

Stem photosynthetic efficiency across woody angiosperms and gymnosperms with contrasting drought tolerance

Sara Natale^{1,2} · Francesco Petruzzellis¹ · Alessandro Alboresi² · Nicoletta La Rocca² · Tomas Morosinotto² · Andrea Nardini¹

Abstract

Key message Stem photosynthesis seems to play an adaptive role for woody plants that prosper in hot and dry ecosystems.

Abstract Stem photosynthesis is thought to be involved in tree resistance/resilience to water shortage. Recent studies have focused on the coordination between stem photosynthesis and hydraulics, but the generality of association of stem photosynthetic efficiency with species-specific adaptation to drought is still unclear. We quantified bark and wood chlorophyll *a* fluorescence (in terms of F_v/F_m) in current-year, 1-year and 2-year-old stems of several woody species harvested in diverse habitats. We ranked species in terms of relative drought tolerance on the basis of their vulnerability to xylem embolism (P_{50}), and compared stem photosynthetic efficiency of drought-tolerant vs drought-sensitive species. F_v/F_m values decreased with increasing stem age, and were generally higher for Angiosperms than Gymnosperms. F_v/F_m both at the bark and wood level was higher for drought-tolerant Angiosperms compared to drought-sensitive ones. Our results highlight the potential adaptive role of stem photosynthesis in drought-tolerant species, thriving under arid conditions likely leading to prolonged stomatal closure and halt of leaf-level carbon gain.

Keywords Bark $\cdot P_{50}$ · Stem photosynthesis · Drought · Wood · F_v/F_m

 Sara Natale sara.natale@phd.units.it
Francesco Petruzzellis fpetruzzellis@units.it
Alessandro Alboresi alessandro.alboresi@unipd.it
Nicoletta La Rocca nicoletta.larocca@unipd.it
Tomas Morosinotto tomas.morosinotto@unipd.it
Andrea Nardini nardini@units.it
Department of Life Sciences, University of Trieste, Trieste, Italy

² Present Address: Department of Biology, University of Padova, Padua, Italy

Introduction

Leaves are the most important photosynthetic organs in most woody plants, but nearly all of them can also perform this fundamental physiological process at stem level. Early observations of 'green stems' date back to the early twentieth century (e.g., Cannon 1908). Since then, photosynthetically competent stems have been detected in a wide range of woody plants typical of Mediterranean ecosystems, subtropical warm deserts, and tropical arid lands worldwide (e.g., Dima et al. 2006; Ávila et al. 2014). Indeed, photosynthetic stems are widely spread throughout different species and biomes (Teskey et al. 2008; Ávila et al. 2014). Stem photosynthesis is obviously common in succulent stems of CAM plants, but it occurs also in bark, wood, and pith of C_3 plants (Pfanz et al. 2002; Lüttge 2008; Kocurek et al. 2015; Wittmann et al. 2016). Stem photosynthetic efficiency is apparently species-specific and varies on a seasonal scale, thus suggesting that it might be very important from a functional and ecological point of view (Rentzou and Psaras 2008).

Although stems are not specialized for photosynthesis, due to their unfavorable surface-to-volume ratio and low light transmittance, it is known that chlorophyll is synthetized in functional chloroplasts even in deep wood layers of aged stems (Pfanz et al. 2002). Most woody species show a greenish, photosynthetically active inner bark (e.g., Pfanz et al. 2002; Wittmann and Pfanz 2008, 2014; Rosell et al. 2017), but chloroplasts can also be found along the xylem rays and in the pith, suggesting that they are not restricted to the relatively more illuminated outer cell layers of the stem (Dima et al. 2006; Berveiller et al. 2007; Yiotis et al. 2009). Among these stem compartments, bark exhibits higher chlorophyll concentration and photochemical efficiency (e.g., Yiotis et al. 2009; Wittmann and Pfanz 2016; Liu et al. 2018).

The amount and quality of light transmitted through the stem might be a key factor governing stem photosynthetic capacity (Manetas et al. 2005; Cernusak and Cheesman 2015; Wittmann and Pfanz 2016). Typically, about 15% of incident PAR can penetrate the periderm (Cernusak and Marshall 2000), but light transmittance changes according to stem anatomy (bark thickness, cell-wall lignification), age, and environment (Cernusak and Marshall 2000; Pfanz et al. 2002; Wittmann and Pfanz 2008). Blue light photons are largely absorbed by the outer bark, while red and far-red light penetrate more in depth, reaching the underlying xylem and pith. In addition, it is also possible that the optical properties of vascular tissue allow some axial light conduction to the pith cells, e.g., from roots (Sun et al. 2003).

Stems covered by an epidermis with stomata and/or periderm with lenticels can rely on net uptake of atmospheric CO_2 (Ávila et al. 2014). Otherwise, internal re-assimilation of CO_2 released by respiration of underlying heterotrophic tissues is also possible (e.g., Cernusak and Marshall 2000; Pfanz et al. 2002; Wittmann and Pfanz 2008; Berveiller et al. 2007). Internal CO_2 might be available at much higher concentrations within the stem (Teskey et al. 2008) compared to leaves, up to levels inhibiting photorespiration, thus increasing quantum yield of photosynthesis (Berveiller et al. 2007).

Previous studies have shown that net photosynthetic rates of stems can be up to 60% of those recorded for leaves (Ávila et al. 2014), and these rates are maintained even throughout the dry season when stomatal closure reduces leaf-level net carbon uptake (Ávila-Lovera et al. 2017). Hence, stem photosynthesis might provide an extra carbon gain under conditions imposing stomatal closure and limiting leaf photosynthesis. The carbon gain assured by stem photosynthesis might also contribute to bud development, flowering, and recovery after herbivores' attack (Saveyn et al. 2010; Kocurek et al. 2020). Stem photosynthesis has also been suggested to contribute to the maintenance and recovery of xylem hydraulic function (Saveyn et al. 2010; Schmitz et al. 2012; Bloemen et al. 2016; De Baerdemaeker et al. 2017; Trifilò et al. 2017, 2021; Tomasella et al. 2021).

