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SHORT COMMUNICATION

Parasitoids from Azores (Hymenoptera: Encyrtidae, Pteromalidae, Braconidae): potential use in integrated pest management against *Ceratitis capitata* (Diptera: Tephritidae)

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Two hymenopterous parasitoids, *Halticoptera patellana* and *Toxeumorpha nigricola*, were recovered from Mediterranean fruit fly pupae on São Miguel Island, Azores. Both are new records for the Azores, and as Medfly parasitoids. Field-collected *Tachinae-phagus zealandicus* was maintained on Medfly for 10 generations, but if it has potential as a biocontrol agent remains an open question.

Keywords: Ceratitis capitata; native parasitoids; Braconidae; Encyrtidae; Pteromalidae; new host

The Mediterranean fruit fly, *Ceratitis capitata* (Weidemann) originated in Africa (White and Elson-Harris 1992) but is now one of the most important fruit pests on a global scale (Liquido, Shinoda, and Cunningham 1991). It was first reported in the Azores (Portugal) in 1829 (MacLeay 1829) and is now a pest of many fruit crops throughout the archipelago (Medeiros 2004). Pesticides are currently the principal means of control but our objective is to develop a more ecologically acceptable integrated management program, including biological control. Headrick and Goeden (1996) consider that the parasitic Hymenoptera are the most promising candidates for Medfly biological control, and a number of candidate species have been identified. Nothing is known about Medfly parasitoids in the Azores so we decided to carry out a survey to see if there were possible candidates for innundative releases before considering the introduction of exotic species.

A survey, using the approach of Falcó et al. (2006), was carried on the Island of São Miguel from September to December 2005, and from August to December 2006. We selected different orchards that harboured Medfly populations, and included the most important host plants of the Medfly in the Azores (Table 1). In each orchard, fruit with signs of Medfly attack were selected and placed beneath the host tree on a plastic tray $(32 \times 47 \times 9 \text{ cm})$ containing 1 cm of sand. The entire surface of each tray was covered with fruits of one species. After 2 weeks, the traps were replaced and the old ones brought back to the laboratory where all larvae and pupae found in the sand were counted, and placed in

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Local	Host plant	Traps per orchard	Survey period
2005			
Rabo de Peixe	Fig (Ficus carica, Moraceae)	2	September
	Cattley guava (<i>Psidium littorale</i> , Myrtaceae)	3	September–November
São Gonçalo	Feijoa (Feijoa sellowiana, Myrtaceae)	3	October–December
2006			
Rabo de Peixe	Cattley guava (<i>Psidium littorale</i> , Myrtaceae)	3	September-November
	Sweet orange (Citrus sinensis, Rutaceae)	1	March–April
	Fig (Ficus carica, Moraceae)	2	August-September
Vila Franca	Cattley guava (<i>Psidium littorale</i> , Myrtaceae)	2	September–November
	Hot pepper (<i>Capsicum annuum</i> , Solanaceae)	1	September
São Gonçalo	Feijoa (Feijoa sellowiana, Myrtaceae)	3	October–December

Table 1. Host fruit traps placed in São Gonçalo $(37^{\circ} 45' 14''N; 25^{\circ} 39' 56''W)$, Rabo de Peixe $(37^{\circ} 48' 25''N; 25^{\circ} 35' 23''W)$ and Vila Franca $(37^{\circ} 43' 02''N; 25^{\circ} 26' 11''W)$, São Miguel Island, to collect larvae and pupae of *C. capitata* and their parasitoids.

plastic containers with moist filter paper until Medfly or parasitoid adults emerged. In addition, all fruits were dissected to collect any Medfly that had remained within the host.

At the time each trap was collected in the field, we captured the adults of all parasitic Hymenoptera present on the fruits. These were brought back to the laboratory $(21 \pm 1^{\circ}C, 70 \pm 10\% \text{ RH})$, under natural photoperiodic conditions) and placed in individual screen cages, each with a source of diluted honey (10%) and an open vial containing 100 different sized Medfly larvae in artificial diet. Each female parasitoid was left with the Medfly until all potential hosts had pupated. The pupae were then collected and kept to monitor the incidence of parasitism.

The number of Medfly pupae recovered varied among host plants, both within and between years (Table 2). Four individual parasitoids were recovered in 2005 (in pupae from figs) but none in 2006. There was one male specimen of *Halticoptera patellana* (Dalman) (Pteromalidae identified by M.J. Verdú) a species with a wide geographic distribution (Noyes 2004). This species has previously been recorded in the Canary Islands but not in the Azores (Koponen and Askew 2002). Furthermore, while members of this genus have been reported from different Diptera (Bouček and Rasplus 1991), it is the first mention of *C. capitata* as a host.

Host fruit	2005		2006	
	Ceratitis capitata	Parasitoids	Ceratitis capitata	Parasitoids
Fig	202	4	96	0
Cattley guava	21	0	24	0
Feijoa	246	0	261	0
Hot pepper	1	0	7	0
Sweet orange	4	0	0	0
Total	474	4	388	0

Table 2. Number of pupae of *C. capitata* and their parasitoids collected in the host fruit traps during 2005 and 2006 placed in different sites on the island of São Miguel, Azores.

We also recovered one male and two female *Toxeumorpha nigricola* (Ferrière) (Pteromalidae identified by M.J. Verdu). This species has been recorded in a number of African countries from Morocco to South Africa, as a parasitoid of several leaf mining Lepidoptera and Diptera (Ferrière 1936; Bouček 1976; Bouček and Rasplus 1991). Koponen and Askew (2002) found this species in the Canary Islands and Madeira, but not in the Azores.

Of all parasitoids foraging on fruits in the field (at least eight species) only two, *Asobara rufescens* (Förster) (European Braconidae) and *Tachinaephagus zealandicus* Ashmead (Encyrtidae, introduced into USA from Australasia), successfully parasitised Medfly larvae and completed a generation under laboratory conditions. These parasitoids were identified by K. van Achterberg and M.J. Verdú, respectively.

Many Asobara species have been recorded as larval parasitoids of drosophilids; however, some species attack Anastrepha spp and C. capitata in Brazil (Fernandes and Zucchi 1990). Asobara rufescens has been found in Germany and The Netherlands (Achterberg 2004). In our rearing A. rufescens only produced males and no additional experiments could be carried out. Whether this was the result of virgin females laying unfertilised eggs or a male biased allocation by mated females in response to hosts of poor quality is not known.

Tachinaephagus zealandicus, a parasitoid of many species of muscoid flies (Geden, Ferreira de Almeida, and Prado 2003), is endemic in Australia and New Zealand (Gold and Dahlsten 1981). However, it was introduced into many countries as USA and is now distributed throughout much of the Southern Hemisphere (Olton 1971), in Italy and in the European Atlantic islands (Koponen and Askew 2002). Neuenschwander (1982) recorded T. zealandicus attacking the olive-fly tephritid Bactrocera oleae Gmelin but thought that it did not regularly attack fruit-infesting Tephritidae. Under laboratory conditions, the culture of *T. zealandicus* was maintained for more than 10 generations on Medfly larvae, indicating this host is acceptable in a no-choice situation. Given that T. zealandicus females have the ability to attack >25 larvae per day (Almeida, Geden, and Prado 2002; Almeida, Prado, and Geden 2002), this species might have some potential in a biological control program using inundative releases. However, it should be noted that no Medfly pupae collected during the survey were parasitised by T. zealandicus (Table 2), suggesting that other preferred hosts exist in the field. Before any attempts of inundative releases against C. capitata, it would be necessary to obtain information about alternative hosts available in the habitat and their abundance relative to Medfly densities.

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