# The Effects of Short Work Versus Longer Work Periods Within Intermittent Exercise on VO2 Kinetics, Muscle Deoxygenation and Energy System Contribution 

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# THE EFFECTS OF SHORT WORK VERSUS LONGER WORK PERIODS WITHIN INTERMITTENT EXERCISE ON $\mathrm{V}_{2}$ KINETICS, MUSCLE DEOXYGENATION AND ENERGY SYSTEM CONTRIBUTION 

Thesis format: Integrated Article
by

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A thesis submitted in partial fulfillment of the requirements for the degree of Master of Science

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#### Abstract

The purpose of this study was to investigate the kinetic response of oxygen uptake $\left(\mathrm{V}_{2}\right)$ and muscle deoxygenation $(\mathrm{HHb})$ in males $(24 \mathrm{yr} \pm 3, \mathrm{n}=10)$ to identical heavy intensity continuous (CONT), and intermittent work; 25 s ( 25 s work: 3 s recovery (20W)), and $10 \mathrm{~s}\left(10 \mathrm{~s}\right.$ work: 3 s recovery (20W)), were compared. The $\tau \dot{\mathrm{V}} \mathrm{O}_{2}$ were similar in all conditions (CONT: $44.2 \mathrm{~s} \pm 9 ; 25 \mathrm{~s}: 38.9 \mathrm{~s} \pm 10 ; 10 \mathrm{~s}: 39.4 \mathrm{~s} \pm 8, \mathrm{p}>0.05$ ), whereas the $\mathrm{HHb} /$ $\dot{\mathrm{V}} \mathrm{O}_{2}$ overshoot decreased across conditions ( $\mathrm{p}<0.05$ ). The increased frequency of recovery periods slowed $\dot{\mathrm{V}}_{2}$ kinetics suggesting increased activity of the creatine kinase enzyme and increased ATP-PCr contribution, while reducing glycolytic and oxidative phosphorylation contributions to perform the identical work rate. Enhanced microvascular blood flow facilitated an accelerated matching of total $\mathrm{O}_{2}$ delivery to $\mathrm{O}_{2}$ utilization as well with increased recovery periods.


Keywords: $\mathrm{O}_{2}$ on-kinetics, near-infrared spectroscopy, intermittent exercise, recovery, energy systems contributions

## CO-AUTHORSHIP STATEMENT

This study was designed by M.C McCrudden ad G.R Belfry with input from the advisory committee (J.M Kowalchuk, D.H Paterson). The data were collected and analyzed by M.C McCrudden with the assistance of G.R Belfry. M.C McCrudden wrote the original manuscript for the study and G.R Belfry provided editoral feedback.

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## LIST OF ABBREVIATIONS

| $\mathrm{A}_{\mathrm{p}}$ | Amplitude |
| :---: | :---: |
| ADP | Adenosine diphosphate |
| ATP | Adenosine Triphosphate |
| CK | Creatine Kinase |
| CI95 | 95\% confidence intervalX |
| $\mathrm{CO}_{2}$ | Carbon dioxide |
| CONT | Continuous exercise |
| Cr | Creatine |
| DCA | Dichloroacetate |
| ETC | Electron transport chain |
| FADH2 | Reduced flavin adenine dinucleotide |
| $\mathrm{H}^{+}$ | Protons |
| Hb | Hemoglobin |
| $\mathrm{HbO}_{2}$ | Oxygenated hemoglobin |
| HHb | Deoxygenated hemoglobin |
| $\mathrm{HHb} / \mathrm{VO}_{2}$ | Normalized deoxygenated hemoglobin to oxygen uptake ratio |
| HI | High intensity exercise |
| HR | Heart rate |
| Hz | Hertz |


| INT | Intermittent exercise |
| :---: | :---: |
| Lac ${ }^{-}$ | Lactate |
| Mb | Myoglobin |
| min | Minute |
| MOD | Moderate intensity exercise |
| $\mathrm{NAD}^{+}$ | Nicotinamide adenine dinucleotide |
| NADH | Reduced nicotinamide adenine dinucleotide |
| NIRS | Near-infrared spectroscopy |
| $\mathrm{O}_{2}$ | Oxygen |
| PCr | Phosphocreatine |
| PDH | Pyruvate dehydrogenase |
| $\mathrm{PCO}_{2}$ | Partial pressure of carbon dioxide |
| $\mathrm{PetO}_{2}$ | End-tidal partial pressure of oxygen |
| $\mathrm{PetCO}_{2}$ | End-tidal partial pressure of carbon dioxide |
| $\mathrm{P}_{\mathrm{i}}$ | Inorganic phosphate |
| $\mathrm{PO}_{2}$ | Partial pressure of oxygen |
| PPO | Peak power output |
| $\mathrm{QO}_{2 \mathrm{C}}$ | Convective oxygen delivery |
| $\mathrm{QO}_{2 \mathrm{D}}$ | Diffusive oxygen delivery |
| RER | Respiratory exchange ratio |


| RI | Ramp-incremental |
| :---: | :---: |
| S | Second |
| SC | Slow component |
| SD | Standard deviation |
| SS | Steady state |
| TD | Time delay |
| $\mathrm{VO}_{2}$ | Oxygen uptake |
| $\mathrm{VO}_{2}$ BSL | Baseline oxygen uptake |
| $\mathrm{VO}_{2 \mathrm{~m}}$ | Skeletal muscle oxygen uptake |
| $\mathrm{VO}_{2 \mathrm{p}}$ | Pulmonary oxygen uptake |
| $\mathrm{VO}_{2 \text { peak }}$ | Peak oxygen uptake |
| $\mathrm{VCO}_{2}$ | Carbon dioxide production |
| Ve | Ventilation |
| WR | Work rate |
| WR ${ }^{\text {d }} 60$ | Work rate associated with $60 \%$ of the difference between $\mathrm{VO}_{2}$ at lactate threshold and $\mathrm{VO}_{\text {2peak }}$ |
| 10s:3s | 10 seconds work; 3 seconds recovery |
| 25s:3s | 25 seconds work; 3 seconds recovery |
| [] | Concentration |
| $\Theta_{L}$ | Lactate threshold |
| $\mu \mathrm{M}$ | Micro-molar |


| $\mathrm{X}^{2}$ | Chi-squared |
| :--- | :--- |
| $\tau$ | Tau (time constant) |

## CHAPTER 1

## 1 REVIEW OF LITERATURE

### 1.1 INTRODUCTION

The effects of short work: short recovery intermittent exercise has been the objective of investigators for many years originating with the pilot work performed by Astrand et al., (1960), and Christensen et al., (1960). The duration of the work : recovery periods may have quite different profiles $5 \mathrm{~min}: 2 \mathrm{~min}$ (Fox et al., 1975); 30 s : 30 s (Astrand et al., 1960) and $10 \mathrm{~s}: 5 \mathrm{~s}$ (Christensen et al., 1960). The latter two studies focused on the effect of the recovery periods on the required oxygen uptake $\left(\mathrm{V}_{\mathrm{V}}^{2}\right)$ to perform an identical work rate. They suggested that a greater contribution from substrate level phosphorylation accompanied the decrease in $\dot{\mathrm{V}} \mathrm{O}_{2}$ observed, thus reflecting a reduced oxidative phosphorylation to perform similar work within intermittent exercise.

With the further development of newer techniques and appropriate statistical power and alpha levels, insight into intermittent exercise and investigation into the postulated increased contribution from substrate level phosphorylation became possible. Complementing the work performed by Christensen \& Saltin, (1960), the effects of short 5 s recovery periods on 10 s work and the accompanying substrate level contributions were investigated by Belfry et al., (2012a; 2012b). The authors concluded that indeed the contribution of substrate level phosphorylation increased as the $\dot{\mathrm{VO}}_{2}$ requirement decreased as increased production of protons $\left(\mathrm{H}^{+}\right)$from both phosphocreatine resynthesis and glycolytic phosphorylation were observed.

Furthermore, Belfry et al., (2012b) observed a reduced in mean oxidative phosphorylation during 10 s work : 5 s recovery exercise bout compared to a continuous exercise bout of the identical work rate.

The work of Belfry et al., (2012a; 2012b) provided further insight into the physiological changes accompanying 5 s of recovery. However these studies have not yet elucidated the effects that these short recovery periods have on the rate of adjustment of oxygen uptake $\left(\dot{\mathrm{VO}}_{2}\right)$ and muscle deoxygenation $(\mathrm{HHb})$ to the reduced steady state (SS) $\dot{\mathrm{VO}}_{2}$ and HHb . The aim of the present study was to elucidate the effects of a further shortening of the recovery period ( 5 s to 3 s ) on $\dot{\mathrm{VO}}_{2 \mathrm{p}}$ and HHb and associated kinetics. Furthermore, an additional intermittent protocol utilizing an increased work period duration ( 25 s work: 3 s recovery) may determine if the responses of these variables are altered with longer work periods.

### 1.2 PULMONARY OXYGEN UPTAKE KINETICS

On the transition from a light intensity baseline work rate (WR) to one of a greater intensity, there is a simultaneous increase in the demand for adenosine triphosphate (ATP) within the working muscle (H. B. Rossiter et al., 1999). This increase in ATP provision occurs via oxidative and substrate level phosphorylation. During the first seconds of exercise onset the majority of ATP re-synthesis is provided via degradation of intramuscular phosphocreatine stores ( PCr ), and glycolysis. As duration of the work increases the contribution from substrate level phosphorylation is supplemented by an increase in oxidative phosphorylation. This increase in oxidative
phosphorylation can be measured at the mouth and the actual pulmonary $\dot{\mathrm{V}}_{2}\left(\dot{\mathrm{VO}}_{2 \mathrm{p}}\right)$ can be calculated (Lamarra et al., 1987).

During exercise in the moderate (MOD) and heavy intensity (HI) domain the kinetic response to the given exercise intensity determines the " $\mathrm{O}_{2}$ deficit", or the use of anaerobic ATP stores in order to sustain exercise until aerobic production can maintain production (Scheuermann \& Barstow, 2003). The faster the kinetic response of $\dot{\mathrm{V}} \mathrm{O}_{2}$, the smaller the corresponding $\mathrm{O}_{2}$ deficit that accompanies the exercise that needs to replenished after exercise (Scheuermann \& Barstow, 2003).

The rate of adjustment of $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}\left(\tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}\right)$ can be described by a mono-exponential response to the increase in ATP demand of MOD exercise which can be separated into three phases (Whipp et al., 1982). The cardio-dynamic (Phase I) defines the initial increase in $\dot{\mathrm{V}}_{2 \text { p }}$ indicative of the temporal delay ( $\sim 20 \mathrm{~s}$ ) between the increase in pulmonary perfusion due to immediate increase in cardiac output, and the deoxygenated blood arriving at the lung from the working muscles. This is followed by a larger, monoexponential increase in $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ that projects towards the new steady state $\left(\dot{\mathrm{V}}_{2 \mathrm{psS}}\right)$ associated with the ATP demand termed Phase II (B J Whipp et al., 1982). The Phase II response has been accepted to indicate oxygen uptake at the muscle $\left(\dot{\mathrm{V}}_{2 \mathrm{~m}}\right)$ (Bangsbo, 2000; Grassi et al., 1996; Krustrup et al., 2009; Rossiter et al., 2002; Rossiter et al., 1999; Whipp et al., 1982). This phase is associated with deoxygenated blood (due to $\mathrm{O}_{2}$ utilization at the muscle) arriving at the lung. The ATP demand for the given WR (below lactate threshold ( $\left.\hat{\theta}_{\mathrm{L}}\right)$ ) dictates the amplitude of the corresponding SS of $\dot{\mathrm{VO}}_{2 \mathrm{p}}$ (Phase III) (Whipp et al., 1982). This is the appropriate model for WR in MOD,
however when exercising above the $\hat{\theta}_{\mathrm{L}}$ in the HI there is a further evolution of $\mathrm{O}_{2}$ requirement reflected by a second exponential, the slow component (SC), that delays the onset of SS (Barstow \& Molé, 1991; Casaburi et al., 1989; Ozyener et al., 2001; Paterson \& Whipp, 1991; Rossiter et al., 2002; Xu \& Rhodes, 1999). The SC is the observed increase in $\dot{\mathrm{V}}_{2 \mathrm{p}}$ above that the predicted by the $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}} /$ WR relationship for work below the $\hat{\theta}_{\mathrm{L}}$ (Barstow \& Mole, 1991; Paterson \& Whipp, 1991; Ozyener et al., 2001).

The kinetic response of Phase II $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ can be described by the time constant $\tau$. This represents $63 \%$ of the temporal relationship between baseline $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ and that required during the $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{pss}}$. At the onset of MOD exercise in healthy young adults, $\tau$ ranges from $20-35 \mathrm{~s}$ (Barstow et al., 1994; DeLorey et al., 2004b). However $\tau$ within the HI domain is controversial. Some have observed the kinetics response to be similar to MOD (Barstow \& Molé, 1991; Ozyener et al., 2001; Scheuermann \& Barstow, 2003) whereas others have observed a slowing of $\tau \dot{\mathrm{V}}{ }_{2 p}$ (Jones et al., 2002; Koppo et al., 2004; Paterson \& Whipp, 1991). This slowing of $\tau$ during HI exercise has been attributed to a limitation in $\mathrm{O}_{2}$ delivery during HI exercise (Hughson et al., 2001; Grassi et al., 2000), the intrinsic slowness of skeletal muscle oxidative metabolism as ATP demand increases (Grassi, 2000), and differing muscle fiber recruitment (Barstow et al., 1996). This suggests that the introduction of short recovery periods, which lowers the mean WR will speed $\tau \dot{\mathrm{V}} \mathrm{O}_{2}$ compared to continuous (CONT) exercise performed at the same work rate as the intermittent work periods.

### 1.3 EFFECTS OF OXYGEN DELIVERY LIMITATIONS ON $\dot{\text { V }} \mathbf{O}_{2}$ KINETICS

$\mathrm{O}_{2}$ is necessary for the aerobic production of ATP via the ETC within the mitochondria (Equ.1) (Murias et al., 2011).

