


## NATURE NOTES

# Early prey intake of a short-finned pilot whale (*Globicephala macrorhynchus* Gray, 1846, Cetacea: Delphinidae) in the Canary Islands

Amanda Luna<sup>1,2</sup>  | Alejandro Escáñez<sup>2,3</sup> | Jacobo Marrero<sup>4</sup> | Eva Íñiguez<sup>3,4,5</sup> | José A. Pérez<sup>6</sup> | Pilar Sánchez<sup>7</sup>

<sup>1</sup>BioCephaLab, Centro de Investigación Mariña de la Universidade de Vigo, Edificio de Ciencias Experimentais, Vigo, Spain

<sup>2</sup>Departamento de Ecoloxía e Bioloxía Animal, Edificio de Ciencias Experimentais, Campus As Lagoas-Marcosende, Universidade de Vigo, Vigo, Spain

<sup>3</sup>MARE-Marine and Environmental Sciences Centre, ARDITI, Edifício Madeira Tecnopolo, Caminho da Penteadá, Funchal, Madeira Island, Portugal

<sup>4</sup>Asociación Tonina, San Cristóbal de La Laguna, Tenerife (Islas Canarias), Spain

<sup>5</sup>Faculty of Life Sciences, University of Madeira, Funchal, Madeira Island, Portugal

<sup>6</sup>Departamento de Biología Animal, Edafología y Geología, Universidad de La Laguna, San Cristóbal de La Laguna, Spain

<sup>7</sup>Department of Marine Renewable Resources, Institute of Marine Sciences-CSIC, Barcelona, Spain

## Correspondence

Amanda Luna, BioCephaLab, Centro de Investigación Mariña de la Universidade de Vigo, Edificio de Ciencias Experimentais, Campus As Lagoas-Marcosende, 36310 Vigo, Spain.  
Email: [amluna@uvigo.gal](mailto:amluna@uvigo.gal)

## Funding information

Universidade de Vigo/CISUG, Grant/Award Number: 33.50.460A.752; European Union NextGeneration EU/PRTR through a Margarita Salas contract of the University of Vigo; Ministry of Science and Innovation, Grant/Award Number: CTM2017-88686-P

## Abstract

This study reveals early prey eating by a short-finned pilot whale (*Globicephala macrorhynchus* Gray, 1846, Cetacea: Delphinidae) in the Canary Islands. Stomach contents, trophic markers, skin isotopic ratios of nitrogen ( $\delta^{15}\text{N}$ : $^{15}\text{N}/^{14}\text{N}$ ) and carbon ( $\delta^{13}\text{C}$ : $^{13}\text{C}/^{12}\text{C}$ ), and fatty acid profiles of the blubber of a short-finned pilot whale of 213 cm size euthanized in free-ranging conditions were analyzed. A total of 15 species of oegopsid squids, mostly diel vertical mesopelagic migrant species of the families Eupoloteuthidae, Ommastrephidae, and Histioteuthidae, as well as mother's milk, were identified in the stomach contents. *Asperoteuthis acanthoderma* (Lu, 1977, Cephalopoda: Chiroteuthidae) was found as first time in this area, suggesting the possibility of its presence on both sides of the subtropical Atlantic, extending its current known distribution. The  $\delta^{15}\text{N}$  value (11.55‰) was higher than expected based on the size range of squid ingested, but lower than that of adult pilot whales, suggesting that mother's milk intake has a significant effect on these values in calves. Similarly, the  $\delta^{13}\text{C}$  values (-17.99‰) were shifted to those of adult pilot whales rather than the ingested squids, also due to the ingestion of high-fat breast milk. The fatty acid (FA) composition of blubber showed a clear stratification. Long-chain polyunsaturated fatty acids (LC-PUFA) were mainly present in the inner layer, while most relevant  $\leq\text{C}20$  monounsaturated fatty acids (MUFA) were more abundant in the outer layer.

## KEYWORDS

fatty acids, *Globicephala macrorhynchus*, Macaronesia, squids, stable isotopes, stomach contents, teuthophagous diet

## TAXONOMY CLASSIFICATION

Conservation ecology

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2024 The Authors. *Ecology and Evolution* published by John Wiley & Sons Ltd.

## 1 | INTRODUCTION

The ontogeny of hunting behavior in cetaceans, which involves the development of cognitive, echolocation and diving skills, remains poorly understood for most cetaceans, especially for deep diving odontocetes. First, prey's intake in calves of deep diving odontocetes remains widely unknown and, to the best of our knowledge, this sort of data is restricted to some of them: the sperm whale (*Physeter macrocephalus* Linnaeus, 1758, Cetacea: Physeteridae), the Risso's Dolphin (*Grampus griseus* G. Cuvier, 1812, Cetacea: Delphinidae), the Baird beaked whale (*Berardius bairdii* Stejneger, 1883, Cetacea: Ziphiidae), and the southern bottlenose whale (*Hyperoodon planifrons* Flower, 1882, Cetacea: Ziphiidae). They are all teuthofagous species, i.e. they feed mainly on oceanic cephalopod species (Dixon et al., 1994; Luna et al., 2022; Santos et al., 2006; Walker et al., 2002). It has been documented that the calves of some marine mammals complement their lactation with solid food during their early life cycle, as occur in Risso's dolphins, which feed on oceanic cephalopods while continuing suckling (Blanco et al., 2006). However, detailed data on cephalopods consumed by these young specimens of deep diving cetaceans are not treated individually in the published articles so hampering the understanding of their early trophic ecology. The first solid feeding in these species seems related with the development of the diving capacities as well as their echolocation skills. In this sense, sperm whales have demonstrated early aptness to perform deep foraging dives below 600m depth (Tønnesen et al., 2018). In addition, Best et al. (1984) found solid food items in the stomach contents of a one-year-old sperm whale, which may reflect independent foraging or food provisioning by adults. Biologging studies have revealed that other deep divers such as Blainville's beaked whale (*Mesoplodon densirostris* de Blainville, 1817, Cetacea: Ziphiidae) show highly synchronization in adult diving behavior, which is consistent with observations made from surface, where all group members dive and re-surface in close coordination (Aguilar de Soto et al., 2020). Young members of these groups, estimated around 3 months, 18 months, and 2 years old, produce clicks in the same way adults and appear to always remain with their mothers, diving and surfacing in synchrony (Dunn et al., 2017), which could indicate a similar capacity for diving and foraging as the adults.

Among deep diving odontocetes, the short-finned pilot whale (*Globicephala macrorhynchus* Gray, 1846, Cetacea: Delphinidae), herein after referred to as pilot whale, has been described as a mainly teuthofagous species that can supplement its diet with fish and crustaceans (Öztürk et al., 2007; Riccialdelli et al., 2013). Thus, its distribution and habitat use have been related to the distribution, abundance and ease of capture of its main food resource, squids (e.g., Copeland et al., 2019; Hui, 1985; Owen et al., 2019; Seagars & Henderson, 1985; Sinclair, 1992). However, the pilot whales have shown a plasticity to adapt their feeding behavior to regional conditions (Shearer et al., 2022). This plasticity explains why they feed in separate regions on ecologically different squid species (from neritic to mesopelagic), and why they can prey on fish in considerable quantities in some areas (Bustamante et al., 2003;

Mintzer et al., 2008). Despite this available information, few stomach contents of this species have been analyzed and published worldwide (Fernández et al., 2009; Mintzer et al., 2008; Overholtz & Waring, 1991; Seagars & Henderson, 1985), which means that there is still limited knowledge of its diet in the different regions where it is found, and even more with regard to the youngest individuals. There are also scarce studies on the diet or trophic ecology of this species using trophic biomarkers such as stable isotopes and fatty acid (FA) profiling, which have been widely used for other cetaceans (e.g., Bowen & Iverson, 2013). Stable isotopes, particularly of  $\delta^{13}\text{C}$  and  $\delta^{15}\text{N}$ , are commonly used in ecological studies, the first to determine the carbon source at the base of the food chain and the feeding habitat of individuals, and latter to study the trophic position of the individuals in relation to their prey (Xavier et al., 2022). This technique, when used on cephalopod beaks, has some advantages, such as that the stable isotopic composition is independent of the period of time during which the beak has been preserved in collections, and that the method of preservation (e.g. dried, frozen, in ethanol, in formalin) does not affect the results, being comparable for different types of studies (Xavier et al., 2022). Fatty acid composition is used to analyze some important dietary parameters of the species, such as final biomass, specific growth rate, or feed intake (Torres et al., 2013) according to the consumption of different kind of food, by being able to compare the nutritional nature of the ecological niche of preys.

In this sense, trophic ecology based on fresh tissues isotopic values of pilot whales have only been reported for few regions, including Gulf of California (Western Pacific), Moorea Island (South Pacific), and the Iberian Peninsula (Northwest Atlantic) (Aurioles-Gamboa et al., 2013; Kiszka et al., 2010; Monteiro et al., 2017).

