



# *Review* **Sickle Cell Disease: Current Drug Treatments and Functional Foods with Therapeutic Potential**

**Elisângela Gonçalves <sup>1</sup> , Slim Smaoui <sup>2</sup> [,](https://orcid.org/0000-0002-8839-7377) Miguel Brito 3,4 [,](https://orcid.org/0000-0001-6394-658X) J. M. Oliveira 5,6,7 [,](https://orcid.org/0000-0003-1564-5728) Ana Paula Arez [1](https://orcid.org/0000-0002-0497-6532) and Loleny Tavares 5,6,7,[\\*](https://orcid.org/0000-0002-2112-4702)**

- <sup>1</sup> Global Health and Tropical Medicine (GHTM), Associate Laboratory in Translation and Innovation Towards Global Health (LA-REAL), Institute of Hygiene and Tropical Medicine, (IHMT), NOVA University of Lisbon (UNL) 1349-008 Lisbon, Portugal; a21001585@ihmt.unl.pt (E.G.); aparez@ihmt.unl.pt (A.P.A.)
- <sup>2</sup> Laboratory of Microbial and Enzymes Biotechnology and Biomolecules (LBMEB), Centre of Biotechnology of Sfax (CBS), University of Sfax-Tunisia, Road of Sidi Mansour Km 6, P.O. Box 1177, Sfax 3018, Tunisia; slim.smaoui@cbs.rnrt.tn
- <sup>3</sup> Health Research Centre of Angola (CISA), Caxito, Angola; miguel.brito@estesl.ipl.pt
- <sup>4</sup> H&TRC—Health & Technology Research Center, Escola Superior de Tecnologia da Saúde, Instituto Politécnico de Lisboa, 1990-092 Lisbon, Portugal
- <sup>5</sup> School of Design, Management and Production Technologies Northern Aveiro, University of Aveiro, Estrada do Cercal, 449, 3810-193 Oliveira de Azeméis, Portugal; martinho@ua.pt
- <sup>6</sup> EMaRT Group—Emerging Materials, Research, Technology, University of Aveiro, 3810-193 Aveiro, Portugal
- <sup>7</sup> CICECO Aveiro—Institute of Materials, University of Aveiro, Campus Universitário de Santiago, 3810-193 Aveiro, Portugal
- **\*** Correspondence: tavaresloleny@ua.pt

**Abstract:** Sickle cell anemia (SCA), the most common form of sickle cell disease (SCD), is a genetic blood disorder. Red blood cells break down prematurely, causing anemia and often blocking blood vessels, leading to chronic pain, organ damage, and increased infection risk. SCD arises from a single-nucleotide mutation in the β-globin gene, substituting glutamic acid with valine in the β-globin chain. This review examines treatments evaluated through randomized controlled trials for managing SCD, analyzes the potential of functional foods (dietary components with health benefits) as a complementary strategy, and explores the use of bioactive compounds as functional food ingredients. While randomized trials show promise for certain drugs, functional foods enriched with bioactive compounds also hold therapeutic potential. Further research is needed to confirm clinical efficacy, optimal dosages, and specific effects of these compounds on SCD, potentially offering a cost-effective and accessible approach to managing the disease.

**Keywords:** sickle cell anemia; functional food; randomized; clinical trials; bioactive compounds

## **1. Introduction**

Sickle cell anemia (SCA), a severe genetic disorder that originated in Africa and spread globally through migration, is one of the world's most common inherited blood diseases [\[1,](#page-15-0)[2\]](#page-15-1). The World Health Organization (WHO) estimates that over 300,000 births occur with this condition every year worldwide [\[3\]](#page-15-2). It results from a mutation on human chromosome 11, in which adenine is replaced by thymine [\[1\]](#page-15-0). This change involves a single DNA base-pair substitution (GAG to GTG) in the beta-globin gene. This mutation alters the amino acid sequence of the beta-globin protein, replacing the hydrophilic glutamic acid at position six with the hydrophobic valine, resulting in hemoglobin S (HbS) with morphological abnormalities in the red blood cells in low oxygen pressure (Figure [1\)](#page-1-0) [\[4\]](#page-15-3).

The change in amino acid properties (hydrophilic to hydrophobic) within the β-globin chain contributes to the reduced solubility of HbS [\[5\]](#page-15-4). Therefore, the abnormal hemoglobin exhibits insolubility and polymerization in response to factors such as decreased oxygen tension, physical trauma, dehydration, stress, acidosis, or exposure to cold temperatures.



**Citation:** Gonçalves, E.; Smaoui, S.; Brito, M.; Oliveira, J.M.; Arez, A.P.; Tavares, L. Sickle Cell Disease: Current Drug Treatments and Functional Foods with Therapeutic Potential. *Curr. Issues Mol. Biol.* **2024**, *46*, 5845–5865. [https://doi.org/](https://doi.org/10.3390/cimb46060349) [10.3390/cimb46060349](https://doi.org/10.3390/cimb46060349)

Academic Editor: Claudiu N. Lungu

Received: 14 May 2024 Revised: 6 June 2024 Accepted: 10 June 2024 Published: 12 June 2024



**Copyright:** © 2024 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license [\(https://](https://creativecommons.org/licenses/by/4.0/) [creativecommons.org/licenses/by/](https://creativecommons.org/licenses/by/4.0/)  $4.0/$ ).

Polymerized hemoglobin leads to the formation of rigid, inflexible, and easily damaged erythrocytes. These facts could lead to a reduction in lifespan, contributing to the development of a range of acute and chronic complications [\[6\]](#page-15-5). Studies examining the haplotypes associated with the sickle cell gene in Africa have revealed compelling evidence suggesting three distinct regions of origin for the mutation within the continent: the Central African Republic, Senegal, and Benin [\[1,](#page-15-0)[7,](#page-16-0)[8\]](#page-16-1). Individuals with SCD, homozygous for the βS mutation, typically exhibit the characteristic symptoms and complications associated with the condition. In very early infancy, SCD often presents as asymptomatic. Symptoms typically emerge with the decline in fetal hemoglobin, around 6 to 8 months of age [\[9\]](#page-16-2). However, as individuals age, various factors may intervene, influencing the symptomatic expression of SCD [\[10\]](#page-16-3). Individuals with SCA experience reduced red blood cell flexibility due to polymerization, resulting in rheological and biochemical alterations that impede blood flow, ultimately leading to vaso-occlusive crises (VOCs) [\[10](#page-16-3)[,11\]](#page-16-4). Studies in genetic mapping and genome-wide association have identified specific genetic loci associated with SCA, including B-cell lymphoma/leukemia 11A (BCL11A), an Xmn1 variant located upstream of the hemoglobin subunit gamma 1 (HBG1), and the HBS1L-MYB intergenic region. These loci have been found to contribute to 20–50% of the variability observed in fetal hemoglobin (HbF) levels among individuals with SCA [\[12\]](#page-16-5).

<span id="page-1-0"></span>

**Figure 1.** Comparison of normal and sickle red blood cells; sickle-shaped red blood cells obstruct **Figure 1.** Comparison of normal and sickle red blood cells; sickle-shaped red blood cells obstruct blood flow in narrow blood vessels. Drawings were made by the authors using BioRender. Adapted blood flow in narrow blood vessels. Drawings were made by the authors using BioRender. Adapted from [4]. from [\[4\]](#page-15-3).

SCD can manifest in various clinical features, including abnormal eye growth in the retina (proliferative retinopathy), tissue death due to blocked blood flow (vascular necrosis), prolonged painful erection (priapism), dilute urine (hyposthenuria), episodes of severely reduced red blood cell production (aplastic crises), kidney problems (nephropathy), and lung disease [5]. For individuals with SCD, episodes of severe pain caused by blocked blood vessels (vaso-occlusive crisis) are the most frequent reason for emergency department visits and hospital admissions [9]. Other symptoms manifested in sickle cell anemia carriers include severe bacterial infections, necrosis (tissue death), acute chest syndrome, ischemic and hemorrhagic stroke, sepsis, hemolytic crises, and functional asplenia [1]. SCD affects various body parts, causing a range of symptoms and complications, which can be acute, chronic, or a combination of both (Figure 2).

<span id="page-2-0"></span>

**Figure 2.** Acute and chronic complications of sickle cell disease in various body systems. Drawings **Figure 2.** Acute and chronic complications of sickle cell disease in various body systems. Drawings were made by the authors (ChemDraw software, version 20.0.1.1). Adapted from  $\mathcal{L}$ were made by the authors (ChemDraw software, version 20.0.0.41). Adapted from [\[13\]](#page-16-6).

cell counts, hemoglobin solubility tests, and high-performance liquid chromatography. Comprenensive information regarding genetic mutations can be obtained using genetic<br>testing options such as restriction fragment length polymorphism, PCR-based methods, and DNA sequencing. Flow cytometry, mechanical sickle cell differentiation, lateral flow mununoassay, and density-based separation are examples of emerging approaches. Further<br>diagnostic capabilities are provided by additional techniques such as pyrosequencing, the Many techniques and assays are used to identify and track SCD. These consist of hemoglobin electrophoresis, isoelectric focusing, peripheral blood smears, complete blood Comprehensive information regarding genetic mutations can be obtained using genetic immunoassay, and density-based separation are examples of emerging approaches. Further

hemoglobin solubility test on paper, and sensor-based technologies including electrical impedance sensors and optofluidic resonators based on fluorescence [\[14\]](#page-16-7). The current approach to SCD management prioritizes symptom relief and complication prevention in the absence of a definitive cure, with efforts to develop effective treatments facing significant challenges and slow progress [\[15,](#page-16-8)[16\]](#page-16-9). In this sense, different drugs targeting diverse biochemical pathways have been developed for SCD [\[17\]](#page-16-10). Despite promising leads in some cases, many fail to demonstrate significant benefits in clinical trials [\[5\]](#page-15-4). Even for those showing potential, limited availability creates a substantial access barrier due to its high costs, particularly in low-income countries [\[5\]](#page-15-4). In Africa, where traditional healing practices are prevalent, many people with SCD rely on plant-based remedies (phytomedicines) as a primary source of healthcare, especially for painful episodes and other SCD complications [\[5](#page-15-4)[,18\]](#page-16-11). Studies have shown that many medicinal plant species from both developing and developed countries possess promising antisickling properties, suggesting their potential for treating and managing sickle cell anemia [\[19\]](#page-16-12). Yembeau et al. [\[19\]](#page-16-12) conducted an ethnobotanical exploration focusing on plants commonly utilized by the populations of Nigeria's southwestern region for disease management. The study identified 44 plant species with promising properties for the development of new medications to manage various diseases [\[20\]](#page-16-13). Plants contain abundant bioactive compounds that hold promise as antisickling agents capable of neutralizing free radicals, which could potentially reduce oxidative stress on red blood cells, leading to a decrease in hemolysis (red blood cell destruction) and a longer lifespan for these cells [\[5\]](#page-15-4). Therefore, incorporating foods rich in bioactive compounds into the diets of people with SCD might offer a complementary approach to manage the condition. These functional foods could potentially improve clinical health by reducing the risk or severity of complications associated with the disease [\[5\]](#page-15-4).

This pilot study presents a comprehensive review analyzing randomized, double-blind placebo-controlled trials (RCTs) across Phase I, II, and III for interventions in SCD, focusing on various treatment modalities. Randomization aims to eliminate both unconscious and deliberate human influence on the assignment of subjects to different groups. Blind assessment ensures that treatment and analysis of outcomes are not colored by prejudice [\[21\]](#page-16-14). This work also summarizes and discusses the findings from these trials while also exploring the potential of functional foods and herbs rich in bioactive compounds as therapeutic agents for SCD management.