## Equation 1. The stoichiometry oxidative phosphorylation

$$
\mathrm{NADH}+0.5 \mathrm{O}_{2}+\mathrm{H}^{+}+3 \mathrm{ADP}+3 \mathrm{Pi} \longrightarrow \mathrm{NAD}^{+}+\mathrm{H}_{2} \mathrm{O}+3 \mathrm{ATP}
$$

Since there are limited stores of $\mathrm{O}_{2}$ within the muscle the exhaustion of these stores increase the reliance of $\mathrm{O}_{2}$ supply upon both convective delivery of $\mathrm{O}_{2}\left(\mathrm{QO}_{2 \mathrm{C}}\right)$ to the working muscles (Grassi et al., 1998a), and diffusive delivery, delivery of $\mathrm{O}_{2}\left(\mathrm{QO}_{2 \mathrm{D}}\right)$ to the mitochondria (Grassi et al., 1998b). The product of $\mathrm{O}_{2}$ delivery and the arterial venous $\mathrm{O}_{2}$ difference equals $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{~m}}\left(\mathrm{~V}_{2}\right.$ at the muscle), known as the Fick's Principle. Altering the convective and diffusive conditions has provided insight into possible rate limiting factors of oxidative phosphorylation (Grassi et al., 1998a; Grassi et al., 1998b). Within MOD, the use of hyperoxia $\left(\mathrm{PO}_{2}: 100 \%\right)$ to enhance peripheral $\mathrm{O}_{2}$ diffusion while maintaining elevated $\mathrm{QO}_{2 \mathrm{C}}$ in canine gastrocnemius showed no effect upon the speeding of $\tau \dot{\mathrm{V}} \mathrm{O}_{2}$ (Grassi et al., 1998b) suggesting that $\mathrm{QO}_{2 \mathrm{D}}$ does not have a limiting role. Similar works on both canine and human have shown that ensuring adequate $\mathrm{QO}_{2 \mathrm{C}}$ does not have an effect upon $\tau \mathrm{V}_{2}$ as there is no overshoot in fractional $\mathrm{O}_{2}$ extraction nor a slowing of $\tau \mathrm{V}_{2}$ within the MOD domain (Grassi et al., 1996; Grassi et al.,1998a; Nyberg et al., 2010). However, at the onset of HI exercise in the presence of elevated $\mathrm{QO}_{2 \mathrm{C}}$ a speeding in $\tau \mathrm{V}_{2}$ was observed in within canine subjects (Grassi et al., 2000). Furthermore, a speeding in $\tau \dot{\mathrm{V}} \mathrm{O}_{2}$ was also observed in studies utilizing human subjects
(Gerbino et al., 1996; Macdonald et al., 1997). Further support that within the HI domain $\mathrm{QO}_{2 \mathrm{C}}$ has a role in determining $\tau \dot{\mathrm{V}} \mathrm{O}_{2}$.

It has also been observed that recovery periods within INT exercise induced a $\sim 4.4$ fold increase in arterial blood velocity within four contraction : recovery cycles in the femoral artery (Rådegran \& Saltin, 1998). The initial rapid increase in blood velocity observed was attributed to muscle mechanical factors (i.e., muscle pump action). The effects of potent vasodilators (i.e., acetylcholine, nitric oxide, adenosine) did not induce similar changes in the femoral artery diameter within similar temporal constraints (Rådegran \& Saltin, 1998). The effect of similar recovery periods upon $\mathrm{QO}_{2 \mathrm{C}}$ within HI INT work and its subsequent effect on $\tau \mathrm{VO}_{2}$ have not been elucidated.

### 1.4 METABOLIC CONTROL OF $\boldsymbol{\tau} \dot{\mathrm{V}} \mathbf{O} 2 \mathrm{p}$

The fluctuations of short high intensity work ( $25 \mathrm{~s}: 10 \mathrm{~s}$ ) and shorter recovery (3 s) INT present unique bioenergetic challenges to the exercising muscles. These include repeated phosphorylation of ADP to perform work and repayment of the $\mathrm{O}_{2}$ deficit during recovery. The ATP required during the work and recovery of INT exercise originates via the CK reaction, and glycolytic and oxidative phosphorylation, thus demonstrating the integrated nature of metabolism during this intermittent exercise (Belfry et al., 2012a).

If availability of any of these substrates (i.e., $\mathrm{NADH}, \mathrm{ADP}, \mathrm{O}_{2}$ ) are limiting, the rate of adjustment of oxidative phosphorylation will be compromised and $\tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ will be
slowed (Murias et al., 2011). Mitochondrial respiration and its regulation are associated with the changes in ATP requirement at the working muscle during exercise (Rossiter et al., 1999). The relationship between the following reactions underpin the activation of mitochondrial activity:

Equation 2. Myosin ATPase

$$
\mathrm{ATP}+\mathrm{H}_{2} \mathrm{O} \longrightarrow \mathrm{ADP}+\mathrm{Pi}+\mathrm{H}^{+}
$$

Equation 3. Creatine Kinase reaction

$$
\mathrm{PCr}+\mathrm{ADP}+\mathrm{H}^{+} \longleftrightarrow \mathrm{Cr}+\mathrm{ATP}
$$

The first reaction (Equ. 2) is controlled by the enzyme Myosin ATPase, whereas the second reaction (Equ. 3) is controlled by the enzyme Creatine Kinase (CK) (Crowther et al., 2002; Kindig et al., 2005; Rossiter et al., 1999). During the on-transient phase, the immediate increase in ATP utilization causes subsequent elevation in intracellular ADP activating the CK reaction leading to the consumption of circulating ADP and resynthesis of ATP for its subsequent breakdown. As intramuscular phosphocreatine (PCr) stores become depleted, both ADP and Pi accumulate (Equ. 2). During short recovery periods some intramuscular Cr and ADP are phosphorylated (Newcomer et al., 1999). It has been suggested that activation of oxidative phosphorylation at the onset of exercise occurs through feedback control in response to increased ADP (Rossiter et al., 2002; Rossiter et al., 1999). Furthermore, the rate of PCr breakdown and re-synthesis has been found to closely match that of $\tau \dot{\mathrm{VO}}_{2 \mathrm{p}}$ (Rossiter et al., 2002; Rossiter et al., 1999). However, studies inhibiting the CK enzyme in both canine (Grassi et al., 2011) and human muscle (Glancy et al., 2008) have shown a speeding of $\tau \mathrm{VO}_{2}$. Conversely, this is
indicative of increased substrate phosphorylation of ADP accompanying increased CK reaction, which down regulates oxidative phosphorylation and slows $\tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$. Whether or not the inclusion of short recovery periods, and thus a greater reduction in ADP throughout INT exercise, has an effect on $\tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ has yet to be elucidated.

The production of reducing equivalents (NADH: nicotinamide adenine dinucleotide) occurs at PDH (pyruvate dehydrogenase) and within the Kreb's cycle (isocitrate dehydrogenase, alpha-ketoglutarate dehydrogenase, malate dehydrogenase). The NADH produced contributes to a proton gradient (via dehydrogenation of NADH) within the electron transport chain (ETC) which ultimately phosphorylates ADP. Increased PDH activity has been observed, simultaneously with increased WR, facilitating increased NADH production for increased ATP production via the ETC (Bangsbo et al., 2002; Howlett et al., 1998; Spriet et al., 2000). However, others have not observed a speeding effect of $\tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ with dichloroacetate (DCA) infusion, a PDH activator (Bangsbo et al., 2002; Grassi et al., 2002; Rossiter et al., 2003). This would suggest that a limitation in substrate provision inducing a slowing of $\tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ lies most downstream of PDH.

### 1.5 MUSCLE FIBRE RECRUITMENT AND $\dot{\mathbf{V}} \mathbf{O}_{2}$ KINETICS

Human skeletal muscle is comprised of three main fibre types: Type I - slow oxidative, Type IIa - fast twitch oxidative glycolytic, and Type $\mathrm{II}_{\mathrm{X}}$ - fast twitch glycolytic (Brooke \& Kaiser, 1970). Each fibre type has its own unique composition with microvascular density and mitochondrial content decreasing respectively. Type I fibres have the greatest mitochondrial content as they are primarily oxidative fibres as
well as the greatest microvascular density, whereas Type IIa fibres and Type IIx fibres have a decreasing mitochondrial content and density with Type IIx having the least (Brooke \& Kaiser, 1970). Previous research suggests that muscle fibre recruitment follows a hierarchy pattern with smaller lower threshold fibres recruited first, followed by larger higher threshold fibres as WR increases (Vollestad \& Blom, 1985).

Furthermore, it has been suggested that recruitment of less oxidative fibres at WR above $\hat{\theta}_{\mathrm{L}}$, particularly Type IIx fibres, are responsible for the development of the $\dot{\mathrm{V}}_{2} \mathrm{SC}$, delaying the onset of $\dot{\mathrm{V}} \mathrm{O}_{2 \text { SS }}$ (Poole et al., 1994; Whipp, 1994). The amplitude of the slow component within HI was observed to be negatively correlated with the per cent Type I fibre composition $(r=-0.83)$ (Barstow et al., 1996a). This supports Whipp (1994) and Poole et al., (1994) observations that fibre type composition correlates with the development of the SC. Consequently, a higher percentage of Type I fibres has been associated with a speeding of $\tau \dot{\mathrm{V}} \mathrm{O}_{2}(\mathrm{r}=-0.67)$ at the onset of HI exercise (Pringle et al., 2003a). Furthermore, a reduced $\dot{\mathrm{VO}}_{2}$ representative of an acute hypoxic environment within the muscle during exercise has been observed to increase Type IIx muscle fibre activity (Gollnick et al., 1974; Nuutinen et al., 1982), thus contributing to a slowed $\dot{\mathrm{V}} \mathrm{O}_{2}$ kinetic response.

As the recruitment of Type IIx muscle fibres is reliant upon WR, it would be expected that the reduction of mean WR with the increased frequency of 3 s recovery periods of the current study would slow $\tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ (Barstow et al., 1996; Pringle et al., 2003) as greater Type IIx fibre involvement has been indicated in INT (Belfry et al., 2012a). However the effect of short recovery periods within INT may have an effect
upon recruitment of specific motor units and muscle fibre activation eliciting a change in the kinetic response.

### 1.6 MUSCLE BIOENERGETICS DURING INTERMITTENT EXERCISE

ATP breakdown (Equation. 2, 1.2) leads to energy release for actin-myosin interactions to elicit muscle shortening, and for the energy required to phosphorylate ADP and Cr during recovery (Crowther et al., 2002). At the onset of HI exercise there is no appreciable change in ATP despite the high-energy requirement due to the simultaneous breakdown of PCr that maintains ATP levels via the CK reaction (Equation. 3, 1.2) (Kindig et al., 2005).

Recently there has been a body of work investigating the acute high-energy phosphate responses (Belfry et al., 2012a; Belfry et al., 2012b) to 10 s work: 5 s recovery periods compared to CONT exercise.

At the onset of work the requirement of ATP is greater than can the quantity that can be provided by oxidative phosphorylation. As oxidative metabolism is being upregulated, ATP is being produced via substrate level phosphorylation either through the aforementioned CK reaction as well as anaerobic glycolytic phosphorylation in the cytosol (Equ. 4).

Equation 4. Anaerobic Glycolytic Phosphorylation

$$
\begin{aligned}
& \text { Glucose }+2 \mathrm{ATP}+4 \mathrm{ADP}+2 \mathrm{Pi}+2 \mathrm{NAD}^{+} \longrightarrow \text { 2Pyruvate }+2 \mathrm{ADP}+4 \mathrm{ATP}+2 \mathrm{NADH}+ \\
& 2 \mathrm{H}^{+}+2 \mathrm{H}_{2} \mathrm{O}
\end{aligned}
$$

It has been suggested that anaerobic glycolytic phosphorylation contributes immediately post exercise for recovery of ATP as well (Belfry et al., 2012a; Burnley et al., 2006; Crowther et al., 2002; Forbes et al., 2009; Rossiter et al., 2002). Furthermore, it has been noted that a substantial resynthesis of $\mathrm{PCr}, \sim 10-20 \%$, occurs within very short 3 - 5 s recovery periods (Newcomer et al., 1999; Belfry et al., 2012a). Utilizing ${ }^{31} \mathrm{P}$ - NMR spectroscopy it has been observed that intramuscular PCr stores replenished $\sim 20 \%$ within 4 s recovery indicated by the increased $\mathrm{H}^{+}$content (Equation 3) accompanied by increased contribution via glycolytic phosphorylation, further increasing $\mathrm{H}^{+}$levels (Newcomer et al., 1999; Belfry et al., 2012a).

Increases in Pi, lactate and $\mathrm{H}^{+}$, and decreased PCr from ATP hydrolysis have been associated with heavy intensity constant load continuous exercise leading to increased metabolic stress and fatigue (Debold et al., 2004; Fitts, 2008; Rossiter et al., 1999). The literature presents uncertainty as to which of these variables has the strongest association with increased metabolic stress (Chance et al., 1980). The comparison of both INT exercise conditions to a CONT exercise performed at a similar work rate as the work periods of INT may differentiate these metabolites as the differences in average power output may require different handling of Pi , lactate and $\mathrm{H}^{+}$and PCr .

### 1.7 NEAR - INFRARED SPECTROSCOPY AND MUSCLE DEOXGENTATION

The Near-Infrared Spectroscopy (NIRS) technology is utilized to observe the changes in deoxyhemoglobin saturation $(\mathrm{HHb})$ within the working muscle. The resulting

HHb signal is indicative of the balance between $\mathrm{QO}_{2}$ and $\dot{\mathrm{V}} \mathrm{O}_{2}$ of the muscle under interrogation. Described below is the theoretical foundation for the NIRS technology.