Tenerife island (Spain) holds one of the few resident populations of pilot whales worldwide, jointly with other oceanic islands such as Madeira and Hawaii (Alves et al., 2013; Heimlich-Boran, 1993; Servidio et al., 2019; Shane & McSweeney, 1990). Previous works on adult pilot whales from Tenerife using motion and digital acoustic recording tags (DTAGs) (Johnson & Tyack, 2003), have shown that these animals feed at average depths between  $810 \pm 92\text{m}$  during the day, and  $141 \pm 81\text{m}$  at night (Aguilar de Soto et al., 2008). It has been suggested a daytime foraging tactic focused on large, calorific, and mobile prey items, such as large squids, that forces pilot whales to perform sprints up to 9 m/s in depths between 538 and 1019 m. In contrast, during night time, foraging dives are shallower and are characterized by steady swimming speeds and multiple prey capture attempts per dive (Aguilar de Soto et al., 2008). However, little is known about the stomach contents of this species in the Canary Islands, being impossible to confirm this hypothesis and, therefore, to have a better understanding of the population feeding habits. To date, only three stomach contents have been analyzed, showing remains of mesopelagic squids, mainly species of the Cranchiidae Prosch, 1847 family (Fernández et al., 2009; Hernández-García & Martín, 1994).

Here, we present data on the simultaneous analyses of the stomach contents, bubbler stable isotopes, and FA signatures of a still

lactating pilot whale calf found in the open sea off Tenerife Island. This case represents a unique opportunity to boost the understanding of the early hunting behavior and prey's selection of the offspring of this pilot whale population.

## 2 | MATERIALS AND METHODS

### 2.1 | Short-finned pilot whale capture and euthanasia

During a survey campaign conducted on 24th March 2019 in the Special Area of Conservation (SAC) "Franja Marina de Teno-Rasca" (European Habitats Directive 92/43/EEC, Figure 1), southwest off Tenerife, the Asociación Tonina received a radio alert from a whale-watching boat about an injured pilot whale calf that had been sighted in the area (28°05'79.13" N, 16°49'53.63" W). Permission for the operation and the handling of the animal was granted by the Spanish Ministry of Environment. The association's team and a professional underwater photographer with administrative permission to photograph cetaceans in the SAC navigated to the position. A pilot whale calf was located resting on surface (28°06'04.0" N, 16°49'12.2" W). The animal had difficulties to swim and to remain upright. Acoustic and photoidentification data, underwater photos and videos (Canon 5D MKIV, Seacam housing), showed that the animal's caudal fin was almost completely cut off near the caudal peduncle (Figure 2; Appendix S1: video link 1). Simultaneously, three adults pilot whales were approaching, presumably alerted by the calls that the calf was producing repeatedly. The team proceeded by sending the audiovisual material to the authorities, notifying them the critical situation of the animal. Once they confirmed that the veterinarian team from the Wildlife Recovery Center "La Tahonilla"

(Cabildo of Tenerife) were moving to the area, we proceeded according to the stranding surveillance protocol. Whale-watching vessels in the area were asked to keep their distance. Four hours later, the veterinarian staff arrived in a Zodiac and assessed the health status of the calf in situ. The severity of the injuries made it impossible for the animal to survive in the wild on its own, so the chief veterinarian decided to euthanize it.

The maneuver was performed with two zodiacs, skippered by the qualified staff of both institutions. Then, the animal was captured with the help of a net stretched underneath the animal between the two boats and euthanized (intraperitoneal injection of 100cc/130 kg of Dolethal (pentobarbital) with xylazine, to facilitate the quick diffusion of the anesthetic into the bloodstream). Few minutes later the animal perished, and his body was transferred to the Wildlife Recovery Center.

### 2.2 | Photoidentification

Dorsal fin photographs of pilot whales accompanying the calf were compared with the Tenerife Island species photo-identification catalog (Pimentel et al., 2023). This catalog is regularly updated and contains photo cards of more than a thousand recognized individuals taken on field cruises from 2014 to date.

### 2.3 | Sample collection and preparation

Necropsy was conducted by staff of the University Institute of Animal Health and Food Safety (IUSA), Veterinary School, University of Las Palmas de Gran Canaria (ULPGC), following standardized protocols (Kuiken & García-Hartmann, 1991).

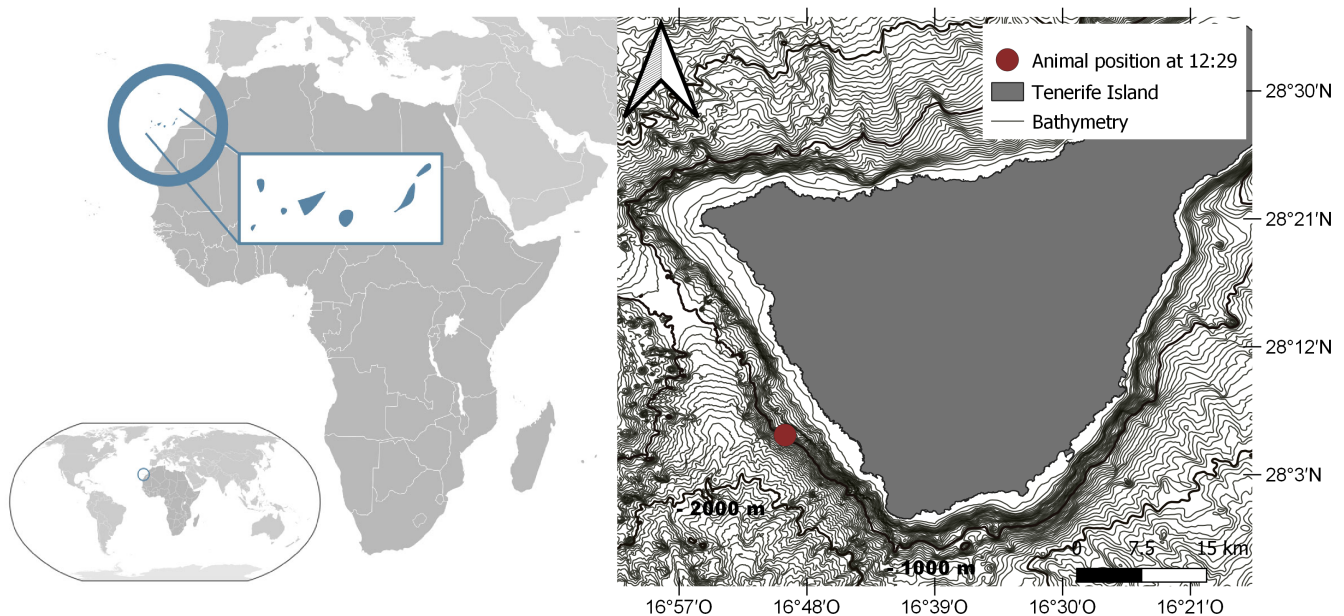


FIGURE 1 Special Area of Conservation (SAC) "Franja Marina de Teno-Rasca" (European Habitats Directive 92/43/EEC), southwest off Tenerife, where the Asociación Tonina caught the injured pilot whale calf (28°05'79.13" N, 16°49'53.63" W).



**FIGURE 2** Injured young specimens of *Globicephala macrorhynchus* named “Hope” found in southwest coast off Tenerife, 24/03/2019. © Francis Pérez.

Samples of skin, blubber and muscle were collected from the surface body in front of the dorsal fin of the animal and were stored at  $-80^{\circ}\text{C}$  until analysis. Skin ( $N=1$ ) was removed and used to analyze stable isotopes, while blubber was separated into three layers: the outer layer closest to the skin (0.9 cm), the middle layer (0.4 cm), and the inner layer (1 cm) closest to the muscle. These samples were used for the FA determination. The stomach contents were temporarily frozen at  $-20^{\circ}\text{C}$  until their analysis in the laboratory.

## 2.4 | Stomach content identification

Cephalopod beaks were cleaned and preserved in 70% ethanol. They were used for specific identification according to Clarke's (1986) and Pérez-Gándaras' (1983) methods. The lower beaks were identified using the available specific literature (Clarke, 1986; Pérez-Gándaras, 1983; Roper, 1966; Xavier & Cherel, 2021), and two cephalopod beaks reference collection from the Institute de Ciencias del Mar (ICM-CSIC, Barcelona), the general collection and the AFOC (Shape Analysis of Cephalopods' Beaks) collection, for corroboration of identifications. The Lower Rostral Length (LRL), Lower Crest Length (LC), Lower Hood Length (LHL), and Lower Width (LW) of the cephalopod lower beaks was measured (vernier calipers, 0.1 mm), and the following indexes were calculated: LC/LHL, LHL/LRL, LRL/LW, LH/LW, LC/LW, to compare them with the described in literature (Clarke, 1986; Pérez-Gándaras, 1983). Allometric equations for dorsal mantle length (ML) and mass (W) estimation for cephalopods were taken from the literature (Clarke, 1986; Lu & Ickeringill, 2002; Pérez-Gándaras, 1983; Rodhouse et al., 1990; Roper, 1966).

