University of Texas Rio Grande Valley

ScholarWorks @ UTRGV

School of Medicine Publications and Presentations

School of Medicine

2-2024

Therapeutic potential of snake venom: Toxin distribution and opportunities in deep learning for novel drug discovery

Anas Bedraoui

Montamas Suntravat Texas A & M University - Kingsville

Salim El Mejjad

Salwa Enezari

Naoual Oukkache

See next page for additional authors

Follow this and additional works at: https://scholarworks.utrgv.edu/som_pub

Part of the Animal Sciences Commons, and the Medicine and Health Sciences Commons

Recommended Citation

Bedraoui, A., Suntravat, M., El Mejjad, S., Enezari, S., Oukkache, N., Sanchez, E. E., ... & Daouda, T. (2023). Therapeutic Potential of Snake Venom: Toxin Distribution and Opportunities in Deep Learning for Novel Drug Discovery. Medicine in Drug Discovery, 100175. https://doi.org/10.1016/j.medidd.2023.100175

This Article is brought to you for free and open access by the School of Medicine at ScholarWorks @ UTRGV. It has been accepted for inclusion in School of Medicine Publications and Presentations by an authorized administrator of ScholarWorks @ UTRGV. For more information, please contact justin.white@utrgv.edu, william.flores01@utrgv.edu.

Authors

Anas Bedraoui, Montamas Suntravat, Salim El Mejjad, Salwa Enezari, Naoual Oukkache, Elda E. Sanchez, Jacob Galan, Rachid El Fatimy, and Tariq Daouda



Contents lists available at ScienceDirect

Medicine in Drug Discovery



journal homepage: www.elsevier.com/locate/medid

Review Article

Therapeutic potential of snake venom: Toxin distribution and opportunities in deep learning for novel drug discovery

Anas Bedraoui^a, Montamas Suntravat^{b,c}, Salim El Mejjad^a, Salwa Enezari^a, Naoual Oukkache^e, Elda E. Sanchez^{b,c}, Jacob A. Galan^d, Rachid El Fatimy^a, Tariq Daouda^{a,*}

^a Institute of Biological Sciences (ISSB-P), UM6P- Faculty of Medical Sciences (FMS), Mohammed VI Polytechnic University (UM6P), Ben Guerir, Morocco

^b National Natural Toxins Research Center (NNTRC), Texas A&M University-Kingsville, Kingsville, TX, USA

^c Department of Chemistry, Texas A&M University-Kingsville, Kingsville, TX, USA

^d Department of Human Genetics, University of Texas Rio Grande Valley, Brownsville, TX, USA

e Laboratoire des Venins et Toxines, Département de Recherche, Institut Pasteur du Maroc, 1, Place Louis Pasteur, Casablanca 20360, Morocco

ARTICLE INFO

Keywords: Snake venom Bioactive molecules Medical applications Drug discovery Toxin distribution Artificial intelligence Deep learning Machine Learning

ABSTRACT

Snake venom is a rich source of bioactive molecules that hold great promise for therapeutic applications. These molecules can be broadly classified into enzymes and non-enzymes, each showcasing unique medicinal properties. Noteworthy compounds such as Bradykinin Potentiating Peptides (BPP) and Three-Finger Toxins (3FTx) are showing therapeutic potential in areas like cardiovascular diseases (CVDs) and pain-relief. Meanwhile, components like snake venom metalloproteinases (SVMP), L-amino acid oxidases (LAAO), and Phospholipase A₂s (PLA₂) are paving new ways in oncology treatments. The full medicinal scope of these toxins is still emerging. In this review, we discuss drugs derived from snake venoms that address CVDs, cancer, diabetes, strokes, and pain. Further, we outline the toxin distribution across 130 snake species, categorized by their genus within the Crotalidae, Viperidae, and Elapidae families. Conclusively, we spotlight the potential of Deep Learning (DL) in discovering groundbreaking drug prospects from these toxins.

1. Introduction

Snake venom is a complex mixture of abundant components, containing a variety of biologically active molecules with peptides and proteins accounting for more than 95 % of snake venom dry weight (the other 5 % contain lipids, carbohydrates, and biogenic amines) [1]. These proteins perform various functions aiding in capturing and digestion of prey after envenomation. Some proteins function as enzymes catalyzing chemical reactions that disrupt coagulation and induce hemorrhage, while others interfere with cellular receptors, causing paralysis. The venom's effects on the body can be hemotoxic, cytotoxic, or neurotoxic, leading to blood cell damage, tissue inflammation, or nervous system disruption, respectively. The specific physiological impacts depend on the snake species and venom composition [2].

Snake venom composition can be divided into two groups of proteins and peptides: enzymatic and non-enzymatic molecules [3]. Enzymatic molecules are proteins with catalytic sites and activity, able to speed up chemical reaction rates in the presence of a substrate. The most common and abundant enzymatic molecules are snake venom phospholipases A₂ (PLA₂), snake venom metalloproteinases (SVMP), snake venom serine proteases (SVSP), and L-amino acid oxidases (LAAO) [4]. The second group is the non-enzymatic molecules found in snake venom. These include neurotoxins, affecting the nervous system; cardiotoxins, affecting the heart; and cytotoxins, damaging cells. The most common non-enzymatic molecules are three-finger toxins (3FTx), Kunitz peptides (KUN), disintegrins (DIS), and cysteine-rich secretory protein (CRiSP) [5] (Fig. 1).

Chronic diseases, including CVDs, cancer, diabetes, and chronic pain, are the leading causes of death and disability worldwide. These conditions have been treated using drugs derived from Viperidae and Elapidae venoms. For example, Captopril, used for hypertension and diabetic nephropathy, was developed from the BPP in snake venom [6,7]. DIS, peptides isolated from snake venoms and act as anti-coagulant agents, inhibiting platelet aggregation and promoting hemorrhage, resulted in the development of Eptifibatide and Tirofiban, two FDA-approved drugs for the treatment of acute coronary syndrome. Among approved venom-derived molecules for the treatment of chronic diseases and pain, we find Captopril, Enalapril, Eptifibatide, Tirofiban, Defibrase, Reptilase,

* Corresponding author. *E-mail address:* Tariq.daouda@um6p.ma (T. Daouda).

https://doi.org/10.1016/j.medidd.2023.100175

Received 4 December 2023; Received in revised form 19 December 2023; Accepted 26 December 2023 Available online 27 December 2023

2590-0986/© 2023 Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).



Fig. 1. 3D structures of toxins used in the treatment of chronic diseases and pain, classified into enzymatic and non-enzymatic components. Labels identify therapeutic effects of each molecule. The 3D structure of BPP was generated using Alphafold2 from its primary structure, and all other 3D structures were obtained from Uniprot [50].

and Cobratide that have either been approved by the FDA (USA), the EMA (Europe), or the National Medical Products Administration (NMPA) (China) [8–12]. Table 1 summarizes the uses of the seven approved drugs, including their UniProt ID, the identified mechanism of action, and the approval jurisdiction.

2. Snake venom composition

According to the reptile database, there are more than 3,900 snake species worldwide [45]. The Viperidae and Elapidae families contain over 700 known species of venomous snakes [45] with venoms that are either hemotoxic, neurotoxic, or cytotoxic. The Viperidae family includes rattlesnakes, copperheads, and cottonmouths, known for their hemotoxic and cytotoxic effects [46]. This family is responsible for approximately 98 % of all envenomations in the United States [47] and nearly 50 % in India [48]. The Elapidae, which include cobras, coral snakes, kraits, and mambas, are known for their neurotoxic effects, which in most cases cause paralysis [49].

Crotalinae and Viperinae are both subfamilies of the Viperidae family and present similar protein distributions (Fig. 2A) with percentages of SVMP, PLA₂, and SVSP being similar between the two subfamilies. The primary distinction between the two is the presence of 3FTx in Elapidae and its complete absence in Viperidae (Fig. 2A). One of the most noticeable similarities between the Viperidae and Elapidae families is the dominant presence of PLA₂. However, the Elapidae has a higher percentage of 3FTx (57 %), PLA₂ (33 %), and a lower percentage of SVMP (3 %) (Fig. 2A).

The proteomic composition of snake venoms varies depending on several factors, such as location, leading to differences in the dominant protein percentages among different populations of the same species. Despite these differences, all populations of *B. atrox* from Brazil, Venezuela, Peru, and Colombia have a higher percentage of SVSP, indicating their relative importance in the venom composition. Interestingly, PLA₂, SVMP, and SVSP are abundant, contributing to the venom's toxicity and other pharmacological effects (Fig. 2B).

PLA₂, SVMP, and SVSP are prevalent in most *Crotalus* species; however, the venom of *Crotalus durissus cascavella* and *C. d. collilineatus* contains a higher abundance of PLA₂ and a comparatively lower abundance of SVSP and SVMP (Fig. 2C). Additionally, the venom of *C. tigris* demonstrates a distinct profile, exhibiting the presence of only SVMP and SVSP with an absence of PLA₂. *Naja* venom contains a significant abundance of 3FTx, one of the most abundant proteins in the Elapidae family (Fig. 2D). Moreover, the presence of PLA₂, a vital protein, in *Naja* venoms underscores the complexity and diversity of its composition. The lower presence of SVMP in *Naja* venom demonstrates the unique composition of this genus compared to other members of the Elapidae family.

The percentage distribution of snake species and protein families among Viperidae, Elapidae, and Crotalinae families is depicted in (Fig. 2A). The abundances of PLA₂, SVMP, and SVSP are similar for both Viperinae and Crotalinae. Conversely, the Elapidae family is distinguished from the others by the predominance of 3FTx proteins in their venom compositions (Fig. 2D). Finally, the high abundance of PLA₂ in all families strongly suggests a primary role in toxicity.

2.1. Bradykinin Potentiating peptides (BPP)

Bradykinin Potentiating Peptides from snake venoms are a group of small peptides that range from 5 to 14 amino acids in length. Based on their structural characteristics, they can be divided into two major groups:

- Bradykinin Potentiating Peptides contain a pyroglutamyl residue at the N-terminus and a proline-rich sequence. The proline-rich sequence is not necessarily located at a specific position within the peptide. Rather, it suggests that the peptide contains a high number of proline residues compared to other amino acids. The exact position of these proline residues can vary depending on the specific BPP [51].
- Bradykinin Potentiating Peptides with a C-terminal extension beyond the proline-rich sequence [15].

The presence of proline residues is crucial for the biological activity of BPP as they confer resistance to degradation by proteases [51,52].

Bradykinin Potentiating Peptides work by inhibiting the ACE (Angiotensin-converting enzyme), a critical enzyme in the reninangiotensin system (RAS) that converts angiotensin I (ATI) to angiotensin II (ATII) [53,54]. ATII is a potent vasoconstrictor and plays a critical role in blood pressure regulation. By inhibiting ACE, BPP effectively potentiate the action of bradykinin, leading to vasodilation and hypotension. This mechanism of action has led to the development of ACE inhibitors as a widely used class of antihypertensive drugs, with the first approved drug, captopril, being inspired by a BPP from the venom of the Brazilian viper *B. jararaca* [55,56].

Bradykinin Potentiating Peptides' primary effect is to lower blood pressure when envenoming the prey [57]. Sequencing studies of BPP highlighted the pharmacologically active Phe-Ala-Pro amino acid structure, which led to its use in developing new drugs [28]. This compound is the origin of Captopril, the first FDA-approved ACE inhibitor for the treatment of hypertension [6]. Captopril functions as an

Table 1

Approved drugs derived from snake venoms for chronic disease and pain treatments.

