

Dual-mode optical microscope based on single-pixel imaging

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Abstract

We demonstrate an inverted microscope that can image specimens in both reflection and transmission modes simultaneously with a single light source. The microscope utilizes a digital micromirror device (DMD) for patterned illumination altogether with two single-pixel photosensors for efficient light detection. The system, a scan-less device with no moving parts, works by sequential projection of a set of binary intensity patterns onto the sample that are codified onto a modified commercial DMD. Data to be displayed are geometrically transformed before written into a memory cell to cancel optical artifacts coming from the diamond-like shaped structure of the micromirror array. The 24-bit color depth of the display is fully exploited to increase the frame rate by a factor of 24, which makes the technique practicable for real samples. Our commercial DMD-based LED-illumination is cost effective and can be easily coupled as an add-on module for already existing inverted microscopes. The reflection and transmission information provided by our dual microscope complement each other and can be useful for imaging non-uniform samples and to prevent self-shadowing effects.

Keywords: Microscopy, Reflection, Transmission, Resolution, Computational imaging, Spatial Light Modulators

1. Introduction

Optical microscopy techniques have become essential tools in basic and applied research. For instance, in the biological realm, fluorescence microscopy is the method of choice for imaging proteins and their nano-scale dynamic organization. Despite impressive improvements in optical resolution during the last decades [1, 2, 3], there is still room to improve a range of other features such as the trade-off between the field of view and the depth of field [4, 5], 3D imaging

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of specimens [6], or the design of cost-effective, light-weight platforms that can operate in resource-limited settings for global health challenges [7, 8], among
10 others.

Structured illumination microscopy (SIM) stands out among the different optical microscopy techniques because it allows to retrieve spatial frequency information inaccessible with standard uniform illumination schemes [9, 10]. It is based on a standard wide-field optical microscope where the specimen is illumin-
15 ated with a set of known sinusoidal patterns with different phase shifts. The object is reconstructed from multiple acquisitions by using dedicated algorithms. Additionally, this fringe-projection method is attractive because of its intrinsic optical sectioning capability that enables whole-field optically sectioned images [11, 12].

In the conventional implementation of a SIM, a focal plane array detector detects variations in light intensities scattered by the sample with spatial resolution. Other structured illumination approaches have also been implemented for microscopy during the past few years. The single-pixel imaging (SPI) scheme [13] has gained considerable attention as a very effective sensing mechanism and
25 has triggered diverse applications where conventional cameras equipped with millions of pixels fail to give an adequate response, including optical microscopy [14]. As a matter of fact, SPI allows imaging at previously undeveloped spectral bands [15, 16], at the photon-counting regime [17], in presence of strong turbulence or scattering [18, 19], and also through multimode optical fibers [20, 21].
30 The combination of wide-field structured illumination altogether with a bucket detector has made hyperspectral imaging across the visible spectrum possible in a fluorescence microscope [22]. Also, a prototype microscope system based on SPI to image simultaneously in the visible and the short-wave infrared has been recently demonstrated [23].

In SPI the problem of spatial resolution is shifted away from the sensor to a set of a microstructured spatial masks that are codified onto a programmable spatial light modulator. The masks are optically projected onto the sample through the microscope objective and the whole intensity is collected onto a bucket (single-pixel) sensor. Measurements are sequential by changing of the
40 spatial mask. If many different masks are used, their shapes and the intensity signals are combined to retrieve the sample. This generalized mask scanning offers several advantages, like signal-to-noise ratio enhancement and the possibility to reduce the acquisition time through compressive sensing, over the traditional raster scan used in confocal microscopy where a single bright pixel
45 is scanned to build up the image [24, 25].

The nature of SPI enforces a reciprocal relationship between the frame rate and image size as the time required to capture an image scales with the number of pixels in the image. Two different approaches can be employed to deal with this issue. On the one hand, given some reasonable assumptions about the
50 sparsity of the signal, compressive sensing dramatically reduces the number of measurements well below the number of pixels of the sample [26, 27]. What is remarkable here is that with the only "a priori" knowledge about the sparsity of the sample it is possible to get rid off the measurement of the full-length signal,

so it saves time. More recently, adaptive sensing has been introduced as a way to
55 circumvent the computational complexity in convex optimization or greedy algo-
rithms used in compressive sensing [23, 28, 29]. On the other hand, SPI usually
relies on the use of fast spatial light modulators such as the digital micromir-
ror device (DMD) to codify the projecting masks. DMDs permit highly flexible
codification of binary masks at frame rates above 20 kHz. Extensive application
60 of the DMD to microscopy has been reported in the past few years including
conventional SIM microscopy with fringe projection [30], super-resolution and
optical sectioning microscopy [31, 32] and the programmable array microscope
[33, 34, 35]. Interestingly, fast DMD and pattern illumination is at the core
of optogenetics, a tool for noninvasive activation and silencing of neurons and
65 muscles [36, 37].