## 2.5 | Stable isotopes analyses

Lipids were extracted from the skin sample using 4 mL of cyclohexane for 1 h; the sample was centrifuged for 5 min at

4000g, and the supernatant containing the lipids discarded (Méndez-Fernández et al., 2012). Then, the sample was dried in an oven at  $45^{\circ}\text{C}$ , for 48 h, and  $0.3 \pm 0.05$  mg subsamples of lipid-free dried powder were finally weighed in tin capsules for stable isotope analyses. These analyses were performed with an isotope ratio mass spectrometer (IRMS-MAT 253, Finnigan™) coupled to a continuous flow interface (ConFlo III) in the lab facilities of CACTI-University of Vigo (Vigo, Spain). The results are presented in the usual  $\delta$  notation relative to Vienna PeeDee Belemnite Standard (VPDB) for  $\delta^{13}\text{C}$  and atmospheric  $\text{N}_2$  for  $\delta^{15}\text{N}$ , in parts per thousand (‰) expressed as  $\delta X^Y = [(R_{\text{sample}} - R_{\text{standard}}) / (R_{\text{standard}})]$ , where  $X$  is the element,  $y$  is the atomic mass of the stable isotope, and  $R$  is the ratio of heavy to light isotopes.

## 2.6 | Fatty acid profile analyses and stratification index

Lipids were independently extracted from each blubber sample (inner, middle, and outer layer) with chloroform-methanol (2:1, v/v) following a modified version of Folch et al. (1957) method as described by Galindo et al. (2022). An aliquot of 1 mg of lipid was subjected to acid-catalyzed transmethylation (Christie, 1982), and the obtained fatty acid methyl esters (FAME) were purified by thin layer chromatography (Christie & Han, 2010). The resultant FAME were resuspended in hexane with butylated hydroxytoluene (BHT) and stored at  $-20^{\circ}\text{C}$  until further analysis.

For the identification and quantification of FA, a TRACE-GC Ultra gas chromatograph (Thermo Scientific, Milan, Italy) equipped with an on-column injection, a flame ionization detector (FID) set at  $250^{\circ}\text{C}$ , and a Supelcowax™ 10 fused silica capillary column (30 m  $\times$  0.32 mm I.D. 0.25  $\mu\text{m}$  thickness; Supelco Inc., Bellefonte, Pennsylvania, USA) was used. Helium was employed as the carrier gas, and temperature programming began at an initial temperature of  $50^{\circ}\text{C}$ , increased at  $40^{\circ}\text{C}/\text{min}$  to  $150^{\circ}\text{C}$ , from  $150^{\circ}\text{C}$  to  $200^{\circ}\text{C}$  at  $2^{\circ}\text{C}/\text{min}$ , to  $214^{\circ}\text{C}$  at  $1^{\circ}\text{C}/\text{min}$ , and to a final temperature of  $230^{\circ}\text{C}$  at  $40^{\circ}\text{C}/\text{min}$ , which was maintained for 5 min. A mixture of authentic standards (Mix C4-C24 and PUFA No. 3 from menhaden oil [Sigma Aldrich, Darmstadt, Germany]) and a well-characterized cod roe oil were used to identify individual FAME. If needed, the identity of FAME was confirmed by GC-MS (DSQ II, Thermo Scientific).

To identify the differences in FAME composition between layers, a stratification index (SI) was calculated, following Guerrero and Rogers (2017) and Olsen and Grahl-Nielsen (2003). This index is based on the composition differences between the outer and inner layers of the blubber showing a possible stratification. SI is calculated taking the percentage concentration of each FA in the outer and inner layers, according to the equation by Olsen and Grahl-Nielsen (2003):  $\text{SI} = (F_o - F_i) / ((F_o + F_i) / 2)$ , where  $F_o$  is the proportion of a FA in the outer layer, and  $F_i$  is the proportion of the same FA in the inner layer.

### 3 | RESULTS

#### 3.1 | Necropsy results and photo-identification

Preliminary necropsy results revealed that the euthanized animal was a 213 cm long, less than one-year-old female (Kasuya & Marsh, 1984). The causes of the injuries remains under investigation. This animal was named 'Hope' (without code) and the accompanying adults were photoidentified as: 'Alejandrina' (GM\_CC\_I\_443) and 'Tere' (GM\_CC\_D\_712), both resident animals that have been repeatedly sighted forming part of well-known groups in the south-west of Tenerife (Pimentel et al., 2023). Hope's presumed mother (animal that stood next to her, traveling together) could not be identified, because her dorsal fin had no marks that helped in its identification.

#### 3.2 | Stomach contents analysis

A total of 144 cephalopod beaks remained in the stomach. It contained 77 upper and 67 lower beaks, a number of cephalopod eye lenses and sucker rings, all of them mixed with milk. 77.6% (52 individuals) of lower beaks were incomplete, with only the hood remaining, broken wings or broken crest, making it difficult to identify them correctly at species level. Nevertheless, a total of 15 different taxa of oegopsid squids were identified from the lower beaks. The list of cephalopod species recovered from the stomach is shown in Table 1.

The family with the highest species diversity in the stomach content was the Ommastrephidae Steenstrup, 1857 with six species (30.29% of the total), followed by Histioteuthidae Verrill, 1881 with four species (22.72%). Other families including Chiroteuthidae Gray, 1849, Cranchiidae, Enoploteuthidae Pfeffer, 1900, Lepidoteuthidae Pfeffer, 1912, and Bathyteuthidae Pfeffer, 1900, were represented only by one species (Table 1). In addition, the three families best represented in number were the enoploteuthids (42.42%), followed by the ommastrephids (30.3%), and the histioteuthids (22.77%). *Enoploteuthis leptura* (Leach, 1817) (Cephalopoda: Enoploteuthidae) was the most abundant species in stomach contents ( $N=22$ ; 33.33% of the total), followed by *Histioteuthis bonnellii* (Férussac, 1834) (Cephalopoda: Histioteuthidae) ( $N=8$ ; 12.12%), and *Hyaloteuthis pelagica* (Bosc, 1802) (Cephalopoda: Ommastrephidae) ( $N=7$ ; 10.60%).

#### 3.3 | Trophic markers analyses

Stable isotopes from skin samples showed values of  $\delta^{15}\text{N}$  of 11.55‰ and  $-17.99\text{‰}$  for  $\delta^{13}\text{C}$ . A total of 31 FA with values  $\geq 0.1\%$  were identified in the blubber samples (Table 2). Among them, five FA including oleic acid (18:1n-9,  $38.09 \pm 3.08\%$ ), palmitic acid (16:0,  $12.38 \pm 1.58\%$ ), palmitoleic acid (16:1n-7,  $11.87 \pm 0.88\%$ ), eicosenoic acid (20:1n-9,  $5.33 \pm 0.21$ ), and docosahexaenoic acid (22:6n-3,  $4.47 \pm 1.22\%$ ), were the most abundant ones representing more than 70% of the total (Table 2). Overall, FA families were dominated by monounsaturated fatty acids (MUFA;  $65.11 \pm 3.57\%$ ); followed

by saturated fatty acids (SFA;  $20.23 \pm 2.16\%$ ), and polyunsaturated fatty acids (PUFA;  $13.17 \pm 3.5\%$ ). Individually, all LC-PUFA except 20:2n-6, were notably more abundant in the inner layer than in the outer one. Thus, the SI values obtained were 20:3n-3 ( $-0.21$ ), 20:4n-3 ( $-0.20$ ), 20:5n-3 ( $-0.40$ ), 22:5n-3 ( $-0.38$ ), 22:6n-3 ( $-0.55$ ), and 20:4n-6 ( $-0.04$ ), 22:2n-6 ( $-2$ ), 22:4n-6 ( $-0.24$ ), and 22:5n-6 ( $-0.26$ ) (Table 2). Interestingly, most  $\leq \text{C}20$  MUFA, including 14:1, 16:1, 17:1, and 18:1 were more abundant in the outer layer. As a consequence, total PUFA (SI =  $-0.29$ ) and total SFA (SI =  $-0.15$ ) were substantially more abundant in the inner layer, whereas total MUFA was in the outer layer (SI =  $0.11$ ).

### 4 | DISCUSSION

#### 4.1 | Stomach content identification

A large number of lower beaks (77.6%) were badly damaged, making correct identification to species level extremely difficult. The results show that the calf had fed on a variety of 15 different species of oceanic squid, including enoploteuthids (13.3%) and several species of histioteuthids (19.4%). All species found in this study have been previously reported in the region, except the chiroteuthid *Asperoteuthis acanthoderma* (Lu, 1977) (Cephalopoda: Chiroteuthidae) (Cherel, 2021; Young & Roper, 2018) which until now had only been cited in the western Atlantic, in the Gulf of Mexico, and Florida. This result suggests the possibility of its presence on both sides of the subtropical Atlantic, extending its currently known distribution.