Based on the above, stem photosynthesis might be particularly useful to maintain carbon uptake and physiological activities of plants during leafless periods, or under environmental conditions limiting leaf gas exchange (Ávila et al. 2014), with specific reference to drought. In particular, locally produced photosynthates might be used to sustain stem non-structural carbohydrates (NSC) pools to buffer carbon starvation under drought stress (De Roo and Salomón 2020), and refill embolized xylem upon drought relief (Schmitz et al. 2012; Liu et al. 2019).

In this study, we investigated possible differences in stem photosynthetic efficiency across a set of woody plants from different habitats. Our analysis was based on 39 woody species (both Angiosperms and Gymnosperms) from a broad phylogenetic range that were categorized in terms of relative drought tolerance on the basis of species-specific vulnerability to xylem embolism retrieved from the literature (see below). We specifically addressed the following questions: (i) are there differences in maximum quantum yield of photosystem II (F_v/F_m) of bark and wood in stems of different ages, and between various Angiosperm and Gymnosperm species? (ii) Are there any significant differences in stem F_v/F_m in drought-tolerant vs drought-sensitive species?

Material and methods

Plant material and experimental design

All experiments and measurements were carried out at the University of Trieste, Italy. Sampling of plant material was done in three different geographical areas to obtain stems from species from diverse families and/or adapted to different environmental conditions: (i) the botanical garden at the University of Trieste; (ii) the Classical Karst, a limestone plateau extending ~ 500 km², nearby Trieste; and (iii) montane forests nearby the village of Forni di Sopra, located in the Friulian Dolomites. Sampling was done between 6 and 8 a.m. in June 2021 for sites (i) and (ii), and in mid-July for site (iii). Three individuals per species were randomly selected, and two sun-exposed branches (about 3-4 years old) were sampled from each tree, for a total of six branches per species. Branches were immediately re-cut for 2-5 cm underwater and stored in a bucket with the cut section immersed in water to avoid dehydration until processing in the laboratory. Three branches (one per individual) were used for chlorophyll fluorescence measurements, while the other three branches were used to measure wood density, as described below. The complete list of species is reported in Fig. 2 and Table S1. Analyses

of photosynthetic efficiency were made on current-year, 1-year and 2-year-old stem segments, both at bark and wood level (Fig. 1).

Current-year stem segment





Bark Wood

Pith

1-year stem segment



2-year stem segment





Fig. 1 Example of a stem sample (Fraxinus ornus) selected for the measurements

Chlorophyll fluorescence

Measurement of chlorophyll *a* fluorescence is a powerful technique for in vivo analysis of different photosynthetic parameters (Maxwell and Johnson 2000). One of the most relevant parameters to estimate the effective yield (or maximal photochemical efficiency) of PSII is the F_v/F_m ratio (Maxwell and Johnson 2000; Schreiber 2004), which indicates the intrinsic efficiency of PSII photochemistry of dark-adapted samples (Aschan et al. 2005). Chlorophyll fluorescence was measured in the laboratory using an imaging PAM chlorophyll fluorometer equipped with a high-resolution camera (6.45 μ m × 6.45 μ m, resolution 1360×1024 px) (Open FluorCam SN-FC800-398, Photon Systems Instruments, Brno, Czech Republic) to quantify $F_{\rm v}/F_{\rm m}$ for bark, wood [outer xylem and xylem longitudinal section (sapwood + pith)] (Fig. 1, S1), and leaves (Tomasella et al. 2022; Natale et al. 2023). Specifically, each sample was placed at a distance of c.18 cm below the camera, and the LED panels were adjusted to an angle of 45° to the center of the measurement area to maintain a homogeneous illumination of samples. The standard protocol " F_v/F_m " was selected from the wizard and the appropriate triggering of the shutter opening of the CCD camera was adjusted. First, the basal fluorescence (F_{α}) was measured, and then, a saturating pulse of actinic light (~4000 μ mol photons m⁻² s⁻¹, 0.8 s) was turned on to induce maximum fluorescence (F_m), and F_v/F_m was calculated according to Maxwell and Johnson (2000) as: $F_v/F_m = (F_m-F_o)/F_m$. Each measurement took about 4 s.

Branches, still maintained with their cut end in water, were dark-adapted for 2 h before measurements to ensure relaxation of photosystems. Three to five mature leaves were detached from each branch and stem segments of different age were sectioned both longitudinally and radially, and bark and wood were manually and carefully separated. Samples were carefully placed on a petri dish with a bottom layer of moisten paper towels, and kept hydrated via partial immersion in a water film during the analysis with the PAM fluorometer.

Vulnerability to xylem embolism and wood density

Data of xylem pressure values inducing 50% loss of xylem hydraulic conductivity (P_{50}) for the study species were derived both from the Xylem Functional Traits (XFT) database (Choat et al. 2012), and integrated with data derived from more recent studies not included in the XFT. In accordance with previous studies (e.g., Petruzzellis et al. 2022), we considered only P_{50} data obtained for stems (excluding those measured on trunk, root, petiole, and leaf) and we discarded values based on r-shaped vulnerability curves (Cochard et al. 2013). When more than one value was available for a given species, we considered the average value from different studies for subsequent analysis. Mean values of traits for each species included in the study, along with the relative references, are reported in Tables S1 and S2.

Based on global relationships between P_{50} and environmental water availability (Maherali et al. 2004; Choat et al. 2012), we set a threshold of $P_{50} = -3.0$ MPa to separate relatively drought-tolerant ($P_{50} < -3.0$ MPa) from relatively drought-sensitive ($P_{50} > -3.0$ MPa) species. We were thus able to separate two groups of Angiosperms based on their P_{50} values. The same approach was not possible for Gymnosperms, due to the relatively low number of species included in the study.

Wood density (WD) is a key functional trait indicating variation in wood structure/carbon investment, and often used as a proxy for vulnerability to embolism due to its general correlation with P_{50} (Hacke and Sperry 2001; Kiorapostolou et al. 2019). Basic WD was measured on 3 cm-long segments from 2-year-old stems (one stem per individual), as

WD = Wood dry weight / Wood fresh volume (g cm⁻³).