Along the same lines than SPI microscopy, here we demonstrate a dual-mode
microscope that can image specimens in reflection and transmission modes si-
multaneously. Many specimens, such as biological samples, are weak reflectors
but produce transmission images with good contrast. Inversely, other samples
70 are very dense and provide poor transmission images or generate self-shadowing
effects. Conventional optical microscope designs make the simultaneous collec-
tion of transmitted and reflected light inefficient, restrictive, or even impossi-
ble. In general they need different light sources for transmission and reflection,
thereby preventing that both images be simultaneously measured in a single
75 sensor. Alternatively they can work with several digital cameras, but then a
careful calibration and geometric adjustment of both sensors is necessary. The
usefulness of dual-mode microscopy for histopathology studies of skin tissue
has been recently reported based on a lensless holographic setup [38, 39] and
near-field scanning optical microscopy [40].

80 Here, we demonstrate that the SPI architecture is particularly well-fitted
for this dual operation recording both reflection and transmission information
simultaneously with a single light source and a simple light sensor configura-
tion constituted by two single-pixel detectors. The field of view, the optical
resolution, and the focused plane are determined by the light projection sys-
85 tem. Therefore, the reflection and transmission images obtained by each light
detector are automatically adjusted geometrically with no need of calibration
procedures. Furthermore both images are focused unequivocally to the same
plane of the sample. In summary, in our microscope, reflection information
complements the transmission one very efficiently, which can be very useful for
90 imaging non-uniform samples and to prevent self-shadowing effects.

For the practical implementation, instead of using a high-end DMD as pre-
vious single-pixel microscopy techniques, we use of an off-the-shelf DMD from
a cost-effective digital light projector. These devices utilize an offset diamond
pixel layout which generates geometrical artifacts. In addition to prove the dual
95 mode operation, in this work we demonstrate an algorithm to precisely allocate
pixels in memory to deal with this problem.

2. Dual-mode single-pixel imaging

In a dual-mode SPI microscope, the DMD plane is relayed by a convenient optics onto the sample plane and the forward and backscattered light components are simultaneously collected onto dual single-pixel photodetectors located at the transmission and reflection ports of the microscope, respectively [see Fig. 1]. A sequence of M sampling patterns is codified onto the DMD so that the irradiance striking the photodetector at the transmission port in the i th timestep is

$$Y_i = \langle \Psi_i(\vec{x}), T(\vec{x}) \rangle, \quad (1)$$

where $T(\vec{x})$ is the transmittance distribution of the sample, $\Psi_i(\vec{x})$ denotes the i th measurement pattern, and $\langle \Psi_i(\vec{x}), T(\vec{x}) \rangle$ represents the inner product between both functions. The state of the sampling projector $\Psi_i(\vec{x})$ is changed from one timestep to the next to implement a full set of measurements

$$\mathbf{Y} = \mathbf{S}\mathbf{T}, \quad (2)$$

where \mathbf{S} is a sensing matrix whose i th row is a one-dimensional reshaping of the i th sampling mask $\Psi_i(\vec{x})$, and \mathbf{T} and \mathbf{Y} are N -dimensional vectors representing a one-dimensional reshaping of the unknown transmittance distribution and the result of the measurements at the transmission port, respectively.

Equivalently, the measurements at the reflection port are concisely represented by the series of linear equations

$$\mathbf{Z} = \mathbf{S}\mathbf{R}, \quad (3)$$

where \mathbf{R} and \mathbf{Z} are N -dimensional vectors representing a one-dimensional reshaping of the unknown reflectance distribution and the result of the measurements at the reflection port, respectively,

The problem of the measurement can be stated as: given the result of the measurements \mathbf{Y} and \mathbf{Z} derive the set of values \mathbf{T} and \mathbf{R} that best represent the transmittance and reflectance distributions $T(\vec{x})$ and $R(\vec{x})$ of the sample.