Chronic disease	Snake species	Proteins/ Peptides	Molecular weight (Da)	UniProt ID [13]	Proteomic identification tools	Potential applications	Approved drugs	Approval	Mechanism of action
Diabetes / CVD	Bothrops jararaca	ВЪЪ	1,059 Da [14]	P0C7J8	Size exclusion chromatography, RP- HPLC chromatography [15]	Treatment of hypertension, cardiac failure, diabetic nephropathy	Captopril [6] and Enalapril [16]	Captopril: FDA- approved to treat hypertension and diabetic nephropathy in 1981 [6]. Enalapril: EMA- approved in 2003 [17]	Inhibits angiotensin- converting enzyme (ACE) [18]
CVD	Sistrurus miliarius barbouri	DIS barbourin	7,701 Da [19]	P22827	RP-HPLC chromatography [10]	Anti-platelet agent, reduce ischemic cardiac events [20]	Eptifibatide [21]	FDA-approved in 1998 and EMA- approved in 1999 for the treatment of acute coronary syndrome [22,23]	Prevents fibrinogen and von Willebrand factor from binding to GP IIb/ IIIa [21]
CVD	Echis carinatus	DIS echistatin	5,425 Da [19,24]	P17347	Gel filtration, cation exchange chromatography, RP-HPLC chromatography [24]	Reduce ischemic heart events [25], anti-platelet agent [26]	Tirofiban [27,28]	FDA-approved in 1998 and EMA-approved in 1999 for the treatment of acute coronary syndrome [29,30]	Inhibits GP IIb/ IIIa receptors [31]
Stroke / CVD	Bothrops moojeni	Batroxobin	25,503 Da [32]	P04971	RP-HPLC chromatography [32]	Treatment of thrombotic diseases, anti- coagulation, inhibit stroke development [33]	Defibrase [34]	Approved for the treatment of stroke and ischemic attack in 2006 in China [35]. Approved for the treatment of deafness in Japan [36,37]	Converts plasminogen into plasmin and cleaves fibrinogen Aα-chain [38]
CVD	Bothrops atrox	SVSP	28,189 Da [39]	P04971	RP-HPLC chromatography [40]	Treatment of thrombotic diseases [41]	Reptilase [41]	Approved for the treatment of both internal and external haemorrhages in Japan, India, and South Korea [2].	Converts fibrinogen to fibrin and activates factor X [41]
Pain	Naja naja atra	3FTx	9,262 Da [42]	P60770	Sheathless capillary electrophoresis-mass spectrometry (CE- MS) [43]	Treatment of chronic pain	Cobratide [43]	Approved for managing moderate to severe pain in China in 1978[44]	Blocks the nicotinic acetylcholine receptor (nAChR)

ACE inhibitor, which is responsible for the conversion of ATI to ATII [58]. Recent studies have shown that Captopril, either alone or in combination with furosemide or hydrochlorothiazide, significantly impacts the treatment of CVDs [6,59].

Captopril also showed promise as a treatment for both type 1 and type 2 diabetes [60]. It prevents renal function deterioration in insulindependent diabetic nephropathy and is more effective than blood pressure control alone [61]. Its effects include reducing ATII levels, leading to vasodilation and lower blood pressure, which is particularly important for diabetic patients as high blood pressure can exacerbate kidney damage and increase cardiovascular risk [7]. Furthermore, captopril indirectly decreases aldosterone production by lowering ATII levels. Aldosterone, a hormone responsible for sodium and water retention in the kidneys, contributes to increased blood volume and blood pressure [54]. Consequently, captopril's ability to reduce both ATII and aldosterone levels contributes to better blood pressure management and kidney protection in diabetic patients [61,62].

2.2. Secreted Phospholipase A₂ (PLA₂)

Secreted Phospholipase A2 are a heterogeneous family of enzymes that play a significant role in the toxicity of snake venoms. These enzymes share a conserved structural scaffold, with a molecular weight of 13–19 kDa, 5–8 disulfide bridges, and an ability to form dimers in

aqueous environments [63]. PLA₂ exert their toxic effects by specifically catalyzing the hydrolysis of the *sn*-2 ester bond in glycerophospholipids, resulting in the release of fatty acids and lysophospholipids [64]. The mechanisms of action of PLA₂ depend on the specific group of enzymes and their target sites. Group I PLA₂ are primarily neurotoxic, acting on neuromuscular junctions and causing paralysis. In contrast, Group II PLA₂ exhibit a wide range of pharmacological effects, such as myotoxicity, anti-coagulant activity, and local tissue damage [65,66].

Studies have shown that BnSP-6, a Lys-49 PLA₂ isolated from the venom of *B. pauloensis*, has anti-tumoral effects on human breast cancer cell line MDA-MB-231, killing cells through a mechanism that combines apoptotic and autophagic components [67]. Another study showed that Asp-49 PLA₂ from the venom of *B. jararacussu* has anti-tumor and antimetastatic effects on the same cell line suggesting that this protein may be used for drug development for the treatment of breast cancer [68]. Additionally, CC-PLA2-1 and CC-PLA2-2, isolated from the venom of *Cerastes cerastes* (Horned desert viper), have been shown to effectively inhibit tumor cell adhesion and migration by interfering with $\alpha 5\beta 1$ and αv integrins, resulting in angiogenesis blocking [69].

Studies have been conducted to investigate the use of PLA_2 enzymes derived from the venom of *N. nigricollis* in treating diabetes. Result suggests hypoglycemic effects, lowering blood sugar levels in diabetes patients [70]. Furthermore, PLA_2 from *N. nigricollis* venom (in two isoforms) increased insulin secretion in BRIN-BD11 cells [71–74]. This



Fig. 2. Relative protein distribution in three snake genera and subfamilies: (A) Distribution of dominant protein families in Crotalinae, Viperinae, and Elapidae subfamilies; (B-D) Comparative analysis of the predominant proteins in the genera Bothrops, Crotalus, and Naja from the Viperidae and Elapidae families. Proteins analyzed are 3FTx, SVMP, SVSP, PLA₂, LAAO (L-amino acid oxidases), NP (natriuretic peptides), KUN, DIS, CTL/SNACLEC (C-type lectins and CTL-like proteins), MVC (minor venom components), and CRiSP (cysteine-rich secretory proteins).

study suggests that PLA_2 may be effective as diabetes treatments, especially for type 2 diabetes mellitus (T2DM) [73].

Acute Ischemic Stroke, which makes up nearly 90 % of all strokes and is a leading cause of death and disability globally, can be averted by inhibiting platelet aggregation to prevent blood clot formation. *N. nigricollis* venom contains PLA₂ that can impede blood clotting by inhibiting the prothrombinase complex and the extrinsic tenase complex (TF-FVIIa). BJ-PLA₂, a subtype of PLA₂ from *B. jararaca*, can block platelet aggregation, thus reducing the risk of acute thrombotic events. This suggests a potential for PLA₂ in preventing and treating CVDs.

Crotoxin is a pre-synaptic β -neurotoxin isolated from the venom of *C. d. terrificus,* known as the South American rattlesnake [75]. Crotoxin is comprised of two main components: enzymatic basic PLA₂ and crotapotin, which is a non-enzymatic acidic counterpart of crotoxin PLA₂ [76]. Crotoxin regulates pain and disease progression in an experimental autoimmune encephalomyelitis (EAE) model induced by myelin oligodendrocyte glycoprotein (MOG), a minor component of CNS myelin [77]. Crotoxin has been shown to alleviate pain in patients with advanced cancer [78–80] and has been proposed as a potential treatment for different pain conditions, including chronic neuropathic pain and cancer pain [81].

2.3. Three Finger toxins (3FTx)

Three Finger Toxins are non-enzymatic proteins with 60-75 amino

acid residues and possess a molecular weight ranging from 6 to 8 kDa [82]. Structurally, 3FTx exhibit a unique fold characterized by three β -stranded loops extending from a central core, which is stabilized by 4–5 conserved disulfide bridges. This three-fingered arrangement grants the toxins their name and confers specificity in interacting with various molecular targets [83]. Found most abundantly in the Elapidae family, they are known for their neurotoxic effects caused by the postsynaptic binding of 3FTx to acetylcholine receptors (AChRs) at neuromuscular junctions [83,84].

3FTx work by primarily modulating the activity of ion channels, receptors, and enzymes within the target organism. Nicotinic acetylcholine receptors (nAChRs), muscarinic acetylcholine receptors (mAChRs), and L-type calcium channels are prominent targets. By binding to these targets, 3FTx can lead to paralysis, impaired neurotransmission, or other physiological disruptions, contributing to the venom's toxicity [85].

3FTx, isolated from the venom of *N. kaouthia*, known as the Indian cobra, was found to inhibit platelet aggregation by inhibiting ADP, thrombin, and arachidonic acid, all of which induce platelet aggregation [86]. Those isolated from the venom of *Hemachatus haemachatus*, known as the ring-necked spitting cobra, form the hemextin AB complex, which inhibits clot initiation and factor VIIa activity [87]. 3FTx' potential antiplatelet and anti-coagulant activities make them a promising target for the development of future cardiovascular drugs.

Cobrotoxin, a subtype of 3FTx, is a short-chain post-synaptic

 α -neurotoxin that exhibits a high affinity for nAChRs at neuromuscular junctions [88]. This neurotoxin is primarily isolated from the venom of *N.n. atra*. The binding of cobrotoxin to nAChRs has been postulated to increase acetylcholine levels in the body by inhibiting acetylcholinesterase, an enzyme responsible for the hydrolysis of acetylcholine. As a result, pain signals may be inhibited, leading to a reduction in pain perception [89]. One such example is Keluoqu, a drug formulation containing cobrotoxin, which has been used for the treatment of chronic cancer pain. Clinical investigations have revealed that Keluoqu may be effective in alleviating moderate to severe cancer-related pain, thereby improving the quality of life for patients [89].

Cobratoxin, a subtype of 3FTx, is a long-chain post-synaptic α -neurotoxin. It was isolated from the venom of *N. n. kaouthia*, known as the Thailand cobra. Cobratoxin works by inhibiting the action of a protein called synaptobrevin, which releases neurotransmitters in the nervous system. This inhibition can lead to the suppression of pain signals. Thus, Cobratoxin has been shown to have analgesic effect, and its use as a painkiller is being investigated [90].

2.4. Disintegrins (DIS)

Disintegrins are a family of small non-enzymatic, cysteine-rich, and RGD (Arg-Gly-Asp)-containing proteins found mostly in the venom of Viperidae family. They are primarily known for their ability to inhibit integrins, which are cell adhesion receptors involved in various biological processes such as cell migration, proliferation, and survival [91].

Disintegrins cause hemorrhage by inhibiting platelet aggregation and induce apoptosis and necrosis in tissues they come in contact with [92]. They have been leveraged in developing thrombolytic drugs to prevent blood clots [27]. The FDA-approved anti-platelet drug Tirofiban (Aggrastat®) was developed from the DIS echistatin found in the venom of the viper *E. carinatus* [27]. Eptifibatide, derived from the DIS of viper *S. m. barbouri*, is also an anti-platelet drug (Integrillin®) [22]. Eptifibatide and Tirofiban, derived from snake venoms, act by blocking the binding of fibrinogen and von Willebrand factor to glycoprotein IIb/IIIa receptor on the platelet surface, thus inhibiting platelet aggregation [20].