## **2. Methods**

This review aimed to identify relevant research on treatments for sickle cell anemia. A comprehensive search strategy was employed across international databases like Scopus, PubMed, and Google Scholar, encompassing the period from 2019 to 2024. The search was divided into two distinct parts (Figure [3\)](#page-4-0). Part I focused on identifying double-blind randomized controlled trials investigating potential treatment compounds. Keywords used in this search included "sickle cell anemia," "randomized controlled trial", and "doubleblind," to ensure robust study design. Additionally, articles were included if they employed an experimental design, had relevance to both food and pharmaceutical applications, and were published in English. This rigorous selection process yielded 15 articles that met the established criteria. Part II of the search strategy explored the use of supplements as functional food additives for patients with sickle cell disease. The keyword search employed terms like "sickle cell anemia", "functional food", "supplement", and "bioactive compounds" to identify relevant studies. The inclusion criteria mirrored those of Part I, ensuring the identification of articles with experimental designs, applicability to both food and pharmaceutical domains, and publication in English. This search identified 10 articles that met the established criteria. The impact of bioactive compounds in SCD management is also present in Section [5.](#page-13-0)

<span id="page-4-0"></span>

**Figure 3.** Flow diagram of systematic review selection criteria. **Figure 3.** Flow diagram of systematic review selection criteria.

## **3. Review of Randomized, Double-Blind Clinical Trials for Sickle Cell Disease Treat-3. Review of Randomized, Double-Blind Clinical Trials for Sickle Cell ments Disease Treatments**

The US Food and Drug Administration (FDA) recognizes four treatment options for  $\frac{1}{2}$ preventing and alleviating complications associated with SCD: hydroxyurea (HD), voxelotor (Oxbryta/GBT440), L-glutamine, and crizanlizumab [\[5\]](#page-15-4). These medications represent significant advancements in managing SCD, but researchers are relentlessly pushing the boundaries to improve treatment effectiveness and patient outcomes. One patiting the boundaries to improve treatment effectiveness and patient outcomes. One key area of exploration involves investigating novel therapeutic compounds, potentially leading to a future where more patients with SCD can experience improved quality of future where more patients with SCD can experience improved quality of life and a reduced burden of complications. Several promising candidates are currently duced burden of complications. Several promising candidates are currently being evalu-being evaluated in rigorous clinical trials, including randomized, double-blind, placebo-controlled Phase I, II, and III studies (Table [1\)](#page-5-0). for preventing and alleviating complications associated with SCD: hydroxyurea (HU),

These trials are designed to ensure the objectivity and generalizability of the findings  $T_{\text{min}}$  trials are designed to the objectivity and unconscious influences by eliminating subset in  $\mathcal{L}$ by eliminating subjectivity and unconscious influences.

<span id="page-5-0"></span>

**Table 1.** Randomized double-blind controlled studies of different products used in the treatment of sickle cell disease.









The addition of HU, L-glutamine, crizanlizumab, and voxelotor to the treatment options for SCD represents a significant step forward  $[16]$ . HU, a drug that effectively increases fetal hemoglobin (HbF) production, has emerged as a pivotal option in managing SCA, demonstrating efficacy and practicality [\[37\]](#page-17-7). HbF, a type of hemoglobin present in fetal red blood cells, has a more flexible structure than adult hemoglobin, allowing it to bend and deform without sickling even under conditions of low oxygenation. This characteristic is crucial in the pathogenesis of SCD [\[38\]](#page-17-8). However, HU can have some side effects, and researchers continue to explore new treatment options. HU is a strong inhibitor of the enzyme ribonucleotide reductase within the cell and converts ribonucleotides into deoxyribonucleotides, which are essential for DNA synthesis and repair. It has been used for several decades in the treatment of various disorders, especially myeloproliferative neoplasms, chronic myeloid leukemia, and the human immunodeficiency virus (HIV) [\[39\]](#page-17-9). Over time, speculation arose that HU could potentially be carcinogenic and harmful to DNA. With this, the widespread use of HU was reduced, and many patients stopped receiving treatment with this drug. Research suggests that HU may not be equally effective for all patients with SCD. Additionally, safety concerns have been raised regarding potential harm to a developing fetus (embryofetal toxicity) and the suppression of bone marrow activity (myelosuppression) [\[5](#page-15-4)[,40\]](#page-17-10).

The guidelines published by the National Institutes of Health, the American Society of Hematology, and the British Society of Hematology all recommend HU as the standard treatment for SCA, but it is important to investigate its genotoxic and carcinogenic potential [\[41\]](#page-17-11). Abdullahi et al. [\[31\]](#page-17-12) conducted a double-blind, randomized controlled trial assessing the efficacy of low-dose (10 mg/kg/day) versus moderate-dose (20 mg/kg/day HU in 220 children with SCA and abnormal transcranial Doppler velocities. The study revealed no significant difference in stroke incidence between the groups over a 20-month period. This finding suggests that a lower dose of HU might be similarly effective for stroke prevention in this pediatric population. The research findings potentially influenced clinical practice guidelines in three Nigerian states, impacting an estimated 40,000 children at risk of stroke.

Fish oil is a rich source of omega-3 fatty acids, specifically two major types: docosahexaenoic acid (DHA) and eicosapentaenoic acid (EPA). These essential fats fall under the broader category of polyunsaturated fatty acids and are vital for maintaining human health [\[22,](#page-16-23)[42\]](#page-17-13). Giriraja et al. [\[22\]](#page-16-23) elaborated a study where ten adults with SCD were given an omega-3-enriched food, daily, for a month. The authors concluded that the omega-3-enriched food reduce sickle cell-related inflammation. Abdelhalim, Murphy, Meabed, Elberry, Gamaleldin, Shaalan, and Hussein [\[23\]](#page-16-24) conducted a randomized, double-blind clinical trial and investigated the effects of omega-3 fish oil supplementation in 165 patients. Participants received either 1000 mg of omega-3 fish oil (containing 400 mg EPA and 300 mg DHA) or a daily dose of 2800 IU vitamin D for 10 months. The study found a significant increase in both serum high-density lipoprotein (HDL) and low-density lipoprotein (LDL) cholesterol levels in the omega-3 group compared to the control group. Additionally, the omega-3 group experienced a notable decrease in the number and severity of pain crises. People with SCD commonly exhibit reduced antioxidant defenses in their bloodstream, potentially due to lipid peroxidation resulting from interactions with ferroptosis and compromised antioxidant capacity. Additionally, EPA and DHA act as powerful anti-inflammatory agents that can modulate pain [\[42\]](#page-17-13).

The human body relies on a multitude of vitamins, and vitamin A stands out for its versatility. It influences a remarkable range of processes throughout life, from enabling healthy development before and after birth to supporting vision, reproduction, and tissue repair. Additionally, vitamin A strengthens our immune system, acting as a key player in overall health [\[24,](#page-16-25)[43\]](#page-17-14). In American children with SCD, inadequate levels of vitamin A are related with increased rates of hospitalization, lower development, and inferior hematologic status (hemoglobin 7.9 vs.  $8.5 \text{ g}/\text{dL}$ ) compared to children with appropriate vitamin A levels [\[44\]](#page-17-15). Brownell et al. [\[24\]](#page-16-25) conducted an 8-week study in which twenty healthy infants with hemoglobin-SS illness received either 3000 or 6000 IU/d of retinol orally. The authors reported minor improvements in exploratory nutritional, hematologic, and muscular results after supplementation. They recommend additional research in the form of a larger-sample longitudinal placebo-controlled study that considers the use of HU.

Folic acid, also known as vitamin B9, is commonly prescribed to SCD patients to support red blood cell production [\[45\]](#page-17-16). Muga et al. [\[25\]](#page-16-26) conducted a randomized doubleblind, active-controlled, clinical trial with 61 individuals for 4 weeks, where the intervention group received 30 capsules of folic acid 500 microgram and 60 capsules of EvenFlo 500 mg. The authors concluded that EvenFlo, a nutritional supplement, shows promise in managing SCD when combined with folic acid. The results suggest that EvenFlo may be beneficial for increasing hemoglobin levels, improving weight management in patients with SCD, and potentially reducing the frequency of crisis episodes.

Vitamin D, a fat-soluble substance essential for the human body, primarily regulates calcium and phosphorus levels to maintain bone health, and also plays important roles in the cardiovascular, immune, and pancreatic systems, as well as in muscle, brain function, and cell cycle control [\[46\]](#page-17-17). Grégoire-Pelchat et al. [\[26\]](#page-16-27) investigated the effectiveness of vitamin D supplementation in raising serum levels of 25-hydroxyvitamin D (25(OH)D) in 38 children with SDC. Participants received either a high-dose vitamin D bolus combined with daily 1000 IU vitamin D3 or daily supplementation alone. Additionally, researchers assessed various health markers, including calcium levels, bone markers, musculoskeletal pain, quality of life, and hematological parameters. The study found that the combined approach was more effective in increasing 25(OH)D levels compared to daily supplementation alone.

Arginine is a nutritional amino acid and the obligate substrate for nitric oxide (NO) production [\[27](#page-16-28)[,28\]](#page-16-29). Studies suggest that impaired L-arginine metabolism and NO availability contribute to the vascular dysfunction observed in SCD [\[28](#page-16-29)[,47\]](#page-17-18). NO acts as a potent vasodilator, impacting various vascular and circulating blood cell functions such as platelet aggregation inhibition, adhesion molecule down-regulation, and ischemia-reperfusion injury modulation, all crucial pathways affected during VOCs [\[27](#page-16-28)[,48](#page-17-19)[,49\]](#page-17-20). Onalo et al. [\[27\]](#page-16-28) investigated the effects of oral L-arginine supplementation in children with sickle cell disease (SCD) hospitalized for vaso-occlusive pain crises (VOC) in Nigeria. For 24 months, participants received either oral L-arginine hydrochloride (100 mg/kg three times a day) or a control treatment. This study provided the evidence that daily oral arginine supplementation may improve pain control and reduce the need for pain medication in children experiencing VOCs associated with SCD. Onalo et al. [\[28\]](#page-16-29) investigated the effects of oral L-arginine supplementation in 47 children (aged 5–17 years) hospitalized with severe pain and/or acute chest syndrome. The study compared daily L-arginine (300 mg/kg/day in three divided doses) to a placebo over 22 months. Notably, treatment with L-arginine led to a greater reduction in key cardiopulmonary hemodynamic markers, including pulmonary artery pressure, compared to a placebo. These findings suggest that L-arginine supplementation may offer benefits for improving cardiovascular health in SCD patients experiencing vaso-occlusive episodes and acute chest syndrome.

Voxelotor is a recently approved drug often used as an alternative for patients unresponsive to HU, with its long-term efficacy and safety currently under investigation [\[50](#page-17-21)[,51\]](#page-17-22). It was recently approved by the FDA for the treatment of SCD in children aged 12 and older [\[50](#page-17-21)[,51\]](#page-17-22). This molecule binds to hemoglobin, boosting its oxygen-carrying capacity (affinity) for better delivery throughout the body. It also stabilizes hemoglobin, preventing the abnormal clumping (sickling) that causes complications. Its daily oral administration has been shown, both in vitro and in vivo, to reduce the sickling of red blood cells, improve blood viscosity, and improve red blood cell deformability [\[30\]](#page-17-23). Additionally, it prolongs the half-life of red blood cells, reducing anemia and hemolysis. The drug is generally well-tolerated, although it may cause moderate adverse effects such as headache, diarrhea, nausea, and arthralgia [\[50\]](#page-17-21). Hutchaleelaha et al. [\[29\]](#page-16-30) evaluated the safety, tolerability, pharmacokinetics, and pharmacodynamics of voxelotor for the first time in humans. The study included both healthy volunteers and individuals with SCD. Twenty-four healthy

volunteers received varying daily doses of voxelotor (300, 600, or 900 mg) or a placebo for 15 days. The findings revealed favorable tolerability profiles in both SCD patients and healthy volunteers. Moreover, the study provided evidence indicating that voxelotor enhances hemoglobin–oxygen affinity, a critical mechanism believed to offer therapeutic benefits in SCD. Furthermore, assessments of erythropoietin levels, exercise tolerance, and hematologic parameters consistently demonstrated normal oxygen delivery during both resting periods and physical exertion. Vichinsky et al. [\[30\]](#page-17-23) also investigated the effects of voxelotor in SCD. The study involved 274 participants randomly assigned to receive either a daily dose of 1500 or 900 mg of voxelotor, or a placebo, for 24 weeks. The researchers found that voxelotor significantly increased hemoglobin levels and reduced the frequency of worsening anemia and hemolysis compared to the placebo group. Notably, despite the rise in hemoglobin, there was no observed increase in VOC rates, suggesting that voxelotor may improve hemoglobin levels without negatively impacting blood viscosity.

