



Presence of *Propionibacterium acnes* in granulomas associates with a chronic disease course in Dutch sarcoidosis patients

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ABSTRACT Several studies demonstrated that *Propionibacterium acnes* may be involved in sarcoidosis pathogenesis. Presence of *P. acnes* was found in granulomas of the majority of Japanese sarcoidosis patients. However, presence of *P. acnes* in tissue has never been related to sarcoidosis phenotypes and clinical outcome. Therefore, the aims of our study were to demonstrate whether *P. acnes* can be detected in granulomas of Dutch sarcoidosis patients and to investigate whether its presence is related to a clinical phenotype and/or course of disease.

Sections of formalin-fixed paraffin-embedded tissue blocks of 76 sarcoidosis patients were examined by immunostaining with a *P. acnes*-specific monoclonal antibody (PAB antibody) using a Ventana BenchMark ULTRA. Clinical outcome status (COS) was determined and classified into two phenotype groups: A: resolved, minimal or persistent disease without treatment (COS 1–6) and B: persistent disease with need for treatment (COS 7–9).

P. acnes was detected in samples of 31 patients (41%) and located within granulomas in samples of 13 patients (17%). The frequency of *P. acnes* detected in granulomas at diagnosis was significantly higher in patients with phenotype B compared to patients with phenotype A (29% *versus* 0%, p=0.021).

Presence of *P. acnes* in granulomas can be confirmed in Dutch sarcoidosis patients. It is intriguing that presence of *P. acnes* in granulomas is more frequently found in patients with chronic disease requiring treatment. This adds to the rationale that a subgroup of sarcoidosis patients might benefit from antibiotic therapy.



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Significantly more sarcoidosis patients with a chronic disease course requiring treatment have presence of *P. acnes* in granulomas. This contributes to the premise that it is relevant to further explore antibacterial therapy in sarcoidosis. https://bit.ly/36wCbbw

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Introduction

Sarcoidosis is a multisystem inflammatory disorder of unknown aetiology that is characterised by the presence of noncaseating granulomas. In over 90% of patients the lungs are involved [1], but the disease can also affect other organs [2]. Patients with symptomatic organ involvement or risk of permanent damage can be treated with immunosuppressive drugs [3], which suppress disease activity but do not cure the disease.

The cause of sarcoidosis is still puzzling scientists for more than a century. Several studies demonstrated that specific microorganisms, including mycobacteria and *Propionibacterium acnes* could be involved in its disease pathogenesis [4–9]. As *P. acnes* is a commensal, Koch's postulates cannot be applied, which makes it difficult to elucidate its aetiological role [10]. A higher number of *P. acnes* genomes have been found in tissue from Chinese, Japanese, Italian, English and German sarcoidosis patients compared to tissue of controls, suggesting involvement in disease pathogenesis [11–13]. Furthermore, an increased immune response to *P. acnes* among patients with sarcoidosis was found in different studies using Japanese or German patients [7, 8, 14]. Remarkably, in recent work from our own group, we found a lower percentage of Dutch sarcoidosis patients than controls with an immunological response to *P. acnes* [15], which is conflicting with the above-mentioned papers.

In addition to its role as possible antigen, *P. acnes* can also act as a mitogen, which is demonstrated by the fact that *P. acnes* enhances immunogenicity of certain vaccines [16] and enhances cytotoxic activity toward different tumour cells [17–19]. A potential mitogenic role of *P. acnes* in sarcoidosis has not previously been studied.

Based on both a possible antigenic as well as mitogenic role for *P. acnes*, we hypothesise that presence of *P. acnes* in tissue could be related to certain clinical phenotypes in sarcoidosis. We therefore examined the presence of *P. acnes* in tissue of Dutch patients with sarcoidosis using an existing *P. acnes*-specific monoclonal antibody (PAB antibody) [20] and related results of immunostaining to clinical characteristics such as organ involvement and course of disease.

Methods and materials

Study patients

Unstained tissue blocks were requested from two sarcoidosis cohorts previously studied at the St Antonius Hospital (Nieuwegein, the Netherlands) [15, 21]. The diagnosis of sarcoidosis had been established according to the criteria of the American Thoracic Society/European Respiratory Society [22]. Patients were included in the study when enough residual tissue was available and when presence of granulomas could be clearly detected in the haematoxylin and eosin (H&E) stained tissue sections. The study was approved by the Medical Research Ethics Committees United (MEC-U) of the St Antonius Hospital (R05-08A) and written consent was obtained from all patients.

