

Gene expression profiling of gilthead sea bream during early development and detection of stress-related genes by the application of cDNA microarray technology

Elena Sarropoulou,^{1,3} Georgios Kotoulas,¹ Deborah M. Power,² and Robert Geisler³

¹Hellenic Center for Marine Research, Institute for Marine Biology and Genetics, Iraklio, Crete, Greece; ²Center for Marine Science, Universidade do Algarve, Faro, Portugal; and ³Max-Planck-Institut für Entwicklungsbiologie, Tübingen, Germany

Submitted 14 June 2005; accepted in final form 25 July 2005

Sarropoulou, Elena, Georgios Kotoulas, Deborah M. Power, and Robert Geisler. Gene expression profiling of gilthead sea bream during early development and detection of stress-related genes by the application of cDNA microarray technology. *Physiol Genomics* 23: 182–191, 2005. First published July 26, 2005; doi:10.1152/physiolgenomics.00139.2005.—Large-scale gene expression studies were performed for one of the main European aquaculture species, the gilthead sea bream *Sparus auratus* L. For this purpose, a cDNA microarray containing 10,176 clones from a cDNA library of mixed embryonic and larval stages was constructed. In addition to its importance for aquaculture, the taxonomic position and the relatively small genome size of sea bream makes it a prospective model for evolutionary biology and comparative genomics. However, so far, no large-scale analysis of gene expression exists for this species. In the present study, gene expression was analyzed in gilthead sea bream during early development, a significant period in the determination of quantitative traits and therefore of considerable interest for aquaculture. Synexpression groups expressed primarily early and late in development were determined and were composed of both known and novel genes. Furthermore, it was possible to identify stress response genes induced by cortisol injections using the cDNA microarray generated. The creation of gene expression profiles for sea bream by microarray hybridization will accelerate identification of candidate genes involved in multifactorial traits and certain regulatory pathways and will also contribute to a better understanding of the genetic background of fish physiology, which may help to improve aquaculture practices.

Sparus auratus; stress response

IN RECENT YEARS, much effort has been invested in developing genome resources. Continuous improvements in technology have allowed rapid progress in the functional interpretation of sequencing data and the investigation of gene expression. The development of technology in model species has paved the way for high-throughput studies at a molecular level of non-model organisms of evolutionary, environmental, or economic interest.

The gilthead sea bream *Sparus auratus* is an organism of both economic and evolutionary interest, and it is one of the main European aquaculture species along with the Atlantic salmon (*Salmo salar*), rainbow trout (*Oncorhynchus mykiss*), oyster (*Crassostrea gigas*), and sea bass (*Dicentrarchus labrax*). In

addition to a basic understanding of the biology and reproductive physiology of this species, well-characterized culture methodologies for its reproduction and rearing also exist. From an evolutionary viewpoint, the gilthead sea bream is of interest as it belongs to the Perciformes, an order that underwent an explosive radiation 60 million years ago and contains a large number of other teleost taxa.

DNA microarray technology has become one of the significant tools in large-scale gene expression studies. For non-model fish species in which sequence data are not extensively available, only a few large-scale gene expression studies exist. For example, in the goby fish *Gillichthys mirabilis*, the common carp *Cyprinus carpio*, and the Atlantic salmon *S. salar*, large-scale gene expression has been explored, respectively, in response to hypoxia, cold, and bacterial infection (19, 20, 49). Microarrays have also been used to study variation in gene expression within and among populations of the genus *Fundulus* (43). In contrast, extensive work setting up gene expression studies for functional analysis has been carried out in model organisms like *Drosophila melanogaster* (15, 41, 55), *Saccharomyces cerevisiae* (23), the mouse *Mus musculus* (54), the frog *Xenopus laevis* (1), *Caenorhabditis elegans* (25), and the zebrafish *Danio rerio* (36, 52).

Here, we report the construction and use of a cDNA microarray for the gilthead sea bream containing 10,176 cDNA clones [several thousand of which are estimated to be unique expressed sequence tags (ESTs)], which were obtained from a mixed embryonic/larval cDNA library (51). In addition, we provide insight into the developmental program of sea bream by characterizing gene expression profiles for five independent stages of sea bream development: neurula (*stage 1*), hatching (*stage 2*), pectoral budding/eye forming (*stage 4*), mouth opening (*stage 6*), and eye pigmentation (*stage 7*). Three main groups of genes were identified by *K* means clustering. More detailed analysis within each of the three groups led to the definition of expression models of up- or downregulated genes during early development as well as genes up- or downregulated before the period of mouth opening. The cDNA microarray chip developed was further validated and used to characterize the gene expression profile in the kidney of juvenile gilthead sea bream after treatment with cortisol. Identification in the present study of several genes previously reported to be up- or downregulated under stress conditions and during development in other teleost species as well as in gilthead sea bream corroborated the data set generated.

The results significantly scale up the exploration of the genome of a commercially important species for which ge-

Article published online before print. See web site for date of publication (<http://physiolgenomics.physiology.org>).

Address for reprint requests and other correspondence: E. Sarropoulou, Hellenic Centre for Marine Research Crete, Thalassocosmos, Former US Base at Gournes, Gournes Peditados, 715 00 Heraklion, Crete, Greece (e-mail: sarris@her.hcmr.gr).

nome information is scarce and provide first insights into the processes during growth and differentiation of the gilthead sea bream as well as during stress conditions.

MATERIALS AND METHODS

(Note that in microarray experiments, the labeled cDNA to be hybridized is called "target" and the DNA samples spotted on the microarray are called "probes.")

Microarray Construction

Sample collection. Embryonic and larval stages of gilthead sea bream were collected from the breeding stock of the aquaculture facility at the Department of Aquaculture of the Hellenic Center of Marine Research (Crete, Greece), snap frozen in liquid nitrogen, and stored at -80°C . Kidneys were collected from untreated juvenile gilthead sea bream ($n = 4$) and from fish ($n = 4$) that received for 72 h a coconut oil implant containing 10 mg/kg (fish wet wt) cortisol. Experiments were carried out at the University of the Algarve (Faro, Portugal) in accordance with national legislation for the welfare of animals. Experiments were conducted in two 125-liter cylindrical tanks supplied with a continuous through flow of oxygenated seawater at $20 \pm 1^{\circ}\text{C}$ using juvenile gilthead sea bream (25 ± 3 g) adapted for 1 wk to the experimental conditions. One tank contained eight untreated fish (control), and the other tank contained eight cortisol-treated fish. At the end of the experiment, fish were removed from tanks and decapitated, and the kidneys rapidly removed and placed in RNAlater (Qiagen) at -20°C . No mortality occurred in the control tank, but two fish died in the cortisol-treated tank.

Bacterial cultures. For microarray production, a cDNA library of mixed embryos/larvae of gilthead sea bream (51) was used. Colonies were obtained by mass excision using the Lambda UNI-ZAP XR cloning Kit (Stratagene; La Jolla, USA) following the manufacturer's instructions and were plated onto a 24×30 -cm agar plate using 3- μm glass beads (Sigma-Aldrich; Munich, Germany). The colonies were picked into 384-square well plates filled with 95 μl of Hogness medium and grown overnight at 37°C without being shaken. A total of 10,176 clones was arrayed in 384-square well plates using the BioPick robotic system (BioRobotics; Cambridge, UK). Clones arrayed in 384-well plates were used to inoculate 4×96 -deep well plates filled with 2 ml TY and 50 $\mu\text{g}/\text{ml}$ kanamycin. They were grown overnight at 37°C with shaking and used for PCRs.

