



## OPEN Studying temperature's impact on *Brassica napus* resistance to identify key regulatory mechanisms using comparative metabolomics

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To investigate the effects of temperature on *Brassica napus* (canola) resistance to *Leptosphaeria maculans* (LM), the causal agent of blackleg disease, metabolic profiles of LM infected resistant (R) and susceptible (S) canola cultivars at 21 °C and 28 °C were analyzed. Metabolites were detected in cotyledons of R and S plants at 48- and 120-h post-inoculation with LM using UPLC-QTOF/MS. The mock-inoculated plants were used as controls. Some of the resistance-related specific pathways, including lipid metabolism, amino acid metabolism, carbohydrate metabolism, and aminoacyl-tRNA biosynthesis, were down-regulated in S plants but up-regulated in R plants at 21 °C. However, some of these pathways were down-regulated in R plants at 28 °C. Amino acid metabolism, lipid metabolism, alkaloid biosynthesis, phenylpropanoid biosynthesis, and flavonoid biosynthesis were the pathways linked to combined heat and pathogen stresses. By using network analysis and enrichment analysis, these pathways were identified as important. The pathways of carotenoid biosynthesis, pyrimidine metabolism, and lysine biosynthesis were identified as unique mechanisms related to heat stress and may be associated with the breakdown of resistance against the pathogen. The increased susceptibility of R plants at 28 °C resulted in the down-regulation of signal transduction pathway components and compromised signaling, particularly during the later stages of infection. Deactivating LM-specific signaling networks in R plants under heat stress may result in compatible responses and deduction in signaling metabolites, highlighting global warming challenges in crop disease control.

**Keywords** Metabolome analysis, *Leptosphaeria maculans*, *Brassica napus*, Temperature sensitive resistance

Blackleg or phoma stem canker is a significant disease of canola (*Brassica napus*) and many other cruciferous species worldwide. Blackleg epidemics frequently occur in canola crops across Europe, Canada, and Australia<sup>1,2</sup>. *Leptosphaeria maculans* (Desm.) Ces. & De Not. (LM) and *L. biglobosa* are two closely related species that can cause blackleg disease on canola, previously addressed as aggressive (Tox<sup>+</sup>, A-group) and non-aggressive (less aggressive, Tox<sup>0</sup>, B-group), respectively<sup>1,3-5</sup>. The two species exist in most canola-growing regions<sup>1,4</sup>. In Iran, few studies have been carried out on canola blackleg. The blackleg disease occurs typically in canola-growing areas with humidity sufficient for disease development. Blackleg exists in 30 regions and 10 provinces in Iran<sup>3</sup>. Like Poland, China, and Lithuania, *L. biglobosa* is the dominant species and causes minor damage across canola growing regions in Iran<sup>3</sup>.

The hemibiotrophic fungus LM produces ascospores acting as primary inoculum while conidia produced on infected leaves act as secondary inoculum. Hyphae from germinated ascospores or conidia enter the plant through the stomata or wounds<sup>5</sup>. The fungus grows between the cells and into the stem after the primary infection

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of the leaf. The intercellular growth is systemic, biotrophic, and visually symptomless<sup>6</sup>. Finally, the fungus invades the stem cortical tissues in the necrotrophic phase, where the blackened cankers are developed after killing the invaded tissues. The blackleg disease can also spread by infected seeds when the fungus infects the seed pods of *B. napus*, but this is far less common<sup>4</sup>.

The most cost-effective and reliable management approach for blackleg is the use of resistant cultivars<sup>7</sup>. Both qualitative (monogenic) and quantitative (polygenic) resistances against LM in *Brassica* species have been reported. Quantitative resistance is conferred by several genes, associated with genomic regions or QTL (quantitative trait loci) and expressed in the adult stage<sup>8</sup>; however, qualitative resistance is mediated by single resistance genes and is effective in protecting plants during the leaf infection stage<sup>7</sup>. Twenty-two major genes for resistance to LM have been identified in *Brassica* species (*B. rapa*, *B. nigra*, *B. oleracea*, *B. napus*, and *B. juncea* lines), among all only five genes (*LepR3*, *Rlm2*, *Rlm4*, *Rlm7*, and *Rlm9*) have been cloned, all of which encode membrane-bound cell surface-localized receptor proteins used to detect apoplastic elicitors<sup>9</sup>.

The effects of temperature on host plant resistance responses have been reported in several pathosystems<sup>8</sup>. Formerly, it has been reported that temperature affects effectiveness of both quantitative and *R* gene-mediated qualitative resistances against LM<sup>6,8–11</sup>. The temperature sensitivity for solanaceous resistance is attributable to the NLR sensor and not to its downstream signaling components in potatoes, which is not yet known in other plants, including *Brassicaceae*<sup>12</sup>. Identifying thermostable resistance is essential for understanding the mechanisms of resistance and for better managing the disease under elevated temperatures<sup>11,13</sup>. *Rlm1* and *Rlm6*-mediated resistance to LM are temperature-sensitive<sup>6,14</sup>. When cotyledons of canola cv. Quinta were inoculated with LM isolate A2; the plant response was resistant at 18 °C but susceptible at 27 °C<sup>14</sup>. The interaction between Quinta and the isolate A2 was characterized as a specific *Rlm1*/*AvrLm1* interaction<sup>15,16</sup>. Results of Huang et al. (2006) under controlled environment experiments indicated that *Rlm6*-mediated resistance in DarmorMX (carrying *Rlm6*) against LM was not effective at 25 °C<sup>6</sup>. Investigations of effects of temperature on effectiveness of *R* gene-mediated resistance (e.g. *Rlm4*) in different genetic backgrounds showed that combining *R* gene with quantitative resistance can maintain stability of resistance at elevated temperature<sup>10</sup>. Results of determining temperature sensitivity and stability of *R* gene-mediated resistance at 28 °C by using four cultivars (*Topas-Rlm1*, *Topas-Rlm2*, *Topas-Rlm3*, and *Topas-Rlm4*) under four temperature regimes (15/10, 20/15, 25/20, and 30/25 °C) showed that lesion size is highly dependent on temperature; however, the influence of temperature on *R*-mediated resistance appears to depend on the specific cultivar harboring the *R* gene<sup>17</sup>.

Different 'omics' have been utilized to study how plants react to biotic or abiotic stress conditions. These tools have provided unique opportunities to examine the study of genomics and transcriptomics in the host and the pathogen<sup>18</sup>. However, metabolomics, defined as the comprehensive analysis of metabolites in a biological specimen, and proteomics, compared to the other two branches, are still lagging. Despite the vast diversity of existing metabolites, few of them have been found in host–pathogen interactions with the *Brassicaceae* family<sup>18,19</sup>. Most studies have focused on specific target groups of metabolites, such as glucosinolates, phenylpropanoids, and a small number of phytoalexins<sup>20</sup>.

Metabolites produced in leaves of *B. napus* and *B. rapa* inoculated with either different races of the biotroph *Albugo candida* or sprayed with CuCl<sub>2</sub> were determined by Pedras et al.<sup>19</sup>. This investigation established consistent phytoalexin (spirobrassinin, cyclobrassinin, and rutalexin) and phytoanticipin (indolyl-3-acetonitrile, arvelxin, caulilexin C, and 4-methoxyglucobrassicin) production and six new phenylpropanoids and two new flavonoids in canola<sup>19</sup>. Abdel-Farid et al. used NMR-based metabolomics to study *B. rapa*'s reactions to infection by *L. maculans*, *Aspergillus niger* and *F. oxysporum*. *F. oxysporum*-infected plants accumulated more phenylpropanoids (sinapoyl-, feruloyl, and 5-hydroxyferuloyl malate), flavonoids (kaempferol and quercetin), and fumaric acid than plants infected with the other two species<sup>20</sup>. Huang et al. transposed *A. thaliana* reduce the synapine composition. The transformation led to a change in derivatives of sinapoyl, quercetin, salicylic acid, and indolyl glucosinolates. Transgenic plants were susceptible to fungal infection, and it was found that these metabolites play an important role in protecting the *Brassicaceae* plants<sup>21</sup>. Then, Botanga et al. inoculated *A. thaliana* C01 plants with *Aternaria brassicola* and found approximately half of the identified metabolites were affected by the inoculation. Due to the infection by *A. brassicola*, several phytohormones, oxylipins, amino acids, synthetics, ligands, and hundreds of potential biomarkers were affected<sup>22</sup>. However, very few studies have been done to evaluate metabolic mechanisms of plant defense against infection by the fungal pathogen LM in the *Brassicaceae* family, especially canola. Currently, there is a lack of metabolomics investigations on canola plants subjected to concurrent exposure to high temperature and LM infection. With the aim to improve our knowledge on plant-fungus interactions challenged with high temperatures, the canola cultivars, Excel (carrying the resistance gene *Rlm7*, resistant) and Drakkar (without any resistance genes, susceptible) were inoculated with the LM isolate ME24 under the optimum temperatures (21/16 °C) and elevated temperatures (28/23 °C), the changes in metabolites at different time intervals after inoculation were investigated.

The resistance gene *Rlm7* in *B. napus* is a valuable source of resistance for controlling blackleg disease on canola<sup>2,23–25</sup>. The *Rlm7* gene was more durable than other resistance genes (e.g., *Rlm1*) for control of LM because it is still used in commercial cultivars to prevent blackleg epidemics, even though it has been used commercially for more than 15 years<sup>2,23</sup>. The major goal of this study was to investigate the effects of temperature on *Rlm7* at the metabolite level to understand the metabolic changes in canola plants under various biotic and abiotic stresses. Unraveling the complex plant-pathogen interactions under biotic and abiotic stresses can lead to better understanding of the molecular and physiological basis of host plant resistance to fungi in response to global warming.