Immunostaining

4-µm thick sections were cut from the formalin-fixed paraffin-embedded tissue sections, which were immunohistochemically stained with the PAB antibody; a P. acnes-specific monoclonal antibody that reacts with cell membrane-bound lipoteichoic acid (LTA) of the bacterium [20]. The PAB antibody was kindly provided by Y. Eishi and colleagues (Department of Human Pathology, Tokyo Medical and Dental University, Tokyo, Japan). We followed the protocol described by Negl et al. [20], but instead of the original manual procedures the sections were stained by the use of a Ventana BenchMark ULTRA (Ventana Medical Systems Inc., Tucson, AZ, USA) using an ultraView Universal Alkaline Phosphatase Red Detection kit (Ventana Medical Systems). We modified the original protocol to optimise the sensitivity and specificity of the staining results for the Ventana BenchMark ULTRA. Shortly, sections were de-paraffinised and rehydrated followed by antigen retrieval using the hot plate heating system of the machine instead of antigen retrieval by microwave. Because mineral oil (Liquid Coverslip, Ventana Medical Systems, Inc.) covering the tissue slides was found to inhibit the reaction with PAB antibody, before the primary antibody reaction with the PAB antibody, a washing step (three times for 5 min each) in the EZ Prep buffer (Roche Nederland B.V., Woerden, the Netherlands) was added under the Antibody Titration program selected, followed by washing for 5 min with tap water and 5 min with reaction buffer (Roche Nederland B.V.). The PAB antibody (crude mouse ascites fluid) was used in a concentration of 1:30 000 (diluted with DAKO REAL antibody diluent, S2022, DAKO, Glostrup, Denmark) and incubated for 16 min at room temperature. Instead of using the peroxidase substrate diaminobenzidine (DAB) to develop the signal, an ultraView Universal Alkaline Phosphatase Red Detection kit (Ventana Medical Systems) was used. Sections were counterstained with Mayer's haematoxylin. Detection of P. acnes in tissue was analysed by a pulmonary pathologist (K. Seldenrijk). The staining was considered positive when

small round/dot-like structures were seen. If such structures were detected, it was determined whether positive PAB staining was present in or outside the granulomas.

Identification of P. acnes-related phenotypes

To determine possible *P. acnes*-related phenotypes, organ involvement, age at diagnosis and Scadding stage at diagnosis and follow-up was collected from medical records of sarcoidosis patients.

The clinical outcome status (COS), a definition of clinical outcome in sarcoidosis established by the World Association of Sarcoidosis and Other Granulomatous Diseases [23]was determined 2 and 5 years after diagnosis. The disease status of patients was retrospectively examined and classified into resolved, minimal or persistent disease. Resolved was classified as patients showing no signs of disease, such as normalisation of chest radiography, pulmonary function tests and laboratory tests. Minimal disease is defined as a disease burden of 25% or less compared with the maximum disease burden experienced by patients measured by pulmonary function tests, chest radiography, biomarkers and skin lesions. For example, the worst pulmonary function test had to be improved by at least 75% to be considered minimal disease.

Furthermore, it was examined whether patients were ever treated, and if so whether they were still using medication or not (defined as no medication >1 year). Patients in whom medication was increased the last year were considered as worsening. Medication included all immunosuppressive systemic therapies used for sarcoidosis, including corticosteroids, disease-modifying antirheumatic drugs and anti-tumour necrosis factor (TNF)- α antibodies, except for nonsteroidal anti-inflammatory drugs. This results in nine COS scores. 1: Resolved disease, never treated; 2: resolved diseases, no treatment >1 year; 3: minimal disease, never treated; 4: minimal disease, no therapy >1 year; 5: persistent disease, never treated; 6: persistent disease, no therapy >1 year; 7: persistent disease, current therapy but no worsening in prior year and asymptomatic; 8: persistent disease, current therapy but no worsening in prior year and 9: persistent disease, current therapy that worsened in the prior year.

Patients who died in the period over which the COS was determined were placed in COS 9.

To analyse whether there was a correlation between presence of *P. acnes* and COS, we classified the COS scores into two phenotype groups: A: resolved, minimal or persistent disease without treatment (COS 1–6) and B: persistent disease with need for treatment (COS 7–9).

