# Cytotoxic, antibacterial, antioxidant, and anticholinesterasic activity of Juniperus brevifolia extracts



Nemésia Oliveira,<sup>a</sup> Sofia Medeiros,<sup>a</sup> Maria Carmo Barreto,<sup>b,c</sup> Ana M. L. Seca<sup>c,d</sup> and José Silvino Rosa<sup>a,e</sup>

<sup>a</sup> Departamento de Biologia, Universidade dos Açores; Ponta Delgada, Açores; <sup>b</sup> Centro de Investigação de Recursos Naturais (CIRN), Universidade dos Açores, 9501-801 Ponta Delgada, Portugal. <sup>c</sup> Departamento de Ciências Tecnológicas e Desenvolvimento (DCTD), Universidade dos Açores, Rua Mãe de Deus, 9501-801 Ponta Delgada, Portugal. <sup>d</sup>QOPNA, Universidade de Aveiro, 3810-193 Aveiro, Portugal

eCIBIO Centro de Investigação em Biodiversidade e Recursos Genéticos, CIBIO-Açores, Departamento de Biologia, Universidade dos Açores.

### Introduction

Juniperus brevifolia, the unique conifer trees endemic to the Azorean archipelago, is locally known as "cedro-do-mato", and is classified as belong to Cupressaceae

family. This species is a typical component of the primitive laurisilva forest and occurs in almost all of the islands, except S. Maria and Graciosa. Old stands of some size are now rare, since J. brevifolia has been widely used in shipbuilding and carved work due to the high quality of its wood. Our interest on the study of J. brevifolia was the search for new bioactive compounds. In this context, the cytotoxic, antibacterial, antioxidant and anticholinesterasic (anti-AChE) activities of the methanol and acetone extracts of *J. brevifolia* wood and bark were determined.

## **Materials and Methods**

#### In vitro Artemia salina toxicity assay

The methodology described by Solis *et al.*<sup>1</sup> with some modifications was used. Briefly, 100 µL of sample with different concentrations and 100 µL of seawater containing 15-20 nauplii were placed in each well of a 96-well microplate. Two types of control were used, one with ethanol and another with potassium dichromate. The microplates were incubated at 25 °C for 24 hrs, then examined under the stereoscopic microscope (12.5 X), where the dead individuals (*i.e.*, without movement) were numbered. After 15 min, 50 µL of acetone were added to each well and total number of nauplii was counted. Each extract was tested in triplicate.

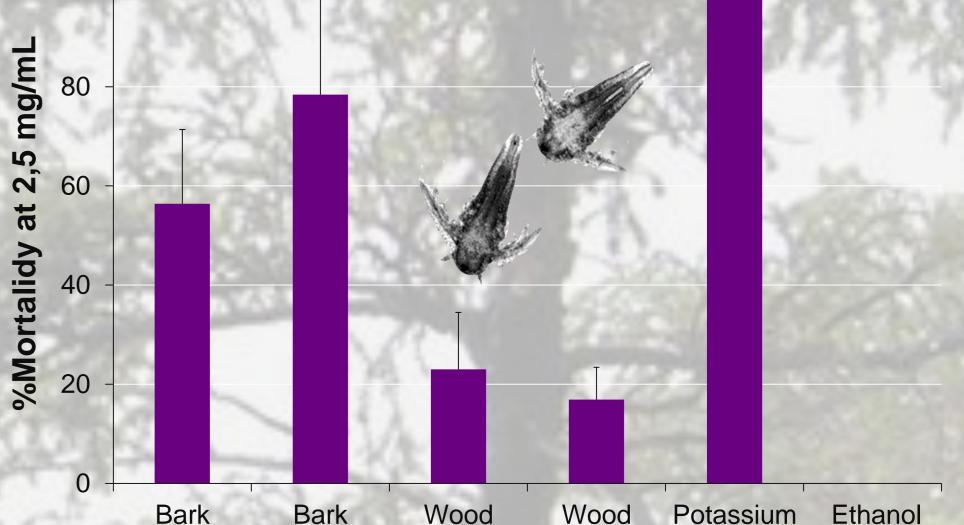
#### **Antibacterial activity**

From a culture of Gram(+) Bacillus subtilis (DSM 10), B. cereus (DSM31), Micrococcus luteus (DS 420030) and Gram(-)

### Results Cytotoxicity on the brine shrimp Artemia salina

The preliminary lethality bioassay showed that only bark extract present considerable activity (> 50% mortality) at 2.5 mg/mL.

LC values of bark methanolic and acetone extracts were found to be 2.03 and 2.12 mg/mL, respectively (Table 1).



Bark Bark Acetone Acetone Methanol Dichromate Methanol

Table 1. LC values for cytotoxicity on Artemia salina

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Escherichia coli (DSM 498) and Enterobacter cloacae (DSM 30850), the inoculums were prepared in 0.85% sterile saline (NaCI). The suspension was compared, using qualitative methods, with a suspension of barium sulphate Tube 0.5 McFarland Scale (bioMerieux®, France).