Bark was removed and samples were rehydrated overnight by immersion in vials filled with tap water. The following morning, samples were weighed, and their fresh volume was measured using a water displacement method (Hughes 2005; Petruzzellis et al. 2019). Samples were then ovendried at 70 °C for 24 h and weighed again to obtain their dry weight (Petruzzellis et al. 2022).

Statistical and phylogenetic analysis

Statistical analyses were performed with R (version 3.4.4) software (R Core Team 2019).

Boxplot panels were made using "ggplot2" package in R. Bar charts were prepared with Origin 9.0 software (Northampton, MA, USA). We set the F_v/F_m as the response variable and the drought sensitiveness (drought-tolerant or drought-sensitive species) and each sample (e.g., bark of current-year stem) as the first and second explanatory variables, respectively. Since homogeneity of variance assumption was violated, generalized least square (GLS) models were run, using the 'nlme' R package (Pinheiro et al. 2019), specifying a 'varPower' variance structure. Differences between groups (drought-tolerant vs drought-sensitive) were tested post hoc with Holm contrasts using the emmeans function ("emmeans" package). For GLS models, the pseudo R^2 was calculated using the Nagelkerke method ("rcompanion" R package; Nagelkerke 1991).

To account for eventual phylogenetic signals from general trends of variation in stem photosynthesis and drought-tolerance traits, we constructed a phylogenetic tree according to Fletcher et al. (2018) and used "*pgls*" function in the "caper" R package to analyze data using a phylogenetic generalized least-squares (PGLS) approach. The PGLS approach enabled calculations of relationships between variables controlling for possible phylogenetic signals (Garland et al., 2005), by taking into account the phylogenetic autocorrelation when testing the relationship between the considered traits. Covariance analysis was based on the Brownian evolution model. Branch length transformation was optimized by calculating Pagel's lambda (λ , Pagel 1999) using the «maximum like-lihood» method (Lambda = 1: phylogenetic co-variation is equal to that estimated by the Brownian evolution model, Lambda = 0: traits' evolution is independent of phylogeny).

We also used the "phylobase" R package to combine the phylogenetic data with the dataset containing the P_{50} values. Then, we used "ggtree" R package to create a phylogenetic tree in which each species is color-coded according the corresponding P_{50} value, to highlight the evolutionary differences between species in terms of resistance to drought stress (Maherali et al. 2004).

Results

We retrieved data of P_{50} for all the species under study (Table S1, references in Table S2). Since our starting hypothesis was that stem photosynthesis might be linked to tree resistance/resilience to drought stress, we mapped P_{50} to the phylogenetic tree to better/visually show that the species "drought-tolerant" or "sensitive" were not phylogenetically close to each other, so that their drought tolerance level was most likely linked to a convergent evolution. Data showed in Fig. 2 exclude the possibility of strong phylogenetic bases for the correlation between P_{50} and PSII function. Indeed, high resistance to embolism (more negative P_{50}) appeared in several species, both Angiosperms and Gymnosperms (Fig. 2), very distant from each other from a phylogenetic point of view. The highest value was found in Sambucus nigra (-1.2 MPa), while Juniperus communis had the lowest value (-5.7 MPa).

Mean values of WD for each species are reported in Table S1. The lowest WD was measured in *Tilia cordata* $(0.30 \pm 0.01 \text{ g cm}^{-3})$, while *Quercus ilex* had the highest value $(0.68 \pm 0.07 \text{ g cm}^{-3})$. A significant negative relationship was found between P₅₀ and WD for all the years investigated (c-yr stem segments: P = 0.03, $R^2 = 0.11$, Lambda = 0.38; 1-yr stem segments: P = 0.02, $R^2 = 0.13$, Lambda = 0.28). We decided to show only the data for the 2-yr stem segments, with lower *P* value and higher R^2 (Fig. 3).

 F_v/F_m of leaves, bark, and wood for each species and each growth year are also reported in Table S3. Values of F_v/F_m for leaves were above 0.80 for all the study species. The lowest F_v/F_m value for wood was recorded in *Pinus halepensis*

Fig. 2 Phylogenetic tree showing species selected for this study, color-coded according to their values of xylem water potential inducing 50% loss of hydraulic conductivity (P_{50}). Note that more negative values characterize drought-tolerant species. Data are retrieved from literature.



 (0.42 ± 0.5) , with *Cercis siliquastrum* scoring the highest one (0.81 ± 0.02) . Instead, the lowest F_v/F_m value at bark level was measured in *Prunus spinosa* (0.62 ± 0.08) , while *Laurus nobilis* had the highest value (0.83 ± 0.004) . In both Angiosperms and Gymnosperms, average F_v/F_m values for leaves were similar across different species, but consistently higher than those measured at bark and wood level in each species. When considering the wood, Angiosperms showed higher values of F_v/F_m than Gymnosperms, especially when considering the wood (Fig. 4). We found a decreasing trend of photosynthetic efficiency with increasing stem age. This trend was more evident for Gymnosperms than for Angiosperms, and more marked when considering the wood, compared to the bark (Fig. 4).

When exploring possible differences in F_v/F_m according to the relative drought tolerance of different species based on the P_{50} threshold, we found that F_v/F_m values for the leaves were similar for drought-tolerant and drought-sensitive species. However, significant differences emerged between the two groups when considering bark of 1-year-old (p=0.02) and 2-year-old stems (p=0.02) (Fig. 5a, Table S4A). When considering wood, we found significant differences between the two groups in the current-year (p = 0.007) and 1-year-old stems (p = 0.023) (Fig. 5b, Table S4B). The trend toward lower F_v/F_m values in drought-sensitive compared to drought-tolerant species was maintained also in the 2-yearold wood, although no significant differences were found in this case due to high variability of data.

Values of F_v/F_m in the bark did not change over different stem ages for drought-tolerant species. On the contrary, in the drought-sensitive species, bark F_v/F_m showed a decreasing trend with age (Table S4A). F_v/F_m values in the wood tended to decrease with age in both drought-tolerant and drought-sensitive species (Table S4B).