In the easiest implementation of the retrieval algorithm, which is possible for well-conditioned measurements systems, a number of measurements M equal to the number of pixels of the sample is required and both the transmission and the reflection images are retrieved through the inverse matrix as

$$\mathbf{T} = S^{-1}\mathbf{Y} \quad , \quad \mathbf{R} = S^{-1}\mathbf{Z}. \quad (4)$$

Concerning the measurement patterns, various matrices can be employed. For instance, raster-scan style masks stem from the well-known raster-scan technique in which spatial pixels are measured sequentially. Random matrices can also be used in which each mask has a random distribution of binary values. On the other hand the Hadamard S matrix provides a convenient codification framework because binary nonnegative elements can easily be displayed onto the DMD [41]. The Hadamard S matrix is defined through the equation

$$\mathbf{S}_N = \frac{1}{2}(1 - \mathbf{H}_N), \quad (5)$$

where

$$\mathbf{H}_N \mathbf{H}_N^T = N \mathbf{I}_N \quad (6)$$

with \mathbf{H}_N the Hadamard matrix of order N , \mathbf{H}_N^T the transposed matrix, and \mathbf{I}_N the $N \times N$ identity matrix. Also the above choice provides minimum variance
 135 least-squares estimation of the unknown variables. Further, Hadamard measurements obtained by subtracting S -matrix measurements from complementary patterns produce a better result in a similar way to the results achieved through balanced detection [42, 43].

2.1. Compressive sensing retrieval

140 Although there are some benefits of using SPI in optical microscopy, it increases image acquisition time over conventional imaging using focal plane array detectors due to the fact that measurements must be taken sequentially, rather than in parallel. To boost up the image acquisition, we take advantage of the ground-breaking theory of compressive sensing (CS), which makes it possible to
 145 recover an N -pixels image from $M < N$ measurements [24, 25]. In CS, one takes samples of the N -dimensional transmission vector \mathbf{T} using a matrix $M \times N$ matrix S' , obtaining undersampled measurements $\mathbf{Y}' = S' \mathbf{T}$. Along this section, and with no lack of generality, we will refer to the transmission signal, where an identical relation holds for the reflection mode.

150 Although the above equation defines an underdetermined linear relation with a number of equations lower than the number of variables ($M < N$), the restricted isometry property guarantees that the right signal can still be reconstructed if we simply assume it has a low information content compared with its physical dimensions. In mathematical terms, we assume that the signal is
 155 sparse in some basis representation. The problem then becomes how to sample properly with a measurement matrix satisfying certain conditions. For random measurement matrices with independent standard Gaussian entries S' , it is known that convex optimization $\min \|\mathbf{T}\|_1$ subject to $\mathbf{Y}' = S' \mathbf{T}$ typically finds the sparsest solution. In the above equation, $\|\cdot\|_1$ denotes the l_1 norm. Yet,
 160 there are different methods to retrieve the image such as basis pursuit methods and greedy pursuit methods.

Also, it has been shown that deterministic measurement ensembles exhibit the same behavior than Gaussian matrices via l_1 minimization. Analytically known matrices avoid the need to store the entire matrix as the entries can be
 165 computed on the fly and permit to use recovery algorithms with lower complexities. Note that operation at video rates is of paramount significance in applications of SPI in optical microscopy [21,27]. Further, partial Hadamard ensembles where the matrix S' is formed by uniform random selection of M rows of the Hadamard transform matrix allow to acquire biologically relevant

170 samples [20]. In this case, it is essential to enforce the sparsity of the reconstruction signal in some representation with a high level of dissimilarity with the elements of the measurement matrix.

Because measurements are noisy, it is better to relax the minimization constraints to

$$\min \|\mathbf{T}\|_1 \quad \text{subject to} \quad \|\mathbf{Y}' = S'\mathbf{T}\|_2 < \epsilon,$$

175 and ask that the fit holds to the noise level ϵ . The full performance analysis of the compressive sensing methodology in terms of the dynamic range of the sensor, the bit depth of the A/D converter, and the amount of noise was conducted in Reference [13].

