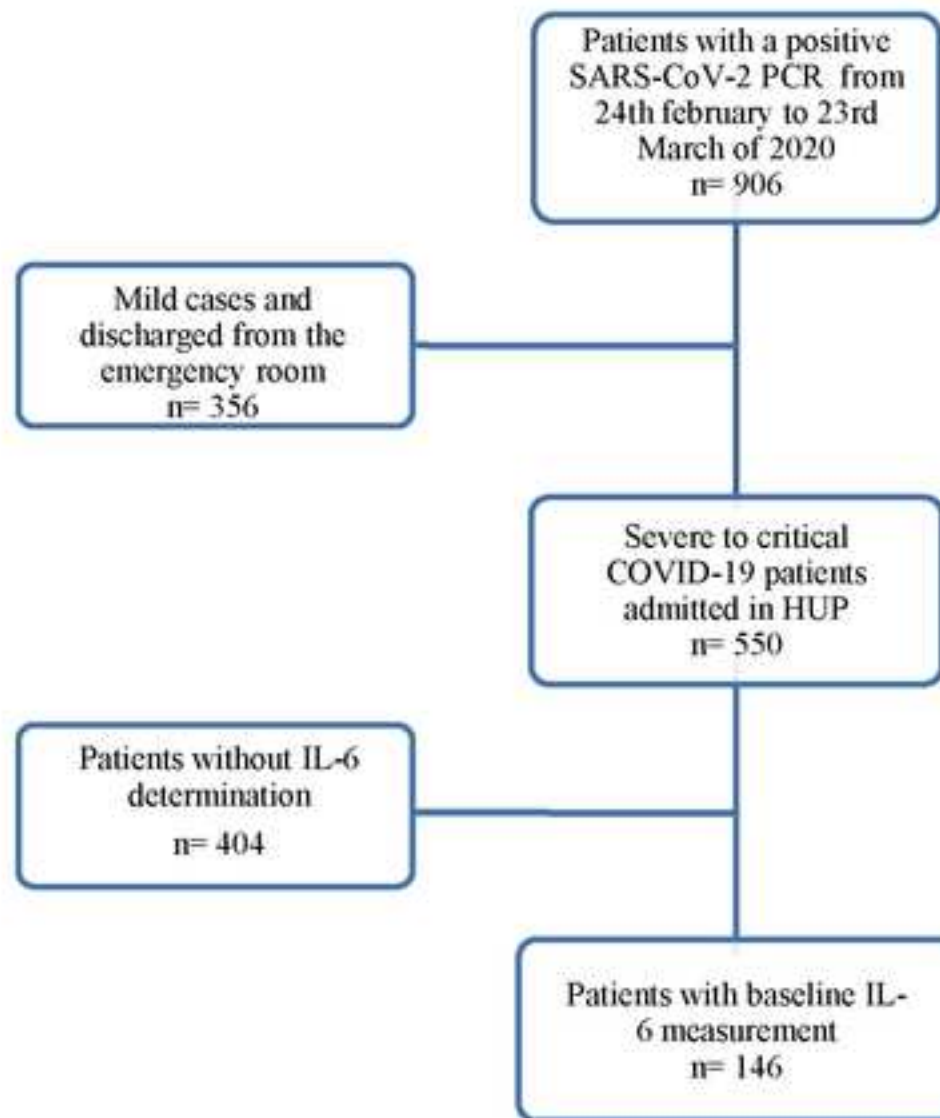
§Includes obstructive sleep apnea syndrome, asthma, hypothyroidism, Ischemic cardiomyopathy, etc

Supplementary Table 4. Variables explaining evolution of PaO₂/FiO₂

	β coeff.	p	95%CI
Baseline PaO ₂ /FiO ₂	0.713	<0.001	0.567 to 0.753
Lymphocyte Count	0.033	<0.001	0.015 to 0.052
Hypertension	-19.059	0.067	-39.432 to 1.314
LDH	- 0.101	0.001	-0.163 to -0.039
CRP	- 1.188	0.038	-2.311 to 0.065
Radiologic Findings			
Clear	ref		
Diffuse ground glass opacities	49.256	0.006	13.785 to 84.727
Unilateral alveolar pattern	31.638	0.131	-9.378 to 72.654
Bilateral alveolar pattern	60.708	0.001	25.986 to 95.431
Others	-123.818	0.113	-276.853 to 29.217
IL-6 serum / TCZ*			
Low IL-6, no TCZ	ref		
Low IL-6, early TCZ	-38.854	0.048	-77.390 to -0.319
Low IL-6, late TCZ	-40.150	0.019	-73.765 to -6.535
High IL-6, no TCZ	-16.513	0.279	-46.440 to 13.413
High IL-6, early TCZ	-8.134	0.672	-45.831 to 29.562
High IL-6, late TCZ	-38.359	0.021	-70.837 to -5.881