Discussion

Stem photosynthesis is thought to play different roles in tree resistance/resilience to water shortage, but the actual correlation of species-specific stem photosynthetic efficiency with adaptation to drought has never been clearly demonstrated.



Fig. 3 Relationship between xylem water potential inducing 50% loss of hydraulic conductivity (P_{50}) and wood density (WD) obtained from "pgls". P_{50} data are retrieved from literature; WD was measured in the species selected for this study. Solid and open circles represent Angiosperm and Gymnosperm species, respectively. Solid line represents the overall regression, while shaded area represents 95% confidence intervals (r^2 =0.24)



Cy: current-year stem; 1y: 1-year stem; 2y: 2-year stem.

Fig. 4 Median values, 25th and 75th percentiles of $F_{\rm v}/F_{\rm m}$ values measured in (**a**, **b**) Angiosperms and (**c**, **d**) Gymnosperms in leaf (same data for both left and right panels) as well as bark and wood of current-year, 1-year-old and 2-year-old stem segments. Each point represents one species. Different letters indicate significant differences between samples (P < 0.05) obtained by gls model

Our data reveal significant differences in F_v/F_m for both bark and wood of species with contrasting levels of drought tolerance, highlighting possible functional roles of stem photosynthesis for woody plants thriving in water-limited habitats. Phylogenetic analysis suggested that such differences were not arising by evolutionary co-variation of stem hydraulic and photosynthetic properties, but rather provide evidence for convergent adaptation to drought involving both functional traits.

Chlorophyll *a* fluorescence is considered an excellent tool to quantify the physiological status of photosynthetic tissues (Govindjee 2004), and F_v/F_m is the parameter most commonly used to this aim. The average F_v/F_m for photosynthetically active and healthy leaves is about 0.83 (Harbinson et al. 2018). Lower values indicate reduced maximum quantum efficiency, due to intrinsic features of the photosynthetic apparatus or to damage resulting from environmental stress (Maxwell and Johnson 2000). Hence, F_v/F_m emerges as a convenient parameter to assess stem photosynthetic efficiency. F_v/F_m for leaves of the study species was above 0.80, and similar among Angiosperms and Gymnosperms. Interestingly, the F_v/F_m of bark for both current-year stems and 1-year-old stems was very close to values measured in leaves (although significantly lower based on statistical analysis), suggesting that the bark of young stems can perform photosynthesis with an efficiency very similar to that of leaves. The current-year wood displayed F_v/F_m particularly high in Angiosperms compared to Gymnosperms, indicating that even this compartment might significantly contribute to carbon uptake in this group of plants.

We found F_v/F_m to be stem age dependent, with relatively high values in current-year stems, but then decreasing in bark and especially wood of older stems. This finding is consistent with the observation that light penetration and CO₂ diffusion generally decrease from the outer surface inside the stem, and as stems age and grow thicker (Saveyn et al. 2010; Wittmann et al. 2006; Wittman and Pfanz 2018). In accordance, blue light is mainly absorbed by the bark, while the wood can be reached by far-red enriched light (Wittmann and Pfanz 2016; Natale et al. 2023) that is far from optimal for the photosynthetic process. However, it is interesting to note that even 2-year-old wood displayed appreciable levels of F_v/F_m (especially in Angiosperms), consistent with possible residual CO₂ fixation even in this compartment. This finding would be in agreement with the role proposed for wood chloroplast as 'recyclers' of CO₂ released by respiration of wood parenchyma (Ávila et al. 2014). This process might help plants to buffer NSC depletion during periods of stress leading to stomatal closure (see below). Another factor possibly influencing the recorded age-dependent trend might be the progressive chloroplast senescence induced in the inner stem portions due to the reduction of incident light, and this hypothesis deserves further studies.



Cy: current-year stem; 1y: 1-year stem; 2y: 2-year stem.

Fig. 5 Mean maximum quantum yield of PSII $(F_{\gamma}/F_{\rm m})$ in drought-tolerant and drought-sensitive Angiosperm species, according to values of xylem water potential inducing 50% loss of hydraulic conductivity (P_{50}) . Values are reported for leaves (same data for both left and right

panels), as well as for bark and wood samples of different age. Vertical bars indicate standard deviation. Asterisks indicate significant differences between groups (P < 0.05)

As expected, statistical analyses revealed that F_v/F_m values of bark and wood differed between Angiosperm and Gymnosperm species. Specifically, Angiosperms had higher F_v/F_m values, especially in the wood, compared to Gymnosperms. These differences are consistent with the xylem anatomical features in the two groups, with specific reference to the percentage wood volume occupied by parenchymatic cells. In fact, Gymnosperms have typically low amounts of parenchyma in their wood (in the range of 5–10% of the total volume), in sharp contrast with Angiosperms where 25–50% of the wood volume can be occupied by living cells (Morris et al. 2016; Kiorapostolou et al. 2019). Hence, Gymnosperm wood can accommodate relatively fewer chloroplasts and is likely less efficient in light transmission below the bark, compared to that of Angiosperms.