The antibacterial assay was performed by a modification of Bauer's diffusion technique<sup>2</sup> on solid medium using 6 mm filter paper discs. The diameter of inhibition caused by the addition of 10 µL of each extract was recorded after 24 h. Gentamicin and chloramphenicol were used as positive control.

#### **DPPH** radical antioxidant activity

Antioxidant activity was assessed by the DPPH radical scavenging activity assay using a modification of the Blois method<sup>3</sup>. Different concentrations of plant extracts were added to the assay medium containing DPPH solution, allowed to stand in the dark for 30 min and the absorbance at 515 nm measured and compared with controls absorbance without the extract samples. Antioxidant activity percentage (AA%) was calculated as AA% = 1 - (Ac-As)/Acx100, where Ac is the absorbance of control and As is the absorbance of each sample. Quercetin was used for comparison.

#### **Acetylcholinesterase inhibition**

The AChE activity determination was assessed following the assay

Park avtract	Concentration (mg/mL)	Artemia salina			
Bark extract		LC <sub>50</sub> (95% CL)	LC <sub>90</sub> (95% CL)	Slope ±SEM (95% CL)	
Methanol	(1, 1.25, 1.5, 2.0, 2.5)	2.03 a (1.87 - 2.25)	4.53 a (3.69 – 6.27)	3.67 ± 0.44 (2.80 – 4.53)	
Acetone	(1, 1.25, 1.5, 2.0, 2.5)	2.12 a (2.02 - 2.25)	3.55 a (3.18– 4.19)	5.72 ± 0.60 (3.69 – 6.27)	

LC values and 95% confidence limits (CL) expressed in mg/mL. LC values and slopes within a column followed by the same letter are not significantly different based on nonoverlapping 95% CL.

Table 2. In vitro antibacterial activity of methanol and acetone extract of J. brevifolia Antibacterial activ   Diameter of inhibition (mm) (mean ± SD) The bark acetone of t								
Test organism	Metha extract (2)	anol	Acet	one	Gentamicin	Cloramphenicol	The bark acetone extract showed activity against <i>Bacillus cereus</i> , <i>B. subtilis</i>	
	bark	wood	bark	wood	(10 µg/disc)	(10 µg/disc)	and Micrococus luteus, while	
B. subtilis	0	0	9 ± 1	0	2012 -	31 ± 0.6	the wood acetone extract	
M. luteus	0	0	10 ± 0	0	-	32 ± 1.5	showed activity only against	
B. cereus	$10 \pm 0.6$	0	11 ± 0.6	10 ± 1	24 ± 0.6	-	B. cereus.	
E. coli	0	0	0	0	-	17 ± 0.6	No activity was observed	
E. cloacae	0	0	0	0	20 ± 0	-	against Gram (-) bacteria.	

#### Table 3. Anti-AChE and antioxidant activity of

described by Arruda et al.<sup>4</sup> at pH 8.0 and using acetylthiocholine iodide as substrate, in the presence of buffer containing DTNB and extract in different concentrations DMSO never exceeded 2.5%. The reaction was followed for 7.5 min at 415 nm, using e Bio-Rad Model 680 Microplate reader. Every experiment was done in triplicate.

### Conclusion

The present study show that bark extracts of *J. brevifolia* are a plant source with interesting biological properties, justifying further detailed investigations of its potential applications.

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metanol and acetone extract of J. brevifolia

Plant extract	AChE inhibitory activity	DPPH scavenging activity	Antioxidant activity The acetone extracts showed good activity when compared to		
(parts used)	IC <sub>50</sub> (mg/mL)	<b>EC</b> <sub>50</sub> (μg/mL)			
Methanol			quercetin (Table 3).		
bark	0.499 ± 0.021	33.0 ± 6.81			
wood	0.742 ± 0.023	49.6 ± 0.14	AChE inhibition		
Acetone			The methanol and acetone extracts		
bark	0.193 ± 0.042	7.0 ± 0.68	from bark and wood presented		
wood	0.825 ± 0.294	15.0 ± 0.74	$IC_{50}$ values between 0.193 and		
Quercetin		3.24 ± 0.07	0.825 mg/mL (Table 3).		

#### References

1. Solis, P.N.; Wright, C.W.; Anderson, M.; Gupta, M.P.; Philhipson, J.D. Planta Med. 1993, 59, 250-252. 2. Brauer, A.W.; Kirby, M.M.; Sherris, J.C.; Truck, M. Am J Clin Pathol 1966, 45, 493-496. 3. Blois, M.S. Nature 1961, 181, 1199-1200. 4. Arruda, M.; Viana, H.; Rainha, N.; Neng, N.R.; Rosa, J.S.; Nogueira, J.M.F.; Barreto, M.C. Molecules 2012, 17, 3082-3092.