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Abstract

# Efficacy of thankuni and grapes in arsenicosis-affected rat

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#### This study was undertaken to observe the effects of Thankuni and grapes on hematological and biochemical parameters against arsenicosis in Long Evans rats. Sixty male rats were selected to perform the study. The experimental rats were randomly divided into five different groups, with n = 12 in each group. Animals in group T<sub>0</sub> were given normal feed and water and kept as control. Rats of group $T_1$ were given arsenic trioxide (a) 100 mg/L of drinking water orally. Rats of group T<sub>2</sub> were given arsenic trioxide @ 100 mg/L drinking water and Thankuni @ 1gm/kg feed. Group T<sub>3</sub> was given arsenic trioxide @ 100 mg/L drinking water with grapes @ 10 mg /kg body weight thrown DW. Group T<sub>4</sub> was given arsenic trioxide, Thankuni, and grapes with the same dose for up to 45 days, respectively. Four randomly selected rats from each group $(T_0, T_1, T_2, T_3, and T_4)$ were sacrificed at 15-day intervals to determine body weight, hematological and biochemical parameters. At the end of the experiment, the result showed that the minimum weight gain group was T1 whereas the maximum weight gain was found in the T3 and T<sub>4</sub> groups. The weight gain of the T<sub>2</sub> group is better than T<sub>1</sub>. Reduction of TEC and Hb values were significant (P < 0.01) in the T<sub>1</sub> group. Whereas in the rest of the groups, the reduction of TEC and Hb was less than in arsenic-treated groups. In conclusion, Thankuni and grapes have a significant effect on body weight and hematological and biochemical parameters.

Keywords: Arsenicosis; Thankuni; grapes; rat.

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#### 1. Introduction

Arsenic is currently one of the most critical environmental global contaminants and toxicants, particularly in developing countries. Arsenic is a potent cardiovascular toxicant associated with numerous biomarkers of cardiovascular diseases in exposed human populations. The toxic effects of arsenic are mediated by mitochondrial dysfunction and related to arsenic's effect on oxidative stress. Therefore, we investigated the effectiveness of antioxidants against arsenic-induced cardiovascular dysfunction (Pace et al., 2017). Arsenic binds to sulfhydryl groups on proteins, modulates protein metabolism, and generates reactive oxygen species (ROS) (Mehta & Hundal, 2016). The free radical-mediated theory becomes more acceptable to explain arsenic-induced damage. Oxidative stress is acceptable as one of the mechanisms of arsenicosis, which can be mediated by excess production of reactive oxygen species (ROS) (Li et al., 2016). For that reason, nutritional antioxidants in diseases related to oxidative stress have gained immense interest in recent years because they could be used as suitable preventive/therapeutic agents (Pineda et al., 2013).

Arsenic can be found in organic and inorganic forms in water, food, air, and soil. The most important inorganic arsenic compounds are arsenic trioxide, sodium arsenite, arsenic trichloride, arsenic acid, arsenites (trivalent forms), and calcium arsenates (pentavalent forms). Common organic arsenic forms are arsanilic acid monomethylarsinate (MMA), dimethylarsinic acid (DMA) (Delnomdedieu et al., 1995), and arsenobetaine. Of these compounds, the inorganic forms of arsenic exhibit the highest toxicity level (Saha, 2003). The safety limit of arsenic accepted by the Bangladesh Government is 0.05 mg/liter for drinking water (Fazal et al., 2001). The World Health Organization limit for drink-Ukrainian Journal of Veterinary and Agricultural Sciences, 2024, Vol. 7, N 1



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ing water is 0.01 mg/liter, and for foodstuffs is 2 mg/liter on a fresh weight basis (Robinson et al., 2003).

Humans are exposed to arsenic predominantly through contaminated drinking water, whereas inhalation and skin absorption are minor routes of exposure (Shi et al., 2004). Chronic arsenic exposure through drinking water to humans leads to carcinogenesis of almost all organs, skin diseases (viz. hyper-pigmentation, hyperkeratosis) leading to cancers of skin and epithelial tissues; hepatic, renal, cardiovascular, respiratory, central nervous system, gastrointestinal, reproductive complications, and children's intellectual impairment; thereby increasing morbidity and mortality (Kapaj et al., 2006; Mazumder, 2008; Khatun et al., 2020).

The toxic effects of arsenic in the human body and their conventional management have been well studied and reviewed previously (Abdul et al., 2015). But there is no comprehensive account of the studies on alternative options for counteracting arsenic toxicity. Chronic arsenic exposure is associated with many human health conditions, including skin lesions and cancers of the liver, lung, bladder, and skin (Uddin & Huda, 2011; Shathy et al., 2020; Basher et al., 2023). Arsenicosis presents with significant changes in the Serum Glutamate Oxaloacetate Transaminase (SGOT), Serum Glutamate Pyruvate Transaminase (SGPT), serum creatinine, urea, uric acid levels and various hematological parameters like TEC, TLC, Hb, blood sugar level in the Swiss albino rats (Ashraf et al., 2010). The clinical features that invariably relate to the gastrointestinal system are nausea, vomiting, colicky abdominal pain and profuse watery diarrhea (Burrows & Tyrl, 2013) and also arsenic can enter into the blood chain (Ulman et al., 1998). Recently arsenic intoxication in experimental animals has been found to be associated with hepatic tumors (Waalkes et al., 2003), Spermatogenesis (Shukla & Pandey, 1984), inhibition of testicular steroidogenic function (Sarkar et al., 1991), and severe metabolic disorders such as diabetes in humans (Tseng et al., 2002). The natural source of human exposure to arsenic occurs through the consumption of drinking water sourced from groundwater that contains dissolved inorganic arsenic (Nandi et al., 2006). Consequently, population in areas where the water supply is contaminated with arsenic (i.e.; areas of Taiwan, China, and parts of Africa) tend to have significantly higher mortality rates from conditions related to arsenic poisoning, such as bladder, lung, liver, kidney, skin, and colon cancers (Azcue & Nriagu, 1995; Meliker et al., 2007; Asaolu & Asaolu, 2010).