## Results

### Disease progress on different cultivars under different temperatures

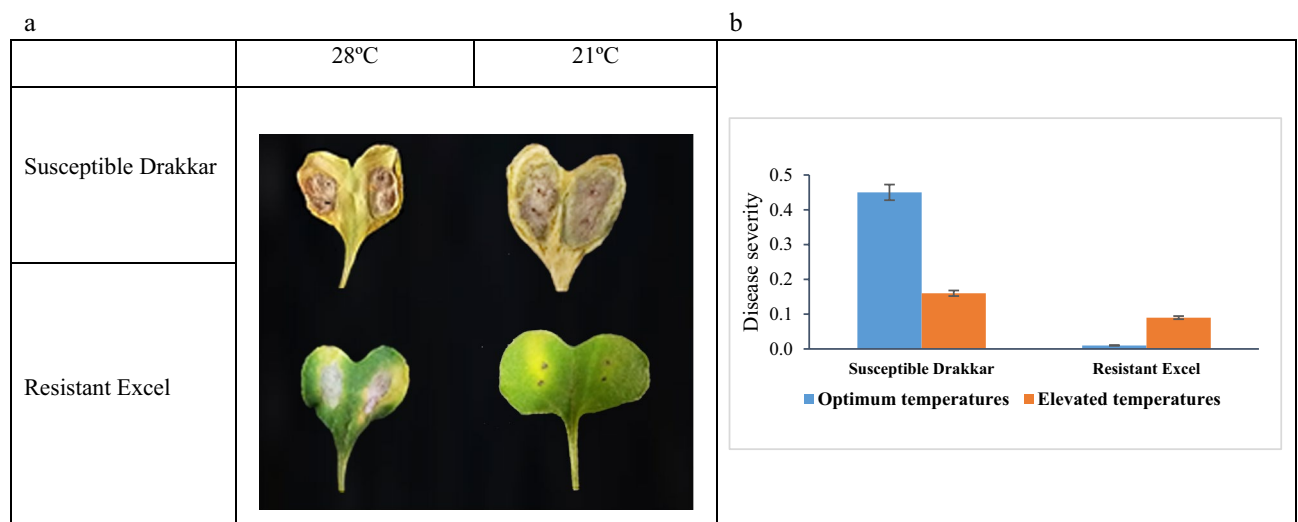
Daily assessment of symptoms showed that disease severity statistically differed between cultivars Excel (resistant, carrying *Rlm7*) and Drakkar (susceptible, lacking any *R* genes) at 21 °C and 28 °C. While large lesions developed on Drakkar at 14 days post inoculation (dpi) at both temperatures, the symptoms differed between temperatures on Excel (Fig. 1a). No visible symptoms were observed on Excel at 21 °C up to 10 dpi and only small dark lesions restricted to the inoculation sites were observed at 14 dpi. However, large lesions were observed on Excel at 28 °C at 14 dpi, suggesting the breakdown of resistance. Assessment of the lesion area of 30 plants for these symptoms in the areas of lesions measured using ImageJ software showed that on the susceptible cultivar Drakkar the lesion size dropped half at 28 °C compared to those at 21 °C. On the other hand, the size of lesions on the resistant cultivar Excel increased three times at 28 °C compared to those at 21 °C (Fig. 1b).

### Metabolic profiles of the *B. napus*-*L. maculans* interactions

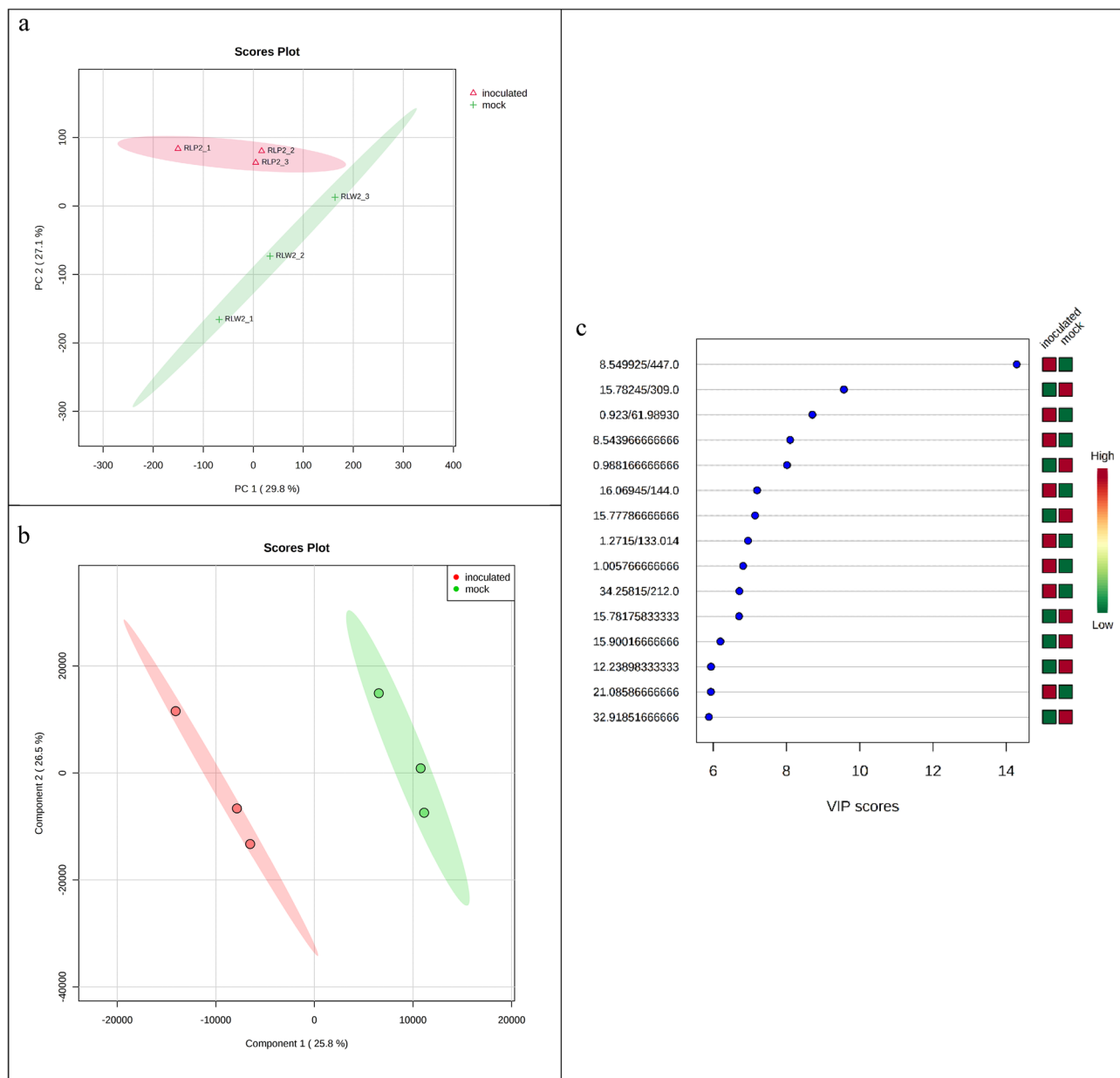
Out of 9966 and 11,652 peaks in positive and negative ionization modes, respectively, 3319 and 7096 metabolites in positive and negative ionization modes were filtered based on a > 30,000 measured mean abundance and used for further multivariate analysis. To obtain candidate metabolites, the selected parent ions were collected using online databases with multivariate analysis. In the positive ionization mode, 2758 metabolites were discovered, while in the negative ionization mode, 6400 metabolites were putatively identified based on matches to publicly accessible databases including METLIN, KEGG, Pubchem, and the Plant Metabolic Network (PMN). In the positive ionization mode, 1369 metabolites at 48 h post inoculation (hpi) and 928 metabolites at 120 hpi and in negative ionization modes, 1988 metabolites at 48 hpi and 1703 metabolites at 120 hpi showed significant ( $p < 0.05$ ) changes between temperatures, cultivars, and/or their interactions. The Supplementary files (Supplementary Tables S1 and S2) provide tables that include information on the annotation and mass error of metabolites, retention time, as well as all transitions and their intensities.

### Univariate and multivariate analysis of the metabolite profile

The score plots obtained from PCA analysis consistently exhibited clustering among the biological replicates for each treatment, indicating a high level of repeatability across various treatments (Figs. 2 and 3; Supplementary Figs. S1–S8). The metabolic profiles of R and S plants were evaluated by creating loading plots to assess the overall differences. To validate the findings of PCA using a more robust pattern recognition technique, we conducted a supervised PLS-DA analysis. The score plots and corresponding VIP (Variable of Importance in Projection) plots derived from the models were built for the metabolic profiles of the canola cotyledon tissues. The results of PLS-DA analyses of the positive and negative ion modes showed that there were differences between cultivars and temperatures in score plots based on the first couple of components (Figs. 2 and 3; Supplementary Figs. S1–S8). The PLS-DA model is believed to be reliable when  $Q^2 > 0.5$  and suitable cross-validation is obtained for  $R^2 > Q^2$ . In this analysis, a  $Q^2 > 0.736$  and  $R^2 > 0.983$  indicated good predictive power. The samples of mock and LM inoculated at both temperatures and at different time points were separated by component 1 only (not component 2) (Figs. 2 and 3). For instance, component 1 measured 22.2% and 21.3% of variations among the samples of Excel inoculated with LM vs mock inoculation, at 48 hpi, at 21 °C and 28 °C, positive ion mode respectively. Our VIP analysis from PLS-DA analysis revealed the top 15 metabolites of negative ion mode for *B. napus*-*L. maculans* pathosystem which showed better sensitivity in terms of peak intensity and numbers (Figs. 2 and 3;