Statistical analysis

Data were analysed using IBM SPSS statistics version 24. An unpaired t-test was used to compare numerical data. Nonparametric tests were used for non-normally distributed data (Mann–Whitney U-test). Categorical data were compared using the Chi-squared test. If expected cell frequencies were below 5, Fisher's exact test was used for categorical data up to two categories. A p-value <0.05 was considered significant.

Results

Characteristics of study patients and tissue samples

Formalin-fixed paraffin-embedded tissue blocks were available from 76 patients. Mean age of included patients was 44 years, 84% were white and 71% had extrapulmonary involvement. Overall, 68% and 67% were classified in phenotype group B 2 and 5 years after diagnosis, respectively (table 1).

P. acnes can be detected in tissue samples of Dutch patients with sarcoidosis

Most tissue sections used for staining originated from the lung, followed by lymph node and skin samples. *P. acnes* was detected in tissue samples from 31 of the 76 sarcoidosis patients (41%) and was located in the granulomas in 13 samples of all patients (17%) (table 2 and figs. 1 and 2). When *P. acnes* was not detected inside granulomas, it was mostly located directly adjacent to granulomas in histiocytes and in a few cases in granuloma-free areas of the tissue. Besides lung, lymph node and skin tissue, we also found presence of *P. acnes* in bone marrow and liver tissue. No significant difference was observed in the frequency of *P. acnes* detected in tissue or granulomas between the different organs used in the study (p=0.583 and p=0.490, respectively). *P. acnes* was detected in tissue or granulomas in three (43%) and three (43%), respectively of the seven lung samples obtained by video-assisted thoracic surgery (VATS), and in seven (28%) and four (16%), respectively of the lung samples obtained by transbronchial lung biopsy (TBLB). No significant difference in presence of *P. acnes* in tissue or granulomas was observed between the different biopsy methods used for the lung samples (p=0.648 and p=0.157, respectively) and lymph node samples (p=0.167 and p=0.588, respectively).

	Sarcoidosis (n=76)
Age at time of biopsy years	43.98±12.22
Male sex	42 (55)
Ever smoker	43 (59)
White	64 (84)
Medication at time of biopsy	6 (8)
Third-line therapy [#]	27 (36)
COS group A/B (2 years follow-up)	23/49 (32/68)
COS group A/B (5 years follow-up)	15/31 (33/67)
Scadding stage at time biopsy (0/I/II/III/IV/unknown)	5/19/28/9/10/5 (7/25/37/12/13/3
Extrapulmonary involvement	54 (71)
Skin	18 (24)
Eyes	7 (9)
Liver	8 (11)
Heart	13 (17)
Spleen	3 (4)
Bones	4 (5)
Nerve system	15 (20)
Central	6 (8)
Peripheral	1 (1)
Small fibre neuropathy	10 (13)

Presence of P. acnes does not correlate with organ involvement

#: third-line therapy consisted of infliximab use at follow-up.

When relating positive *P. acnes* staining results to clinical characteristics, no relation with age, sex, ethnicity, organ involvement or Scadding stage was observed (table 3).

with pulmonary infiltrates; III: pulmonary infiltrates without BHL; IV: fibrosis. COS: clinical outcome status.

Presence of P. acnes correlates with clinical outcome

Presence of *P. acnes* in tissue was compared between the two COS groups defined 2 and 5 years after diagnosis. Two years after diagnosis, a trend (p=0.093) towards a higher prevalence of *P. acnes* in tissue was seen in COS group B (47%, 23 of 49) compared to COS group A (26%, 6 of 23). When comparing presence of *P. acnes* specifically inside granulomas, also a trend (p=0.050) towards a higher prevalence was seen in COS group B patients (25%, 12 of 49) compared to COS group A patients (4%, 1 of 23) (fig. 3a). COS defined after 5 years revealed a trend (p=0.072) towards a higher prevalence of *P. acnes* in tissue in COS group B (55%, 17 of 31) compared to COS group A (27%, 4 of 15). A significantly higher percentage of patients within COS group B (29%, 9 of 31) were positively stained for *P. acnes* inside granulomas compared to COS group A patients (0, 0 of 15) (p=0.021, fig. 3b).