Some intravenous (IV) medications also play a role in managing pain associated with SCD [\[32\]](#page-17-24). Notably, the FDA has approved IV acetaminophen for children over two years old to manage mild to severe pain, both with and without opioids [\[52,](#page-17-25)[53\]](#page-17-26). While IV diclofenac remains the current standard treatment for managing skeletal VOCs in SCD, its use requires careful monitoring due to potential side effects [\[32\]](#page-17-24). These include respiratory depression, severe constipation, and the risk of dependence with frequent use.

Opioid medications, although effective for pain relief, often have limited availability and require similar monitoring due to their side effects [\[32](#page-17-24)[,52\]](#page-17-25). Panda, Mishra, Patra, Nayak, and Panda [\[32\]](#page-17-24) compared the effectiveness of IV acetaminophen and diclofenac sodium in managing skeletal VOCs in 104 children with SCD. The study found that IV acetaminophen (10 mg/kg/dose) led to several improvements compared to IV diclofenac sodium (1 mg/kg/dose) administered over two months. These included a greater reduction in pain scores (50%), a decrease in the number of medication doses needed to achieve pain relief within 24 h, and a faster reduction in pain scores within the first hour. Based on these findings, the researchers concluded that IV acetaminophen may be a preferable alternative to IV diclofenac [\[37\]](#page-17-7). Other studies are currently in progress to explore therapeutic alternatives, aiming to identify new pharmacological agents of different natures and potential drug candidates for managing this disease. Meanwhile, a wide range of compounds are being investigated to complement treatment and promote significant improvements in the quality of life of individuals affected by this condition. One example of these advances is the monoclonal antibodies crizanlizumab (adakveo) and canakinumab [\[33](#page-17-27)[,54\]](#page-17-28). Crizanlizumab, a humanized monoclonal antibody, has been approved for the treatment of SCD in patients aged 16 and older. Its mechanism of action includes attenuating VOCs by binding to P-selectin and blocking its interaction with other molecules, preventing the adhesion of red blood cells. Crizanlizumab received FDA approval in 2019 to reduce VOC frequency in patients with SCD in adults and children SCD patients 16 years of age and older [\[15](#page-16-8)[,37\]](#page-17-7). This approval was based on positive results from the SUSTAIN trial, which demonstrated a 45.3% reduction in annual pain crises for patients receiving crizanlizumab compared to a placebo group [\[55\]](#page-17-29). Notably, the benefit was even greater for patients not taking HU, with a 50% reduction in annual crisis rate compared to the placebo. However, preliminary data from the phase III STAND clinical trial, released in early 2023, did not show a statistically significant difference in annual vaso-occlusive crisis rates between crizanlizumab and placebo groups. This lack of efficacy in the STAND trial led to the European Medicines Agency's decision to remove crizanlizumab from their list of approved SCD medications [\[56\]](#page-18-0).

*Canakinumab* is a human monoclonal antibody that acts by blocking interleukin-1 beta  $(IL-1\beta)$ , a pro-inflammatory protein involved in various chronic inflammatory conditions. In addition to being beneficial in the treatment of various rheumatic diseases, a reduction in cardiovascular events has been observed after myocardial infarction. Regarding SCA, it is being investigated as a possible therapeutic option due to its potential to modulate

the inflammatory response, which plays an important role in the pathogenesis of this disease [\[33\]](#page-17-27).

Rees, Kilinc, Unal, Dampier, Pace, Kaya, Trompeter, Odame, Mahlangu, and Unal [\[33\]](#page-17-27) investigated the effects of *canakinumab* in adolescents and young adults with SCA (HbSS or HbSβ0-thalassemia). The study involved 49 participants aged 8 to 20 years with a history of acute pain episodes. The participants received either 300 mg subcutaneous *canakinumab* or a placebo, administered six times over a set period. The study found *canakinumab* to be well-tolerated with no serious adverse events (SAEs) attributable to the treatment and no unexpected safety concerns. Notably, the authors observed that canakinumab's selective blockade of IL-1 $\beta$  inflammation appeared to yield therapeutic benefits, particularly by reducing fatigue, a significant symptom in SCA patients.

Ticagrelor is an antiplatelet drug widely used as a P2Y12 receptor antagonist, essential for platelet activation [\[57\]](#page-18-1). It is employed to prevent cardiovascular events like heart attacks and strokes in adults with coronary or cerebrovascular artery disease [\[58\]](#page-18-2). Its action involves blocking the P2Y12 ADP receptor on platelets, inhibiting their activation and aggregation, thereby reducing blood clot formation, and helping to prevent adverse cardiovascular events [\[58\]](#page-18-2). Currently, clinical studies are underway to investigate the use of ticagrelor in SCD treatment, reducing and contributing to complications such as VOCs and strokes, which have shown satisfactory tolerance and safety [\[59](#page-18-3)[,60\]](#page-18-4). Heeney, Abboud, Githanga, Inusa, Kanter, Michelson, Nduba, Musiime, Apte, and Inati [\[34\]](#page-17-30) investigated ticagrelor's effectiveness in reducing VOCs in children with SCD. Participants received ticagrelor or a placebo for 12 months, with varying doses based on weight. However, the trial was stopped early after 8 months due to safety concerns. The data monitoring committee determined that the potential risks of continuing outweighed any potential benefits of ticagrelor. This unfortunately adds ticagrelor to a growing list of unsuccessful medications for SCD, which includes senicapoc, prasugrel, regadenoson, sevuparin, poloxamer 188, olinciguat, and rivipansel [\[35\]](#page-17-31).

Ketamine, a medication that blocks a specific receptor in the nervous system (NMDA receptor), shows promise in reducing a condition called opioid-induced hyperalgesia. This occurs by potentially preventing nerve cells in the spine from becoming overly sensitive to pain signals [\[35](#page-17-31)[,61\]](#page-18-5). It operates on glutamate and NMDA receptors, influencing the peripheral pain sensitization process along pain pathways [\[61,](#page-18-5)[62\]](#page-18-6). Additionally, ketamine is theorized to impact neural plasticity in both the NMDA receptors and spinal pathways, thereby impeding the transmission of stimuli generated towards the central nervous system [\[62\]](#page-18-6). Alshahrani, AlSulaibikh, ElTahan, AlFaraj, Asonto, AlMulhim, Al-Abbad, Almaghraby, AlJumaan, AlJunaid, Darweesh, AlHawaj, Mahmoud, Alossaimi, Alotaibi, AlMutairi, AlSulaiman Pharm, Alfaraj, Alhawwas, Mbuagbaw, Lewis, Verhovsek, Crowther, Guyatt, and Alhazzani [\[35\]](#page-17-31) investigated the effectiveness and safety of a single, low-dose ketamine infusion (at 0.3 mg/kg) for managing acute VOC in adults with SCD. The study involved 278 adults with SCD over a 12-month period. The findings suggest that ketamine provided significant pain relief, allowing patients to require lower cumulative morphine doses. Importantly, the study found no major safety concerns with ketamine use. The authors recommend further research to explore the effectiveness and safety of repeated ketamine dosing or continuous infusion for managing VOCs in adults with SCD. Additionally, they suggest investigating the potential benefits of combining ketamine with opioids compared to opioid use alone.

Another treatment under study is the use of isoquercetin, a flavonoid found in various sources such as citrus fruits, onions, apples, and teas [\[63\]](#page-18-7). It is the orally available form of quercetin and it possesses the glycosidic form [\[63\]](#page-18-7). It has been studied for its antioxidant, anti-inflammatory, antimicrobial, and antiviral properties, and its use has improved thrombosis biomarkers in cancer patients without inducing bleeding [\[36](#page-17-32)[,64\]](#page-18-8). Therefore, some studies are testing its efficacy and safety in regulating thromboinflammatory processes in SCD [\[36,](#page-17-32)[64\]](#page-18-8). *Sevuparin*, a drug derived from heparin that exhibits broad preclinical activity, targets relevant targets in VOCs, such as P- and L-selectin, thrombospondin, von

Willebrand factor, and fibronectin [\[65\]](#page-18-9). Although the anticoagulant activity of this drug has been removed, its anti-adhesive properties have been preserved. This compound has been shown to be a potent anti-adhesive agent, both in in vitro and in vivo studies, which is crucial in the context of VOCs in SCD. It helps prevent or reduce blood vessel obstruction, a common phenomenon in this condition due to the abnormal adherence of sickle cells to each other and vessel walls [\[66,](#page-18-10)[67\]](#page-18-11). Lizarralde-Iragorri, Parachalil Gopalan, Merriweather, Brooks, Hill, Lovins, Pierre-Charles, Cullinane, Dulau-Florea, Lee, Villasmil, Jeffries, and Shet [\[36\]](#page-17-32) investigated the effectiveness of isoquercetin in reducing inflammation associated with blood clots (thromboinflammation) in SCD. The study involved 46 patients receiving either a placebo or 1000 mg of isoquercetin daily for 28 to 35 months. The researchers observed a significant reduction in several biomarkers linked to thromboinflammation in patients taking isoquercetin compared to the placebo group. These findings suggest that future trials are warranted to explore the potential benefits of higher isoquercetin doses and longer treatment durations for managing thromboinflammation in SCD patients.

## **4. Functional Food Exploited in SCD Management**

Functional foods, often referred to as nutraceuticals, are foods that are rich in bioactive substances that improve the health of their consumers [\[68\]](#page-18-12). According to Granato, Barba, Bursać Kovačević, Lorenzo, Cruz, and Putnik [\[69\]](#page-18-13), functional foods encompass both natural and processed options that, when incorporated regularly into a varied diet, may offer additional health benefits beyond basic nutrition. The following requirements are the primary conditions for an ingredient or product meeting a certain functional claim on a food label [\[69\]](#page-18-13): (i) food security; (ii) accessible without a prescription (or guidance from a doctor); and (iii) proof that consuming it on a regular basis in a balanced diet has health benefits. Luvián-Morales, Varela-Castillo, Flores-Cisneros, Cetina-Pérez, and Castro-Eguiluz [\[70\]](#page-18-14) define a functional food as one that delivers a variety of health benefits, including enhancing overall physical well-being and reducing the risk of diseases.

The exploration of functional foods represents a dynamic and rapidly evolving field within nutritional science. This section explores the most current research and study results about bioactive substances present in nutrients and their possible health advantages in SCD. Referred to as antisickling factors, biological compounds from foods and comestible and medicinal plants could enhance the health-related quality of life in SCD. These matrixes are abundant in phenolics, vitamins, minerals, proteins, amino acids, and unsaturated fats, and possess antisickling activity [\[71,](#page-18-15)[72\]](#page-18-16). Researchers are exploring the potential of antioxidant-rich compounds in managing SCD. These substances may operate by reducing the body's oxidative stress levels, a factor believed to intensify the severity of the illness [\[73\]](#page-18-17). Their ability to lower oxidative stress makes them a promising treatment option for sickle cell complications [\[42,](#page-17-13)[74\]](#page-18-18).