Culture PCR. The total volume of one PCR was 50 μl and contained 43.5 μl of reaction mix (2 μl of $10\times$ PCR buffer; 0.04 μl of 100 mM of each dATP, dCTP, dGTP, and dTTP; and 12.12 μl distilled H_2O), 1.5 μl of T7, 1.5 μl of T3 primer (20 μM), 1 μl of *Taq* polymerase (5 U/ μl), and 2.5 μl of bacterial culture. The thermocycle utilized was 94°C for 1 min 20 s, followed by 35 cycles of 94°C for 1 min, 50°C for 1 min, 72°C for 1 min 30 s, and a final step of 72°C for 5 min. PCRs were performed in duplicate to decrease the failure rate of reactions; duplicates were pooled, separated by agarose gel (2%) electrophoresis, and visualized by transillumination in the presence of ethidium bromide to check the insert size. Purification of PCR products was performed using the GFX-96 PCR Purification Kit (Amersham Pharmacia) according to the manufacturer's instructions. In brief, 300 μl of binding buffer were added to the PCR products, mixed thoroughly, and transferred to the wells of the GFX-96 plate. The assembled plates were centrifuged at 1,800 g for 2 min, the liquid in the wash plate discarded, and the GFX-96 plate was reassembled on the top of the same wash plate. A second wash was conducted by adding 400 μl of wash buffer to each well of the GFX-96 plate and centrifuging for 5 min at 1,800 g; wash liquid was then discarded, and the GFX-96 plate was centrifuged for a further 30 s at 1,800 g to remove the remaining wash buffer. To recover the PCR products, the GFX-96 plate was assembled on the top of the sample collection plate, 50 μl of elution buffer were pipetted into the center of each well, and

samples were incubated for 1 min and centrifuged at 1,800 g for 2 min. The purified products were dried under vacuum, reeluted in $3\times$ SSC and 1.5 M betaine to a volume of 10 μl , and transferred back to 384-well plates.

Spotting. Three rounds of spotting were performed: one round of 192 spots to test hybridization and spotting conditions, one round of 3,456 cDNA clones spotted in duplicate, and finally a third round of 10,176 cDNA clones again spotted in duplicate.

Test slides were spotted with the Affymetrix 417 Arrayer configured with 12 pins and rings. The final spotting was performed with the OmniGrid 100 robotic system (San Carlos, CA) configured with 48 pins (TeleChem SMP3; Sunnyvale, CA).

In total, 100 GAPII glass slides were spotted, with each glass slide contained 10,176 PCR-amplified inserts of the clones picked from the embryonic/larval cDNA library. The amplified cDNA products were spotted randomly and in duplicate in two separate subarrays, such that duplicate probe spots were not in the same quadrant but the next but one. As a control for hybridization and background, only buffer was spotted at the end of each spotting group. The spotted glass slides were baked at 80°C for 2 h to cross-link the DNA onto the glass slide. The slides were then incubated in blocking buffer at 50°C for 30 min, washed twice with Milli-Q water at room temperature for 2 min, and dried by spinning in an Eppendorf 5810R centrifuge for 7 min at 800 rpm.

Data Acquisition

RNA extraction. Total RNA was extracted from five larval stages (neurula, newly hatched, pectoral budding, open mouth, and eye pigmentation stages) and also from cortisol-treated and untreated juvenile gilthead sea bream kidneys using the mini-RNAeasy extraction kit (Qiagen). The quality of the total RNA obtained was checked by gel electrophoresis on a 1% agarose gel containing 0.05% ethidium bromide (10 mg/ml), and the concentration was determined by spectrophotometry.

Labeling. Ten micrograms of intact total RNA were indirectly labeled following the TIGR protocol (<http://www.tigr.org/tdb/microarray/protocolsTIGR.shtml>) with the red fluorescent dye ester cyanine 5 (Cy5) and green fluorescent dye ester cyanine 3 (Cy3) (Amersham Pharmacia). In brief, cDNA was synthesized by incorporating aminoallyl dUTP (AA-dUTP). Synthesis took place at 42°C for 3 h. RNA was hydrolyzed with 10 μl of 1 M NaOH and 10 μl of 0.5 M EDTA for 15 min at 65°C and neutralized by adding 10 μl of 1 M HCl. To remove unincorporated AA-dUTP and free amines, the reactions were purified with Qiagen columns using phosphate wash buffer and phosphate elution buffer. The purified sample was dried in a Speed Vac. AA-cDNA was resuspended in 0.1 M sodium carbonate buffer, and coupling to the Cy dye ester was performed for 1 h in the dark at room temperature. The removal of uncoupled dye was performed using the Qiagen PCR Purification Kit. Finally, samples were tested for successful dye ester incorporation by spectrophotometry. The labeling was considered successful when total dye incorporation was over 200 pmol and the ratio of nucleotides to dye molecules was <50 .

Hybridization. The two targets (Cy3 and Cy5 labeled) were mixed, and the volume was increased to 500 μl with $1\times$ Tris-EDTA buffer (TE). The mixture was applied to a microcon 30 spin filter column and centrifuged for 13 min at 8,000 rpm to reduce the target volume to 15 μl . To the target DNA, 2 μl of 10 mg/ml herring sperm DNA were added, and the volume was increased to 15 μl with $1\times$ TE. The target was denatured for 2 min at 95°C , incubated on ice for 1 min, and finally left at room temperature for 5 min. An equal volume (15 μl) of hybridization buffer was added to the target; the solution was vortexed, spun down, and carefully pipetted onto a coverslip; and the microarray was inverted and placed in a hybridization chamber containing a prewetted Whatman paper strip. Hybridization took place overnight in a water bath at 42°C .

Washing. After hybridization, the chip was carefully taken out of the chamber, and the coverslip was removed by immersion in $4\times$ SSC. The chip was then washed for 5 min in wash *buffer 1* ($2\times$ SSC, 0.1% SDS, and Milli-Q water), 5 min in wash *buffer 2* ($0.1\times$ SSC, 0.1% SDS, and Milli-Q water), and twice for 2.5 min in wash *buffer 3* ($0.1\times$ SSC and Milli-Q water). All washing steps were carried out at room temperature with vigorous shaking. Finally, the slides were quickly dipped in $0.05\times$ SSC and in Milli-Q water. Drying of the slides was performed by centrifugation at $1,000\text{ g}$ for 12 min in a 50-ml Falcon tube at room temperature. All the above steps were performed in the dark.

Data processing and analysis. Microarray gene expression data for embryonic and larval stages were obtained by comparing the results of each stage with each other using a loop design (Fig. 1). All experiments were performed by dye swap hybridizations of the sea bream cDNA microarray chip. Taking into account all possible combinations, 20 hybridizations were performed in total within this experimental setup.

For the study of treated versus untreated kidneys, a total of four hybridizations was performed. The labeling of total RNA from the kidney was performed using a sample from one individual.

For each cDNA microarray chip, two series of digital images were produced by dual laser scanning with a GMS 418 Array Scanner (MWG). The first series was obtained by illuminating the array at 635 nm (excitation of Cy5), and the second series was obtained by illuminating the array at 532 nm (Cy3). For both illuminations, three to four different settings of the photomultiplier tube (gain and light amplification) and two settings (90% and 100%) of the laser power were used. All images were captured in TIFF format.

Spot finding, image segmentation, and quantification was carried out with ImaGene 5.0 microarray image analysis software (BioDiscovery; Los Angeles, CA). Briefly, a grid was created taking into account the number of spots, the print pattern, and the size of the spots. Spot finding was performed using a semiautomatic method in which each spot is checked for proper alignment within the grid manually, as this gives more accurate data collection than fully automatic methods. Finally, spot quantification was performed to obtain the expression level of each gene deposited on the array. For downstream analysis, Cy3 and Cy5 scans were chosen according to the distribution of the Cy3-to-Cy5 ratio after spot quantification (ImaGene 5.0 software package). A ratio of around 1 is expected because the majority of genes do not change expression.

Data set preparation and quality control. The quantified data were analyzed using the software program GeneSight lite included within the ImaGene 5.0 software package. For each data set, the background was subtracted, and poor, empty, or negative spots (see the ImaGene

5.0 manual) were flagged as unreliable and were excluded from downstream analysis. For each data set, the ratio was calculated and normalization was performed using the z transform logarithm of GeneSight lite. The normalized data were stored in a relational database using FileMaker Pro5 software (FileMaker; Santa Clara, CA). Genes selected as being differentially regulated were those in which the ratio of experiment to control was at least 2σ away from the mean value obtained.

The correlation of measurements was determined using the results for duplicate spots on the array.