Thankuni (*Centella asiatica*) is an herbaceous, frosttender perennial plant in the flowering family Apiaceae (Ploenkutham, 2018). It is native to the wetlands in Asia and it is used as a culinary vegetable and medicinal herb (Ploenkutham, 2018). Centella grows in temperate and tropical swampy areas in many world regions (Ploenkutham, 2018). Thankuni extract improves the medicinal value and these properties have been ascribed to the active principles viz., Asiaticoside, madecassic acid and madecassocide. These are pentacyclic triterpenes, found to display venous insufficiency, and various vein and wound healing properties (Kant et al., 2019). Thankuni is essential for numerous intrinsic processes. A study of arsenic toxicity and long-term treatment with thankuni in animals could establish unequivocal decisions about the impacts of arsenic.

Resveratrol is the beneficial compound found in red wine that is associated with life extension and some of the health benefits in wine. It is produced in grapes as a defense against toxins, and biological activity, cardio protective effects, antidiabetic and antioxidant effects (Sharma et al., 2018).

Grape Seed Extract (GSE), which is rich in polyphenols, has been demonstrated to possess potent antioxidant properties and is a safe and effective antioxidant compound. Previous studies have shown that the antioxidant activity of GSE is greater than that of vitamins C and E and  $\beta$ -carotene (Bagchi et al., 2000). GSE may exert its antioxidant and anti-inflammatory effects by scavenging oxygen free radicals, inhibiting lipid per oxidation and the formation of inflammatory cytokines, altering cell membrane receptors and intracellular signaling pathway proteins and modulating gene expression (Li et al., 2001; Kris-Etherton et al., 2004). In a previous study, we demonstrated that GSE may inhibit rat liver injury (Pan et al., 2011).

Bangladesh is available for arsenic toxicity only on tubewell water; however, data on the specific treatment for the prevention of arsenic toxicity in both humans and animals is little. Therefore, data on the effective prevention of arsenic toxicity with thankuni and grapes and their comparative efficacy will be the expected new findings, especially for Bangladesh and the world.

#### 2. Materials and methods

#### Experimental animals

A total of 60 male rats of 6 weeks of age were purchased from the International Centre for Diarrheal Disease Research in Bangladesh (ICDDRB). The animals were housed in compartmented rectangular metallic cage under standard laboratory conditions (12 h light: 12 h dark,  $25 \pm 2^{0}$ C and humidity  $60 \pm 5\%$ ). The rats were acclimatized for 15 days in the laboratory before the experiment started.

# Preparation of house

At first, the room and the wire cages were washed by sweeping and washing with tap water using a hose pipe connected to a tap. The room was disinfected with a phenolic disinfectant. The room was allowed to dry by leaving an electric fan and bulb switched on. Proper ventilation was provided.

## Body weight (BW)

The rats were weighed first on Day 0 (Day 0 = immediate previous day of starting treatment) after grouping and marking. Then again on, Day 15, Day 30, and finally on Day 45. All results were recorded.

#### Clinical signs

Experimental rats were closely observed after feeding arsenic trioxide and Thankuni daily at three times (morning, afternoon, and evening), looking for the appearance of any toxic signs in them. The observations were conducted throughout the entire experimental period (from Day 1 to Day 45), and the findings were recorded.

#### Experimental trial

The experimental trial was conducted for 45 days. The rats in Group  $T_0$  were maintained with only normal pellet feed and water ad libitum as control. The rats in Group  $T_1$  were treated with arsenic trioxide at a dose of 100 mg/L in drinking water. The rats in Group  $T_2$  were treated with arsenic trioxide at 100 mg/L in drinking water daily and Thankuni simultaneously, at a dose of 1 gm/kg feed. The Thankuni used in this experiment was collected from the local market. The rats in Group  $T_3$  were treated with arsenic trioxide at 100 mg/L in drinking water daily and 2 market. The rats in Group Ta were treated with arsenic trioxide at 100 mg/L in drinking water daily and grapes

from a local supermarket, simultaneously, at a dose of 10 mg/kg per bodyweight. The animals of Group  $T_4$  were treated with arsenic trioxide at 100 mg/L in drinking water daily and grapes at a dose of 10 mg/kg body weight and Thankuni simultaneously at a dose of 1gm/kg feed. All treatments were given for 45 days.