The vulnerability of xylem to embolism (P_{50}) is considered one of the most reliable proxies to quantify plant tolerance to drought stress (e.g., Choat et al. 2012; Petruzzellis et al. 2022). P_{50} is a measure of the apoplastic vulnerability to dehydration, since drought stress can affect the soil-toleaf pathway by causing dysfunction of water columns in xylem conduits (e.g., Nardini and Luglio 2014), potentially inducing embolism formation and further desiccation (e.g., Nardini et al. 2018; McDowell et al. 2008; Adams et al. 2009). Indeed, P_{50} values show large inter-specific variation, and are generally coordinated with climatic conditions of the species' natural range with specific reference to precipitation, evapotranspiration, and temperature (Maherali et al. 2004; Choat et al. 2012). Values of P_{50} in our dataset spanned a twofold range for both Angiosperms and Gymnosperms, and allowed us to separate two groups of relatively drought-tolerant (11 species) and drought-sensitive (20 species) Angiosperms. The same classification was not possible for Gymnosperms, due to the relatively small number of species analyzed. Drought-tolerant and drought-sensitive species were distributed along independent phylogenetic trajectories; therefore, their level of drought adaptation is not due to phylogenetic proximity (Fig. 2). A possible pitfall of our analysis lies in the fact that P_{50} values were retrieved from the literature, and not measured on the same individuals used for F_v/F_m measurements. Although species-specific vulnerability to xylem embolism displays limited genotypic variability (Wortemann et al. 2011; Unterholzner et al. 2020), we decided to measure wood density in the same plants used for analysis of stem photosynthetic efficiency to further test the overall reliability of the P_{50} dataset as based on literature data. WD is recognized to be positively linked to hydraulic safety in some species, and negatively related to hydraulic efficiency in most species (Chave et al. 2009; Markesteijn et al. 2011; Scholz et al. 2013; Nardini et al. 2013; Kiorapostolou et al. 2019), and this trait was correlated with P_{50} values across our dataset, confirming the overall validity of our classification of Angiosperms according to their relative drought tolerance. We found that woody species with more negative P_{50} values were also characterized by higher F_v/F_m values in bark and wood, although not for all age ranges. This finding might suggest that drought-tolerant species have greater stem photosynthetic efficiency, as a possible adaptation to environmental conditions imposing prolonged leaf stomatal closure.

It is also noteworthy that drought-tolerant species have slightly lower leaf F_v/F_m values than drought-sensitive species, but the former have generally greater stem F_v/F_m than the latter. This finding might indicate a possible trade-off between leaf and stem photosynthetic efficiencies. These conclusions would be in accordance with the previous studies, suggesting that photosynthetic stems could have evolved as an adaptation to life in tropicaldry and Mediterranean-like ecosystems (Ávila-Lovera and Ezcurra 2016; Ávila-Lovera and Garcillán 2020). In fact, under prolonged drought, leaf stomata close to prevent excessive tissue dehydration (Brodribb and Holbrook 2003; Bartlett et al. 2016; Choat et al. 2018). Stomatal closure cannot fully prevent residual water loss and progressive drop of xylem pressure, so that these species gain an adaptive advantage from developing xylem conduits highly resistant to xylem embolism that can assure the integrity of the root-to-leaf water pathway. Under such conditions, plants face a reduction of leaf-level photosynthetic carbon gain (Meinzer et al. 2009; Choat et al. 2012; Manzoni et al. 2013; Nardini et al. 2018), potentially leading to depletion of stored non-structural carbohydrates (NSC) to maintain primary and secondary metabolism (McDowell et al. 2022). Prolonged drought stress might provoke exhaustion of NSC pools (McDowell et al. 2008; Adams et al. 2009), so that additional carbon gain at the stem level might represent a significant adaptive advantage for such species.

In this study, we used P_{50} values to disentangle droughtsensitive from drought-tolerant species, and to compare stem photosynthetic efficiency in these two groups. Hence, our data cannot provide any insight into possible mechanistic relationships between vulnerability to xylem embolism and stem photosynthesis. However, some observations lead to interesting speculations. In particular, it is worth noting that wood photosynthetic efficiency was higher in Angiosperms than in Gymnosperms. Locally produced non-structural carbohydrates have been suggested to be involved in the post-drought recovery of stem hydraulics, pending refilling of embolized conduits (Schmitz et al. 2012; Bloemen et al. 2016; Nardini et al. 2018). In this light, it is important to remember that embolism repair has been observed more frequently in Angiosperms compared to Gymnosperms (Johnson et al. 2012; Klein et al. 2018). Thus, it is tempting to speculate that the limited ability of Gymnosperms to recover their hydraulic functions after drought partly depends on their low photosynthetic efficiency at wood level, that would prevent adequate production of non-structural carbohydrates to be invested in the generation of osmotic forces which are thought to be required for embolism reversal (Nardini et al. 2011; Secchi and Zwieniecki 2016; Secchi et al. 2021). It is also possible to speculate that PSII function in drought-tolerant species is important for some oxygen production at wood level, thus favoring respiration and ATP production based on consumption of starch reserves under conditions of impaired photosynthesis, but further studies are required to support or discard such speculative conclusions.

To conclude, our data suggest an adaptive role for stem photosynthesis for woody plants thriving in hot and dry ecosystems. Considering that this is the first report showing the connection between stem photosynthesis and drought-related functional traits, we call for more studies aimed at verifying these findings at larger geographical, ecological, and taxonomic scales, to better understand the ecophysiological importance of photosynthetic stems for plants facing ongoing climate changes.

Author contributions AN, SN, and FP planned and designed the research. SN performed experimental measurements. SN, FP, and AN analyzed the data. TM, NLR, and AA contributed to the interpretation of the data. SN and AN wrote the manuscript, with contribution by all co-authors.

Funding This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Data availability The data supporting the findings of this study are either reported in Supplementary Information files, or available from the corresponding author upon reasonable request.

Declarations

Conflict of interest We declare the absence of any competing interests at the time of submission.

References

- Adams HD, Guardiola-Claramonte M, Barron-Gafford GA, Villegas JC, Breshears DD, Zou CB, Troch PA, Huxman TE (2009) Reply to Leuzinger et al.: drought-induced tree mortality temperature sensitivity requires pressing forward with best available science. Proc Natl Acad Sci USA 106:E107
- Aschan G, Pfanz H, Vodnik D, Batič F (2005) Photosynthetic performance of vegetative and reproductive structures of green hellebore (*Helleborus viridis* L. agg.). Photosynthetica 43:55–64
- Ávila E, Herrera A, Tezara W (2014) Contribution of stem CO₂ fixation to whole-plant carbon balance in nonsucculent species. Photosynthetica 52:3–15
- Ávila-Lovera E, Ezcurra E (2016) Stem-succulent trees from the old and new world tropics. In: Goldstein G, Santiago LS (eds) Tropical tree physiology. Springer International, Cham, pp 45–65
- Ávila-Lovera E, Garcillán PP (2020) Phylogenetic signal and climatic niche of stem photosynthesis in the mediterranean and desert regions of California and Baja California Peninsula. Am J Bot 108:334–345
- Ávila-Lovera E, Zerpa AJ, Santiago LS (2017) Stem photosynthesis and hydraulics are coordinated in desert plant species. New Phytol 216:1119–1129
- Ávila-Lovera E, Haro R, Ezcurra E, Santiago LS (2018) Costs and benefits of photosynthetic stems in desert species from southern California. Funct Plant Biol 46:175–186