Raw and normalized data were submitted to ArrayExpress, a public repository for microarray gene expression data at the European Bioinformatics Institute (experiment: E-MEXP-181, array: A-MEXP-110), as well as to the National Center for Biotechnology Information (NCBI) Omnibus under Accession Nos. GSE 2064 and GSE 1887; all clones are available on request. All sequence data have been submitted to the NCBI EST database (Accession Nos. CB184056–CB184594 and CV133223–CV133736).

Analysis. Normalized data were further analyzed using K means clustering and univariate analysis with the statistical software package SPSS 12.0 (SPSS; Chicago, IL).

The K means clustering method is the most appropriate clustering method when analyzing populations of more than 200 data points and was used to group all data points obtained setting cluster numbers from 2 to 5. Distances were computed using simple Euclidean distance. The cluster-cluster distance was calculated by determining the distance between centroids. Expression patterns of genes over the course of the developmental stages were obtained by univariate analysis using the statistical package SPSS.

RESULTS

Microarray Construction

As a first approach toward gene expression studies in sea bream, a cDNA microarray of an embryonic/larval cDNA library was constructed. First, preliminary cDNA microarrays were constructed with 192 cDNA clones spotted 5 times each (data not shown); a second round of test slides containing 3,392 cDNA clones was also prepared. These slides were used to establish and validate the microarray generated. Probes containing zebrafish PCR products of *distalless* (*dlx3*) and genomic sea bream DNA were included as controls. Hybridizations were carried out with the following targets: 1) labeled 50-mer oligonucleotides designed from previously sequenced gilthead sea bream clones, 2) labeled PCR products of gilthead sea bream *dlx3*, 3) gilthead sea bream ribosomal protein L13a, 4) zebrafish *nacre*, 5) zebrafish *dlx3*, and 6) labeled total RNA extracted from two different larval gilthead sea bream stages. The zebrafish targets hybridized only with the spotted probes from zebrafish, indicating the stringency of the hybridization conditions utilized. Cross-hybridization between gilthead sea bream cDNA probes and the zebrafish target was not observed. On the basis of these preliminary tests, a final cDNA microarray containing a total of 10,176 cDNAs was printed in duplicate onto aminosilane-coated slides from Corning. Of the 10,176 spotted cDNA clones, 768 had previously been sequenced (51). It is expected that several thousand different cDNA clones are represented on the chip, as around 75% of the previously sequenced clones were unique. The correlation, determined before normalization, between duplicate spots used for downstream analysis in each hybridization subsequently was found on average to be 85%. The quality of the array was indicated by the high R^2 values obtained after univariate

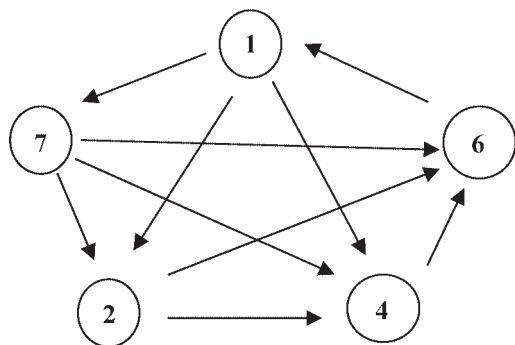


Fig. 1. Experimental design of the microarray experiment in this study (direct or loop design). Five different developmental stages, neurula (*stage 1*), hatching (48 h postfertilization; *stage 2*), pectoral fin budding (96 h postfertilization; *stage 4*), mouth opening (144 h postfertilization; *stage 6*), and eye pigmentation (168 h postfertilization; *stage 7*), were compared directly with each other in all possible combinations.

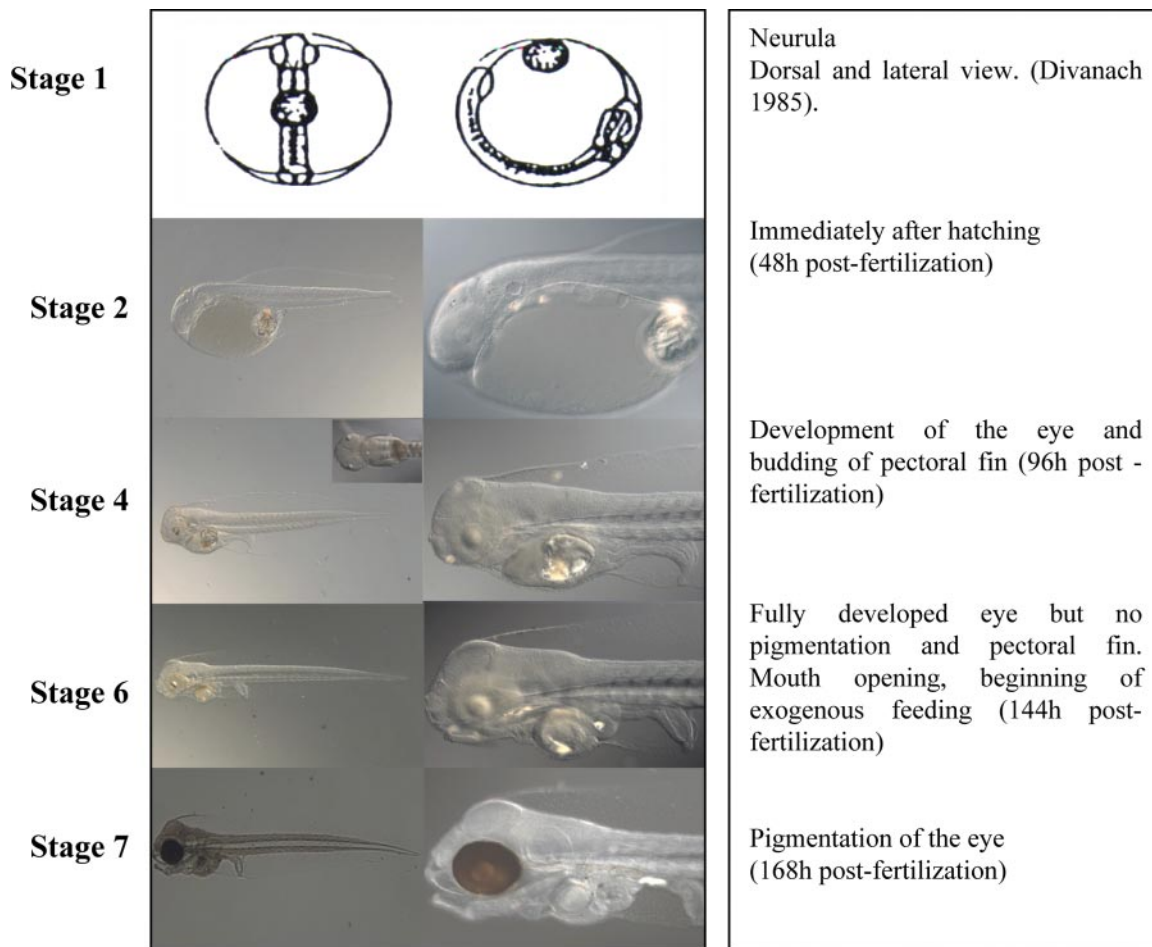


Fig. 2. Pictures of gilthead sea bream larvae at the significant time points during development that were chosen for transcription profiling.

analysis of genes represented more than once in the array (e.g., Fig. 6B) and by the comparable expression results of known genes in the present study and previously published studies.