The diel vertical migrant species of the families Pyroteuthidae Pfeffer, 1912, Enoploteuthidae, and Onychoteuthidae Gray, 1847 represent the core of the cephalopod community in shallow (<200 m) nocturnal oceanic waters in the subtropical Atlantic (Ariza et al., 2016; Escáñez et al., 2022). Although they are also abundant in deeper layers, non-migratory phases or mesopelagic species are notable in this area (Escáñez et al., 2022). This is in line with the high percentage of enoploteuthids found in the stomach, which suggests a shallow foraging habitat of this young pilot whale. *Enoploteuthis leptura*, the largest species of the family, performs vertical migrations from mesopelagic to epipelagic depths, where they concentrated in the first 20 m during the night (Jereb & Roper, 2010). The ML range for two of the *E. leptura* encompasses larger animals than those cited in the literature (maximum ML of 92 mm; Jereb & Roper, 2010; see Table 1). This can be due to the fact that there are no specific allometric equations for this particular species, having used equations pertaining to *Enoploteuthis* spp. (Lu & Ickeringill, 2002) in the present study. In addition, most beaks were quite eroded, preventing the measurement of the total length of their wings, which might distort the results of the calculated indices and, therefore, the correct identification of the specimens.

*Enoploteuthis leptura* is a foraging species for several predators such as fishes (e.g. *Thunnus obesus* (Lowe, 1839), Teleostei: Scombridae, *Thunnus alalunga* (Bonnaterre, 1788), Teleostei: Scombridae, *Ruvettus pretiosus* Cocco, 1833, Teleostei: Gempylidae),

TABLE 1 Cephalopod species identified from the beaks found in the stomach contents of a calf pilot whale euthanized on the island of Tenerife.

Family/species	N	% N	ML (mm)	W (gr)	Geog. Distrib.	Reference	Depth (m)
<b>Chiroteuthidae</b>							
<i>Asperoteuthis acanthoderma</i> (Lu, 1977)	1	1.51	56.36	184.39	Indo-Pacific, Atl.	Clarke (1986) for <i>Mastigoteuthis</i> sp.	200–1100 <sup>a</sup>
<b>Cranchiidae</b>							
Cranchid not id.	1	1.51	63.11	4.24		Clarke (1986) for Cranchiinae spp.	>2000
<b>Enoploteuthidae</b>							
<i>Enoploteuthis leptura</i> (Leach, 1817)	22	33.3	31.71–223.83	1.80–429.23	Tropical-subtropical Atl.	Lu and Ickeringill (2002) for <i>Enoploteuthis</i> spp.	0–1620 <sup>b</sup>
<b>Histioteuthidae</b>							
Histioteuthid not id.	1	1.51	40.81	39.02		Clarke (1986) for Histioteuthidae.	0–4000
<i>Histioteuthis bonnellii</i> (Férussac, 1835)	8	12.1	26.34–159.13	11.27–27.64	Atl., SW Pacific, S Indian, Mediterranean	Pedà et al. (2022)	150–4000
<i>Histioteuthis corona</i> (N. A. Voss & G. L. Voss, 1962)	2	3.03	23.27–57.47	15.87–72.3	Tropical-subtropical Atl.	Clarke (1986) for Histioteuthidae.	100 to >1500
<i>Stigmatoteuthis arcturi</i> (G. C. Robson, 1948)	4	6.06	25.32–66.54	68.67–294.23	Tropical-subtropical Atl.	Clarke (1986; for ML >180 mm)	200–1000
<b>Lepidoteuthidae</b>							
<i>Lepidoteuthis grimaldi</i> Joubin, 1895	6	9.09	30.87–86.49	120.30–203.86	Cosmopolitan tropical-subtropical	Lu and Ickeringill (2002)	200–4000
<b>Bathyteuthidae</b>							
<i>Bathyteuthis abyssicola</i> Hoyle, 1885	1	1.51	32.63	3.09	Cosmopolitan	Clarke (1962)	700–2500
<b>Ommastrephidae</b>							
Ommastrephid not id.	5	7.57	0.94–6.60	187.96–1570.92		Clarke (1962)	0–2000
<i>Hyaloteuthis pelagica</i> (Bosc, 1802)	7	10.6	44.47–75.10	15.80–170.99	Subtropical Atl., Pacific	Clarke (1986)	15–800
<i>Ommastrephes caroli</i> (Furtado, 1887)	2	3.03	114.47–321.83	40.50–113.86	N Atl.	Agus et al. (2021)	0 to >1000 <sup>c</sup>
<i>Sthenoteuthis pteropus</i> (Steenstrup, 1855)	2	3.03	48.99–98.24	15.92–184.30	Tropical-subtropical Atl.	Guerra-Marrero (2017)	0–1200
<i>Todarodes sagittatus</i> (Lamarck, 1798)	2	3.03	27.58–51.15	31.29–157.01	NE and S. Atl.	Clarke (1962)	0 to >1000. In North Africa, 350–700m.

TABLE 1 (Continued)

Family/species	N	% N	ML (mm)	W (gr)	Geog. Distrib.	Reference	Depth (m)
<i>Todaropsis eblanae</i> (Ball, 1841)	2	3.03	55.99–66.16	6.58–7.50	E Atl.	Pérez-Gándaras (1983)	20–850
N TOTAL	66						

Abbreviations: Atl, Atlantic; Geog. Distrib., Geographic distribution of the species from Jereb and Roper (2010); ML, Estimated dorsal mantle length in mm; N %, percentages of beaks found for this species; N total, total number of beaks in the stomach; N, total number of beaks found for this species; W, Estimated weight in grams. References used for cephalopod ML regressions.

<sup>a</sup>In Judkins et al. (2009); Joseph et al. (2015).

<sup>b</sup>In Roper (1966).

<sup>c</sup>In Young and Vecchione (2018).

cetaceans (*Kogia* spp.), or seabirds (e.g. petrels) (Bello, 1999; Ravache et al., 2020; Staudinger et al., 2013; Vaske et al., 2012). Similarly, the majority of the histioteuthids (*H. bonnellii*, *H. corona*, *Stigmatoteuthis arcturi* G. C. Robson, 1948) (Cephalopoda: Histioteuthidae) were represented in the stomach content by small individuals (23–57 mm of ML). These sizes correspond to juvenile individuals of each species as ML maximum sizes of these species are between 50 and 330 mm. This young pilot whale fed mostly on small cephalopods (small species and juvenile of large species), probably due to the animal's small size and therefore its ability to catch prey more in line with its size, as well as its swimming and diving abilities. Histioteuthids are distributed throughout the water column showing ontogenetic migration to deeper waters during their growth (Judkins & Vecchione, 2020; Quetglas et al., 2010) and, predictability, the individuals registered here (23–57 mm of ML) were at shallowest depths.

Including our present results, a total of 22 cephalopod species and three genera belonging to 10 families of Teuthida have been identified so far in the stomach contents of four pilot whales from the Canary Islands. Among them, ommastrephids, histioteuthids, and cranchids contain the higher number of species (Table S1). The Canary Islands holds a rich cephalopod community with 85 species belonging to 31 families (Escáñez et al., 2021), which means that pilot whales' prey on about 25.9% of the total known cephalopod species in the Canaries. Compared to other regions where stomach contents of this cetacean species have been analyzed, Canary Islands present the highest cephalopods richness with only four specimens, followed by 27 pilot whales stranded in North Carolina (USA) of which a total of 11 cephalopods species were reported (Mintzer et al., 2008), and seven species identified in four pilot whales from California (USA) (Sinclair, 1992) (Table S1). This suggests that Canary Islands pilot whales are mainly teuthophagous predators, largely preying on a wide variety of squid species present in the area. However, predation on other types of prey, such as fish, cannot be completely ruled out for this species. The present study verifies that the diet of this young pilot whale was mainly based on squid.

Isotopic values of  $\delta^{15}\text{N}$  (11.55‰) of the young pilot whale were higher than expected values derived from the small oceanic squids found in its stomach (enoploteuthids and histioteuthids), but slightly lower than that of adult pilot whales from the region ( $12.2 \pm 0.5$ ; Escáñez, 2019) and from the Iberian Peninsula ( $11.9 \pm 0.3$ ‰, Monteiro et al., 2017). The  $\delta^{15}\text{N}$  values for small squids of the families Enoploteuthidae and Histioteuthidae, dominants of stomach contents, ranged generally between 5.7 and 10‰ in Atlantic temperate waters (Cherel et al., 2009). The isotopic values of the nursing young pilot whale analyzed here ( $\delta^{15}\text{N}$  11.55‰;  $\delta^{13}\text{C}$  -17.99‰) differ from adult individuals from Tenerife ( $\delta^{15}\text{N}$   $12.2 \pm 0.5$ ‰ and  $\delta^{13}\text{C}$  -15.8  $\pm$  0.02‰). The results suggested that the isotopic values of the still-suckling pilot whale are depleted in both,  $\delta^{15}\text{N}$  ( $\Delta^{15}\text{N} = -0.65$ ‰) and  $\delta^{13}\text{C}$  ( $\Delta^{13}\text{C} = -2.19$ ‰), compared with the mean values of adult individuals. Depletion in the  $\delta^{13}\text{C}$  in suckling calves (relative to the mother's tissues) is a common issue in several marine mammals that feed on lipid-rich milks, since lipids are also depleted in  $^{13}\text{C}$  during biochemical discrimination (Borrell et al., 2016; Cherel et al., 2015;