PaO₂/FiO₂: arterial oxygen tension - fraction of inspired oxygen ratio; β coeff.: Beta-coefficient; 95%CI: 95% Confidence Interval; LDH: Lactate Dehydrogenase; CRP: C-reactive protein; IMV: Invasive Mechanical Ventilation; ref: reference; IL-6: interleukin 6; TCZ: Tocilizumab

* IL-6 serum / TCZ: refers to the interaction between both high or low IL-6 (cut-off 30 pg/ml) and TCZ (No treatment, Early or late treatment; cut-off 11 days since symptoms onset) as an independent predictor within the model.

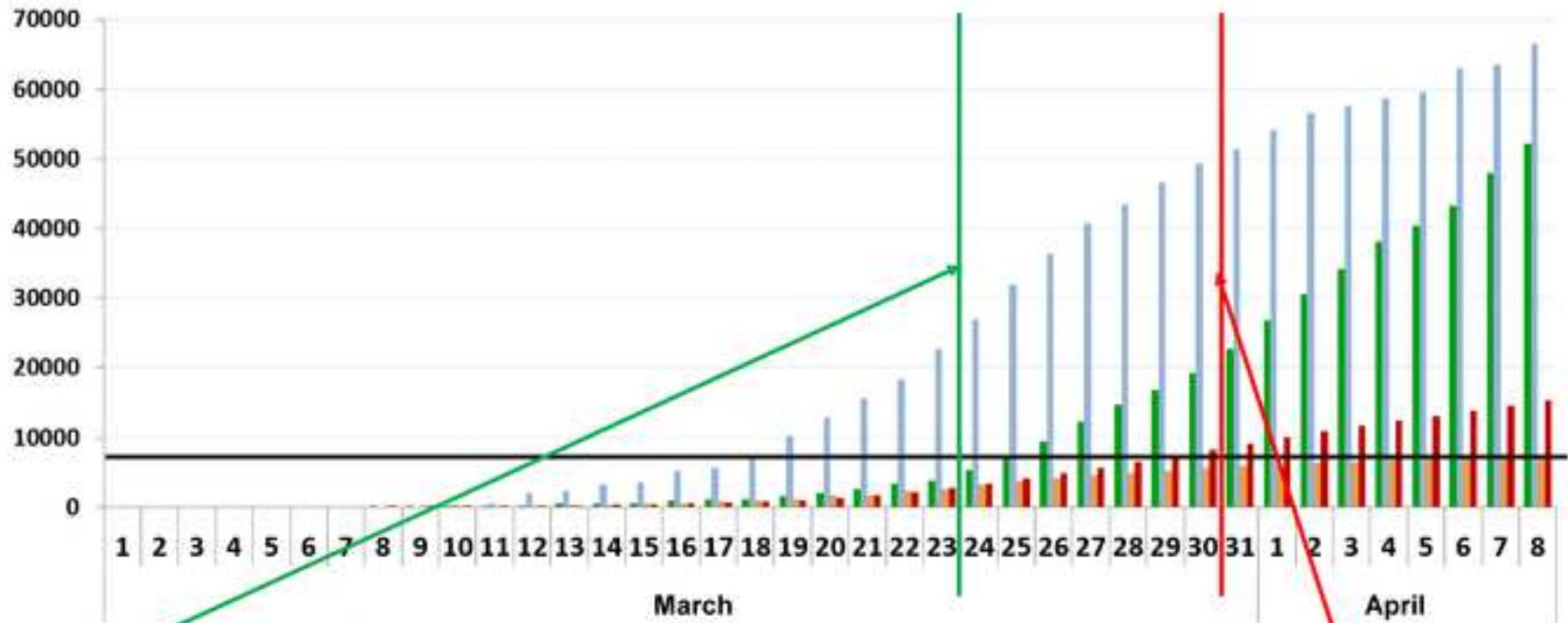


Sociodemographic characteristics of patients hospitalized due to COVID-19 until 24th March 2020.