As COS group B can be a very heterogeneous group of patients, ranging from patients with remission of symptoms due to long-term corticosteroid treatment to severe therapy-refractory patients requiring third-line medication, we also determined whether presence of *P. acnes* in tissue and granulomas was specifically related to patients requiring third-line medication at follow-up. In a significantly higher percentage of patients using infliximab, *P. acnes* was present in granulomas compared to patients not requiring third-line therapy (33% *versus* 8%, p=0.009) (fig. 4).

Discussion

In this study we demonstrated that *P. acnes* is present in granulomas of Dutch patients with sarcoidosis, which is in line with previous studies in Japanese and German sarcoidosis patients [20]. Furthermore, to the best of our knowledge, our study is the first to show a relation between presence of *P. acnes* and clinical outcome in sarcoidosis. Regardless of whether the role of *P. acnes* might be antigenic or mitogenic, these data add to the rationale that *P. acnes* might be involved in the disease pathogenesis of sarcoidosis.

NEGI et al. [20] previously examined the presence of *P. acnes* in tissue samples from sarcoidosis patients, using the same PAB antibody as used in the current study. A remarkable difference was found in percentage of *P. acnes*-positive sarcoidosis patients. They found *P. acnes* in granulomas in 57% of the 77 lung samples from Japanese sarcoidosis patients, whereas we found *P. acnes* in granulomas in 22% of the 32 lung samples from Dutch sarcoidosis patients. This difference could be partially explained by the lower

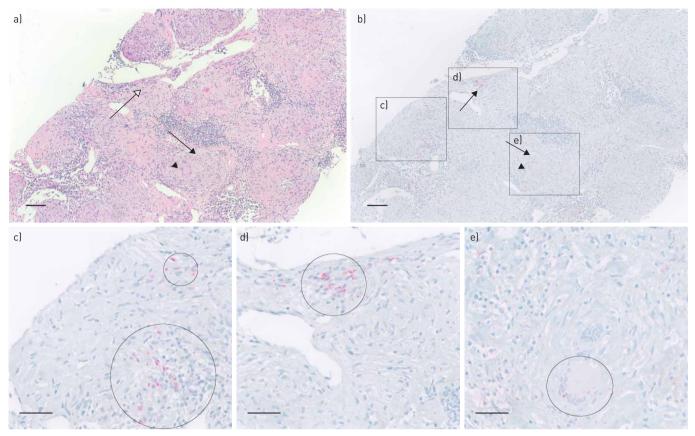


FIGURE 1 Presence of *Propionibacterium acnes* in and around an immature granuloma of the lymph node. a) Haematoxylin and eosin (H&E) staining of a needle lymph node biopsy, including several mature granulomas. Arrow: a mature granuloma including a multinucleated giant cell (arrow head). Open arrow: an immature granuloma. b) *P. acnes*-specific monoclonal antibody (PAB) staining corresponding with the area in a, including several areas with *P. acnes*-positive red signals (c and d) and a mature granuloma negative for *P. acnes* (e). c) Higher magnification of positive *P. acnes* staining (circles) around granulomas. d) Higher magnification of positive *P. acnes*. Scale bars=100 µm (a and b) and 50 µm (c-e)

proportion of VATS samples (35% in their study compared to 22% in our study). Negi et al. [20] described that *P. acnes* in granulomas was more frequently detected in VATS samples compared to TBLB samples, probably due to the larger size of VATS samples. Similar to the lung samples, the percentage of Dutch patients with presence of *P. acnes* in granulomas of lymph node samples (8%) was lower compared to Japanese and German patients (88% and 89%, respectively). Although the difference between Japanese and Dutch patients may be explained by difference in ethnicity, this is not a plausible explanation for the difference between German and Dutch patients. The difference in the detection frequency of *P. acnes* in granulomas can possible be caused by a lower sensitivity of immunostaining method used in our study. Therefore, future studies will have to compare the detection sensitivity of *P. acnes* in granulomas using the different immunostaining methods. It is however important to state that the localisation and pattern of *P. acnes* within and outside granulomas was comparable with the study of Negi *et al.* [20]. Specifically, we observed that presence of *P. acnes* outside granulomas was most frequently detected directly adjacent to granulomas but also in granuloma-free parts of the tissue in a few cases. Furthermore, the pattern within granulomas was comparable, with more intense dot-like structures in immature granulomas and more sparsely distributed staining or even no staining in mature granulomas.