The electromagnetic spectrum refers to the diverse collection of radiant energy considered as a wave or particle traveling at the speed of light. These waves differ from each other in length and frequency. Infrared refers to that part of the electromagnetic spectrum which falls between the visible and microwave regions. During infrared spectroscopy organic molecules, such as hemoglobin $(\mathrm{Hb})$ and myoglobin $(\mathrm{Mb})$, are exposed to infrared radiation (wavelength of $690-850 \mathrm{~nm}$ ). In doing so, NIRS can provide a measure of both skeletal muscle oxygenation $\left(\mathrm{HbO}_{2}\right)$ and deoxygenation (HHb) (Koga et al., 2011). This measurement is detected based on the absorption of the near-infrared (NIR) light projected by the diode refracted back to the optode from the organic tissue. The NIR spectrum is utilized due to its ability to penetrate the organic tissue for absorption by Hb and Mb and any $\mathrm{O}_{2}$ bound to these molecules will alter its absorption (McCully \& Hamaoka, 2000). The wavelength emitted by the NIRS diode provides insight into the HHb , and $\mathrm{HbO}_{2}$ content within the microvasculature by the difference in detected light by the optode (McCully \& Hamaoka, 2000). This measured difference occurs due to the variances in NIR absorption by HHb and $\mathrm{HbO}_{2}$ molecules which absorb light of distinct wavelengths 690-760 nm and $800-850 \mathrm{~nm}$, respectively (due to the presence/absence of $\mathrm{O}_{2}$ ) (McCully \& Hamaoka, 2000).

When the radiant energy matches the energy of a specific molecular vibration, absorption occurs. This reduces the light returning to the spectrophotometer. The NIRS information obtained from the muscle provides insight to the level of oxygenation at the microvascular level and allows for an estimate of $\mathrm{QO}_{2}$ and muscle $\dot{\mathrm{V}} \mathrm{O}_{2}$ (DeLorey et al.,

2003; Mancini et al., 1994). An observed decrease in HHb for a given WR and $\dot{\mathrm{VO}}_{2}$ suggests an increase in $\mathrm{O}_{2}$ provision. Conversely, an increase in HHb for a given $\dot{\mathrm{VO}}_{2}$ suggests a greater reliance on $\mathrm{O}_{2}$ extraction versus provision to meet the oxidative requirements (Grassi et al., 2003).

Within both HI and MOD changes in HHb are presumed to be indicative of changes in $\mathrm{O}_{2}$ extraction from the Hb (DeLorey et al., 2003a; Saitoh et al., 2009). The rate of adjustment of muscle deoxygenation $(\tau \mathrm{HHb})$ has been observed to be faster than that of $\tau \dot{\mathrm{V}} \mathrm{O}_{2}$ reflecting increased $\mathrm{O}_{2}$ extraction to meet the $\mathrm{O}_{2}$ demands of the working muscle (DeLorey et al., 2003; Grassi et al., 2003; Saitoh et al., 2009). A faster adjustment of $\tau \mathrm{HHb}$ suggests that microvascular blood flow distribution is inadequate for the metabolic $\mathrm{O}_{2}$ demands at the (Bangsbo, 2000; DeLorey et al., 2003; DeLorey et al., 2004a). It has been suggested that a hyperperfusion of the surrounding microvasculature of inactive tissue exacerbates this inadequacy (Saitoh et al., 2009). The speeding of $\tau \mathrm{HHb}$ observed within Type II diabetes mellitus has been attributed to vasodilatory dysfunction and its limiting effect on $\mathrm{QO}_{2}$ (Bauer et al., 2007). A similar speeding in $\tau \mathrm{HHb}$ has been observed in older adults who exhibit an impaired ability to redistribute blood flow from the periphery to exercising muscles (DeLorey et al., 2004a). Conversely, priming exercise has been observed to improve muscle blood flow distribution within HI exercise, which slows $\tau \mathrm{HHb}$ (Saitoh et al., 2009)

Within the present study it is possible that despite the brevity of the 3 s recovery periods, an improvement of microvascular blood flow distribution, as a function of the increased frequency of the 3 s recovery may elicit a slowing of $\tau \mathrm{HHb}$.

### 1.8 BREATH BY BREATH ANALYSIS BY MASS SPECTROSCOPY

The determinations of $\dot{\mathrm{V}} \mathrm{O}_{2} \mathrm{p}, \dot{\mathrm{VCO}}_{2}$ and ventilation rates were acquired with measuring the inspired and expired flow rates using a low-dead-space bidirectional turbine (calibrated before each test by using a syringe of known volume). Inspired and expired gases were sampled continuously at the mouth and analyzed for concentrations of $\mathrm{O}_{2}, \mathrm{CO}_{2}$, and $\mathrm{N}_{2}$ by mass spectrometry after calibration with precision-analyzed gas mixtures. Changes in gas concentration were aligned with gas volumes by measuring the time delay for a square-wave bolus of gas passing through the turbine to the resulting changes in fractional gas concentrations as measured by the mass spectrometer. Data was collected every 20 ms and transferred to a computer, which aligned concentrations with volume data to build a profile of each breath. Breath-by-breath alveolar gas exchange was calculated by using algorithms of Swanson et al., (1980).

These algorithms were developed to estimate breath-by-breath alveolar gas exchange by accounting for changes in both functional residual capacity and alveolar gas concentrations during ventilation. These corrections are computed to give accurate total lung gas exchange.

### 1.9 STUDY RATIONALE

Short work: shorter recovery durations may be present in a variety of sporting competitions. Sports such as hockey, basketball, and swimming involve such alternating bouts. Differing pool lengths in swimming dictate the work and recovery periods of the main propulsive muscles in freestyle swimming. These include the shoulder flexors and internal rotator muscles (Clarys 1985). When the swimmers change direction and push
off the wall these propulsive muscles are inactive for a period of approximately 3 s . In the United States the majority of swimming competitions take place in 50 m and 25 yd pools. International athletes may swim these lengths in $\sim 25 \mathrm{~s}$ and 10 s respectively. There are also open water swimming competitions in which the propulsive muscles are activated continuously. The comparison of the physiological responses to the differing durations of work associated with these swimming venues (CONT, 25 s and 10 s ) is the applied physiology objective of this study.

Therefore the primary purposes of this study were to contrast the effects of increasing the recovery frequency from CONT exercise to 25 s , and 10 s of work on both $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ and HHb kinetics, overall changes of $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ and HHb and blood lactate concentration. This will provide insight into the responses of $\mathrm{V}_{\mathrm{O}} \mathrm{O}_{2 \mathrm{p}}$ and HHb kinetics, $\mathrm{O}_{2}$ provision and utilization, and energy system contribution between each exercise condition

## REFERENCES:

Astrand, I., Astrand, P. O., Christensen, E. H., \& Hedman, R. (1960). Intermittent muscular work. Acta Physiologica Scandinavica, 48, 448-453. doi:10.1111/j.17481716.1960.tb01879.x

Bangsbo, J. (2000). Muscle oxygen uptake in humans at onset of and during intense exercise. Acta Physiologica Scandinavica, 168, 457-464. doi:10.1046/j.1365201X.2000.00697.x

Bangsbo, J., Bangsbo, J., Gibala, M. J., Gibala, M. J., Krustrup, P., Krustrup, P., ... Saltin, B. (2002). Enhanced pyruvate dehydrogenase activity does not affect muscle O2 uptake at onset of intense exercise in humans. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology, 282(1), R27380. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/11742848

Barstow, T. J., Buchthal, S., Zanconato, S., \& Cooper, D. M. (1994). Muscle energetics and pulmonary oxygen uptake kinetics during moderate exercise. Journal of Applied Physiology, 77(4), 1742-1749.

Barstow, T. J., Jones, a M., Nguyen, P. H., \& Casaburi, R. (1996a). Influence of muscle fiber type and pedal frequency on oxygen uptake kinetics of heavy exercise. Journal of Applied Physiology (Bethesda, Md.: 1985), 81, 1642-1650.

Barstow, T. J., Jones, a M., Nguyen, P. H., \& Casaburi, R. (1996b). Influence of muscle fiber type and pedal frequency on oxygen uptake kinetics of heavy exercise. Journal of Applied Physiology, 81(4), 1642-1650.

Barstow, T. J., \& Molé, P. a. (1991). Linear and nonlinear characteristics of oxygen uptake kinetics during heavy exercise. Journal of Applied Physiology (Bethesda, Md. : 1985), 71, 2099-2106.

Bauer, T., Reusch, J., Levi, M., \& Regensteiner, J. (2007). Skeletal Muscle Deoxygenation After the Slowed Microvascular Blood Flow Kinetics. Diabetes Care, 30(11), 2880-2885. doi:10.2337/dc07-0843.Abbreviations

Belfry, G. R., Raymer, G. H., Marsh, G. D., Paterson, D. H., Thompson, R. T., \& Thomas, S. G. (2012A). Muscle metabolic status and acid-base balance during 10-s work:5-s recovery intermittent and continuous exercise. Journal of Applied Physiology, 113(3), 410-417. doi:10.1152/japplphysiol.01059.2011

Belfry, G. R., Paterson, D. H., Murias, J. M., \& Thomas, S. G. (2012B). The effects of short recovery duration on VO2 and muscle deoxygenation during intermittent exercise. European Journal of Applied Physiology, 112(112), 1907-1915. doi:10.1007/s00421-011-2152-4

Brooke, M. H., \& Kaiser, K. K. (1970). Muscle fiber types: how many and what kind? Archives of Neurology, 23(4), 369-379. doi:10.1001/archneur.1970.00480280083010

Burnley, M., Doust, J. H., Jones, A. M., \& Time, A. M. J. (2006). O2 kinetics Time required for the restoration of normal heavy exercise V following prior heavy exercise. Journal of Applied Physiology, (101), 1320-1327. doi:10.1152/japplphysiol.00475.2006.

Casaburi, R., Barstow, T. J., Robinson, T., \& Wasserman, K. (1989). Influence of work rate on ventilatory and gas exchange kinetics. Journal of Applied Physiology (Bethesda, Md. : 1985), 67, 547-555.

Chance, B., Eleff, S., \& Leigh, J. S. (1980). Noninvasive, nondestructive approaches to cell bioenergetics. Proceedings of the National Academy of Sciences of the United States of America, 77(12), 7430-7434. doi:10.1073/pnas.77.12.7430

Christensen, E. H., Hedman, R., \& Saltin, B. (1960). Intermittent and Continuous Running. Acta Physiologica Scandinavica, 50, 269-286.

Clarys, J. P. (1985). Hydrodynamics and electromyography: ergonomics aspects in aquatics. Applied Ergonomics, 16(1), 11-24.

Crowther, G. J., Kemper, W. F., Carey, M. F., \& Conley, K. E. (2002). Control of glycolysis in contracting skeletal muscle. II. Turning it off. American Journal of Physiology. Endocrinology and Metabolism, 282(1), E74-E79.

Debold, E. P., Dave, H., \& Fitts, R. H. (2004). Fiber type and temperature dependence of inorganic phosphate: implications for fatigue. American Journal of Physiology. Cell Physiology, 287(3), C673-C681. doi:10.1152/ajpcell.00044.2004

DeLorey, D. S., Kowalchuk, J. M., \& Paterson, D. H. (2003). Relationship between pulmonary O 2 uptake kinetics and muscle deoxygenation during moderateintensity exercise. Journal of Applied Physiology (Bethesda, Md. : 1985), 95(April 2003), 113-120. doi:10.1152/japplphysiol.00956.2002

DeLorey, D. S., Kowalchuk, J. M., \& Paterson, D. H. (2004a). Effect of age on O(2) uptake kinetics and the adaptation of muscle deoxygenation at the onset of moderate-intensity cycling exercise. Journal of Applied Physiology, 97(1), 165172. doi:10.1152/japplphysiol.01179.2003

DeLorey, D. S., Kowalchuk, J. M., \& Paterson, D. H. (2004b). Effects of prior heavyintensity exercise on pulmonary O 2 uptake and muscle deoxygenation kinetics in young and older adult humans. Journal of Applied Physiology (Bethesda, Md. : 1985), 97(3), 998-1005. doi:10.1152/japplphysiol.01280.2003

Fitts, R. H. (2008). The cross-bridge cycle and skeletal muscle fatigue. Journal of Applied Physiology, 104(2), 551-558. doi:10.1152/japplphysiol.01200.2007

Forbes, S. C., Paganini, A. T., Slade, J. M., Towse, T. F., \& Meyer, R. a. (2009). Phosphocreatine recovery kinetics following low- and high-intensity exercise in human triceps surae and rat posterior hindlimb muscles. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology, 296(1), R161R170. doi:10.1152/ajpregu. 90704.2008

Fox, E., Bartels, R., Billings, C., O’Brien, R., Bason, R., \& Mathews, D. (1975). Frequency and duration of interval training programs and changes in aerobic power. Journal of Applied Physiology, 38(3), 481-484.

Gerbino, A., Ward, S. A., \& Whipp, B. J. (1996). Effects of prior exercise on pulmonary gas-exchange kinetics during high-intensity exercise in humans. Journal of Applied Physiology, 80(1), 99-107.

Glancy, B., Barstow, T., \& Willis, W. T. (2008). Linear relation between time constant of oxygen uptake kinetics, total creatine, and mitochondrial content in vitro. American Journal of Physiology. Cell Physiology, 294(1), C79-C87. doi:10.1152/ajpcell.00138.2007

Gollnick, P., Karlsson, J., Piehl, K., \& Saltin, B. (1974). Selective glycogen depletion in skeletal muscle fibres of man following sustained contractions. Journal of Physiology, 241, 59-67.

Grassi, B. (2000). Skeletal muscle VO2 on-kinetics: set by O2 delivery or by O2 utilization? New insights into an old issue. Medicine and Science in Sports and Exercise, 32(1), 108-116.

Grassi, B., Gladden, L. B., Samaja, M., Stary, C. M., \& Hogan, M. C. (1998a). Faster adjustment of O 2 delivery does not affect $\mathrm{V}(\mathrm{O} 2)$ on-kinetics in isolated in situ canine muscle. Journal of Applied Physiology, 85(4), 1394-1403.

Grassi, B., Gladden, L. B., Stary, C. M., Wagner, P. D., C, M., Rossiter, H. B., ... Physiol, C. (1998b). Peripheral O2 diffusion does not affect Vo2 on-kinetics in isolated in situ canine muscle inhibition Peripheral O 2 diffusion does not affect V in isolated in situ canine muscle. Journal of Applied Physiology, 85, 1404-1412.

Grassi, B., Hogan, M. C., Greenhaff, P. L., Hamann, J. J., Kelley, K. M., Aschenbach, W. G., ... Gladden, L. B. (2002). Oxygen uptake on-kinetics in dog gastrocnemius in situ following activation of pyruvate dehydrogenase by dichloroacetate. The Journal of Physiology, 538(Pt 1), 195-207. doi:10.1113/jphysiol.2001.012984

Grassi, B., Hogan, M. C., Kelley, K. M., Aschenbach, W. G., Hamann, J., Evans, R. K., Ronald, K. (2000). Role of convective O 2 delivery in determiningV ? o 2 onkinetics in canine muscle contracting at peak V ? o 2 Role of convective O 2 delivery in determining V on-kinetics in canine muscle contracting at peak V . Journal of Applied Physiology, 89, 1293-1301.