#### Sampling

After administering the treatment for 15 days, four rats from each group were anesthetized using chloroform anesthesia, and they were then sacrificed. Blood samples from each rat (about six milliliters) were collected directly from cardiac punctures, using disposable plastic syringe. The blood from each rat was then transferred into two tubes to determine biochemical parameters, and hematological tests. For the biochemical test, four milliliters (4 ml) of blood sample were taken into pre-marked centrifuge glass test tubes immediately after collection. The collected blood was kept at room temperature to allow it to clot properly, then stored in a refrigerator overnight. The serum was separated the next morning by centrifugation and the supernatant serum was taken into pre-marked Eppendorf tubes. The harvested serum was kept at -20 °C until used. One milliliter (1 ml) of blood was taken into EDTA-coated tubes to run hematological and arsenic concentration tests. All lungs, livers, and kidneys were collected aseptically, washed with physiologic saline, and kept in pre-marked zipper polythene bag. Blood samples for hematological investigation were preserved at 4 °C temperature. The blood samples were taken first on Day 15, then on Day 30, and lastly on Day 45.

Biochemical tests

Sera was thawed on the laboratory bench, and the SGOT and, SGPT activity, as well as the serum creatinine were determined by using Thankuni (Deneke & Rittersdorf, 1984; Deneke et al., 1985).

# Statistical analysis

The collected data were statistically analyzed as per Steel and Torrie (1980), using the Completely Randomized Design (CRD). An analysis of variance (ANOVA) and Duncan's Multiple Range Test (DMRT) were performed on SPSS 20 software to determine the difference among the treatments.

# 3. Results and discussion

# Body weight (BW) of the rats

The body weight (BW) of the experimental rats in all groups were taken at fifteen days intervals, on Day 0, Day 15, Day 30, and Day 45. Table 1 shows that the body weight gain was the highest (298.40  $\pm$  3.72) in T<sub>2</sub> group rats at 45 days, but the lowest body weight gain (97.60  $\pm$  2.63) was found in the arsenic treated T<sub>1</sub> group at 45 days. The body weight gain in T<sub>0</sub>, T<sub>3</sub> and T<sub>4</sub> were (256.80  $\pm$  5.13), (283.80  $\pm$  7.17), and (274.80  $\pm$  2.84), respectively, which were better than in the arsenic treated T<sub>1</sub> group. The initial body weight of the groups was not significant (P > 0.05), but then in 15, 30, and 45 days, the mean value of body weight was significant (P < 0.01).

The body weight of the treated group increased with their age, but in the  $T_1$  group it decreased compared to other groups. In the present study, arsenic reduced body weight as the age increased. The highest body weight gain was found in the  $T_2$  group, where Thankuni were treated with arsenic. This suggests that Thankuni acts against arsenic by decreasing body weight. Sharma et al. (2007) reported that reduced body weight was observed in an arsenic treated group of Swiss albino mice, and it also significantly (P < 0.01) decreased the body weight of rats (Sharma et al., 2007; Wu et al., 2008).

#### Table 1

Effects of Thankuni and grapes on the body weight of arsenic-fed rats

Treatment	To	T1	$T_2$	T3	T4	P. value
Initial	$78.20\pm3.28$	$82.60\pm2.50$	$88.46\pm3.32$	$84.20\pm4.32$	$85.00\pm2.78$	NS
15 days	$171.00 \pm 5.56$ <sup>b</sup>	$87.40 \pm 4.02$ a	$181.80 \pm 5.21$ bc	$188.40 \pm 4.97^{\circ}$	$195.00 \pm 5.17$ °	**
30 days	$220.20 \pm 4.32^{\ b}$	$94.80\pm2.63~^{\mathrm{a}}$	$261.20 \pm 2.92$ <sup>d</sup>	$255.40 \pm 2.04^{\ d}$	$239.40\pm4.74^{\circ}$	**
45 days	$256.80 \pm 5.13^{\ b}$	$97.60\pm2.63~^{\mathrm{a}}$	$298.40 \pm 3.72^{\; d}$	$283.80 \pm 7.17^{\rm c}$	$274.80\pm2.84^{\text{c}}$	**

*Note:* within each row, figures with the same or without superscripts do not differ significantly as per the DMRT. The data were calculated at a 99 % level of significance (P < 0.01)

# Hematological parameter

# Total Erythrocyte Count (TEC)

In Table 2, Total Erythrocyte Count (TEC) values were the highest ( $8.54 \pm 0.15$ ) in the T<sub>4</sub> group at 45 days, where Thankuni and grapes were used against arsenic toxicity, but

the lowest value (6.34  $\pm$  0.26) was found in the T<sub>1</sub> group, where only arsenic was given. The TEC values at 15 days (7.12  $\pm$  0.03) and 45 days were significant at (P < 0.01), and the values found at 30 days (7.64  $\pm$  0.27) were also significant at (P < 0.05).

# Table 2

Effects of Thankuni and grapes on Total Erythrocyte Count (TEC) values of arsenic-fed rats

Treatment	T <sub>0</sub>	$T_1$	$T_2$	T3	$T_4$	P. value
15 Days	$6.40\pm0.13$ $^{\rm a}$	$6.20\pm0.04$ a	$6.48\pm0.13^{\text{ a}}$	$6.43\pm0.13$ $^{\mathrm{a}}$	$7.12\pm0.03$ <sup>b</sup>	**
30 Days	$6.48\pm0.18^{\ ab}$	$6.23\pm0.04~^{a}$	$6.98 \pm 0.18^{b}$	$6.98 \pm 0.13$ <sup>b</sup>	$7.64\pm0.27^{\circ}$	*
45 Days	$6.71\pm0.25^{\ ab}$	$6.34\pm0.26^{a}$	$7.34\pm0.26^{\text{c}}$	$7.55\pm0.17^{\text{c}}$	$8.54\pm0.15^{\rm \ d}$	**

*Note:* within each row, figures with the same, or without superscripts do not differ significantly as per the DMRT. The data was calculated at a 99 % level of significance (P < 0.01)

# Total Leukocyte Count (TLC):

In Table 3, total leukocyte counts on day 30 were found to be the highest (10.86  $\pm$  0.004) in the control group rats, and lowest in the T<sub>4</sub> group rats, where Thankuni and grapes were administered. The differences were statistically significant among all groups of rats (P < 0.01). Hence, it can be deduced that Thankuni and grapes decrease the TLC level.