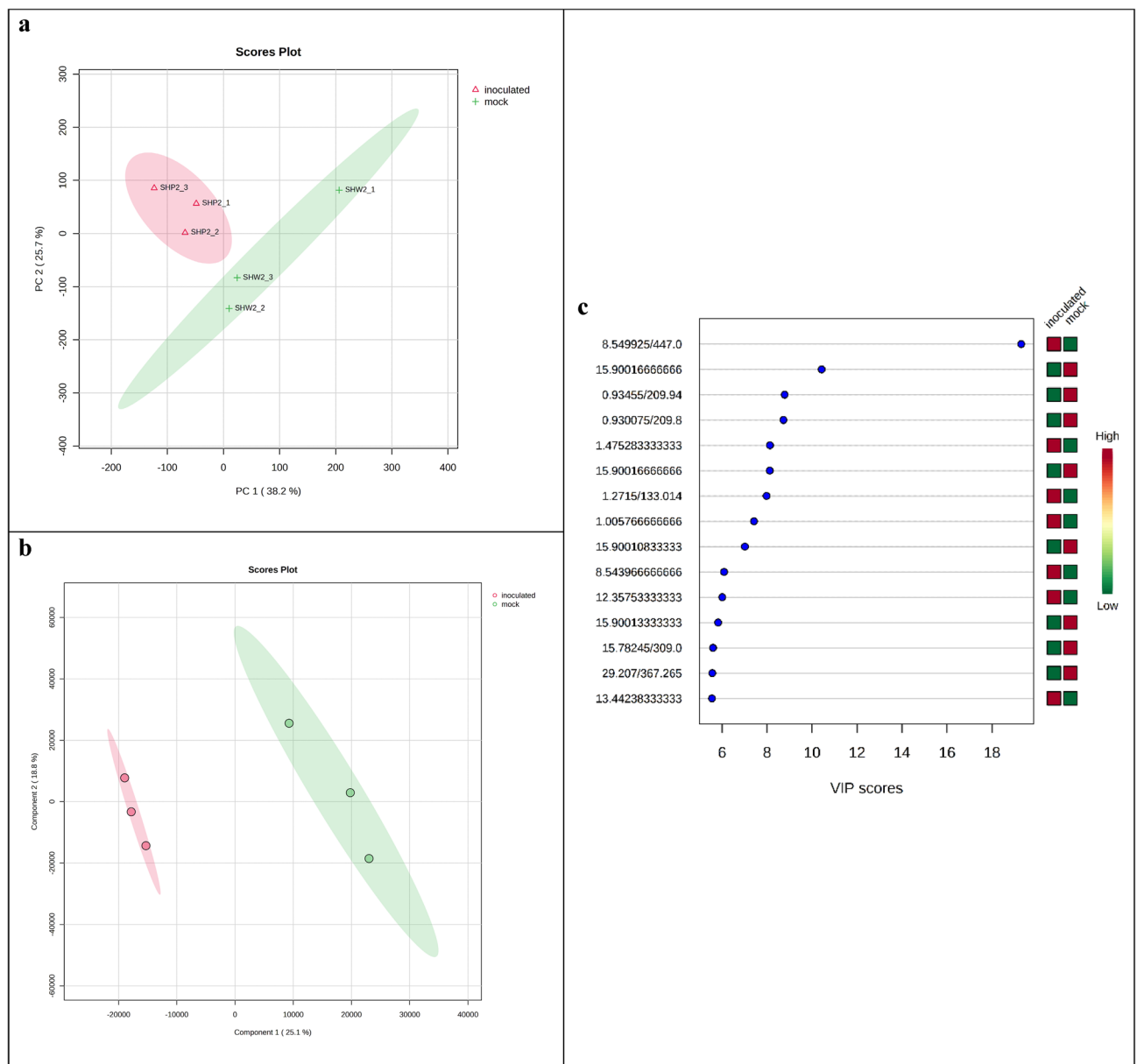
**Figure 1.** (a) The symptoms on cotyledons of cultivars Excel (resistant, carrying *Rlm7*) and Drakkar (susceptible, lacking any *R* genes) at 14 dpi. Plants were inoculated with *Leptosphaeria maculans* isolate ME24 at 21 °C and 28 °C. (b) disease severity on cultivars seedling Excel and Drakkar at 21 °C (optimum temperatures) and 28 °C (Elevated temperatures). Each bar represents the mean of disease severity (on a 0–1 ratio scale) 30 leaf samples and the error bars are the standard error of the mean (SEM).



**Figure 2.** PCA and PLS-DA analysis; **(a)** Principal component analysis PCA score plot showing distinct relationship between samples and differences in resistant cultivar inoculated with the pathogen LM (RP) vs resistant cultivar with mock inoculation (RM) at 48 hpi/21 °C in negative ionization mode with the MetaboAnalystR 6.0 software package along the x-axis (PC1) and within groups along the y-axis (PC2). PC1 and PC2 respectively summarize 29.8% and 27.1% of the observed variance in the metabolites, green and red shaded areas are the 95% confidence regions of each group; **(b)** Scores plot of metabolite features in PLS-DA (partial least squares-discriminant analysis) model representing covariance between component 1 (25.8%) at x-axis and component 2 (26.5%) at y-axis; **(c)** Variable of Importance in Projection (VIP) score plots derived from PLS-DA analysis, a higher VIP score (>2.0) for metabolites represent higher contribution in the PLS-DA model.

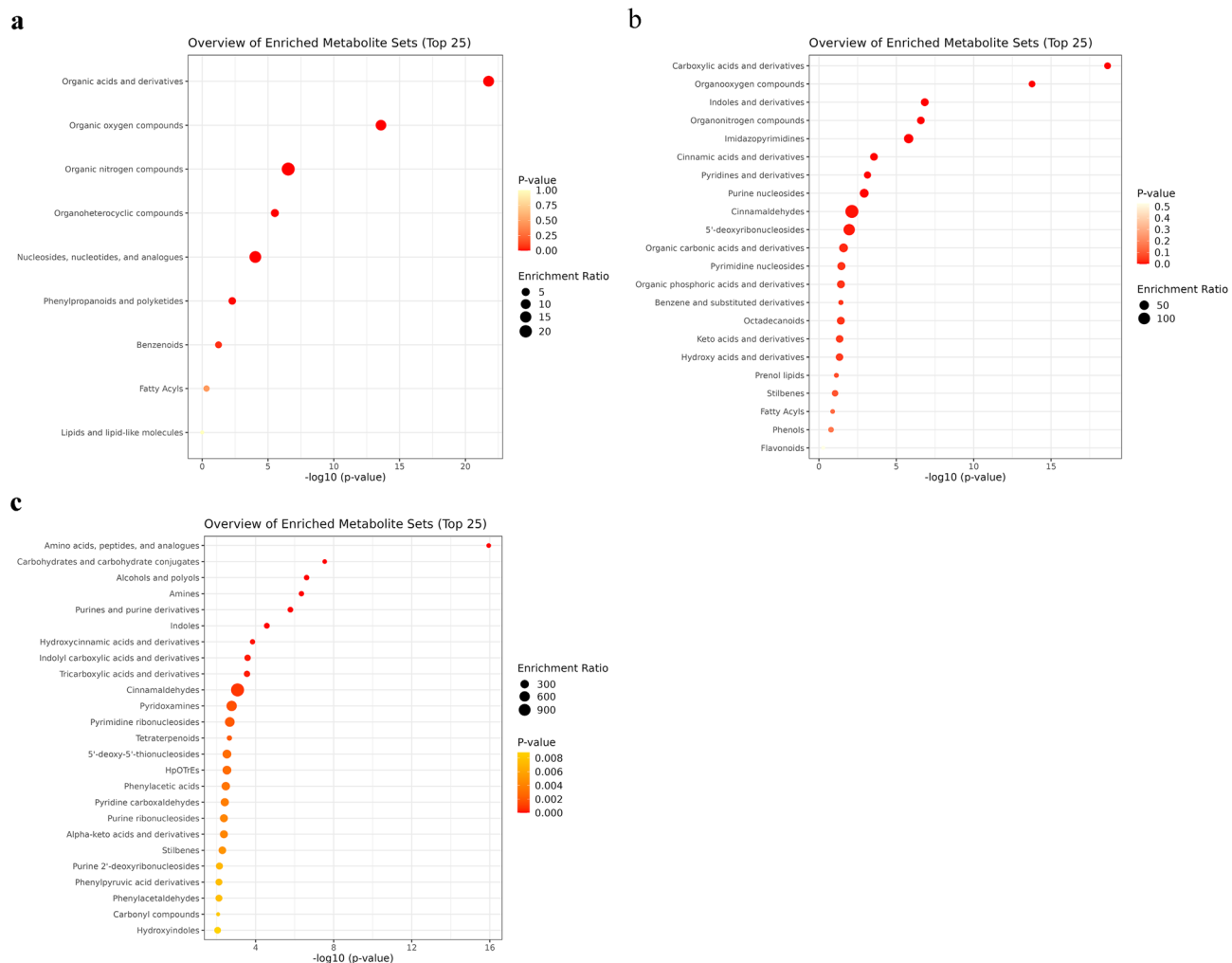
Supplementary Figs. S9–S16). The enrichment analysis module performs metabolite set enrichment analysis (MSEA) on several libraries containing ~6300 groups of metabolite sets. We selected chemical structures as a metabolite set library, which includes: 1. super-class (39 super chemical class metabolite sets or lipid sets), 2. main-class (617 main chemical class metabolite sets or lipid sets), and 3. sub-class (1250 sub-chemical class metabolite sets or lipid sets). The results of enrichment analysis for chemical structure classification of the key metabolites in the *B. naupus*-*L. maculans* pathosystem showed that metabolites are placed in 9 super classes (Fig. 4a and Supplementary Table S3), in 22 main classes (Fig. 4b and Supplementary Table S4), and 39 subclasses (Fig. 4c and Supplementary Table S5).