- Ávila-Lovera E, Garcillán PP, Silva-Bejarano C, Santiago LS (2020) Functional traits of leaves and photosynthetic stems of species from a sarcocaulescent scrub in the southern Baja California Peninsula. Am J Bot 107:1410–1422
- Bartlett MK, Scoffoni C, Sack L (2012) The determinants of leaf turgor loss point and prediction of drought tolerance of species and biomes: a global meta-analysis. Ecol Lett 15:393–405
- Bartlett MK, Zhang Y, Kreidler N, Sun S, Ardy R, Cao K, Sack L (2014) Global analysis of plasticity in turgor loss point, a key drought tolerance trait. Ecol Lett 17:1580–1590
- Bartlett MK, Klein T, Jansen S, Choat B, Sack L (2016) The correlations and sequence of plant stomatal, hydraulic and wilting responses to drought. PNAS USA 113:13098–13103
- Berveiller D, Kierzkowski D, Damesin C (2007) Interspecific variability of stem photosynthesis among tree species. Tree Physiol 27:53–61
- Bloemen J, Overlaet-Michiels L, Steppe K (2013) Understanding plant responses to drought: how important is woody tissue photosynthesis? Acta Hortic 991:149–157
- Bloemen J, Vergeynst L, Overlaet-Michiels L, Steppe K (2016) How important is woody tissue photosynthesis in poplar during drought stress? Trees 30:63–72
- Brodersen CR, McElrone AJ (2013) Maintenance of xylem network transport capacity: a review of embolism repair in vascular plants. Front Plant Sci 4:108
- Brodribb TJ, Holbrook NM (2003) Stomatal closure during leaf dehydration, correlation with other leaf physiological traits. Plant Physiol 132:2166–2173
- Cannon W (1908) The topography of the chlorophyll apparatus in desert plants. Carnegie Inst Wash Publ 98:2
- Cernusak LA, Cheesman AW (2015) The benefits of recycling: how photosynthetic bark can increase drought tolerance. New Phytol 208:995–997
- Cernusak LA, Marshall JD (2000) Photosynthetic refixation in branches of western white pine. Funct Ecol 14:300–311
- Chave J, Coomes D, Jansen S, Lewis SL, Sewnson NG, Zanne AE (2009) Towards a worldwide wood economics spectrum. Ecol Lett 12:351–366
- Choat B, Jansen S, Brodribb TJ, Cochard H, Delzon S, Bhaskar R, Bucci SJ, Feild TS, Gleason SM, Hacke UG, Jacobsen AL, Lens F, Maherali H, Martnez-Vilalta J, Mayr S, Mencuccini M, Mitchell PJ, Nardini A, Pittermann J, Pratt RB, Sperry JS, Westoby M, Wright IJ, Zanne AE (2012) Global convergence in the vulnerability of forests to drought. Nature 491:752–755
- Choat B, Brodribb TJ, Brodersen CR, Duursma RA, López R, Medlyn BE (2018) Triggers of tree mortality under drought. Nature 558:531–539
- Cochard H, Badel E, Herbette S, Delzon S, Choat B, Jansen S (2013) Methods for measuring plant vulnerability to cavitation: a critical review. J Exp Bot 64:4779–4791
- De Baerdemaeker NJ, Salomón RL, De Roo L, Steppe K (2017) Sugars from woody tissue photosynthesis reduce xylem vulnerability to cavitation. New Phytol 216:720–727
- De Roo L, Salomón RL, Steppe K (2020) Woody tissue photosynthesis reduces stem CO₂ efflux by half and remains unaffected by drought stress in young *Populus tremula* trees. Plant Cell Environ 43:981–991
- Dima E, Manetas Y, Psaras GK (2006) Chlorophyll distribution pattern in inner stem tissues: evidence from epifluorescence microscopy and reflectance measurements in 20 woody species. Trees 20:515–521
- Fletcher LR, Cui H, Callahan H, Scoffoni C, John GP, Bartlett MK, Burge DO, Sack L (2018) Evolution of leaf structure and drought tolerance in species of Californian *Ceanothus*. Am J Bot 105:1672–1687