# Hemoglobin (Hb):

The highest Hb concentration  $(15.26 \pm 0.76)$  was found in the T<sub>4</sub> group at 30 days, and the lowest concentration was found in the T<sub>0</sub> group (Table 4). The difference among the values of Hb concentration at 30 days was statistically significant (P < 0.01), and the difference among the Hb concentration values at 15 and 45 days was statistically significant (P < 0.05). It might be concluded that Thankuni and grapes might slightly increase the values of Hb against arsenic toxicity in rats.

# Erythrocyte Sedimentation Rate (ESR):

The highest ESR value was observed in the  $T_4$  group, and the lowest value was observed in the control group

(Table 5). The differences between the mean values of different groups were found to be significant (P < 0.05). It was found that hemoglobin and hematocrit values were reduced in arsenic toxicities in rats as observed in the present study, but in other groups, it was increased. The cause of the change in hematological values might be due to the toxic effect of arsenic on the hematopoietic system which is responsible for such alterations in hematological parameters. However, it is assumed that the toxic effects of arsenic trioxide on bone marrow may be responsible for erythrocytopenia (Islam et al., 2009).

# Packed Cell Volume (PCV):

The PCV values decreased in the arsenic-treated group. The highest values were found in the  $T_4$  group, and the lowest values were found in the arsenic treated  $T_1$  group (Table 6). The differences between the mean values of the different groups were found to be non-significant. The cause of the change in hematological values might be due to the toxic effect of arsenic on the hematopoietic system, which is responsible for such alterations in hematological parameters.

#### Table 3

Effects of Thankuni and grapes on Total Leukocyte Count (TLC) values of arsenic-fed rats

Treatment	T <sub>0</sub>	T1	$T_2$	$T_3$	T4	P. value
15 Days	$9.56 \pm 0.003 \ ^{d}$	$9.09\pm0.005~^{\mathrm{a}}$	$9.41 \pm 0.005$ <sup>b</sup>	$9.37 \pm 0.030^{\ b}$	$9.51 \pm 0.006$ °	**
30 Days	$10.86 \pm 0.004$ °	$10.24 \pm 0.005$ a	$10.42 \pm 0.005$ °	$10.27 \pm 0.005 \ ^{\rm b}$	$10.76 \pm 0.006$ <sup>d</sup>	**
45 Days	$10.82 \pm 0.003$ °	$9.79\pm0.008~^{\rm a}$	$9.87\pm0.005^{\text{ ab}}$	$9.88 \pm 0.010^{\ b}$	$10.80\pm 0.058{}^{\rm c}$	**
	a					

*Note*: within each row, figures with the same, or without superscripts do not differ significantly as per the DMRT. The data was calculated at a 99 % level of significance (P < 0.01)

#### Table 4

Effects of Thankuni and grapes on Hemoglobin concentration (Hb) (gm/dl) values of arsenic-fed rats

Treatment	To	$T_1$	$T_2$	T3	T4	P. Value
15 Days	$13.50 \pm 0.65$ <sup>b</sup>	$11.13 \pm 0.43$ a	$14.00 \pm 0.41$ <sup>b</sup>	$13.13 \pm 0.31$ <sup>b</sup>	$12.88\pm0.43^{\text{ b}}$	*
30 Days	$11.03 \pm 0.89^{\ b}$	$8.45\pm0.65^{\text{ a}}$	$15.26 \pm 0.76$ °	$14.98\pm0.72^{\text{ c}}$	$15.25\pm0.78^{\rm c}$	**
45 Days	$15.13 \pm 0.97^{\ b}$	$7.75\pm1.78^{\ a}$	$15.28 \pm 0.83 \ ^{\rm b}$	$15.05 \pm 1.05$ <sup>b</sup>	$17.20 \pm 1.54^{\text{ b}}$	*

*Note*: within each row, figures with the same, or without superscripts do not differ significantly as per the DMRT. The data was calculated at a 99 % level of significance (P < 0.01)

#### Table 5

Effects of Thankuni and grapes on Erythrocyte Sedimentation Rate (ESR) (gm/dl) values of arsenic-fed rats

Treatment	T <sub>0</sub>	$T_1$	T2	T3	Τ4	P. value
15 Days	$43.88 \pm 2.78^{\ b}$	$39.75\pm1.03^{\text{ ab}}$	$38.38 \pm 0.85{}^{\rm a}$	$36.25 \pm 0.85^{\;a}$	$36.50 \pm 0.65  ^{a}$	*
30 Days	$42.00 \pm 3.58^{b}$	$31.00 \pm 1.47^{a}$	$45.50 \pm 2.10^{b}$	$45.13 \pm 2.56^{b}$	$41.50 \pm 1.32^{\ b}$	*
45 Days	$42.25 \pm 2.81^{\ b}$	$24.25 \pm 2.17^{\ a}$	$47.00 \pm 3.45^{\ b}$	$49.63 \pm 3.90^{\; b}$	$50.25 \pm 7.16^{b}$	*