Fold change ( $FC \geq 2.0$ ) analysis was conducted to identify up- and down-regulated discriminant annotated metabolites by univariate feature analysis in Metaboanalyst 6.0. Fold change ( $FC \geq 2.0$ ) analysis in positive

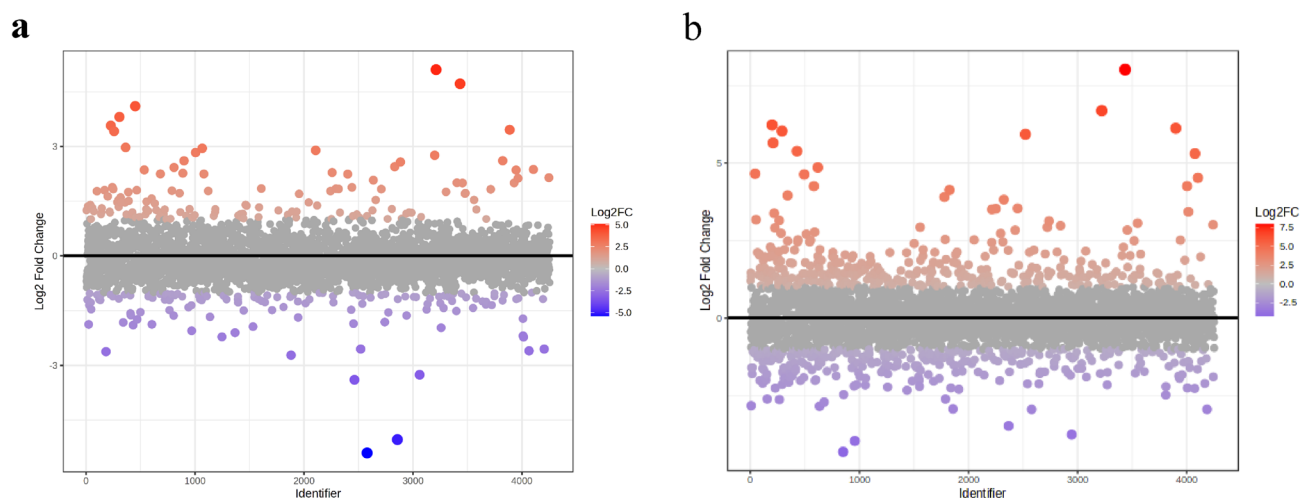


**Figure 3.** PCA and PLS-DA analysis; **(a)** principal component analysis PCA score plot showing distinct relationship between samples and differences in susceptible cultivar inoculated with the pathogen LM (SP) vs susceptible cultivar with mock inoculation (SM) at 48 hpi/28 °C in negative ionization mode with the MetaboAnalystR 6.0 software package along the x-axis (PC1) and within groups along the y-axis (PC2). PC1 and PC2 respectively summarize 38.2% and 25.7% of the observed variance in metabolites, green and red shaded areas are the 95% confidence regions of each group; **(b)** Scores plot of metabolite features in PLS-DA (partial least squares-discriminant analysis) model representing covariance between component 1 (25.1%) at x-axis and component 2 (18.8%) at y-axis; **(c)** Variable of Importance in Projection (VIP) score plots derived from PLS-DA analysis, a higher VIP score (>2.0) for metabolites represent higher contribution in the PLS-DA model.

ionization in resistant cultivar inoculated with the pathogen LM (RP) vs. resistant cultivar with mock inoculation (RM) at 48 hpi /21 °C suggested 95 metabolites significantly up- and 56 down-regulated metabolite (Fig. 5a). In negative ionization, in the RP vs. RM at 48 hpi /21 °C suggested 129 metabolites significantly up- and 142 down-regulated metabolite. Fold change ( $FC \geq 2.0$ ) analysis in positive ionization in RP vs. RM at 48 hpi /28 °C suggested 310 metabolites significantly up- and 109 down-regulated metabolite. In negative ionization, in RP vs. RM at 28 °C/48 hpi suggested 366 metabolites significantly up- and 315 down-regulated metabolite (Fig. 5b). In RP vs. RM at 48hpi/21 °C under  $VIP \geq 2$ , in positive mode, 81 features showed significant differences, 38 up-regulated and 53 down-regulated. In negative mode, RP vs. RM at 48hpi/21 °C, with  $VIP \geq 2$ , 171 features showed significant differences, 76 up-regulated and 95 down-regulated. In elevated temperature, RP vs. RM at 48hpi/28 °C with  $VIP \geq 2$ , in positive mode, 72 features showed significant differences, 42 up-regulated and 30



**Figure 4.** Enrichment analysis for chemical structure classification of the key metabolic pathways in the *Brassica napus*–*Leptosphaeria maculans* pathosystem with the MetaboAnalystR 6.0 software package. **(a)** Supper class, **(b)** Main class, **(c)** Subclass. The color is based on the p-value (darker colors indicate more significance) and the size of each circle is based on the enrichment ratio (the larger indicates a higher ratio). The top 25 crucial chemical groups were studied in each class. The details of the enrichment analysis results for the identified metabolites in each class were presented in supplementary tables (Tables S3, S4, S5).



**Figure 5.** Univariate feature analysis in Metaboanalyst 6.0. fold change (FC threshold  $\geq 2$ ) analysis of metabolite in **(a)** resistant cultivar inoculated with the pathogen LM (RP) vs resistant cultivar with mock inoculation (RM) at 48 hpi/21 °C in positive ionization and **(b)** RP vs RM at 48 hpi/28 °C in negative ionization.

down-regulated. In negative mode, RP vs. RM at 48hpi /28 °C, in VIP  $\geq 2$ , in positive mode, 164 features showed significant differences, 76 up-regulated and 88 down-regulated.

### Effects of temperature on metabolic pathways related to *L. maculans* infection

Only 128 out of 1000 significant metabolites at 21 °C and 28 °C (FDR < 0.05) were recognized in 66 Arabidopsis pathways using the database of Arabidopsis metabolic pathways from KEGG or HMDB as a reference for a global test algorithm (Table 1 and Supplementary Table S6). Most of the 54 and 63 pathways identified in the positive and negative ionization modes were common. By comparing pathways impacted in LM-infected plants of R (resistant) and S (susceptible) cultivars at 21 °C, several pathways down-regulated in S cultivar were up-regulated in the R cultivar at both periods (2 and 5 dpi) and interestingly these pathways were among those down-regulated in R cultivars at 28 °C. In this case, the pathways were classified as either pathogen stress-related or host resistance-related. These pathways included alpha-Linolenic acid metabolism, nicotinate and nicotinamide metabolism, galactose metabolism, histidine metabolism, aminoacyl tRNA biosynthesis, porphyrin and chlorophyll metabolism, fatty acid elongation in mitochondria, fatty acid metabolism, and fatty acid biosynthesis. Some pathways were down-regulated at 21 °C and at the same time up-regulated at 28 °C (Table 1). They could be classified as heat stress-specific related pathways. Lysine biosynthesis, carotenoid biosynthesis, and pyrimidine metabolism were grouped in this category. On the other hand, following pathogen inoculation at 21 °C some pathways were up-regulated in the R cultivar at all-time points and remained up-regulated at 28 °C. Consequently, these pathways that were related to both pathogen and heat stresses were categorized as general or non-specific stress-related pathways (Table 1). Given the fact that a metabolite could be a component of different pathways and influence multiple biological processes, as a result, detecting plant metabolic networks becomes a challenging task. To solve this problem and simplify the process, the standardized overview of pathways was achieved by adapting the KEGG classification for canola pathways (Figs. 6 and 7). Phenylalanine metabolism, tryptophan metabolism, alpha-Linolenic acid metabolism, beta-Alanine metabolism, isoquinoline alkaloid biosynthesis, pantothenate and CoA biosynthesis, and tyrosine metabolism had the most effect on the *B. napus*-*L. maculans* interaction (Figs. 6 and 7). Also, according to the results of network analysis based on the KEGG global metabolic network with the MetaboAnalystR 6.0 software package, 15 significant pathways were identified (Table 2).

### Discussion

Elevated temperature is the primary abiotic stress for plants under climate change. With increased temperatures due to global warming, future plant disease epidemics are anticipated to be more frequent and more severe. In nature, temperature-sensitive resistance in plants has been regularly reported<sup>10–13,26</sup>. Metabolic pathways down/up-regulated under pathogen attack to reinforce the defense mechanisms at elevated temperatures can neutralize defense through altering these regulations, may also be exploited to engineer new crop cultivars and to understand how temperature affects plant innate immunity and discover elite genes that confer disease resistance at elevated temperature<sup>27</sup>. Nevertheless, little is known about the molecular mechanisms of plant defense against pathogen infections and how climate change influences them<sup>26,27</sup>. To better comprehend the physiological responses arbitrated by the resistance gene *Rlm7* when confronted with the pathogen *L. maculans* (LM) under incompatible and compatible interactions, a comparative metabolomic study was done. Our study is the first to examine the effectiveness of the resistance gene *Rlm7* at elevated temperatures using metabolomic techniques.

Large blackleg lesions were observed on the susceptible cultivar Drakkar at both 21 °C and 28 °C. However, the lesion size at 28 °C was half of that at 21 °C. This may be due to reduced growth of the pathogen and drop of humidity at 28 °C which may have slowed down the spread of the lesion. No pycnidia were observed on cotyledons of Drakkar at 28 °C may also be due to low humidity at 28 °C since low humidity is hostile for pathogen sporulation. Small dark lesions around the inoculation areas were observed on the resistance cultivar Excel at 21 °C, while large lesions were observed at 28 °C. The size of lesions on Excel increased three times at 28 °C in comparison with those at 21 °C, suggesting the *Rlm7* gene-mediated resistance in Excel is sensitive to temperature. This is consistent with a recent study of *Rlm7* in the Topas introgression line (Topas-*Rlm7*) which showed that the *Rlm7* is temperature-sensitive in Topas-*Rlm7*<sup>13</sup>. Previous studies showed that the genetic background plays a crucial role in single *Rlm* gene-mediated resistance<sup>10,17,23</sup>, this study reconfirmed that the *Rlm7* resistance gene specific for LM in the Excel cultivar was temperature-sensitive. Similarly, other studies suggest that *Rlm7* is the next best candidate resistance gene to be introduced into the canola-host gene pool<sup>23–25</sup>. The *Rlm9*, *Rlm4*, and *Rlm7* encode wall-associated kinase-like (WAKL) proteins<sup>28,29</sup>. Cloning these three genes has shown them to be allelic variants of one another. WAKLs are a newly emerging class of *Avr*-responsive, ETI-inducing *R* genes.

The findings of this study demonstrate that metabolic profiling is a highly effective approach for investigating the underlying mechanisms of the pathogenesis of the hemibiotrophic pathogen LM and the mechanisms of host resistance in canola. Our results showed that LM could cause up-regulation of a range of metabolic pathways. Some metabolites associated with different pathways were likely to be involved in canola resistance to LM infection. Results of our study also indicate a hierarchical structure in the control of multi-dimensional metabolic networks responding to numerous simultaneous stressors, such as pathogen (biotic) and heat (abiotic). The observed activation of metabolic pathways may indicate the rapid inhibition of the fungal infection, as shown in other plant-pathogen interactions<sup>30</sup>. The results of metabolic network analysis showed three classes of pathways: resistance-related specific pathways, non-specific stress-related pathways, and heat stress-related specific pathways.