Fatty acid	Inner	Middle	Outer	Mean $\pm$ SD	SI
14:0	3.64	3.28	3.29	3.41 $\pm$ 0.21	-0.10
14:1n-5	0.55	0.68	0.59	0.61 $\pm$ 0.07	0.08
15:0	0.61	0.51	0.63	0.59 $\pm$ 0.06	0.04
16:0	14.17	11.17	11.81	12.38 $\pm$ 1.58	-0.18
16:1n-9	1.13	1.13	1.42	1.23 $\pm$ 0.17	0.23
16:1n-7	11.07	12.82	11.72	11.87 $\pm$ 0.88	0.06
16:2n-4	0.77	0.89	0.91	0.86 $\pm$ 0.08	0.17
17:0	0.67	0.56	0.60	0.61 $\pm$ 0.05	-0.10
17:1n-7	1.09	1.24	1.32	1.22 $\pm$ 0.12	0.20
18:0	3.55	2.95	3.23	3.24 $\pm$ 0.3	-0.09
18:1n-9	34.79	38.59	40.89	38.09 $\pm$ 3.08	0.16
18:1n-7	3.80	3.11	3.86	3.59 $\pm$ 0.42	0.02
18:1n-5	0.41	0.40	0.37	0.39 $\pm$ 0.02	-0.11
18:2n-6	0.99	1.11	1.11	1.07 $\pm$ 0.07	0.11
18:3n-4	0.31	0.33	0.34	0.33 $\pm$ 0.02	0.10
18:3n-3	0.33	0.38	0.34	0.35 $\pm$ 0.02	0.03
20:1n-11	1.38	1.38	1.55	1.44 $\pm$ 0.1	0.12
20:1n-9	5.57	5.23	5.19	5.33 $\pm$ 0.21	-0.07
20:1n-7	0.35	0.30	0.29	0.31 $\pm$ 0.04	-0.20
20:2n-6	nd	0.44	0.40	0.28 $\pm$ 0.24	2.00
20:4n-6	1.15	1.29	1.11	1.18 $\pm$ 0.10	-0.04
20:3n-3	0.40	0.35	0.32	0.36 $\pm$ 0.04	-0.21
20:4n-3	0.35	0.29	0.29	0.31 $\pm$ 0.04	-0.20
20:5n-3	1.87	1.81	1.25	1.64 $\pm$ 0.34	-0.40
22:1n-11	0.70	0.57	0.63	0.64 $\pm$ 0.06	-0.10
22:1n-9	0.34	0.27	0.31	0.31 $\pm$ 0.04	-0.10
22:2n-6	0.46	nd	nd	0.15 $\pm$ 0.27	-2.00
22:4n-6	0.34	0.31	0.27	0.31 $\pm$ 0.04	-0.24
22:5n-6	0.46	0.31	0.36	0.38 $\pm$ 0.08	-0.26
22:5n-3	1.70	1.57	1.16	1.48 $\pm$ 0.28	-0.38
22:6n-3	5.63	4.60	3.20	4.47 $\pm$ 1.22	-0.55
UK	1.05	1.85	1.23		
$\Sigma$ SFA	22.64	18.48	19.57	20.23 $\pm$ 2.16	
$\Sigma$ MUFA	61.18	66.01	68.14	65.11 $\pm$ 3.57	
$\Sigma$ PUFA	14.78	13.66	11.06	13.17 $\pm$ 1.90	
$\Sigma$ n-6PUFA	3.41	3.46	3.24	3.37 $\pm$ 0.11	
$\Sigma$ n-3PUFA	10.29	8.99	6.56	8.61 $\pm$ 1.89	
n-3/n-6	3.02	2.60	2.03	2.55 $\pm$ 0.49	
$\Sigma$ n-3LC-PUFA	9.96	8.61	6.22	8.26 $\pm$ 1.89	

Abbreviations: nd, not detected; SI, stratification index; UK, Unknown FA;  $\Sigma$ MUFA, total monounsaturated FA;  $\Sigma\omega$ -3 LC-PUFA, total omega-3 long-chain PUFA;  $\Sigma\omega$ -3, total omega-3 PUFA;  $\Sigma\omega$ -6, total omega-6 PUFA;  $\Sigma$ PUFA, total polyunsaturated FA;  $\Sigma$ SFA, Total saturated FA.

DeNiro & Epstein, 1977). In this case, pilot whales' (*Globicephala* spp.) milk has around 15–31% of lipid (Lockyer, 2007). On the other hand,  $\delta^{15}\text{N}$  values of nursing young that only feed milk should be one trophic level higher than mothers (Cherel et al., 2015). Unfortunately, we lack the information about  $\delta^{15}\text{N}$  values from the mother in our

study. In any case, the isotopic values of the nursing young individual are the result of the isotopic mixing signatures of breast milk, food intake, and biochemical discrimination factors. It is to be expected that the influence from the maternal milk isotopic signature decreases over time with respect to isotopic influence of the food

TABLE 2 Fatty acid (FA) profiles of the inner, middle, and outer blubber layers (% of total FA) from a lactating *Globicephala macrorhynchus* from Tenerife.



intake (mainly squids). Oftedal (1997) estimated ages of first ingestions of major solids for pilot whales in 6–12 months, while milk has been found in the stomach up to three years old individuals. These data emplace our nursing young animal with an estimated age of 6–12 months in its very early first food intakes.

The FA profile of the external blubber of this animal did not differ greatly from FA profiles previously reported in adult individuals from the same area during 2015 and 2017, where the same dominant FA (18:1n-9, 16:0, 16:1n-7, 20:1n-9, and 22:6n-3) were present in similar proportions, except 22:6n-3, which was three times higher in the calf (Table 2) (Gil, 2018; work in preparation). Totals of MUFA and SFA were also found in similar amounts, but total PUFA was slightly higher in this calf (Table 2), possibly related to the importance of these FA during neurological, immune, and growth development of young animals (Parzanini et al., 2018). The great proportion of MUFA, primarily 18:1n-9, in this species is probably related to its endogenous biosynthesis through the introduction of a first unsaturation into stearic acid (18:0) by means of a  $\Delta 9$  desaturase activity (Castro et al., 2016; Hooker et al., 2001). Similarly, other major FA such as 16:1n-7 has been previously reported to be endogenously biosynthesized from 16:0 in other aquatic mammals (Kirsch et al., 2000). MUFA have a key function in thermic isolation due to their lower melting point and their capacity for improving the cellular membrane fluidity in cold environments, being more abundant in the external layers in adults and juveniles. Furthermore, MUFA is an adaptive response to the pressure, an important adaptation for deep diving cetaceans as pilot-whales (Burgess et al., 2018; Parrish, 2013). In addition, they are used as energy storage in many cetaceans. Similarly, short and medium chain SFA can be endogenously biosynthesized, also presenting thermic isolation functions, serving as energy storage and, as previously stated, as substrate to produce MUFA (Guerrero & Rogers, 2017). Notable variations in the percentage of FA in the inner and outer layers indicate a selective FA deposition during the development and growth of this young animal. Thus, all n-3 and n-6 LC-PUFA that are typically ingested through the diet, are more prevalent in the inner blubber layer suggesting that this layer is more metabolically active in terms of fat storage and lipid deposition from the diet. By contrast, biosynthesized  $\leq C20$  MUFA such as 14:1n-5, 16:1n-7, 18:1n-9 and 20:1n-11, were more abundant in the outer blubber layer.

Little is known about the diet of this species and even less about the ontogenetic changes during its early life stages. Opportunities such as the one described in this work are extremely rare. Our present results allow deepening our knowledge on the life cycle and bioecology of deep-diving cetaceans, especially during their first life stages. This information is a valuable contribution to conservation policies. However, future studies, based on trophic markers such as isotopes and FA of both pilot whales and their potential preys in the area, together with the analysis of the stomach contents of a larger number of specimens, will help to better understand the needs and use of food resources on which depend the populations present in the oceanic islands of this region.

## AUTHOR CONTRIBUTIONS

**Amanda Luna:** Conceptualization (equal); data curation (equal); formal analysis (equal); investigation (equal); methodology (equal); resources (equal); writing – original draft (equal); writing – review and editing (equal). **Alejandro Escáñez:** Conceptualization (equal); investigation (equal); methodology (equal); writing – original draft (equal); writing – review and editing (equal). **Jacobo Marrero:** Methodology (equal); writing – original draft (equal); writing – review and editing (equal). **Eva Íñiguez:** Formal analysis (equal); writing – original draft (equal). **José A. Pérez:** Methodology (equal); resources (equal); supervision (equal). **Pilar Sánchez:** Data curation (equal); methodology (equal); resources (equal); supervision (equal); writing – review and editing (equal).

## ACKNOWLEDGMENTS

The authors would like to thank Francis Pérez for his help in obtaining underwater photographs and videos of the short-pilot whale, as well as his help and support in capturing it. We would also like to thank the support of the technicians of the Centro de Recuperación de Fauna Silvestre "La Tahonilla", as well as Santiago Mayans for his quick response and coordination of the veterinary team. In addition, the authors would like to thank the veterinary team of the IUSA-ULPGC for providing the samples for study and the BIOECOMAC research group of the University of La Laguna for their support during the campaign. We are very grateful for the cession of the reference collection of cephalopod beaks AFOC (ICM-CSIC, Barcelona; Sánchez-Márquez et al., 2023). Alejandro Escáñez has been funded by the Action funded by the Ministry of Universities under the application 33.50.460A.752 and by the European Union NextGeneration EU/PRTR through a Margarita Salas contract of the University of Vigo, as well as by the Ministry of Science and Innovation project DeepCom CTM2017-88686-P. Funding for open access charge: Universidade de Vigo/CISUG. The authors would like to thank the reviewers for their fine work in revising and improving the manuscript.