Increasing antioxidant intake may help SCD patients by supporting cellular renewal and promoting the formation of red blood cells [\[75\]](#page-18-19). For example, significant components in vegetables, phenolics have promising impact on human health. As an illustration, the *Carica papaya* was employed as a substitute therapeutic agent for SCD [\[74\]](#page-18-18). Polar phenolic compounds viz. phenolic acids kaempferol and quercetin were detected and characterized in papaya leaves [\[76\]](#page-18-20). The extract has an antisickling impact and a substantial membrane stabilizing (protective) effect. Another study conducted by Famojuro, Adeyemi, Ajayi, Fasola, Fukushi, Omotade, and Moody [\[77\]](#page-18-21) reported that two isolated compounds from the root of *Combretum racemosum* P. beauv (*Combretaceae*) showed effective SCD management. These compounds are proved to trigger red cells' membrane-bound enzymes  $\mathrm{Na^+}$ , K<sup>+</sup>-ATPase, and  $\mathrm{Ca^{2+}}$ -ATPase, which are engaged in the sickling process. Ahajumobi and Asika [\[78\]](#page-18-22) demonstrated that 2-hydroxymethylbenzoic acid has an antisickling impact and could stabilize the erythrocytes membrane. It has been revealed that some amino acids could avoid sickling by disturbing the erythrocyte membrane, provoking intensification in the cell volume of the erythrocyte, and consequently decreasing the intracellular hemoglobin level [\[79](#page-19-0)[,80\]](#page-19-1). Among the described amino acids, phenylalanine has been revealed to be the most efficient [\[81\]](#page-19-2). L-arginine reduces

oxidative stress and L-Glutamine diminishes inactive energy expenditure [\[82\]](#page-19-3). Niihara [\[83\]](#page-19-4) reported a phase 3 trial demonstrating efficacy in reducing acute complications of SCD, leading the FDA to approve pharmaceutical-grade L-glutamine (Endari, Emmaus Medical) as a prescription medication for adults and children aged 5 and above.

Daak et al. [\[84\]](#page-19-5) concluded that omega-3 fatty acids reduce thrombotic activities and the level of episode pain. In this sense, elevated protein and L-Arginine supplements and n-3 fatty acids to Heinz body (denatured Hb) have shown a noteworthy lowering in inflammation, oxidative stress, red cell density, and pain episodes, while enhancing microvascular function. In addition, numerous dietary supplements, like thiocyanate, possess a vast ability to prevent erythrocytic deformations in the management of sickle cell disease [\[76\]](#page-18-20).

Vitamin E and β-carotene are obtainable in foods such as vegetable oils, nuts, seeds, breakfast cereal, and fortified fruit juices, margarine, and spreads. Some investigations have stated that high levels of vitamin E decrease oxidative stress-induced erythrocyte injury [\[85,](#page-19-6)[86\]](#page-19-7). Physiologically, vitamin E lessens lipid peroxidation and amends erythrocyte membrane stability [\[87\]](#page-19-8). On the other hand, ascorbic acid is lengthily circulated in nature, typically abundant in leafy vegetables and fresh fruits. Vitamin C normalizes attenuated hemodynamic alterations brought on by postural corrections and inhibits the development of Heinz body (denatured Hb) in vitro in sickle red blood cells [\[76\]](#page-18-20). Similarly, vitamin B9 (folic acid) could manage HbS; this management is predicated on stopping the absence from the amplified folate turnover [\[88\]](#page-19-9). In this way, vitamin B9 supplementation could reduce the high risk of endothelial damage.

Regarding minerals, Mg adjunct could reduce the number of dense erythrocytes and improve the erythrocyte membrane transport abnormalities of individuals with SCD [\[5,](#page-15-4)[89](#page-19-10)[,90\]](#page-19-11). Research suggests that maintaining magnesium levels within the recommended range for patients, with regular monitoring, might be associated with a lower risk of mortality [\[91\]](#page-19-12). Zn can be found in plentiful foods, including seafood (as herring and oysters), beans and peas, and grains. Zn could enhance thymulin activity, reduce bacterial contamination occurrence, and cure painful disasters [\[92\]](#page-19-13).

Fruits and vegetables contain a diverse range of functional compounds, including carotenoids, vitamin C, fiber, magnesium, and potassium. These components work together, sometimes in cooperation (synergistically) and sometimes in opposition (antagonistically), to contribute to overall health benefits. Strong evidence supports current dietary recommendations that encourage increased consumption of fruits and vegetables, aiming for at least five servings daily [\[76\]](#page-18-20). In addition, a wide range of constituents found in functional food, each with unique physiological effects, has been demonstrated by recent research. For example, black beans (*Phaseolus vulgaris* L.), *Fragaria vesca* L., bitter *Garcinia kola*, *Annona muricata* L., and *Azadirachta indica* J. have been shown to be able to manage SCD [\[76,](#page-18-20)[93\]](#page-19-14). Some studies have concluded that vitamins C and E and minerals viz. Zn and Mg could reduce the percentage of irreversibly sickled cells [\[94](#page-19-15)[,95\]](#page-19-16).

#### <span id="page-13-0"></span>**5. Impact of Bioactive Compounds in Sickle Cell Disease Management**

Researchers are exploring the potential of various natural plant compounds for SCD (Table [2\)](#page-14-0). Before hydroxyurea was available in Africa, people with SCD depended on hospital-based supportive treatments and home-based herbal medicines, with several medicinal plants identified that reactivated  $\gamma$ -gene transcription through different cellular mechanisms [\[96\]](#page-19-17). Commonly used herbs possess antisickling properties and are primarily utilized to prevent VOCs [\[96\]](#page-19-17). The potent components found in medicinal plants and natural compounds, termed antisickling agents, abundant in aromatic amino acids, phenolic compounds, and antioxidants, serve as antioxidant therapy to alleviate the complexities of sickle cell anemia, exhibiting protective effects such as shielding against lipid peroxidation in red blood cells, enhancing glutathione levels (GSH), and diminishing levels of reactive oxygen species (ROS) [\[97\]](#page-19-18). Therefore, it was concluded that herbal medicines have substantial potential in the therapeutic management of sickle cell disease, with the possibility of addressing both the underlying causes and the symptoms of the condition [\[74\]](#page-18-18).



<span id="page-14-0"></span>**Table 2.** Potential therapeutic effects of bioactive compounds from natural products in sickle cell disease.

One promising example is niprisan, a plant-based medicine with antisickling properties. Studies suggest it may inhibit the polymerization of HbS, a key factor in SCD complications [\[112,](#page-20-2)[113\]](#page-20-3).

Patent (US5800819A) describes the phytochemical composition of a product containing a mixture of four plant materials: *Piper guineense* seeds and *Pterocarpus osun*, *Eugenia caryophyllus*, and *Sorghum bicolor* extracts. The mixture has been used for the treatment of SCD [\[114\]](#page-20-4). The extracted fraction from Ciklavit (*Cajanus cajan*) extract, rich in essential amino acids, vitamin C, and Zn, may decrease the occurrence of painful crises and mitigate the negative impact of SCA on the liver [\[71\]](#page-18-15). The authors reported that the mechanism of action remains to be determined [\[115\]](#page-20-5).

The rising consumer demand for functional foods, those offering health benefits beyond basic nutrition, has spurred a surge of innovation within the food industry [\[116](#page-20-6)[,117\]](#page-20-7). Nevertheless, a significant challenge persists ensuring the consistent quality and effectiveness of these enriched products throughout their entire journey, from initial preparation and processing to transportation, storage, and, finally, consumption by the end user. Bioactive compounds are chemically unstable, prone to degradation, volatilization, and oxidation, leading to a loss of their biological activities when exposed to harsh environmental conditions like high temperatures and the presence of oxygen and light [\[116,](#page-20-6)[117\]](#page-20-7). Hence, their vulnerability to degradation constrains their direct application in food products and limits their potential health benefits. Encapsulation technology stands out as a potentially transformative solution. By encapsulating bioactive compounds within a protective shell, this technology not only shields them but also enhances their essential properties, such as solubility, stability, and bioavailability, and their antioxidative effects. Moreover, encapsulation facilitates the controlled release of these beneficial compounds, thereby maximizing their positive impact on human health [\[116](#page-20-6)[,117\]](#page-20-7). The field of natural products for sickle

cell disease (SCD) management holds immense promise. Advancements in nutrigenomics, digital health, omics technologies, personalized nutrition, and sustainability offer exciting avenues for future research and development. Exploring the nutritional properties of functional foods and harnessing their therapeutic potential can establish a path toward a healthier, more resilient future. Including health-related statements on functional food product labels is a key method of educating consumers about the relationship between nutrition and health. However, regulations are essential to oversee these claims, as they significantly influence consumer food choices.

#### **6. Conclusions**

Current treatments for SCD have limitations. Therefore, well-designed clinical trials are crucial to validate the effectiveness of new therapies before widespread use. Recent trials with pharmaceutical interventions for both acute and chronic symptoms of SCD show promise. Despite the encouraging findings from these trials, there remains a necessity for further investigations. Functional foods enriched with bioactive compounds hold particular interest for SCD management, especially in resource-limited regions where traditional herbal remedies are prevalent. These foods may offer therapeutic, antisickling, antioxidant, and inhibitory effects, acting as potential combatants against sickle cell disease crises. This highlights the urgent need for clinical studies to assess the potential benefits of such nutrition-based approaches, focusing on commonly used medicinal plants to maximize their feasibility and biological impact. Research into the role of food in managing SCD demands further research. Plants with a long history of medicinal use present a promising and practical avenue for exploration. Rigorous evaluation of functional foods enriched with beneficial compounds could lead to the discovery of new and accessible treatment options for individuals living with SCD, particularly in regions with limited resources.

**Author Contributions:** Conceptualization: E.G., S.S., M.B., J.M.O., A.P.A. and L.T.; Methodology: E.G., S.S., M.B., J.M.O., A.P.A. and L.T.; Visualization: E.G., S.S. and L.T.; Validation: E.G., S.S., M.B., J.M.O., A.P.A. and L.T.; Formal Analysis: E.G., S.S., M.B., J.M.O., A.P.A. and L.T.; Resources: M.B., A.P.A. and L.T.; Data Curation: E.G., S.S. and L.T.; Writing—Original Draft Preparation: E.G., S.S., M.B., J.M.O., A.P.A. and L.T.; Writing—Review and Editing: E.G., S.S., M.B., J.M.O., A.P.A. and L.T.; Supervision: M.B., A.P.A. and L.T.; Project Administration: S.S., M.B., A.P.A. and L.T. All authors have read and agreed to the published version of the manuscript.

**Funding:** Miguel Brito gratefully acknowledges the FCT/MCTES national support through the projects H&TRC UIDB/05608/2020, UIDP/05608/2020, and IPL/IDI&CA2023/Ipasthma\_ESTeSL. J.M. Oliveira acknowledges the financial support of CICECO–Aveiro Institute of Materials, UIDB/50011/2020 (DOI 10.54499/UIDB/50011/2020), UIDP/50011/2020 (DOI 10.54499/UIDP/ 50011/2020) & LA/P/0006/2020 (DOI 10.54499/LA/P/0006/2020), financed by national funds through the FCT/MCTES (PIDDAC). Ana Paula Arez would like to acknowledge Fundação para a Ciência e a Tecnologia for funds to GHTM—UID/04413/2020 and LA-REAL LA/P/0117/2020.

**Data Availability Statement:** The data presented in this study are available on request from the corresponding authors.