Isolation of Candidate Genes Involved in Development

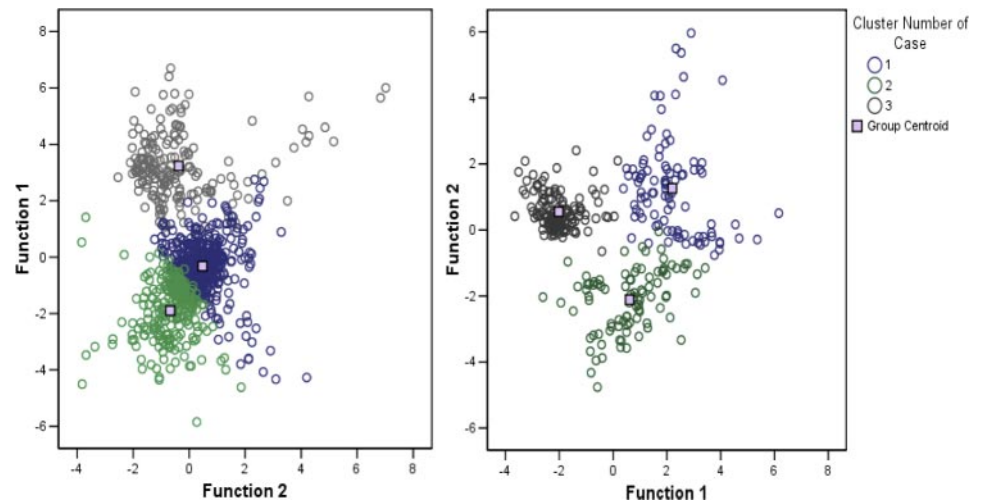
To obtain an overview of genes involved in sea bream development, targets were chosen that represented five time points covering different developmental events from neurula through to pigmentation of the eye (Fig. 2). A group of cDNA clones out of each hybridization, for which z scores of $x \leq -2$ or $x \geq 2$ were detected, were picked, sequenced, and analyzed using BLASTN and BLASTX searches against the NCBI nr database. Sequence alignments resulting from the BLASTN search against the nr database were examined, and a sequence identity of $>65\%$ over a minimum range of 80 bp and an E value of $<10^{-5}$ were chosen as the criteria for designation of putative gene identity (cf. Supplement A; available at the *Physiological Genomics* web site).¹ When the minimum alignment was <80 bp, BLASTX analysis was also performed to confirm the BLASTN result. Some cDNA clones with a low BLASTN e value and no hit in BLASTX were also selected as

they shared a similar expression pattern, also showing the quality of the hybridizations [e.g., SA-E33D_A08, SA-E42D_C10, SA-E60B_A11, and SA-E60C_B11 (all clones assigned to bj AU301079.1 *C. carpio* cDNA clone: 3-033, expressed in barbel)]. Subsequent sequence analysis revealed that the four cDNA clones encoded the same gene, and, in BLASTX, the assembled sequence shared sequence similarity with lipocalin-type prostaglandin D synthase-like protein (*D. rerio*). The above procedure was performed for all clones with a low e value and a similar expression pattern which by BLASTN were assigned to a specific clone.

Frequently used methods for grouping genes by their expression patterns over the experimental variables are hierarchical clustering and K means clustering. In this study, K means clustering was performed on the data points obtained after z score normalization, as this is more suitable for analyzing populations of more than 200 genes (21). Clustering of all sequenced clones on the chip [clones already known from Sarropoulou et al. (51) and clones selected by z score values] before the selection step by microarray hybridization revealed the largest F values throughout the 10 experiments when the cluster number was set to 3, indicating the greatest separation between clusters (Fig. 3) (SPSS). When only genes showing significant differential expression using the z transform method in at least 1 of the 10 experiments were taken into

¹ The Supplemental Material for this article (Supplements A and B) is available online at <http://physiolgenomics.physiology.org/cgi/content/full/00139.2005/DC1>.

Fig. 3. Canonical discrimination functions. *A* and *B*: clustering of all sequenced clones (*A*) and clustering of clones with differential expression identified via a z score of $x \leq -2$ or $x \geq 2$ by the K means method (*B*). The data points given (*cases 1–3*) were grouped in three main clusters (colored blue, green, and gray, respectively). The group centroid represents the average value of the cases contained in each cluster.



account, K means cluster analysis again revealed the largest F values when the cluster number was set to 3 (Table 1 and Fig. 3).

The three clusters represent, respectively, genes that are mainly expressed early in development, genes mainly expressed in later developmental stages, and genes showing differential regulation in the middle of the stages selected for hybridization. Expression profiles were visualized using the software TreeView (12), which plots the experiments on the x -axis and the differentially expressed genes on the y -axis. The colors displayed indicate whether the gene is up- or downregulated. In this study, as shown in Fig. 4, the green color indicates that genes are downregulated in the later developmental stages considered and upregulated in the earlier developmental stages.

Isolation of Candidate Genes Involved in Stress Responses

Microarray experiments were performed with kidneys of cortisol-injected fish versus control fish to identify known and novel genes potentially involved in the stress response induced by increased circulating cortisol levels. These experiments revealed 257 significantly regulated genes, including several genes related to carbohydrate metabolism as well as genes involved in Na^+/K^+ transport, polyamine biosynthesis, and iron homeostasis. In total, 206 clones were sequenced, and 35% did not show a significant BLAST hit (Supplement B).

DISCUSSION

In the present study, insight into the developmental program of the gilthead sea bream is established by the construction and

Table 1. F values of K means clustering setting K to 2–5

	$F(K=5)$	$F(K=4)$	$F(K=3)$	$F(K=2)$
st7_1	687.125	916.502	746.112	1,438.439
st6_1	506.404	678.135	604.256	1,174.903
st4_1	446.923	599.907	714.731	769.158
st2_1	437.775	569.543	552.328	1,172.915
st7_2	304.721	399.919	319.417	142.207
st6_2	230.470	296.790	160.452	281.984
st4_2	94.199	126.572	130.262	306.470
st7_4	186.944	235.830	267.165	31.486
st6_4	189.521	215.469	156.491	55.146
st7_6	68.071	95.263	151.784	4.003

Largest F values (>100) are indicated in bold.

application of a cDNA microarray from embryos and larvae of the gilthead sea bream. Furthermore, this platform was also validated for use in studies of the stress response in gilthead sea bream.

To identify genes involved in the development of sea bream, five different developmental stages (neurula, hatching, pectoral budding/eye development, mouth opening, and eye pigmentation) were chosen for the first round of experiments. Microarray studies providing specific gene expression patterns have been widely used to isolate genes and as a sensitive indicator of gene function (for a review, see Ref. 57). Most of the microarray experiments use an indirect design (also called reference design) as it has the advantage of allowing easy analysis and interpretation without the need for extensive statistical tools. However, if subtle variations between the experiments are of interest, then a design with direct comparison (also called a balanced or loop design) is preferable (56) as it allows comparison of experiments with much greater precision (26, 27). In the present study, a direct experimental design (Fig. 1) for the five different developmental stages of the gilthead sea bream (Fig. 2) were used. To corroborate microarray data, it is desirable to confirm the specificity of a microarray chip as well as the hybridization and analysis method used by analyzing the expression profiles of known genes (50). In this context, several approaches were taken, including comparison of expression profiles with previous studies in other fish species and comparison from a functional genomics viewpoint of the results for developmental stages with those of cortisol-injected fish as well as specific functional studies of genes found in the literature. As expected, genes encoding the translation machinery (according to the gene ontology classification of cellular components) were only scantily represented among the significantly regulated genes in the experiment, which is in direct contrast to results previously obtained with an EST-based study of the same library (51). Similar microarray experiments performed for developmental stages of zebrafish (36, 52, 53) indicated the increased expression of genes involved in myogenesis at the 24-h stage, corresponding to the period of pharyngula, and the 72-h stage, corresponding to the hatching period. These results are congruent with the present results in which there is an upregulation of genes encoding myogenic proteins at *stage 2*, when hatching occurs (Fig. 5). The results obtained from our stress response