Grassi, B., Pogliaghi, S., Rampichini, S., Quaresima, V., Ferrari, M., Marconi, C., \& Cerretelli, P. (2003). Muscle oxygenation and pulmonary gas exchange kinetics during cycling exercise on-transitions in humans. Journal of Applied Physiology (Bethesda, Md. : 1985), 95(February 2003), 149-158. doi:10.1152/japplphysiol.00695.2002

Grassi, B., Poole, D. C., Richardson, R. S., Knight, D. R., Erickson, B. K., \& Wagner, P. D. (1996a). Muscle O2 uptake kinetics in humans: implications for metabolic control. Journal of Applied Physiology (Bethesda, Md. : 1985), 80, 988-998.

Grassi, B., Poole, D. C., Richardson, R. S., Knight, D. R., Erickson, B. K., \& Wagner, P. D. (1996b). Muscle O2 uptake kinetics in humans: implications for metabolic control. Journal of Applied Physiology, 80(3), 988-998.

Grassi, B., Rossiter, H. B., Hogan, M. C., Howlett, R. a, Harris, J. E., Goodwin, M. L., ... Gladden, L. B. (2011). Faster $\mathrm{O}_{2}$ uptake kinetics in canine skeletal muscle in situ after acute creatine kinase inhibition. The Journal of Physiology, 589(Pt 1), 221-233. doi:10.1113/jphysiol.2010.195164

Howlett, R. a, Parolin, M. L., Dyck, D. J., Hultman, E., Jones, N. L., Heigenhauser, G. J., \& Spriet, L. L. (1998). Regulation of skeletal muscle glycogen phosphorylase and PDH at varying exercise power outputs. The American Journal of Physiology, 275, R418-R425.

Jones, A. M., Carter, H., Pringle, J. S. M., \& Campbell, I. T. (2002). Effect of creatine supplementation on oxygen uptake kinetics during submaximal cycle exercise. Journal of Applied Physiology, 92(6), 2571-2577. doi:10.1152/japplphysiol.01065.2001

Kindig, C. a, Howlett, R. a, Stary, C. M., Walsh, B., \& Hogan, M. C. (2005). Effects of acute creatine kinase inhibition on metabolism and tension development in isolated single myocytes. Journal of Applied Physiology, 98(2), 541-549. doi:10.1152/japplphysiol.00354.2004

Koga, S., Poole, D. C., Fukuoka, Y., Ferreira, L. F., Kondo, N., Ohmae, E., \& Barstow, T. J. (2011). Methodological validation of the dynamic heterogeneity of muscle deoxygenation within the quadriceps during cycle exercise. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology, 301(2), R534R541. doi:10.1152/ajpregu.00101.2011

Koppo, K., Bouckaert, J., \& Jones, A. M. (2004). Effects of Training Status and Exercise Intensity on Phase II VO2 Kinetics. Medicine \& Science in Sports \& Exercise, 36(2), 225-232.

Krustrup, P., Jones, a M., Wilkerson, D. P., Calbet, J. a L., \& Bangsbo, J. (2009). Muscular and pulmonary O 2 uptake kinetics during moderate- and high-intensity sub-maximal knee-extensor exercise in humans. The Journal of Physiology, 587(Pt 8), 1843-1856. doi:10.1113/jphysiol.2008.166397

Lamarra, N., Whipp, B. J., Ward, S. a, \& Wasserman, K. (1987). Effect of interbreath fluctuations on characterizing exercise gas exchange kinetics. Journal of Applied Physiology, 62(5), 2003-2012.

Macdonald, M., Pedersen, P. K., \& Hughson, R. L. (1997). Acceleration of Vo2 kinetics in heavy submaximal exercise by hyperoxia and prior high-intensity exercise humans to contractions subsequent heavy exercise ' O 2 kinetics in heavy submaximal exercise Acceleration of V by hyperoxia and prior high-intensi. Journal of Applied Physiology, 83(4), 1318-1325.

Mancini, D., Bolinger, L., Li, H., Kendrick, K., Chance, B., \& Wilson, J. R. (1994). Validation of near-infrared spectroscopy in humans. Journal of Applied Physiology, 77(6), 2740-2747.

McCully, K., \& Hamaoka, T. (2000). Near-Infrared Spectroscopy: What Can It Tell Us about Oxygen Saturation in Skeletal Muscle?

Murias, J. M., Spencer, M. D., Kowalchuk, J. M., \& Paterson, D. H. (2011). Muscle deoxygenation to VO 2 relationship differs in young subjects with varying ??vO 2. European Journal of Applied Physiology, 11 1(12), 3107-3118. doi:10.1007/s00421-011-1937-9

Newcomer, B. R., Boska, M. D., \& Hetherington, H. (1999). T 1 Measurements of 31 P Metabolites in Resting and Exercising Human Gastrocnemius / Soleus Muscle at 1 . 5 Tesla. NMR in Biomedicine, 494(12), 486-494.

Nuutinen, E. M., Nishiki, K., Erecińska, M., \& Wilson, D. F. (1982). Role of mitochondrial oxidative phosphorylation in regulation of coronary blood flow. The American Journal of Physiology, 243(2), H159-H169.

Nyberg, M., Mortensen, S. P., Saltin, B., Hellsten, Y., \& Bangsbo, J. (2010). Low blood flow at onset of moderate-intensity exercise does not limit muscle oxygen uptake. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology, 298(3), R843-R848. doi:10.1152/ajpregu.00730.2009

Ozyener, F., Rossiter, H. B., Ward, S. a, \& Whipp, B. J. (2001). Influence of exercise intensity on the on- and off-transient kinetics of pulmonary oxygen uptake in humans. The Journal of Physiology, 533(Pt 3), 891-902. doi:10.1111/j.1469-7793.2001.t01-1-00891.x

Paterson, B. Y. D. H., \& Whipp, B. J. (1991). Asymmetries of oxygen uptake transients at the on-and offset of heavy exercise in humans. Journal of Physiology, 443, 575586.

Poole, D. C., Barstow, T. J., Gaesser, G., \& Willis, W. T. (1994). VO2 Slow component: physiological and functional significance. Medicine \& Science in Sports \& Exercise, 26(11), 1354-1358.

Pringle, J. S. M., Doust, J. H., Carter, H., Tolfrey, K., Campbell, I. T., Sakkas, G. K., \& Jones, A. M. (2003a). Oxygen uptake kinetics during moderate, heavy and severe intensity "submaximal" exercise in humans: the influence of muscle fibre type and capillarisation. European Journal of Applied Physiology, 89(3-4), 289-300. doi:10.1007/s00421-004-1101-x

Pringle, J. S. M., Doust, J. H., Carter, H., Tolfrey, K., Campbell, I. T., Sakkas, G. K., \& Jones, A. M. (2003b). Oxygen uptake kinetics during moderate, heavy and severe intensity "submaximal" exercise in humans: the influence of muscle fibre type and capillarisation. European Journal of Applied Physiology, 89, 289-300. doi:10.1007/s00421-004-1101-x

Rådegran, G., \& Saltin, B. (1998). Muscle blood flow at onset of dynamic exercise in humans. The American Journal of Physiology, 274, H314-H322.

Rossiter, H. B., Ward, S. a, Howe, F. a, Wood, D. M., Kowalchuk, J. M., Griffiths, J. R., \& Whipp, B. J. (2003). Effects of dichloroacetate on VO2 and intramuscular 31P metabolite kinetics during high-intensity exercise in humans. Journal of Applied Physiology (Bethesda, Md. : 1985), 95(3), 1105-1115. doi:10.1152/japplphysiol.00964.2002

Rossiter, H. B., Ward, S. a, Kowalchuk, J. M., Howe, F. a, Griffiths, J. R., \& Whipp, B. J. (2002). Dynamic asymmetry of phosphocreatine concentration and O(2) uptake between the on- and off-transients of moderate- and high-intensity exercise in humans. The Journal of Physiology, 541, 991-1002. doi:10.1113/jphysiol.2001.012910

Rossiter, H. B., Ward, S. a., Doyle, V. L., Howe, F. a., Griffiths, J. R., \& Whipp, B. J. (1999). Inferences from pulmonary O2 uptake with respect to intramuscular [phosphocreatine] kinetics during moderate exercise in humans. Journal of Physiology, 518, 921-932. doi:10.1111/j.1469-7793.1999.0921p.x

Saitoh, T., Ferreira, L. F., Barstow, T. J., Poole, D. C., Ooue, A., Kondo, N., \& Koga, S. (2009). Effects of prior heavy exercise on heterogeneity of muscle deoxygenation
kinetics during subsequent heavy exercise. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology, 297(3), R615-R621. doi:10.1152/ajpregu.00048.2009

Scheuermann, B. W., \& Barstow, T. J. (2003). O2 uptake kinetics during exercise at peak O2 uptake. Journal of Applied Physiology, 95(5), 2014-2022. doi:10.1152/japplphysiol.00590.2002

Spriet, L. L., Howlett, R. a, \& Heigenhauser, G. J. (2000). An enzymatic approach to lactate production in human skeletal muscle during exercise. Medicine and Science in Sports and Exercise, 32(4), 756-763.
Swanson, G. D. (1980). Breath-to-breath considerations for gas exchange kinetics. Exercise, bioenergetics and gas exchange. Amsterdam: Elsevier/North Holland, 211-222.

Vollestad, N. K., \& Blom, C. . (1985). Effect of varying intensity on glyocgen depletion in human muscle fibres. Acta Physiologica Scandinavica, 125, 395-405.

Whipp, B. J. (1994). The slow component of O2 uptake kinetics during heavy exercise, 26(11), 1319-1326.

Whipp, B. J., Ward, S. a, Lamarra, N., Davis, J. a, \& Wasserman, K. (1982). Parameters of ventilatory and gas exchange dynamics during exercise. Journal of Applied Physiology (Bethesda, Md. : 1985), 52(6), 1506-1513.

Xu, F., \& Rhodes, E. C. (1999). Oxygen uptake during exercise, 27(5), 313-327.

## CHAPTER 2

## 2 THE EFFECTS OF SHORT WORK VERSUS LONGER WORK PERIODS WITHIN INTERMITTENT EXERCISE ON $\dot{\mathrm{V}} \mathbf{O}_{2}$ KINETICS, MUSCLE DEOXYGENATION AND ENERGY SYSTEM CONTRIBUTION

### 2.1 INTRODUCTION

Continuous heavy intensity exercise (CONT) has been compared with short work ( 10 s ), shorter recovery ( 5 s ) intermittent exercise in which the work performed is identical to that of the continuous exercise regardless of a reduction in mean work rate (Belfry et al., 2012a; Belfry et al., 2012b). They demonstrated that the phosphocreatine (PCr) synthesis at 4 s of recovery during intermittent exercise (Belfry et al., 2012a), was followed by increased PCr breakdown during the subsequent work period. This increased substrate phosphorylation contribution resulted in an observed reduction in mean oxidative phosphorylation as indicated by a reduced $\dot{\mathrm{V}} \mathrm{O}_{2}$ compared to that necessary to perform the same work rate performed continuously (Belfry et al., 2012b).

By increasing the frequency of the recovery periods there is consequently a shortening of the accompanying work periods (CONT, to 25 s work ( 25 s ), to 10 s work $(10 \mathrm{~s})$ ). The increased capacity of PCr breakdown associated with the recovery periods provides adequate adenosine triphosphate (ATP) to perform the identical WR while reducing the glycolytic phosphorylation contribution necessary during work. This reduction, in part, should be reflected in a lower $\dot{\mathrm{V}} \mathrm{CO}_{2}$ as a consequence of the reduced ventilatory buffering associated with the reduction in $\left[\mathrm{H}^{+}\right]$release form the carbonic anhydrase reaction as work duration decreases (Beaver et al., 1986b).

Comparing the effects of short work: shorter recovery exercise to CONT exercise on the rate of adjustment $\dot{\mathrm{VO}}_{2}$ and $\mathrm{HHb}\left(\dot{\mathrm{VO}}_{2}\right.$ and HHb kinetics) has not been studied. The previously observed speeding of $\dot{\mathrm{V}} \mathrm{O}_{2}$ kinetics as work rate decreases has been associated with the decrease in metabolic demand (Paterson \& Whipp, 1991; Rossiter et al., 1999; Whipp \& Wasserman, 1972; Whipp et al., 1994; Barstow \& Mole 1991), whereas others have observed a slowing of $\dot{\mathrm{V}} \mathrm{O}_{2}$ kinetics with a similar decrease in work rate that has been ascribed to increased creatine kinase (CK) activity which, as a buffering system, decreases key metabolites (ADP) that drive oxidative phosphorylation (Grassi et al., 2011; Korzeniewski \& Zoladz, 2004). It is suggested that the increased PCr contribution and concomitant increased CK activity would slow $\dot{\mathrm{VO}}_{2}$ kinetics.

A slowing of the rate of adjustment of muscle deoxygenation (HHb kinetics) has previously been observed which was attributed to a disproportionate increase in microvascular blood flow to $\dot{\mathrm{V}} \mathrm{O}_{2}$ as $\mathrm{O}_{2}$ delivery is improved (DeLorey et al., 2004a; 2005). A similar increase in blood flow during the $10 \mathrm{~s}: 5 \mathrm{~s}$ recovery period was observed. This was attributed to the decrease in intramuscular pressures and impedance within the contracting muscle group during the recovery period (Rådegran \& Saltin, 1998). This increase in regional blood flow associated with the inclusion of short recovery periods may elicit a similar slowing of HHb kinetics.