**Conflicts of Interest:** The authors declare no conflicts of interest.

#### **References**

- <span id="page-15-0"></span>1. Tebbi, C.K. Sickle Cell Disease, a Review. *Hemato* **2022**, *3*, 341–366. [\[CrossRef\]](https://doi.org/10.3390/hemato3020024)
- <span id="page-15-1"></span>2. Serjeant, G.R. The emerging understanding of sickle cell disease. *Br. J. Haematol.* **2001**, *112*, 3–18. [\[CrossRef\]](https://doi.org/10.1046/j.1365-2141.2001.02557.x) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11167776)
- <span id="page-15-2"></span>3. WHO. Sickle Cell Disease. Available online: <https://www.afro.who.int/health-topics/sickle-cell-disease> (accessed on 9 June 2024).
- <span id="page-15-3"></span>4. Sanyaolu, A.; Agiri, E.; Bertram, C.; Brookes, L.; Choudhury, J.; Datt, D.; Ibrahim, A.; Maciejko, A.; Mansfield, A.; Nkrumah, J. Current modalities of sickle cell disease management. *Blood Sci.* **2020**, *2*, 109–116. [\[CrossRef\]](https://doi.org/10.1097/BS9.0000000000000056) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35400022)
- <span id="page-15-4"></span>5. Alabi, O.J.; Adegboyega, F.N.; Olawoyin, D.S.; Babatunde, O.A. Functional foods: Promising therapeutics for Nigerian Children with sickle cell diseases. *Heliyon* **2022**, *8*, e09630. [\[CrossRef\]](https://doi.org/10.1016/j.heliyon.2022.e09630) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35677416)
- <span id="page-15-5"></span>6. Adigwe, O.P.; Onoja, S.O.; Onavbavba, G. A Critical Review of Sickle Cell Disease Burden and Challenges in Sub-Saharan Africa. *J. Blood Med.* **2023**, *14*, 367–376. [\[CrossRef\]](https://doi.org/10.2147/JBM.S406196) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37284610)
- <span id="page-16-0"></span>7. Trabuchet, G.; Elion, J.; Baudot, G.; Pagnier, J.; Bouhass, R.; Nigon, V.M.; Labie, D.; Krishnamoorthy, R. Origin and spread of β-globin gene mutations in India, Africa, and Mediterranea: Analysis of the 5′flanking and intragenic sequences of β s and β c genes. *Hum. Biol.* **1991**, *63*, 241–252. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/1676014)
- <span id="page-16-1"></span>8. Zago, M.A.; Silva, W.A., Jr.; Dalle, B.; Gualandro, S.; Hutz, M.H.; Lapoumeroulie, C.; Tavella, M.H.; Araujo, A.G.; Krieger, J.E.; Elion, J.; et al. Atypical βs haplotypes are generated by diverse genetic mechanisms. *Am. J. Hematol.* **2000**, *63*, 79–84. [\[CrossRef\]](https://doi.org/10.1002/(SICI)1096-8652(200002)63:2%3C79::AID-AJH4%3E3.0.CO;2-D)
- <span id="page-16-2"></span>9. Jang, T.; Poplawska, M.; Cimpeanu, E.; Mo, G.; Dutta, D.; Lim, S.H. Vaso-occlusive crisis in sickle cell disease: A vicious cycle of secondary events. *J. Transl. Med.* **2021**, *19*, 397. [\[CrossRef\]](https://doi.org/10.1186/s12967-021-03074-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34544432)
- <span id="page-16-3"></span>10. da Guarda, C.C.; Yahouédéhou, S.; Santiago, R.P.; Neres, J.; Fernandes, C.F.L.; Aleluia, M.M.; Figueiredo, C.V.B.; Fiuza, L.M.; Carvalho, S.P.; Oliveira, R.M.; et al. Sickle cell disease: A distinction of two most frequent genotypes (HbSS and HbSC). *PLoS ONE* **2020**, *15*, e0228399. [\[CrossRef\]](https://doi.org/10.1371/journal.pone.0228399) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31995624)
- <span id="page-16-4"></span>11. Tonin, F.S.; Ginete, C.; Ferreira, J.; Delgadinho, M.; Santos, B.; Fernandez-Llimos, F.; Brito, M. Efficacy and safety of pharmacological interventions for managing sickle cell disease complications in children and adolescents: Systematic review with network meta-analysis. *Pediatr. Blood Cancer* **2023**, *70*, e30294. [\[CrossRef\]](https://doi.org/10.1002/pbc.30294)
- <span id="page-16-15"></span><span id="page-16-5"></span>12. Starlard-Davenport, A.; Gu, Q.; Pace, B.S. Targeting Genetic Modifiers of HBG Gene Expression in Sickle Cell Disease: The miRNA Option. *Mol. Diagn. Ther.* **2022**, *26*, 497–509. [\[CrossRef\]](https://doi.org/10.1007/s40291-022-00589-z) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35553407)
- <span id="page-16-16"></span><span id="page-16-6"></span>13. Change, S.S.C. Impact of Sickle Cell on the Body. 2024. Available online: [https://www.sparksicklecellchange.com/what-is](https://www.sparksicklecellchange.com/what-is-sickle-cell/symptoms-complications)[sickle-cell/symptoms-complications](https://www.sparksicklecellchange.com/what-is-sickle-cell/symptoms-complications) (accessed on 9 June 2024).
- <span id="page-16-7"></span>14. Arishi, W.A.; Alhadrami, H.A.; Zourob, M. Techniques for the Detection of Sickle Cell Disease: A Review. *Micromachines* **2021**, *12*, 519. [\[CrossRef\]](https://doi.org/10.3390/mi12050519) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34063111)
- <span id="page-16-17"></span><span id="page-16-8"></span>15. Salinas Cisneros, G.; Thein, S.L. Recent Advances in the Treatment of Sickle Cell Disease. *Front. Physiol.* **2020**, *11*, 435. [\[CrossRef\]](https://doi.org/10.3389/fphys.2020.00435) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32508672)
- <span id="page-16-18"></span><span id="page-16-9"></span>16. Abdel-Hadi, L.; Ventura Carmenate, Y.; Castillo-Aleman, Y.M.; Sheikh, S.; Zakaria, A.; Phillips, J. Treatment of sickle cell disease—Options and perspective. *Am. J. Blood Res.* **2023**, *13*, 61–70. [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37214647)
- <span id="page-16-10"></span>17. Eaton, W.A.; Bunn, H.F. Treating sickle cell disease by targeting HbS polymerization. *Blood* **2017**, *129*, 2719–2726. [\[CrossRef\]](https://doi.org/10.1182/blood-2017-02-765891) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28385699)
- <span id="page-16-19"></span><span id="page-16-11"></span>18. Oniyangi, O.; Cohall, D.H. Phytomedicines (medicines derived from plants) for sickle cell disease. *Cochrane Database Syst. Rev.* **2020**, *9*, Cd004448. [\[CrossRef\]](https://doi.org/10.1002/14651858.CD004448.pub7) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32977351)
- <span id="page-16-20"></span><span id="page-16-12"></span>19. Yembeau, N.L.; Biapa Nya, P.C.; Pieme, C.A.; Tchouane, K.D.; Kengne Fotsing, C.B.; Nya Nkwikeu, P.J.; Feudjio, A.F.; Telefo, P.B. Ethnopharmacological Study of the Medicinal Plants Used in the Treatment of Sickle Cell Anemia in the West Region of Cameroon. *Evid. Based Complement. Altern. Med.* **2022**, *2022*, 5098428. [\[CrossRef\]](https://doi.org/10.1155/2022/5098428) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35518347)
- <span id="page-16-21"></span><span id="page-16-13"></span>20. Famojuro, T.; Moody, J. Survey of medicinal plants used in the management of sickle cell disease by traditional medical practitioners of gbonyin local government area of Ekiti state, Nigeria. *Niger. J. Nat. Prod. Med.* **2015**, *19*, 78–84. [\[CrossRef\]](https://doi.org/10.4314/njnpm.v19i0.8)
- <span id="page-16-14"></span>21. Kaptchuk, T.J. The double-blind, randomized, placebo-controlled trial: Gold standard or golden calf? *J. Clin. Epidemiol.* **2001**, *54*, 541–549. [\[CrossRef\]](https://doi.org/10.1016/S0895-4356(00)00347-4) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11377113)
- <span id="page-16-23"></span><span id="page-16-22"></span>22. Giriraja, K.V.; Bhatnagar, S.K.; Tomlinson, L.; Sancilio, F. An open-label, multicenter, phase 2 study of a food enriched with docosahexaenoic acid in adults with sickle cell disease. *Prostaglandins Leukot. Essent. Fat. Acids* **2023**, *193*, 102574. [\[CrossRef\]](https://doi.org/10.1016/j.plefa.2023.102574) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37121179)
- <span id="page-16-24"></span>23. Abdelhalim, S.M.; Murphy, J.E.; Meabed, M.H.; Elberry, A.A.; Gamaleldin, M.M.; Shaalan, M.S.; Hussein, R.R.S. Comparative effectiveness of adding Omega-3 or Vitamin D to standard therapy in preventing and treating episodes of painful crisis in pediatric sickle cell patients. *Eur. Rev. Med. Pharmacol. Sci.* **2022**, *26*, 5043–5052. [\[CrossRef\]](https://doi.org/10.26355/eurrev_202207_29290) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35916800)
- <span id="page-16-25"></span>24. Brownell, J.N.; Schall, J.I.; McAnlis, C.R.; Smith-Whitley, K.; Norris, C.F.; Stallings, V.A. Effect of High-dose Vitamin A Supplementation in Children With Sickle Cell Disease: A Randomized, Double-blind, Dose-finding Pilot Study. *J. Pediatr. Hematol. Oncol.* **2020**, *42*, 83–91. [\[CrossRef\]](https://doi.org/10.1097/mph.0000000000001673) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31764511)
- <span id="page-16-26"></span>25. Muga, R.; Ajwang, A.; Ouma, J.; Ojigo, J.; Otieno, J.; Okoth, P.; Wafula, C.; Ajwang, S.; Ogolla, D.; Hollist, A. Efficacy of the Nutritional Supplement, EvenFlo, in the Management of Sickle Cell Disease: A Randomized Controlled Trial. *Nurs. Health Sci. Res. J.* **2020**, *3*, 35–45. [\[CrossRef\]](https://doi.org/10.55481/2578-3750.1058)
- <span id="page-16-27"></span>26. Grégoire-Pelchat, P.; Pastore, Y.; Robitaille, N.; LeMay, S.; Khamessan, A.; Kleiber, N.; Nyalendo, C.; Gagné, N.; Alos, N.; Mailhot, G. Comparison of two vitamin D supplementation strategies in children with sickle cell disease: A randomized controlled trial. *Br. J. Haematol.* **2021**, *192*, 385–394. [\[CrossRef\]](https://doi.org/10.1111/bjh.17119) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33169863)
- <span id="page-16-28"></span>27. Onalo, R.; Cooper, P.; Cilliers, A.; Vorster, B.C.; Uche, N.-A.; Oluseyi, O.O.; Onalo, V.D.; Zubairu, Y.; Ayodele-Kehinde, A.U.; Damilare, O.M.; et al. Randomized control trial of oral arginine therapy for children with sickle cell anemia hospitalized for pain in Nigeria. *Am. J. Hematol.* **2021**, *96*, 89–97. [\[CrossRef\]](https://doi.org/10.1002/ajh.26028) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33075179)
- <span id="page-16-29"></span>28. Onalo, R.; Cilliers, A.; Cooper, P.; Morris, C.R. Arginine therapy and cardiopulmonary hemodynamics in hospitalized children with sickle cell anemia: A prospective, double-blinded, randomized placebo-controlled clinical trial. *Am. J. Respir. Crit. Care Med.* **2022**, *206*, 70–80. [\[CrossRef\]](https://doi.org/10.1164/rccm.202108-1930OC) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35426778)