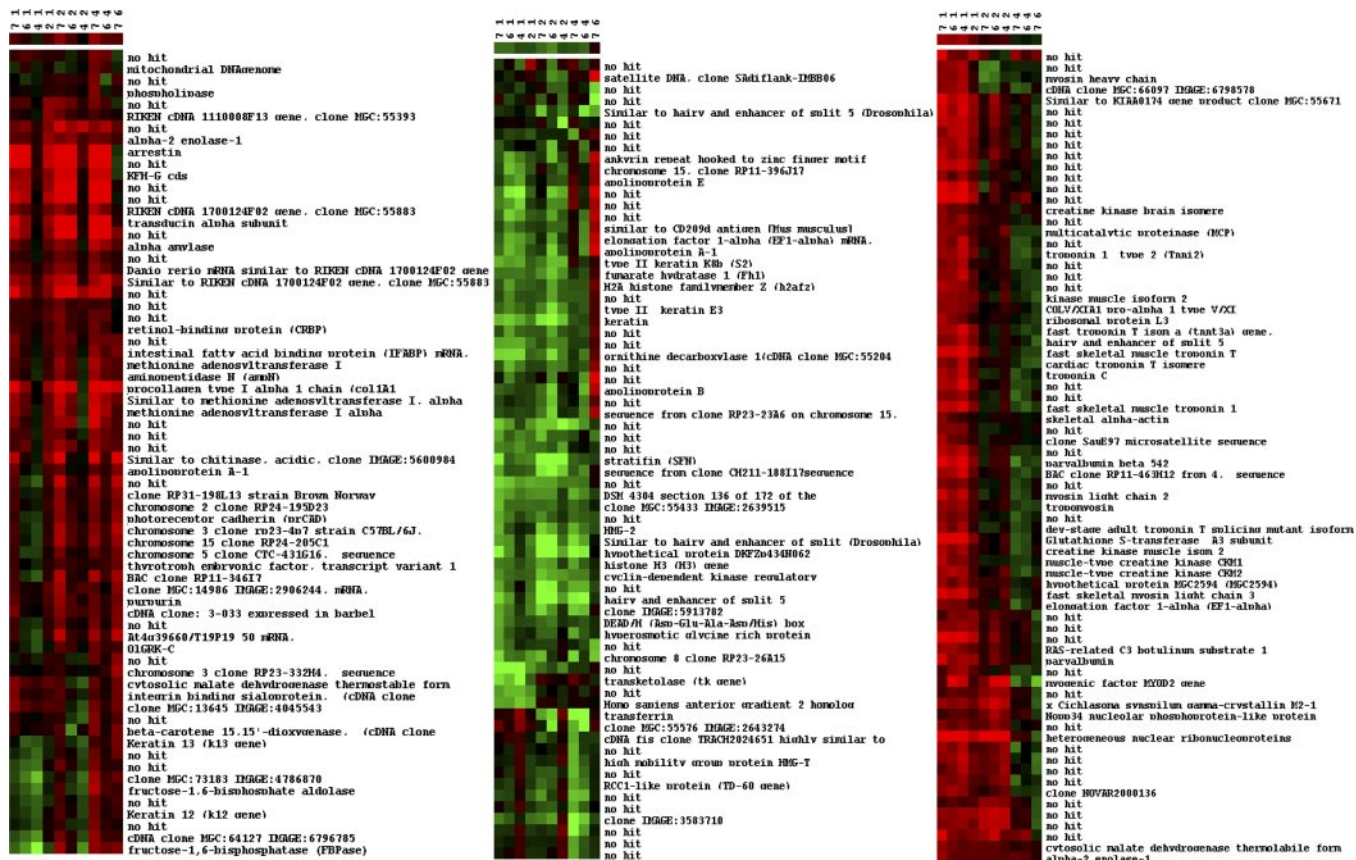


Fig. 4. Heat map images showing the genes grouped by the *K* means clustering method in *cluster 1* (genes expressed mainly in the middle and later ones of the selected developmental stages), *cluster 2* (genes expressed mainly in the early ones of the selected developmental stages), and *cluster 3* (genes up- or downregulated transiently in the middle of the selected developmental stages). Stages are indicated above each column. Gene expression is shown in rows. The quantitative changes in gene expression are represented in color: red indicates upregulation in the later stage, whereas green indicates upregulation in the earlier stage of the hybridization experiment.

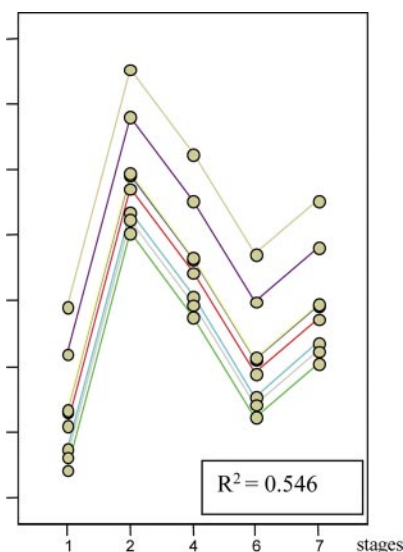


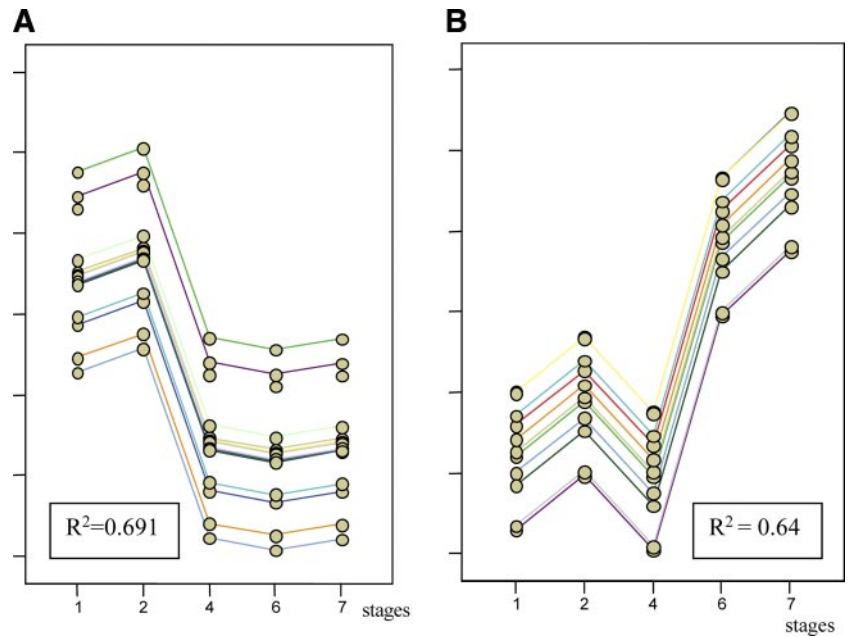
Fig. 5. Genes involved in myogenesis, showing upregulation at *stage 2*, when the larva has just hatched. On the *x*-axis, the stages are plotted, and, on the *y*-axis, the estimated marginal means are plotted. The univariate analysis of variance for randomized design uses the criterion of the multiple R^2 statistics measure. If all the independent variables fit on the regression line, then R^2 is >0.5 .

hybridizations (discussed further below) also corroborate the microarray data for developmental stages. In addition, bibliographic corroboration of the data obtained in this study was also found, e.g., a gene belonging to the enolase superfamily was upregulated in the stress microarray study, as reported in a previous study (48) of α -enolase expression under stress conditions. Furthermore, as a nonnormalized cDNA library was spotted, several genes were represented as multiple duplicates on the same chip, and the results showed good reproducibility of the up- or downregulation of specific genes after clustering and applying statistical analysis by univariate analysis of variance (21).

Cluster Analysis

Cluster analysis revealed three main groups of genes with stage-specific expression. *Cluster 1* contains genes showing upregulation in the last two stages as well as genes showing upregulation in *stage 4*, with a sustained high level of expression in the later stages. Among the genes showing upregulation in the last two stages, *stage 6* (mouth opening) and *stage 7* (eye pigmentation), many candidate genes for visual pigmentation were found, which is in line with phenotypic observations of eye pigmentation [168 hours postfertilization (hpf) (Fig. 6)]. In contrast, many of the genes that are found to be upregulated in

Fig. 6. *A*: expression of genes upregulated early in development. *B*: genes upregulated at stages 6 and 7, encoding proteins involved in eye pigmentation. On the *x*-axis, the stages are plotted, and, on the *y*-axis, the estimated marginal means are plotted. The univariate analysis of variance for randomized design uses the criterion of the multiple R^2 statistics measure. This is the square of the correlation between the dependent and independent variables. If all the independent variables fit on the regression line, then R^2 is >0.5 .