Sociodemographic Characteristics	All COVID-19 hospitalized population (n=550)	Patients with IL-6 measurement		p value
		Yes (n=146)	No (n=404)	
Age	66.6 [55.0 – 77.9]	63.0 [54.0 – 72]	68.9 [55.2 – 80.0]	0.0001
Male sex	329 (59.8)	97 (66.4)	232 (57.4)	0.057
Mortality during hospitalization	115 (20.9)	30 (20.6)	85 (21.0)	0.900

Categorical variables are expressed as number (%) and quantitative variables as median [interquartile range]

Daily evolution of COVID-19 in Spain according to clinical situation

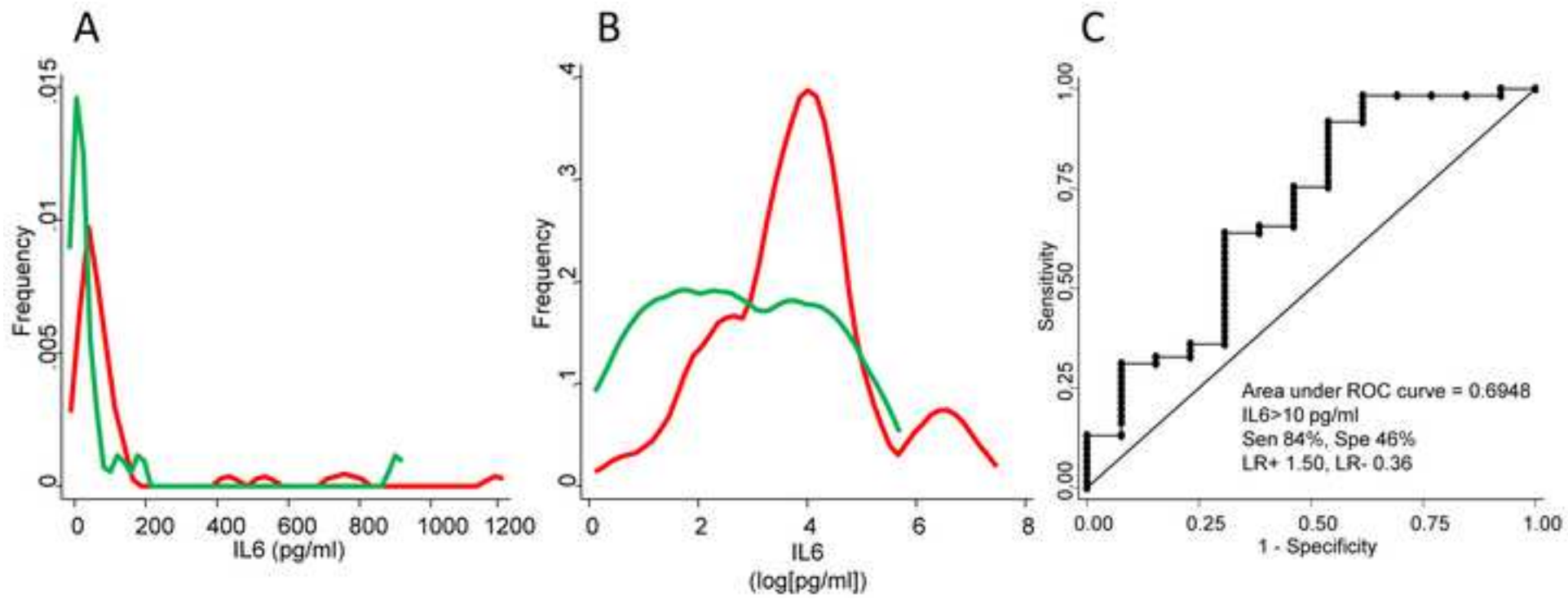


Deadline for inclusion of COVID-19 patients in the study

■ Discharged ■ Hospitalized ■ ICU ■ Exitus

Source: RENAVE. ISCIII-CCAES

Time to Intensive Care Unit (ICU Orange bars) overrun



LEGENDS FOR THE SUPPLEMENTARY FIGURES

Supplementary Figure 1. Flow chart of patients included in the study.

Supplementary Figure 2. Histogram showing the daily evolution of COVID-19 cases in Spain according to their clinical situation.

Source: <https://covid19.isciii.es/resources/CURVASTATUS.png>. Accessed on April 9th 2020.

Supplementary Figure 3. (A) Plots showing the distribution of raw IL-6 serum levels in patients (red) vs healthy donors (green). (B) Distribution of log-transformed serum IL-6 levels in the former groups. (C) ROC curve showing the ability of IL-6 serum levels to classify COVID-19 patients vs healthy donors. (A) Plots showing the distribution of raw IL-6 serum levels in patients (red) vs healthy donors (green). (B) Distribution of log-transformed serum IL-6 levels in the former groups. (C) ROC curve showing the ability of IL-6 serum levels to classify COVID-19 patients vs healthy donors.