A new and clinically relevant finding, in our opinion, is the fact that this is the first study that shows an association between presence of *P. acnes* and clinical outcome in patients with sarcoidosis. We observed that presence of *P. acnes* in tissue and in granulomas is more frequently found in patients with a chronic disease course requiring chronic treatment. Several studies demonstrated the mitogenic properties of *P. acnes* on different immune cells [16, 24, 25]. Our study was not aimed to prove that *P. acnes* has a specific aetiological role in sarcoidosis; however, in our opinion, the results do suggest that at least a potential mitogenic role of *P. acnes* may contribute to sarcoidosis disease pathogenesis. It is tempting to speculate that presence of *P. acnes* in or around granulomas can enhance an ongoing inflammatory reaction in sarcoidosis, contributing to perpetuation of the inflammatory granulomatous response seen in some

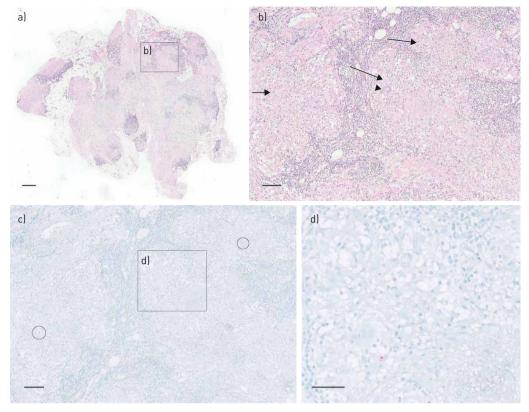


FIGURE 2 Presence of *Propionibacterium acnes* in a mature granuloma in s.c. tissue of the lower eyelid. a) Overview of haematoxylin and eosin (H&E) staining of s.c. tissue of the lower eyelid. b) Higher magnification of area b from picture a, including mature granulomas (arrows) and multinucleated giant cells (arrow head). c) *P. acnes*-specific monoclonal antibody (PAB) staining of corresponding area of b, including several areas of *P. acnes*-positive signals (circles and rectangle d), in mature granulomas. d) Higher magnification of area d from picture c, with positive *P. acnes* staining in a mature granuloma. Scale bars=500 μ m (a), 100 μ m (b, c) and 50 μ m (d)

patients. If this holds true, it is interesting to see whether decreasing the bacterial load of *P. acnes* using antibiotics can be beneficial in a subgroup of patients with sarcoidosis.

A case report already described a good effect of clarithromycin on fever, joint pain, 2-fluoro-2-deoxy-D-glucose uptake on positron emission tomography/computed tomography, C-reactive protein and s-interleukin-2R levels in a sarcoidosis patient in whom *P. acnes* was present in granulomas [26]. Moreover, a retrospective study described a good response on treatment with minocycline in *P. acnes*-positive cutaneous sarcoidosis patients [27]. When we examined severe therapy-refractory patients

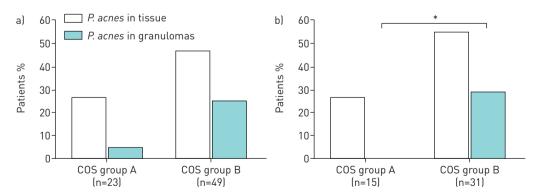


FIGURE 3 Frequency of *Propionibacterium acnes* detected in tissue samples from sarcoidosis patients with a different clinical outcome status (COS) 2 years (a) and 5 years (b) after diagnosis. COS group A indicates resolved, minimal or persistent disease without treatment (COS 1–6). COS group B indicates persistent disease with need for treatment (COS 7–9). PAB: *P. acnes*-specific monoclonal antibodies that react with cell membrane-bound lipoteichoic acid.