Integrins are transmembrane receptors that have been shown to play a significant role in cancer progression and metastasis [93]. Integrins are divided into two subunits, α and β [94]. Many DIS have been shown to bind to numerous integrins, such as $\alpha 2\beta 1$, which are expressed by a wide variety of cells, namely those involved in tumor development [91]. An RGD-DIS isolated from the *Porthidium lansbergii*, also known as Lansberg's hognosed pitviper, inhibited the adhesion and migration of MCF7 and MDA-DB 231 breast cancer cells by binding to integrins $\alpha 2$ and/or $\beta 1$ [95]. Moreover, the RGD-DIS DisBa-01 isolated from the *B. alternatus* binds to $\alpha\nu\beta3$ integrin, which could prevent cell migration and adhesion in breast tumor cell 4T1BM2 [96,97].

Disintegrins have shown great promise in various diagnostic and therapeutic applications, including stroke treatment. Rhodostomin, a DIS purified from the *Calloselasma rhodostoma* venom and known as the Malayan Pit Viper, could aid in the treatment of arterial ischemic stroke [98]. Many studies strongly suggest that DIS, which act as α IIb β 3 antagonists, have anti-thrombotic therapy potential [99–101]. Nevertheless, the use of α IIb β 3 antagonists for stroke treatment is still being investigated [100].

2.5. Snake venom serine proteases (SVSP)

Snake Venom Serine Proteases are a class of proteolytic enzymes, possess a molecular weight ranging between 20 and 35 kDa and are comprised of approximately 180 to 240 amino acids, including a highly conserved catalytic triad of histidine, aspartate, and serine residues [102,103]. Structurally, SVSP adopt a globular fold similar to other serine proteases, featuring two β -barrel domains connected by a stretch of polypeptide chain, with the catalytic triad positioned at the interface

of these two domains [104].

Snake Venom Serine Proteases work by hydrolyzing specific peptide bonds in target proteins, such as those involved in blood coagulation, platelet aggregation, and fibrinolysis. The serine residue in the catalytic triad acts as a nucleophile, attacking the peptide bond's carbonyl carbon to form a tetrahedral intermediate, which collapses and releases the cleaved peptide fragments [105].

Batroxobin, a SVSP derived from the venom of *B. atrox*, is a singlechain glycoprotein composed of approximately 255 amino acids. It exhibits a conserved fold characteristic of serine proteases, with two β -barrel domains connected by a single peptide chain [106]. The active site of batroxobin contains a catalytic triad comprising histidine, aspartate, and serine residues, which confer its proteolytic activity. Batroxobin exerts its action by selectively cleaving the $\mbox{A}\alpha$ chain of fibrinogen, a key blood clotting protein, at Arg16-Gly17 peptide bond. This cleavage event results in the generation of fibrin monomers, which then spontaneously polymerize to form a fibrin clot, ultimately promoting hemostasis [2,37,102]. Studies have shown that batroxobin could decrease stroke recurrence rates and substantially improve neurological analyses [107,108]. A study using 1 h of Transcranial Doppler (TCD) Monitoring in patients with acute stroke found that batroxobin treatment can aid in neurological function recovery and reduce the risk of advancing stroke [107]. Batroxobin has demonstrated excellent results in stroke treatment when used alone or in combination with other molecules [109,110].

The potential applications of SVSP beyond hemostasis, considering intrinsic pathways involved in the wide range of biological activities of these molecules, may benefit from expanding their beneficial applications. It has been reported, a SVSP called collinein-1, derived from *C. durissus collilineatus* venom, blocking ion channels activity. SVSP toxin blocks hEAG1, a channel that plays a role in proliferation, migration, and apoptosis and contributes to cancer progression. Research into new specific inhibitors of hEAG1 channels could lead to controlling cell proliferation. Additionally, these drugs can be used in conjunction with chemotherapy to increase the patient's survival rate [102,111,112]. Another example is thrombin-like toxin called gyroxin toxin is found in the *C. d. terrificus* venom. This toxin displays hemostatic effects but also causes the "Gyroxin syndrome", a series of aberrant motor behavior in mice, suggesting SVSP can affect the blood brain barrier [113].

Ancrod, another enzymatic protein purified from *C. rhodostoma*, has previously shown great promise in treating strokes [114–116]. However, studies found that Ancrod did not improve outcomes for stroke patients compared to a placebo [117,118]. Ancrod's mechanism of action is unclear, and studies have cast doubts on its efficacy as a treatment for stroke [119].

2.6. Snake venom metalloproteinases (SVMP)

Snake Venom Metalloproteinases are Zn^{2+} -dependent enzymes found in high abundance in the Viperidae snake family. The molecular sizes of these enzymes range from 20 to 100 kDa, with amino acid sequences ranging from 200 to 600 residues [120,121]. Based on their domain composition, SVMP can be classified into three main classes (P-I, P-II, and P-III), with P-III being the most complex, containing multiple additional domains. SVMP' mechanism of action is generally characterized by the hydrolysis of extracellular matrix components such as collagen and laminin, resulting in local tissue damage and hemorrhage [96,122].

P-I SVMP class is the simplest, comprising just the metalloproteinase domain. This classification exhibits the smallest molecular sizes within the range, typically clustering around 20 kDa. Despite their relative simplicity, they can exert significant local tissue damage due to their hydrolytic action on extracellular matrix components, particularly on collagen and laminin. Their damaging activity is mostly localized, contributing predominantly to hemorrhagic outcomes [123].

P-II SVMP contain both a metalloproteinase domain and a DIS domain. These dual-domain proteins have a slightly larger molecular size compared to P-I, generally ranging from 30 to 60 kDa. The inclusion of a DIS domain augments their biological activity; the DIS domain is known for its interaction with integrin receptors, playing a significant role in inhibiting platelet aggregation. Hence, P-II SVMP not only induce local tissue damage and hemorrhage but also modulate blood coagulation, leading to an anti-coagulant effect [124].

The most complex class is P-III SVMP, which contain the metalloproteinase domain, the DIS-like domain, and a cysteine-rich domain. With this broad array of functional domains, their molecular size is correspondingly larger, usually spanning between 60 and 100 kDa. The presence of multiple additional domains endows P-III SVMP with a wide variety of biological activities, encompassing not only the functions of P-I and P-II classes but also additional effects. These may include activation of complement, inducing inflammation, and interfering with the immune response. Consequently, P-III SVMP can provoke systemic responses, resulting in more severe and multifaceted clinical manifestations upon envenomation [125].

Jararhagin, a P-III SVMP isolated from *B. jararaca*, has been shown in an in vivo treatment to reduce the incidence of nodules, metastasis, and antiproliferative inhibition capacity, and has been proposed as a potential anti-neoplastic drug [126,127].

2.7. L-amino acid oxidases (LAAO)

L-amino acid oxidases are a class of flavoproteins found in various sources, including snake venoms, and have garnered significant interest due to their multifaceted pharmacological properties. Structurally, LAAO are homodimeric proteins with a molecular weight of about 60–70 kDa per monomer and a non-covalently bound flavin adenine dinucleotide (FAD) as a prosthetic group [128]. LAAO have two domains that work together to facilitate the enzyme's catalytic activity: a FAD-binding domain and a substrate-binding domain.

L-amino acid oxidases mechanism of action involves the stereospecific oxidation of L-amino acids, converting them into their corresponding α -keto acids, along with the concomitant production of hydrogen peroxide (H₂O₂) and ammonia [129]. This enzymatic activity has been implicated in cytotoxicity, apoptosis induction, edema formation, and anti-parasitic, anti-microbial, and anti-viral activities [130–132].

The cytotoxic effects of LAAO are primarily attributed to the generation of H_2O_2 , which can cause oxidative stress, leading to cellular damage and apoptosis. Furthermore, LAAO have been reported to exhibit selective cytotoxicity towards certain cancer cells, making them potential candidates for the development of novel anti-cancer therapies [133].

The LAAO isolated from the South American rattlesnake, *Crotalus durissus terrificus*, showed anti-tumor activity in several cancer cell lines [134]. Moreover, the LAAO extracted from *Ophiophagus hannah*, known as the king cobra, has been shown to have potent anti-proliferative properties against both human breast and lung cancer cells [135].

2.8. Other promising snake venom proteins and peptides

Hannalgesin is a long neurotoxin, analgesic compound derived from the venom of *O. hannah*. It has demonstrated substantial pain-relieving capabilities in preclinical studies. The mechanism of action involves an interruption in the activity of nitric oxide synthase in neurons, leading to a reduction in nitric oxide production, a known player in pain perception [136]. It also appears to interact with both the opioid and nitric oxide systems in its pain-alleviating action, suggesting a complex mode of activity [137]. Hannalgesin, exhibits analgesic activity in mice being 2,700 times more effective on a molar basis than morphine [137].

Kunitz peptides are small proteins comprised of roughly 60 amino acid residues organized in a unique $\alpha + \beta$ domain architecture, featuring

two β-strands and two short α-helices [138]. This domain is rich in cysteines and is stabilized by three conserved disulfide bridges. The molecular weight of these peptides is around 6 kDa [139]. KUN have been implicated in blood clotting, fibrinolysis, response to injury or infection, and the regulation of ion channels [140]. They are primarily known as serine protease inhibitors [141].

Cysteine-rich secretory proteins are a group of non-enzymatic proteins characterized by a unique molecular structure: an N-terminal pathogenesis-related (PR-1) domain and a C-terminal cysteine-rich domain (CRD), linked by a hinge region [142]. This structure, having a molecular weight ranging between 20 and 30 kDa, is highly conserved across various snake species [143]. The mechanism of action of CRISP primarily involves blocking ion channels, particularly those associated with smooth muscle contraction and glandular secretion [144]. This leads to a variety of physiological effects, including paralysis, hypotension, and inhibition of platelet aggregation.

Natriuretic peptides (NP) are non-enzymatic peptides with a length of 25–30 amino acids and a size of 2–3 kDa, with their structures stabilized by intra-chain disulfide bonds [145]. These peptides primarily interact with natriuretic peptide receptors (NPR), including NPR-A and NPR-B, found on the surfaces of cells in various tissues. When these receptors bind, they initiate intracellular signaling cascades involving cyclic guanosine monophosphate (cGMP) [146]. This activation process then results in a series of physiological changes, including vasodilation.

Snake venom C-type lectin-like proteins (SNACLEC), are a group of non-enzymatic proteins found in snake venoms. These proteins typically consist of 120–140 amino acids and exhibit a common carbohydraterecognition domain (CRD) of approximately 130 residues folded into a characteristic "double-loop" structure stabilized by two disulfide bridges [147]. They function primarily by interacting with specific receptors on the cell surface, disrupting hemostasis and immune responses, often leading to systemic effects like edema, hemorrhage, and necrosis [148].

3. Future perspectives on deep learning application in snake venom for drug discovery

Recent advancements in venomics have been made using Artificial Intelligence (AI). The AI Haemorrhage Analysis (AHA) tool, designed for quantifying venom-induced hemorrhage in mice, especially in snakebite scenarios, exemplifies this progress [149]. AHA can automatically identify hemorrhagic lesions using Machine Leaning (ML) methods, adjust for lighting variations, and compute Hemorrhagic Units (HaU). Its main benefits include a significant reduction in analysis time, a web interface for global laboratory use, and improved antivenom efficacy assessment. AHA stands out for its reliability, accessibility, and reproducibility, marking a notable contribution to toxinology research.