The ratio of $\mathrm{HHb} / \mathrm{VO}_{2}$ is utilized to provide insight into $\mathrm{O}_{2}$ delivery and $\mathrm{O}_{2}$ utilization within the microvasculature (Murias et al., 2012). With an improvement within microvascular $\mathrm{O}_{2}$ delivery a corresponding decrease in the mismatching "overshoot" is observed (Murias et al., 2012). Further studies have shown that INT exercise of similar temporal relations have observed a decrease in the $\mathrm{HHb} / \mathrm{V}_{2}$
"overshoot" due to the improvements of $\mathrm{O}_{2}$ delivery as a consequence of improved blood flow (Belfry et al., 2012; Rådegran \& Saltin, 1998). If 25 s and 10 s work periods are paired with a shorter recovery period (3 s ) this recovery-dependent effect on $\mathrm{O}_{2}$ delivery and/or $\mathrm{O}_{2}$ utilization and its influence on both HHb and $\mathrm{HHb} / \mathrm{VO}_{2}$ may be mitigated as the time frame of the recovery period may not allow for a similar response within the truncated recovery period. Furthermore, the previously observed decrease in HHb during the recovery period of 10 s work: 5 s may be eliminated by reducing the recovery periods from 5 s to 3 s as recovery dependent improvements in blood flow may not be to the same extent with a shorter recovery periods.

Therefore the primary purposes of this study were to contrast the effects of increasing the recovery frequency from CONT exercise to 25 s , and 10 s protocols on; 1) the $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ and HHb kinetics, 2) the overall changes of $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ and HHb and 3) blood lactate concentration. This will provide insight into the responses of $\dot{\mathrm{VO}}_{2 \mathrm{p}}$ and HHb kinetics, $\mathrm{O}_{2}$ provision and utilization, and energy system contribution between each exercise condition. It was hypothesized that 1) the $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ and HHb kinetics would be fastest in the CONT bout and be sequentially slower with the inclusion of more frequent recovery periods, and 2) the $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}, \mathrm{HHb}$ and blood lactate concentration would be lower with the more frequent inclusion of 3 s recovery periods.

### 2.2 METHODS

Subjects. Ten males (see Table 1) volunteered and gave written informed consent to participate in this study. All procedures were approved by The University of Western Ontario's Ethics Board for Health Sciences Research Involving Human Subjects.

Subjects were recreationally active and non-smokers, and had no known cardiovascular, respiratory, metabolic or musculoskeletal diseases. Subjects were not taking any medications that might affect the cardiorespiratory and hemodynamic response to exercise. Subjects were instructed not to perform any strenuous exercise for up to twenty-four hours prior to visits to the laboratory, nor were they to consume any food or caffeine two hours prior.

Pre-experimental Protocol. Each subject reported to the laboratory to perform a fatiguelimited ramp incremental (RI) exercise test ( 20 W baseline for 4 min followed by a 25 $\mathrm{W} \cdot \mathrm{min}^{-1} \mathrm{ramp}$ ) on an electronically-braked cycle ergometer (H-300-R Lode; Lode B. V., Groningen, Holland) for determination of peak $\dot{\mathrm{V}}_{2 \mathrm{p}}\left(\dot{\mathrm{VO}}_{2 \text { peak }}\right)$, peak power output (PPO) and estimated lactate threshold ( $\hat{\theta}_{\mathrm{L}}$ ). Subjects were asked to maintain a cadence between 60-70 rpm during the test. $\dot{\mathrm{V}} \mathrm{O}_{\text {2peak }}$ was defined as the highest 20-s $\dot{\mathrm{V}} \mathrm{O}_{2 \text { p }}$. PPO was defined as the WR achieved at the termination of the RI test.

The $\hat{\theta}_{\mathrm{L}}$ was estimated by visual inspection using previously described standard gas exchange and ventilatory parameters (Beaver et al., 1986a). Briefly this point was determined by observing the $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ at which $\mathrm{CO}_{2}$ output $\left(\dot{\mathrm{V} C O}{ }_{2 \mathrm{p}}\right)$ began to increase out of proportion to $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$, the non-linear increase in minute ventilation $(\dot{\mathrm{V} e})-$ to $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$, while the initial increase in end-tidal $\mathrm{PO}_{2}\left(\mathrm{PetO}_{2}\right)$ to $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$, and the $\dot{\mathrm{V}} \mathrm{e}-$ to $\dot{\mathrm{V} C O} \mathrm{Cl}_{2}$ ratio and end-tidal $\mathrm{PCO}_{2}\left(\mathrm{PetCO}_{2}\right)$ to $\dot{\mathrm{V}}_{2 \mathrm{p}}$ were unchanged (Beaver et al., 1986a). A heavy intensity work rate (WR) of delta ( $\Delta$ ) $60 \%$ was calculated $(\mathrm{WR} \Delta 60)$ from the results of the RI test. The WR was $60 \%$ of the difference between $\hat{\theta}_{\mathrm{L}}$ and $\dot{\mathrm{V}} \mathrm{O}_{2 \text { peaak }}$. Two
experienced exercise physiologists evaluated each graph to identify $\hat{\theta}_{\mathrm{L}}$ (Beaver et al., 1986a). If a discrepancy arose between the two investigators a mean of the identified points was utilized.

Experimental Protocol. Subjects returned to the laboratory on six separate occasions and performed one of three cycle ergometer exercise protocols on each visit. The exercise protocols were as follows: 1) Continuous protocol (CONT): 4 min baseline cycling at 20 W , followed by 8 min of exercise at $\mathrm{WR} \Delta 60,2$ ) a 25 s work : 3 s recovery protocol (25 s): 4 min baseline cycling at 20 W followed by a series of square wave exercise intervals alternating with a 3 s recovery period at 20 W followed by 25 s at WR $\Delta 60$ for 8 min , and 3) a 10 s work : 3 s recovery protocol ( 10 s ): 4 min baseline cycling at 20 W followed by a series of square wave exercise intervals alternating from 3 s recovery at 20 W to 10 s at WR $\Delta 60$ for 8 min . Subjects performed 2 trials of each protocol. During each trial the subjects were asked to maintain a cadence of 60-70 rpm that was on display visually.

Data Collection. Breath-by-breath gas-exchange measurements were made continuously during each exercise protocol and were similar to those previously described (Keir et al., 2014b) . During each trial subjects breathed through a mouthpiece and wore a noseclip. Inspired and expired volumes and flow rates were measured using a pneumotach (Hans Rudolph, Model 4813) and a low dead space ( 90 mL ) bidirectional turbine (Alpha Technologies, VMM 110) positioned in series with the mouthpiece. The pneumotach was adjusted for zero flow whereas the volume turbine was calibrated before each test using a syringe of known volume (3 L). Expired gas was sampled continuously at the
mouth by mass spectrometry (PerkinElmer, Medical Gas Analyzer 1100, Massachusetts, United States; Innovision, AMIS 2000, Lindvedvej, Denmark) and analyzed for concentrations of $\mathrm{O}_{2}$ and $\mathrm{CO}_{2}$. Gas concentrations were calibrated with precisionanalyzed gas mixtures. The time delay between an instantaneous, square-wave change in fractional gas concentration at the sampling inlet, and its detection by the mass spectrometer was measured electronically. Respiratory volumes, flow, and gas concentrations were recorded in real-time at a sampling frequency of 100 Hz and transferred to a computer. Gas concentrations were aligned with respiratory flow using the measured delay to the mass spectrometer. The computer executed a peak-detection program to determine $\mathrm{PetO}_{2}$ and $\mathrm{PetCO}_{2}$, as well as inspired and expired volumes and durations to build a profile of each breath. Breath-by-breath gas exchange at the pulmonary capillary was calculated using the Swanson (1980) algorithms.

Muscle deoxygenation ( HHb ) of the vastus lateralis muscle was measured using a frequency domain multi-distance near-infrared spectroscopy (NIRS) system (Oxiplex TS, Model 92505, ISS, Champaign, USA) as described elsewhere (Spencer et al., 2012). Briefly, the system was comprised of a single channel consisting of eight laser diodes operating at two wavelengths ( $\lambda=690$ and 828 nm , four at each wavelength), which were pulsed in a rapid succession down a photomultiplier tube. A rigid plastic NIRS probe (connected to laser diodes and photomultiplier tube by optical fibres) consisted of two parallel rows of light emitter fibres and one detector fibre bundle; the sourcedetector separations for this probe were $2.0,2.5,3.0$, and 3.5 cm for both wavelengths. The probe was placed on the greatest circumference of the vastus lateralis muscle. NIRS measurements were collected continuously for the entire duration of each trial. The

NIRS unit was calibrated at the beginning of each testing session after the unit had been on for at least 20 min . Continuous measurements of the absolute scattering $\left(\mu_{\mathrm{s}}\right)$ and absorption $\left(\mu_{\mathrm{a}}\right)$ coefficients were determined from the measured intensity and phase shift of the light entering and traversing the tissue (at both wavelengths). Absolute concentrations of HHb and $\mathrm{HbO}_{2}(\mu \mathrm{M})$ were then derived. Data were collected at a frequency of 25 Hz , then reduced to 1-s bins for storage and subsequent analyses.

Heart rate (HR) was collected using a Polar Wearlink Chest Strap, H1 Heart Rate Sensor and SP0180 Polar Transmitter (Polar Electro Inc., Lachine, QC, Canada) linked to a PowerLab Chart data collection system (v.7.3.1 ADInstruments Inc., Colorado, CO, USA).

Arterialized-capillary blood samples $(\sim 5 \mu \mathrm{~L})$ were taken from the index finger 6 min before and 2 min after all trials for blood lactate measurments ([Lac $\left.{ }^{-}\right] ; \mathrm{mM}$ ). An Accu-Chek (Safe-T-Pro Plus) lancet was utilised after the finger had been sterilised using alcohol swabs. The researcher wore sterile latex gloves for this procedure. A lactate analyzer (Lactate Scout; Sports Resource Group, Hawthorne, NY) analysed the samples immediately.

Data Analysis. Breath-by-breath $\dot{\mathrm{VO}}_{2 \mathrm{p}}$ data were edited on an individual basis by removing aberrant data 3 SD and greater from the local mean (Lamarra et al., 1987). After each subject's individual trial was edited, the two like-trials were linearly interpolated on a second-by-second basis, time aligned and ensemble-averaged such that
time "zero" represented the onset of the transition to WR $\Delta 60$. The on-transient of each profile was modeled with the following mono-exponential function (Equation 1):

$$
\begin{equation*}
\mathrm{y}(t)=\mathrm{y}_{\mathrm{BSL}}+\mathrm{A}_{\mathrm{p}} \cdot\left(1-e^{-(t-\mathrm{TD}) / \tau}\right) \tag{1}
\end{equation*}
$$

where, $\mathrm{y}(t)$ is the value of the dependent variable at any time during the transition, $\mathrm{y}_{\mathrm{BSL}}$ is the pre-transition baseline value, $\mathrm{A}_{\mathrm{p}}$ is the steady-state increase in y above the baseline value, $\tau$ is the time constant of the response or the time for $y$ to increase to $63 \%$ of the new steady-state, and TD is the time delay. The details of the fitting procedure are described elsewhere (Keir et al., 2014a). Briefly, the Levenberg-Marquardt algorithm was applied to find the minimum sum of squared residuals between the monoexponential function and the experimental data using specialized software (Origin 8.5; OriginLab, Northampton, MA). The phase I-phase II transition was determined by visual inspection of the second-by-second and 5-s averaged data as the point at which there was a sharp decrease from baseline values ( 20 W cycling) in both respiratory exchange ratio (RER) and PetO2. The end of the phase II fitting window was determined by examining the change in $\tau, \mathrm{CI}_{95}, \chi^{2}$, and plotted residuals in response to progressive increases in the end of the fitting window. The point at which there was a systematic increase in $\tau, \mathrm{CI}_{95}$, and $\chi^{2}$ was considered as the end of phase II.

The TD for the HHb response was determined using second-by-second data and corresponded to the time, after the onset of exercise, at which the HHb signal began a systematic increase from its nadir value. Determination of the TD of HHb was made on each individual's ensemble-averaged response and the data were modeled using Equation 1. Different fitting strategies ranging from $90-180 \mathrm{~s}$ into a transition resulted
in no differences in $\tau \mathrm{HHb}(\mathrm{p}>0.05)$. Baseline values for $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}\left(\dot{\mathrm{V}}_{2 \mathrm{pBSI}}\right)$, and HHb $\left(\mathrm{HHb}_{\mathrm{BSL}}\right)$ values were fixed as the mean value of the 60 s "window" prior to the work rate transition. The HHb data of each condition was then normalized to a zero baseline ultimately showing the progression of the HHb response above baseline.

CONT, 25 s , and 10 s HHb data were divided into a series of work and recovery blocks between $120-240$ s corresponding to the specific condition. These blocks were then overlaid to give a mean HHb response over the specific work; recovery cycle for each subject (Figure 5). CONT and 25 s HHb data were divided into four time blocks to match the 28 s intervals; 3 s recovery, $4-8 \mathrm{~s}$ work, $9-13 \mathrm{~s}$ work, and $14-28 \mathrm{~s}$ work. Whereas CONT and 10 s data were divided into three time blocks to match the 13 s intervals; 3 s recovery period, 4 - 8 s work, and $9-13 \mathrm{~s}$ work.
$\mathrm{O}_{2}$ pulse $\left(\mathrm{O}_{2}\right.$ pulse $\left.=\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}} / \mathrm{HR}\right)\left(\right.$ Whipp et al., 1996), a measure of bulk $\mathrm{O}_{2}$ delivery, was calculated at 180 s . At this time point end Phase II (4 $\tau$ ) had been reached in all conditions. The ratio of $\mathrm{HHb} / \mathrm{VO}_{2}$ was obtained from normalized HHb and $\mathrm{VO}_{2}$ data to a zero baseline value. The normalized values were then converted to percentages of the peak values of the final 30 s of exercise $\left(\mathrm{HHb}_{\text {End }}\right.$ and $\left.\dot{\mathrm{V}} \mathrm{O}_{\text {2End }}\right)$, within each of the conditions of the corresponding measures.

Statistical Analysis. Table data are presented as means $\pm$ SD, bar graphs are presented as means $\pm$ SE. A one-way analysis of variance (ANOVA) for repeated measures was used to compare kinetics responses between conditions. A two-way ANOVA for repeated measures was used to compare responses across time between the three conditions. Where significant main effects were found, a Tukey post hoc analysis was performed for
multiple comparisons testing. All ensemble-averaged data is presented in 5 s averages unless otherwise stated. All statistical analyses were performed using SigmaPlot Version 12.3, (Systat Software Inc., San Jose, CA). Statistical significance was accepted at an alpha level of $5 \%$.