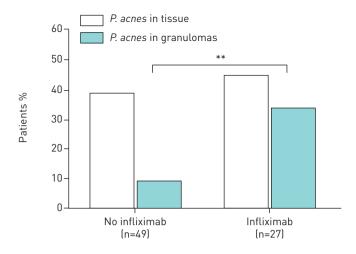


FIGURE 4 Frequency of *Propionibacterium acnes* detected in biopsy samples from sarcoidosis patients with or without third-line therapy (infliximab) during follow-up. A significantly higher percentage of third-line therapy-requiring patients (all using infliximab) showed presence of *P. acnes* in granulomas than patients who did not require third-line therapy (p=0.009). PAB: *P. acnes*-specific monoclonal anti-bodies that react with cell membrane-bound lipoteichoic acid.

for the presence of *P. acnes*, we observed that patients requiring infliximab treatment at follow-up, often had more presence of *P. acnes* in granulomas at diagnosis compared to patients without treatment or treated with first- or second-line treatment. If antibacterial treatment in sarcoidosis patients with presence of *P. acnes* in tissue and granulomas is beneficial, the requirement of third-line treatments (*e.g.* infliximab) may be prevented.

Currently, in a randomised controlled clinical trial (J-ACNES), the effect of antibacterial drugs in addition to standard corticosteroid therapy in cardiac sarcoidosis patients is examined [28]. However, in this trial presence of *P. acnes* in myocardial tissue was not an inclusion criterion. Therefore, in future studies it would be interesting to have information on the presence or absence of *P. acnes* in these patients, while investigating the effect of antibiotic therapy.

TABLE 2 Detection of <i>Propionibacterium acnes</i> and origin of tissue used								
	n	Detection	Detection of <i>P. acnes</i> in:					
		Tissue	Granulomas					
Total patients#	76	31 (41)	13 (17)					
Total tissue sections	80	32 (40)	14 (18)					
Lung	32	10 (31)	7 (22)					
VATS	7	3 (43)	3 (43)					
TBLB	25	7 (28)	4 (16)					
Lymph node	25	12 (48)	2 (8)					
Lymphadenectomy	14	9 (64)	1 (7)					
Mediastinoscopy	5	1 (20)	0					
EBUS-TBNA/needle	6	2 (33)	1 (17)					
Skin	17	7 (41)	4 (24)					
Other	6	3 (50)	1 (17)					
Bone marrow	2	2 (100)	0					
Liver	2	1 (50)	1 (50)					
Nasal concha	1	0	0					
Salivary gland	1	0	0					

Data are presented as n [%], unless otherwise stated. #: In 4 out of 76 patients, two tissue sections of different organs were stained namely: lymph node (negative) and lung (P. acnes-positive in tissue and granulomas), liver (negative) and lymph node (P. acnes-positive in tissue), skin (P. acnes-positive in tissue and granulomas) and lung (negative), skin (P. acnes-positive in tissue and granulomas) and lung (P. acnes-positive in tissue and granulomas). VATS: video-assisted thoracoscopic surgery; TBLB: transbronchial lung biopsy; EBUS-TBNA: endobronchial ultrasound-transbronchial needle aspiration.

TABLE 3 Presence of *Propionibacterium acnes* in tissue and granulomas related to organ involvement and Scadding stage

	PAB staining			PAB staining present in granulomas		
	Negative (n=45)	Positive (n=31)	p-value#	No (n=63)	Yes (n=13)	p-value#
Age [¶] years	44.59±12.51	43.08±11.94	0.601	44.76±12.17	40.18±12.26	0.222
Male sex	23 (51)	19 (61)	0.380	33 (52)	9 (69)	0.266
White	38 (84)	26 (84)	1.000	52 (83)	12 (92)	0.679
Extrapulmonary	32 (71)	22 (71)	0.989	46 (73)	8 (62)	0.504
involvement						
Skin	12 (27)	6 (19)	0.461	15 (24)	3 (23)	1.000
Eyes	4 (9)	3 (10)	1.000	6 (10)	1 (8)	1.000
Liver	2 (4)	6 (19)	0.057	6 (10)	2 (15)	0.619
Heart	8 (18)	5 (16)	0.851	12 (19)	1 (8)	0.446
Spleen	2 (4)	1 (3)	1.000	2 (3)	1 (8)	0.435
Bones	3 (7)	1 (3)	0.641	4 (6)	0	1.000
Nerve system	11 (24)	4 (13)	0.214	14 (22)	1 (8)	0.444
Central	5 (11)	1 (3)	0.391	6 (10)	0	0.582
Peripheral	1 (2)	0	1.000	1 (2)	0	1.000
SFN	7 (16)	3 (10)	0.514	9 (14)	1 (8)	1.000
Scadding stage at	(N=44), 3/13/	(N=29), 0/10/	0.585	(N=61), 3/19/	(N=12), 0/4/	0.277
diagnosis (0/I/	19/5/4 (7/30/	13/2/4 (0/35/		29/5/5 (5/31/	3/2/3 (0/33/	
II/III/IV)	43/11/9)	45/7/14)		48/8/8)	25/17/25)	
Scadding stage	(N=39), 11/4/	(N=24), 6/3/	0.794	(N=53), 14/6/	(N=10), 3/1/	0.408
after 2 years (0/	12/5/7 (28/	8/1/6 (25/13/		19/5/9 (26/	1/1/4 (30/10/	
I/II/III/IV)	10/31/13/18)	33/4/25)		11/36/9/17)	10/10/40)	
Scadding stage	(N=25) 5/2/4/	(N=16) 3/1/4/	0.878	(N=34), 7/2/	(N=7), 1/1/0/	0.441
after 5 years (0/	6/8 (20/8/16/	2/6 (19/6/25/		8/7/10 (21/6/	1/4 14/14/0/	
I/II/III/IV)	24/32)	13/38)		24/21/29)	14/57	