In our study, we utilized the l_1 -magic solver [44] to reconstruct the images with partial Hadamard ensemble to form the measurement matrix. Yet, as the power spectrum of most biological images is generally concentrated at low frequencies, we employed a sampling strategy that favours this frequency range. While this is true, biological samples also have fine details that could be lost due to this sampling strategy. However, in this work, for low frequencies we refer to low Hadamard frequencies, which also include high frequencies in Fourier space.
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3. Experimental set-up

In this section we describe how to setup the dual-mode single-pixel microscope by attaching a low-cost commercial DMD and two photodetectors to an inverted microscope. In order to have a low-cost DMD with standard video input control, we remove the optics and the illumination system of an off-the-shelf digital light projector (DLP). We use the illumination system of the commercial microscope and we project the display through the epifluorescence port of the microscope and transfer a demagnified image to the sample plane [Fig. 1]. The inverted microscope used in the experiment is a Nikon Eclipse Ti-U. The system can be divided in two parts: the illumination system and the collection system. The illumination system is composed by a mercury lamp (Nikon, Intensilight C-HGFI), a DMD (DLP LightCrafterTM EVM 0.3 WVGA), two relay lenses ($\varnothing = 2''$ achromatic doublets, with focal lengths 100mm (L1) and 200mm (L2)) producing a 2X magnification, a tube lens and the projection objective (Nikon LU Plan 20X/0.40 WD 13). The DMD is illuminated with a collimated light beam. An intermediate image of the DMD is projected onto the focal plane of the tube lens by the relay lenses and then demagnified by the projection objective onto the sample plane. The two collection systems for reflection and transmission are composed by a condenser lens and a photomultiplier tube PMT (PMM01, Thorlabs Inc.). The reflection PMT is placed in a lateral port and the transmission PMT is placed above the stage of the microscope.
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3.1. Pattern codification on diamond shaped DMDs

The direct implementation of an $N \times N$ image onto the diamond pixel layout differs from the implementation onto the conventional orthogonal pixel layout.

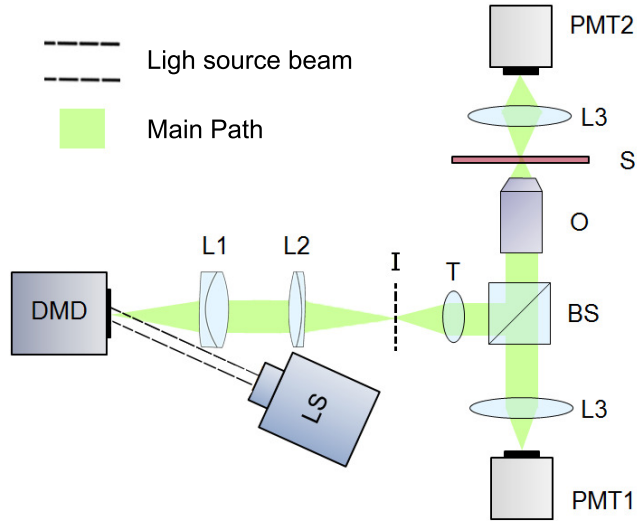


Figure 1: Experimental set-up. (LS) Light Source. (DMD) Digital micromirror device. (L1, L2) Relay lenses. (I) Intermediate image of the DMD. (T) Tube lens. (BS) Beam splitter. (S) Sample plane. (L3) Condenser lenses. (O) Projection objective. Reflection and transmission photomultiplier tubes (PMT1 and PMT2 respectively).

210 In an orthogonal layout the rows and the columns are straight lines, so the aspect ratio is preserved. However in the diamond layout meanwhile the rows are straight lines, the columns are zigzag lines. Thereby the direct implementation changes the original aspect ratio of the image. Moreover, straight edges are transformed on zigzag edges. Both consequences are shown in Fig. 2.

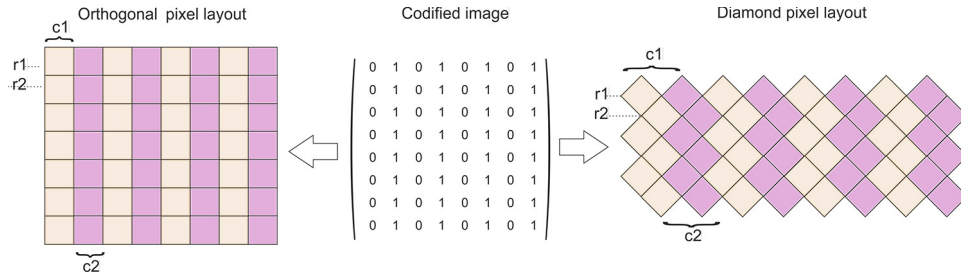


Figure 2: Graphical representation of the different coordinate system for orthogonal pixel layout (left) and diamond pixel layout (right).