- Govindjee, (2004) Chlorophyll a fluorescence: a bit of basics and history. In: Papageorgiou GC, Govindjee SS (eds) Chlorophyll a fluorescence a signature of photosynthesis. Springer, Dordrecht, pp 1–41
- Hacke UG, Sperry JS (2001) Functional and ecological xylem anatomy. Perspect Plant Ecol Evol 4:97–115
- Harbinson J, Croce R, van Grondelle R, van Amerongen H, van Stokkum I (2018) Chlorophyll fluorescence as a tool for describing the operation and regulation of photosynthesis in vivo. Light harvesting in photosynthesis. CRC Press, Boca Raton, pp 539–571
- Hughes SW (2005) Archimedes revisited: a faster, better, cheaper method of accurately measuring the volume of small objects. Phys Educ 40:468
- Johnson DM, McCulloh KA, Woodruff DR, Meinzer FC (2012) Hydraulic safety margins an embolism reversal in stems and leaves: why are conifers and angiosperms so different? Plant Sci 195:48–53
- JrT G, Bennett AF, Rezende EL (2005) Phylogenetic approaches in comparative physiology. J Exp Biol 208:3015–3035
- Kiorapostolou N, Da Sois L, Petruzzellis F, Savi T, Trifilò P, Nardini A, Petit G (2019) Vulnerability to xylem embolism correlates to wood parenchyma fraction in angiosperms but not in gymnosperms. Tree Physiol 39:1675–1684
- Klein T, Zeppel MJ, Anderegg WR, Bloemen J, De Kauwe MG, Hudson P et al (2018) Xylem embolism refilling and resilience against drought-induced mortality in woody plants: processes and tradeoffs. Ecol Res 33:839–855
- Kocurek M, Kornas A, Pilarski J, Tokarz K, Lüttge U, Miszalski Z (2015) Photosynthetic activity of stems in two *Clusia species*. Trees 29:1029–1040
- Kocurek M, Kornas A, Wierzchnicki R, Lüttge U, Miszalski Z (2020) Importance of stem photosynthesis in plant carbon allocation of *Clusia minor*. Trees 34:1009–81020
- Lamy JB, Delzon S, Bouche PS, Alia R, Vendramin GG, Cochard H (2014) Limited genetic variability and phenotypic plasticity detected for cavitation resistance in a Mediterranean pine. New Phytol 201:874–886
- Liu J, Gu L, Yu Y, Ju G, Sun Z (2018) Stem photosynthesis of twig and its contribution to new organ development in cutting seedlings of *Salix Matsudana* Koidz. Forests 9:207
- Liu J, Gu L, Yu Y, Huang P, Wu Z, Zhang Q, Qian Y, Wan X, Sun Z (2019) Corticular photosynthesis drives bark water uptake to refill embolized vessels in dehydrated branches of *Salix Matsudana*. Plant Cell Environ 42:2584–2596
- Lüttge U (2008) Stem CAM in arborescent succulents. Trees 22:139–148
- Maherali H, Pockman WT, Jackson RB (2004) Adaptive variation in the vulnerability of woody plants to xylem cavitation. Ecology 85:2184–2199
- Manetas Y, Pfanz H (2005) Spatial heterogeneity of light penetration through periderm and lenticels and concomitant patchy acclimation of corticular photosynthesis. Trees 19:409–414
- Manzoni S, Vico G, Katul G, Palmroth S, Jackson RB, Porporato A (2013) Hydraulic limits on maximum plant transpiration and the emergence of the safety–efficiency trade-off. New Phytol 198:169–178
- Markesteijn L, Poorter L, Paz H, Sack L, Bongers F (2011) Ecological differentiation in xylem cavitation resistance is associated with stem and leaf structural traits. Plant, Cell & Environ 34:137–148
- Maxwell K, Johnson GN (2000) Chlorophyll fluorescence—a practical guide. J Exp Bot 51:659–668
- McDowell NG (2011) Mechanisms linking drought, hydraulics, carbon metabolism, and vegetation mortality. Plant Physiol 155:1051–1059

- McDowell N, Pockman WT, Allen CD, Breshears DD, Cobb N, Kolb T, Plaut J, Sperry J, West A, Williams DG et al (2008) Mechanisms of plant survival and mortality during drought: why do some plants survive while others succumb to drought? New Phytol 178:719–739
- McDowell NG, Sapes G, Pivovaroff A, Adams HD, Allen CD, Anderegg WR et al (2022) Mechanisms of woody-plant mortality under rising drought, CO₂ and vapour pressure deficit. Nat Rev Earth Environ 3:294–308
- Meinzer FC, Johnson DM, Lachenbruch B, McCulloh KA, Woodruff DR (2009) Xylem hydraulic safety margins in woody plants: coordination of stomatal control of xylem tension with hydraulic capacitance. Funct Ecol 23:922–930
- Méndez-Alonzo R, Paz H, Zuluaga RC, Rosell JA, Olson ME (2012) Coordinated evolution of leaf and stem economics in tropical dry forest trees. Ecology 93:2397–2406
- Mitchell PJ, O'Grady AP, Tissue DT, White DA, Ottenschlaeger ML, Pinkard EA (2013) Drought response strategies define the relative contributions of hydraulic dysfunction and carbohydrate depletion during tree mortality. New Phytol 197:862–872
- Morris H, Plavcová L, Cvecko P, Fichtler E, Gillingham MAF, Martínez Cabrera HI, McGlinn DJ, Wheeler E, Zheng J, Ziemińska K, Jansen S (2016) A global analysis of parenchyma tissue fractions in secondary xylem of seed plants. New Phytol 209:1553–1565
- Nagelkerke NJD (1991) A note on a general definition of the coefficient of determination. Biometrika 78:691–692
- Nardini A, Luglio J (2014) Leaf hydraulic capacity and drought vulnerability: possible trade-offs and correlations with climate across three major biomes. Funct Ecol 28:810–818
- Nardini A, Lo Gullo MA, Salleo S (2011) Refilling embolized xylem conduits: is it a matter of phloem unloading? Plant Sci 180:604–611
- Nardini A, Battistuzzo M, Savi T (2013) Shoot desiccation and hydraulic failure in temperate woody angiosperms during an extreme summer drought. New Phytol 200:322–329
- Nardini A, Savi T, Trifilò P, Lo Gullo MA (2018) Drought stress and the recovery from xylem embolism in woody plants. Prog Bot 79:197–231
- Natale S, La Rocca N, Battistuzzi M, Morosinotto T, Nardini A, Alboresi A (2023) Structure and function of bark and wood chloroplasts in a drought tolerant tree (Fraxinus ornus L.). Tree Physiol. https://doi.org/10.1093/treephys/tpad013
- Pagel M (1999) Inferring the historical patterns of biological evolution. Nature 401:877–884
- Petruzzellis F, Nardini A, Savi T, Tonet V, Castello M, Bacaro G (2019) Less safety for more efficiency: water relations and hydraulics of the invasive tree *Ailanthus altissima* (Mill.) Swingle compared with native *Fraxinus ornus* L. Tree Physiol 39:76–87
- Petruzzellis F, Tordoni E, Di Bonaventura A, Tomasella M, Natale S, Panepinto F, Bacaro G, Nardini A (2022) Turgor loss point and vulnerability to xylem embolism predict species-specific risk of drought-induced decline of urban trees. Plant Biol 24:1198–1207
- Pfanz H, Aschan G, Langenfeld-Heyser R, Wittmann C, Loose M (2002) Ecology and ecophysiology of tree stems: corticular and wood photosynthesis. Naturwissenschaften 89:147–162
- Pinheiro J, Bates D, DebRoy S, Sarkar D. R Core Team. Nlme: Linear and Nonlinear Mixed Effects Models. 2019. Available online: https://CRAN.R-project.org/package=nlme.
- Pivovaroff AL, Cook VM, Santiago LS (2018) Stomatal behaviour and stem xylem traits are coordinated for woody plant species under exceptional drought conditions. Plant Cell Environ 41:2617–2626
- Rentzou A, Psaras GK (2008) Green plastids, maximal PSII photochemical efficiency and starch content of inner stem tissues of three Mediterranean woody species during the year. Flora: Morphol. Distrib Funct Ecol Plants 203:350–357