- <span id="page-16-30"></span>29. Hutchaleelaha, A.; Patel, M.; Washington, C.; Siu, V.; Allen, E.; Oksenberg, D.; Gretler, D.D.; Mant, T.; Lehrer-Graiwer, J. Pharmacokinetics and pharmacodynamics of voxelotor (GBT440) in healthy adults and patients with sickle cell disease. *Br. J. Clin. Pharmacol.* **2019**, *85*, 1290–1302. [\[CrossRef\]](https://doi.org/10.1111/bcp.13896) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30743314)
- <span id="page-17-23"></span><span id="page-17-6"></span><span id="page-17-5"></span><span id="page-17-4"></span><span id="page-17-3"></span><span id="page-17-2"></span><span id="page-17-1"></span><span id="page-17-0"></span>30. Vichinsky, E.; Hoppe, C.C.; Ataga, K.I.; Ware, R.E.; Nduba, V.; El-Beshlawy, A.; Hassab, H.; Achebe, M.M.; Alkindi, S.; Brown, R.C. A phase 3 randomized trial of voxelotor in sickle cell disease. *N. Engl. J. Med.* **2019**, *381*, 509–519. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa1903212) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31199090)
- <span id="page-17-12"></span>31. Abdullahi, S.U.; Jibir, B.W.; Bello-Manga, H.; Gambo, S.; Inuwa, H.; Tijjani, A.G.; Idris, N.; Galadanci, A.; Hikima, M.S.; Galadanci, N.; et al. Hydroxyurea for primary stroke prevention in children with sickle cell anaemia in Nigeria (SPRING): A double-blind, multicentre, randomised, phase 3 trial. *Lancet Haematol.* **2022**, *9*, e26–e37. [\[CrossRef\]](https://doi.org/10.1016/s2352-3026(21)00368-9)
- <span id="page-17-24"></span>32. Panda, P.C.; Mishra, N.R.; Patra, C.S.; Nayak, B.K.; Panda, S.K. Intravenous Acetaminophen vs Intravenous Diclofenac Sodium in Management of Skeletal Vaso-occlusive Crisis Among Children with Homozygous Sickle Cell Disease: A Randomized Controlled Trial. *Indian Pediatr.* **2021**, *58*, 229–232. [\[CrossRef\]](https://doi.org/10.1007/s13312-021-2160-3)
- <span id="page-17-27"></span>33. Rees, D.C.; Kilinc, Y.; Unal, S.; Dampier, C.; Pace, B.S.; Kaya, B.; Trompeter, S.; Odame, I.; Mahlangu, J.; Unal, S. A randomized, placebo-controlled, double-blind trial of canakinumab in children and young adults with sickle cell anemia. *Blood J. Am. Soc. Hematol.* **2022**, *139*, 2642–2652. [\[CrossRef\]](https://doi.org/10.1182/blood.2021013674) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35226723)
- <span id="page-17-30"></span>34. Heeney, M.M.; Abboud, M.R.; Githanga, J.; Inusa, B.P.D.; Kanter, J.; Michelson, A.D.; Nduba, V.; Musiime, V.; Apte, M.; Inati, A.; et al. Ticagrelor vs placebo for the reduction of vaso-occlusive crises in pediatric sickle cell disease: The HESTIA3 study. *Blood* **2022**, *140*, 1470–1481. [\[CrossRef\]](https://doi.org/10.1182/blood.2021014095) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35849650)
- <span id="page-17-31"></span>35. Alshahrani, M.S.; AlSulaibikh, A.H.; ElTahan, M.R.; AlFaraj, S.Z.; Asonto, L.P.; AlMulhim, A.A.; AlAbbad, M.F.; Almaghraby, N.; AlJumaan, M.A.; AlJunaid, T.O.; et al. Ketamine administration for acute painful sickle cell crisis: A randomized controlled trial. *Acad. Emerg. Med.* **2022**, *29*, 150–158. [\[CrossRef\]](https://doi.org/10.1111/acem.14382) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34449939)
- <span id="page-17-32"></span>36. Lizarralde-Iragorri, M.A.; Parachalil Gopalan, B.; Merriweather, B.; Brooks, J.; Hill, M.; Lovins, D.; Pierre-Charles, R.; Cullinane, A.; Dulau-Florea, A.; Lee, D.-Y.; et al. Isoquercetin for thromboinflammation in sickle cell disease: A randomized double-blind placebo-controlled trial. *Blood Adv.* **2024**, *8*, 172–182. [\[CrossRef\]](https://doi.org/10.1182/bloodadvances.2023011542) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38157227)
- <span id="page-17-7"></span>37. Raghuraman, A.; Lawrence, R.; Shetty, R.; Chaithanya, A.; Jhaveri, S.; Pichardo, B.V.; Mujakari, A. Role of gene therapy in sickle cell disease. *Dis. Mon.* **2024**, 101689. [\[CrossRef\]](https://doi.org/10.1016/j.disamonth.2024.101689) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38326171)
- <span id="page-17-8"></span>38. Dong, M.; McGann, P.T. Changing the Clinical Paradigm of Hydroxyurea Treatment for Sickle Cell Anemia Through Precision Medicine. *Clin. Pharmacol. Ther.* **2021**, *109*, 73–81. [\[CrossRef\]](https://doi.org/10.1002/cpt.2028) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32869281)
- <span id="page-17-9"></span>39. Yahouédéhou, S.C.M.A.; Adorno, E.V.; da Guarda, C.C.; Ndidi, U.S.; Carvalho, S.P.; Santiago, R.P.; Aleluia, M.M.; de Oliveira, R.M.; Gonçalves, M.d.S. Hydroxyurea in the management of sickle cell disease: Pharmacogenomics and enzymatic metabolism. *Pharmacogenomics J.* **2018**, *18*, 730–739. [\[CrossRef\]](https://doi.org/10.1038/s41397-018-0045-1) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30206297)
- <span id="page-17-10"></span>40. Jain, D.; Atmapoojya, P.; Colah, R.; Lodha, P. Sickle Cell Disease and Pregnancy. *Mediterr. J. Hematol. Infect. Dis.* **2019**, *11*, e2019040. [\[CrossRef\]](https://doi.org/10.4084/mjhid.2019.040) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31308916)
- <span id="page-17-11"></span>41. McGann, P.T.; Ware, R.E. Hydroxyurea therapy for sickle cell anemia. *Expert Opin. Drug Saf.* **2015**, *14*, 1749–1758. [\[CrossRef\]](https://doi.org/10.1517/14740338.2015.1088827)
- <span id="page-17-13"></span>42. Bell, V.; Varzakas, T.; Psaltopoulou, T.; Fernandes, T. Sickle Cell Disease Update: New Treatments and Challenging Nutritional Interventions. *Nutrients* **2024**, *16*, 258. [\[CrossRef\]](https://doi.org/10.3390/nu16020258)
- <span id="page-17-14"></span>43. Erkelens, M.N.; Mebius, R.E. Retinoic Acid and Immune Homeostasis: A Balancing Act. *Trends Immunol.* **2017**, *38*, 168–180. [\[CrossRef\]](https://doi.org/10.1016/j.it.2016.12.006) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28094101)
- <span id="page-17-15"></span>44. Schall, J.I.; Zemel, B.S.; Kawchak, D.A.; Ohene-Frempong, K.; Stallings, V.A. Vitamin A status, hospitalizations, and other outcomes in young children with sickle cell disease. *J. Pediatr.* **2004**, *145*, 99–106. [\[CrossRef\]](https://doi.org/10.1016/j.jpeds.2004.03.051) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/15238915)
- <span id="page-17-16"></span>45. Chandrakar, D.; Patel, S.; Wasnik, P.N.; Mohapatra, E.; Nanda, R.; Shah, S.; Gupta, D.L. Effect of Unmetabolized Folic Acid on Immunoinflammatory Markers in Sickle Cell Disease Patients Taking Folic Acid Supplementation. *Indian J. Clin. Biochem.* **2024**, *2024*, 1–8. [\[CrossRef\]](https://doi.org/10.1007/s12291-024-01204-0)
- <span id="page-17-17"></span>46. Soe, H.H.K.; Abas, A.B.; Than, N.N.; Ni, H.; Singh, J.; Said, A.; Osunkwo, I. Vitamin D supplementation for sickle cell disease. *Cochrane Database Syst. Rev.* **2020**, *5*, Cd010858. [\[CrossRef\]](https://doi.org/10.1002/14651858.CD010858.pub3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32462740)
- <span id="page-17-18"></span>47. Bakshi, N.; Morris, C.R. The role of the arginine metabolome in pain: Implications for sickle cell disease. *J. Pain Res.* **2016**, 167–175.
- <span id="page-17-19"></span>48. Morris, S.M. Chapter 11—Regulation of Arginine Availability and Its Impact on NO Synthesis. In *Nitric Oxide*; Ignarro, L.J., Ed.; Academic Press: San Diego, CA, USA, 2000; pp. 187–197.
- <span id="page-17-20"></span>49. Morris, C.R.; Hamilton-Reeves, J.; Martindale, R.G.; Sarav, M.; Ochoa Gautier, J.B. Acquired Amino Acid Deficiencies: A Focus on Arginine and Glutamine. *Nutr. Clin. Pract.* **2017**, *32*, 30S–47S. [\[CrossRef\]](https://doi.org/10.1177/0884533617691250) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/28388380)
- <span id="page-17-21"></span>50. Tayyaba Rehan, S.; Hussain, H.U.; Malik, F.; Usama, R.M.; Tahir, M.J.; Asghar, M.S. Voxelotor versus other therapeutic options for sickle cell disease: Are we still lagging behind in treating the disease? *Health Sci. Rep.* **2022**, *5*, e713. [\[CrossRef\]](https://doi.org/10.1002/hsr2.713) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35774831)
- <span id="page-17-22"></span>51. Barriteau, C.M.; Badawy, S.M. Practical Guidance for the Use of Voxelotor in the Management of Sickle Cell Disease. *J. Blood Med.* **2022**, *13*, 739–745. [\[CrossRef\]](https://doi.org/10.2147/jbm.S362222) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36471678)
- <span id="page-17-25"></span>52. Shastri, N. Intravenous acetaminophen use in pediatrics. *Pediatr. Emerg. Care* **2015**, *31*, 444–448. [\[CrossRef\]](https://doi.org/10.1097/PEC.0000000000000463)
- <span id="page-17-26"></span>53. Dampier, C.D.; Setty, B.; Logan, J.; Ioli, J.G.; Dean, R. Intravenous morphine pharmacokinetics in pediatric patients with sickle cell disease. *J. Pediatr.* **1995**, *126*, 461–467. [\[CrossRef\]](https://doi.org/10.1016/S0022-3476(95)70472-8)
- <span id="page-17-28"></span>54. Riley, T.R.; Riley, T.T. Profile of crizanlizumab and its potential in the prevention of pain crises in sickle cell disease: Evidence to date. *J. Blood Med.* **2019**, *10*, 307–311. [\[CrossRef\]](https://doi.org/10.2147/JBM.S191423)
- <span id="page-17-29"></span>55. Kutlar, A.; Kanter, J.; Liles, D.K.; Alvarez, O.A.; Cançado, R.D.; Friedrisch, J.R.; Knight-Madden, J.M.; Bruederle, A.; Shi, M.; Zhu, Z.; et al. Effect of crizanlizumab on pain crises in subgroups of patients with sickle cell disease: A SUSTAIN study analysis. *Am. J. Hematol.* **2019**, *94*, 55–61. [\[CrossRef\]](https://doi.org/10.1002/ajh.25308) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30295335)
- <span id="page-18-0"></span>56. Chan, K.H.; Buddharaju, R.; Idowu, M. Real-world experience of patients with sickle cell disease treated with crizanlizumab. *J. Investig. Med.* **2024**, *72*, 242–247. [\[CrossRef\]](https://doi.org/10.1177/10815589231220592) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38073007)
- <span id="page-18-1"></span>57. Wei, P.; Wang, X.; Fu, Q.; Cao, B. Progress in the clinical effects and adverse reactions of ticagrelor. *Thromb. J.* **2024**, *22*, 8. [\[CrossRef\]](https://doi.org/10.1186/s12959-023-00559-3) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38200557)