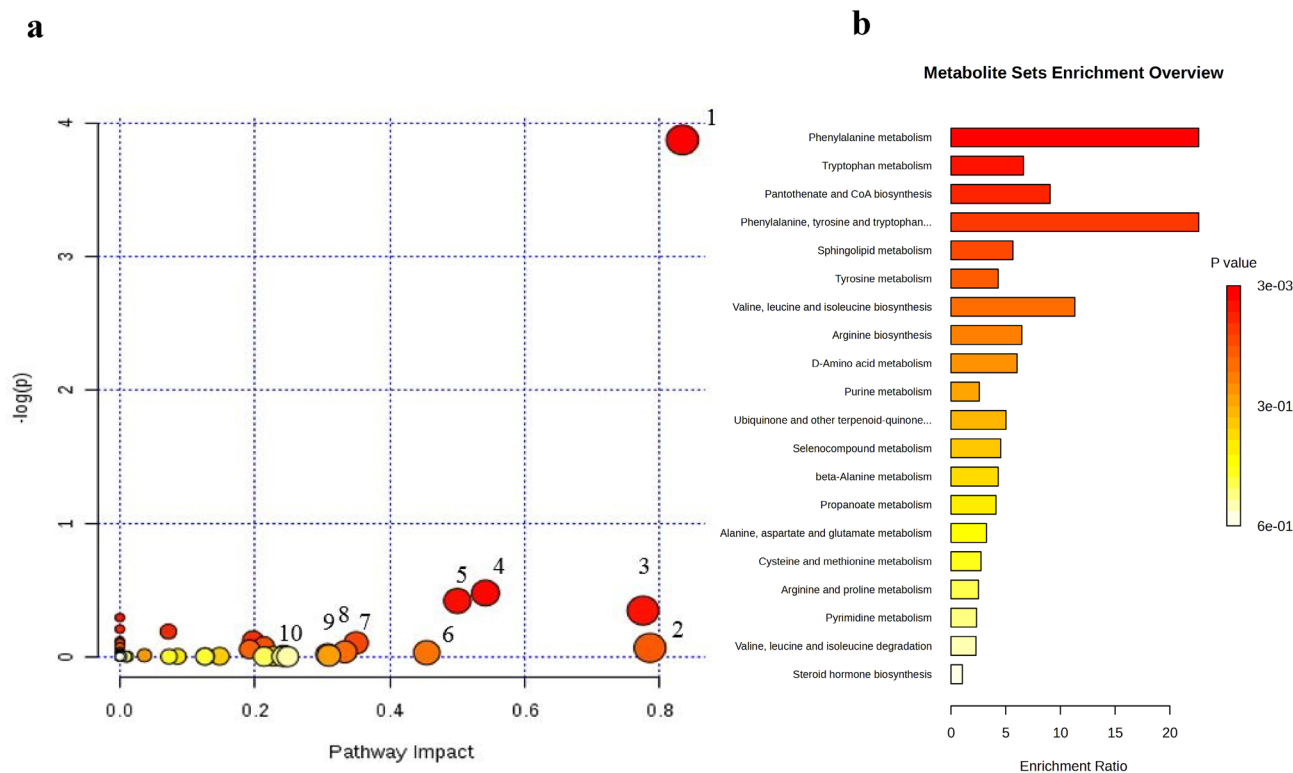
### Resistance-related specific pathways

Metabolic pathways for fatty acids, alpha-linolenic acid, nicotinate and nicotinamide, galactose, histidine, porphyrin and chlorophyll, fatty acid elongation in mitochondria, and aminoacyl tRNA biosynthesis were considered

	Pathway name	21°C				28°C			
		R cultivar		S cultivar		R cultivar		S cultivar	
		48	120	48	120	48	120	48	120
1	Phenylalanine metabolism								
2	beta-Alanine metabolis								
3	Isoquinoline alkaloid biosynthesis								
4	Indole alkaloid biosynthesis								
5	Tropane, piperidine and pyridine alkaloid biosynthesis								
6	alpha-Linolenic acid metabolism								
7	Nicotinate and nicotinamide metabolism								
8	Phenylpropanoid biosynthesis								
9	Phenylalanine, tyrosine and tryptophan biosynthesis								
10	Caffeine metabolism								
11	Stilbenoid, diarylheptanoid and gingerol biosynthesis								
12	Pantothenate and CoA biosynthesis								
13	Glycine, serine and threonine metabolism								
14	Vitamin B6 metabolism								
15	Tryptophan metabolism								
16	Fructose and mannose metabolism								
17	Anthocyanin biosynthesis								
18	Sphingolipid metabolism								
19	Tyrosine metabolism								
20	Flavone and flavonol biosynthesis								
21	Nitrogen metabolism								
22	Limonene and pinene degradation								
23	Lysine biosynthesis								
24	Arginine and proline metabolism								
25	Valine, leucine and isoleucine biosynthesis								
26	Glyoxylate and dicarboxylate metabolism								
27	Pentose and glucuronate interconversions								
28	Butanoate metabolism								
29	Carotenoid biosynthesis								
30	Zeatin biosynthesis								
31	Inositol phosphate metabolism								
32	Cysteine and methionine metabolism								
33	Citrate cycle (TCA cycle)								
34	Glutathione metabolism								
35	Galactose metabolism								
36	Histidine metabolism								
37	Glucosinolate biosynthesis								
38	Ubiquinone and other terpenoid-quinone biosynthesis								
39	Pyrimidine metabolism								
40	Pentose phosphate pathway								
41	Glycerophospholipid metabolism								
42	Glycolysis or Gluconeogenesis								
43	Terpenoid backbone biosynthesis								
44	Selenoamino acid metabolism								
45	Aminoacyl-tRNA biosynthesis								
46	Amino sugar and nucleotide sugar metabolism								
47	Carbon fixation in photosynthetic organisms								
48	Valine, leucine and isoleucine degradation								
49	Alanine, aspartate and glutamate metabolism								
50	Starch and sucrose metabolism								
51	Biosynthesis of unsaturated fatty acids								
52	Porphyrin and chlorophyll metabolism								
53	Purine metabolism								
54	Flavonoid biosynthesis								
55	C5-Branched dibasic acid metabolism								
56	Riboflavin metabolism								
57	Ascorbate and aldarate metabolism								
58	Cyanoamino acid metabolism								
59	Pyruvate metabolism								
60	Sulfur metabolism								
61	Fatty acid elongation in mitochondria								
62	Propanoate metabolism								
63	Lysine degradation								
64	Diterpenoid biosynthesis								
65	Fatty acid metabolism								
66	Fatty acid biosynthesis								

**Table 1.** Metabolic pathways differentially regulated in R (resistant) and S (susceptible) cultivars at 48- and 120-h post inoculation at 21 °C and 28 °C in the *Brassica napus*—*Leptosphaeria maculans* pathosystem. Pathway alterations were shown using colors. Up and down-regulated pathways are shown by light blue and red colors, respectively ( $p < 0.05$ ).



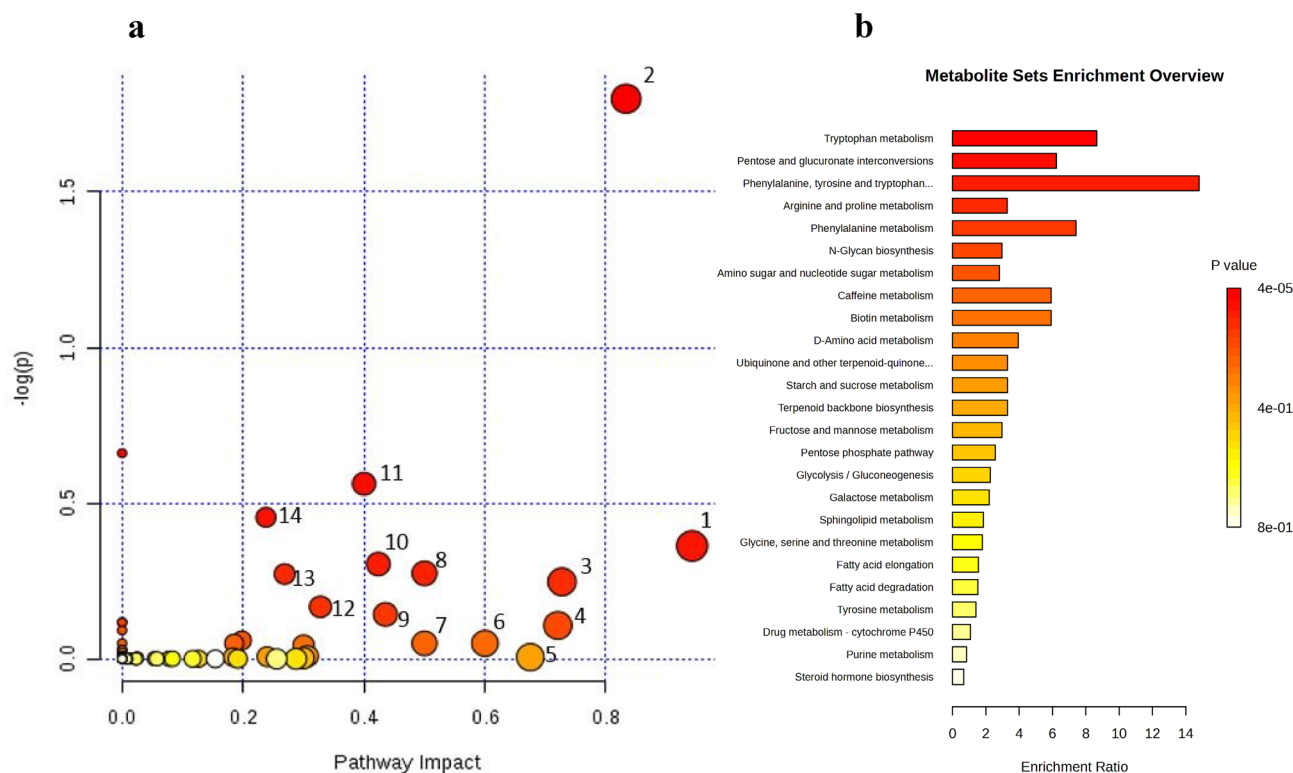


**Figure 6.** Overview of pathways in the *Brassica napus*–*Leptosphaeria maculans* pathosystem. (a) Metabolic pathway analysis (MetPA) from positive ionization mode with the MetaboAnalystR 6.0 software package. All the matched pathways are displayed as circles. The color and size of each circle are based on p-value (darker colors indicate more significant changes of metabolites in the corresponding pathway) and pathway impact value (the larger size the more impact), respectively. The most impacted pathways are annotated. 1: Phenylalanine metabolism, 2: Tryptophan metabolism, 3: alpha-Linolenic acid metabolism, 4: beta-Alanine metabolism, 5: Isoquinoline alkaloid biosynthesis, 6: Tyrosine metabolism, 7: Pantothenate and CoA biosynthesis, 8: Sphingolipid metabolism, 9: Glyoxylate and dicarboxylate metabolism, 10: Limonene and pinene degradation. (b) Metabolite sets enrichment overview. The horizontal bar (white-to-red) reflects increasing statistical significance.