- Rosell JA, Olson ME, Anfodillo T, Martínez-Méndez N (2017) Exploring the bark thickness-stem diameter relationship: clues from lianas, successive cambia, monocots and gymnosperms. New Phytol 215:569–581
- Saveyn A, Steppe K, Ubierna N, Dawson TE (2010) Woody tissue photosynthesis and its contribution to trunk growth and bud development in young plants. Plant Cell Environ 33:1949–1958
- Schmitz N, Egerton JJG, Lovelock CE, Ball MC (2012) Light-dependent maintenance of hydraulic function in mangrove branches: do xylary chloroplasts play a role in embolism repair? New Phytol 195:40–46
- Scholz A, Klepsch M, Karimi Z, Jansen S (2013) How to quantify conduits in wood? Front Plant Sci 4:56
- Schreiber U (2004) Pulse-amplitude-modulation (PAM) fluorometry and saturation pulse method: an overview. In: Papageorgiou GC, Govindjee DD (eds) Chlorophyll a fluorescence: a signature of photosynthesis advances in photosynthetic respiration, vol 19. Springer, Dordrecht, pp 279–319
- Secchi F, Zwieniecki MA (2016) Accumulation of sugars in the xylem apoplast observed under water stress conditions is controlled by xylem pH. Plant Cell Environ 39:2350–2360
- Secchi F, Pagliarani C, Cavalletto S, Petruzzellis F, Tonel G, Savi T et al (2021) Chemical inhibition of xylem cellular activity impedes the removal of drought-induced embolisms in poplar stems–new insights from micro-CT analysis. New Phytol 229:820–830
- Steppe K, Sterck F, Deslauriers A (2015) Diel growth dynamics in tree stems: linking anatomy and ecophysiology. Trends Plant Sci 20:335–343
- Sun Q, Kiyotsugu Y, Mitsuo S, Hitoshi S (2003) Vascular tissue in the stem and roots of woody plants can conduct light. J Exp Bot 54:1627–1635
- Teskey RO, Saveyn A, Steppe K, McGuire MA (2008) Origin, fate and significance of CO_2 in tree stems. New Phytol 177:17–32
- Tomasella M, Petrussa E, Petruzzellis F, Nardini A, Casolo V (2019) The possible role of non-structural carbohydrates in the regulation of tree hydraulics. Int J Mol Sci 21:144
- Tomasella M, Casolo V, Natale S, Petruzzellis F, Kofler W, Beikircher B et al (2021) Shade-induced reduction of stem nonstructural carbohydrates increases xylem vulnerability to embolism and impedes hydraulic recovery in *Populus nigra*. New Phytol 231:108–121
- Tomasella M, Natale S, Petruzzellis F, Di Bert S, D'Amico L, Tromba G, Nardini A (2022) No evidence for light-induced embolism repair in cut stems of drought-resistant Mediterranean species under soaking. Plants 11:307
- Trifilò P, Casolo V, Raimondo F, Petrussa E, Boscutti F, Lo Gullo MA, Nardini A (2017) Effects of prolonged drought on stem non- structural carbohydrates content and post-drought hydraulic recovery in *Laurus nobilis* L.: The possible link between carbon starvation and hydraulic failure. Plant Physiol Biochem 120:232–241
- Trifilò P, Kiorapostolou N, Petruzzellis F, Vitti S, Petit G, Lo Gullo MA, Nardini A, Casolo V (2019) Hydraulic recovery from xylem embolism in excised branches of twelve woody species: relationships with parenchyma cells and non-structural carbohydrates. Plant Physiol Biochem 139:513–520
- Trifilò P, Natale S, Gargiulo S, Abate E, Casolo V, Nardini A (2021) Stem photosynthesis affects hydraulic resilience in the deciduous *Populus alba* but not in the evergreen *Laurus nobilis*. Water 13:2911
- Unterholzner L, Carrer M, Bär A, Beikircher B, Dämon B et al (2020) *Juniperus communis* populations exhibit low variability in hydraulic safety and efficiency. Tree Physiol 40:1668–1679
- Vandegehuchte MW, Bloemen J, Vergeynst LL, Steppe K (2015) Woody tissue photosynthesis in trees: salve on the wounds of drought? New Phytol 208:998–1002

- Wittmann C, Pfanz H (2008) General trait relationships in stems: a study on the performance and interrelationships of several functional and structural parameters involved in corticular photosynthesis. Physiol Plant 134:636–648
- Wittmann C, Pfanz H (2014) Bark and woody tissue photosynthesis: a means to avoid hypoxia or anoxia in developing stem tissues. Funct Plant Biol 41:940–953
- Wittmann C, Pfanz H (2016) The optical, absorptive and chlorophyll fluorescence properties of young stems of five woody species. Environ Exp Bot 121:83–93
- Wortemann R, Herbette S, Barigah TS, Fumanal B, Alia R, Ducousso A, Gomory D, Roeckel-Drevet P, Cochard H (2011) Genotypic variability and phenotypic plasticity of cavitation resistance in Fagus sylvatica L. across Europe. Tree Physiol 31:1175–1182
- Yiotis C, Petropoulou Y, Manetas Y (2009) Evidence for light-independent and steeply decreasing PSII efficiency along twig depth in four tree species. Photosynthetica 47:223–231

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Springer Nature or its licensor (e.g. a society or other partner) holds exclusive rights to this article under a publishing agreement with the author(s) or other rightsholder(s); author self-archiving of the accepted manuscript version of this article is solely governed by the terms of such publishing agreement and applicable law.