## CONFLICT OF INTEREST STATEMENT

The authors declare no conflicts of interest.

## DATA AVAILABILITY STATEMENT

Data available on request from the authors.

## ORCID

Amanda Luna  <https://orcid.org/0000-0002-8006-1035>

## REFERENCES

- Aguilar de Soto, N., Johnson, M. P., Madsen, P. T., Daz, F., Domínguez, I., Brito, A., & Tyack, P. (2008). Cheetahs of the deep sea: Deep foraging sprints in short-finned pilot whales off Tenerife (Canary Islands). *Journal of Animal Ecology*, 77, 936–947. <https://doi.org/10.1111/j.1365-2656.2008.01393.x>
- Aguilar de Soto, N., Visser, F., Tyack, P. L., Alcazar, J., Ruxton, G., Arranz, P., Madsen, P. T., & Johnson, M. (2020). Fear of killer whales drives extreme synchrony in deep diving beaked whales.

- Scientific Reports, 10(1), 13. <https://doi.org/10.1038/s41598-019-55911-3>
- Agus, B., Carugati, L., Bellodi, A., Cannas, R., Cau, A., Cera, J., Coluccia, E., Melis, R., Ruiu, S., & Cuccu, D. (2021). Molecular and biological analysis on *Ommastrephes caroli* findings in the central western Mediterranean Sea (Sardinian waters) including first age investigation using eye lenses and beaks. *Frontiers in Marine Science*, 8, 683856. <https://doi.org/10.3389/fmars.2021.683856>
- Alves, F., Quérouil, S., Dinis, A., Nicolau, C., Ribeiro, C., Freitas, L., Kaufmann, M., & Fortuna, C. (2013). Population structure of short-finned pilot whales in the oceanic archipelago of Madeira based on photo-identification and genetic analyses: Implications for conservation. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 23(5), 758–776. <https://doi.org/10.1002/aqc.2332>
- Ariza, A., Landeira, J. M., Escáñez, A., Wienerroither, R., de Soto, N. A., Røstad, A., Kaartvedt, S., & Hernández-León, S. (2016). Vertical distribution, composition and migratory patterns of acoustic scattering layers in the Canary Islands. *Journal of Marine Systems*, 157, 82–91. <https://doi.org/10.1016/j.jmarsys.2016.01.004>
- Aurioles-Gamboa, D., Rodríguez-Pérez, M. Y., Sánchez-Velasco, L., & Lavin, M. F. (2013). Habitat, trophic level, and residence of marine mammals in the Gulf of California assessed by stable isotope analysis. *Marine Ecology Progress Series*, 488, 275–290. <https://doi.org/10.3354/meps10369>
- Bello, G. (1999). Cephalopods in the diet of albacore, *Thunnus alalunga*, from the Adriatic Sea. *Journal of Molluscan Studies*, 65(2), 233–240. <https://doi.org/10.1093/mollus/65.2.233>
- Best, P. B., Canham, P. A. S., & Macleod, N. (1984). Patterns of reproduction in sperm whales, *Physeter macrocephalus*. *Report of the International Whaling Commission*, 6, 51–79.
- Blanco, C., Raduán, Á., & Raga, J. A. (2006). Diet of Risso's dolphin (*G. griseus*) in the Western Mediterranean Sea. *Scientia Marina*, 70(3), 407–411. <https://doi.org/10.3989/scimar.2006.70n3407>
- Borrell, A., Gómez-Campos, E., & Aguilar, A. (2016). Influence of reproduction on stable-isotope ratios: Nitrogen and carbon isotope discrimination between mothers, fetuses, and milk in the fin whale, a capital breeder. *Physiological and Biochemical Zoology*, 89(1), 41–50. <https://doi.org/10.1086/684632>
- Bowen, W. D., & Iverson, S. J. (2013). Methods of estimating marine mammal diets: A review of validation experiments and sources of bias and uncertainty. *Marine Mammal Science*, 29(4), 719–754. <https://doi.org/10.1111/j.1748-7692.2012.00604.x>
- Burgess, K. B., Guerrero, M., Marshall, A. D., Richardson, A. J., Bennett, M. B., & Couturier, L. I. (2018). Novel signature fatty acid profile of the giant manta ray suggests reliance on an uncharacterised mesopelagic food source low in polyunsaturated fatty acids. *PLoS One*, 13(1), e0186464. <https://doi.org/10.1371/journal.pone.0186464>
- Bustamante, P., Garrigue, C., Breau, L., Caurant, F., Dabin, W., Greaves, J., & Dodemont, R. (2003). Trace elements in two odontocete species (*Kogia breviceps* and *Globicephala macrorhynchus*) stranded in New Caledonia (South Pacific). *Environmental Pollution*, 124(2), 263–271. [https://doi.org/10.1016/S0269-7491\(02\)00480-3](https://doi.org/10.1016/S0269-7491(02)00480-3)
- Castro, L. F. C., Tocher, D. R., & Monroig, O. (2016). Long-chain polyunsaturated fatty acid biosynthesis in chordates: Insights into the evolution of fads and Elovl gene repertoire. *Progress in Lipid Research*, 62, 25–40. <https://doi.org/10.1016/j.plipres.2016.01.001>
- Cherel, Y. (2021). ?Mastigoteuthis B Clarke, 1980, is a junior synonym of *Asperoteuthis acanthoderma* (Lu, 1977) (Cephalopoda, Oegopsida, Chiroteuthidae), a rare cosmopolitan deep-sea squid. *Marine Biodiversity*, 51(1), 1–8. <https://doi.org/10.1007/s12526-020-01155-8>
- Cherel, Y., Hobson, K. A., & Guinet, C. (2015). Milk isotopic values demonstrate that nursing fur seal pups are a full trophic level higher than their mothers. *Rapid Communications in Mass Spectrometry*, 29(16), 1485–1490. <https://doi.org/10.1002/rcm.7243>
- Cherel, Y., Ridoux, V., Spitz, J., & Richard, P. (2009). Stable isotopes document the trophic structure of a deep-sea cephalopod assemblage including giant octopod and giant squid. *Biology Letters*, 5(3), 364–367. <https://doi.org/10.1098/rsbl.2009.0024>
- Christie, W. W. (1982). *Lipid analysis*, 207 (p. 337). Pergamon Press.
- Christie, W. W., & Han, X. (2010). *Lipid analysis: Isolation, separation, identification and lipidomic analysis*. Oily Press, An Imprint of PJ Barnes & Associates.
- Clarke, M. R. (1962). Significance of cephalopod beaks. *Nature*, 193, 560–561. <https://doi.org/10.1038/193560a0>
- Clarke, M. R. (1986). *A handbook for the identification of cephalopod beaks* (p. 273). Clarendon Press.
- Copeland, A. M., Au, W. W., & Polovina, J. (2019). Influences of temporal changes in pelagic scattering layers on short-finned pilot whales behavior. *Oceanography & Fisheries Open Access Journal*, 9, 555758. <https://doi.org/10.19080/OFOAJ.2019.09.555758>
- DeNiro, M. J., & Epstein, S. (1977). Mechanism of carbon isotope fractionation associated with lipid synthesis. *Science*, 197(4300), 261–263. <https://doi.org/10.1126/science.327543>
- Dixon, J. M., Frigo, L. F., & Moyle, R. L. C. (1994). New information on the southern bottlenose whale, *Hyperoodon planifrons* (Cetacea: Ziphiidae), from a recent stranding in Victoria, Australia. *Australian Mammalogy*, 17, 85–95. <https://doi.org/10.1071/AM94009>
- Dunn, C., Claridge, D., Durban, J., Shaffer, J., Moretti, D., Tyack, P., & Rendell, L. (2017). Insights into Blainville's beaked whale (*Mesoplodon densirostris*) echolocation ontogeny from recordings of mother-calf pairs. *Marine Mammal Science*, 33(1), 356–364. <https://doi.org/10.1111/mms.12351>
- Escáñez, A. (2019). *Diversidad y ecología de los cefalópodos oceánicos de la región macaronésica* (PhD Dissertation). Universidad de Vigo.
- Escáñez, A., Guerra, Á., Riera, R., Ariza, A., González, Á. F., & de Soto, N. A. (2022). New contribution to the knowledge of the mesopelagic cephalopod community off the western Canary Islands slope. *Regional Studies in Marine Science*, 55, 102572. <https://doi.org/10.1016/j.rsma.2022.102572>
- Escáñez, A., Guerra, Á., Riera, R., & Rocha, F. J. (2021). Revised species records reveal the Canary Islands as a cephalopod biodiversity hotspot. *Regional Studies in Marine Science*, 41, 101541. <https://doi.org/10.1016/j.rsma.2020.101541>
- European Habitats Directive 92/43/EEC Council Directive 92/43/EEC of 21 May 1992 on the conservation of natural habitats and of wild fauna and flora. Document 31992L0043.
- Fernández, R., Santos, M. B., Carrillo, M., Tejedor, M., & Pierce, G. J. (2009). Stomach contents of cetaceans stranded in the Canary Islands 1996–2006. *Journal of the Marine Biological Association of the United Kingdom*, 89(5), 873–883. <https://doi.org/10.1017/S0025315409000290>
- Folch, J., Lees, M., & Sloane Stanley, G. H. (1957). A simple method for the isolation and purification of total lipids from animal tissues. *Journal of Biological Chemistry*, 226(1), 497–509. [https://doi.org/10.1016/S0021-9258\(18\)64849-5](https://doi.org/10.1016/S0021-9258(18)64849-5)
- Galindo, A., Reis, D. B., Rodríguez, I., Pérez, J. A., Abdul-Jalbar, B., Zárate, R., Nunes, N., Pinheiro de Carvalho, M. A. A., Acosta, N. G., & Rodríguez, C. (2022). Lipid characterization of 14 macroalgal species from Madeira archipelago: Implications for animal and human nutrition. *Botanica Marina*, 65(1), 51–67. <https://doi.org/10.1515/bot-2021-0024>
- Gil, P. (2018). *Dieta del calderón tropical (Globicephala macrorhynchus) en la costa de Tenerife basada en el estudio de ácidos grasos (QFASA)* (Master thesis). Universidad de La Laguna. <http://riull.ull.es/xmlui/handle/915/11598>
- Guerra-Marrero, A. M. (2017). *Contribution to the bioecological knowledge of the orange-back squid Sthenoteuthis pteropus (family Ommastrephidae Steenstrup, 1857) for Madeira and canary waters* (Master's thesis). Universidad de Las Palmas de Gran Canaria.