*Note*: within each row, figures with the same or without superscripts do not differ significantly as per the DMRT. The data was calculated at a 99 % level of significance (P < 0.01)

#### Table 6

Effects of Thankuni and grapes on Packed Cell Volume (PCV) (gm/dl) values of arsenic-fed rats

Treatment	T <sub>0</sub>	$T_1$	$T_2$	T3	T4	P. value
15 Days	$13.67\pm1.33$	$9.00\pm0.58$	$12.00\pm1.53$	$15.00\pm2.52$	$16.00\pm3.21$	NS
30 Days	$12.00\pm0.58$	$11.00\pm0.58$	$13.33\pm0.88$	$15.67\pm0.67$	$19.00\pm2.08$	NS
45 Days	$18.33\pm3.84$	$9.00\pm0.58$	$15.33\pm0.33$	$21.67\pm6.01$	$20.00\pm2.89$	NS

*Note:* within a row, figures with the same or without superscripts do not differ significantly as per DMRT. The data were calculated at a 99 % level of significance (P < 0.01)

# **Biochemical parameters**

# Serum Glutamate Oxaloacetate Transaminase activity (SGOT)

The highest values of SGOT were observed in the control group  $(T_0)$ , while the lowest values were observed in the

*tate Transaminase activity* were observed in the convalues were observed in the CV and F sponse to prolonged administration of arsenic, the combination of Thankuni and grapes produced a more significant reduction in SGOT level comparable to the control group (P < 0.01). However, this finding disagreed with the previous findings that SGOT was reduced by as alone (Mahaffey et al., 1981; Ashraf et al., 2010). In Thankuni-treated (T<sub>2</sub>), grapes-treated (T<sub>3</sub>), and Thankuni plus grapes-treated (T<sub>4</sub>) experimental arsenicosis groups, significantly decreased values of arsenic were recorded (P < 0.01).

# Table 7

Effects of Thankuni and grapes on SGOT values of arsenic-fed rats

Treatment	$T_0$	$T_1$	$T_2$	T <sub>3</sub>	T4	P. value
15 Days	$115.00 \pm 2.89$ °	$110.00 \pm 289^{bc}$	$100.00 \pm 2.52$ a	$106.67 \pm 4.41$ abc	$102.00 \pm 0.58^{ab}$	*
30 Days	$109.00 \pm 3.79^{\rm \ bc}$	$117.00 \pm 8.50$ °	$97.33 \pm 1.45^{\text{ ab}}$	$98.67\pm1.86^{ab}$	$88.00 \pm 1.53{}^{\rm a}$	**
45 Days	$109.67 \pm 3.18^{\ b}$	$141.67 \pm 6.23$ °	$106.67 \pm 1.67$ <sup>b</sup>	$105.00 \pm 2.89^{\ b}$	$84.33\pm2.33~^{\mathrm{a}}$	**

*Note*: within a row, figures with the same or without superscripts do not differ significantly as per DMRT. The data was calculated at a 99 % level of significance (P < 0.01)

# Serum Glutamate Pyruvate Transaminase activity (SGPT)

Continuous administration of arsenic to Long-Evans rats caused a significant increase in the blood SGPT level. The highest values of SGPT were observed in the  $T_1$  group, where the rats were treated with only arsenic. There were insignificant differences within the groups during days 15 and 30, but this difference became statistically significant (P < 0.01) by day 45. The lowest values of SGPT were observed in the  $T_4$  group, where the combination of Thankuni and grapes was administered against arsenic toxicity. In 15 days and 30 days, blood SGPT levels were increased however it is not statistically significant (P > 0.05). In 45 days, the level of blood SGPT decreased at a statistically significant (P < 0.01) (Table 8). Overall, SGPT values have a decreasing trend with the progress of time in all groups, which agreed with the previous research findings (Islam, 2008). It may be concluded that prolonged treatment with Thankuni and grapes may reduce the blood SGPT level.

# Table 8

Effects of Thankuni and grapes on SGPT values of arsenic-fed rats

Treatment	T <sub>0</sub>	$T_1$	T <sub>2</sub>	T <sub>3</sub>	T4	P. value
15 Days	$71.67\pm0.89$	$77.00 \pm 1.53$	$64.33\pm7.17$	$72.67 \pm 1.45$	$79.00 \pm 1.52$	NS
30 Days	$73.00\pm0.58$	$76.00\pm3.51$	$68.67\pm8.95$	$64.33 \pm 2.33$	$70.67\pm0.67$	NS
45 Days	$74.00 \pm 0.00^{\; b}$	$89.33\pm4.84^{\text{ c}}$	$68.67 \pm 7.36^{b}$	$42.33\pm0.89^{\text{ a}}$	$45.67 \pm 2.33$ a	**

*Note*: Within a row, figures with the same or without superscripts do not differ significantly as per DMRT. The data was calculated at a 99 % level of significance (P < 0.01).

Figures indicate the Mean  $\pm$  SE (standard error); NS means not significant.

\*\* = Significant at P < 0.01 level of probability

\*= Significant at P < 0.05 level of probability.