Data are presented as mean±sD or n [%], unless otherwise stated. Chest radiographs to determine Scadding stages were missing from three sarcoidosis patients at diagnosis, from 13 patients 2 years after diagnosis and from 35 patients 5 years after diagnosis. Scadding stages: 0: Normal chest radiograph; I: BHL; II: BHL with pulmonary infiltrates; III: pulmonary infiltrates without BHL; IV: fibrosis. SFN: Small fibre neuropathy, PAB: *P. acnes*-specific monoclonal antibodies that react with cell membrane-bound lipoteichoic acid; BHL: bilateral hilar lymphadenopathy. #: p-values regarding age was calculated using an independent samples t-test. Other p-values were calculated using a Fisher's exact test, except for sex (both p-values), extrapulmonary involvement in the *P. acnes* score group, skin, heart and nerve system involvement in the *P. acnes* score group and Scadding stages; ¶: Age at time of biopsy.

A limitation of the study was that the COS after 5 years could not be determined in all patients. However, as the proportion of patients within COS group A and B was quite similar after 2 and 5 years and also the results regarding association with *P. acnes* staining, we assume that this analysis on a smaller group of patients has not introduced a bias. A disadvantage of using COS is the fact that disease status and medication use have been retrospectively assessed. As we only scored whether medication was used or not, we have no information on patients who declined the use of medication while they actually needed it.

Another limitation is that we probably have a more severe patient group than other general hospitals, as the St Antonius Hospital is a national referral centre for interstitial lung disease and sarcoidosis. As a consequence, we had very few patients in the resolved and minimal disease COS groups. For this reason, we did not have enough patients in every group to adequately analyse whether disease status alone, irrespective of use of medication, was associated with presence of *P. acnes* in tissue and granulomas. However, instead of disease status we think that need for treatment could be of more value regarding the presence of *P. acnes*. If we want to further explore the use of antibacterial therapy, in our opinion, it is more relevant to focus on patients who actually need treatment.

In future studies, it would be interesting to add a second detection method, such as PCR, to the immunostaining. To accurately compare the results, development and use of a primer specific to LTA of the *P. acnes* bacteria would be valuable.

To conclude, this study confirms the presence of *P. acnes* in tissues and granulomas in 40% and 17%, respectively of Dutch patients with sarcoidosis. Interestingly, the presence of *P. acnes* inside granulomas of

Dutch sarcoidosis patients was associated with a chronic disease phenotype and requirement of treatment. Regardless of whether the role of *P. acnes* is antigenic or mitogenic, our data contribute to the premise that it is relevant to further explore antibacterial therapy as a treatment option for a subset of sarcoidosis patients.

Author contributions: E. Beijer contributed to data acquisition and analysis, statistical analysis, and drafting and editing of the manuscript. K. Seldenrijk participated in the design of the study, scored the staining results and edited the manuscript. Y. Eishi developed the PAB antibody, advised on study design and edited the manuscript. K. Uchida reviewed the methods and advised on the automated staining protocol. J. Damen carried out all the immunohistochemical staining and developed the automated staining protocol. J.C. Grutters supervised the project and edited the manuscript. M. Veltcamp designed the study, supervised the project and edited the manuscript.

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Conflict of interest: None declared.

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