- Guerrero, A. I., & Rogers, T. L. (2017). Blubber fatty acid composition and stratification in the crabeater seal, *Lobodon carcinophaga*. *Journal of Experimental Marine Biology and Ecology*, 491, 51–57. <https://doi.org/10.1016/j.jembe.2017.03.004>
- Heimlich-Boran, J. R. (1993). *Social organisation of the short-finned pilot whale, Globicephala macrorhynchus, with special reference to the comparative social ecology of delphinids* (PhD Dissertation). University of Cambridge, Department of Zoology, United Kingdom.
- Hernández-García, V., & Martín, V. (1994). Stomach contents of two short-finned pilot whales (*Globicephala macrorhynchus* Gray, 1846) (Cetacea, Delphinidae) off the Canary Islands: A preliminary note. ICES Document C.M.1994/N:16: 9 pp.
- Hooker, S. K., Iverson, S. J., Ostrom, P., & Smith, S. C. (2001). Diet of northern bottlenose whales inferred from fatty-acid and stable-isotope analyses of biopsy samples. *Canadian Journal of Zoology*, 79(8), 1442–1454. <https://doi.org/10.1139/z01-096>
- Hui, C. A. (1985). Undersea topography and the comparative distributions of two pelagic cetaceans. *Fishery Bulletin*, 83(3), 472–475.
- Jereb, P., & Roper, C. F. (2010). *Cephalopods of the world—an annotated and illustrated catalogue of cephalopod species known to date: Myopsid and oegopsid squids, 2*. FAO species catalogue for fishery purposes, 4 (2) (p. 649). FAO.
- Johnson, M. P., & Tyack, P. L. (2003). A digital acoustic recording tag for measuring the response of wild marine mammals to sound. *IEEE Journal of Oceanic Engineering*, 28(1), 3–12. <https://doi.org/10.1109/JOE.2002.808212>
- Joseph, R., Ravi, R., & Remesan, M. P. (2015). First record of *Asperoteuthis acanthoderma* (Lu, 1977) (Cephalopoda: Oegopsida: Chiroteuthidae), from the Arabian Sea. *International Journal of Scientific and Research Publications*, 5(9), 1–5.
- Judkins, H., Ingrao, D. A., & Roper, C. F. (2009). First records of *Asperoteuthis acanthoderma* (Lu, 1977) (Cephalopoda: Oegopsida: Chiroteuthidae), from the North Atlantic Ocean, straits of Florida. *Proceedings of the Biological Society of Washington*, 122(2), 162–170. <https://doi.org/10.2988/08-30.1>
- Judkins, H., & Vecchione, M. (2020). Vertical distribution patterns of cephalopods in the northern Gulf of Mexico. *Frontiers in Marine Science*, 7, 47. <https://doi.org/10.3389/fmars.2020.00047>
- Kasuya, T., & Marsh, H. (1984). Life history and reproductive biology of the short-finned pilot whale, *Globicephala macrorhynchus*, off the Pacific coast of Japan. *Reports of the International Whale Commission*, 6, 239–310.
- Kirsch, P. E., Iverson, S. J., & Bowen, W. D. (2000). Effect of diet on body composition and blubber fatty acids in captive harp seals (*Phoca groenlandica*). *Physiological and Biochemical Zoology*, 73, 45–59. <https://doi.org/10.1086/316723>
- Kiszka, J., Oremus, M., Richard, P., Poole, M., & Ridoux, V. (2010). The use of stable isotope analyses from skin biopsy samples to assess trophic relationships of sympatric delphinids off Moorea (French Polynesia). *Journal of Experimental Marine Biology and Ecology*, 395(1–2), 48–54. <https://doi.org/10.1016/j.jembe.2010.08.010>
- Kuiken, T., & García-Hartmann, M. (1991). Proceedings of dissection techniques and tissue sampling. In *Proceedings of the First ECS Workshop on Cetacean Pathology*. Leiden (1991), 13–4.
- Lockyer, C. (2007). All creatures great and smaller: A study in cetacean life history energetics. *Journal of the Marine Biological Association of the United Kingdom*, 87(4), 1035–1045. <https://doi.org/10.1017/S0025315407054720>
- Lu, C. C., & Ickeringill, R. (2002). Cephalopod beak identification and bio-mass estimation techniques: Tools for dietary studies of southern Australian finfishes. *Museum Victoria Science Reports*, 6, 1–65.
- Luna, A., Sánchez, P., Chicote, C., & Gazo, M. (2022). Cephalopods in the diet of Risso's dolphin (*Grampus griseus*) from the Mediterranean Sea: A review. *Marine Mammal Science*, 38(2), 725–741. <https://doi.org/10.1111/mms.12869>
- Méndez-Fernández, P., Bustamante, P., Bode, A., Chouvelon, T., Ferreira, M., López, A., Pierce, G. J., Santos, M. B., Spitz, J., Vingada, J. V., & Caurant, F. (2012). Foraging ecology of five toothed whale species in the Northwest Iberian Peninsula, inferred using carbon and nitrogen isotope ratios. *Journal of Experimental Marine Biology and Ecology*, 413, 150–158. <https://doi.org/10.1016/j.jembe.2011.12.007>
- Mintzer, V. J., Gannon, D. P., Barros, N. B., & Read, A. J. (2008). Stomach contents of mass-stranded short-finned pilot whales (*Globicephala macrorhynchus*) from North Carolina. *Marine Mammal Science*, 24(2), 290–302. <https://doi.org/10.1111/j.1748-7692.2008.00189.x>
- Monteiro, S. S., Caurant, F., López, A., Cedeira, J., Ferreira, M., Vingada, J. V., Eira, C., & Méndez-Fernández, P. (2017). Sympatric *Globicephala* species: Feeding ecology and contamination status based on stable isotopes and trace elements. *Marine Ecology Progress Series*, 563, 233–247. <https://doi.org/10.3354/meps11965>
- Oftedal, O. T. (1997). Lactation in whales and dolphins: Evidence of divergence between baleen-and toothed-species. *Journal of Mammary Gland Biology and Neoplasia*, 2, 205–230. <https://doi.org/10.1023/A:1026328203526>
- Olsen, E., & Grahl-Nielsen, O. (2003). Blubber fatty acids of minke whales: Stratification, population identification and relation to diet. *Marine Biology*, 142, 13–24. <https://doi.org/10.1007/s00227-002-0934-2>
- Overholtz, W. J., & Waring, G. T. (1991). Diet composition of pilot whales *Globicephala* sp. and common dolphins *Delphinus delphis* in the mid-Atlantic bight during spring 1989. *Fishery Bulletin*, 89(4), 723–728.
- Owen, K., Andrews, R. D., Baird, R. W., Schorr, G. S., & Webster, D. L. (2019). Lunar cycles influence the diving behavior and habitat use of short-finned pilot whales around the main Hawaiian islands. *Marine Ecology Progress Series*, 629, 193–206. <https://doi.org/10.3354/meps13123>
- Öztürk, B., Salman, A., Öztürk, A., & Tonay, A. (2007). Cephalopod remains in the diet of striped dolphins (*Stenella coeruleoalba*) and Risso's dolphins (*Grampus griseus*) in the eastern Mediterranean Sea. *Vie et Milieu, Life and Environment*, 57(1/2), 53–59.
- Parrish, C. C. (2013). Lipids in marine ecosystems. *International Scholarly Research Notices*, 2013, 604045. <https://doi.org/10.5402/2013/604045>
- Parzanini, C., Parrish, C. C., Hamel, J. F., & Mercier, A. (2018). Functional diversity and nutritional content in a deep-sea faunal assemblage through total lipid, lipid class, and fatty acid analyses. *PLoS One*, 13(11), e0207395. <https://doi.org/10.1371/journal.pone.0207395>
- Pedà, C., Battaglia, P., Romero, T., Stipa, M. J., Longo, F., Malara, D., Consoli, P., & Andaloro, F. (2022). *Photographic atlas of cephalopod beaks from the Mediterranean Sea* (p. 107). Etabeta.
- Pérez-Gándaras, G. (1983). *Estudio de los cefalópodos ibéricos: Sistemática y Bionomía mediante el estudio morfológico comparado de sus mandíbulas* (PhD Dissertation). Universidad Complutense de Madrid.
- Pimentel, A., Piñero, M., Crespo, A., & Marrero, J. (2023). *Catálogo de fotoidentificación del calderón tropical (Globicephala macrorhynchus)*. Asociación Tonina. [https://asociaciontonina.com/wp-content/uploads/2020/03/Catálogo\\_GM\\_10\\_01\\_2023.pdf](https://asociaciontonina.com/wp-content/uploads/2020/03/Catálogo_GM_10_01_2023.pdf)
- Quetglas, A., de Mesa, A., Ordines, F., & Grau, A. (2010). Life history of the deep-sea cephalopod family Histioteuthidae in the western Mediterranean. *Deep Sea Research Part I: Oceanographic Research Papers*, 57(8), 999–1008. <https://doi.org/10.1016/j.dsr.2010.04.008>
- Ravache, A., Bourgeois, K., Weimerskirch, H., Pagenaud, A., De Grissac, S., Miller, M., Dromzéé, S., Lorrain, A., Allain, V., Bustamante, P., Bylemans, J., Gleeson, D., Letourneau, Y., & Vidal, E. (2020). Behavioral and trophic segregations help the Tahiti petrel to cope with the abundance of wedge-tailed shearwater when foraging in oligotrophic tropical waters. *Scientific Reports*, 10(1), 1–18. <https://doi.org/10.1038/s41598-020-72206-0>