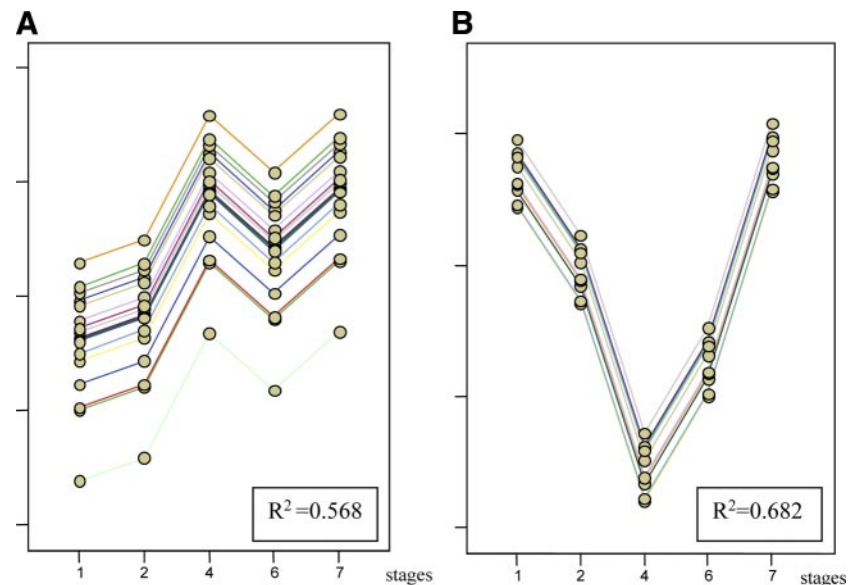
stage 4 encode metabolic enzymes, studies of which are of special interest for fish physiology and aquaculture.

In fish larvae, nutrients are initially supplied only by the yolk reserve, although at mouth opening and the onset of exogenous feeding nutrients are absorbed across the rudimentary digestive tract. A major problem for aquaculture is the low acceptance of artificial diets by larval fish. Large variations in ingestion rates are observed in feeding experiments, and, in some cases, first-feeding marine fish larvae ingest artificial feed but fail to grow. Advances in this area have produced better results (9, 13, 29, 31, 34), but, in most species, problems arising from the use of formulated feeds at the onset of exogenous feeding still persist. The larvae of gilthead sea bream hatch from pelagic eggs, and full development of the digestive system only occurs at metamorphosis (40). Determination of the onset of enzyme production is therefore essential

for choosing appropriate larval diets and establishing the earliest possible time for weaning.

In the present study, many genes encoding metabolic enzymes including amylase are upregulated before mouth opening, namely, in *stage 4* (Fig. 7), which corresponds to 4 days posthatched. At this time, the pectoral fin is budding and the eye is beginning to form. Studies of the amylase gene in the winter flounder *Pleuronectes americanus* show that the onset of amylase expression is at a similar stage, 5 days posthatched (10). Interestingly, in the present study, both fructose-1,6-bisphosphatase and fructose-1,6-aldolase B, enzymes involved in gluconeogenesis, are downregulated at *stage 4*, just when amylase is upregulated (Fig. 7). This may indicate a concerted switch from gluconeogenesis, when nutrients come from yolk lipids, to the glycolytic pathway as soon as exogenous carbohydrates become available. Although amylase activity in *D.*

Fig. 7. *A*: genes encoding enzymes with an expression peak at *stage 4*. *B*: expression profile of two of the key enzymes involved in the production of glucose, fructose-1,6-bisphosphatase and fructose-1,6-bisphosphate aldolase B (data from 8 and 4 measurements, respectively). On the *x*-axis, the stages are plotted, and, on the *y*-axis, the estimated marginal means are plotted. The univariate analysis of variance for randomized design uses the criterion of the multiple R^2 statistics measure. This is the square of the correlation between the dependent and independent variables. If all the independent variables fit on the regression line, then R^2 is >0.5 .



melanogaster is repressed by glucose and induced by starch (7, 24), the expression changes observed in this study must be developmentally controlled as they occur before the onset of exogenous feeding at stage 6. Isolation and expression profiling of further genes encoding digestive enzymes throughout development, and, in particular, peptidases, will generate knowledge about the metabolic capabilities of different larval stages and contribute to the development of suitable diets. Furthermore, microarray analysis of feeding experiments should help to clarify the role played by inductive processes during the development of the digestive system and help to monitor the acceptance of artificial food to larval fish.

Cluster 2 contains genes that are downregulated in the later stages. Compared with the other clusters, genes encoding proteins involved in epithelial development were more abundant (Fig. 6). Among them are proteins of the largest subfamily of intermediate filament proteins, type I and type II keratins. Heterodimers of type I and type II keratins act as the main meshwork for the keratin cytoskeleton.

Cluster 3 (Figs. 4 and 5) contains genes transiently upregulated around the time of hatching (stage 2). A comparison of *cluster 3* to *clusters 1* and *2* revealed a greater number of genes involved in myogenesis, such as myosin light chain 2 and 3, myosin heavy chain, and tropomyosin, which were also reported to be upregulated during the hatching period in expression profiling studies of zebrafish (36, 52).

Expression of these genes coincides with the period in which abnormalities in intensively reared sea bream become evident. Moreover, it has been shown that abnormalities, such as opercular deficiency (8), morphological deformities (2, 30), and failure to form a functional swimbladder (28, 45), are to a large extent responsible for reduced growth and a higher mortality rate (46). Identification and further study of genes expressed during this period in healthy and abnormal larvae may help to improve the performance of subsequent rearing stages.

Isolation of Genes Exposed to Stress Conditions

Fish in aquaculture are frequently exposed to a range of stressors. Stress can be induced by overcrowding, inappropriate culture temperatures, overfeeding, poor water quality, and exposure to toxicants (22). Under stress conditions, fish are more susceptible to infection, e.g., by *Streptococcus iniae* (14), and to other diseases. Stress increases the metabolic rate and oxygen uptake in fish (6), and serum cortisol is elevated. The rise in cortisol associated with stress is responsible for activation of the central nervous system, increasing blood glucose concentration and elevating mean blood pressure, all of which contribute to coping with stress (6, 44). Corticosteroids, in addition to their influence on gluconeogenesis and glycolysis, also affect the activity of enzymes involved in metabolic regulation (for a review, see Ref. 38), influence osmoregulation (33), and induce profound immunosuppression (6).

In the present study, the administration of cortisol was used to simulate stress, and it was possible to establish for the first time in gilthead sea bream the transcriptional response in the kidney. Among the genes upregulated in this study are several genes involved in gluconeogenesis, glycolysis, and the respiratory chain, including fructose-1,6-bisphosphatase, fructose 1,6-bisphosphate aldolase B, and α_2 -enolase-1. This is in

agreement with previous findings showing that fructose-1,6-bisphosphatase activity is upregulated in carp treated with cortisol (11) and that α -enolase of *G. mirabilis* as well as α -enolase of gilthead sea bream is upregulated under stress conditions (18, 48). Genes encoding proteins involved in Na^+/K^+ transport were also found to be upregulated, which is in concordance with studies showing that cortisol stimulates branchial Na^+/K^+ -ATPase activity, an enzyme crucial for osmoregulation (37).

A number of genes encoding proteins of interest were found to be up- or downregulated under stress conditions in the present study. One such transcript encodes ornithine decarboxylase (ODC), a key enzyme of polyamine biosynthesis that is elevated in rapidly proliferating cells. Overexpression of ODC has been associated with carcinogenesis in mammals (4, 39). ODC is also a marker of liver neoplasia in winter flounder (*P. americanus*) (32), and its upregulation in stressed gilthead sea bream may cause a predisposition to tumors, an issue of some importance for aquaculture. Interestingly, two of the main proteins in iron metabolism, ferritin heavy and light subunit and the transferrin receptor, were downregulated by cortisol. Oxidative stress and iron metabolism are interrelated in multiple ways. Ferritin is an intracellular molecule that stores iron in a soluble nontoxic form and was found to be induced in response to hypoxia in a gene expression profiling study in *G. mirabilis* (18) and repressed in response to quinone-induced oxidative stress in a study of murine B6 fibroblasts (17). The transferrin receptor is an iron transport protein that was likewise found to be repressed in murine fibroblasts in response to oxidative stress (17). Because it is known that intracellular iron can be a rate-limiting factor for cell growth and proliferation (35), a downregulation of intracellular iron levels may be associated with the suppression of growth observed under stress conditions. Furthermore, elevated levels of transferrin receptor were found in Atlantic salmon macrophages activated by infection with *Piscirickettsia salmonis* (49), suggesting that its downregulation could contribute to immunosuppression. In humans, transferrin is used as a clinical marker as its concentration level correlates with the total mass of immature erythroid cells (47). Potentially, it could also be employed as a diagnostic marker in aquaculture. For this purpose, it will be important to determine whether the decrease of the transferrin receptor seen during stress primarily reflects a decrease in the number of hematopoietic cells or a lowering of intracellular iron levels.