Another innovative tool is the Venom Induced Dermonecrosis Analysis tool (VIDAL) [150]. Using ML, VIDAL assesses dermonecrosis in snakebite envenomations through in vivo mouse models. It automates lesion detection, corrects for lighting inconsistencies, and measures severity with a unique Dermonecrotic Unit (DnU). VIDAL's accuracy and reproducibility are on par with histopathological analysis, making it valuable in developing treatments for this neglected tropical disease.

Furthermore, a recent systematic review focusing on AI in snakebite identification analyzed 26 studies [151]. The findings indicate that machine learning and deep learning algorithms have achieved 72–98 % accuracy in snake image classification, 80-100 % in wound image classification, and 71-97 % in other related areas. This highlights the ongoing need for better data quality and decision support systems to enhance snakebite treatment.

Recently, major developments have been realized in DL for molecular discovery (Fig. 3). Molecular contrastive learning (MolCLR) has shown significant improvements in performance on molecular property benchmarks according to recent studies [152,153]. A recent study introduces MolCLR, a self-supervised framework using Graph Neural Networks (GNNs) for molecular property prediction, overcoming



Fig. 3. Potential DL applications in venomics. DL models have been successfully applied to many aspects of drugs and molecular discovery. Several types of DL models could be applied to venomics. RNN and transformers can identify long and short-ranging patterns and molecular sequences. Diffusion models are generative networks that can generate molecular structures. MCL is a special type of training that learns to identify differences between molecules.

challenges of limited labeled data in drug discovery [154]. Using around 10 million unique unlabeled molecules, MolCLR uses innovative graph augmentations (atom masking, bond deletion, and subgraph removal) and contrastive learning, significantly boosting GNN performance on various benchmarks, achieving top results post fine-tuning, and effectively discerning molecular similarities.

This technology has potential applications in areas like snake venombased drug discovery where it could be instrumental in developing drugs that target and inhibit snake venom toxins' receptors. The method itself involves building molecule graphs and using augmentations and a contrastive estimator. The aim is to increase the similarity between augmentations from the same molecule while decreasing the similarity between different molecules. It operates on a large set of unlabeled molecular data, aiming to boost the performance of graph neural network encoders. The underlying principle of this method is to train the model to efficiently identify and differentiate between various molecules, capitalizing on the concept of self-supervised learning where the data itself provides supervision.

Transformer-based models, like AlphaFold2, are DL models that use self-attention to process sequential data, such as protein sequences [155]. Self-attention, also referred to as the attention mechanism, enables the model to weigh and prioritize different parts of the input sequence differently. This allows it to focus more on relevant parts and less on irrelevant ones, thereby capturing long-range dependencies and intricate patterns within the data [156]. AlphaFold2 is a protein structure prediction system that uses a deep neural network based on transformer architecture to predict the 3D structure of a protein from its amino acid sequence [157]. Models such as Alphafold2 can be applied to predict the 3D structure of snake venom proteins. Such applications could, for example, help accelerate the design of new antivenoms inhibiting the effects of snake venom toxins, for instance, singlemolecule antibodies structurally designed to bind to the SVMP or SVSP catalytic sites. Applications in 3D modeling would also facilitate the development of toxins as therapeutics, able to mimic the desired activity of the toxins without the associated toxicity or to mitigate "offtarget" effects. A recent study evaluated AlphaFold2 and ColabFold's performance on more than a thousand snake venom toxins [157-159]. The results indicated that while caution must be exercised with proteins that have limited reference data, these tools significantly contribute to understanding protein functions and their potential applications. This implies that despite certain limitations, such models are proving to be valuable in toxinology and related fields, offering new perspectives on protein structure and function.

A recent study proposed a DL method that uses data augmentation to predict novel spider neurotoxic peptides [160]. The method combines a generative adversarial network (GAN) with Convolutional Neural Networks (CNN) to predict the neurotoxicity of virtual peptides generated by the GAN. Data augmentation methods were applied to the training dataset of known neurotoxic spider peptides to increase the data's diversity and improve the CNN's performance. The resulting model was able to accurately predict the neurotoxicity of virtual peptides generated by the GAN, demonstrating the potential of this method for discovering novel spider neurotoxic peptides [160]. (Perpetuo, L et al.) discusses the use of Artificial Intelligence (AI) in peptidomics for developing therapeutic peptides, crucial biomarkers and treatment agents for various diseases [161]. It focuses on data-driven AI methods like support vector machines, random forests, and deep learning for efficient peptide-based drug discovery. It also emphasizes AI's role in advancing peptidomics and selective peptide therapies, highlighting its importance in the prediction and development of successful peptide-based drugs.

Generative models are a class of Machine Learning (ML) algorithms aiming to learn the true data distribution of the training set to generate new data points with some variations. They have found notable applications in the field of molecular discovery [162,163]. Variational Autoencoders (VAEs) are a type of generative model used for unsupervised learning tasks. It combines an autoencoder, a neural network trained to reconstruct its input, and Bayesian variational approximation, a method for approximating complex probability distributions. A recent study proposed a DL framework that uses CNNs, VAEs, and attention mechanisms to predict drug-protein interactions using VAEs [164,165]. VAEs can be used in a variety of ways for venom-based drug discovery. For example, generative models can be trained on a known compound dataset and then used to generate new, chemically valid compounds with similar properties to the training set. This could be used to generate new leads for snake venom-based drug discovery. VAEs can also be trained on a dataset of known protein-ligand complexes and then used to predict the binding affinity of new compounds for a given protein target [166]. This can be used for the virtual screening of large venomics

datasets of compounds to identify potential hits for further development.

Another type of DL that could hold potential in the exploration and interpretation of snake venom compounds is Recurrent Neural Networks (RNNs). RNNs are comparable to transformer-based models in their ability to identify and leverage extensive, long-range patterns within biological sequences. Specifically, Bidirectional Long Short-Term Memory (BiLSTM) networks, a distinct subclass of RNNs, have demonstrated versatility across a range of applications, with drug discovery being a notable example [166-169]. "DeepLPI" is a novel DL-based model for predicting protein-ligand interactions, which can be used in drug repurposing [170]. The model uses a BiLSTM network to analyze the molecular structures of proteins and ligands to predict their interactions. The model was trained and tested on a large dataset and was found to have high accuracy and good performance compared to other existing methods [170]. These results suggest that BiLSTM models could also be applied to snake venom protein-ligand binding affinity prediction.

4. Conclusions

Snake venoms' multivalent nature, with hemotoxic, cytotoxic, and neurotoxic effects, makes it a potential source for drug discovery and development. The therapeutic potential of these venoms is fundamentally linked to their composition, the specific snake species, and the interplay of toxins present within the venom.

The integration of AI methods in venomics research has the potential to revolutionize the field of venom-based drug discovery. As venomics expands, using AI can aid in discovering new potential drugs and exploring venom evolution across various venomous species like snakes, spiders, scorpions, bees, and cone snails. This can also forecast the emergence of new venom components.

The use of antivenoms as a therapeutic intervention for venomous bites has been widely acknowledged for its efficacy in neutralizing the toxic effects of venom [171,172]. However, using antivenoms, if available due to global shortages, has serious limitations, such as efficacy and impurities can cause anaphylactic shock or serum sickness and can be costly [173]. Interestingly, toxicity prediction using ML and DL models can advance next-generation antivenoms by supporting the design of single antibodies, bioengineered antibodies, and structure-guided antivenoms against toxins. DL models can predict toxicity scores and lethal dose 50 s (LD50) by analyzing massive amounts of venomics data, which can help develop more effective and specific antivenoms. DL models can also be used to analyze blood samples from snakebite-envenomed patients to identify biomarkers indicative of the presence of specific toxins and thus allow for accurate antivenom selection and dosage needed for toxin neutralization.

Finally, 130 species represent only approximately 15 % of all venomous snake species. More efforts are needed to characterize snake venom composition across species and within the same species. To be effective these data-gathering studies should encompass different levels of biological information, including transcriptomics, proteomics, and metabolomics [174].

CRediT authorship contribution statement

Anas Bedraoui: Writing – review & editing, Writing – original draft, Visualization, Validation, Software, Methodology, Investigation, Formal analysis, Conceptualization. Montamas Suntravat: Writing – review & editing, Data curation. Salim El Mejjad: Visualization, Software, Formal analysis. Salwa Enezari: Visualization, Software, Formal analysis. Naoual Oukkache: Writing – review & editing. Elda E. Sanchez: Writing – review & editing, Validation, Resources, Data curation. Jacob A. Galan: Writing – review & editing, Visualization, Validation, Supervision, Methodology, Investigation, Formal analysis, Data curation, Conceptualization. Rachid El Fatimy: Writing – review & editing, Visualization, Validation, Supervision, Software, Resources, Project administration, Methodology, Investigation, Funding acquisition, Conceptualization. **Tariq Daouda:** Writing – review & editing, Visualization, Validation, Supervision, Software, Resources, Project administration, Methodology, Investigation, Formal analysis, Conceptualization.

Declaration of competing interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