- Riccialdelli, L., Newsome, S. D., Dellabianca, N. A., Bastida, R., Fogel, M. L., & Goodall, R. N. P. (2013). Ontogenetic diet shift in Commerson's dolphin (*Cephalorhynchus commersonii commersonii*) off Tierra del Fuego. *Polar Biology*, 36, 617–627. <https://doi.org/10.1007/s00300-013-1289-5>
- Rodhouse, P. G., Prince, P. A., Clarke, M. R., & Murray, A. W. A. (1990). Cephalopod prey of the grey-headed albatross *Diomedea chrysostoma*. *Marine Biology*, 104, 353–362. <https://doi.org/10.1007/BF01314337>
- Roper, C. F. (1966). A study of the genus *Enoploteuthis* (Cephalopoda: Oegopsida) in the Atlantic Ocean with a redescription of the type species, *E. leptura* (Leach, 1817). *Dana-Report*, 66, 45.
- Sánchez-Márquez, A., Navarro, J., Kaliontzopoulou, A., Farré, M., Taite, M., Escolar, O., Villanueva, R., Allcock, A. L., & Fernández-Álvarez, F. Á. (2023). Unravelling the phylogenetic and ecological drivers of beak shape variability in cephalopods. *Reviews in Fish Biology and Fisheries*, 33, 221–239. <https://doi.org/10.1007/s11160-022-09744-5>
- Santos, M. B., Berrow, S. D., & Pierce, G. J. (2006). Stomach contents of a sperm whale calf *Physeter macrocephalus* L. found dead in Co Clare, Ireland. *The Irish Naturalists' Journal*, 28(7), 272–275.
- Seagars, D. J., & Henderson, J. R. (1985). Cephalopod remains from the stomach of a short-finned pilot whale collected near Santa Catalina Island, California. *Journal of Mammalogy*, 66, 777–779. <https://doi.org/10.2307/1380806>
- Servidio, A., Pérez-Gil, E., Pérez-Gil, M., Cañadas, A., Hammond, P. S., & Martín, V. (2019). Site fidelity and movement patterns of short-finned pilot whales within the Canary Islands: Evidence for resident and transient populations. *Aquatic Conservation: Marine and Freshwater Ecosystems*, 29, 227–241. <https://doi.org/10.1002/aqc.3135>
- Shane, S. H., & McSweeney, D. (1990). Using photo-identification to study pilot whale social organization. *Report of the International Whaling Commission*, 12, 259–263.
- Shearer, J. M., Jensen, F. H., Quick, N. J., Friedlaender, A., Southall, B., Nowacek, D. P., Bowers, M., Foley, H. J., Swaim, Z. T., Waples, D. M., & Read, A. J. (2022). Short-finned pilot whales exhibit behavioral plasticity in foraging strategies mediated by their environment. *Marine Ecology Progress Series*, 695, 1–14. <https://doi.org/10.3354/meps14132>
- Sinclair, E. (1992). Stomach contents of four short-finned pilot whales (*Globicephala macrorhynchus*) from the southern California bight. *Marine Mammal Science*, 8(1), 76–81. <https://doi.org/10.1111/j.1748-7692.1992.tb00127.x>
- Staudinger, M. D., Juanes, F., Salmon, B., & Teffer, A. K. (2013). The distribution, diversity, and importance of cephalopods in top predator diets from offshore habitats of the Northwest Atlantic Ocean. *Deep Sea Research Part II: Topical Studies in Oceanography*, 95, 182–192. <https://doi.org/10.1016/j.dsr2.2012.06.004>
- Tønnesen, P., Gero, S., Ladegaard, M., Johnson, M., & Madsen, P. T. (2018). First-year sperm whale calves echolocate and perform long, deep dives. *Behavioral Ecology and Sociobiology*, 72, 1–15. <https://doi.org/10.1007/s00265-018-2570-y>
- Torres, C., Londoño, J., Hincapié, S., & Ruales, C. (2013). Extracción y caracterización de aceite de pescado derivado de subproductos de trucha arco iris *Oncorhynchus mykiss*. *Chilean Journal of Agricultural and Animal Sciences*, 2(2), 34–41.
- Vaske, T., Jr., Travassos, P. E., Hazin, F. H. V., Tolotti, M. T., & Barbosa, T. M. (2012). Forage fauna in the diet of bigeye tuna (*Thunnus obesus*) in the western tropical Atlantic Ocean. *Brazilian Journal of Oceanography*, 60, 89–97. <https://doi.org/10.1590/S1679-87592012000100009>
- Walker, W. W., Mead, J. G., & Brownell, R. L., Jr. (2002). Diets of Bairds beaked whales, *Berardius bairdii*, in the southern Sea of Okhotsk and off the Pacific coast of Honshu, Japan. *Marine Mammal Science*, 18(4), 902–919. <https://doi.org/10.1111/j.1748-7692.2002.tb01081.x>
- Xavier, J., & Cherel, Y. (2021). *Cephalopod beak guide for the Southern Ocean: An update on taxonomy (revised edition)* (p. 129). British Antarctic Survey.
- Xavier, J. C., Golikov, A. V., Queirós, J. P., Perales-Raya, C., Rosas-Luis, R., Abreu, J., Bello, G., Bustamante, P., Capaz, J. C., Dimkovikj, V. H., González, A. F., Guimaro, H., Guerra-Marrero, A., Gomes-Pereira, J. N., Hernández-Urcera, J., Kubodera, T., Laptikhovskiy, V., Lefkaditou, E., Lishchenko, F., ... Cherel, Y. (2022). The significance of cephalopod beaks as a research tool: An update. *Frontiers in Physiology*, 13, 1038064. <https://doi.org/10.3389/fphys.2022.1038064>
- Young, R. E., & Roper, C. F. E. (2018). *Asperoteuthis acanthoderma* (Lu, 1977). Version 31 October 2018 (under construction). [http://tolweb.org/Asperoteuthis\\_acanthoderma/19466/2018.10.31](http://tolweb.org/Asperoteuthis_acanthoderma/19466/2018.10.31)
- Young, R. E., & Vecchione, M. (2018). *Ommastrephes Orbigny 1834. Ommastrephes bartramii* (Lesueur 1821). Red squid. Version 20 February 2018 (under construction). [http://tolweb.org/Ommas\\_trephes\\_bartramii/19947/2018.02.20](http://tolweb.org/Ommas_trephes_bartramii/19947/2018.02.20)

## SUPPORTING INFORMATION

Additional supporting information can be found online in the Supporting Information section at the end of this article.

**How to cite this article:** Luna, A., Escáñez, A., Marrero, J., Íñiguez, E., Pérez, J. A., & Sánchez, P. (2024). Early prey intake of a short-finned pilot whale (*Globicephala macrorhynchus* Gray, 1846, Cetacea: Delphinidae) in the Canary Islands. *Ecology and Evolution*, 14, e11139. <https://doi.org/10.1002/ece3.11139>