Another area to be explored in future work is the use of whole fry as an effective sample of stress response as well as the effect of stress on specific developmental stages of sea bream as several stress-responding genes isolated in this study were also found to be regulated during development.

In conclusion, the present study provides a first glimpse of how the global analysis of gene expression, using material from appropriately chosen conditions or tissues as a target, can yield insight into the molecular basis of development and stress in aquaculture species. The data set presented here will be a basis for future expression profiling work in this area. It has been proposed that genes with similar biological function constitute synexpression groups with coordinated expression throughout development (16, 41), an expectation that is borne out by our clustering studies, e.g., of proteins encoding for epithelial tissue. This opens up the possibility of focusing

sequencing efforts on unknown genes with expression profiles similar to known genes of interest to identify new genes that have similar functions. Such a strategy will enrich genomic resources for the gilthead sea bream in a more efficient way than random sequencing of ESTs could and promises to provide specific candidate genes for quantitative trait loci analysis and marker-assisted selection. The identification of candidate genes is of particular importance for marine fish species in which only very little information about the genetic source of phenotypic variation exists, in contrast to other agricultural animals. Finally, expression profiling will facilitate studies of comparative physiology and comparative genomics beyond the traditional model species, possibly yielding new insights into the evolution of animal physiology (19).

ACKNOWLEDGMENTS

We thank Dr. M. Pankratz and lab for providing the microarray spotting facilities and Dr. C. Seiler for support in generating pictures of the developmental stages of sea bream.

Sequences reported in this article have been submitted to the National Center for Biotechnology Information (NCBI) EST database under Accession Nos. CB184056–CB184594 and CV133223–CV133736. Microarray expression data have been submitted to ARRAYExpress under Accession Nos. E-MEXP-181 (experiment) and A-MEXP-110 (array) as well as to the NCBI Omnibus under Accession Nos. GSE 2064 and GSE 1887.

GRANTS

This study was funded by the BRIDGEMAP-project (EU Contract Q5R-2001-01797) as part of the European Commission's Fifth Framework Programme "Quality of Life and Management of Living Resources."

REFERENCES

- Altmann CR, Bell E, Sczyrba A, Pun J, Bekiranov S, Gaasterland T, and Brivanlou AH. Microarray-based analysis of early development in *Xenopus laevis*. *Dev Biol* 236: 64–75, 2001.
- Andrades JA, Becerra J, and Fernandez-Llebrez P. Skeletal deformities in larval, juvenile and adult stages of cultured gilthead sea bream (*Sparus aurata*). *Aquaculture* 141: 1–11, 1995.
- Ashburner M, Ball CA, Blake JA, Botstein D, Butler H, Cherry JM, Davis AP, Dolinski K, Dwight SS, Eppig JT, Harris MA, Hill DP, Issel-Tarver L, Kasarskis A, Lewis S, Matese JC, Richardson JE, Ringwald M, Rubin GM, and Sherlock G. Gene ontology: tool for the unification of biology. The Gene Ontology Consortium. *Nat Genet* 25: 25–29, 2000.
- Auvinen M, Paasinen A, Andersson LC, and Holtta E. Ornithine decarboxylase activity is critical for cell transformation. *Nature* 360: 355–358, 1992.
- Bamberger CM, Schulte HM, and Chrousos GP. Molecular determinants of glucocorticoid receptor function and tissue sensitivity to glucocorticoids. *Endocr Rev* 17: 245–261, 1996.
- Barton BA, Schreck CB, and Barton LD, and Baanante IV. Effects of chronic cortisol administration and daily acute stress on growth, physiological conditions, and stress responses in juvenile rainbow trout. *Dis Aquat Org* 2: 173–185, 1987.
- Benkel BF and Hickey DA. Glucose repression of amylase gene expression in *Drosophila melanogaster*. *Genetics* 114: 943–954, 1986.
- Beraldo P, Pinosa M, Tibaldi E, and Canavese B. Abnormalities of the operculum in gilthead sea bream (*Sparus aurata*): morphological description. *Aquaculture* 220: 89–99, 2003.
- Cahu CL and Zambonino Infante JL. Substitution of live food by formulated diets in marine fish larvae. *Aquaculture* 200: 161–180, 2001.
- Douglas SE, Mandla S, and Gallant JW. Molecular analysis of the amylase gene and its expression during development in the winter flounder, *Pleuronectes americanus*. *Aquaculture* 190: 247–260, 2000.
- Dziewulska-Szwajkowska D, Lozinska-Gabska M, Adamowicz A, Wojtaszek J, and Dzugaj A. The effect of high dose of cortisol on glucose-6-phosphatase and fructose-1,6-bisphosphatase activity, and glucose and fructose-2,6-bisphosphate concentration in carp tissues (*Cyprinus carpio* L.). *Comp Biochem Physiol B Biochem Mol Biol* 135: 485–491, 2003.
- Eisen MB, Spellman PT, Brown PO, and Botstein D. Cluster analysis and display of genome-wide expression patterns. *Proc Natl Acad Sci USA* 95: 14863–14868, 1998.
- Fernández-Díaz C and Yüfera M. Detecting growth in gilthead sea-bream, *Sparus aurata* L.; larvae fed microcapsules. *Aquaculture* 153: 193–202, 1997.
- Fuller JD, Bast DJ, Nizet V, Low DE, and de Azavedo JC. *Streptococcus iniae* virulence is associated with a distinct genetic profile. *Infect Immun* 69: 1994–2000, 2001.
- Furlong EE, Andersen EC, Null B, White KP, and Scott MP. Patterns of gene expression during *Drosophila* mesoderm development. *Science* 293: 1629–1633, 2001.
- Gawantka V, Pollet N, Delius H, Vingron M, Pfister R, Nitsch R, Blumenstock C, and Niehrs C. Gene expression screening in *Xenopus* identifies molecular pathways, predicts gene function and provides a global view of embryonic patterning. *Mech Dev* 77: 95–141, 1998.
- Gehring NH, Hentze MW, and Pantopoulos K. Inactivation of both RNA binding and aconitase activities of iron regulatory protein-1 by quinone-induced oxidative stress. *J Biol Chem* 274: 6219–6225, 1999.
- Gracey AY, Troll JV, and Somero GN. Hypoxia-induced gene expression profiling in the euryoxic fish *Gillichthys mirabilis*. *Proc Natl Acad Sci USA* 98: 1993–1998, 2001.
- Gracey AY and Cossins AR. Application of microarray technology in environmental and comparative physiology. *Annu Rev Physiol* 65: 231–259, 2003.
- Gracey AY, Fraser EJ, Li W, Fang Y, Taylor RR, Rogers J, Brass A, and Cossins AR. Coping with cold: an integrative, multitissue analysis of the transcriptome of a poikilothermic vertebrate. *Proc Natl Acad Sci USA* 101: 16970–16975, 2004.
- Hair JF, Anderson RE, Tatham RL, and Black WC. *Multivariate Data Analysis* (5th ed.). Upper Saddle Hall, NJ: Prentice-Hall, 1998.
- Hontela A, Rasmussen JB, Audet C, and Chevalier G. Impaired cortisol stress response in fish from environments polluted by PAHs, PCBs, and mercury. *Arch Environ Contam Toxicol* 22: 278–283, 1992.
- Hughes TR, Mao M, Jones AR, Burchard J, Marton MJ, Shannon KW, Lefkowitz SM, Ziman M, Schelter JM, Meyer MR, Kobayashi S, Davis C, Dai H, He YD, Stephanianis SB, Cavet G, Walker WL, West A, Coffey E, Shoemaker DD, Stoughton R, Blanchard AP, Friend SH, and Linsley PS. Expression profiling using microarrays fabricated by an ink-jet oligonucleotide synthesizer. *Nat Biotechnol* 19: 342–347, 2001.
- Inomata N, Kanda K, Cariou ML, Tachida H, and Yamazaki T. Evolution of the response patterns to dietary carbohydrates and the developmental differentiation of gene expression of alpha-amylase in *Drosophila*. *J Mol Evol* 41: 1076–1084, 1995.
- Jiang M, Ryu J, Kiraly M, Duke K, Reinke V, and Kim SK. Genome-wide analysis of developmental and sex-regulated gene expression profiles in *Caenorhabditis elegans*. *Proc Natl Acad Sci USA* 98: 218–223, 2001.
- Kerr MK and Churchill GA. Experimental design for gene expression microarrays. *Biostatistics* 2: 183–201, 2001.
- Kerr MK and Churchill GA. Statistical design and the analysis of gene expression microarray data. *Genet Res* 77: 123–128, 2001.
- Kitajima C, Tsukashima Y, Fujita S, Watanabe T, and Yone Y. Relationship between uninflated swimbladders and lordotic deformity in hatchery reared sea bream *Pagrus major*. *Bull Jpn Soc Sci Fish* 47: 1289–1294, 1981.
- Kolkovski S. Digestive enzymes in fish larvae and juveniles—implications and applications to formulated diets. *Aquaculture* 200: 181–200, 2001.
- Koumoundouros G, Gagliardi F, Divanach P, Boglione C, Cataudella S, and Kentouri M. Normal and abnormal osteological development of caudal fin in *Sparus aurata* L. fry. *Aquaculture* 149: 215–226, 1997.
- Koven W, Kolkovski S, Hadas E, and Gamsiz KTA. Advances in the development of microdiets for gilthead sea bream, *Sparus aurata*: a review. *Aquaculture* 194: 107–121, 2001.
- Koza RA, Moore MJ, and Stegeman JJ. Elevated ornithine decarboxylase activity, polyamines and cell proliferation in neoplastic and vacuolated liver cells of winter flounder (*Pleuronectes americanus*). *Carcinogenesis* 14: 399–405, 1993.
- Laiz-Carrión R, Martín Del Río MP, Miguez JM, Mancera JM, and Soengas JL. Influence of cortisol on osmoregulation and energy metabolism in gilthead sea bream *Sparus aurata*. *J Exp Zool* 298: 105–118, 2003.
- Lazo JP, Dinis MT, Holt JG, Faulk C, and Arnold CR. Co-feeding microparticulate diets with algae: towards eliminating the need of zoo-