215 To appreciate the effects due to miscoding of the patterns in image retrieval within the framework of SPI, we reconstructed the transmission mode of a negative USAF test chart (USAF 1951 2x2) that was used as a sample in the optical setup of Fig. 1. The number of projected test patterns (Hadamard patterns) was 4096, with a resolution of 64×64 pixels, using no compression techniques. The result is shown in Figure 3.

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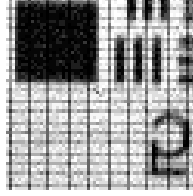


Figure 3: Experimental image reconstructed without pattern predistortion (64×64 pixels) for the transmission mode. Geometrical artifacts (phantom grid) due to the diamond layout are clearly noticeable.

To avoid these artifacts, before sending an image to the DMD, the information must be codified in a different way. The aspect ratio could be preserved pre-distorting the original image but this operation do not correct the zigzag edges artifact. In this work a subtler coding is presented that preserves both the aspect ratio and edges of the original image. An example of the operation of our coding algorithm is presented in Fig. 4(a) and (b) for the case of a 2×2 image. Without correction, information codified in a 2×2 array provides a distorted image in the diamond layout as is shown in Fig. 4(a). Our algorithm codifies the information in an array in such way that original image appears rotated but preserves the aspect ratio and straight edges as shown in Fig. 4(b).

The array transformation produced by the algorithm is shown in Fig. 4(c). In general, the algorithm transforms an $N \times N$ image (where N is even) into a $(2N - 1) \times N$ array, following the next steps:

1. Diagonals are extracted from the initial image in the order shown in Fig. 4(c).
2. Each row of the $(2N - 1) \times N$ image is built by zero-padding each diagonal. The order of the rows matches the order of the extracted diagonals. If the row is even, it is padded to left and right of the diagonal with the same number of zeros. If the row is odd, an additional zero is needed to the left.

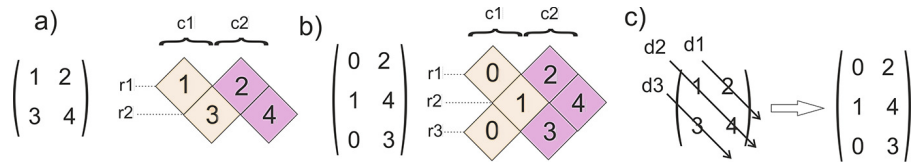


Figure 4: Graphical representation of the matrix transformation applied to the projected patterns to avoid artifacts.

In this way, by means of this previous transformation, the coding of a square image is displayed as a square image rotated 45 degrees, preserving the original aspect ratio and edges.

3.2. Temporal multiplexing of binary patterns

Commercial DMDs from video projectors are slower than scientific DMDs
 245 created specifically for scientific applications, as they work only at video rates
 (60-120 Hz). To improve speed performance, we take advantage of the codifica-
 tion procedure used by the DLP for color video projection. In the standard
 video projection mode, the light changes sequentially to blue, red and green col-
 ors. For each color, the display codifies 8 binary patterns. An improvement in
 250 speed can be done by codifying 24 binary patterns within a single 24-bit depth
 image. This technique allows an improvement in speed rate of 24, going from
 60-120 Hz to 1440-2880 Hz. A schematic overview of the codification is shown
 in Fig. 5. The DMD was fully controlled by software designed by our group in
 LabviewTM. The image processing was performed using MatlabTM.