References

- Tasoulis T, Isbister GK. A review and database of snake venom proteomes. Toxins 2017;9:290. https://doi.org/10.3390/toxins9090290.
- [2] Oliveira AL, Viegas MF, da Silva SL, Soares AM, Ramos MJ, Fernandes PA. The chemistry of snake venom and its medicinal potential. Nat Rev Chem 2022;6: 451–69. https://doi.org/10.1038/s41570-022-00393-7.
- [3] Kang TS, Georgieva D, Genov N, Murakami MT, Sinha M, Kumar RP, et al. Enzymatic toxins from snake venom: structural characterization and mechanism of catalysis. FEBS J 2011;278:4544–76. https://doi.org/10.1111/j.1742-4658.2011.08115.x.
- [4] Guerra-Duarte C, Lopes-Peixoto J, Fonseca-de-Souza BR, Stransky S, Oliveira D, Schneider FS, et al. Partial in vitro analysis of toxic and antigenic activities of eleven peruvian pitviper snake venoms. Toxicon 2015;108:84–96. https://doi. org/10.1016/j.toxicon.2015.09.007.
- [5] Bocian A, Hus KK. Antibacterial properties of snake venom components. Chem Pap 2020;74:407–19. https://doi.org/10.1007/s11696-019-00939-y.
- [6] Marte F, Sankar P, Cassagnol M. Captopril. Treasure Island (FL): In StatPearls; StatPearls Publishing; 2022.
- [7] D'Angelo A, Giannini S, Benetollo P, Castrignano R, Lodetti MG, Malvasi L, et al. Efficacy of captopril in hypertensive diabetic patients. Am J Med 1988;84:155–8. https://doi.org/10.1016/0002-9343(88)90225-2.
- [8] Bordon K. de C.F., Cologna CT, Fornari-Baldo EC, Pinheiro-Júnior EL, Cerni FA, Amorim FG, et al. From animal poisons and venoms to medicines: achievements, challenges and perspectives in drug discovery. Front Pharmacol 2020; 11.
- [9] Hartman GD, Egbertson MS, Halczenko W, Laswell WL, Duggan ME, Smith RL, et al. Non-peptide fibrinogen receptor antagonists. 1. Discovery and design of exosite inhibitors. J Med Chem 1992;35:4640–2. https://doi.org/10.1021/ im00102a020.
- [10] Scarborough RM, Rose JW, Hsu MA, Phillips DR, Fried VA, Campbell AM, et al. A GPIIb-IIIa-specific integrin antagonist from the venom of sistrurus m. Barbouri J Biol Chem 1991;266:9359–62.
- [11] Patchett A. The chemistry of enalapril. Br J Clin Pharmacol 1984;18:2018–78. https://doi.org/10.1111/j.1365-2125.1984.tb02599.x.
- [12] Shen H. Deadly snake venom delivers pain relief. Nature 2012. https://doi.org/ 10.1038/nature.2012.11526.
- [13] The UniProt Consortium UniProt. The Universal Protein Knowledgebase in 2021. Nucl Acids Res 2021;49:D480–9. https://doi.org/10.1093/nar/gkaa1100.
- [14] Bradykinin-Potentiating Peptide 11 Bothrops Jararaca (Jararaca) | UniProtKB | UniProt Available online: https://www.uniprot.org/uniprotkb/P0C7J8/entry (accessed on 18 November 2022).
- [15] Ianzer D, Konno K, Marques-Porto R, Vieira Portaro FC, Stöcklin R, Martins de Camargo AC, et al. Identification of Five New Bradykinin Potentiating Peptides (BPPs) from Bothrops Jararaca Crude venom by using electrospray ionization tandem mass spectrometry after a two-step liquid chromatography. Peptides 2004;25:1085–92. https://doi.org/10.1016/j.peptides.2004.04.006.
- [16] Faisal M, Cawello W, Laeer S. Clinical pharmacokinetics of enalapril and enalaprilat in pediatric patients—a systematic review. Front Pediatr 2021;9: 611322. https://doi.org/10.3389/fped.2021.611322.
- [17] EMA Renitec Available online: https://www.ema.europa.eu/en/medicines/ human/referrals/renitec (accessed on 11 December 2022).
- [18] Odaka C, Mizuochi T. Angiotensin-converting enzyme inhibitor captopril prevents activation-induced apoptosis by interfering with T cell activation signals. Clin Exp Immunol 2000;121:515–22. https://doi.org/10.1046/j.1365-2249.2000.01323.x.
- [19] Rivas-Mercado EA, Garza-Ocañas L. Disintegrins obtained from snake venom and their pharmacological potential. Medicina Universitaria 2017;19:32–7. https:// doi.org/10.1016/j.rmu.2017.02.004.
- [20] Bansal AB, Sattar Y, Jamil RTE. StatPearls Publishing 2022.[21] Shah I, Khan SO, Malhotra S, Fischell T. Eptifibatide: the evidence for its role in
- the management of acute coronary syndromes. Core Evid 2009;4:49–65.
 [22] Drug Approval Package: Integrilin (Eptifibatide) NDA# 20-718 Available online: https://www.accessdata.fda.gov/drugsatfda_docs/nda/98/20718_Integrilin.cfm (accessed on 30 October 2022).
- [23] EMA Integrilin Available online: https://www.ema.europa.eu/en/medicines/ human/EPAR/integrilin (accessed on 11 December 2022).
- [24] Gan ZR, Gould RJ, Jacobs JW, Friedman PA, Polokoff MAE. A potent platelet aggregation inhibitor from the venom of the viper, Echis Carinatus. J Biol Chem 1988;263:19827–32. https://doi.org/10.1016/S0021-9258(19)77710-2.

- [25] Valgimigli M, Biondi-Zoccai G, Tebaldi M, van 't Hof AWJ, Campo G, et al. Tirofiban as adjunctive therapy for acute coronary syndromes and percutaneous coronary intervention: a meta-analysis of randomized trials. Eur Heart J 2010; 31, 35-49, doi:10.1093/eurheartj/ehp376.
- [26] Guo Y, Zhao Z, Li S, Chen L. Clinical efficacy and safety of tirofiban combined with conventional dual antiplatelet therapy in ACS Patients Undergoing PCI. Sci Rep 2021;11:17144. https://doi.org/10.1038/s41598-021-96606
- Lazarovici P, Marcinkiewicz C, Lelkes PI. From snake venom's disintegrins and C-[27] Type lectins to anti-platelet drugs. Toxins 2019;11:303. https://doi.org/10.3390/ toxins11050303.
- [28] Hayashi MAF, Camargo ACM. The Bradykinin-potentiating peptides from venom gland and brain of bothrops jararaca contain highly site specific inhibitors of the somatic angiotensin-converting enzyme. Toxicon 2005;45:1163-70. https://doi. org/10.1016/j.toxicon.2005.02.017.
- [29] Drug Approval Package: Aggrastat (Tirofiban Hydrochloride) NDA# 20912/S1 & 20913/S1 Available online: https://www.accessdata.fda.gov/drugsatfda_docs/ nda/99/20912S001_Aggrastat.cfm (accessed on 30 October 2022)
- [30] EMA Aggrastat Available online: https://www.ema.europa.eu/en/medicines/ human/referrals/aggrastat (accessed on 11 December 2022).
- [31] Hashemzadeh M, Furukawa M, Goldsberry S, Movahed MR. Chemical structures and mode of action of intravenous glycoprotein IIb/IIIa receptor blockers: a review. Exp Clin Cardiol 2008;13:192-7.
- [32] Itoh N, Tanaka N, Mihashi S, Yamashina I. Molecular cloning and sequence analysis of cDNA for batroxobin, a thrombin-like snake venom enzyme. J Biol Chem 1987;262:3132-5.
- [33] Guo Y, Zuo Y, Wang Q, Tang B, Li F, Sun Y. Meta-analysis of defibrase in treatment of acute cerebral infarction. Chin Med J (Engl) 2006;119:662-8.
- [34] Fischer T, Riedl R. Paracelsus' legacy in the faunal realm: drugs deriving from animal toxins. Drug Discov Today 2022;27:567-75. https://doi.org/10.1016/j. drudis.2021.10.003.
- Cooperative Group for Reassessment of Defibrase Reassessment of Defibrase in [35] Treatment of Acute Cerebral Infarction: A Multicenter, Randomized, Double-Blind, Placebo-Controlled Trial. Chin Med Sci J 2005, 20, 151-158.
- [36] Kubo T, Matsunaga T, Asai H, Kawamoto K, Kusakari J, Nomura Y, et al. Efficacy of defibrinogenation and steroid therapies on sudden deafness. Archives of Otolaryngology-Head & Neck Surgery 1988;114:649-52. https://doi.org/ 10.1001/archotol.1988.01860180063031.
- [37] Masuda H, Sato A, Shizuno T, Yokoyama K, Suzuki Y, Tokunaga M, et al. Batroxobin accelerated tissue repair via neutrophil extracellular trap regulation and defibrinogenation in a murine ischemic hindlimb model. PLoS One 2019;14: e0220898
- [38] Takacs Z. Nathan S. Animal venoms in medicine. In Encyclopedia of Toxicology (Third Edition); Wexler, P., Ed.; Academic Press: Oxford, 2014; pp. 252-259 ISBN 978-0-12-386455-0.
- Amorim FG, Menaldo DL, Carone SEI, Silva TA, Sartim MA, De Pauw E, et al. New [39] Insights on Moojase, a Thrombin-Like Serine Protease from Bothrops Moojeni Snake Venom, Toxins (Basel) 2018:10:500, https://doi.org/10.3390/ toxins10120500
- [40] Moura-da-Silva AM, Contreras-Bernal JC, Gimenes SNC, Freitas-de-Sousa LA, Portes-Junior JA, da Peixoto P, et al. The relationship between clinics and the venom of the Causative Amazon Pit Viper (Bothrops Atrox). PLoS Negl Trop Dis 2020:14:e0008299.
- [41] Reptilase® Haemocoagulase Pentapharm Available online: https://www. pentapharm.com/markets-and-products/products/pharma/haemocoagulase/ (accessed on 11 December 2022).
- [42] Roly ZY, Islam MM, Reza MA. A comparative in silico characterization of functional and physicochemical properties of 3FTx (Three Finger Toxin) Proteins from Four Venomous Snakes, Bioinformation 2014:10:281-7, https://doi.org/ 0.6026/97320630010281.
- [43] Liu B, Wang W, Gao T, Huang L, Fan H, Chen H-X. Separation, Identification and Quantification of Associated Impurities in Cobratide Using Sheathless CE-MS and CE-UV. Anal Methods 2021;13:3845-51. https://doi.org/10.1039/D1AY0071
- [44] Lin F, Reid PF, Qin Z. Cobrotoxin Could Be an Effective Therapeutic for COVID-19. Acta Pharmacol Sin 2020;41:1258-60. https://doi.org/10.1038/s41401-020-00501-7
- [45] Uetz P, Koo M, Aguilar R, Brings E, Catenazzi A, Chang A, et al. A Quarter Century of Reptile and Amphibian Databases. Herpetological Review 2021;52:246-55.
- [46] Roddy, M.; Freishtat, R.J. Snake Bites. In Pediatric Clinical Advisor (Second Edition); Garfunkel, L.C., Kaczorowski, J.M., Christy, C., Eds.; Mosby: Philadelphia, 2007; p. 533 ISBN 978-0-323-03506-4.
- [47] Bites, Snake - ClinicalKey Available online: https://www.clinicalkey.com/ #!/content/book/3-s2.0-B9780323755702001260 (accessed on 29 October 2022)
- [48] Gopalakrishnan M, Vinod KV, Dutta TK, Shaha KK, Sridhar MG, Saurabh S. Exploring Circulatory Shock and Mortality in Viper Envenomation: A Prospective Observational Study from India. QJM: An International Journal of Medicine 2018;111:799-806. https://doi.org/10.1093/qjmed/hcy175.
- Ratanabanangkoon K. A Quest for a Universal Plasma-Derived Antivenom Against [49] All Elapid Neurotoxic Snake Venoms. Front Immunol 2021;12.
- [50] Ahdritz, G.; Bouatta, N.; Kadyan, S.; Xia, Q.; Gerecke, W.; O'Donnell, T.J.; Berenberg, D.; Fisk, I.; Zanichelli, N.; Zhang, B.; et al. OpenFold: Retraining AlphaFold2 Yields New Insights into Its Learning Mechanisms and Capacity for Generalization 2022, 2022.11.20.517210.
- Murayama N, Hayashi MAF, Ohi H, Ferreira LAF, Hermann VV, Saito H, et al. [51] Cloning and Sequence Analysis of a Bothrops Jararaca cDNA Encoding a Precursor of Seven Bradykinin-Potentiating Peptides and a C-Type Natriuretic

Peptide. Proc Natl Acad Sci 1997;94:1189-93. https://doi.org/10.1073/ pnas.94.4.1189