as host resistance-related specific pathways or pathogen stress-related specific pathways. Fatty acids (FA) metabolic pathways have significant roles in plant defense against pathogens. Previous studies on FAs reveal that they play passive roles in plant defense, serving as biosynthetic precursors for cuticular components or the phytohormone jasmonic acid. Studies of FA metabolic mutants demonstrated an active signaling role for the cuticle in plant defense<sup>31</sup>.

Our study showed that alpha-linolenic acid metabolism has significant effects in both positive and negative ionization modes (Figs. 6 and 7). Alpha-linolenic acid, as a plant hormone, responds to plant infection by pathogens, cooperates with other plant hormones to stimulate growth, adapt to stresses, and regulate metabolic pathways<sup>32</sup>. Its application enhances the activity of various antioxidant enzymes, including sodium dismutase, catalase, ascorbate peroxidase, NADH peroxidase, and glutathione reductase<sup>33</sup>. It is speculated that oxidation regulation of unsaturated fatty acids can be used for their long-distance transport in plants<sup>34</sup>. The metabolisms of 12-OPDA, 13(S)-HPOT, and alpha-linolenic acids were up-regulated in the LM-canola interaction at 120 hpi in the R cultivar under 21 °C. The 12-OPDA is a precursor for synthesizing jasmonic acid (JA), the increasing JA at 120 hpi is probably in response to the fungal necrotrophic stage. The 12-Oxo-9(Z)-dodecenoic acid strongly inhibited mycelial growth and spore germination of eukaryotic microbes<sup>35</sup>. Oxylipins have several functions including direct antimicrobial effect, stimulation of plant defense gene expression, and regulation of plant cell death<sup>36</sup>.

The abundance of palmitic acid (hexadecanoic acid) a precursor for JA<sup>37</sup>, was significantly reduced in both R and S plants inoculated with the pathogen LM compared to their control at 48 hpi at 21 °C. This reduction is probably related to suppressing the production of JA and increasing the expression of salicylic acid (SA) synthesis. The increase of palmitic acid at 120 hpi suggests higher demands for the production of JA. Palmitic acid also affects the pathways of fatty acid biosynthesis, fatty acid elongation in mitochondria, and fatty acid metabolism. The study by Šašek et al. revealed that the expression of multiple marker genes linked to different plant hormones was elevated in resistant and susceptible *B. napus* plants infected with LM<sup>38</sup>. Additionally, the study revealed that the expression of genes related to the ET pathway increased at later stages of the infection (8–10 dpi), while genes related to SA were induced earlier (4–6 dpi)<sup>38</sup>. Similarly, the study by Lowe et al. (2014) demonstrated that the marker genes for SA and ABA were up-regulated in *B. napus* inoculated with LM at 7 and 14 dpi<sup>39</sup>. Haddadi



**Figure 7.** Overview of pathways in *Brassica napus*–*Leptosphaeria maculans* pathosystem. **(a)** Metabolic pathway analysis (MetPA) from negative ionization mode with the MetaboAnalystR 6.0 software package. All the matched pathways are displayed as circles. The color and size of each circle are based on p-value (darker colors indicate more significant changes of metabolites in the corresponding pathway) and pathway impact value (the larger the value, the more impact), respectively. The most impacted pathways are annotated. 1: Alanine, aspartate, and glutamate metabolism, 2: Phenylalanine metabolism, 3: Tyrosine metabolism, 4: Tryptophan metabolism, 5: alpha-Linolenic acid metabolism, 6: Riboflavin metabolism, 7: Isoquinoline alkaloid biosynthesis, 8: Stilbenoid, diarylheptanoid, and gingerol biosynthesis, 9: Glyoxylate and dicarboxylate metabolism, 10: Citrate cycle (TCA cycle), 11: Indole alkaloid biosynthesis, 12: Phenylpropanoid biosynthesis, 13: Porphyrin and chlorophyll metabolism, 14: Pantothenate and CoA biosynthesis. **(b)** Metabolite sets enrichment overview. The horizontal bar (white-to-red) reflects increasing statistical significance.

No	Name	Hits	P-value
1	Alanine, aspartate and glutamate metabolism	11	3.41e-7
2	Valine, leucine and isoleucine biosynthesis	6	0.00000143
3	Phenylalanine metabolism	5	0.000045
4	Arginine biosynthesis	6	0.000112
5	Amino sugar and nucleotide sugar metabolism	9	0.000898
6	Phenylalanine, tyrosine and tryptophan biosynthesis	3	0.000949
7	Citrate cycle (TCA cycle)	6	0.00106
8	Arginine and proline metabolism	8	0.00137
9	Tryptophan metabolism	8	0.00331
10	Butanoate metabolism	4	0.0121
11	Nicotinate and nicotinamide metabolism	4	0.0121
12	Glyoxylate and dicarboxylate metabolism	6	0.0132
13	Cysteine and methionine metabolism	6	0.0153
14	Galactose metabolism	5	0.0246
15	Fructose and mannose metabolism	4	0.0337

**Table 2.** Pathways with significant ( $p < 0.05$ ) high impact in *Brassica napus*–*Leptosphaeria maculans* pathosystem based on the KEGG global metabolic network with the MetaboAnalystR 6.0 software package.

et al. provided a more detailed picture of the global transcriptome that is active in *B. napus* during cotyledon infection by LM. The gene expression profile associated with different plant hormones revealed the importance of SA at the earlier stages of infection (until 4 dpi) and the up-regulation of genes related to the JA pathway at the later stages (6–8 dpi)<sup>40</sup>. SA is more effective in defending against biotrophic and hemibiotrophic pathogens, while ET/JA signaling is more capable in resisting against necrotrophic pathogens and herbivorous insects. Gene-for-gene interaction studies in the *B. napus*–*L. maculans* pathosystem showed early induction of SA/JA responsive factors, indicating the importance of those genes in the incompatible interaction. The early activation of SA-responsive factors in resistant *B. napus* genotypes guarantees effective slowdown of the pathogen infection due to the reinforcement of the defense throughout the plant via the associated SAR (systemic acquired resistance). Each hormone has certain responsive factors and signaling pathways, where the responsive pathways of different hormones also have different potential connections, building up an integrated and systemic signaling network to cope with various challenges<sup>41</sup>. SA–JA relationship is not always antagonistic. Tamaoki et al. explained that the defense system is activated by both SA and JA signaling interaction during the induction of defense response<sup>42</sup>. There may be at least a partly shared signal transduction pathway used for signaling both JA and SA.

The increased activity of nicotinate and nicotinamide metabolic pathway in the R cultivar compared to the control at 48 hpi under 21 °C (48 hpi/21 °C) might be considered as an indication of a greater supply of these two vital precursors for the generation of coenzymes, NAD<sup>+</sup> and NADP<sup>+</sup>, which are crucial for redox reactions and electron transportation from one reaction to another. These coenzymes are essential for many metabolic pathways including the TCA cycle, glycolysis, fatty acid biosynthesis, pentose phosphate cycle, and many other metabolic pathways. Among these redox carriers, NAD is the keystone of cellular oxidations along catabolism and is essential for plant growth and development<sup>43</sup>. In addition to its redox role, there is compelling evidence that NAD is a signal molecule controlling crucial functions in primary and secondary carbon metabolism. Studies using integrative omics approaches combined with molecular plant pathology have shown that manipulating NAD biosynthesis and recycling led to an alteration of metabolite pools and developmental processes and changes in resistance to various pathogens<sup>43</sup>. NAD levels could be used as a potential target to improve tolerance to biotic stress and crop improvement.

The increased histidine metabolism in the R cultivar compared to control at 48 hpi/21 °C suggesting the importance of histidine in host resistance since histidine biosynthesis pathway is integrated with several metabolic pathways, including tryptophan. Tryptophan plays a significant role in regulating plant development and defense responses and is the precursor for indoleacetic acid, a plant hormone necessary for cell expansion<sup>44</sup>. Liquid fertilizer originated from a yeast cell extract containing histidine exerted inhibitory effects on the development of bacterial wilt disease in tobacco plants caused by the soil-borne bacterial pathogen *Ralstonia solanacearum*<sup>45</sup>.

An increase in the abundance of galactose metabolism at 48 hpi compared to control in the R cultivar at 21 °C showed the importance of the central carbohydrate metabolism as one of the key sources for protecting cells from stresses<sup>46</sup>. Galactose has a defensive function and is involved in synthesizing hemicellulose<sup>47</sup>. Sugars play a crucial role in the interaction between plants and pathogens, serving as a vital energy source for defense and functioning as signaling molecules for the regulation of defense genes<sup>48</sup>. The concept of “Sweet Immunity” and sugar-enhanced defense has emerged due to the potential involvement of certain sugars in plant immunity<sup>47–49</sup>. Our network analysis and Enrichment analysis also identified galactose metabolism as an important pathway. The elevated levels of aminoacyl-tRNA biosynthesis in the R cultivar compared to control at 48 hpi/21 °C support previous study that the function of aminoacyl-tRNA synthesis is to precisely match amino acids with tRNAs containing the corresponding anticodon<sup>50</sup>.