- <span id="page-18-2"></span>58. Tao, L.; Ren, S.; Zhang, L.; Liu, W.; Zhao, Y.; Chen, C.; Mao, X.; Chen, Z.; Gu, X. A Review of the Role of the Antiplatelet Drug Ticagrelor in the Management of Acute Coronary Syndrome, Acute Thrombotic Disease, and Other Diseases. *Med. Sci. Monit.* **2022**, *28*, e935664. [\[CrossRef\]](https://doi.org/10.12659/msm.935664) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35570383)
- <span id="page-18-3"></span>59. Heeney, M.M.; Abboud, M.R.; Amilon, C.; Andersson, M.; Githanga, J.; Inusa, B.; Kanter, J.; Leonsson-Zachrisson, M.; Michelson, A.D.; Berggren, A.R.; et al. Ticagrelor versus placebo for the reduction of vaso-occlusive crises in pediatric sickle cell disease: Rationale and design of a randomized, double-blind, parallel-group, multicenter phase 3 study (HESTIA3). *Contemp. Clin. Trials* **2019**, *85*, 105835. [\[CrossRef\]](https://doi.org/10.1016/j.cct.2019.105835) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31446143)
- <span id="page-18-4"></span>60. Ribeiro-Filho, J.; Yahouédéhou, S.C.M.A.; Pitanga, T.N.; Santana, S.S.; Adorno, E.V.; Barbosa, C.G.; Ferreira, J.R.D.; Pina, E.T.G.; Neres, J.S.d.S.; Leite, I.P.R.; et al. An evaluation of ticagrelor for the treatment of sickle cell anemia. *Expert Rev. Hematol.* **2020**, *13*, 1047–1055. [\[CrossRef\]](https://doi.org/10.1080/17474086.2020.1817736) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32972255)
- <span id="page-18-5"></span>61. Zhang, Y.; Ye, F.; Zhang, T.; Lv, S.; Zhou, L.; Du, D.; Lin, H.; Guo, F.; Luo, C.; Zhu, S. Structural basis of ketamine action on human NMDA receptors. *Nature* **2021**, *596*, 301–305. [\[CrossRef\]](https://doi.org/10.1038/s41586-021-03769-9) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34321660)
- <span id="page-18-6"></span>62. Davoudian, P.A.; Wilkinson, S.T. Chapter Four—Clinical overview of NMDA-R Antagonists and Clinical Practice. In *Advances in Pharmacology*; Duman, R.S., Krystal, J.H., Eds.; Academic Press: San Diego, CA, USA, 2020; Volume 89, pp. 103–129.
- <span id="page-18-7"></span>63. Syed, M.M.; Doshi, P.J.; Dhavale, D.D.; Doshi, J.B.; Kate, S.L.; Kulkarni, G.; Sharma, N.; Uppuladinne, M.; Sonavane, U.; Joshi, R.; et al. Potential of isoquercitrin as antisickling agent: A multi-spectroscopic, thermophoresis and molecular modeling approach. *J. Biomol. Struct. Dyn.* **2020**, *38*, 2717–2736. [\[CrossRef\]](https://doi.org/10.1080/07391102.2019.1645735)
- <span id="page-18-8"></span>64. Lizarralde, M.A.; Merriweather, B.; Conrey, A.; Saxena, A.; Shet, A.S. Effects of Flavonoid Quercetin on Thrombo-Inflammatory Processes in Patients with Sickle Cell Disease. *Blood* **2021**, *138*, 2020. [\[CrossRef\]](https://doi.org/10.1182/blood-2021-148601)
- <span id="page-18-9"></span>65. Telen, M.J.; Batchvarova, M.; Shan, S.; Bovee-Geurts, P.H.; Zennadi, R.; Leitgeb, A.; Brock, R.; Lindgren, M. Sevuparin binds to multiple adhesive ligands and reduces sickle red blood cell-induced vaso-occlusion. *Br. J. Haematol.* **2016**, *175*, 935–948. [\[CrossRef\]](https://doi.org/10.1111/bjh.14303) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/27549988)
- <span id="page-18-10"></span>66. White, J.; Lindgren, M.; Liu, K.; Gao, X.; Jendeberg, L.; Hines, P. Sevuparin blocks sickle blood cell adhesion and sickle-leucocyte rolling on immobilized L-selectin in a dose dependent manner. *Br. J. Haematol.* **2019**, *184*, 873–876. [\[CrossRef\]](https://doi.org/10.1111/bjh.15188) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/29767405)
- <span id="page-18-11"></span>67. Biemond, B.J.; Tombak, A.; Kilinc, Y.; Al-Khabori, M.; Abboud, M.R.; Nafea, M.; Inati, A.; Wali, Y.A.M.S.; Kristensen, J.; Donnelly, E.; et al. Efficacy and Safety of Sevuparin, a Novel Non-Anti-Coagulant Heparinoid, in Patients with Acute Painful Vaso-Occlusive Crisis; A Global, Multicenter Double-Blind, Randomized, Placebo-Controlled Phase 2 Trial (TVOC01). *Blood* **2019**, *134*, 614. [\[CrossRef\]](https://doi.org/10.1182/blood-2019-124653)
- <span id="page-18-12"></span>68. Subramanian, P.; Anandharamakrishnan, C. Chapter One—Introduction to Functional Foods and Nutraceuticals. In *Industrial Application of Functional Foods, Ingredients and Nutraceuticals*; Anandharamakrishnan, C., Subramanian, P., Eds.; Academic Press: San Diego, CA, USA, 2023; pp. 3–43.
- <span id="page-18-13"></span>69. Granato, D.; Barba, F.J.; Bursać Kovačević, D.; Lorenzo, J.M.; Cruz, A.G.; Putnik, P. Functional Foods: Product Development, Technological Trends, Efficacy Testing, and Safety. *Annu. Rev. Food Sci. Technol.* **2020**, *11*, 93–118. [\[CrossRef\]](https://doi.org/10.1146/annurev-food-032519-051708) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31905019)
- <span id="page-18-14"></span>70. Luvián-Morales, J.; Varela-Castillo, F.O.; Flores-Cisneros, L.; Cetina-Pérez, L.; Castro-Eguiluz, D. Functional foods modulating inflammation and metabolism in chronic diseases: A systematic review. *Crit. Rev. Food Sci. Nutr.* **2022**, *62*, 4371–4392. [\[CrossRef\]](https://doi.org/10.1080/10408398.2021.1875189) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33506690)
- <span id="page-18-15"></span>71. Ibrahim, A.; Muhammad, S.A. Antioxidant-Rich Nutraceutical as a Therapeutic Strategy for Sickle Cell Disease. *J. Am. Nutr. Assoc.* **2023**, *42*, 588–597. [\[CrossRef\]](https://doi.org/10.1080/27697061.2022.2108930) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36069788)
- <span id="page-18-16"></span>72. Teguem Tchoulegheu, A.; Nya Nkwikeu, P.J.; Lena Yembeau, N.; Choupo, A.C.; Nkenmeni Djamnou, C.; Feudjio, A.F.; Chetcha Chemegni, B.; Biapa Nya, P.C.; Pieme, C.A. Antisickling and Antihemolytic Mechanism of *Spirulina platensis* (*Oscillatoriaceae*): A Nutraceutical Commonly Used in Cameroon. *Evid. Based Complement. Altern. Med.* **2023**, *2023*, 1260169. [\[CrossRef\]](https://doi.org/10.1155/2023/1260169)
- <span id="page-18-17"></span>73. Ejiofor, E.U.; Ako, A.C.; Kube, M.T.; Agwamba, E.C.; Alala, C.; Maduabuchi, K.; Ejiofor, M. Phytochemistry, Mineral Estimation, Nutritional, and the In Vitro Anti-Sickling Potentials of Oil Extracted from the Seeds of Mucuna Flagellipes. *J. Mex. Chem. Soc.* **2024**, *68*, 220–233. [\[CrossRef\]](https://doi.org/10.29356/jmcs.v68i2.1898)
- <span id="page-18-18"></span>74. de Paula, R.G.; Ribeiro, H.M.; de Melo Borges, L.; Barreto, O.A.C.; Montel, A.L.B.; Scapin, E.; Silva, K.L.F.; Seibert, C.S. The Use of Natural Products in the Treatment of Sickle Cell Disease. *Rev. Bras. Farmacogn.* **2024**, *2024*, 1–13. [\[CrossRef\]](https://doi.org/10.1007/s43450-024-00535-6)
- <span id="page-18-19"></span>75. Sadowska-Bartosz, I.; Bartosz, G. Peroxiredoxin 2: An Important Element of the Antioxidant Defense of the Erythrocyte. *Antioxidants* **2023**, *12*, 1012. [\[CrossRef\]](https://doi.org/10.3390/antiox12051012)
- <span id="page-18-20"></span>76. Kotue, T. Functional foods and nutraceuticals in the primary prevention of sickle cell disease crises. *Indian J. Nutr.* **2018**, *5*, 186.
- <span id="page-18-21"></span>77. Famojuro, T.I.; Adeyemi, A.A.; Ajayi, T.O.; Fasola, F.A.; Fukushi, Y.; Omotade, O.O.; Moody, J.O. Anti-sickling activities of two isolated compounds from the root of Combretum racemosum *P. beauv.* (*Combretaceae*). *J. Ethnopharmacol.* **2021**, *273*, 113992. [\[CrossRef\]](https://doi.org/10.1016/j.jep.2021.113992) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33677007)
- <span id="page-18-22"></span>78. Ahajumobi, N.E.; Asika, J.C. Afro Medicinal Plants a Promising Remedy for Sickle Cell Anemia. *Int. Blood Res. Rev.* **2024**, *15*, 26–37. [\[CrossRef\]](https://doi.org/10.9734/ibrr/2024/v15i1332)
- <span id="page-19-0"></span>79. Wang, Q.; Zennadi, R. The Role of RBC Oxidative Stress in Sickle Cell Disease: From the Molecular Basis to Pathologic Implications. *Antioxidants* **2021**, *10*, 1608. [\[CrossRef\]](https://doi.org/10.3390/antiox10101608) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34679742)
- <span id="page-19-1"></span>80. Ilboudo, Y. The genetics of red blood cell density, a biomarker of clinical severity in sickle cell disease. Master's Thesis, Université de Montréal, Montreal, QC, Canda, 2017.
- <span id="page-19-2"></span>81. Liu, M.; Huang, Y.; Zhang, H.; Aitken, D.; Nevitt, M.C.; Rockel, J.S.; Pelletier, J.-P.; Lewis, C.E.; Torner, J.; Rampersaud, Y.R. Restricting branched-chain amino acids within a high-fat diet prevents obesity. *Metabolites* **2022**, *12*, 334. [\[CrossRef\]](https://doi.org/10.3390/metabo12040334) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35448521)
- <span id="page-19-3"></span>82. Jiang, L.; Liu, Y.; Zhou, Y.; Xu, Q.; Cheng, S.; Yan, J.; Xiao, Y.; Han, L.; Wang, Y.; Cai, W. Targeted metabolomics unravels altered phenylalanine levels in piglets receiving total parenteral nutrition. *Faseb J.* **2023**, *37*, e23014. [\[CrossRef\]](https://doi.org/10.1096/fj.202300261RR) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37261736)
- <span id="page-19-4"></span>83. Niihara, Y.; Miller, S.T.; Kanter, J.; Lanzkron, S.; Smith, W.R.; Hsu, L.L.; Gordeuk, V.R.; Viswanathan, K.; Sarnaik, S.; Osunkwo, I.; et al. A Phase 3 Trial of l-Glutamine in Sickle Cell Disease. *N. Engl. J. Med.* **2018**, *379*, 226–235. [\[CrossRef\]](https://doi.org/10.1056/NEJMoa1715971) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/30021096)
- <span id="page-19-5"></span>84. Daak, A.A.; Lopez-Toledano, M.A.; Heeney, M.M. Biochemical and therapeutic effects of Omega-3 fatty acids in sickle cell disease. *Complement. Ther. Med.* **2020**, *52*, 102482. [\[CrossRef\]](https://doi.org/10.1016/j.ctim.2020.102482) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/32951732)
- <span id="page-19-6"></span>85. Vona, R.; Sposi, N.M.; Mattia, L.; Gambardella, L.; Straface, E.; Pietraforte, D. Sickle Cell Disease: Role of Oxidative Stress and Antioxidant Therapy. *Antioxidants* **2021**, *10*, 296. [\[CrossRef\]](https://doi.org/10.3390/antiox10020296) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/33669171)