- [52] Cintra ACO, Vieira CA, Giglio JR. Primary Structure and Biological Activity of Bradykinin Potentiating Peptides fromBothrops Insularis Snake Venom. J Protein Chem 1990;9:221-7. https://doi.org/10.1007/BF01025312.
- [53] Sivieri DO, Bispo-da-Silva LB, Oliveira EB, Resende AC, Salgado MCO. Potentiation of Bradykinin Effect by Angiotensin-Converting Enzyme Inhibition Does Not Correlate with Angiotensin-Converting Enzyme Activity in the Rat Mesenteric Arteries. Hypertension 2007;50:110-5. https://doi.org/10.1161/ HYPERTENSIONAHA.106.085761.
- [54] Goyal A, Cusick AS, Inhibitors TBA. StatPearls Publishing 2022.
- [55] Péterfi O, Boda F, Szabó Z, Ferencz E, Bába L. Hypotensive Snake Venom Components-A Mini-Review. Molecules 2019;24:2778. https://doi.org/ 10.3390/molecules24152778.
- Sciani JM, Pimenta DC. The Modular Nature of Bradykinin-Potentiating Peptides Isolated from Snake Venoms. J Venomous Anim Toxins Incl Trop Dis 2017;23:45. ttps://doi.org/10.1186/s40409-017-0134-
- [57] Munawar A, Trusch M, Georgieva D, Hildebrand D, Kwiatkowski M, Behnken H, et al. Elapid Snake Venom Analyses Show the Specificity of the Peptide Composition at the Level of Genera Naja and Notechis. Toxins 2014;6:850-68. https://doi.org/10.3390/toxins6030850.
- [58] Wishart DS, Knox C, Guo AC, Shrivastava S, Hassanali M, Stothard P, et al. DrugBank: A Comprehensive Resource for in Silico Drug Discovery and Exploration. Nucleic Acids Res 2006;34:D668-72. https://doi.org/10.1093/nar/
- [59] Bryniarski P, Nazimek K, Marcinkiewicz J. Captopril Combined with Furosemide or Hydrochlorothiazide Affects Macrophage Functions in Mouse Contact Hypersensitivity Response. Int J Mol Sci 2021;23:74. https://doi.org/10.3390/ ijms23010074.
- [60] Frangieh J, Rima M, Fajloun Z, Henrion D, Sabatier J-M, Legros C, et al. Snake Venom Components: Tools and Cures to Target Cardiovascular Diseases. Molecules 2021;26:2223. https://doi.org/10.3390/molecules26082223.
- Lewis EJ, Hunsicker LG, Bain RP, Rohde RD. The Effect of Angiotensin-[61] Converting-Enzyme Inhibition on Diabetic Nephropathy. N Engl J Med 1993;329: 1456-62. https://doi.org/10.1056/NEJM199311113292004.
- Zatz R, Dunn BR, Meyer TW, Anderson S, Rennke HG, Brenner BM. Prevention of [62] Diabetic Glomerulopathy by Pharmacological Amelioration of Glomerular Capillary Hypertension. J Clin Invest 1986;77:1925-30. https://doi.org/ 10 1172/JCI112521
- [63] Marcussi S, Sant'Ana CD, Oliveira CZ, Rueda AQ, Menaldo DL, Beleboni RO, et al. Snake venom phospholipase a2 inhibitors: medicinal chemistry and therapeutic potential. Curr Top Med Chem 2007, 7, 743-756, doi:10.2174/ 156802607780487614.
- [64] Burke JE, Dennis EA. Phospholipase A2 Biochemistry. Cardiovasc Drugs Ther 2009;23:49–59. https://doi.org/10.1007/s10557-008-6132-9. Vardjan N, Mattiazzi M, Rowan EG, Križaj I, Petrovič U, Petan T. Neurotoxic
- [65] Phospholipase A2 Toxicity Model. Commun Integr Biol 2013;6:e23600.
- [66] Kallajoki M, Alanen KA, Nevalainen M, Nevalainen TJ. Group II Phospholipase A2 in Human Male Reproductive Organs and Genital Tumors. Prostate 1998;35: 263-72. https://doi.org/10.1002/(sici)1097-0045(19980601)35:4<263::aidpros5>3.0.co:2-h.
- [67] Azevedo FVPV, Lopes DS, Cirilo Gimenes SN, Achê DC, Vecchi L, Alves PT, et al. Human Breast Cancer Cell Death Induced by BnSP-6, a Lys-49 PLA2 Homologue from Bothrops Pauloensis Venom. Int J Biol Macromol 2016;82:671-7. https:// doi.org/10.1016/j.ijbiomac.2015.10.080.
- [68] de Vasconcelos Azevedo FVP, Zóia MAP, Lopes DS, Gimenes SN, Vecchi L, Alves PT, et al. Antitumor and Antimetastatic Effects of PLA2-BthTX-II from Bothrops Jararacussu Venom on Human Breast Cancer Cells. Int J Biol Macromol 2019;135:261-73. https://doi.org/10.1016/j.ijbiomac.2019.05.164.
- Zouari-Kessentini R, Luis J, Karray A, Kallech-Ziri O, Srairi-Abid N, Bazaa A, et al. [69] Two Purified and Characterized Phospholipases A2 from Cerastes Cerastes Venom, That Inhibit Cancerous Cell Adhesion and Migration. Toxicon 2009;53: 444-53. https://doi.org/10.1016/j.toxicon.2009.01.003.
- [70] Sarmiento BE, Santos Menezes LF, Schwartz EF. Insulin Release Mechanism Modulated by Toxins Isolated from Animal Venoms: From Basic Research to Drug Development Prospects. Molecules 2019;24:1846. https://doi.org/10.3390/ molecules24101846.
- [71] Yamamoto S, Nakaki T, Nakadate T, Kato R. Insulinotropic Effects of Exogenous Phospholipase A2 and C in Isolated Pancreatic Islets. Eur J Pharmacol 1982;86: 121-4. https://doi.org/10.1016/0014-2999(82)90409-5.
- A. Coulter-Parkhill S. McClean V.A. Gault N. Irwin Therapeutic Potential of [72] Peptides Derived from Animal Venoms: Current Views and Emerging Drugs for Diabetes Clin Med Insights Endocrinol Diabetes 14 2021 11795514211006071 10.1177/11795514211006071.
- [73] Conlon JM, Attoub S, Musale V, Leprince J, Casewell NR, Sanz L, et al. Isolation and Characterization of Cytotoxic and Insulin-Releasing Components from the Venom of the Black-Necked Spitting Cobra Naja Nigricollis (Elapidae). Toxicon: X 2020;6:100030. https://doi.org/10.1016/j.toxcx.2020.100030.
- [74] Nogueira TCA, Ferreira F, Toyama MH, Stoppiglia LF, Marangoni S, Boschero AC, et al. Characterization of the Insulinotropic Action of a Phospholipase A2 Isolated from Crotalus Durissus Collilineatus Rattlesnake Venom on Rat Pancreatic Islets. Toxicon 2005;45:243-8. https://doi.org/10.1016/j.toxicon.2004.10.017
- [75] Seki C, Vidal JC, Barrio A. Purification of Gyroxin from a South American Rattlesnake (Crotalus Durissus Terrificus) Venom. Toxicon 1980;18:235-47. https://doi.org/10.1016/0041-0101(80)90002-1.

- [76] Cecchini AL, Soares AM, Cecchini R, de Oliveira AHC, Ward RJ, Giglio JR, et al. Effect of Crotapotin on the Biological Activity of Asp49 and Lys49 Phospholipases A2 from Bothrops Snake Venoms. Comp Biochem Physiol C: Toxicol Pharmacol 2004;138:429–36. https://doi.org/10.1016/j.cca.2004.07.010.
- [77] Teixeira NB, Sant'Anna, M.B., Giardini, A.C., Araujo, L.P., Fonseca, L.A., Basso, A. S., Cury, Y., Picolo, G. Crotoxin Down-Modulates pro-Inflammatory Cells and Alleviates Pain on the MOG35-55-Induced Experimental Autoimmune Encephalomyelitis, an Animal Model of Multiple Sclerosis. Brain Behav Immun 2020;84:253–68. https://doi.org/10.1016/j.bbi.2019.12.009.
- [78] Cura JE, Blanzaco DP, Brisson C, Cura MA, Cabrol R, Larrateguy L, et al. Phase I and Pharmacokinetics Study of Crotoxin (Cytotoxic PLA(2), NSC-624244) in Patients with Advanced Cancer. Clin Cancer Res 2002;8:1033–41.
- [79] Sampaio SC, Hyslop S, Fontes MRM, Prado-Franceschi J, Zambelli VO, Magro AJ, et al. Crotoxin: Novel Activities for a Classic Beta-Neurotoxin. Toxicon 2010;55: 1045–60. https://doi.org/10.1016/j.toxicon.2010.01.011.
- [80] Almeida CF, Amaral C, Augusto TV, Correia-da-Silva G, Marques de Andrade C, Torqueti MR, et al. The Anti-Cancer Potential of Crotoxin in Estrogen Receptor-Positive Breast Cancer: Its Effects and Mechanism of Action. Toxicon 2021;200: 69–77. https://doi.org/10.1016/j.toxicon.2021.07.003.
- [81] Nogueira-Neto F. de S, Amorim RL, Brigatte P, Picolo G, Ferreira WA, Gutierrez VP, et al. The analgesic effect of crotoxin on neuropathic pain is mediated by central muscarinic receptors and 5-lipoxygenase-derived mediators. Pharmacol Biochem Behav 2008, 91, 252–260.
- [82] Tsetlin V. Snake Venom α-Neurotoxins and Other 'Three-Finger' Proteins. Eur J Biochem 1999;264:281–6. https://doi.org/10.1046/j.1432-1327.1999.00623.x.
- [83] Utkin YN. Three-Finger Toxins, a Deadly Weapon of Elapid Venom-Milestones of Discovery. Toxicon 2013;62:50–5. https://doi.org/10.1016/j. toxicon.2012.09.007.
- [84] Toxins for Decoding Interface Selectivity in Nicotinic Acetylcholine Receptors | Biochemical Journal | Portland Press Available online: https://portlandpress. com/biochemj/article-abstract/476/10/1515/219570/Toxins-for-decodinginterface-selectivity-in?redirectedFrom=fulltext (accessed on 30 October 2022).
- [85] Utkin YN. Last Decade Update for Three-Finger Toxins: Newly Emerging Structures and Biological Activities. World J Biol Chem 2019;10:17–27. https:// doi.org/10.4331/wjbc.v10.i1.17.
- [86] Chanda C, Sarkar A, Sistla S, Chakrabarty D. Anti-Platelet Activity of a Three-Finger Toxin (3FTx) from Indian Monocled Cobra (Naja Kaouthia) Venom. Biochem Biophys Res Commun 2013;441:550–4. https://doi.org/10.1016/j. bbrc.2013.10.125.
- [87] Banerjee Y, Mizuguchi J, Iwanaga S, Kini RM. Hemextin AB Complex, a Unique Anticoagulant Protein Complex from Hemachatus Haemachatus (African Ringhals Cobra) Venom That Inhibits Clot Initiation and Factor VIIa Activity*. J Biol Chem 2005;280:42601–11. https://doi.org/10.1074/jbc.M508987200.
- [88] Yang CC. Cobrotoxin: Structure and Function. J Nat Toxins 1999;8:221–33.
- [89] Xu JM, Song ST, Feng FY, Huang FL, Yang Y, Xie GR, et al. Cobrotoxin-Containing Analgesic Compound to Treat Chronic Moderate to Severe Cancer Pain: Results from a Randomized, Double-Blind, Cross-over Study and from an Open-Label Study. Oncol Rep 2006;16:1077–84. https://doi.org/10.3892/or.16.5.1077.
- [90] Reid, P.F.; Qin, Z.H. (54) USE OF COBRATOXINAS AN ANALGESIC.
- [91] Arruda Macèdo JK, Fox JW, Souza Castro M. Disintegrins from Snake Venoms and Their Applications in Cancer Research and Therapy. Curr Protein Pept Sci 2015; 16:532–48. https://doi.org/10.2174/1389203716666150515125002.
- [92] McLane MA, Sanchez EE, Wong A, Paquette-Straub C, Perez JC. Disintegrins. Curr Drug Targets Cardiovasc Haematol Disord 2004;4:327–55. https://doi.org/ 10.2174/1568006043335880.
- [93] Hamidi H, Ivaska J. Every Step of the Way: Integrins in Cancer Progression and Metastasis. Nat Rev Cancer 2018;18:533–48. https://doi.org/10.1038/s41568-018-0038-z.
- [94] Alberts B, Johnson A, Lewis J, Raff M, Roberts K, Integrins WP. Molecular Biology of the Cell. 4th edition 2002.
- [95] Montealegre-Sánchez, L.; Gimenes, S.N.C.; Lopes, D.S.; Teixeira, S.C.; Solano-Redondo, L.; Rodrigues, V. de M.; Jiménez-Charris, E. Antitumoral Potential of Lansbermin-I, a Novel Disintegrin from Porthidium Lansbergii Lansbergii Venom on Breast Cancer Cells. *Current Topics in Medicinal Chemistry* 19, 2069–2078.
- [96] Olaoba OT, Karina dos Santos P, Selistre-de-Araujo HS, Ferreira de Souza DH. Snake Venom Metalloproteinases (SVMPs): A Structure-Function Update. Toxicon: X 2020;7:100052. https://doi.org/10.1016/j.toxcx.2020.100052.
- [97] R.L.B. Lino P.K. dos Santos G.F.D. Pisani W.F. Altei M.R. Cominetti H.S. Selistrede-Araújo Alphavbeta3 Integrin Blocking Inhibits Apoptosis and Induces Autophagy in Murine Breast Tumor Cells. Biochimica et Biophysica Acta (BBA) -Molecular Cell Research 118536 2019, 1866, doi:10.1016/j. bbamcr.2019.118536.
- [98] Beeton, C. Chapter 64 Targets and Therapeutic Properties. In Handbook of Biologically Active Peptides (Second Edition); Kastin, A.J., Ed.; Academic Press: Boston, 2013; pp. 473–482 ISBN 978-0-12-385095-9.
- [99] Kuo Y-J, Chung C-H, Pan T-Y, Chuang W-J, Huang T-F. A Novel αIIbβ3 Antagonist from Snake Venom Prevents Thrombosis without Causing Bleeding. Toxins (Basel) 2019;12:11. https://doi.org/10.3390/toxins12010011.
- [100] Kuo Y-J, Chung C-H, Huang T-F. From discovery of snake venom disintegrins to a safer therapeutic antithrombotic agent. Toxins 2019;11:372. https://doi.org/ 10.3390/toxins11070372.
- [101] Bentur OS, Coller BS. In vitro effects of the novel platelet αIIbβ3 receptor antagonist RUC-4 on the verifynow assays: potential for point-of-care monitoring of RUC-4 therapy. Blood 2019;134:166. https://doi.org/10.1182/blood-2019-124548.