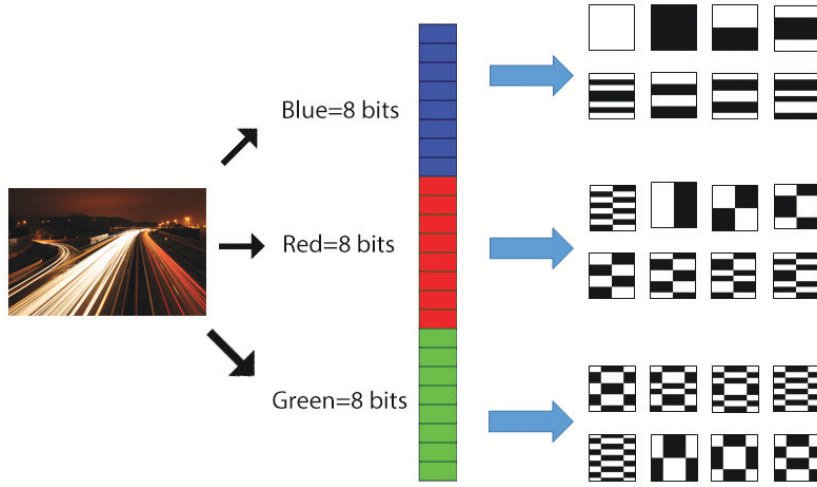


Figure 5: Codification of a 24-bit image with 24 Hadamard patterns of 1-bit depth. This technique allows an improvement in speed rate of 24, going from 60-120 Hz to 1440-2880 Hz.

255 In Fig. 6 we plot the temporal response of the transmission photomultiplier
 tube for a single 24-bit image. The patterns projected and the object under
 measure were the same as in the experiment seen in Fig. 3. Each value for
 Y_i is obtained by averaging the values within the ranges marked by a red line.
 Because the number of patterns exceeds the projector's internal memory we
 260 have to send the patterns through commercial video stream. Through this
 channel, the video projector repeats each image 6 times (for visual purposes).
 The maximum speed rate reached was $1440/6 = 240$ Hz, allowing us to capture
 a 64×64 image (with no compression techniques) in 17 seconds.

4. Results

265 With the aim of analyzing the optical resolution in both transmission and
 reflection modes, a negative USAF test chart (USAF 1951 2x2) is employed as

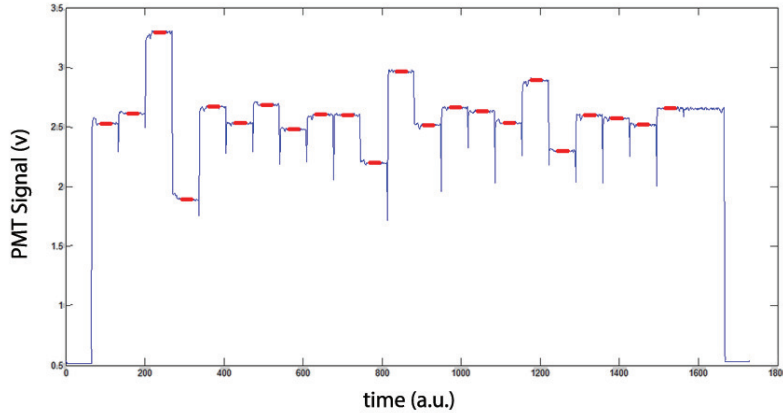


Figure 6: Photocurrent values for a single 24-bit frame. In the graphic the different values for each 1-bit depth pattern codified in the 24-bit image are shown in red color. In this case, the first and last bit are use as reference.

an object in the setup shown in Fig. 1. The microscope objective is a 20X with 0.40 NA. Only the smallest groups of the USAF test, 6 and 7, were used to bound the resolution of the system. The scales and dimensions of the bars are given by the expression R (lp/mm) = $2^{Group+(element-1)/6}$. For example, elements 4 and 5 of group 7 have a resolution of 181.0 lp/mm and 203.2 lp/mm, equivalent to a feature size of $2.76\mu\text{m}$ and $2.46\mu\text{m}$ respectively. The images were obtained as follows. The patterns of the basis were projected onto the object plane by L1 and O, then the intensity transmitted and reflected by the object for each pattern were captured, simultaneously by the two PMTs. The experimental results are shown in Fig. 7. To boost the sensing stage, compressive sensing algorithms were used. Binning is performed by codifying each pixel of the Hadamard patterns with a group of 4×4 micromirrors in the DMD. Pictures (a) and (b) show transmission and reflection images, respectively, by using only 25% of the total number of patterns ($M = 1,012$) for a resolution of 64×64 pixels while (c) and (d) have a 50% compression ($M = 2,046$) for the same resolution. Pictures (e) and (f) were obtained by projecting the total number of patterns, without compression ($M = 4,092$). The results show that the resolution obtained by transmission and reflection are almost identical ($2.46\mu\text{m}$ for the images with no compression) and that reducing the projected patterns to 50% does not significantly affect resolution ($2.76\mu\text{m}$) but allows measuring twice as fast. These two resolutions values are obtained from the last resolvable element of group 7 (5 and 4 respectively). In our experimental set-up, the PMT can detect light signals at frequencies as high as 20 kHz. However, the acquisition frequency is limited to 60 Hz by the graphic card and, therefore, to 1440 Hz by the frame rate of the commercial grade DMD.