- <span id="page-19-7"></span>86. Karafin, M.S.; Field, J.J.; Ilich, A.; Li, L.; Qaquish, B.F.; Shevkoplyas, S.S.; Yoshida, T. Hypoxic storage of donor red cells preserves deformability after exposure to plasma from adults with sickle cell disease. *Transfusion* **2023**, *63*, 193–202. [\[CrossRef\]](https://doi.org/10.1111/trf.17163) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/36310401)
- <span id="page-19-8"></span>87. Hajizamani, S.; Atarodi, K.; Deyhim, M.R.; Kermani, F.R.; Hosseini, K.M. Antioxidative effects of α-tocopherol on stored human red blood cell units. *Asian J. Transfus. Sci.* **2023**. [\[CrossRef\]](https://doi.org/10.4103/ajts.ajts_130_22)
- <span id="page-19-9"></span>88. Wong-Roushar, J. Biochemical and Biophysical Characterization of Multi-Targeted Inhibitors of One-Carbon Metabolism for Cancer Treatment. Ph.D. Thesis, Indiana University, Bloomington, IN, USA, 2021.
- <span id="page-19-10"></span>89. Bhatt, S.; Argueta, D.A.; Gupta, K.; Kundu, S. Red Blood Cells as Therapeutic Target to Treat Sickle Cell Disease. *Antioxid. Redox. Signal* **2024**. [\[CrossRef\]](https://doi.org/10.1089/ars.2023.0348) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/37975291)
- <span id="page-19-11"></span>90. Ballas, S.K. The Evolving Pharmacotherapeutic Landscape for the Treatment of Sickle Cell Disease. *Mediterr. J. Hematol. Infect. Dis.* **2020**, *12*, e2020010. [\[CrossRef\]](https://doi.org/10.4084/mjhid.2020.010) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31934320)
- <span id="page-19-12"></span>91. Doganci, M.; Doganay, G.E. Magnesium levels and mortality relationship in patients with Acinetobacter baumannii detected in the Intensive Care Unit. *Eur. Rev. Med. Pharmacol. Sci.* **2024**, *28*, 1295–1305. [\[CrossRef\]](https://doi.org/10.26355/eurrev_202402_35451)
- <span id="page-19-13"></span>92. Aliev, G.; Li, Y.; Chubarev, V.N.; Lebedeva, S.A.; Parshina, L.N.; Trofimov, B.A.; Sologova, S.S.; Makhmutova, A.; Avila-Rodriguez, M.F.; Klochkov, S.G.; et al. Application of Acyzol in the Context of Zinc Deficiency and Perspectives. *Int. J. Mol. Sci.* **2019**, *20*, 2104. [\[CrossRef\]](https://doi.org/10.3390/ijms20092104) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/31035445)
- <span id="page-19-14"></span>93. Kotue, T.; Pieme, A.; Fokou, E. Ethnobotanicals usages in the management of sickle cell disease (SDC) in some localities of Cameroon. *Pharmacophore* **2016**, *7*, 192–200.
- <span id="page-19-15"></span>94. Öztaş, Y.; Boşgelmez, İ.İ. Oxidative Stress in Sickle Cell Disease and Emerging Roles for Antioxidants in Treatment Strategies. In *Pathology*; Elsevier: Amsterdam, The Netherlands, 2020; pp. 65–75.
- <span id="page-19-16"></span>95. Bleizgys, A. Zinc, Magnesium and Vitamin K Supplementation in Vitamin D Deficiency: Pathophysiological Background and Implications for Clinical Practice. *Nutrients* **2024**, *16*, 834. [\[CrossRef\]](https://doi.org/10.3390/nu16060834) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38542745)
- <span id="page-19-17"></span>96. Awor, S.; Bongomin, F.; Kaggwa, M.M.; Pebalo, F.P.; Musoke, D. Prevalence of Use of Herbal Medicines for the Treatment of Sickle Cell Disease in Africa: A Systematic Review and Meta-analysis. *J. Herb. Med.* **2023**, *42*, 100735. [\[CrossRef\]](https://doi.org/10.1016/j.hermed.2023.100735)
- <span id="page-19-18"></span>97. Cotoraci, C.; Ciceu, A.; Sasu, A.; Hermenean, A. Natural Antioxidants in Anemia Treatment. *Int. J. Mol. Sci.* **2021**, *22*, 1883. [\[CrossRef\]](https://doi.org/10.3390/ijms22041883)
- <span id="page-19-19"></span>98. Ameh, S.J.; Tarfa, F.D.; Ebeshi, B.U. Traditional herbal management of sickle cell anemia: Lessons from Nigeria. *Anemia* **2012**, *2012*, 607436. [\[CrossRef\]](https://doi.org/10.1155/2012/607436)
- 99. Takasu, J.; Uykimpang, R.; Sunga, M.A.; Amagase, H.; Niihara, Y. Aged Garlic Extract Is a Potential Therapy for Sickle-Cell Anemia13. *J. Nutr.* **2006**, *136*, 803S–805S. [\[CrossRef\]](https://doi.org/10.1093/jn/136.3.803S) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/16484568)
- <span id="page-19-20"></span>100. Ohnishi, S.T.; Ohnishi, T. In Vitro Effects of Aged Garlic Extract and Other Nutritional Supplements on Sickle Erythrocytes. *J. Nutr.* **2001**, *131*, 1085S–1092S. [\[CrossRef\]](https://doi.org/10.1093/jn/131.3.1085S) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/11238822)
- <span id="page-19-21"></span>101. Fokou, E.; Arumugam, N. HPLC profiling, in vitro antisickling and antioxidant activities of phenolic compound extracts from black bean seeds (*Phaseolus vulgarus* L.) used in the management of sickle cell disease in the West Region of Cameroon. *Nutr. Res.* **2019**, *3*, 30.
- <span id="page-19-22"></span>102. Afolabi, I.S.; Osikoya, I.O.; Fajimi, O.D.; Usoro, P.I.; Ogunleye, D.O.; Bisi-Adeniyi, T.; Adeyemi, A.O.; Adekeye, B.T. Solenostemon monostachyus, Ipomoea involucrata and *Carica papaya* seed oil versus Glutathione, or *Vernonia amygdalina*: Methanolic extracts of novel plants for the management of sickle cell anemia disease. *BMC Complement. Altern. Med.* **2012**, *12*, 262. [\[CrossRef\]](https://doi.org/10.1186/1472-6882-12-262) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/23259718)
- 103. Oduola, T.; Adeniyi, F.; Ogunyemi, E.; Idowu, T.; Bello, I. Evaluation of the effects of intake of extract of unripe pawpaw (*Carica papaya*) on liver function in sickle cell patients. *World J. Med. Sci.* **2007**, *2*, 28–32.
- <span id="page-19-23"></span>104. Imaga, N.; Gbenle, G.; Okochi, V.; Akanbi, S.; Edeoghon, S.; Oigbochie, V.; Kehinde, M.; Bamiro, S. Antisickling property of *Carica papaya* leaf extract. *Afr. J. Biochem. Res.* **2009**, *3*, 102–106.
- <span id="page-19-24"></span>105. Christianah, C.-O.M.; Ajayi, D.O.; Odunowo, O.O. Ethno medicinal survey and evaluation of two recipes used in managing sickle cell disease in Ile-Ife community of Osun-State, Nigeria. *Afr. J. Tradit. Complement. Altern. Med.* **2020**, *17*, 37–54.
- <span id="page-19-25"></span>106. Anorue, E.C.; Joshua, P.E. Evaluation of anti-sickling effects of two varieties of *Cajanus cajan* (L.) Huth on sickle cell beta thalassemia. *J. Ethnopharmacol.* **2024**, *331*, 118280. [\[CrossRef\]](https://doi.org/10.1016/j.jep.2024.118280) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/38714239)
- 107. Elemo, G.N.; Erukainure, O.L.; Okafor, J.N.C.; Banerjee, P.; Preissner, R.; Nwachukwu Nicholas-Okpara, V.A.; Atolani, O.; Omowunmi, O.; Ezeanyanaso, C.S.; Awosika, A.; et al. Underutilized legumes, *Cajanus cajan* and Glycine max may bring about antisickling effect in sickle cell disease by modulation of redox homeostasis in sickled erythrocytes and alteration of its functional chemistry. *J. Food Biochem.* **2022**, *46*, e14322. [\[CrossRef\]](https://doi.org/10.1111/jfbc.14322) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/35894096)
- 108. Yang, S.-E.; Vo, T.-L.T.; Chen, C.-L.; Yang, N.-C.; Chen, C.-I.; Song, T.-Y. Nutritional Composition, Bioactive Compounds and Functional Evaluation of Various Parts of *Cajanus cajan* (L.) Millsp. *Agriculture* **2020**, *10*, 558. [\[CrossRef\]](https://doi.org/10.3390/agriculture10110558)
- <span id="page-20-0"></span>109. Ohiagu, F.; Chikezie, P.; Chikezie, C. Sickle hemoglobin polymerization inhibition and antisickling medicinal plants. *J. Phytopharm.* **2021**, *10*, 126–133. [\[CrossRef\]](https://doi.org/10.31254/phyto.2021.10209)
- 110. Pauline, N.; Cabral, B.N.P.; Anatole, P.C.; Jocelyne, A.M.V.; Bruno, M.; Jeanne, N.Y. The in vitro antisickling and antioxidant effects of aqueous extracts Zanthoxyllum heitzii on sickle cell disorder. *BMC Complement. Altern. Med.* **2013**, *13*, 162. [\[CrossRef\]](https://doi.org/10.1186/1472-6882-13-162)
- <span id="page-20-1"></span>111. Okagu, I.U.; Ndefo, J.C.; Aham, E.C.; Udenigwe, C.C. Zanthoxylum Species: A Review of Traditional Uses, Phytochemistry and Pharmacology in Relation to Cancer, Infectious Diseases and Sickle Cell Anemia. *Front. Pharmacol.* **2021**, *12*, 713090. [\[CrossRef\]](https://doi.org/10.3389/fphar.2021.713090) [\[PubMed\]](https://www.ncbi.nlm.nih.gov/pubmed/34603027)
- <span id="page-20-2"></span>112. Iyekowa, O.; Okieimen, F.; Ehisuoria, C.O. In-vitro Antisickling Activity of *Pergularia daemia*, *Canna indica* and *Petiveria alliacea* Plants used in the Treatment of Sickle Cell Anaemia in Edo State, Nigeria. *Tanzan. J. Sci.* **2023**, *49*, 433–445. [\[CrossRef\]](https://doi.org/10.4314/tjs.v49i2.14)
- <span id="page-20-3"></span>113. Danladi, B.Y.; Geetha, P. Ways to improve Life Expectancy in Sickle Cell Anaemia Patients using Herbs. *Asian J. Res. Pharm. Sci.* **2017**, *7*, 205. [\[CrossRef\]](https://doi.org/10.5958/2231-5659.2017.00031.5)
- <span id="page-20-4"></span>114. Wambebe, C.; Ogunyale, P.; Gamaniel, K.; Nasipuri, R.; Okogun, J.; Samuel, B.; Olusola, A.; Orisadipe, A. Piper Guineense, Pterocarpus Osun, Eugenia Caryophyllata, and Sorghum Bicolor Extracts for Treating Sickle Cell Disease. U.S. Patent 5,800,819A, 1 September 1998.
- <span id="page-20-5"></span>115. Akinsulie, A.; Temiye, E.; Akanmu, A.; Lesi, F.; Whyte, C. Clinical evaluation of extract of *Cajanus cajan* (Ciklavit®) in sickle cell anaemia. *J. Trop. Pediatr.* **2005**, *51*, 200–205. [\[CrossRef\]](https://doi.org/10.1093/tropej/fmh097)
- <span id="page-20-6"></span>116. Tavares, L.; Smaoui, S.; Pinilla, C.M.B.; Ben Hlima, H.; Lopes Barros, H. Ginger: A systematic review of clinical trials and recent advances in encapsulation of its bioactive compounds. *Food Funct.* **2022**, *13*, 1078–1091. [\[CrossRef\]](https://doi.org/10.1039/D1FO02998C)
- <span id="page-20-7"></span>117. Tavares, L.; Santos, L.; Zapata Noreña, C.P. Bioactive compounds of garlic: A comprehensive review of encapsulation technologies, characterization of the encapsulated garlic compounds and their industrial applicability. *Trends Food Sci. Technol.* **2021**, *114*, 232–244. [\[CrossRef\]](https://doi.org/10.1016/j.tifs.2021.05.019)

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.