#### Serum creatinine

Serum creatinine with the highest values was found in the T<sub>3</sub> group at 45 days, and the lowest values were observed in the control groups. The differences were found to be significant (P < 0.05) on day 15. The differences between the mean values from the 30 and 45-day groups were found to be significant (P < 0.01). On day 30, the lowest mean values were observed in the control group. The highest mean values were observed in the T<sub>3</sub> group rats, and the differences were statistically significant (P < 0.01). On day 45, the lowest mean values were observed in the T<sub>2</sub> group, and the highest mean values were observed in the T<sub>3</sub> group rats. These differences were statistically significant (P < 0.01). The differences between groups T<sub>2</sub> and T<sub>3</sub> were statistically significant (P < 0.01). However, the contents increased in the T<sub>1</sub>, T<sub>3</sub>, and T<sub>4</sub> groups, but decreased in the T<sub>2</sub> group on day 30, compared to day 45. On day 45, the values of serum creatinine were the highest in the T<sub>3</sub> group rats and lowest in the T2 group. The differences were observed to be significant (P < 0.01) on day 45 (Table 9). There was a significant difference in serum creatinine level observed between the control group and all other treatment group rats throughout the whole study period, which depicts dissimilar results with previous studies (Nurun Nabi et al., 2005) involving human beings. The study showed that the patients with arsenicosis had significantly lower levels of serum creatinine compared to the control and it was also observed that there was a relationship between arsenic level and the degree of chronic renal insufficiency in men (Zheng et al., 2015). There were no significant rises in the serum creatinine levels of arsenic-treated mice (Mitchell et al., 2000; Islam et al., 2009).

#### Table 9

Effects of Thankuni and grapes on Serum creatinine values of arsenic-fed rats

Treatment	T <sub>0</sub>	$T_1$	T2	T3	T4	P. value
15 Days	$0.51\pm0.005$ a	$0.62\pm0.003^{\circ}$	$0.52\pm0.005$ $^{a}$	$0.59\pm0.042^{\circ}$	$0.53\pm0.003$ $^{a}$	*
30 Days	$0.52\pm0.003$ $^{\mathrm{a}}$	$0.65\pm0.005^{\text{c}}$	$0.54 \pm 0.008^{\;b}$	$0.64\pm0.005^{\circ}$	$0.52\pm0.006^{\text{ a}}$	**
45 days	$0.52\pm0.006^{\text{ a}}$	$0.67\pm0.003$ $^{\rm c}$	$0.51 \pm 0.003$ a	$0.69 \pm 0.012$ °	$0.57 \pm 0.023^{\ b}$	**

*Note:* within a row, figures with the same or without superscripts do not differ significantly as per DMRT. The data was calculated at a 99% significance level (P < 0.01)

# 4. Conclusions

A study of arsenic toxicity and long-term treatment with Thankuni and grapes in animals could be able to establish unequivocal decisions about the impacts of arsenic, Thankuni, and grapes on biochemical parameters, as well as provide strong evidence of the efficacy of Thankuni treatment for arsenic toxicities. Treatment with Thankuni and grapes might increase body weight. Thankuni and grapes alone can reduce the effects of arsenic toxicity, but if both are used in combination, it may be more effective. Arsenic toxicity has adverse effects on hematological and biochemical parameters in rats. Thankuni and grapes have a protective effect in improving these parameters. Combined Thankuni and grapes treatment is more effective for arsenic toxicity. This study suggested that Thankuni and grapes have significantly reduced the arsenic concentration of inorganic arsenic toxicity in rats. Further investigation in this line may make more clear evidence to use Thankuni as a treatment for arsenic toxicity.

# **Conflict of interest**

The authors declare that there is no conflict of interest.

#### References

Abdul, K. S. M., Jayasinghe, S. S., Chandana, E. P., Jayasumana, C., & De Silva, P. M. C. (2015). Arsenic and human health effects: A review. Environmental Toxicology and Pharmacology, 40(3), 828-846.

[Crossref] [Google Scholar]

Asaolu, S., & Asaolu, M. (2010). Trace metal distribution in Nigerian leafy vegetables. Pakistan Journal of Nutrition, 9(1), 91-92.

[Crossref] [Google Scholar]

- Ashraf, M., Hayat, M. Q., Jabeen, S., Shaheen, N., Khan, M. A., & Yasmin, G. (2010). Artemisia L. species recognized by the local community of northern areas of Pakistan as folk therapeutic plants. Journal of Medicinal Plants Research, 4(2), 112–119.
  - [Abstract] [Google Scholar]
- Azcue, J. M., & Nriagu, J. O. (1995). Impact of abandoned mine tailings on the arsenic concentrations in Moira Lake, Ontario. Journal of Geochemical Exploration, 52(1-2), 81-89. [Crossref] [Google Scholar]
- Bagchi, D., Bagchi, M., Stohs, S. J., Das, D. K., Ray, S. D., Kuszynski, C. A., Joshi, S. S., & Pruess, H. G. (2000). Free radicals and grape seed proanthocyanidin extract: importance in human health and disease prevention. Toxicology, 148(2-3), 187-197.
- [Crossref] [Google Scholar] Basher, M. K., Sarkar, S., Mobarak, M. H., & Islam, M. R. (2023). Inorganic arsenic exposure during pregnancy affects post-natal growth, blood parameters, and organ development of mice offspring. Journal of Advanced Biotechnology Experimental Therapeutics, 6(2), 445–457.