- [102] Boldrini-França J, Pinheiro-Junior EL, Peigneur S, Pucca MB, Cerni FA, Borges RJ, et al. Beyond hemostasis: a snake venom serine protease with potassium channel blocking and potential antitumor activities. Sci Rep 2020;10:4476. https://doi. org/10.1038/s41598-020-61258-x.
- [103] Carone SEI, Menaldo DL, Sartim MA, Bernardes CP, Caetano RC, da Silva RR, et al. BjSP, a novel serine protease from bothrops jararaca snake venom that degrades fibrinogen without forming fibrin clots. Toxicol Appl Pharmacol 2018; 357:50–61. https://doi.org/10.1016/j.taap.2018.08.018.
- [104] Serrano SMT. The long road of research on snake venom serine proteinases. Toxicon 2013;62:19–26. https://doi.org/10.1016/j.toxicon.2012.09.003.
- [105] Ferraz CR, Arrahman A, Xie C, Casewell NR, Lewis RJ, Kool J, et al. Multifunctional toxins in snake venoms and therapeutic implications: from pain to hemorrhage and necrosis. Front Ecol Evol 2019:7.
- [106] Thrombin-like Enzyme Batroxobin Bothrops Atrox (Barba Amarilla) | UniProtKB | UniProt Available online: https://www.uniprot.org/uniprotkb/P04971/entry (accessed on 12 November 2023).
- [107] Yitao H, Kefu M, Bingshan T, Xuejun F, Ying Z, Zhili C, et al. Effects of batroxobin with continuous transcranial doppler monitoring in patients with acute cerebral stroke: a randomized controlled trial. Echocardiography 2014;31:1283–92. https://doi.org/10.1111/echo.12559.
- [108] Song S, Wu H, Ji X, Meng R. The BE COOL Treatments (Batroxobin, oxygEn, Conditioning, and cOOLing): Emerging Adjunct Therapies for Ischemic Cerebrovascular Disease. J Clin Med 2022;11:6193. https://doi.org/10.3390/ jcm11206193.
- [109] Lan D, Song S, Liu Y, Jiao B, Meng R. Use of batroxobin in central and peripheral ischemic vascular diseases: a systematic review. Front Neurol 2021;12:716778. https://doi.org/10.3389/fneur.2021.716778.
- [110] The Efficacy and Safety of Batroxobin in Combination with Anticoagulation on Cerebral Venous Sinus Thrombosis | SpringerLink Available online: https://link. springer.com/article/10.1007/s11239-018-1718-y (accessed on 4 November 2022).
- [111] Cázares-Ordoñez V, Pardo LA. Kv10.1 potassium channel: from the brain to the tumors. Biochem Cell Biol 2017;95:531–6. https://doi.org/10.1139/bcb-2017-0062.
- [112] Gustina AS, Trudeau MC. HERG potassium channel regulation by the N-terminal eag domain. Cell Signal 2012;24:1592–8. https://doi.org/10.1016/j. cellsig.2012.04.004.
- [113] Ferrari CZ, Ribeiro R, Lima AM, Soares AM, Cavalcante WLG, Vieira LB. Gyroxin, a toxin from crotalus durissus terrificus snake venom, induces a calcium dependent increase in glutamate release in mice brain cortical synaptosomes. Neuropeptides 2020;83:102081. https://doi.org/10.1016/j.npep.2020.102081.
- [114] Hossmann V, Heiss W-D, Bewermeyer H, Wiedemann G. Controlled trial of ancrod in ischemic stroke. Arch Neurol 1983;40:803–8. https://doi.org/10.1001/ archneur.1983.04050120053007.
- [115] Olinger CP, Brott TG, Barsan WG, Hedges JR, Glas-Greenwalt P, Pollak VE, et al. Use of ancrod in acute or progressing ischemic cerebral infarction. Ann Emerg Med 1988;17:1208–9. https://doi.org/10.1016/S0196-0644(88)80071-4.
- [116] Sherman DG, Atkinson RP, Chippendale T, Levin KA, Ng K, Futrell N, et al. for the STAT participants intravenous ancrod for treatment of acute ischemic strokeThe STAT Study: a randomized controlled trial. JAMA 2000;283:2395–403. https:// doi.org/10.1001/jama.283.18.2395.
- [117] Ancrod for the treatment of acute ischemic brain infarction. The ancrod stroke study investigators. *Stroke* 1994, 25, 1755–1759, doi:10.1161/01.str.25.9.1755.
- [118] Caplan, L.R. Chapter 5 Treatment. In Caplan's Stroke (Fourth Edition); Caplan, L. R., Ed.; W.B. Saunders: Philadelphia, 2009; pp. 146–217 ISBN 978-1-4160-4721-6.
- [119] Liu S, Marder VJ, Levy DE, Wang S-J, Yang F, Paganini-Hill A, et al. Ancrod and fibrin formation. Stroke 2011;42:3277–80. https://doi.org/10.1161/ STROKEAHA.111.622753.
- [120] Miyamoto JG, Kitano ES, Zelanis A, Nachtigall PG, Junqueira-de-Azevedo I, Sant'Anna SS, et al. A novel metalloproteinase-derived cryptide from bothrops cotiara venom inhibits angiotensin-converting enzyme activity. Biochimie 2023, doi:10.1016/j.biochi.2023.10.010.
- [121] Oyama E, Takahashi H. Structures and Functions of Snake Venom Metalloproteinases (SVMP) from protobothrops venom collected in Japan. Molecules 2017;22:1305. https://doi.org/10.3390/molecules22081305.
- [122] Takeda S, Takeya H, Iwanaga S. Snake venom metalloproteinases: structure, function and relevance to the mammalian ADAM/ADAMTS Family Proteins. Biochimica et Biophysica Acta (BBA) - Proteins and Proteomics 2012;1824: 164–76. https://doi.org/10.1016/j.bbapap.2011.04.009.
- [123] Okamoto DN, Kondo MY, Oliveira LCG, Honorato RV, Zanphorlin LM, Coronado MA, et al. P-I Class metalloproteinase from bothrops moojeni venom is a post-proline cleaving peptidase with kininogenase activity: insights into substrate selectivity and kinetic behavior. Biochim Biophys Acta 2014;1844: 545–52. https://doi.org/10.1016/j.bbapap.2013.12.014.
- [124] Han Y-P, Lu X-Y, Wang X-F, Xu J. Isolation and Characterization of a Novel P-II Class Snake Venom Metalloproteinase from Trimeresurus Stejnegeri. Toxicon 2007;49:889–98. https://doi.org/10.1016/j.toxicon.2006.11.030.
- [125] Takeda, S. Structure-Function Relationship of Modular Domains of P-III Class Snake Venom Metalloproteinases. In Venom Genomics and Proteomics; Gopalakrishnakone, P., Calvete, J.J., Eds.; Toxinology; Springer Netherlands: Dordrecht, 2016; pp. 185–209 ISBN 978-94-007-6416-3.
- [126] Maria DA, Silva MGL, Correia Junior MC, Ruiz IRG. Antiproliferative effect of the jararhagin toxin on B16F10 Murine Melanoma. BMC Complement Altern Med 2014;14:446. https://doi.org/10.1186/1472-6882-14-446.