- plankton at first feeding red drum (*Sciaenops ocellatus*). *Aquaculture* 188: 339–351, 2000.
35. **Li ZR, Hromchak R, Mudipalli A, and Bloch A.** Tumor suppressor proteins as regulators of cell differentiation. *Cancer Res* 58: 4282–4287, 1998.
 36. **Lo J, Lee S, Xu M, Liu F, Ruan H, Eun A, He Y, Ma W, Wang W, Wen Z, and Peng J.** 15000 unique zebrafish EST clusters and their future use in microarray for profiling gene expression patterns during embryogenesis. *Genome Res* 13: 455–466, 2003.
 37. **McCormick SD, Dickhoff WW, Duston J, Nishioka RS, and Bern HA.** Developmental differences in the responsiveness of gill Na⁺,K⁺-ATPase to cortisol in salmonids. *Gen Comp Endocrinol* 84: 308–317, 1991.
 38. **Mommsen TP, Vijayan MM, and Moon TW.** Cortisol in teleosts: dynamics, mechanisms of action and metabolic regulation. *Rev Fish Biol Fisheries* 9: 211–268, 1999.
 39. **Moshier JA, Dosecu J, Skunca M, and Luk GD.** Transformation of NIH/3T3 cells by ornithine decarboxylase overexpression. *Cancer Res* 53: 2618–2622, 1993.
 40. **Moyano FJ, Diaz M, Alarcon FJ, and Sarasquete MC.** Characterization of digestive enzyme activity during larval development of gilthead seabream *Sparus aurata*. *Fish Physiol Biochem* 15: 121–130, 1996.
 41. **Neal SJ, Gibson ML, So AK, and Westwood JT.** Construction of a cDNA-based microarray for *Drosophila melanogaster*: a comparison of gene transcription profiles from SL2 and Kc167 cells. *Genome* 46: 879–892, 2003.
 42. **Niehrs C and Pollet N.** Synexpression groups in eukaryotes. *Nature* 402: 483–487, 1999.
 43. **Oleksiak MF, Churchill GA, and Crawford DL.** Variation in gene expression within and among natural populations. *Nat Genet* 32: 261–266, 2002.
 44. **Olivotto I, Mosconi G, Maradonna F, Cardinali M, and Carnevali O.** *Diplodus sargus* interrenal-pituitary response: chemical communication in stressed fish. *Gen Comp Endocrinol* 127: 66–70, 2002.
 45. **Paperna I.** Swimbladder and skeletal deformations in hatchery bred *Sparus aurata*. *J Fish Biol* 12: 109–114, 1978.
 46. **Parson-LeRuyet J and Verillaud P.** Technique d'élevage intensif de la daurade dorée (*Sparus aurata* L.) de la naissance à l'âge de deux mois. *Aquaculture* 20: 1980.
 47. **Ponka P and Lok CN.** The transferrin receptor: role in health and disease. *Int J Biochem Cell Biol* 31: 1111–1137, 1999.
 48. **Ribas LPJV, Barton B, Monetti C, Bernadini G, Saroglia M, Tort L, and MacKenzie S.** A differentially expressed enolase gene isolated from the gilthead sea bream (*Sparus aurata*) under high-density conditions is up-regulated in brain after in vivo lipopolysaccharide challenge. *Aquaculture* 241: 195–206, 2004.
 49. **Rise ML, Jones SR, Brown GD, Von Schalburg KR, Davidson WS, and Koop BF.** Microarray analyses identify molecular biomarkers of Atlantic salmon macrophage and hematopoietic kidney response to *Piscirickettsia salmonis* infection. *Physiol Genomics* 20: 21–35, 2004.
 50. **Rockett JC and Hellmann GM.** Confirming microarray data—is it really necessary? *Genomics* 83: 541–549, 2004.
 51. **Sarropoulou E, Power DM, Magoulas A, Geisler R, and Kotoulas G.** Comparative analysis and characterization of expressed sequence tags in gilthead sea bream (*Sparus aurata*) liver and embryos. *Aquaculture* 243: 69–81, 2005.
 52. **Ton C, Stamatiou D, Dzau VJ, and Liew CC.** Construction of a zebrafish cDNA microarray: gene expression profiling of the zebrafish during development. *Biochem Biophys Res Commun* 296: 1134–1142, 2002.
 53. **Ton C, Stamatiou D, and Liew CC.** Gene expression profile of zebrafish exposed to hypoxia during development. *Physiol Genomics* 13: 97–106, 2003.
 54. **Wang QT, Piotrowska K, Ciemerych MA, Milenkovic L, Scott MP, Davis RW, and Zernicka-Goetz M.** A genome-wide study of gene activity reveals developmental signaling pathways in the preimplantation mouse embryo. *Dev Cell* 6: 133–144, 2004.
 55. **White KP, Rifkin SA, Hurban P, and Hogness DS.** Microarray analysis of *Drosophila* development during metamorphosis. *Science* 286: 2179–2184, 1999.
 56. **Yang YH and Speed T.** Design issues for cDNA microarray experiments. *Nat Rev Genet* 3: 579–588, 2002.
 57. **Zhou XJ and Gibson G.** Cross-species comparison of genome-wide expression patterns. *Genome Biol* 5: 232, 2004.