[Crossref] [Google Scholar]

- Burrows, G. E., & Tyrl, R. J. (2013). Toxic plants of north America. John Wiley & Sons. [Abstract] [Google Scholar]
- Delnomdedieu, M., Styblo, M., & Thomas, D. J. (1995). Time dependence of accumulation and binding of inorganic and organic arsenic species in rabbit erythrocytes. Chemico-Biological Interactions, 98(1), 69-83. [Crossref] [Google Scholar]

- Deneke, U., & Rittersdorf, W. (1984). Evaluation of the Refloquant GPT (ALT) reagent carriers with Reflotron. Clinical Chemistry, 30, 1009. [Google Scholar]
- Deneke, U., Rittersdorf, W., & Werner, W. (1985). Performance data of Reflotron-GOT (AST) dry chemistry test for Reflotron. Clinical Chemistry, 31(6), 921–921. [Google Scholar]
- Fazal, M. A., Kawachi, T., & Ichion, E. (2001). Extent and severity of groundwater arsenic contamination in Bangladesh. Water International, 26(3), 370-379. [Google Scholar]
- Islam, M. (2008). Comparative efficacy of spirulina and spinach extract against arsenic toxicity in rats. Thesis, Department of Pharmacology, Faculty of Veterinary Science, Bangladesh Agricultural University, Mymensingh, Bangladesh. [Google Scholar]
- Islam, M., Awal, M., Mostofa, M., Begum, F., Khair, A., & Myenuddin, M. (2009). Effect of spirulina on toxic signs, body weight and hematological parameters in arsenic induced toxicities in ducks. International Journal of Poultry Science, 8(1), 75–79.

[Crossref] [Google Scholar]

- Kant, R., Srivastav, P. P., & Datta, A. K. (2019). The medicinal role of Centella asiatica and its applications in the Dahi: a research review. Journal of Pharmaceutical Research International, 28(6), 1-9. [Crossref] [Google Scholar]
- Kapaj, S., Peterson, H., Liber, K., & Bhattacharya, P. (2006). Human health effects from chronic arsenic poisoning-a review. Journal of Environmental Science and Health, Part A, 41(10), 2399-2428.

[Crossref] [Google Scholar]

- Khatun, M. F., Hasan, M. M., Islam, R., Sarkar, S., & Haque, M. A. (2020). Effect of spirulina (Spirulina platensis) and vitamin E on arsenic induced toxicity in Quail. Asian Journal of Medical and Biological Research, 6(1), 93–98. [Crossref] [Google Scholar]
- Kris-Etherton, P. M., Lefevre, M., Beecher, G., Gross, M., Keen, C. L., & Etherton, T. D. (2004). Bioactive compounds in nutrition and health-research methodologies for establishing biological function: the antioxidant and anti-inflammatory effects of flavonoids on atherosclerosis. Annual Review of Nutrition, 24, 511–538. [Crossref] [Google Scholar]
- Li, S., Xu, S., Niu, Q., Ding, Y., Pang, L., Ma, R., Jing, M., Wang, K., Ma, X., & Feng, G. (2016). Lutein alleviates arsenicinduced reproductive toxicity in male mice via Nrf2 signaling. Human & Experimental Toxicology, 35(5), 491–500. [Crossref] [Google Scholar]
- Li, W.-G., Zhang, X., Wu, Y., & Tian, X. (2001). Antiinflammatory effect and mechanism of proanthocyanidins from grape seeds. Acta Pharmacologica Sinica, 22(12), 1117-1120. [Abstract] [Google Scholar]
- Mahaffey, K. R., Capar, S. G., Gladen, B. C., & Fowler, B. A. (1981). Concurrent exposure to lead, cadmium, and arsenic: effects on toxicity and tissue metal concentrations in the rat. The Journal of laboratory and clinical medicine, 98(4), 463-481. [Abstract] [Google Scholar]
- Mazumder, D. G. (2008). Chronic arsenic toxicity & human health. Indian Journal of Medical Research, 128(4), 436–447. [Abstract] [Google Scholar]
- Mehta, M., & Hundal, S. (2016). Effect of sodium arsenite on reproductive organs of female Wistar rats. Archives of Environmental & Occupational Health, 71(1), 16-25. [Crossref] [Google Scholar]

and

- Meliker, J. R., Wahl, R. L., Cameron, L. L., & Nriagu, J. O. (2007). Arsenic in drinking water and cerebrovascular disease, diabetes mellitus, and kidney disease in Michigan: a standardized mortality ratio analysis. *Environmental Health*, 6(1), 1–11.
  [Crossref] [Google Scholar]
- Mitchell, R. D., Ayala-Fierro, F., & Carter, D. E. (2000). Systemic indicators of inorganic arsenic toxicity in four animal species. *Journal of Toxicology and Environmental Health Part A*, 59(2), 119–134.

[Crossref] [Google Scholar]

- Nandi, D., Patra, R., & Swarup, D. (2006). Oxidative stress indices and plasma biochemical parameters during oral exposure to arsenic in rats. *Food and Chemical Toxicology*, 44(9), 1579–1584. [Crossref] [Google Scholar]
- Nurun Nabi, A., Rahman, M. M., & Islam, L. N. (2005). Evaluation of biochemical changes in chronic arsenic poisoning among Bangladeshi patients. *International journal of environmental research and public health*, 2(3), 385–393. [Crossref] [Google Scholar]
- Pace, C., Dagda, R., & Angermann, J. (2017). Antioxidants protect against arsenic induced mitochondrial cardio-toxicity. *Toxics*, 5(4), 38.