The quality of the compressed images was tested using the standard peak signal-to-noise ratio, $PSNR = 10 \log(I_{max}^2/MSE)$, where I_{max} is the maximum

pixel intensity value of the reference image and $MSE = (\frac{1}{N})\|\mathbf{I}_{ref}^{in} - \mathbf{I}_{rec}^{in}\|^2$, where
 295 \mathbf{I}_{rec}^{in} is the undersampled image and \mathbf{I}_{ref}^{in} is the image recovered from the whole
 measurement. The result of this analysis for the case of the reflection mode
 is shown in Fig 8. The figure shows a plot of the PSNR together with the
 reconstructed image for different compression ratios.

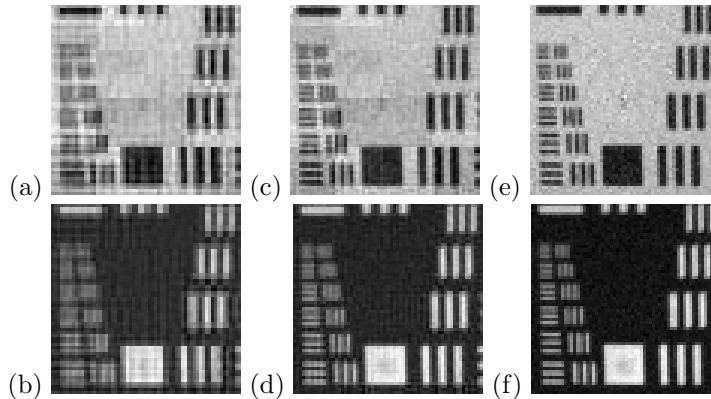


Figure 7: Experimental images of the USAF test (groups 6 and 7) obtained with the dual-mode single-pixel microscope in Fig. 1 using compressive sensing algorithms with a different compression ratio. Pictures (a) and (b) show transmission and reflection images, respectively, by using only 25% of the total number of patterns for this resolution (64×64 pixels) and 50% (c-d) of the whole number of patterns. (e-f) Transmission and reflection images with no compression. In all cases, the size of the image was 64×64 pixels and the field of view $192\mu m \times 192\mu m$.

As a proof of concept, we have also tested the capability of the dual-mode SPI
 300 microscope to image biological samples with different reflective and transmissive
 imaging profiles. The results can be seen in Fig. 9. The first row show images
 of an Anaphothrips obscurus in both reflection (a) and transmission (b) modes
 taken simultaneously. The images have a resolution of 64×64 pixels ($M =$
 4,096, binning = 4×4 micromirrors) and were obtained with a 20X microscope
 305 objective. In this case the transmission image shows only the silhouette of the
 insect because of its low transparency. The second row show images of a Zeuzera
 pyrina wing taken with a single-pixel camera in reflection (c) and transmission
 (d) modes, respectively. Both images have a resolution of 128×128 pixels
 ($M = 16,384$, binning = 2×2 micromirrors). In this case, the transmission
 310 image shows structures of the other side of the wing (hairs), hidden in the
 reflection images. The images of both modes are focused to the same plane
 of the sample and are automatically adjusted geometrically. This can be an
 advantage for biological structural studies. Image (e) show the same area of the
 sample taken in reflection with a CMOS sensor located at the output port of
 315 the microscope. This time the resolution of the image is 200×200 pixels.