- [127] Tanjoni I, Weinlich R, Della-Casa MS, Clissa PB, Saldanha-Gama RF, de Freitas MS, et al. Jararhagin, a snake venom metalloproteinase, induces a specialized form of apoptosis (Anoikis) selective to endothelial cells. Apoptosis 2005;10:851–61. https://doi.org/10.1007/s10495-005-2945-1.
- [128] Costa TR, Burin SM, Menaldo DL, de Castro FA, Sampaio SV. Snake Venom Lamino acid oxidases: an overview on their antitumor effects. J Venom Anim Toxins Incl Trop Dis 2014;20:23. https://doi.org/10.1186/1678-9199-20-23.
- [129] Moustafa IM, Foster S, Lyubimov AY, Vrielink A. Crystal Structure of LAAO from Calloselasma Rhodostoma with an L-phenylalanine substrate: insights into structure and mechanism. J Mol Biol 2006;364:991–1002. https://doi.org/ 10.1016/j.jmb.2006.09.032.
- [130] Yong Y, Hiu JJ, Yap MKK. Chapter Seven the secretory phenotypes of envenomed cells: insights into venom cytotoxicity. In: advances in protein chemistry and structural biology; Donev, R., Ed.; Secretory Proteins; Academic Press, 2023; Vol. 133, pp. 193–230.
- [131] Costal-Oliveira F, Stransky S, Guerra-Duarte C, Naves de Souza DL, Vivas-Ruiz DE, Yarlequé A, et al. L-Amino acid oxidase from bothrops atrox snake venom triggers autophagy, apoptosis and necrosis in normal human keratinocytes. Sci Rep 2019;9:781. https://doi.org/10.1038/s41598-018-37435-4.
- [132] Zhang Y-J, Wang J-H, Lee W-H, Wang Q, Liu H, Zheng Y-T, et al. Molecular characterization of trimeresurus stejnegeri venom L-amino acid oxidase with potential anti-HIV Activity. Biochem Biophys Res Commun 2003;309:598–604. https://doi.org/10.1016/j.bbrc.2003.08.044.
- [133] Tan KK, Bay BH, Gopalakrishnakone P. L-amino acid oxidase from snake venom and its anticancer potential. Toxicon 2018;144:7–13. https://doi.org/10.1016/j. toxicon.2018.01.015.
- [134] Teixeira, T.L.; Oliveira Silva, V.A.; da Cunha, D.B.; Polettini, F.L.; Thomaz, C.D.; Pianca, A.A.; Zambom, F.L.; da Silva Leitão Mazzi, D.P.; Reis, R.M.; Mazzi, M.V. Isolation, Characterization and Screening of the in Vitro Cytotoxic Activity of a Novel L-Amino Acid Oxidase (LAAOcdt) from Crotalus Durissus Terrificus Venom on Human Cancer Cell Lines. *Toxicon* 2016, 119, 203–217, doi:10.1016/j. toxicon.2016.06.009.
- [135] Li Lee M, Chung I, Yee Fung S, Kanthimathi M, Hong Tan sN. Antiproliferative Activity of King Cobra (Ophiophagus Hannah) Venom I-Amino Acid Oxidase. Basic Clin Paharmacol Toxicol 2014;114:336–43. https://doi.org/10.1111/ bcpt.12155.
- [136] Peng SS, Kumar TK, Jayaraman G, Chang CC, Yu C. Solution Structure of Toxin b, a Long Neurotoxin from the Venom of the King Cobra (Ophiophagus Hannah). J Biol Chem 1997:272:7817–23. https://doi.org/10.1074/jbc.272.12.7817.
- [137] Pu XC, Wong PT, Gopalakrishnakone P. A Novel Analgesic Toxin (Hannalgesin) from the Venom of King Cobra (Ophiophagus Hannah). Toxicon 1995;33: 1425–31. https://doi.org/10.1016/0041-0101(95)00096-5.
- [138] Hernández-Goenaga J, López-Abán J, Protasio AV, Vicente Santiago B, del Olmo E, Vanegas M, et al. Peptides derived of kunitz-type serine protease inhibitor as potential vaccine against experimental schistosomiasis. Front Immunol 2019;10.
- [139] Durani, V.; Magliery, T.J. Chapter Eleven Protein Engineering and Stabilization from Sequence Statistics: Variation and Covariation Analysis. In *Methods in Enzymology*; Keating, A.E., Ed.; Methods in Protein Design; Academic Press, 2013; Vol. 523, pp. 237–256.
- [140] Ranasinghe SL, McManus DP. Protease inhibitors of parasitic flukes: emerging roles in parasite survival and immune defence. Trends Parasitol 2017;33:400–13. https://doi.org/10.1016/j.pt.2016.12.013.
- [141] Sintsova O, Gladkikh I, Monastyrnaya M, Tabakmakher V, Yurchenko E, Menchinskaya E, et al. Sea Anemone Kunitz-Type Peptides Demonstrate Neuroprotective Activity in the 6-Hydroxydopamine Induced Neurotoxicity Model. Biomedicines 2021;9:283. https://doi.org/10.3390/ biomedicines9030283.
- [142] Tadokoro T, Modahl M, C., Maenaka, K., Aoki-Shioi, N. Cysteine-Rich Secretory Proteins (CRISPs) from Venomous Snakes: An Overview of the Functional Diversity in a Large and Underappreciated Superfamily. Toxins (Basel) 2020;12: 175. https://doi.org/10.3390/toxins12030175.
- [143] Badari JC, Díaz-Roa A, Teixeira Rocha MM, Mendonça RZ, da Silva Junior PI. Patagonin-CRISP: Antimicrobial Activity and Source of Antimicrobial Molecules in Duvernoy's Gland Secretion (Philodryas Patagoniensis Snake). Front Pharmacol 2021;11:586705. https://doi.org/10.3389/fphar.2020.586705.
- [144] Zhang X, Zhao L, Jin R, Li M, Li M-S, Li R, et al. CRISPR/Cas9-Mediated α-ENaC Knockout in a Murine Pancreatic β-Cell Line. Front Genet 2021;12.
- [145] Potter LR, Yoder AR, Flora DR, Antos LK, Dickey DM. Natriuretic Peptides: Their Structures, Receptors, Physiologic Functions and Therapeutic Applications. Handb Exp Pharmacol 2009:341–66. https://doi.org/10.1007/978-3-540-68964-5_15.
- [146] Ang WF, Koh CY, Kini RM. From Snake Venoms to Therapeutics: A Focus on Natriuretic Peptides. Pharmaceuticals (Basel) 2022;15:1153. https://doi.org/ 10.3390/ph15091153.
- [147] Arlinghaus FT, Eble JA. C-Type Lectin-like Proteins from Snake Venoms. Toxicon 2012;60:512–9. https://doi.org/10.1016/j.toxicon.2012.03.001.
- [148] Teixeira C, Fernandes CM, Leiguez E, Chudzinski-Tavassi AM. Inflammation Induced by Platelet-Activating Viperid Snake Venoms: Perspectives on Thromboinflammation. Front Immunol 2019;10.

- [149] Jenkins TP, Laprade WM, Sánchez A, Tulika T, O'Brien C, Sørensen CV, et al. AHA: AI-Guided Tool for the Quantification of Venom-Induced Haemorrhage in Mice. Frontiers in Tropical Diseases 2022;3.
- [150] Laprade W, Bartlett KE, Christensen CR, Kazandjian TD, Patel RN, Crittenden E, et al. Machine-learning guided venom induced dermonecrosis analysis tool: VIDAL. Sci Rep 2023;13:21662. https://doi.org/10.1038/s41598-023-49011-6.
- [151] Zhang J, Chen X, Song A, Li X. Artificial intelligence-based snakebite identification using snake images, snakebite wound images, and other modalities of information: a systematic review. Int J Med Inf 2023;173:105024. https://doi. org/10.1016/j.ijmedinf.2023.105024.
- [152] Moon K, Im H-J, Kwon S. 3D graph contrastive learning for molecular property prediction. Bioinformatics 2023;39:btad371. https://doi.org/10.1093/ bioinformatics/btad371.
- [153] Zhang Z, Xie A, Guan J, Zhou S. Molecular property prediction by semanticinvariant contrastive learning; 2023.
- [154] Wang Y, Wang J, Cao Z, Barati Farimani A. Molecular contrastive learning of representations via graph neural networks. Nat Mach Intell 2022;4:279–87. https://doi.org/10.1038/s42256-022-00447-x.
- [155] Eisenstein M. Artificial intelligence powers protein-folding predictions. Nature 2021;599:706–8. https://doi.org/10.1038/d41586-021-03499-y.
- [156] Vaswani A, Shazeer N, Parmar N, Uszkoreit J, Jones L, Gomez AN, et al. Attention Is All You Need 2017.
- [157] Jumper J, Evans R, Pritzel A, Green T, Figurnov M, Ronneberger O, et al. Highly accurate protein structure prediction with alphaFold. Nature 2021;596:583–9. https://doi.org/10.1038/s41586-021-03819-2.
- [158] Kalogeropoulos K, Bohn M-F, Jenkins DE, Ledergerber J, Sørensen CV, Hofmann N, et al. A comparative study of protein structure prediction tools for challenging targets: snake venom toxins. Bioinformatics 2023.
- [159] Mirdita M, Schütze K, Moriwaki Y, Heo L, Ovchinnikov S, Steinegger M. ColabFold: making protein folding accessible to all. Nat Methods 2022;19: 679–82. https://doi.org/10.1038/s41592-022-01488-1.
- [160] Lee B, Shin MK, Hwang I-W, Jung J, Shim YJ, Kim GW, et al. A deep learning approach with data augmentation to predict novel spider neurotoxic peptides. Int J Mol Sci 2021;22:12291. https://doi.org/10.3390/ijms222212291.
- [161] Perpetuo L, Klein J, Ferreira R, Guedes S, Amado F, Leite-Moreira A, et al. How can artificial intelligence be used for peptidomics? Expert Rev Proteomics 2021; 18:527–56. https://doi.org/10.1080/14789450.2021.1962303.
- [162] Bilodeau C, Jin W, Jaakkola T, Barzilay R, Jensen KF. Generative models for molecular discovery: recent advances and challenges. WIREs Comput Mol Sci 2022;12:e1608.
- [163] Merz Jr KM, De Fabritiis G, Wei G-W. Generative models for molecular design. J Chem Inf Model 2020;60:5635–6. https://doi.org/10.1021/acs.jcim.0c01388.
- [164] Zhang Y, Hu Y, Li H, Liu X. Drug-protein interaction prediction via variational autoencoders and attention mechanisms. Front Genet 2022;13:1032779. https:// doi.org/10.3389/fgene.2022.1032779.
- [165] Seo S, Choi J, Park S, Ahn J. Binding affinity prediction for protein-ligand complex using deep attention mechanism based on intermolecular interactions. BMC Bioinf 2021;22:542. https://doi.org/10.1186/s12859-021-04466-0.
- [166] Li T, Zhao X-M, Li L. Co-VAE: drug-target binding affinity prediction by coregularized variational autoencoders. IEEE Trans Pattern Anal Mach Intell 2022; 44:8861–73. https://doi.org/10.1109/TPAMI.2021.3120428.
- [167] Xuan P, Ye Y, Zhang T, Zhao L, Sun C. Convolutional neural network and bidirectional long short-term memory-based method for predicting drug-disease associations. Cells 2019;8:705. https://doi.org/10.3390/cells8070705.
- [168] Askr H, Elgeldawi E, Aboul Ella H, Elshaier YAMM, Gomaa MM, Hassanien AE. Deep learning in drug discovery: an integrative review and future challenges. Artif Intell Rev 2023;56:5975–6037. https://doi.org/10.1007/s10462-022-10306-1.
- [169] Talat A, Khan AU. Artificial intelligence as a smart approach to develop antimicrobial drug molecules: a paradigm to combat drug-resistant infections. Drug Discov Today 2023;28:103491. https://doi.org/10.1016/j. drudis.2023.103491.
- [170] Wei B, Zhang Y, Gong X. DeepLPI: a novel deep learning-based model for proteinligand interaction prediction for drug repurposing. Sci Rep 2022;12:18200. https://doi.org/10.1038/s41598-022-23014-1.
- [171] Sánchez EE, Migl C, Suntravat M, Rodriguez-Acosta A, Galan JA, Salazar E. The neutralization efficacy of expired polyvalent antivenoms: an alternative option. Toxicon 2019;168:32–9. https://doi.org/10.1016/j.toxicon.2019.06.216.
- [172] Szteiter SS, Diego IN, Ortegon J, Salinas EM, Cirilo A, Reyes A, et al. Examination of the efficacy and cross-reactivity of a novel polyclonal antibody targeting the disintegrin domain in SVMPs to neutralize snake venom. Toxins 2021;13:254. https://doi.org/10.3390/toxins13040254.
- [173] Sriapha C, Rittilert P, Vasaruchapong T, Srisuma S, Wananukul W, Trakulsrichai S. Early adverse reactions to snake antivenom: poison center data analysis. Toxins 2022;14:694. https://doi.org/10.3390/toxins14100694.
- [174] Bedraoui A., El Mejjad S., Alouani Z., Enezari S., A.Galan J., Daouda T., Machine Learning Analysis Suggests Relative Protein Abundance is Weakly Correlated with Snake Venom Toxicity, in Proceedings of the 2nd International Electronic Conference on Toxins, 14–28 July 2023, MDPI: Basel, Switzerland, doi: 10.3390/ IECT2023-14785.