### 2.3 RESULTS

Absolute and relative workloads of $\Delta 60$. The mean WR between all conditions were different ( $\mathrm{p}<0.05$; Table 2).

Oxygen Uptake $\left(\dot{V} O_{2}\right)$ and $\dot{V} O_{2}$ kinetics. The mean $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ response profile for all subjects during the exercise conditions is displayed in Figure 1. Mean $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ from 61 s to end exercise was different in all conditions $(\mathrm{p}<0.05) . \tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ was not different between conditions ( $\mathrm{p}>0.05$; Table 2). $\dot{\mathrm{V}} \mathrm{O}_{2}$ at the end of phase two $\left(\dot{\mathrm{VO}}_{2 \mathrm{pu}}\right)$ during CONT and 25 s was higher than $10 \mathrm{~s}(\mathrm{p}<0.05)$, whereas CONT and 25 s were similar $(\mathrm{p}>0.05)$. The amplitude of the $\dot{\mathrm{V}} \mathrm{O}_{2}$ slow component $\left(\dot{\mathrm{V}}_{2 \mathrm{sc}}\right)$ and $\dot{\mathrm{V}} \mathrm{O}_{2}$ at the end of exercise $\left(\dot{\mathrm{V}}_{2 \text { End }}\right)$ was different between conditions ( $\mathrm{p}<0.05$; Table 2 ).
$\dot{V} \mathrm{CO}_{2}$ Production and RER . The $\dot{\mathrm{V}} \mathrm{CO}_{2}$ (Figure 2) and respiratory exchange ratio (RER $=\dot{\mathrm{V}} \mathrm{CO}_{2} / \dot{\mathrm{V}}_{2}$ ) at end exercise were different between all conditions ( $\mathrm{p}<0.05$; Figure 6; Table 2). Whereas the amplitude of the $\dot{\mathrm{V} C O}{ }_{2}$ slow component $\left(\dot{\mathrm{V} C O}_{2 \mathrm{sc}}\right)$ was different between CONT, and both the intermittent conditions ( $\mathrm{p}<0.05$ ). No difference was observed between the two intermittent conditions $(\mathrm{p}>0.05)($ Table 2$)$.
$\Delta H H b / \Delta \dot{V} O_{2 . \text {. }}$ The matching between conditions of the $\mathrm{HHb} / \mathrm{V}_{2}$ overshoot were significantly faster with the increased frequency of the 3 s recovery periods ( $\mathrm{p}<0.05$ ) (Figure 4).

Muscle deoxygenation parameters and NIRS-derived HHb kinetics. The mean HHb response during the CONT, 25 s and 10 s trials are presented in Figure 3. A summary of parameter estimates for the on-transient muscle deoxygenation is presented in Table 2. $\tau \mathrm{HHb}$ and HHb , at the end of phase 2 , and HHb at end exercise $\left(\mathrm{HHb}_{\text {End }}\right)$ did not differ between conditions ( $\mathrm{p}>0.05$; Table 2). Oscillations in HHb (Figure 5) during 25 s were not observed between the 3 s recovery and $4-8 \mathrm{~s}$ work, whereas differences between 9 -13 s to $14-28 \mathrm{~s}$, and 3 s recovery to 5 s work were observed ( $\mathrm{p}<0.05$ ). Within the 10 s: 3 s condition HHb was similar between 3 s recovery and $4-8 \mathrm{~s}$ of work, whereas both were lower than $9-13 \mathrm{~s}$ work ( $\mathrm{p}<0.05$ ). There were no oscillations in CONT, within similar 25 s and 10 s cycles, over the same period ( $\mathrm{p}>0.05$ ).

Heart rate and $O_{2}$ Pulse. The mean HR response during the CONT, 25 s and 10 s conditions are presented in Figure 7. HR differed across conditions ( $p>0.05$ ) from 60 s to end exercise. $\mathrm{O}_{2}$ pulse was similar across conditions at 180 s of exercise ( $\mathrm{p}>$ 0.05 ;Table 2).

Blood lactate concentration. Pre-exercise mean blood [Lac] for CONT, 25 s , and 10 s were similar between conditions ( $p>0.05$ ). Post exercise measurements showed that [Lac ${ }^{-}$] differed between all conditions (Table 2; $\mathrm{p}<0.05$ ).

Table 1.
Subject characteristics and results from incremental test

|  | Subject | Age | Height (cm) | Weight $(\mathrm{kg})$ | $\begin{aligned} & \text { PPO } \\ & \text { (W) } \end{aligned}$ | $\begin{aligned} & \dot{\mathrm{VO}_{2 \max }(\mathrm{~mL}} \\ & \mathrm{O}_{2} * \mathrm{~kg}^{-1} * \\ & \left.\min ^{-}\right) \end{aligned}$ | $\begin{aligned} & \hline \hat{\theta}_{\mathrm{L}}(\mathrm{~L} \\ & \mathrm{O}_{2} / \mathrm{mi} \\ & \mathrm{n}) \\ & \hline \end{aligned}$ | Work Rate ( $\Delta 60 \mathrm{~W}$ ) |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | 1 | 24 | 178 | 83.9 | 317 | 43.9 | 1.8 | 248 |
|  | 2 | 24 | 176 | 71.4 | 324 | 50.4 | 1.7 | 242 |
|  | 3 | 22 | 173 | 72.3 | 343 | 54.6 | 2.1 | 262 |
|  | 4 | 25 | 183 | 82.6 | 367 | 49.8 | 2.1 | 277 |
|  | 5 | 24 | 173 | 69.2 | 286 | 45.9 | 1.3 | 203 |
|  | 6 | 31 | 190 | 85.0 | 385 | 48.5 | 2.1 | 293 |
|  | 7 | 24 | 185 | 91.5 | 305 | 40.3 | 1.7 | 231 |
|  | 8 | 24 | 173 | 76.8 | 282 | 41.9 | 1.2 | 203 |
|  | 9 | 24 | 188 | 79.2 | 383 | 52.1 | 2.3 | 300 |
|  | 10 | 21 | 183 | 78.1 | 329 | 51.9 | 2.1 | 262 |
| Mean |  | 24 | 180 | 79 | 332 | 47.9 | 1.8 | 252 |
| SD |  | 3 | 6 | 7 | 37 | 4.7 | 0.4 | 34 |

Values given as means (SD). $\dot{\mathrm{V}} \mathrm{O}_{2 \text { max: }}$ maximal oxygen uptake, PPO: peak power output at $\dot{\mathrm{V}} \mathrm{O}_{2 \text { max, }}, \hat{\theta}_{\mathrm{L}}$ : estimated lactate threshold, $\Delta 60$ Work rate: work rate equivalent to $60 \%$ of the difference between $\hat{\theta}_{\mathrm{L}}$ and $\dot{\mathrm{V}} \mathrm{O}_{2 \text { max }}$

Table 2.
Results from CONT, $25 \mathrm{~s}: 3 \mathrm{~s}, 10 \mathrm{~s}: 3 \mathrm{~s}$ conditions on measured parameters.

|  |  | Mean | SD |  |  | Mean | SD |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Work Rate | CONT | 252 | 33.6 | $\dot{\mathrm{V}}_{2} \mathrm{p}^{\mathrm{S}} \mathrm{SC}$ | CONT | 0.69 | 0.20 |
| (W) | 25 s | 225* | 29.7 | $\left(\mathrm{L} * \min ^{-1}\right)$ | 25 s | 0.38* | 0.18 |
|  | 10 s | 198* $\beta$ | 25.9 |  | 10 s | $0.21 * \beta$ | 0.09 |
| Pre - [ $\mathrm{Lac}^{-}$] | CONT | 1.8 | 0.36 | HHb BSL | CONT | 25.8 | 5.8 |
| (mM) | 25 s | 1.97 | 0.36 | ( $\mu \mathrm{M}$ ) | 25 s | 26.1 | 5.7 |
|  | 10 s | 1.99 | 0.36 |  | 10 s | 26.6 | 5.7 |
| Post - [ $\mathrm{Lac}^{-}$] | CONT | 14.28 | 3.08 | HHb TD (s) | CONT | 5.6 | 1.17 |
| (mM) | 25 s | 10.23* | 1.9 |  | 25 s | 8.1 | 0.99 |
|  | 10 s | 7.0* $\beta$ | 1.71 |  | 10 s | 8.8 | 0.92 |
| $\dot{\mathrm{V}}^{2}$ BSL | CONT | 0.969 | 0.09 | $\tau \mathrm{HHb}$ (s) | CONT | 10.4 | 4.8 |
| $\left(\mathrm{L} * \min ^{-1}\right)$ | 25 s | 0.962 | 0.13 |  | 25 s | 9.3 | 4.9 |
|  | 10 s | 0.977 | 0.10 |  | 10 s | 8.5 | 3.9 |
| $\dot{\mathrm{V}}_{2} \mathrm{TD}(\mathrm{s})$ | CONT | 13.0 | 5.16 | $\mathrm{HHb}_{\text {END }}$ | CONT | 16.1 | 12.0 |
|  | 25 s | 17.0 | 4.08 | (a.u) | 25 s | 15.2 | 11.5 |
|  | 10 s | 14.5 | 3.5 | (Normalized) | 10 s | 15.0 | 12.1 |
| $\dot{\mathrm{V}} \mathrm{O}_{2 \text { EndPhase2 }}$ | CONT | 3.01 | 0.30 | RER | CONT | 1.13 | 0.03 |
| $\left(\mathrm{L} * \min ^{-1}\right)$ | 25 s | 2.91 | 0.35 |  | 25 s | 1.09* | 0.02 |
|  | 10 s | $2.67 * \beta$ | 0.31 |  | 10 s | 1.06 * $\beta$ | 0.04 |
| $\tau \dot{\mathrm{V}}_{2}(\mathrm{~s})$ | CONT | 44.2 | 9.44 | $\mathrm{O}_{2}$ - Pulse | CONT | 19.7 | 1.7 |
|  | 25 s | 38.9 | 10.06 | (mL/beat) | 25 s | 19.6 | 1.7 |
|  | 10 s | 39.4 | 8.31 |  | 10 s | 19.0 | 1.8 |
| $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{pEND}}$ | CONT | 3.69 | 0.37 | $\dot{\mathrm{V}} \mathrm{CO}_{2 \text { EndPhase2 }}$ | CONT | 3.79 | 0.44 |
| $\left(\mathrm{L} * \min ^{-1}\right)$ | 25 s | 3.29* | 0.36 | $\left(\mathrm{L} * \min ^{-1}\right)$ | 25 s | 3.53* | 0.36 |
|  | 10 s | $2.89 * \beta$ | 0.33 |  | 10 s | $2.95 * \beta$ | 0.29 |

Values are given as means (SD). CONT: continuous condition; $25 \mathrm{~s}: 25 \mathrm{~s}$ work : 3 s recovery intermittent exercise condition; $10 \mathrm{~s}: 10 \mathrm{~s}$ work : 3 s recovery intermittent exercise; $\dot{\mathrm{VO}}_{2}$ : oxygen uptake; $\dot{\mathrm{VCO}}_{2}$ : carbon dioxide production; HHb : muscle deoxygenation; $\mathrm{O}_{2}$ - Pulse: proxy measure of oxygen delivery per heart beat; SC : slow component; $\tau$ : time constant representing $63 \%$ of time to reach steady state; BSL: baseline; TD: time delay; RER: respiratory exchange ratio, * different from CONT ( $\mathrm{p}<$ $0.05), \beta$ different from to $25 \mathrm{~s}: 3 \mathrm{~s}(\mathrm{p}<0.05)$


Figure 1. Mean $\dot{\mathrm{VO}}_{2}$ response during CONT (continuous $\Delta 60$ work rate) (filled circle), $25 \mathrm{~s}(25 \mathrm{~s}$ work $(\Delta 60): 3 \mathrm{~s}$ recovery (20W)) (grey circle), and 10 s ( 10 s work ( $\mathrm{\Delta} 60$ ) : 3 s recovery (20W)) (open circle) (Overall $\dot{\mathrm{V}}_{2}$ was lower as increased frequency of recovery periods were introduced within INT; p $<0.05$ )


Figure 2. Mean $\dot{\mathrm{V}} \mathrm{CO}_{2}$ response during CONT (continuous $\Delta 60$ work rate) (filled circle), $25 \mathrm{~s}(25 \mathrm{~s}$ work ( $\Delta 60$ ) : 3 s recovery (20W)) (grey circle), and 10 s ( 10 s work ( $\Delta 60$ ) : 3 s recovery (20W)) (open circle) (Overall $\mathrm{V}^{2} \mathrm{CO}_{2}$ was lower as increased frequency of recovery periods were introduced within INT; $\mathbf{p}<0.05$ )


Figure 3. Mean HHb response during CONT (continuous $\Delta 60$ work rate) (filled circle), $25 \mathrm{~s}(25 \mathrm{~s}$ work $(\Delta 60): 3 \mathrm{~s}$ recovery (20W)) (grey circle), and 10 s ( 10 s work ( 460 ):3 s recovery (20W)) (open circle) ( HHb among conditions was not significantly different; $\mathrm{p}>0.05$ )


Figure 4. $\Delta \mathrm{HHb} / \Delta \dot{\mathrm{V}} \mathrm{O}_{2}$ for CONT (continuous $\Delta 60$ work rate) (A), 25 s ( 25 s work $(\Delta 60): 3$ s recovery (20W)) (B) and 10 s ( 10 s work ( $\Delta 60$ ) : 3 s recovery (20W)) (C). Time elapsed until overshoot diminished was significantly different; ( $\mathrm{p}<0.05$ ) Dashed lines represent time point of matching.


Figure 5. Average acute HHb oscillations $( \pm \mathrm{SE})$ in $25 \mathrm{~s}: 3 \mathrm{~s}(\mathrm{~B})$ and $10 \mathrm{~s}: 3 \mathrm{~s}(\mathrm{D})$ at $0-$ 3 s recovery, $4-8 \mathrm{~s}$ work, $9-13 \mathrm{~s}$ work, and $14-28 \mathrm{~s}$ work (B only). $0-3 \mathrm{~s}$ recovery and $4-8 \mathrm{~s}$ work different from $9-13 \mathrm{~s}$ work, and $14-28 \mathrm{~s}$ (B) (noted by asterisk) ( $\mathrm{p}<$ 0.05 ; from $120 \mathrm{~s}-240 \mathrm{~s}$ ) Both A and C represent the CONT condition.