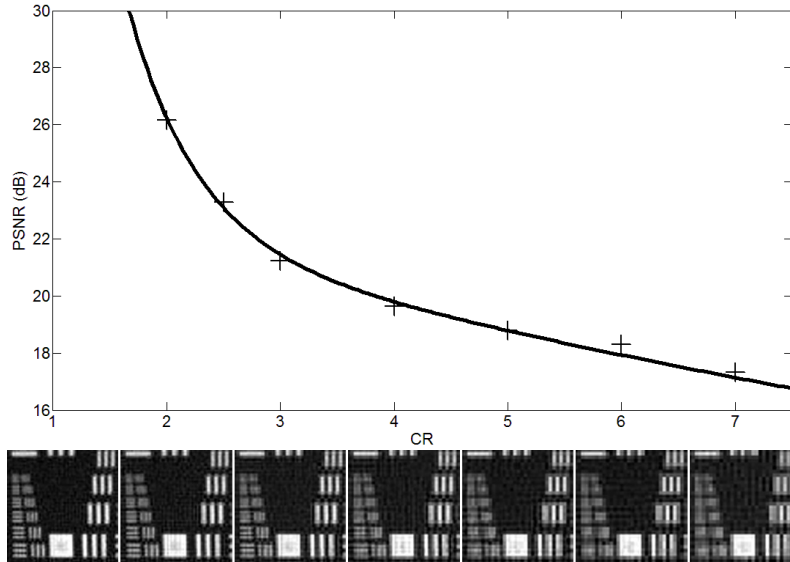


Figure 8: (Top) Plot of the standard peak signal-to-noise ratio (PSNR) in dB as a function of the compression ratio ($CR = N/M$) to show the quality of the recovered images for the case of the reflection mode. (Bottom) Images reconstructed with the different compression ratios associated with the PSNR plotted in the top figure with increasing CR from left to right.

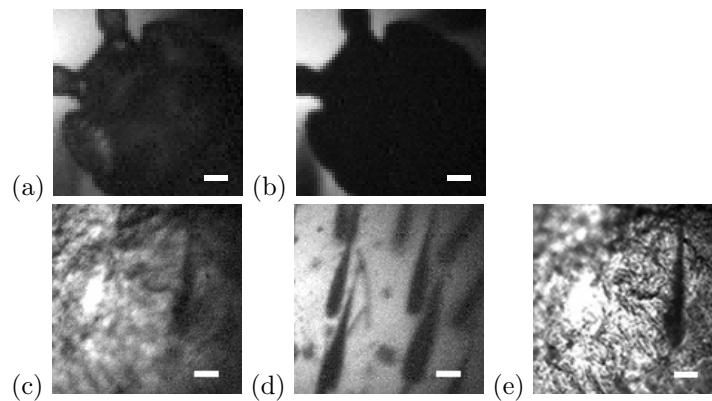


Figure 9: Experimental results. Anaphothrips obscurus in reflection (a) and transmission (b) modes obtained with the optical setup in Fig. 1 using a 20X objective. The images have 64×64 px. Images of a Zeuzera pyrina wing in reflection (c) and transmission (d) modes. In this case the images have a resolution of 128×128 px. (e) Image in reflection taken with a CMOS sensor located at the output port of the microscope. (200×200 px). Scale bar: $25\mu m$.

5. Conclusion

In conclusion, we have developed a dual-mode optical microscope by attaching a commercial DMD and two single-pixel detectors to an inverted microscope. We have designed an algorithm to modify the shape of the projected patterns that improves the resolution and prevents the artifacts produced by the so-called diamond pixel architecture. We have demonstrated that our system is able to retrieve images of reflection and transmission modes with the same field of view and from the same plane of the sample, revealing structures not visible easily by conventional microscopy techniques. This is done with no need of calibration procedures because the field of view, resolution, and focusing plane are controlled exclusively by the properties of the light patterns generated by the optical projection system. By recording transmitted and reflected information loss of data by self-shadowing is minimized which can be advantageous in non-uniform samples containing transparent and opaque regions. Both imaging modes benefit from the advantages present in single-pixel imaging as low-light sensitivity as well as multidimensional capability through the use of specialized sensors. Single-pixel imaging can also be advantageous in the presence of optical aberrations and optical imperfections in between the sample and the sensor. In our case, this is a clear advantage as the mode of transmission has to go through the entire sample before reaching the sensor. Our system benefits also from the advantages of compressive sensing, reducing the measure time by 50% without strongly affecting the resolution of the system. This fact allows us to configure the system with a compromise between acquisition speed and resolution. Additionally, the simplicity of the detection system will allow us to record different spectral channels or polarization states in each imaging mode very easily, increasing the versatility of the system.

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