### Non-specific stress-related pathways

The network analysis and enrichment analysis identified the importance of phenylalanine metabolism, tryptophan metabolism, beta-Alanine metabolism, isoquinoline alkaloid biosynthesis, tyrosine metabolism, pantothenate and CoA biosynthesis, sphingolipid metabolism, glyoxylate, and dicarboxylate metabolism as general or non-specific stress-related pathways. Phenylalanine metabolism and tyrosine metabolism as non-specific stress-related pathways were increased in all treatments compared to control at 21 °C and in resistance cultivar at 48 hpi under the elevated temperature regime. Tyrosine metabolism was up-regulated in soybean at 1 and 2 dpi in response to *R. solani*, which is consistent with our findings<sup>51,52</sup>. Tyrosine can undergo catabolism, resulting in the production of diverse secondary metabolites such as tocopherols, tocotrienols, and tyramine. These metabolites serve as precursors for the synthesis of hydroxycinnamic acid amides that are linked to the cell wall<sup>53</sup>. Phenylalanine, tyrosine, and tryptophan, as precursors for a wide variety of secondary metabolites, play a critical role in plant defense against biotic stressors and resistance to pathogen infections<sup>54</sup>. It has been hypothesized that tyrosine decarboxylated precursors (tyramine and dopamine) are used in the synthesis of isoquinolines in plants due to the up-regulation of the isoquinoline alkaloids pathway. Historically, the physiological activity of these metabolites has a role in plant defense systems, either by a direct toxic impact or by suppressing certain processes of invading pathogens<sup>55,56</sup>.

Pantothenate (vitamin B5) is the precursor to coenzyme A (CoA), which in turn, plays an essential role in fatty acid and pyruvate metabolism<sup>57</sup>. The main function of vitamins is to act as a cofactor in diverse metabolic pathways, facilitate the production of essential compounds for plants, induce resistance against pathogens, directly promote plant growth, and participate in energy conservation in the plant from stored compounds<sup>57</sup>. This is supported by the results that sphingolipid metabolism was increased at 120 hpi compared to control in the R cultivar under both optimum/elevated temperature regimes. Sphingolipids could act as second messengers with effects on cellular homeostasis.

### Heat stress-related specific pathways

Carotenoid biosynthesis, pyrimidine metabolism, and lysine biosynthesis were classified as heat stress-specific related pathways. This is supported by the results that carotenoid biosynthesis pathway in pathogen-inoculated plants of the resistant cultivar compared to those of mock up-regulated at 120 hpi/28 °C but down-regulated at 120 hpi/21 °C. The functional metabolism of carotenoids encompasses various processes, such as the stability of membrane lipid bilayers, the elimination of free radicals induced by reactive oxygen species, and the safeguarding of membrane lipid integrity<sup>58,59</sup>. The plant hormones abscisic acid and strigolactones as well as other apocarotenoids that are involved in different developmental processes and stress responses, are all synthesized from carotenoids<sup>60</sup>. Quan et al. revealed that glutamic acid and poly- $\gamma$ -glutamic acid boosted the heat resistance of Chinese cabbage (*Brassica rapa* L. ssp. *Pekinensis*) by refining carotenoid biosynthesis, photosynthesis, and ROS Signaling<sup>61</sup>. Exogenous application of biostimulants, such as carotenoids, can alleviate the damage caused by elevated temperatures in canola and improve growth parameters and physiological characteristics.

Nucleotide metabolism is a crucial component of plant metabolism and affects many metabolic pathways. Specifically, pyrimidine nucleotides participate directly in nucleic acid synthesis<sup>52,62</sup>. The observed up-regulation of the pyrimidine metabolism pathway at 28 °C, with the increase in this pathway more pronounced in the S cultivar than in the R cultivar, suggests that genes of the pyrimidine catabolic pathway are important in plant abiotic stress responses. The role of pyrimidine catabolism in stress tolerance may be related to the production of proline biosynthesis precursors<sup>62</sup>. In plants, a complete characterization of this pathway is lacking<sup>63,64</sup>. The accumulation of purine and pyrimidine bases in R cultivars at 48 hpi/28 °C might be interpreted as the plant's reaction to mixed pathogen and heat stress. Furthermore, pyrimidine derivatives have broad-spectrum biological activities, including antibacterial, antifungal, insecticidal, herbicidal, and antiviral activities<sup>65</sup>.

The lysine biosynthesis was increased at 48 and 120 hpi compared to the control in both R and S cultivars under elevated temperatures, which suggests the importance of lysine biosynthesis in plant heat stress response. Lysine is an important precursor for the synthesis of glutamate, which is an essential final energy source for the production of carbohydrates, lipids, peptides, and secondary metabolites. Lysine levels increase under abiotic stress, such as salt, water, heavy metals, and nutritive stress<sup>66</sup>. Endogenous lysine metabolism or its exogenous application helps reduce abiotic stress<sup>67</sup>. It has also been suggested that the jasmonate signaling pathway and tryptophan metabolism through stress responses are induced by lysine metabolism<sup>68</sup>.

### Global metabolic network

KEGG mapping of the global metabolic network displayed enriched biosynthetic pathways for the biosynthesis of phenylalanine, tyrosine, tryptophan, pantothenate and CoA, sphingolipid, valine, leucine, isoleucine, arginine, proline, caffeine, biotin, glycan, starch and sucrose and galactose, fructose and mannose, ubiquinone and terpenoids-quinones, as well as selenocompounds (Figs. 6 and 7). A significant number of these pathways are related to amino acid metabolism, which serves as a common source for secondary metabolites such as phenolics and alkaloids<sup>69</sup>. Enrichment of amino acid metabolism is essential for influencing metabolic processes after pathogen contact. Pantothenic acid, a precursor for some cofactors, is crucial for regulating enzyme activity and hence influencing plant development via metabolic pathways. Selenocompounds protect plants from a variety of abiotic stresses<sup>70</sup>. Metabolites that enhance the ubiquinone pathway can act as electron transporters in the process of photosynthesis<sup>71</sup>. Caffeine, classified as a xanthine alkaloid, may protect the young and soft tissues from infections caused by pathogens<sup>72</sup>. Glycans function as signaling molecules in stress and plant defense responses, as well as being involved in the energy metabolism of a plant<sup>73</sup>. In summary, the enrichment analysis showed a significant presence of metabolite hits in many pathways. The prevalence of these hits suggests that plant responses are influenced in terms of defense, resistance, and/or survival.

Network analysis provides a clear snapshot of the pathways in which the annotated metabolites are engaged, enhancing our knowledge of their significance in these processes<sup>74</sup>. Network analysis of different pathways in canola cotyledons in response to *L. maculans* infection identified 15 significant pathways (Table 2) that are mostly associated with the production of amino acids and subsequently carbohydrates. Cardoso et al. performed a meta-analysis on a wide range of species and various stress circumstances such as water deficit, cold, salt, and nitrogen deprivation stress<sup>75</sup>. Their findings emphasized that amino acid metabolism plays a vital role in plant stress adaptation. Amino acids are not the only source of carbon and nitrogen for different pathways but also good stress signal messengers that aid the entire plant in acclimating to stress conditions<sup>75</sup>.

The specific outcomes of disease severity when a plant is simultaneously challenged by an invading pathogen and heat stress, depend on many factors, including host and pathogen genetics. Most studies have described negatively impacted host resistance under elevated temperatures, while others noticed positive or neutral effects of elevated temperatures on host resistance. Development of strategies to protect food security under global warming will require an understanding of how plants deal, both positively and negatively, with simultaneous abiotic and biotic stresses<sup>76</sup>. Our results obtained from *B. napus*-*L. maculans* (LM) interaction suggest that the elevated temperatures might negatively affect the resistance to LM. However, this work was done using canola cotyledons and the effects of temperature on resistance to LM might be different at various canola growth stages. There is a need to further investigate the effects of temperature on the effectiveness of *Rlm7* at different canola growth stages. One of the most effective approaches to developing crops that are adaptable to climate change is to understand the diverse plant immune mechanisms impacted by temperature and identify key genes that can enhance disease resistance through genome editing techniques. This study identified important alterations in a diversity of metabolite classes, suggesting an infection-related reprogramming of particular pathways. Our results indicate the importance of hierarchical regulation of multidimensional metabolic networks in the face of several concurrent challenges, such as pathogen and heat stress. To identify other mechanisms in physiological

processes under biotic and abiotic stresses, further research is needed using RNA sequencing, proteomics, and metabolomics with other platforms.

## Conclusion

This work profiles the metabolisms of canola cotyledons at the initial infection stage (biotrophic stage) and the necrotrophic stage, in both compatible and incompatible (with the resistance gene *Rlm7*) interactions at normal (21 °C) and elevated (28 °C) temperatures using UPLC-QTOF/MS. Resistance-related specific pathways, heat stress-related specific pathways, and pathways associated with combined heat and pathogen stresses have been identified. The identified pathways associated with the breakdown of resistance at elevated temperatures, such as carotenoid biosynthesis, pyrimidine metabolism, and lysine biosynthesis will play an important role in developing climate-resilient crops by using effective techniques to manipulate these pathways. However, to fully understand the multidimensional nature of plant-pathogen interactions and produce disease-resistant cultivars that are resilient to climate change, there is a need for future research investigating a wider range of environmental conditions.

## Materials and methods

### Plant growth and infection conditions

Two canola cultivars (Drakkar—susceptible without any resistance genes and Excel—resistant carrying the resistance gene *Rlm7*) and the LM isolate EM24 carrying three known effectors (*AvrLm1*, *AvrLm6*, and *AvrLm7*), were used in this study. Before planting, seeds were distributed in a 9-cm-diameter sterile Petri dish with one layer of saturated filter paper. The petri dish was placed in an incubator at 20 °C ± 1 with 45% relative humidity, for two days. The germinated seeds were sown in plastic pots containing pasteurized soil (1:3 mixture of peat and soil). Pots were maintained in a growth chamber under controlled environmental conditions at two different temperatures: 21 °C (light)/16 °C (dark) and 28 °C (light)/23 °C (dark), with 75% relative humidity and 16 h photoperiod with 13,000 lx light intensity at canopy level.

After 14 days, the cotyledons of individual seedlings were wounded, using a sterile needle at each of the four lobes. Each of the wounded sites was inoculated with a 10 µl drop of conidial suspension or sterile distilled water (control). Spore suspensions of *L. maculans* were prepared from cultures grown on V8-agar. The fungal inoculum was prepared according to Chen and Fernando<sup>77</sup>. The concentration of spores was diluted to a final spore concentration of 1 × 10<sup>7</sup> spores mL<sup>-1</sup>. Each treatment included three biological replicates (each replicate consists of five pots with five plants per pot). The treatments were arranged in a completely randomized design. At 48, 120 h post-inoculation (hpi), 300 mg of whole cotyledon tissues per time point, corresponding to 3 cotyledons (3 cotyledons from 3 plants with one cotyledon from each plant), were randomly collected for metabolomics analysis from pathogen- and mock-inoculated plants. The leaf samples were shock-frozen in liquid nitrogen and stored at -80 °C until processing. The rest of the seedlings were kept under the same growth conditions and disease progression was monitored.

### Disease severity assessment

After sampling cotyledon tissues for metabolite analysis, the remaining plants were kept in the growth chamber for disease symptom evaluation up to 14 dpi in 3-day intervals by rating the infected cotyledons on a 0 to 9 scale (where 0 = no symptom and 9 = tissue collapse and appearance of conidia)<sup>78</sup>. The 0–9 original ordinal score of disease severity data was converted to a ratio scale normalized to the 0–1 range using the following equation:

$$y = (\sum [(0 \times a) + (1 \times b) + (2 \times c) + (3 \times d) + (4 \times e) + (5 \times f) + (6 \times g) + (7 \times h) + (8 \times i) + (9 \times j)])/(n \times 9)$$

where  $y$  = blackleg severity on a ratio scale,  $i$  to  $j$  are the number of infected canola plants scored between 0 and 9, respectively according to their disease severities and  $n$  is the total number of plants scored. The normalized blackleg severity ( $y$ ) values over time were used to construct a disease progress curve. The disease severity over time was subsequently reduced to a single summary variable by calculating the area under the disease progress curve (AUDPC) based on the following formulae:

$$AUDPC = \sum_{i=1}^{n-1} \left( \frac{y_i + y_{i+1}}{2} \right) (t_{i+1} - t_i)$$

where  $i$  is the number of individual observations,  $n$  is the total number of observations,  $t$  is time after inoculation (days) and  $y$  is the normalized blackleg severity. At 14 dpi, the lesions were digitally imaged and the areas of lesions were measured using ImageJ software<sup>79</sup>.

### Metabolite extraction and UPLC-QTOF/MS analysis

To prepare 200 ml of extraction mixture, 250 µL of 80% methanol–water was added to 40 mg freeze-dried samples that were already grounded into a fine powder using an electric mill. After the extraction solvent was added, the samples were sonicated for 15 min and subsequently centrifuged at a speed of 14,000 rpm for 10 min. Finally, supernatants were transferred to new 1.5 mL micro-tubes after filtration with a 0.2 µm micropore PTFE membrane<sup>54</sup>. For each sample, 5 µL was injected into an ultra-high performance liquid chromatography system (Thermo Dionex Ultimate 3000 LC) connected to a quadrupole time of flight detector (Bruker Compact™ QTOF-MS) with a heated-electrospray ionization source (ESI). Chromatographic separation was performed as described in Tortosa et al. using a binary gradient solvent mode consisting of 0.1% formic acid in water (solvent A) and acetonitrile (solvent B). The following gradient was used: 3% B (0–3 min), from 3 to 25% B (3–10 min),

from 25 to 80% B (10–18 min), from 80 to 100% B (18–22 min), and to hold 100% B until 24 min. The flow rate was established at 0.3 ml/min and the column temperature was controlled at 35 °C<sup>80</sup>.

MS data were acquired using an acquisition rate of 2 Hz over the mass range of 50–1200 m/z. Both polarities ( $\pm$ ) ESI mode were used under the following specific conditions: gas flow 8 l/min; nebulizer pressure 38 psi; dry gas 7 l/min; dry temperature 220 °C. Capillary and endplate offsets were set to 4500 and 500 V, respectively. To monitor the performance of data acquisition, the run sequence was started with 3 blanks (methanol, the solvent used in sample extraction), and a standard compound (triphenyl phosphate for positive ionization mode and chloramphenicol for negative ionization mode) followed by every 10 samples to ensure instrument drift was minimal<sup>81</sup>.

### Metabolite identification and quantification

Total ion chromatograms generated by LC/MS were imported straight into Data Analysis (Bruker) and then exported as mzXML using CompassXport (Bruker). Peaks were aligned and identified using the ProfileAnalysis 2.1 program<sup>82</sup>. The peaks were aligned using a 10-s, 8-mDa frame and the "split buckets with multiple features" setting. Buckets were created using the molecular characteristics option, which takes into consideration the retention duration, m/z isotopic pattern, and the charge state of the probable compounds. The investigation focused on metabolites with an average abundance larger than 30,000. The intensity of peaks that exhibit a larger change on RT was added by hand to the bucket table. Metabolites reported in commercial databases including METLIN, KEGG, Pubchem, HMDB, and Plant Metabolic Network (PMN) were used for tentative identification using accurate mass (accurate mass error (AME) = 5 ppm).

### Statistical analysis

Multivariate analysis including principal components analysis (PCA) and Partial least squares discriminant analysis (PLS-DA) was carried out using the online software Metaboanalyst 6.0 (2023). The data from each day under different temperatures were uploaded separately for analysis. The data was Pareto scaled to reduce the influence of intense peaks while emphasizing weaker peaks that may have more biological relevance. The corresponding loadings of intense peaks were reduced and loadings from weak peaks were increased due to Pareto scaling. PLS-DA is a supervised approach that aims to differentiate between classes (Y) in highly complex data sets, despite within-class variability in the observed variables (X), in other words, to find the metabolic differences between the inoculated groups and mock groups. PLS-DA models were cross-validated using  $R^2$  and  $Q^2$  parameters. The quality evaluation ( $Q^2$ ) and R-squared ( $R^2$ ) statistics give a quantitative measure of consistency between the anticipated and original data and provide estimates of the predictive power of the model, as well. The PLS-DA model utilizing the first principal component of VIP (variable importance in the projection) values paired with univariate analysis (Fold change) ( $P < 0.05$ ) to detect differentially expressed metabolites. By computation of log twofold change ( $\log_2$  FC (inoculated/mock)) for each metabolite in both genotypes (resistant and susceptible) and based on  $VIP^2$ , metabolites linked to resistance or susceptibility or heat stress were identified. In many studies, a VIP value  $> 2$  is a suitable threshold for feature selection; however, this cut-off depends on the number of variables employed<sup>83</sup>. To identify the metabolites responsible for discrimination among the metabolomic profiles, the variable importance in the projection (VIP) score was used to select features with the most significant contribution in a PLS-DA model. VIP scores are a weighted sum of PLS weights for each variable and measure the contribution of each predictor variable to the model. Further, the VIP statistic summarizes the importance of the metabolites in differentiating the time points in multivariate space. Metabolites exhibiting high VIP scores ( $\geq 2$ ) are the more influential variables.

### Metabolic pathway analysis

Metabolite set enrichment analysis (MSEA) using metabolites significantly altered was conducted to identify the altered metabolic pathways (metabolite sets) associated with phenotypes of interest. Metabolomics Pathway Analysis (MetPA) was used for the analysis and visualization of metabolomics data within the biological context of metabolic pathways according to KEGG and HMDB metabolic databases. Arabidopsis metabolic pathway databases as references were used for pathway analysis. The final result is a ranking of the relative importance of pathways in the KEGG and HMDB databases together with their associated p-values from enrichment analysis<sup>82,84</sup>. Network analysis based on the KEGG global metabolic network according to the KEGG and HMDB metabolic database was also done<sup>85</sup>. The enrichment analysis, pathway analysis, and network analysis were done using MetaboAnalystR 6.0 software package.

### Ethical approval

The study was conducted in accordance with relevant guidelines and legislation.

### Data availability

Data are available from the corresponding author upon reasonable request. The datasets used and/or analyzed during the current study are also available from the corresponding author on reasonable request or downloadable from Google Drive. All LC-Qtof MS files in mzXML format have been converted to two archive files (positive and negative ionization modes). The addresses of links: Positive mode ionization ([https://drive.google.com/file/d/1RXcMzDDkRy7QCCx\\_UrtK93FZq-PGmkQv/view?usp=drive\\_link](https://drive.google.com/file/d/1RXcMzDDkRy7QCCx_UrtK93FZq-PGmkQv/view?usp=drive_link)). Negative mode ionization ([https://drive.google.com/file/d/1Pz\\_yC6pBFnmt23qo3UksYRp4Ph-W8gKc/view?usp=drive\\_link](https://drive.google.com/file/d/1Pz_yC6pBFnmt23qo3UksYRp4Ph-W8gKc/view?usp=drive_link)).

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## Author contributions

H. Hamzehzarghani contributed in conceptualization, methodology, and visualization of the investigation, defining the general hypothesis, designing the experiments based on the hypothesis, supervising the research project throughout, analyzing and interpreting the results, administering the chemical and statistical analysis and writing the original draft of the main manuscript and revising, reviewing and/or editing of the text. Z. Amjadi performed the research throughout her Ph.D. project, contributed in analyzing the results, the chemical and statistical analysis, as well as revising, reviewing and editing of the text. V.M. Rodriguez helped us with resources and hyphenated instrumentation by providing UPLC-QTOF/MSm methodology, and formal analysis. Y. Huang helped with methodology and providing resources [two canola cultivars (Drakkar—susceptible without any resistance genes and Excel-resistant carrying the resistance gene Rlm7) and one Lm isolate EM24, which possesses three known effectors (AvrLm1, AvrLm6, and AvrLm7)] F. Farahbakhsh helped with chemical and statistical analysis.

## Competing interests

The authors declare no competing interests.

## Additional information

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