Figure 6. Mean $\dot{\mathrm{V}}_{2}$ (open circles) and $\dot{\mathrm{V}} \mathrm{CO}_{2}$ (filled circles) for CONT (continuous $\Delta 60$ work rate) (A), $25 \mathrm{~s}(25 \mathrm{~s}$ work ( $\Delta 60$ ) : 3 s recovery (20W)) (B) and 10 s ( 10 s work ( $\Delta 60$ ) : 3 s recovery (20W)) (C). Respiratory exchange ratio (RER) indicated by $\dot{\mathrm{VCO}} \mathrm{Cl}_{2} / \mathrm{VO}_{2}$ was significantly different between all conditions; $(\mathrm{p}<0.05)$


Figure 7. Mean heart rate (HR) response during CONT (continuous $\Delta 60$ work rate) (filled circle), 25 s ( 25 s work ( $\Delta 60$ ) : 3 s recovery (20W)) (grey circle), and $10 \mathrm{~s}(10 \mathrm{~s}$ work ( $\Delta 60$ ) : 3 s recovery ( 20 W )) (open circle) (Overall HR was lower as increased frequency of recovery periods were introduced within INT; $\mathrm{p}<0.05$ )

### 2.4 DISCUSSION

It was hypothesized that 1) the $\tau \dot{\mathrm{VO}}{ }_{2 \mathrm{p}}$ and $\tau \mathrm{HHb}$ would be fastest in the continuous bout and slow with the increased frequency of the recovery periods, 2) the overall amplitude of both $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ and HHb would be lower with the increased frequency of the recovery periods and 3) blood lactate would decrease with the increased frequency of the recovery periods. The main findings of this study were: 1 ) the increased frequency of the recovery periods $\tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ was similar although the $\dot{\mathrm{V}} \mathrm{O}_{2}$ kinetics were slowed whereas $\tau \mathrm{HHb}$ and HHb kinetics were similar across conditions, and 2) $\dot{\mathrm{VO}}_{2 \mathrm{p}}$, and blood lactate concentrations were lower whereas HHb was similar at end exercise.

The $\dot{\mathrm{V}} \mathrm{O}_{2}$ kinetic response to decreasing work rates above the lactate threshold has previously been observed to speed $\dot{\mathrm{V}} \mathrm{O}_{2}$ kinetics (Whipp \& Wasserman, 1972; Jones et al., 2004; Casaburi et al, 1989). However, the increased frequency of 3 s recovery periods, while lowering mean WR (Table 2) and $\dot{\mathrm{V}}_{2 \mathrm{p}}$ (Figure 1), did not result in a speeding of $\tau \dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ (Table 2). The slowing of the $\dot{\mathrm{V}} \mathrm{O}_{2}$ kinetic response to a lower $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{pEND}}$ Phase II suggests the increased frequency of 3 s recovery periods elicited an increase in substrate phosphorylation contribution to replace the reduction in oxidative ATP associated with the decline in $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ brought forth by the decreased mean WR (Gollnick et al., 1974). Increases in PCr after 3 s rest (Newcomer et al., 1999) and 4 s of recovery within a similar protocol (Belfry et al., 2012a) have been observed. Moreover, decreased $\dot{\mathrm{V}}_{2 p}$ (Belfry et al., 2012b) and a greater contribution from substrate level phosphorylation ( PCr ) (Belfry et al., 2012a) have been observed previously during subsequent work periods. This previously observed increase in ADP phosphorylation
via PCr substitutes the observed reduction of ATP derived from oxidative phosphorylation. Furthermore, it has been suggested that increased activation of creatine kinase (CK) results in the slowing of $\dot{\mathrm{V}} \mathrm{O}_{2}$ kinetics (Korzeniewski \& Zoladz, 2004; Grassi et al., 2011). Moreover, the increase in PCr during recovery, and thus increased PCr breakdown during work has been shown to down regulate oxidative phosphorylation and slow the kinetic response of $\dot{\mathrm{V}}_{2 \mathrm{p}}$ (Grassi et al., 2011). As well, this slower $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ kinetic response has been associated to the increase in phosphate potential during the recovery period in relation to the increased activity of the CK enzyme (Grassi et al., 2011; Greenhaff, 2001; Korzeniewski, 2003; Meyer et al., 1984; Rossiter et al., 2005).

It has been suggested that decreased activity of the pyruvate dehydrogenase complex (PDH) that accompanies lower WR (Howlett et al., 1998; Spriet et al., 2002) associated with the increased recovery frequency in the present study would slow $\dot{\mathrm{V}}_{2}$ kinetics (Gurd et al., 2009). This would reduce availability of reducing equivalents $\left(\mathrm{NADH} / \mathrm{FADH}_{2}\right)$ intended for the Kreb's cycle, which limits the rate of oxidative phosphorylation, and, in part, could account for the observed slowing of the kinetic response of $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ in the present study. Conversely it has been shown that an increase in PDH activity does not affect $\mathrm{V}_{2}$ kinetics (Bangsbo et al., 2002), even when activated pharmacologically via dichloroacetate (DCA) (Grassi et al., 2002). The increased recovery frequency conditions could, through decreased PDH activity, contribute to the slowing of $\dot{\mathrm{V}}_{2}$ kinetics in the present study.

It has been observed previously that the amplitude of the $\dot{\mathrm{V}} \mathrm{O}_{2}$ slow component (Table 2) decreases as mean WR decreases (Barstow \& Molé, 1991; Casaburi et al.,

1989; Ozyener et al., 2001; Bartsow et al., 1996). The observed decrease in [Lac] (Table 2) has been positively correlated to smaller SC development (Barstow et al., 1993). The continued reduction in [ $\mathrm{Lac}^{-}$] with increasing recovery periods again suggests an increase in ATP - PCr phosphorylation contribution as opposed to glycolytic phosphorylation during these intermittent conditions (Belfry et al. 2012a). The concomitant decrease in RER corroborates this reduction in observed blood [ $\mathrm{Lac}^{-}$] as the necessity for bicarbonate buffering and accompanying $\dot{\mathrm{V}} \mathrm{CO}_{2}$ has been reduced (Beaver et al., 1986b; Beaver \& Wasserman, 1991). This supports the contention that the reduced oxidative phosphorylation for the identical ATP demand increases primarily the ATP - PCr contribution.

It has been suggested that $\mathrm{HHb} / \mathrm{VO}_{2 \mathrm{p}}$ provides insight into the matching of $\mathrm{O}_{2}$ provision to $\mathrm{O}_{2}$ utilization within the muscle under investigation (Murias et al., 2010). The inclusion and increased frequency of the 3 s recovery periods during of the intermittent conditions in the present study resulted in a progressively faster matching of the $\mathrm{HHb} / \dot{\mathrm{VO}}_{2 \mathrm{p}}$ (Figure. 4). It has been suggested that during the muscle relaxation phase of INT exercise there is a marked increase in blood velocity (reflecting blood flow) in comparison to the contraction phase (Walløe \& Wesche, 1988). It is suggested that this phenomena also exists during the 3 s recovery period of the INT conditions as intramuscular pressures (Rådegran \& Saltin, 1998) and impedance to flow is decreased within the working muscle (Barcroft \& Dornhorst, 1949; Folkow et al., 1970; Lutjemeier et al., 2005). This would increase microvasculature perfusion and improve $\mathrm{O}_{2}$ delivery as recovery frequency increases, thus reducing the $\mathrm{HHb} / \dot{\mathrm{V}}_{2 \mathrm{O}}$ overshoot.

This differs from the work by Essen et al.,, (1977) comparing CONT work (157 W) with matched mean power output yet markedly different 15 s work (299 W): 15 s recovery intermittent condition. They observed a decrease in blood flow yet higher HR during their intermittent condition. Presumably the higher HR they observed, compared to their CONT, was necessary to increase blood pressures and subsequent leg blood flow to contest the increased impedance to blood flow of the higher work rate during the 15 s work periods compared to the present study.
$\mathrm{O}_{2}$ pulse, a proxy measure of bulk $\mathrm{O}_{2}$ delivery (Whipp et al., 1996), was measured at 180 s (Table 2). At this time point, the end of Phase II had been attained in all exercise conditions. $\mathrm{O}_{2}$ pulse was equivalent across conditions (Table 2), whereas HR (Figure 7) and $\dot{\mathrm{V}}_{2 \mathrm{p}}$ (Figure 1) decreased with recovery frequency. This suggests that the decreased $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ across conditions was associated with the decrease in bulk $\mathrm{O}_{2}$ delivery. The decrease in bulk $\mathrm{O}_{2}$ delivery in CONT $\left(0.218 \mathrm{~L} \mathrm{O}_{2}\right)$ was well matched with the associated decrease in $\dot{\mathrm{V}}_{2 \mathrm{p}}$ in the 25 s conditions $\left(0.210 \mathrm{~L} \mathrm{O}_{2} / \mathrm{min}\right)$. However, the 27 W decrease in mean WR from 25 s to 10 s , eliciting a decrease in $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ of $0.300 \mathrm{~L} / \mathrm{min}$, was not matched by a similar decrease in bulk $\mathrm{O}_{2}$ delivery $\left(0.153 \mathrm{~L} \mathrm{O}_{2}\right)$. A similar increase in $\mathrm{O}_{2}$ delivery to $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ has been shown to slow the $\tau \mathrm{HHb}$ response (DeLorey et al., 2003;

DeLorey et al., 2005). This slowing was not observed in the present study and suggests a maldistribution of microvasculature blood flow within the area of interrogation as similar HHb and $\tau \mathrm{HHb}$ were observed regardless of recovery period inclusion (Figure 3) (DeLorey et al., 2003) $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ decreases (Nuutinen et al., 1982).

Finally, the within cycle fluctuations in HHb observed during both 25 s and 10 s , (Figure. 5) is similar to that observed previously during 10 s work: 5 s recovery
intermittent exercise (Belfry et al., 2012b). It is suggested that the decrease in HHb during recovery reflects either the increased $\mathrm{O}_{2}$ delivery originating from increased muscle pump activity (Rådegran \& Saltin, 1998; Walløe \& Wesche, 1988) and/or fluctuations in $\dot{\mathrm{VO}}_{2 \mathrm{p}}$ over the work recovery cycles (Belfry et al., 2012b). The modest oscillations in HHb observed over the work: recovery cycles within the current and previous work were well above baseline (Belfry et al., 2012b; Lutjemeier et al., 2008). This suggests that ATP that has been phosphorylated oxidatively for work has been directed to PCr synthesis during recovery. Furthermore, the similar HHb at 3 s of recovery and 5 s of work in both intermittent conditions in the current study (Figure 5) suggests a delay of oxidative phosphorylation due to ATP contribution from PCr breakdown (Belfry et al. 2012a).

It has been suggested that as recovery frequency increases across conditions, the reduced oxidative phosphorylation must be balanced by increases from the ATP -PCr energy system to sustain the identical work rate. Concomitantly, a reduction in citric acid cycle $\dot{\mathrm{V}} \mathrm{CO}_{2 \mathrm{p}}$ results. However, the disproportionate reduction in $\dot{\mathrm{V}} \mathrm{CO}_{2 \mathrm{p}}$ to $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ as recovery frequency increases, suggests a reduction in $\dot{\mathrm{V}} \mathrm{CO}_{2 \mathrm{p}}$ associated with ventilatory buffering. This reduced buffering of $\left[\mathrm{H}^{+}\right]$via the carbonic anhydrase reaction reflects the observed decrease in lactic acid production from glycolytic phosphorylation (Table 2) (Spriet et al., 2000).

The findings from this study may have particular training and performance implications to the sport of swimming in the United States. In this country, competitors at NCAA, National and International competitions will race and train in a variety of venues. These pools will range in size from $25 \mathrm{yd}, 50 \mathrm{~m}$, to open water. This dictates
differential work: recovery durations of the predominant propulsive muscles in freestyle swimming, the shoulder flexors and upper limb internal rotator muscles (Clarys 1985). In a middle distance race ( $500 \mathrm{yd}, 400 \mathrm{~m}$ ), elite freestyle swimmers will perform work periods of $\sim 10 \mathrm{~s}$ to $\sim 25 \mathrm{~s}$, in 25 yd and 50 m pools respectively, to continuous work in open water competitions. Notwithstanding the physiologic responses imposed by the hypoxic nature of the $\sim 3 \mathrm{~s}$ push and glide off the wall, and the imposed breathing patterns of swimming freestyle, this current study has shown that the physiology associated with these differing work: recovery durations dictated by the different pool sizes on the musculature could be markedly different. Exercise physiologists and coaches must be cognizant of the incongruence of the physiologic responses to the different pool lengths, despite the similar duration work bouts during training or competition. The effects of the increased anaerobic contributions and reduced aerobic contributions at similar power outputs, as pool size decreases, must be taken into account when training programs, and race strategies are designed.

### 2.5 CONCLUSIONS

Increasing the frequency of 3 s recovery periods elicited changes in energy system contributions that included a greater contribution from the ATP - PCr system and reduced glycolytic and oxidative phosphorylation contributions to perform the identical work rate. The concurrent slowing of the rate of adjustment of $\dot{\mathrm{V}} \mathrm{O}_{2 \mathrm{p}}$ of the different exercise conditions may be associated the role of different permutations of the substrate phosphorylation energy systems and the activity associated with the CK reaction. Finally, the increased frequency of recovery periods established enhanced microvascular blood flow, which facilitated an accelerated recovery dependent matching
of total $\mathrm{O}_{2}$ delivery to $\mathrm{O}_{2}$ utilization similar to that observed by longer recovery periods (5 s).

### 2.6 FUTURE DIRECTIONS AND LIMITATIONS

The current study differs methodologically from previous studies in which PCr resynthesis data was collected. During exercise in the current study participants would perform exercise upon a cycle ergometer whereas the previous participants executed plantar flexion exercise (Belfry et al., 2012a). Furthermore, the recovery periods during the present study were performed as active recovery rather than full rest as this may have effects upon PCr resynthesis.