[Crossref] [Google Scholar]

- Pan, X., Dai, Y., Li, X., Niu, N., Li, W., Liu, F., Zhao, Y., & Yu, Z. (2011). Inhibition of arsenic induced-rat liver injury by grape seed exact through suppression of NADPH oxidase and TGF-β/Smad activation. *Toxicology and Applied Pharmacology*, 254(3), 323–331.
  [Crossref] [Google Scholar]
- Pineda, J., Herrera, A., & Antonio, M. T. (2013). Comparison between hepatic and renal effects in rats treated with arsenic and/or antioxidants during gestation and lactation. *Journal of Trace Elements in Medicine and Biology*, 27(3), 236–241. [Crossref] [Google Scholar]
- Ploenkutham, R. (2018). Development of herbal tea incorporated with asiatic pennywort extract for elderly. A Thesis Submitted to the Department of Food Biotechnology, Assumption University in pati of fulfillment of the requirement for the degree of Master of Science in Food Biotechnology. [Abstract] [Google Scholar]
- Robinson, B., Duwig, C., Bolan, N., Kannathasan, M., & Saravanan, A. (2003). Uptake of arsenic by New Zealand watercress (Lepidium sativum). Science of the total environment, 301(1–3), 67–73. [Crossref] [Google Scholar]
- Saha, K. (2003). Review of arsenicosis in west Bengal, India—a clinical perspective. *Critical Reviews in Environmental Science* and Technology, 33(2), 127–163. [Crossref] [Google Scholar]
- Sarkar, M., Biswas, N., & Ghosh, D. (1991). Effect of sodium arsenite on testicular Δ5-3β and 17β-hydroxysteroid dehydrogenase activities in albino rats: dose-and duration-dependent responses. *Medical Science Research*, 19(22), 789–790.
   [Abstract] [Google Scholar]

- Sharma, A., Sharma, M. K., & Kumar, M. (2007). Protective effect of Mentha piperita against arsenic-induced toxicity in liver of Swiss Albino mice. *Basic & Clinical Pharmacology & Toxicology*, 100(4), 249–257.
  [Crossref] [Google Scholar]
- Sharma, N., Vashist, H., & Kumar, A. (2018). Reputation of herbal drugs in treatments of liver cancer. *Innovat International Journal of Medical & Pharmaceutical Sciences*, 3, 9–16. [Google Scholar]
- Shathy, M. A., Hasan, M. M., Aziz, F. B., Islam, R., & Sarkar, S. (2020). Prevention of arsenic toxicity in quail by using spirulina and garlic. *International Journal of Natural and Social Sciences*, 7, 26–31. [Abstract] [Google Scholar]

Shi, H., Shi, X., & Liu, K. J. (2004). Oxidative mechanism of arsenic toxicity and carcinogenesis. *Molecular and cellular biochemistry*, 255(1–2), 67–78. [Crossref] [Google Scholar]

- Shukla, J. P., & Pandey, K. (1984). Impaired spermatogenesis in arsenic treated freshwater fish, Colisa fasciatus (Bl. and Sch.). *Toxicology Letters*, 21(2), 191–195. [Crossref] [Google Scholar]
- Tseng, C.-H., Tseng, C.-P., Chiou, H.-Y., Hsueh, Y.-M., Chong, C.-K., & Chen, C.-J. (2002). Epidemiologic evidence of diabetogenic effect of arsenic. *Toxicology Letters*, 133(1), 69–76. [Crossref] [Google Scholar]
- Uddin, R., & Huda, N. H. (2011). Arsenic poisoning in Bangladesh. *Oman Medical Journal*, 26(3), 207. [Crossref] [Google Scholar]
- Ulman, C., Gezer, S., Anal, Ö., Töre, I. R., & Kirca, Ü. (1998). Arsenic in human and cow's milk: a reflection of environmental pollution. *Water, Air, and Soil Pollution*, 101, 411–416. [Crossref] [Google Scholar]
- Waalkes, M. P., Ward, J. M., Liu, J., & Diwan, B. A. (2003). Transplacental carcinogenicity of inorganic arsenic in the drinking water: induction of hepatic, ovarian, pulmonary, and adrenal tumors in mice. *Toxicology and Applied Pharmacology*, 186(1), 7–17. [Crossref] [Google Scholar]
- Wu, J., Liu, J., Waalkes, M. P., Cheng, M.-L., Li, L., Li, C.-X., & Yang, Q. (2008). High dietary fat exacerbates arsenic-induced liver fibrosis in mice. *Experimental Biology and Medicine*, 233(3), 377–384.

[Crossref] [Google Scholar]

Zheng, L. Y., Umans, J. G., Yeh, F., Francesconi, K. A., Goessler, W., Silbergeld, E. K., Bandeen-Roche, K., Guallar, E., Howard, B. V., & Weaver, V. M. (2015). The association of urine arsenic with prevalent and incident chronic kidney disease: evidence from the Strong Heart Study. *Epidemiology (Cambridge, Mass.)*, 26(4), 601.

[Crossref] [Google Scholar]