Future studies may include the use of magnetic resonance spectroscopy in order to provide insight to $\left[\mathrm{H}^{+}\right], \mathrm{ADP}, \mathrm{PCr}$ status under the exercise conditions of the present study. Furthermore, the use of muscle biopsy samples of the vastus lateralis to provide insight into the muscle fibre type distribution within the area of interrogation may elucidate factors in relation to the $\dot{\mathrm{V}} \mathrm{O}_{2}$ kinetic response to the INT exercise conditions.

## REFERENCES

Bangsbo, J., Bangsbo, J., Gibala, M. J., Gibala, M. J., Krustrup, P., Krustrup, P., ... Saltin, B. (2002). Enhanced pyruvate dehydrogenase activity does not affect muscle O2 uptake at onset of intense exercise in humans. American Journal of Physiology. Regulatory, Integrative and Comparative Physiology, 282(1), R27380. Retrieved from http://www.ncbi.nlm.nih.gov/pubmed/11742848

Barcroft, B. Y. H., \& Dornhorst, A. C. (1949). The blood flow through the human calf during rhythmic exercise. Journal of Physiology, (109), 402-412.

Barstow, T. J., Casaburi, R., \& Wasserman, K. (1993). O2 uptake kinetics and the O2 deficit as related to exercise intensity and blood lactate. Journal of Applied Physiology (Bethesda, Md. : 1985), 75, 755-762.

Barstow, T. J., \& Molé, P. a. (1991). Linear and nonlinear characteristics of oxygen uptake kinetics during heavy exercise. Journal of Applied Physiology (Bethesda, Md. : 1985), 71, 2099-2106.

Beaver, W. L., \& Wasserman, K. (1991). Muscle RQ and Lactate Accumulation from Analysis of the VCO2-VO2 Relationship during exercise. Clinical Journal of Sport Medicine, 27-34.

Beaver, W. L., Wasserman, K., \& Whipp, B. J. (1986a). A new method for detecting anaerobic threshold by gas exchange A new method for detecting threshold by gas exchange anaerobic. Journal of Applied Physiology, 60(6), 2020-2027.

Beaver, W. L., Wasserman, K., \& Whipp, B. J. (1986b). Bicarbonate buffering of lactic acid generated during exercise. Journal of Applied Physiology (Bethesda, Md. : 1985), 60(2), 472-478.

Belfry, G. R., Raymer, G. H., Marsh, G. D., Paterson, D. H., Thompson, R. T., \& Thomas, S. G. (2012A). Muscle metabolic status and acid-base balance during 10-s
work:5-s recovery intermittent and continuous exercise. Journal of Applied Physiology, 113(3), 410-417. doi:10.1152/japplphysiol.01059.2011

Belfry, G. R., Paterson, D. H., Murias, J. M., \& Thomas, S. G. (2012B). The effects of short recovery duration on VO 2 and muscle deoxygenation during intermittent exercise. European Journal of Applied Physiology, 112(112), 1907-1915. doi:10.1007/s00421-011-2152-4

Casaburi, R., Barstow, T. J., Robinson, T., \& Wasserman, K. (1989). Influence of work rate on ventilatory and gas exchange kinetics. Journal of Applied Physiology (Bethesda, Md. : 1985), 67, 547-555.

DeLorey, D. S., Kowalchuk, J. M., \& Paterson, D. H. (2003). Relationship between pulmonary O 2 uptake kinetics and muscle deoxygenation during moderateintensity exercise. Journal of Applied Physiology (Bethesda, Md. : 1985), 95(April 2003), 113-120. doi:10.1152/japplphysiol.00956.2002

DeLorey, D. S., Kowalchuk, J. M., \& Paterson, D. H. (2004). Effects of prior heavyintensity exercise on pulmonary O 2 uptake and muscle deoxygenation kinetics in young and older adult humans. Journal of Applied Physiology (Bethesda, Md. : 1985), 97(3), 998-1005. doi:10.1152/japplphysiol.01280.2003

DeLorey, D. S., Kowalchuk, J. M., \& Paterson, D. H. (2005). Adaptation of pulmonary O2 uptake kinetics and muscle deoxygenation at the onset of heavy-intensity exercise in young and older adults. Journal of Applied Physiology (Bethesda, Md. : 1985), 98(5), 1697-1704. doi:10.1152/japplphysiol.00607.2004

Essen, B., Hagenfeldt, L., \& Kaijser, L. (1977). Utilization of blood-borne and intramuscular substrates during continuous and intermittent exercise in man. Journal of Physiology, 265, 489-506.

Folkow, B., Gaskell, P., \& Waaler, B. a. (1970). Blood flow through limb muscles during heavy rhythmic exercise. Acta Physiologica Scandinavica, 80(1), 61-72.

Grassi, B., Hogan, M. C., Greenhaff, P. L., Hamann, J. J., Kelley, K. M., Aschenbach, W. G., ... Gladden, L. B. (2002). Oxygen uptake on-kinetics in dog gastrocnemius in situ following activation of pyruvate dehydrogenase by dichloroacetate. The Journal of Physiology, 538(Pt 1), 195-207. doi:10.1113/jphysiol.2001.012984

Grassi, B., Rossiter, H. B., Hogan, M. C., Howlett, R. a, Harris, J. E., Goodwin, M. L., Gladden, L. B. (2011). Faster $\mathrm{O}_{2}$ uptake kinetics in canine skeletal muscle in situ after acute creatine kinase inhibition. The Journal of Physiology, 589(Pt 1), 221233. doi:10.1113/jphysiol.2010.195164

Greenhaff, P. L. (2001). The creatine-phosphocreatine system: there's more than one song in its repetoire. Journal of Physiology, 537(3), 657.

Gurd, B., Peters, S., Heigenhauser, G., Leblanc, P., Doherty, T., Paterson, D., \& Kowalchuk, J. (2009). Prior heavy exercise elevates pyruvate dehydrogenase activity and muscle oxygenation and speeds O 2 uptake kinetics during moderate exercise in older adults. American Journal of ..., 297, 877-884. doi:10.1152/ajpregu.90848.2008.

Howlett, R., Parolin, M. L., Dyck, D. J., Hultman, E., Jones, N. L., Heigenhauser, G. J., \& Spriet, L. L. (1998). Regulation of skeletal muscle glycogen phosphorylase and PDH at varying exercise power outputs. The American Journal of Physiology, 275, R418-R425.

Keir, D. A., Murias, J. M., Paterson, D. H., \& Kowalchuk, J. M. (2014a). Breath-bybreath pulmonary O 2 uptake kinetics: effect of data processing on confidence in estimating model parameters. Experimental Physiology, 99(11), 1511-1522. doi:10.1113/expphysiol.2014.080812

Keir, D. A., Nederveen, J. P., Paterson, D. H., \& Kowalchuk, J. M. (2014b). Pulmonary O2 uptake kinetics during moderate-intensity exercise transitions initiated from low versus elevated metabolic rates: insights from manipulations in cadence. European Journal of Applied Physiology, 114(12), 2655-2665. doi:10.1007/s00421-014-2984-9

Korzeniewski, B. (2003). Regulation of oxidative phosphorylation in different muscles and various experimental conditions. The Biochemical Journal, 375(Pt 3), 799804. doi:10.1042/BJ20030882

Korzeniewski, B., \& Zoladz, J. A. (2004). Factors determining the oxygen consumption rate (VO2) on-kinetics in skeletal muscles. The Biochemical Journal, 379(Pt 3), 703-710. doi:10.1042/BJ20031740

Lamarra, N., Whipp, B. J., Ward, S. A, \& Wasserman, K. (1987). Effect of interbreath fluctuations on characterizing exercise gas exchange kinetics. Journal of Applied Physiology, 62(5), 2003-2012.

Lutjemeier, B. J., Ferreira, L. F., Poole, D. C., Townsend, D., \& Barstow, T. J. (2008). Muscle microvascular hemoglobin concentration and oxygenation within the contraction-relaxation cycle. Respiratory Physiology and Neurobiology, 160(2), 131-138. doi:10.1016/j.resp.2007.09.005

Lutjemeier, B. J., Miura, A., Scheuermann, B. W., Koga, S., Townsend, D. K., \& Barstow, T. J. (2005). Muscle contraction-blood flow interactions during upright knee extension exercise in humans. Journal of Applied Physiology (Bethesda, Md. : 1985), 98(November 2004), 1575-1583. doi:10.1152/japplphysiol.00219.2004

Meyer, R. A., Sweeney, H. L., \& Kushmerick, M. J. (1984). A simple analysis of the "phosphocreatine shuttle". The American Journal of Physiology, 246(5 Pt 1), C365-C377.

Murias, J. M., Kowalchuk, J. M., \& Paterson, D. H. (2010). Speeding of VO2 kinetics with endurance training in old and young men is associated with improved matching of local O2 delivery to muscle O 2 utilization. Journal of Applied Physiology (Bethesda, Md. : 1985), 108(February 2010), 913-922.
doi:10.1152/japplphysiol.01355.2009

Murias, J. M., Spencer, M. D., Pogliaghi, S., \& Paterson, D. H. (2012). Noninvasive estimation of microvascular O 2 provision during exercise on-transients in healthy
young males. AJP: Regulatory, Integrative and Comparative Physiology, 303(8), R815-R823. doi:10.1152/ajpregu.00306.2012

Newcomer, B. R., Boska, M. D., \& Hetherington, H. (1999). T 1 Measurements of 31 P Metabolites in Resting and Exercising Human Gastrocnemius / Soleus Muscle at 1 . 5 Tesla. NMR in Biomedicine, 494(12), 486-494.

Ozyener, F., Rossiter, H. B., Ward, S. a, \& Whipp, B. J. (2001). Influence of exercise intensity on the on- and off-transient kinetics of pulmonary oxygen uptake in humans. The Journal of Physiology, 533(Pt 3), 891-902. doi:10.1111/j.1469-7793.2001.t01-1-00891.x

Paterson, B. Y. D. H., \& Whipp, B. J. (1991). Asymmetries of oxygen uptake transients at the on-and offset of heavy exercise in humans. Journal of Physiology, 443, 575586.

Rådegran, G., \& Saltin, B. (1998). Muscle blood flow at onset of dynamic exercise in humans. The American Journal of Physiology, 274, H314-H322.

Rossiter, H. B., Ward, S. a., Doyle, V. L., Howe, F. a., Griffiths, J. R., \& Whipp, B. J. (1999). Inferences from pulmonary O2 uptake with respect to intramuscular [phosphocreatine] kinetics during moderate exercise in humans. Journal of Physiology, 518, 921-932. doi:10.1111/j.1469-7793.1999.0921p.x

Spencer, M. D., Murias, J. M., \& Paterson, D. H. (2012). Characterizing the profile of muscle deoxygenation during ramp incremental exercise in young men. European Journal of Applied Physiology, 112(9), 3349-3360. doi:10.1007/s00421-012-2323y

Spriet, L. L., \& Heigenhauser, G. J. F. (2002). Regulation of pyruvate dehydrogenase (PDH) activity in human skeletal muscle during exercise. Exercise and Sport Sciences Reviews, 30(2), 91-95.

Spriet, L. L., Howlett, R. a, \& Heigenhauser, G. J. (2000). An enzymatic approach to lactate production in human skeletal muscle during exercise. Medicine and Science in Sports and Exercise, 32(4), 756-763.
Swanson, G. D. (1980). Breath-to-breath considerations for gas exchange kinetics. Exercise, bioenergetics and gas exchange. Amsterdam: Elsevier/North Holland, 211-222.

Walløe, L., \& Wesche, J. (1988). Time course and magnitude of blood flow changes in the human quadriceps muscles during and following rhythmic exercise. The Journal of Physiology, 405, 257-273.

Whipp, B. J., Higgenbotham, M. B., \& Cobb, F. C. (1996). Estimating exercise stroke volume from asymptotic oxygen pulse in humans. Journal of Applied Physiology, 81(6), 2674-2679.

Whipp, B. J., \& Wasserman, K. (1972). Oxygen uptake kinetics for various intensities of constant-load work. Journal of Applied Physiology (Bethesda, Md. : 1985), 33(September), 351-356.

# APPENDICES 

## Appendix A 1. Ethics Approval Notice

## Western

## Researchse of Human Participanns - Intital Ethics Approval Notice

Principal Investigator:Dr. Glen Belfry
File Number:104580
Review Level:Full Board
Protocol Title:The effects of short work versus longer work periods within intermittent exercise on the acute
responses of energy system contribution, lactate threshold, cardiac output and regional blood flow
distribution
Department \& Institution:Health Sciences K Kinesiology, Western University
Sponsor:
Ethics Approval Date:January 02,2014
Ethics Expiry Date:May 31,2015
Documents Reviewed \& Approved \& Documents Received for Information:

| Document Name | Comments | Version Date |
| :--- | :--- | :--- |
| Western University Protocol |  |  |
| Response to Board Recommendations |  | $2013 / 11 / 21$ |
| Advertisement |  | $2013 / 11 / 21$ |
| Advertisement | $2013 / 12 / 11$ |  |
| Response to Board Recommendations |  | $2013 / 12 / 11$ |
| Revised Letter of Information \& Consent | $2013 / 12 / 11$ |  |

This is to notify you that the University of Western Ontario Health Sciences Research Ethics Board (HSREB which is organized and operates according to the Tri-Council Policy Statement: Ethical Conduct of Research Involving Humans and the Health Canada/ICH Good Clinical Practice Practices: Consolidated Guidelines; and the applicable laws and regulations of Ontario has reviewed and granted approval to the above referenced study on the approval date noted above. The membership of this HSREB also complies with the membership requirements for REB's as defined in Division 5 of the Food and Drug Regulations.

The ethics approval for this study shall remain valid until the expiry date noted above assuming timely and acceptable responses to the HSREB's periodic requests for surveillance and monitoring information. If you require an updated approval notice prior to that time you must request it using the University of Western Ontario Updated Approval Request form

Member of the HSREB that are named as investigators in research studies, or declare a conflict of interest, do not participate in discussions related to, nor vote on, such studies when they are presented to the HSREB.

The Chair of the HSREB is Dr. Joseph Gilbert. The HSREB is registered with the U.S. Department of Health \& Human Services under the IRB registration number IRB 00000940.

| Etyics Officer to Contact for Further Information |  |  |  |
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## Publications:

