

May 8th, 9:00 AM - 11:00 AM

The Influence of a Ubiquitous Filter Feeder on Coastal Microbial Communities.

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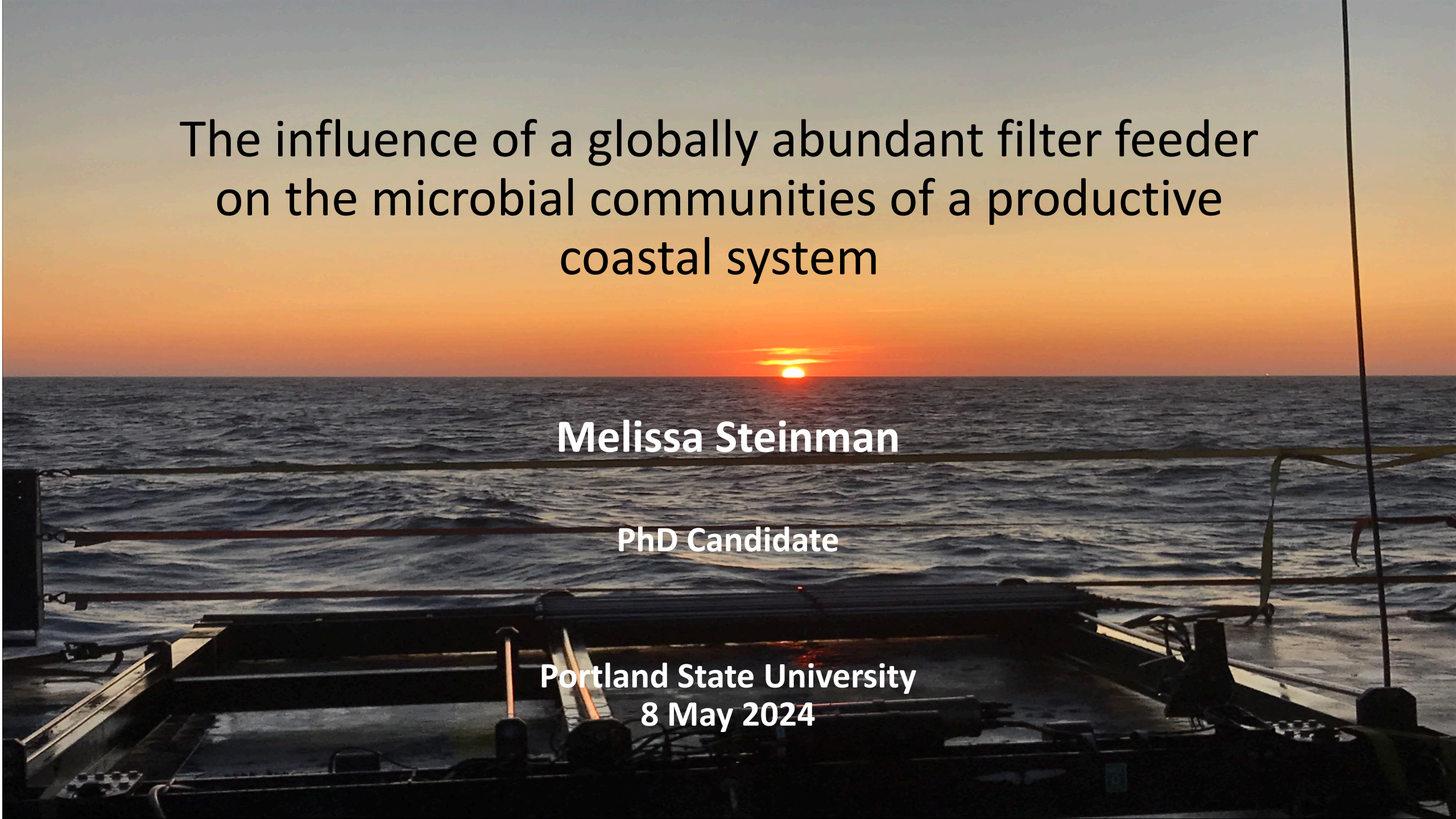
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Presenter Information

Melissa Steinman, Moritz S. Schmid, Robert K. Cowen, Su Sponaugle, Kelly R. Sutherland, and Anne W. Thompson

A photograph of a sunset over the ocean, taken from the deck of a ship. The sun is low on the horizon, creating a bright orange and yellow glow in the sky. The water is dark blue with small waves. In the foreground, the ship's deck and railings are visible, silhouetted against the bright sky.

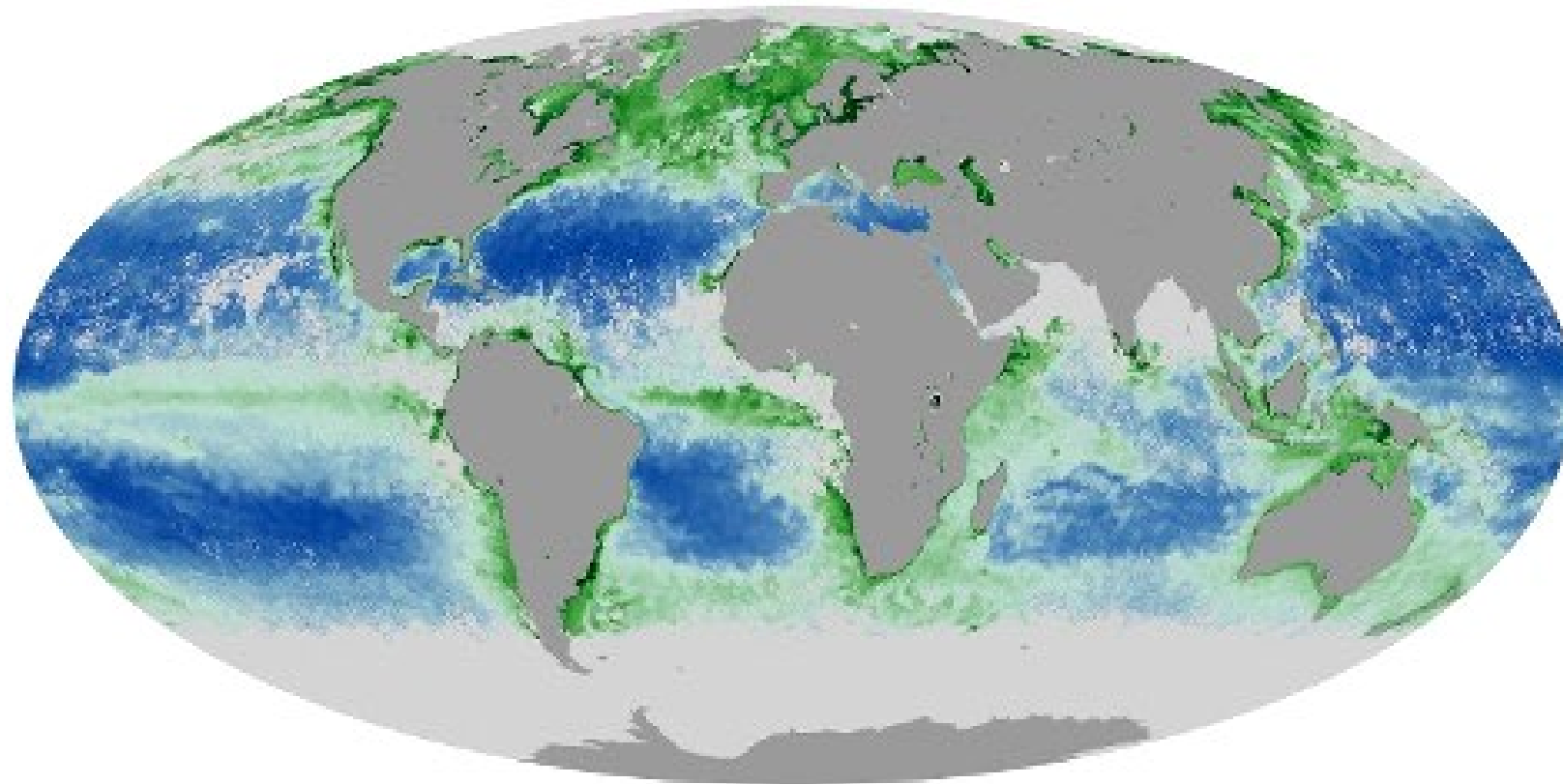
The influence of a globally abundant filter feeder on the microbial communities of a productive coastal system

Melissa Steinman

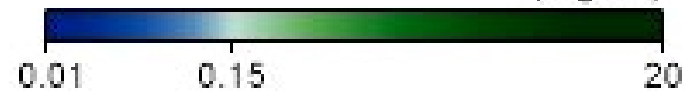
PhD Candidate

**Portland State University
8 May 2024**

Photosynthetic microbes have a global impact

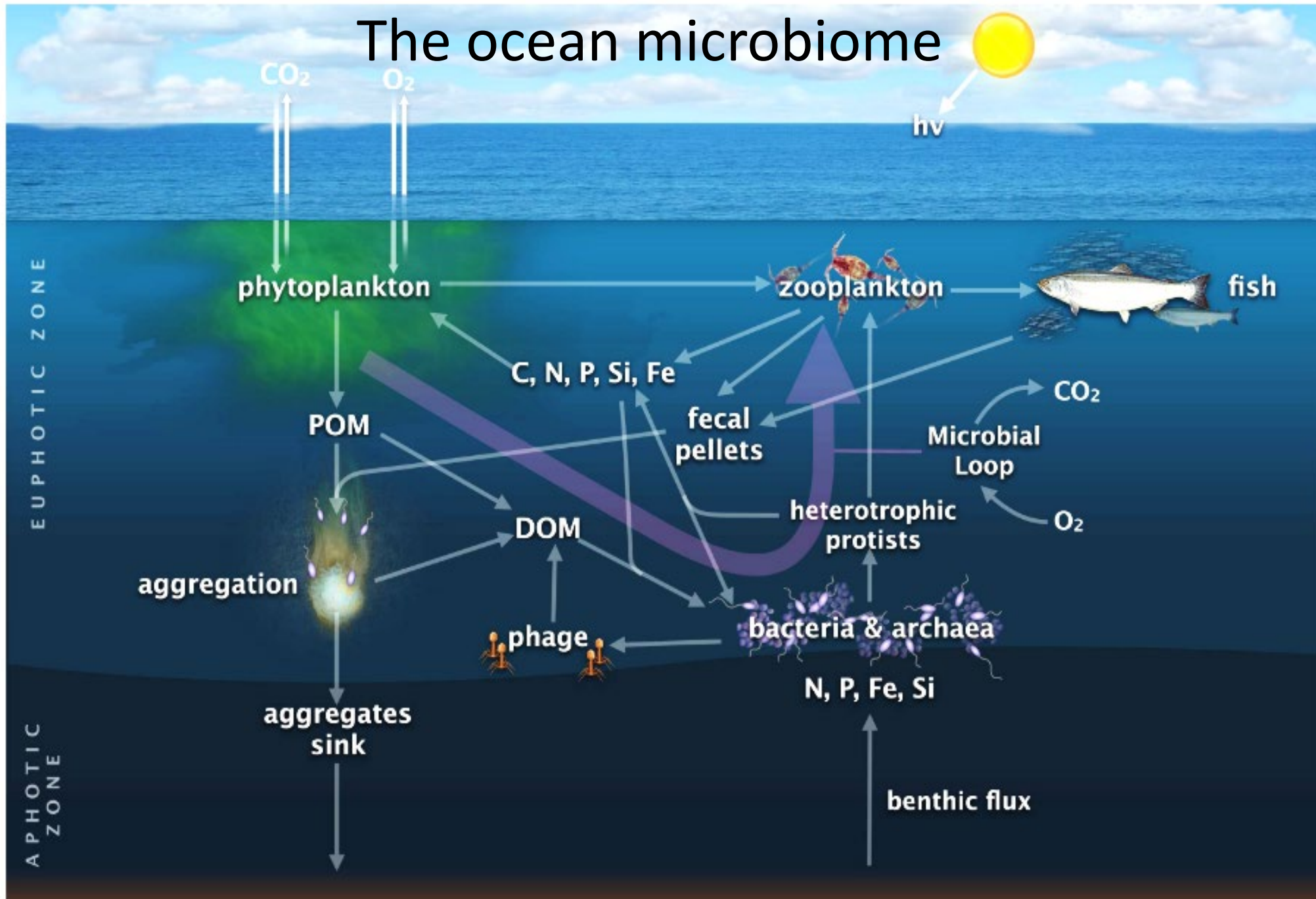


Chlorophyll Concentration
(mg/m³)

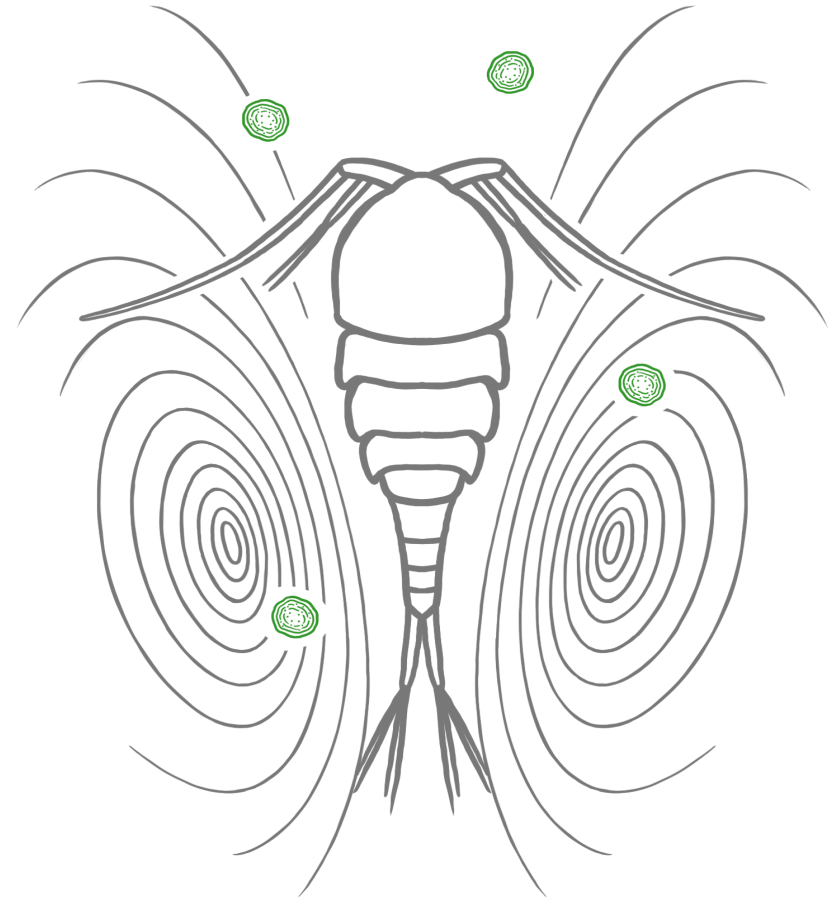
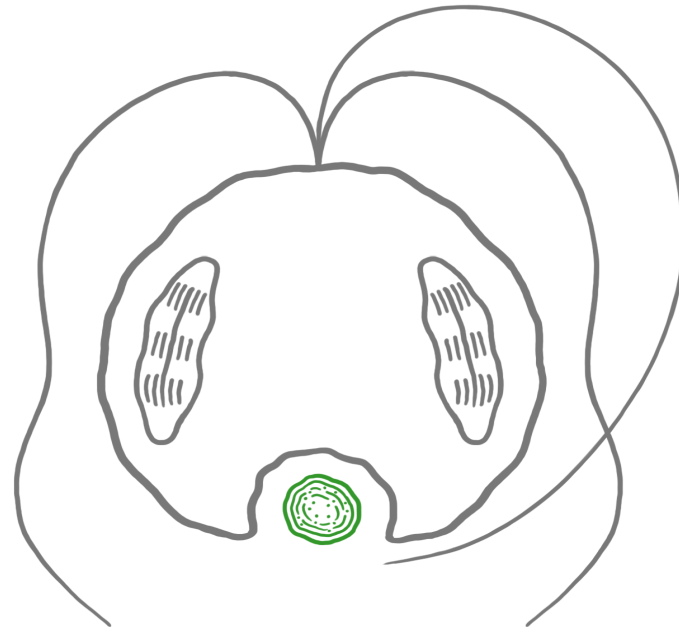
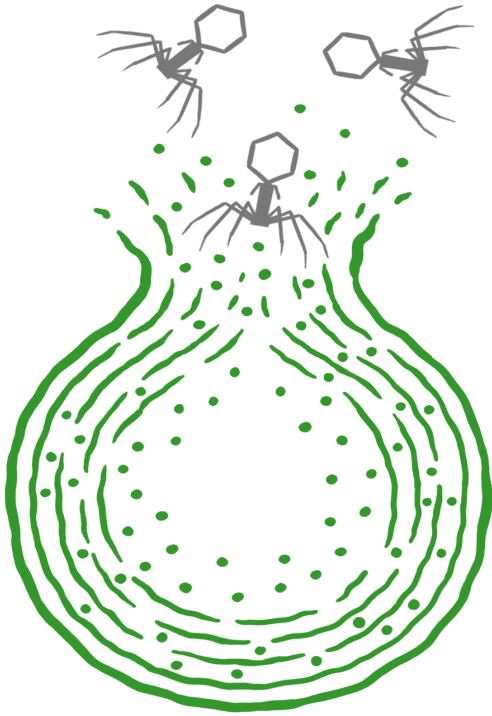


July 2002

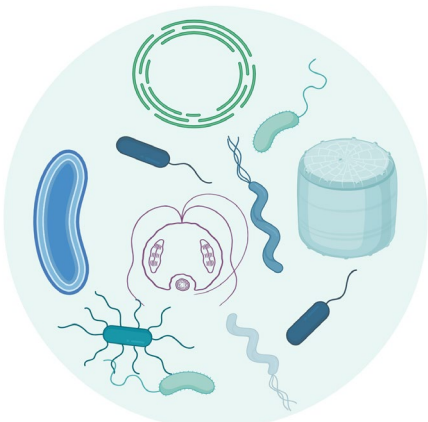
The ocean microbiome



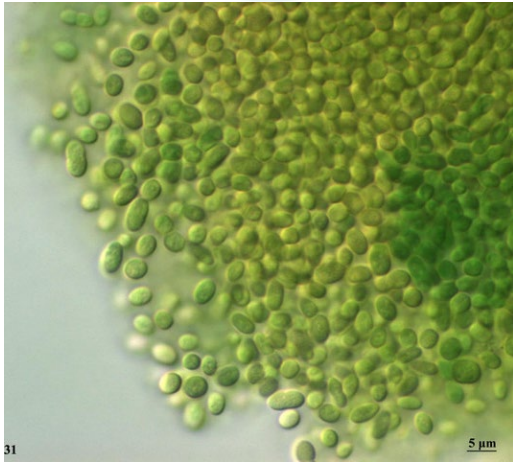
Microbial mortality is not fully understood



26% of microbial mortality is unaccounted for



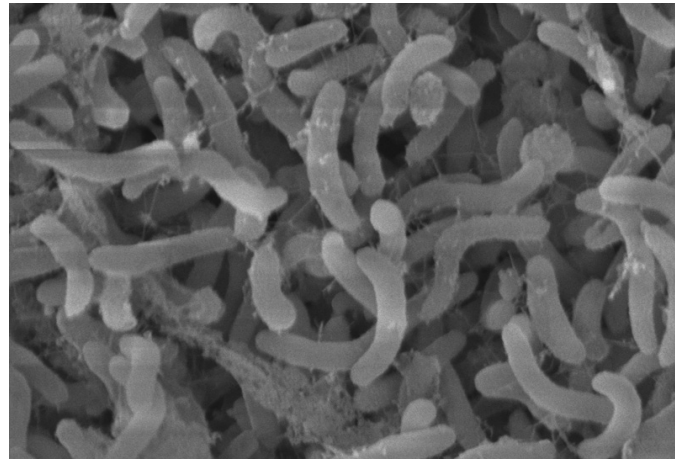
Dominant oceanic microorganisms



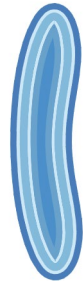
Synechococcus



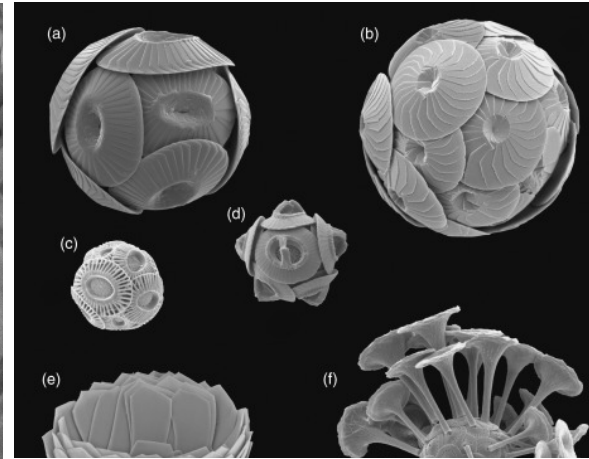
Critical to global oxygen production



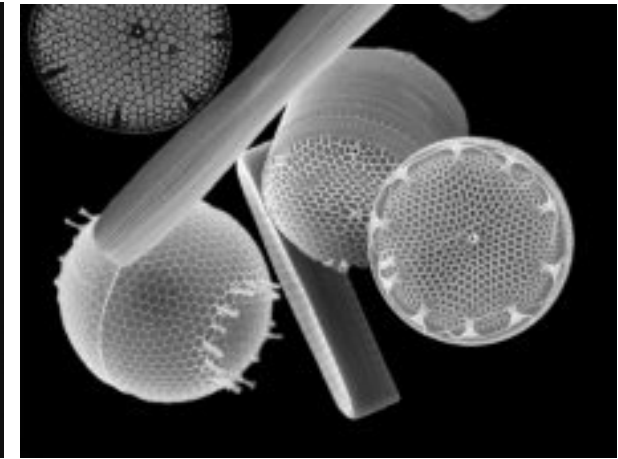
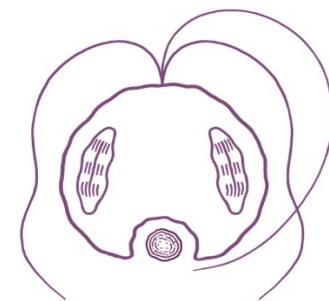
SAR11



Most abundant cell in the ocean



Prymnesiophytes

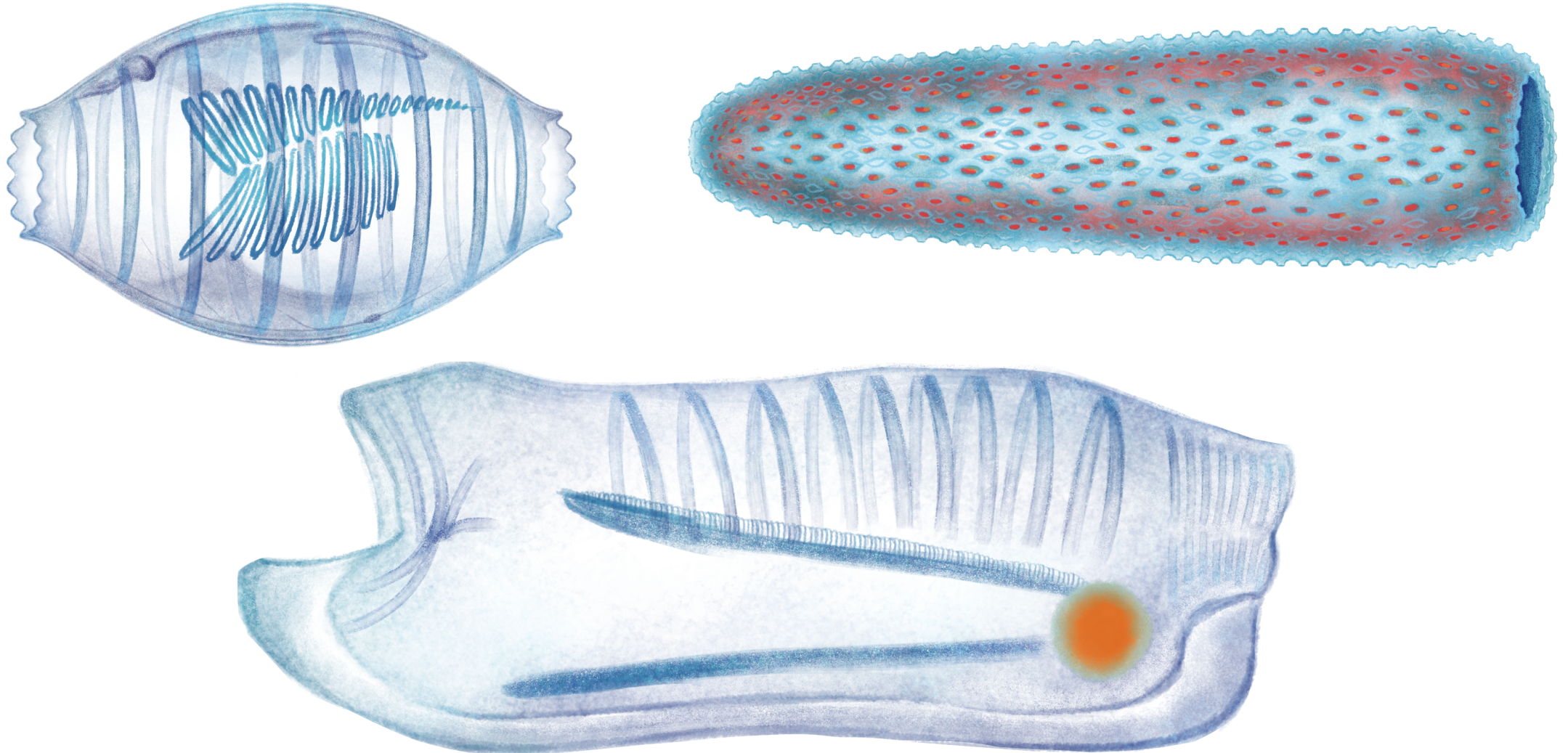


Diatoms

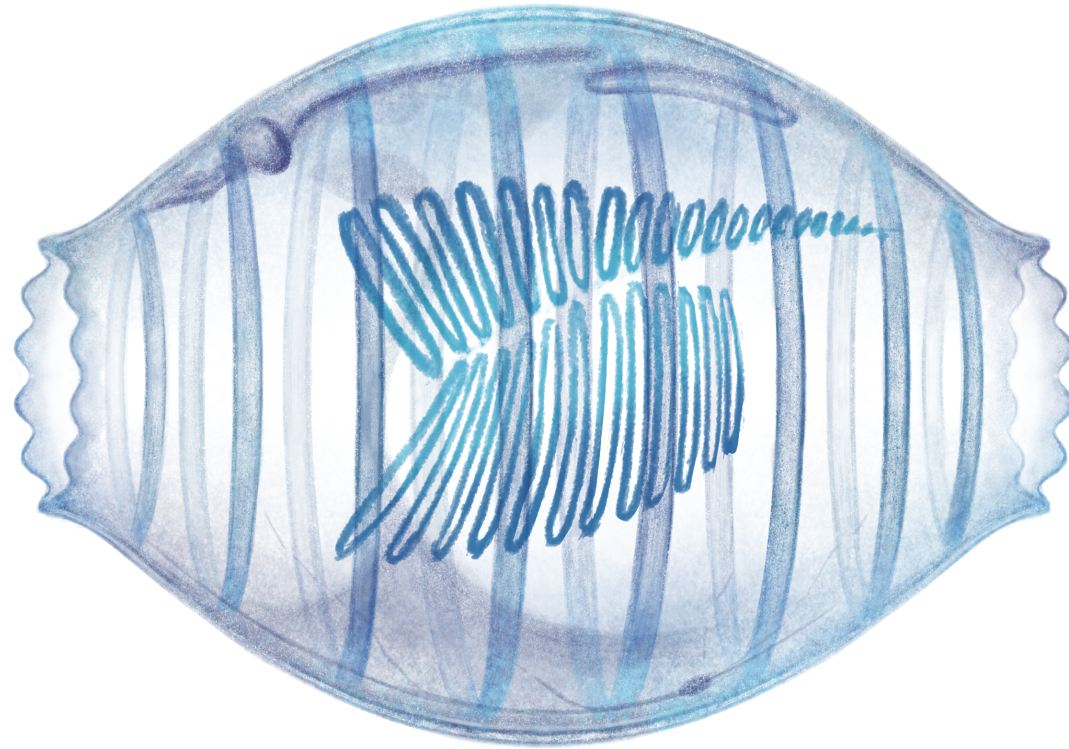


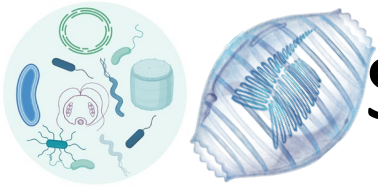
Maximum size

Thaliaceans – powerful but poorly understood



How do doliolids impact the microbial community?





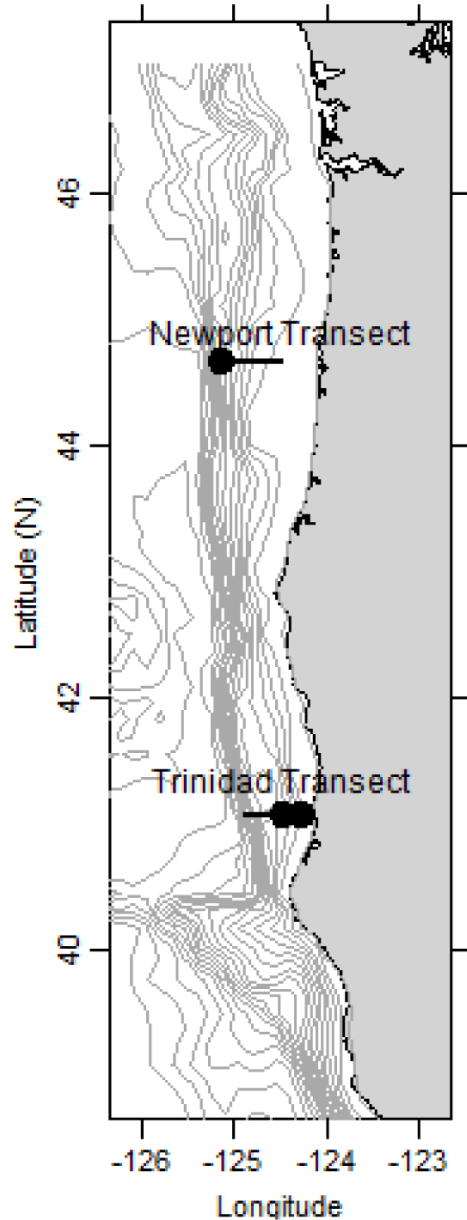
Sampling in the California Current System

Stations:

- On-shelf
- Shelf Break
- Off-Shelf

Transects:

- Newport Hydrographic Line (NH)
- Trinidad Head Transect (TR)



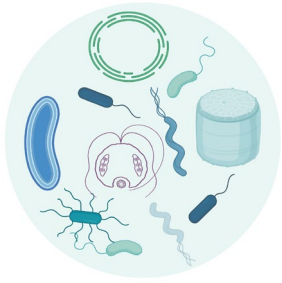
Su Sponaugle
Oregon State University
Plankton Ecology Lab



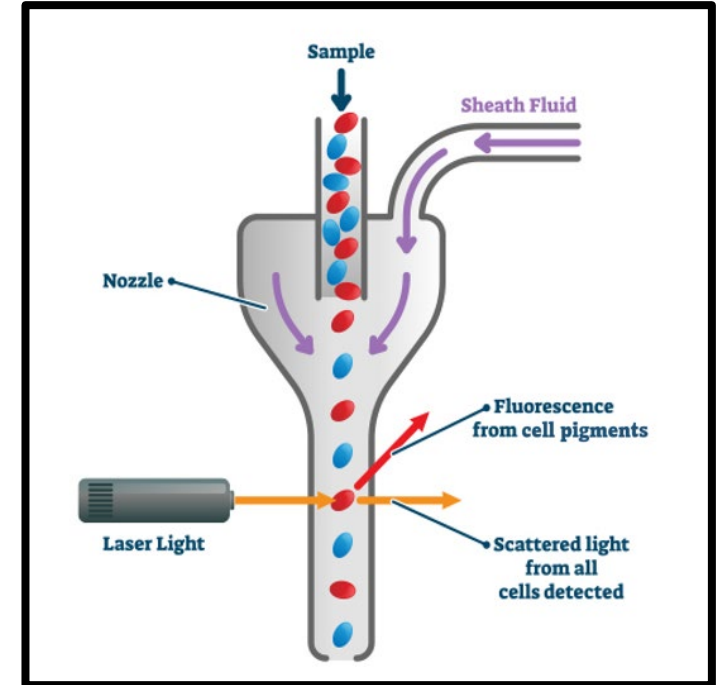
Robert Cowen
Oregon State University
Hatfield Marine Science Center



Kelly Sutherland
University of Oregon
Form, Function, Flow of Plankton
<https://www.sutherlandlab.org/>

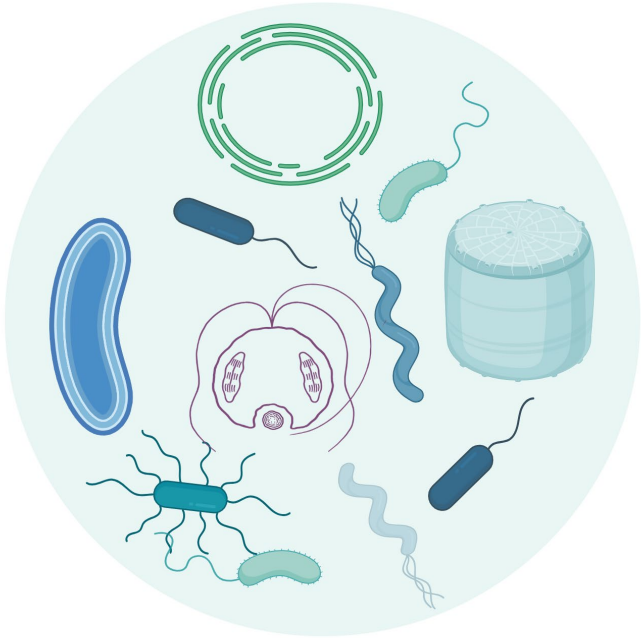


Sampling in the California Current System



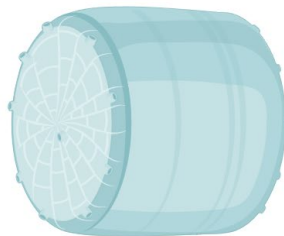
- Size Fractions
- Four Depths
- Fixed for Flow Cytometry

Microbial identification



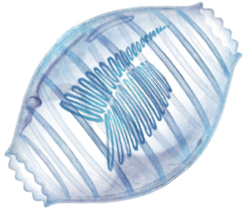
16S rRNA gene sequencing & identification of ASVs

- Identify amplicon sequence variants using *dada2*
- Taxonomically identify microbial sequences using BLASTn (NCBI)
- Relative abundances using R
- Hypothetical functional roles
 - Free-living (prey), potential symbionts/pathogens/neutral
 - Comparison to published functionality

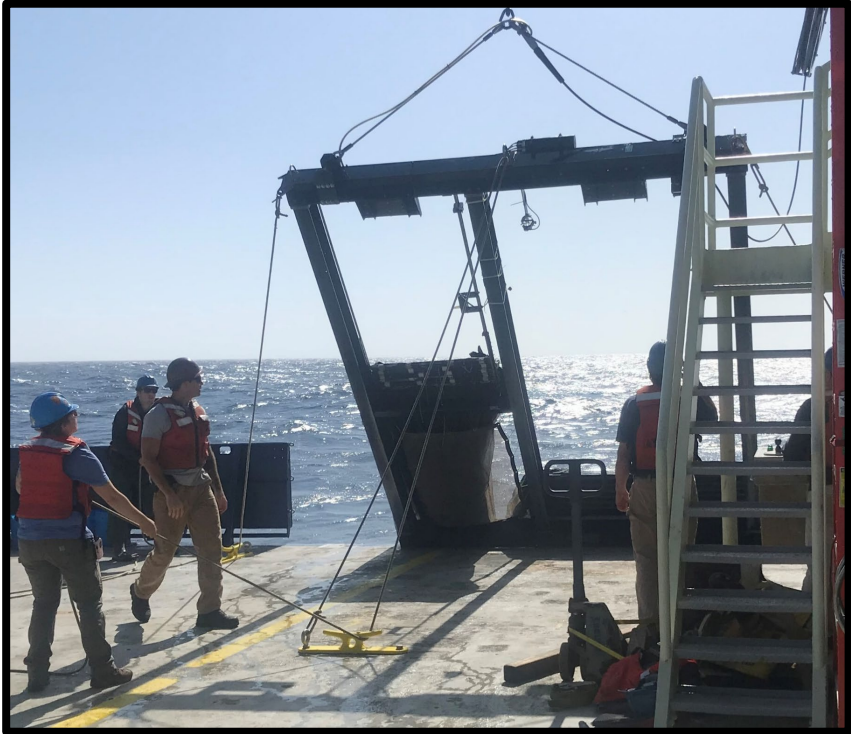


qPCR

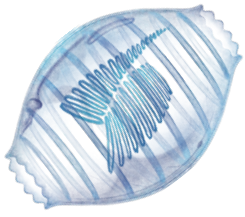
- Quantify specific taxa



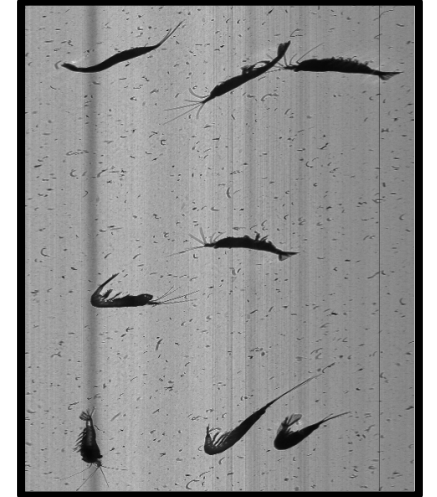
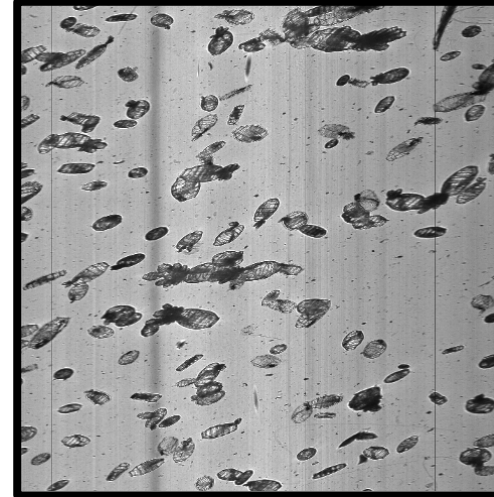
Sampling in the California Current System



- Multiple Opening Closing Net Environmental Sampling System (MOCNESS)
- Coupled asymmetrical net
- Minimizes damage to gentle bodies



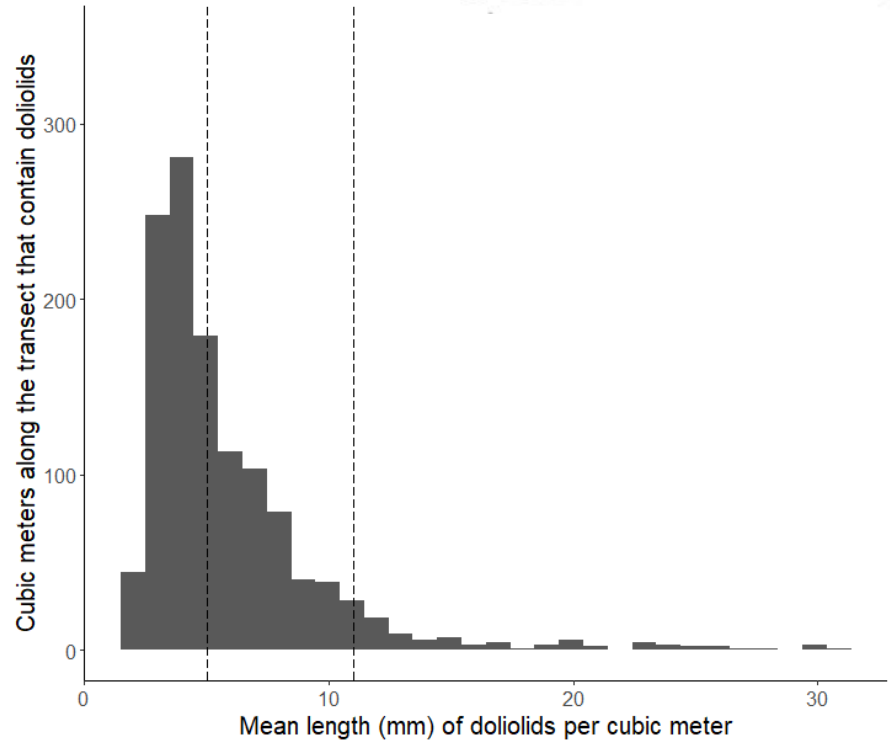
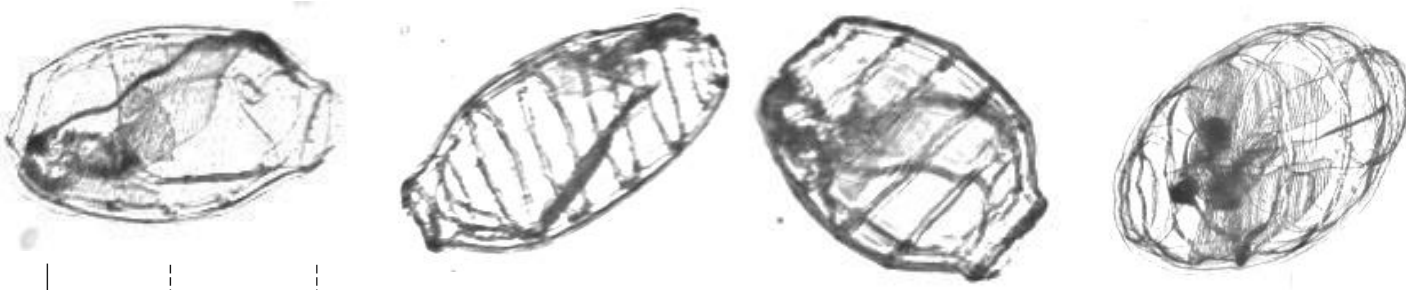
In Situ Ichthyoplankton Imaging System (ISIIS)



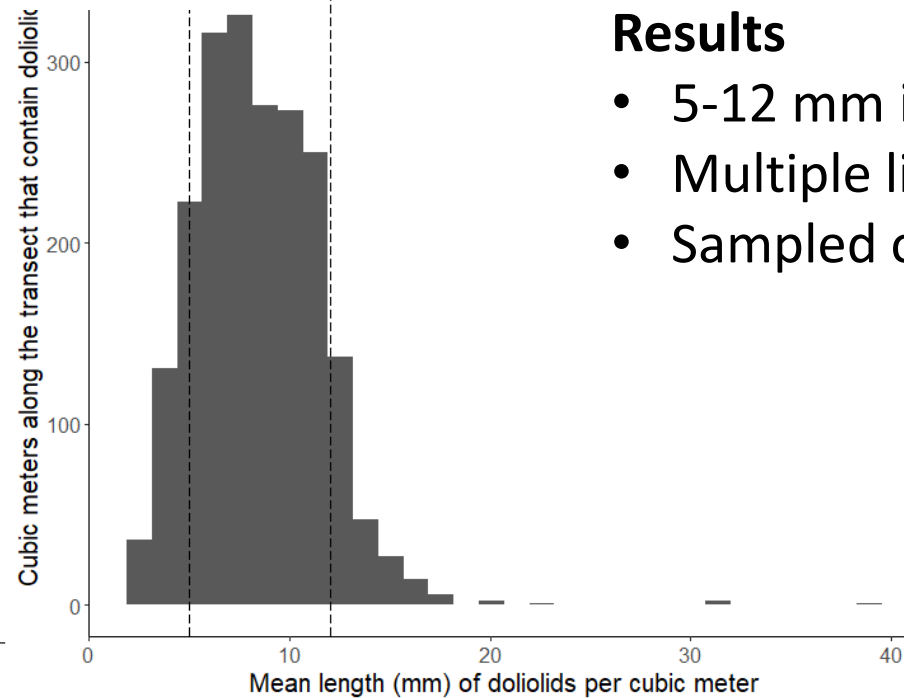
Moritz Schmid

Oregon Department of Fish and Wildlife
Oregon Marine Reserves

Doliolid identification and abundance



Newport Hydrographic (NH) Line Transect



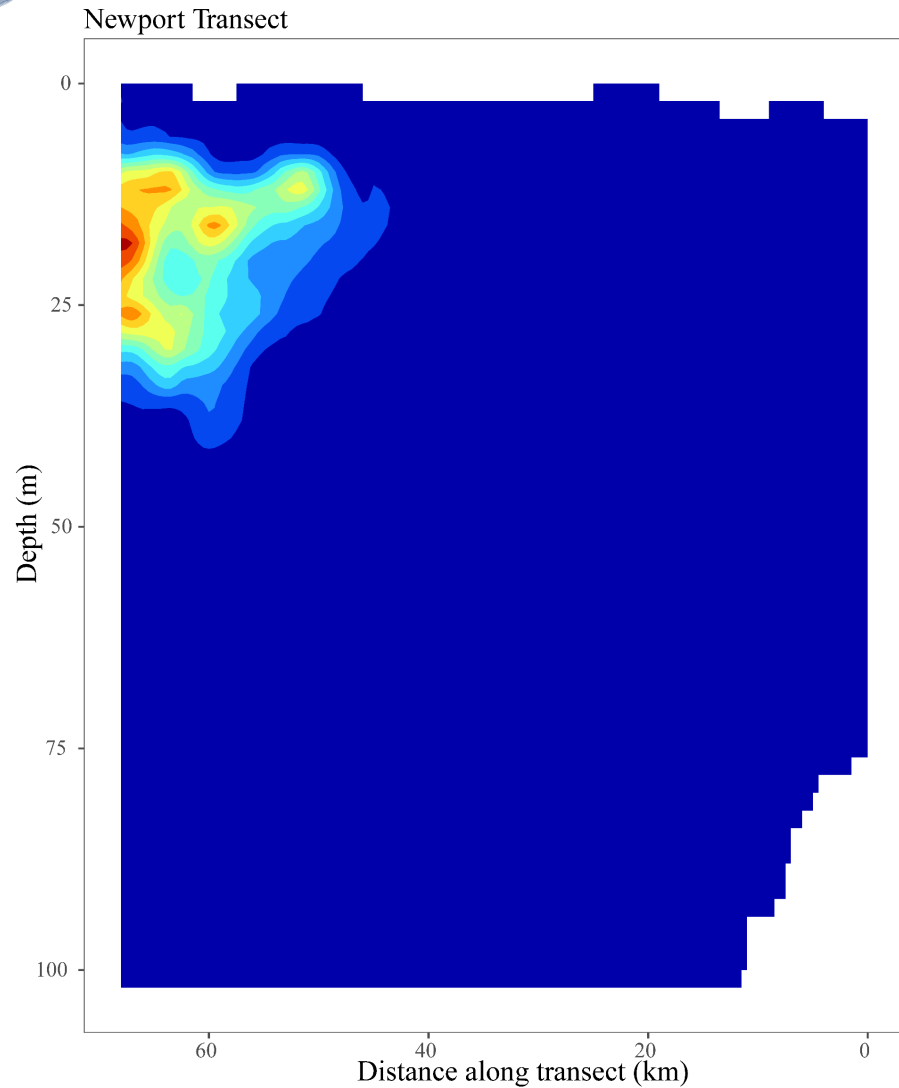
Trinidad Head (TR) Transect

Results

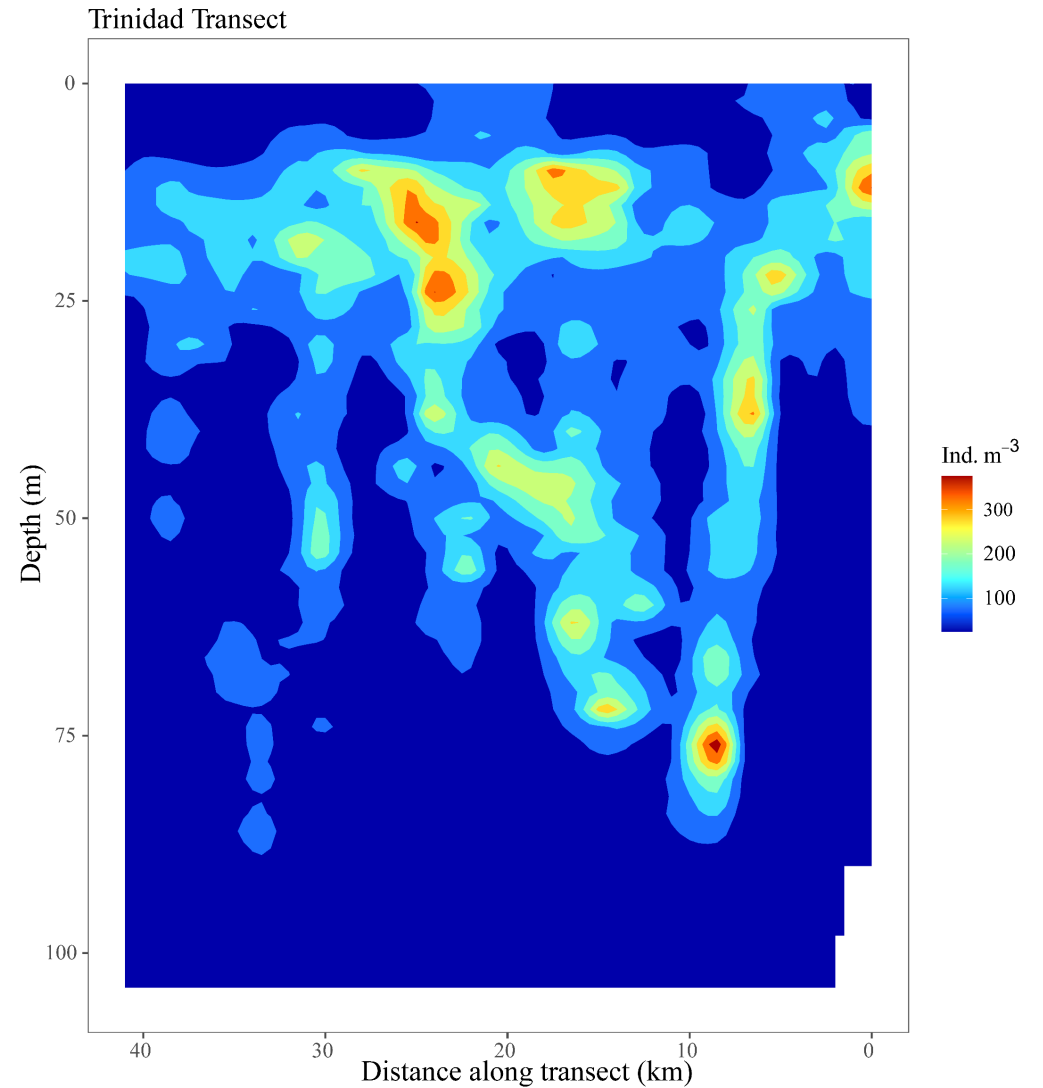
- 5-12 mm in size
- Multiple life stages on transect
- Sampled only gonozooid life stage



Sometimes very abundant, always patchy!

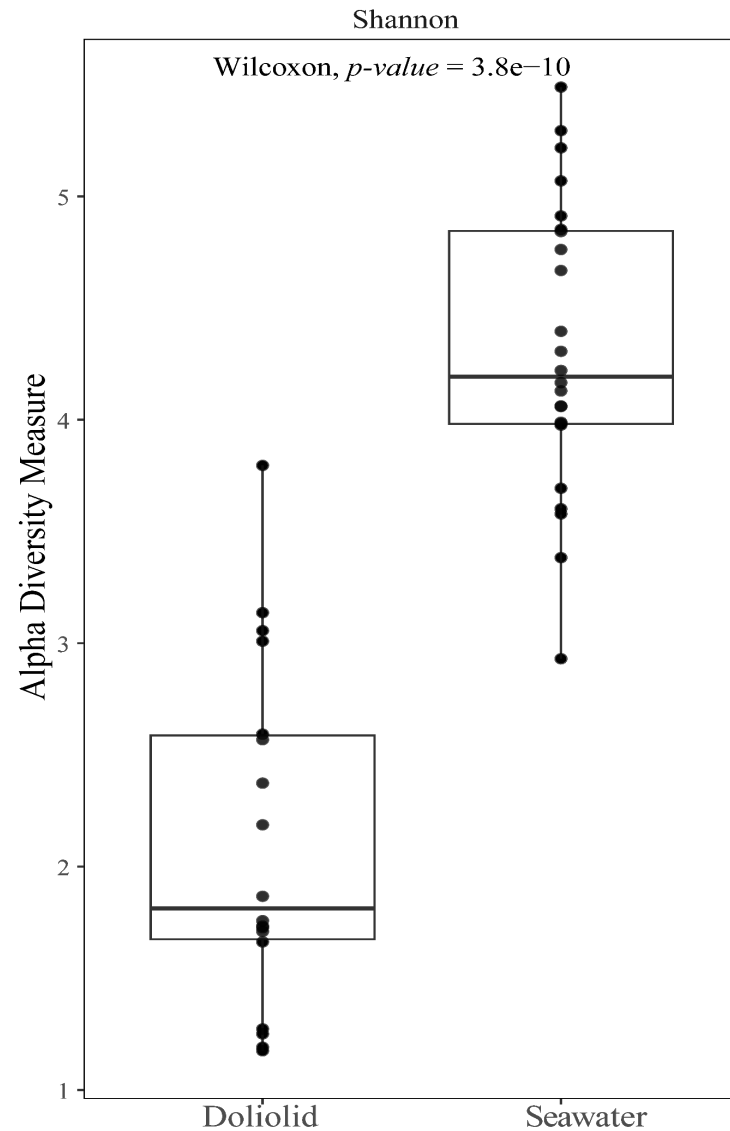


227 individuals per m^3



533 individuals per m^3

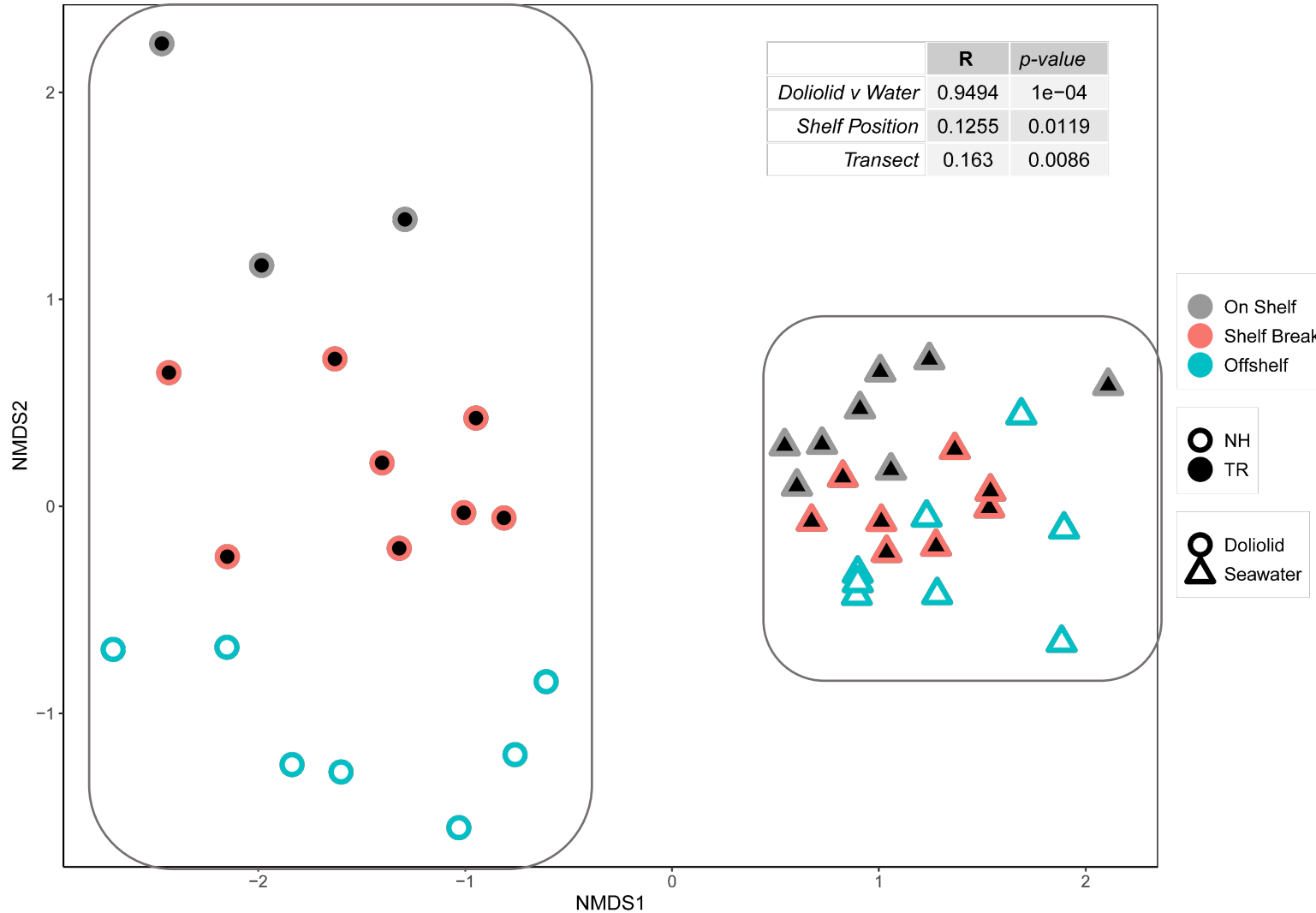
Seawater vs doliolid microbial diversity?



Results

- 6801 unique ASVs
- Doliolid diversity low
- Doliolid microbes are different than seawater
- Consistent with other microbiomes

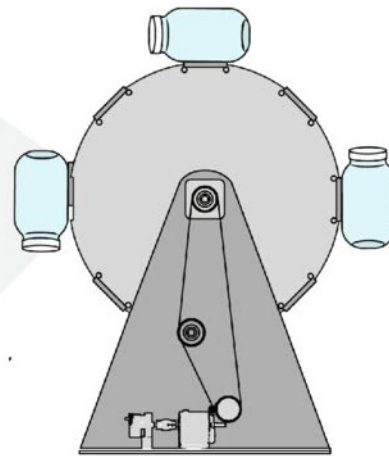
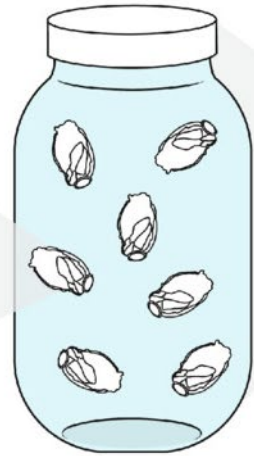
Seawater vs doliolid microbial diversity?



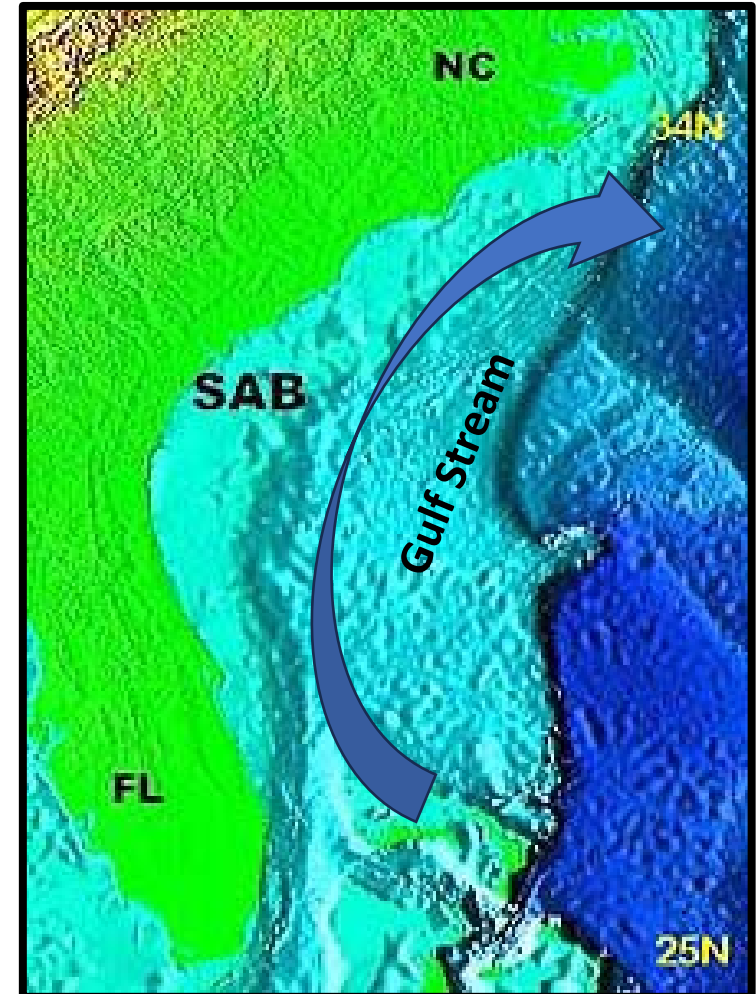
Results

- Microbial communities are distinct
- Differences exist in between locations
- Doliolid microbiome by location

Compare: Atlantic vs Pacific doliolid microbiomes?



Pereira et al., 2022

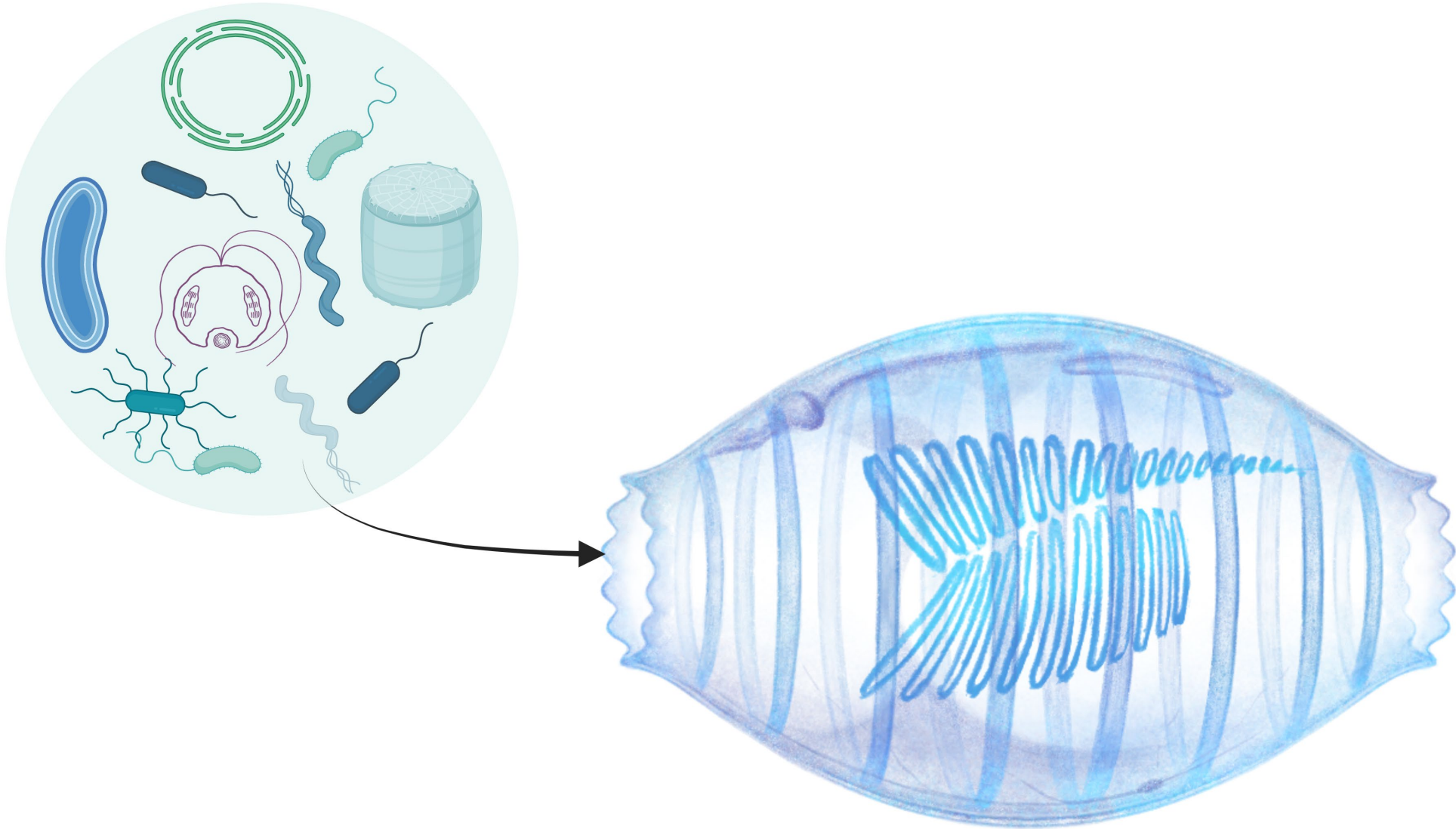


Noaa.gov

Chapter 2 Conclusions – doliolid microbiomes

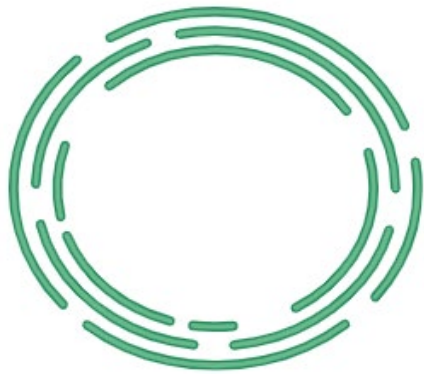
- Doliolids host a unique microbiome.
- Specific microorganisms not shared Atlantic vs Pacific.
- Four possible reasons:
 - Seawater microbial community?
 - Host genotype?
 - Host factors?
 - Body compartments?
- Hypotheses and targets generated:
 - Identify doliolid symbionts or pathogens?
 - Microbiome could be supporting unique doliolid survival strategies

Do doliolids shape the microbial community through feeding?

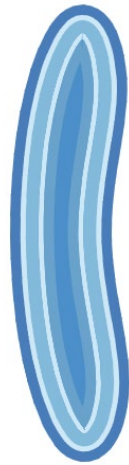


Known free-living microorganisms as potential prey

Synechococcus



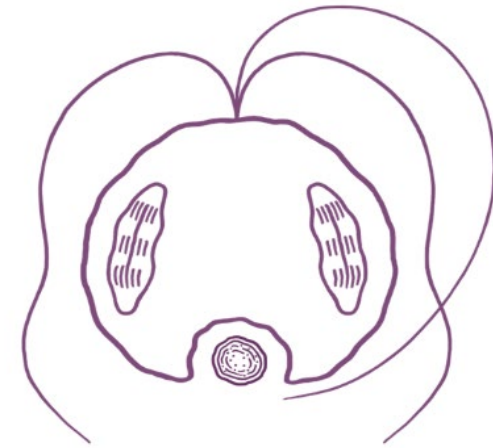
SAR11



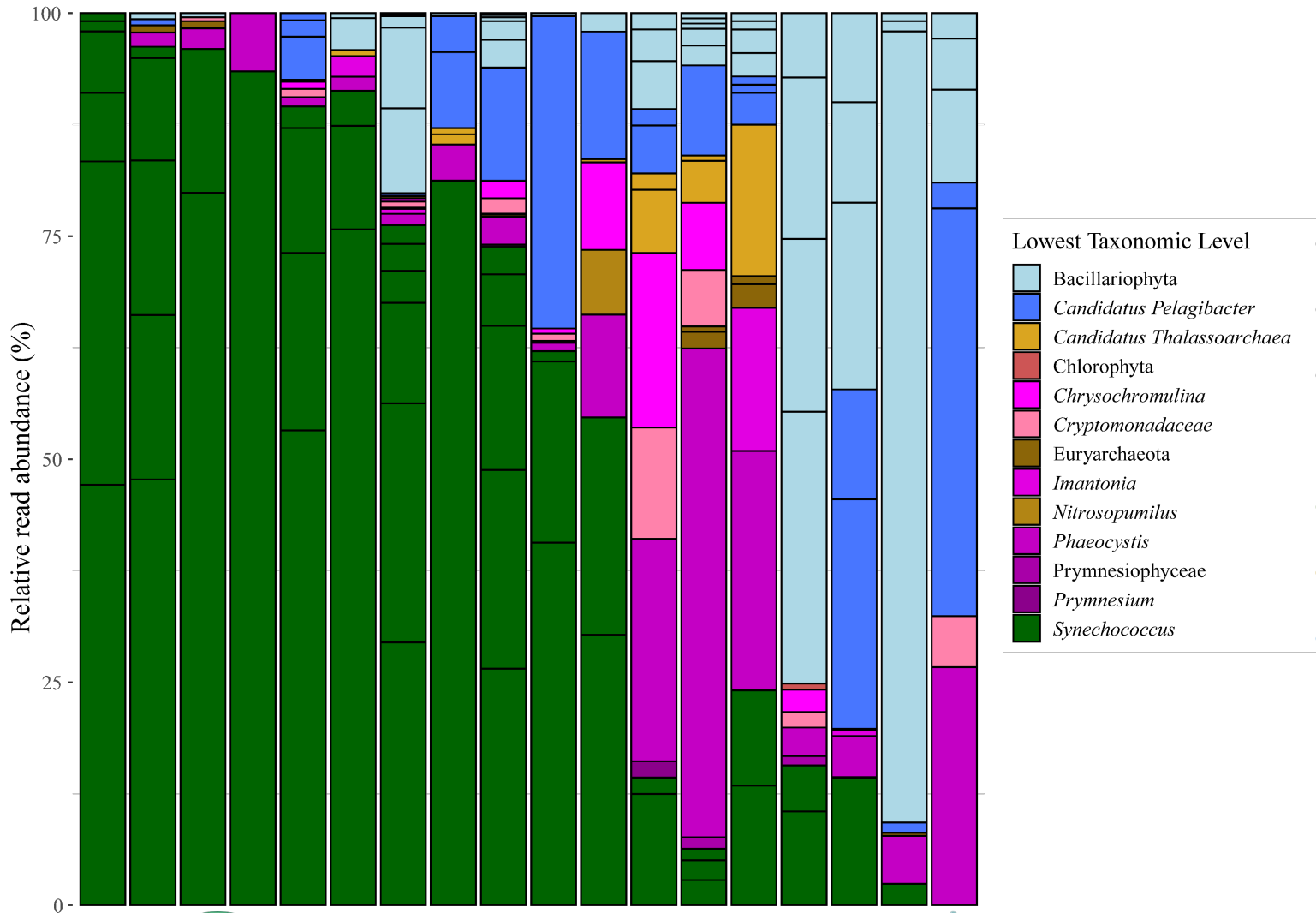
Diatoms



Prymnesiophytes



Which microbial prey are found in doliolids?



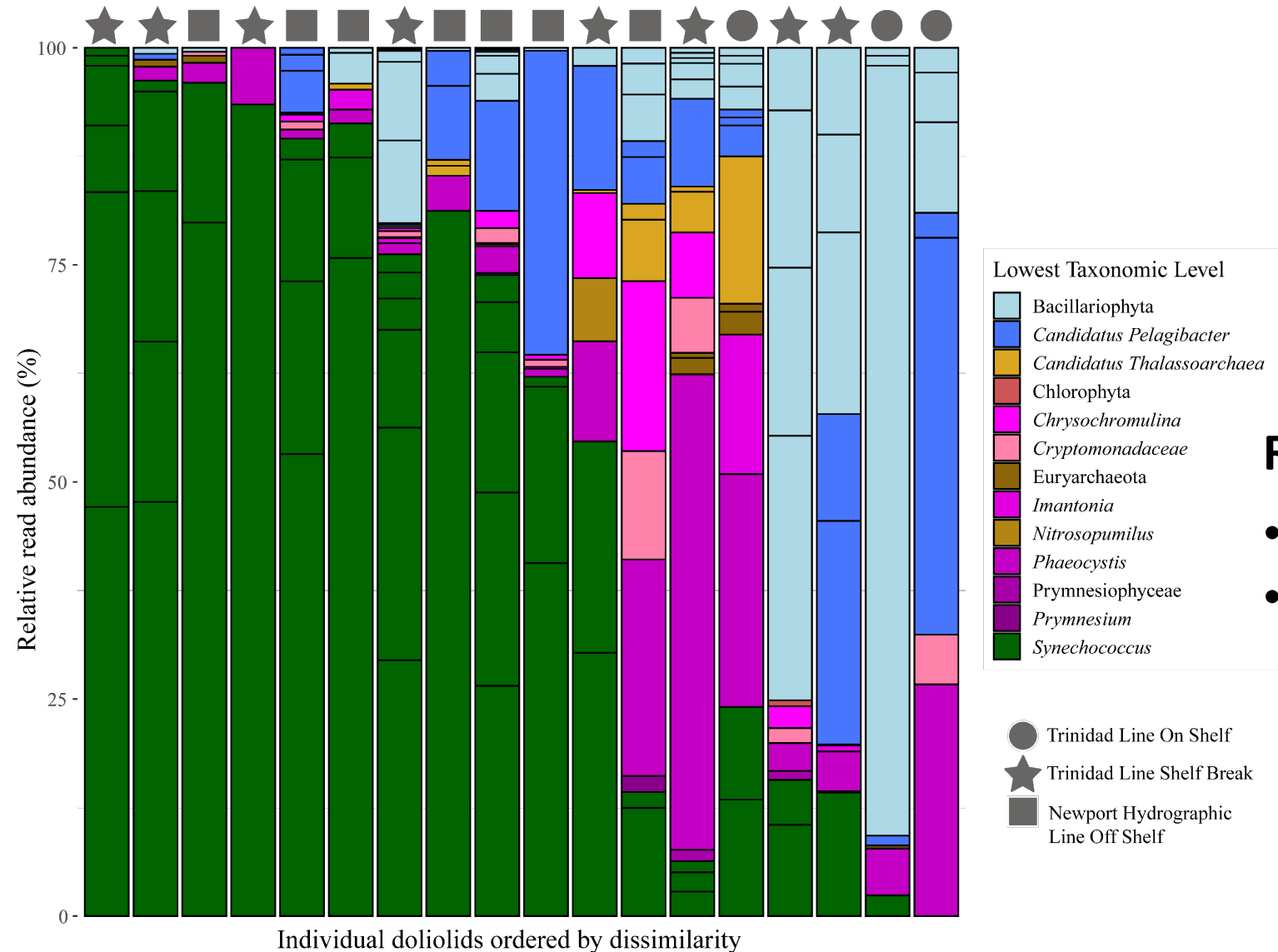
Individual doliolids ordered by dissimilarity



Results

- Effective predator of prokaryotes
- Many microbial functional groups
- Wide range of cell sizes
- *Synechococcus* is present
- Archaeal is present
- SAR11 is present

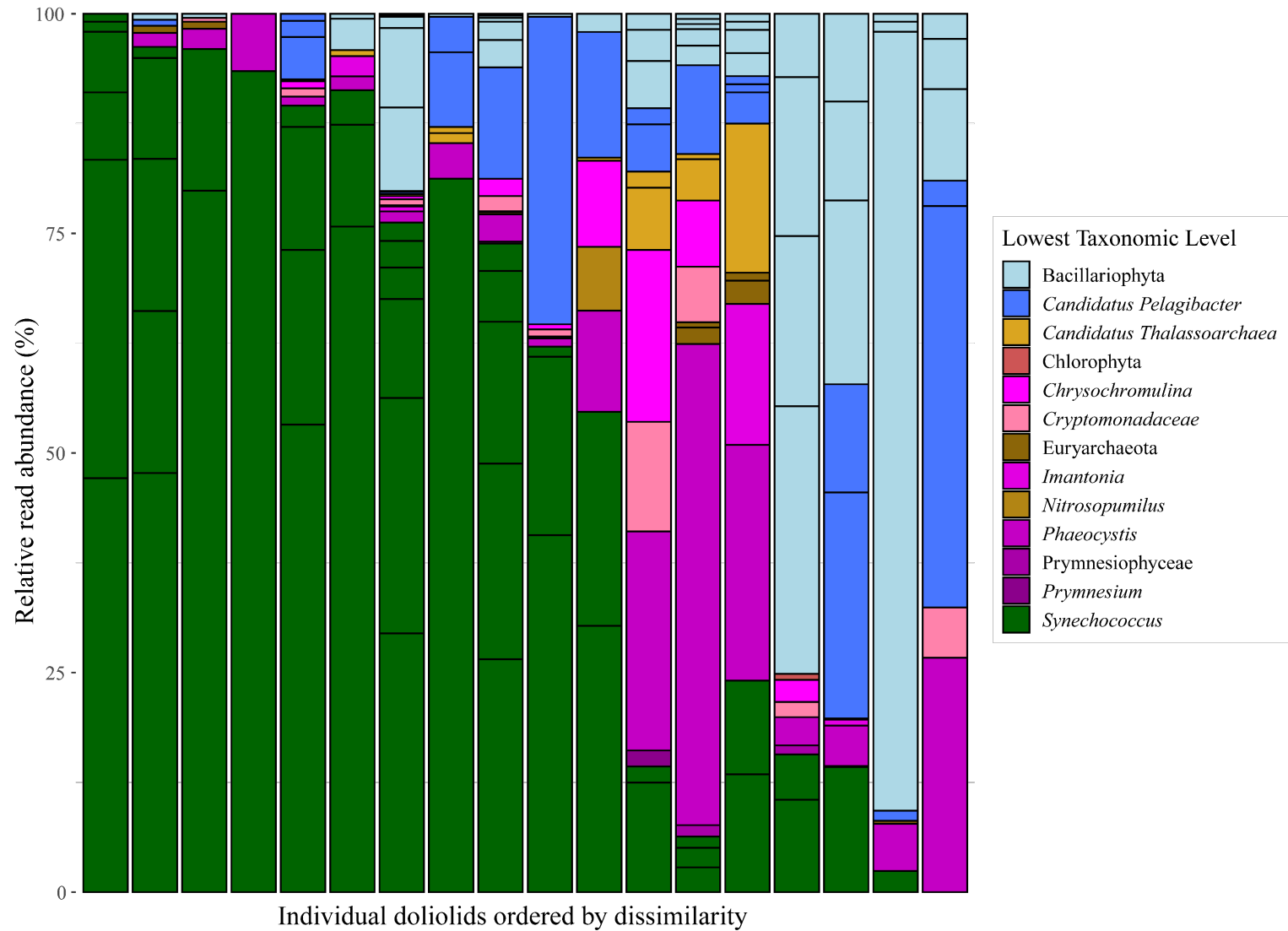
What is driving the differences in feeding between doliolids?



Results

- Moderately by shelf position
- Not impacted by transect

Which microbial prey are found in doliolids?

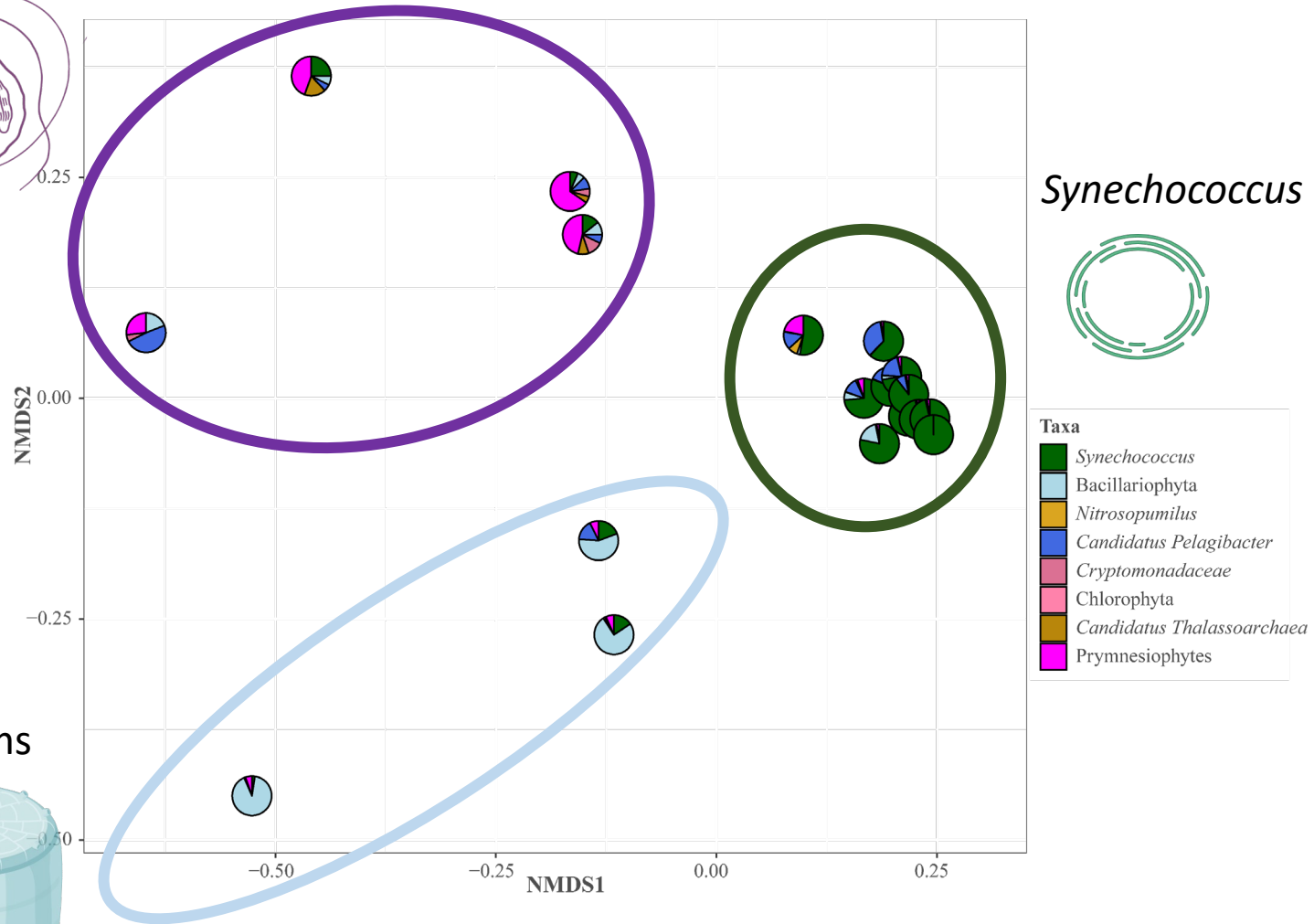


Results

- Individual differences

Feeding differences among doliolids?

Prymnesiophytes



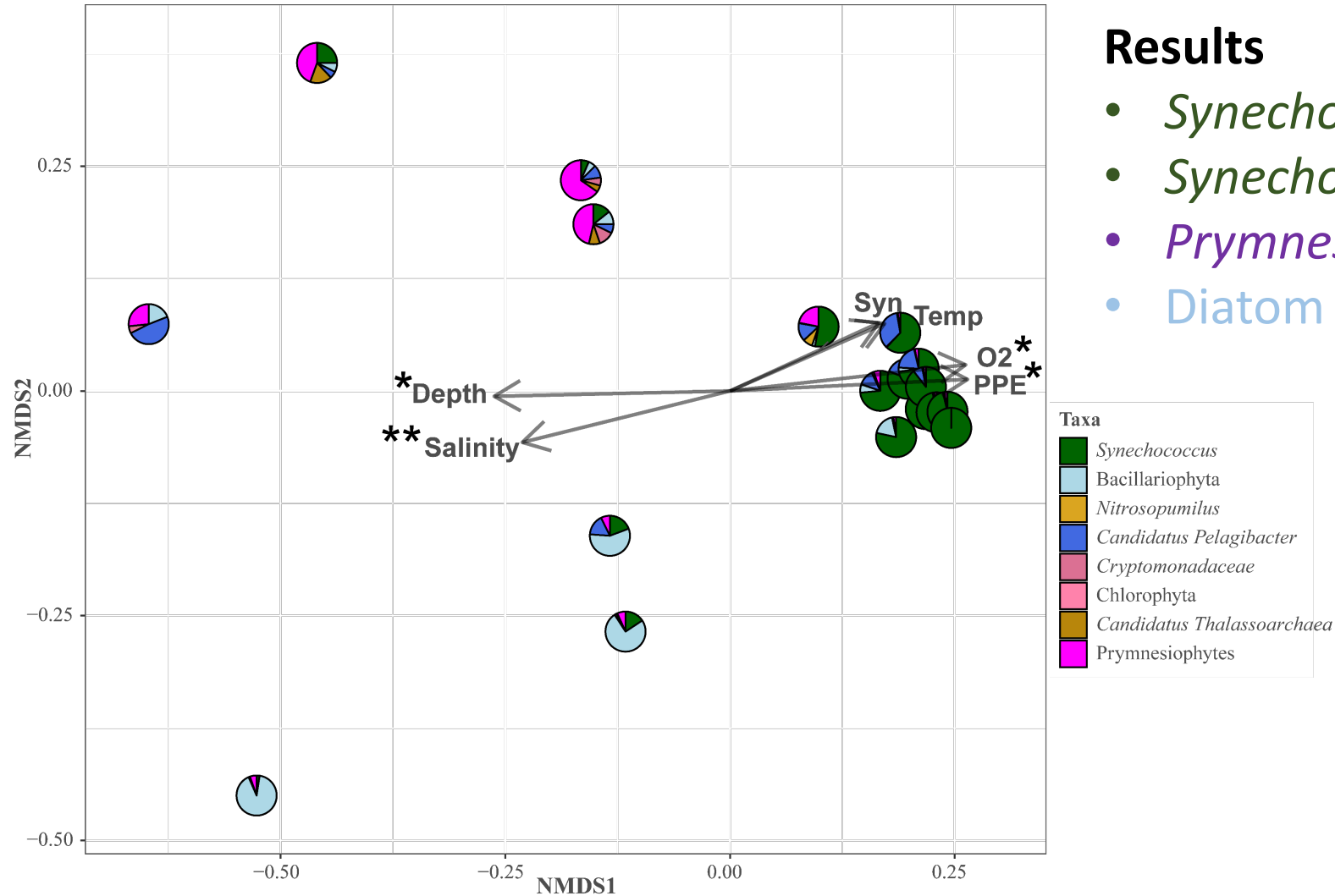
Diatoms



Results

- Three feeding groups

What is driving differences in feeding among doliolids?



Results

- *Synechococcus* group -> increased O₂
- *Synechococcus* group -> increased PPE
- *Prymnesiophytes* group -> depth
- *Diatom* group -> increased salinity

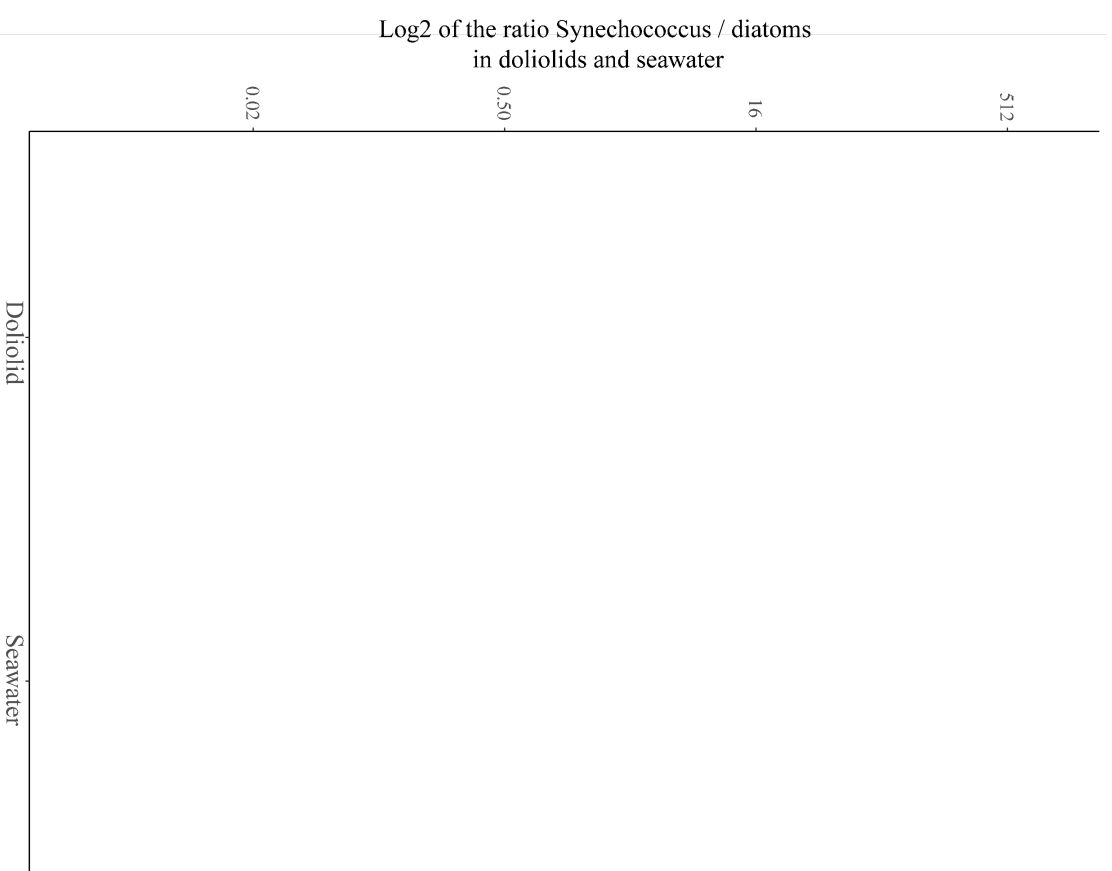
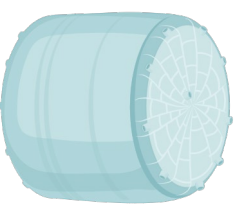
* p -value < 0.05 ** p -value < 0.01

Selective feeding? Diatoms vs. *Synechococcus*

Synechococcus



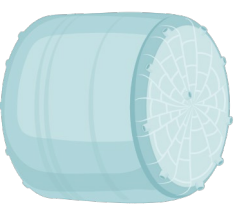
Diatoms



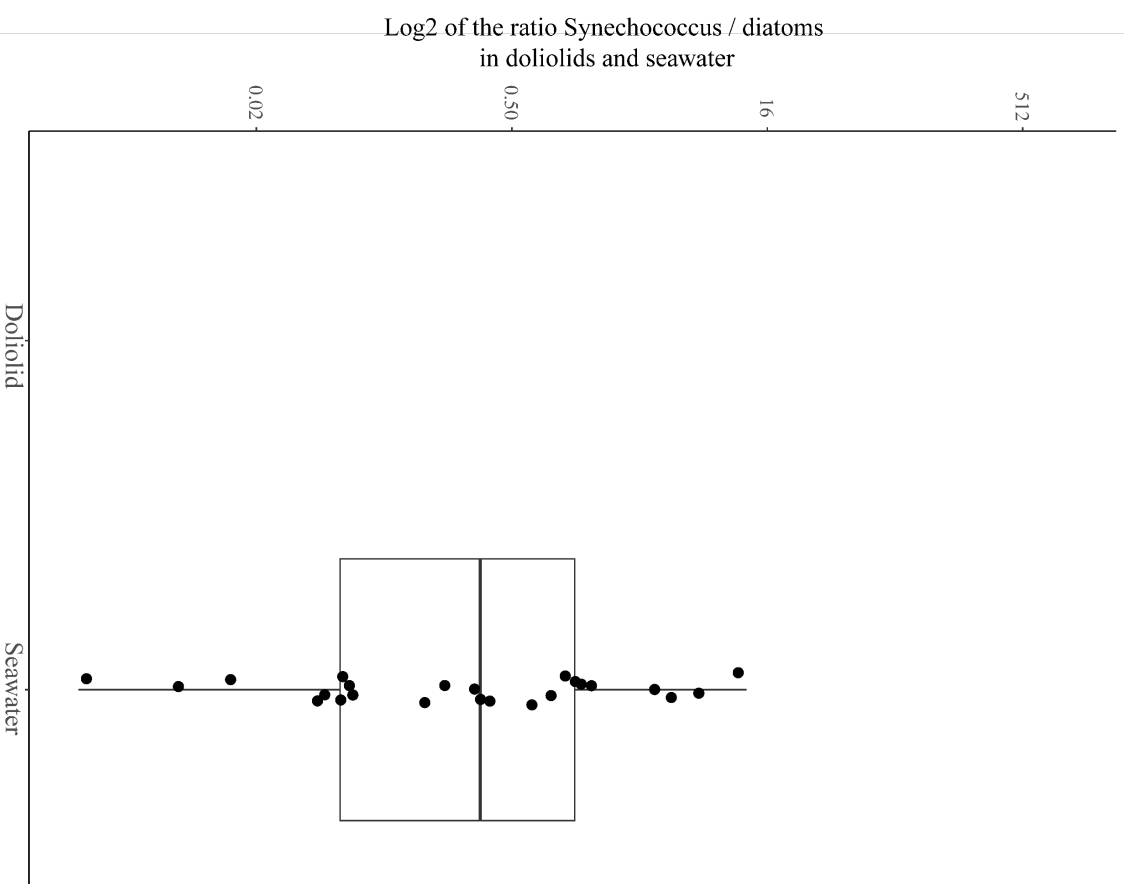
Selective feeding? Diatoms vs. *Synechococcus*



Synechococcus



Diatoms

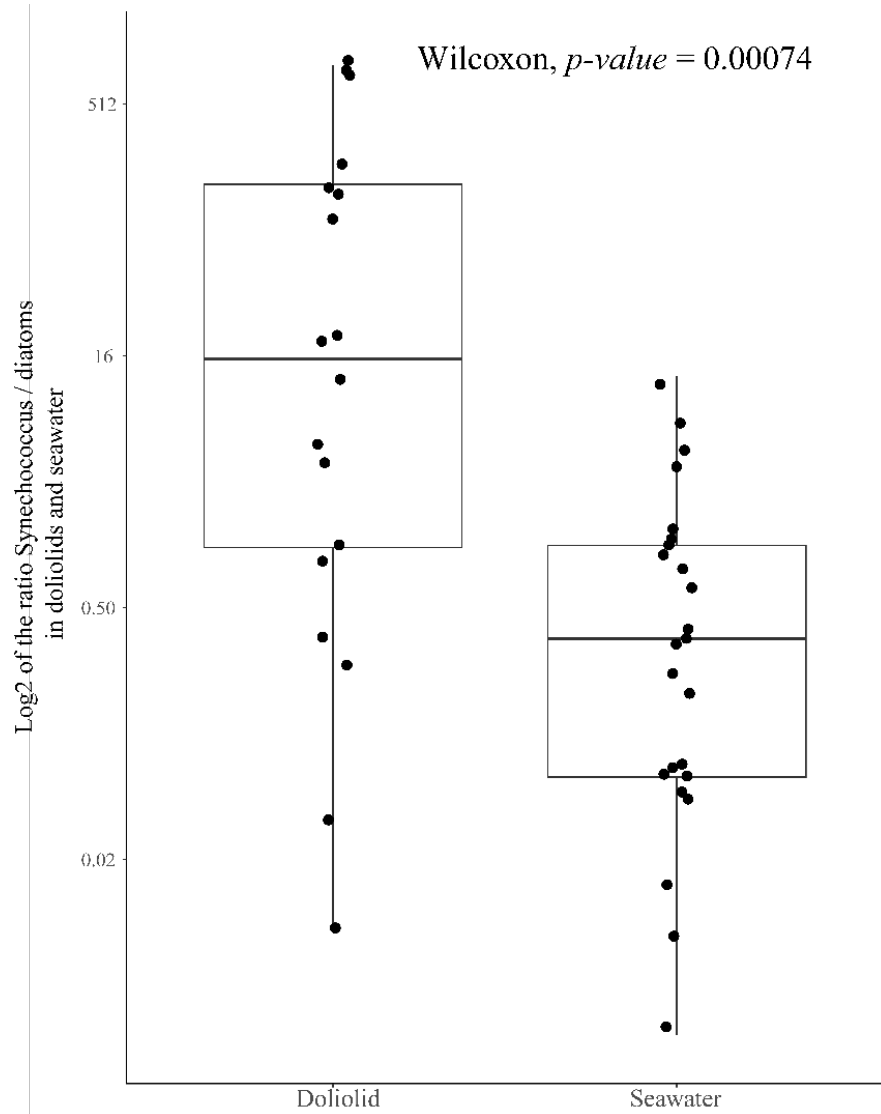


Selective feeding? Diatoms vs. *Synechococcus*

Synechococcus



Diatoms



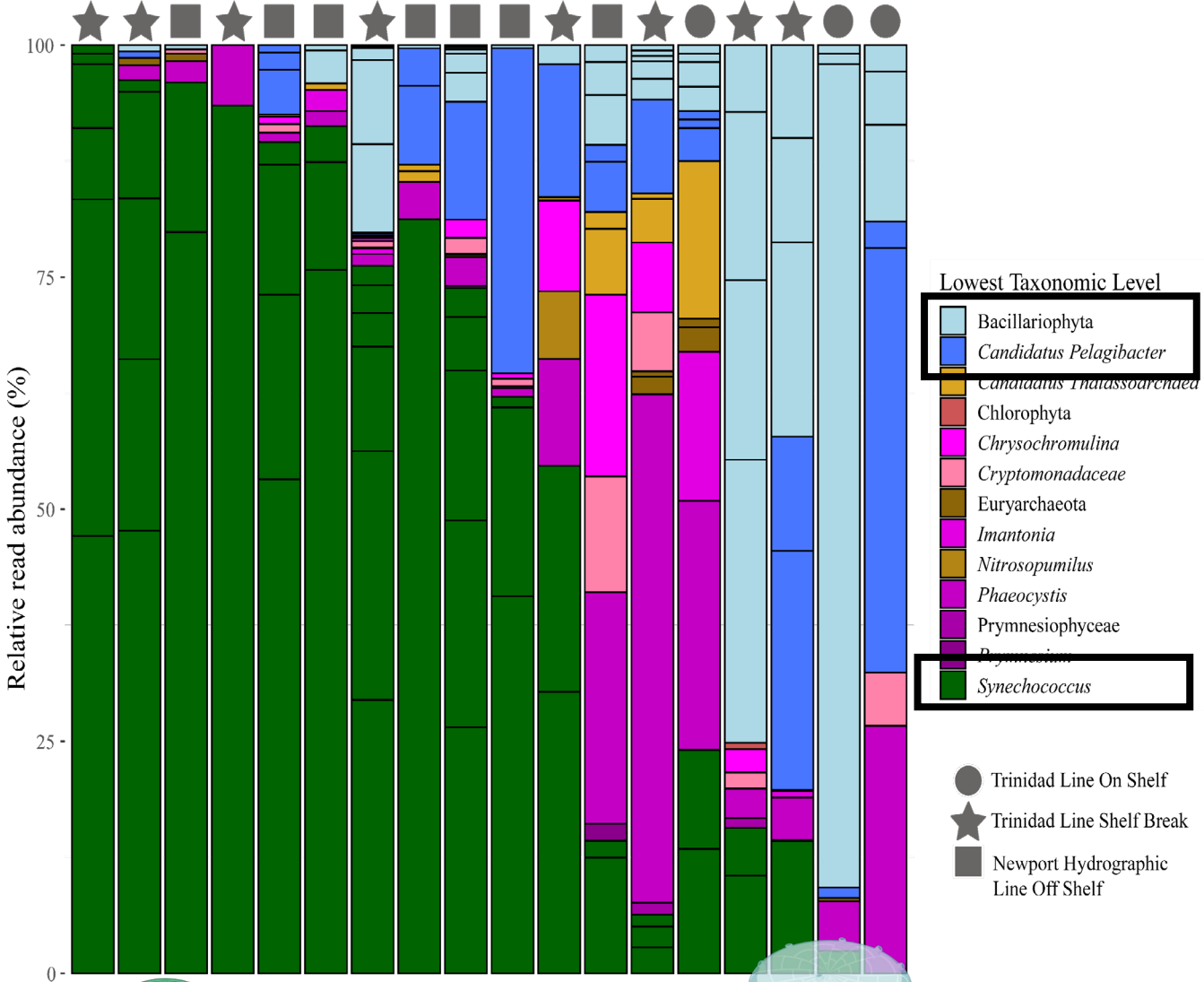
Results

- Selection for *Synechococcus* over diatoms
- Feeding may alter size structure

Chapter 3 Conclusions – doliolid feeding

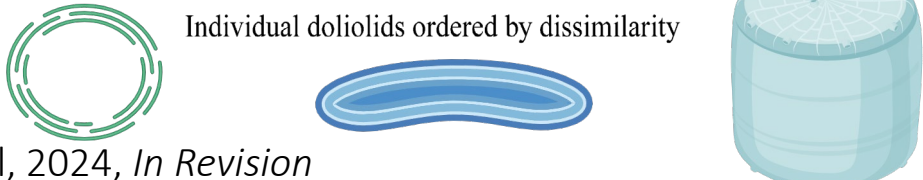
- Prokaryotes present in doliolids
- Feed on more microbial groups than previously known
- Doliolids select *Synechococcus* over diatoms
- Restructure microbial communities?
- Doliolids may impact the prey of higher trophic levels

Synechococcus, diatoms and SAR11 are abundant in doliolids.



Prey differences?

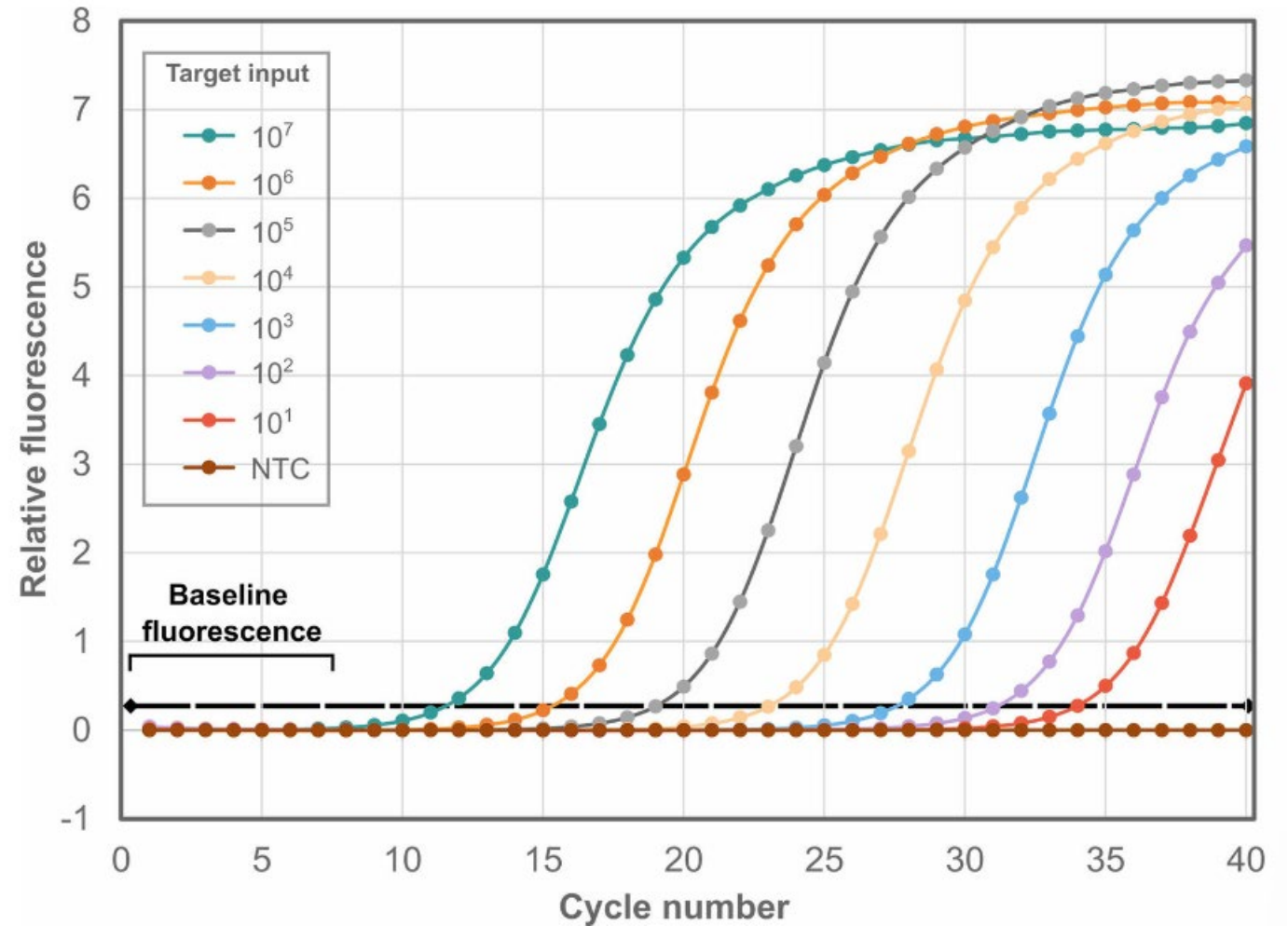
- Unique surface properties
- Abundant cells in seawater
- Selective feeding



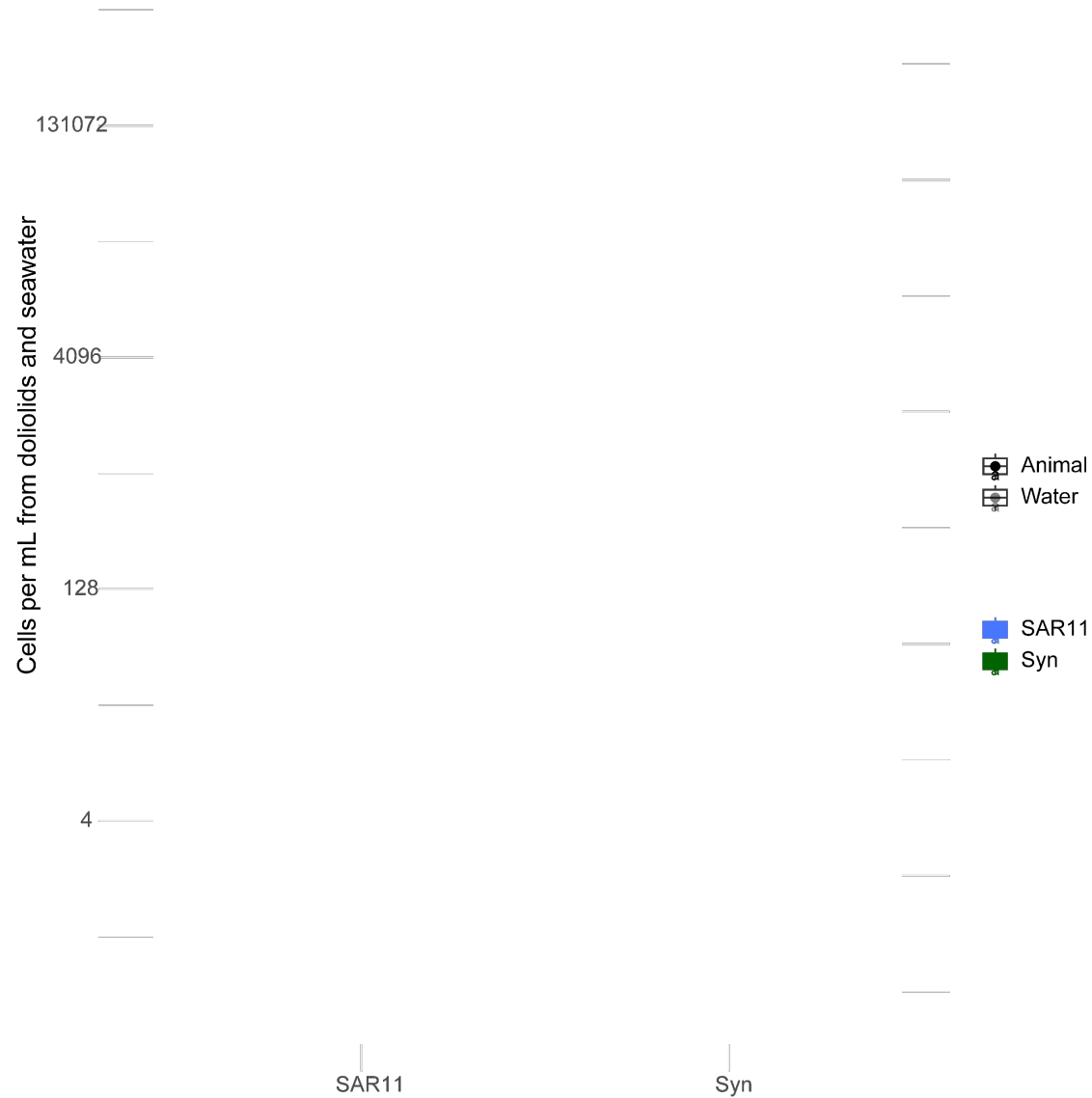
Exact quantities of targeted prey taxa with qPCR

Three published qPCR assays:

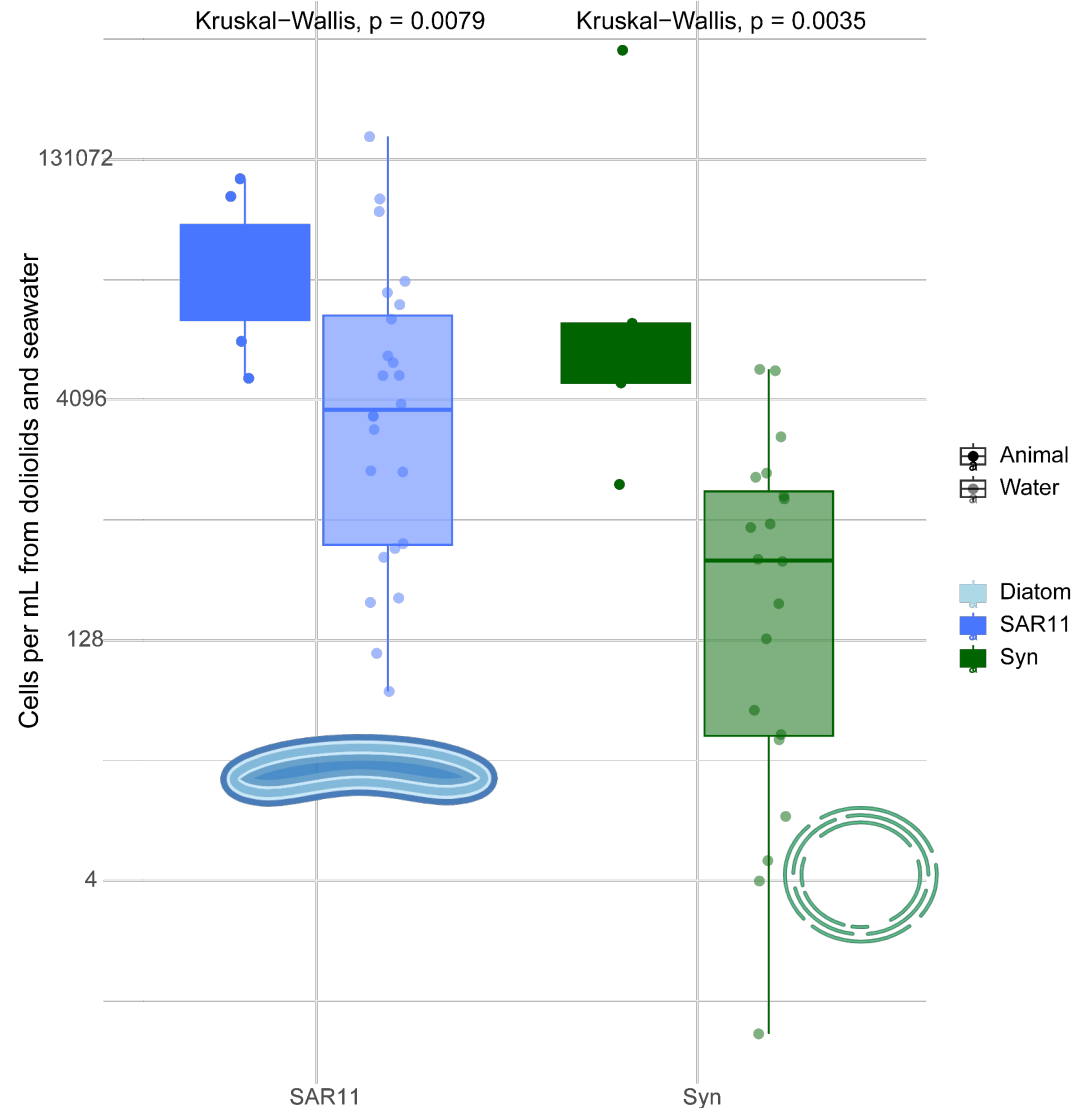
- *Synechococcus* Clade 1
 - Ahlgren, 2012
- Diatoms
 - Frischer et al. 2014; 2021
- SAR11
 - Suzuki et al. 2001



Doliolids enrich SAR11 and *Synechococcus* – *active feeding*



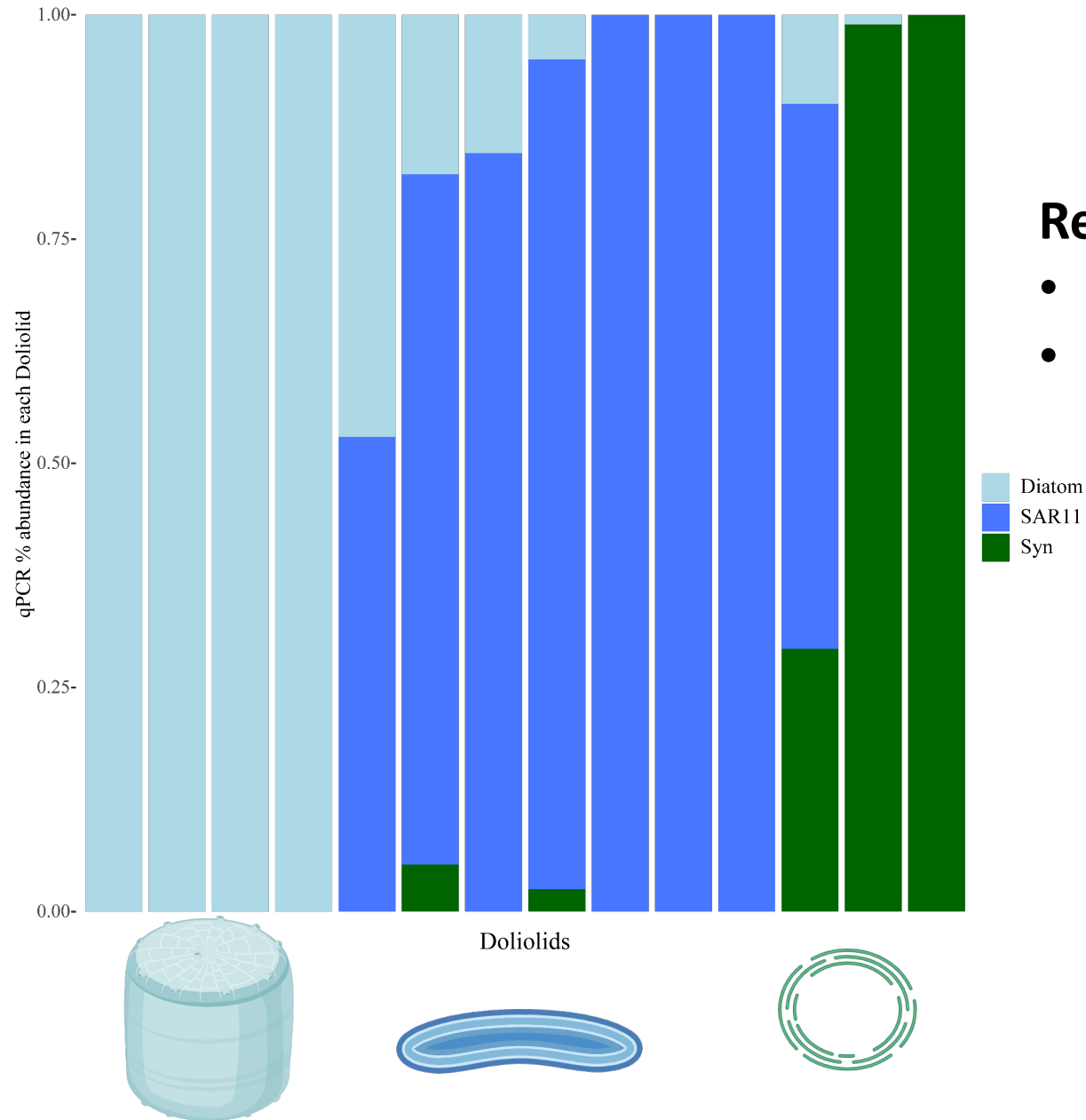
Doliolids feed selectively on SAR11 and *Synechococcus*



Results

- First evidence of feeding on SAR 11
- SAR11 is enriched 2-fold
- First study to quantify feeding on *Synechococcus*
- *Synechococcus* is enriched 118-fold

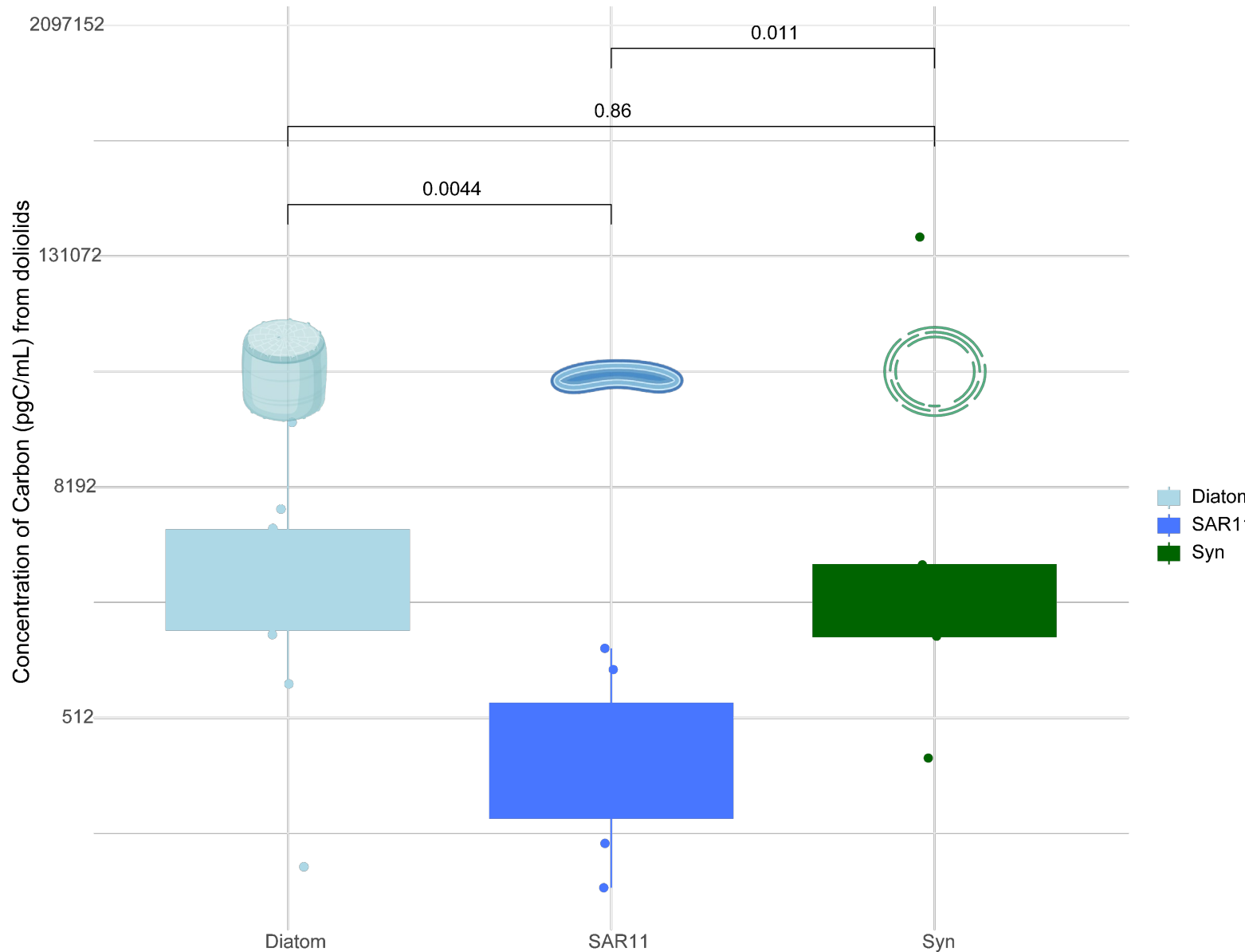
Feeding differences among doliolids?



Results

- Doliolid feeding varies between individuals
- Supports results from Chapter 3

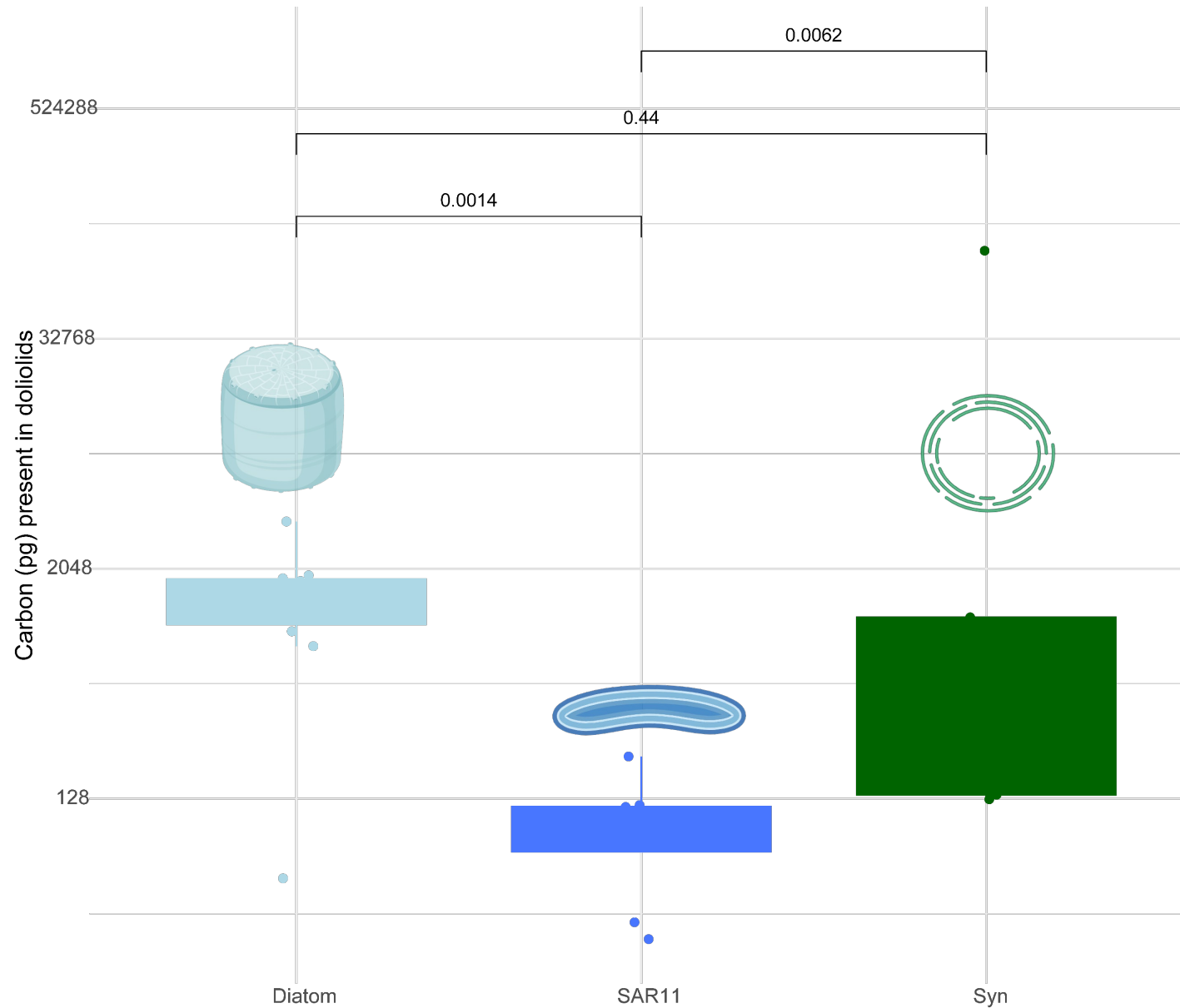
Carbon contribution is different between taxa



Results

- SAR11
 - Most abundant cell
 - Least carbon biomass
- Diatoms
 - Least abundant
 - Most carbon biomass
- *Synechococcus*
 - More carbon than SAR11

Snapshot of carbon contained in doliolids



Results

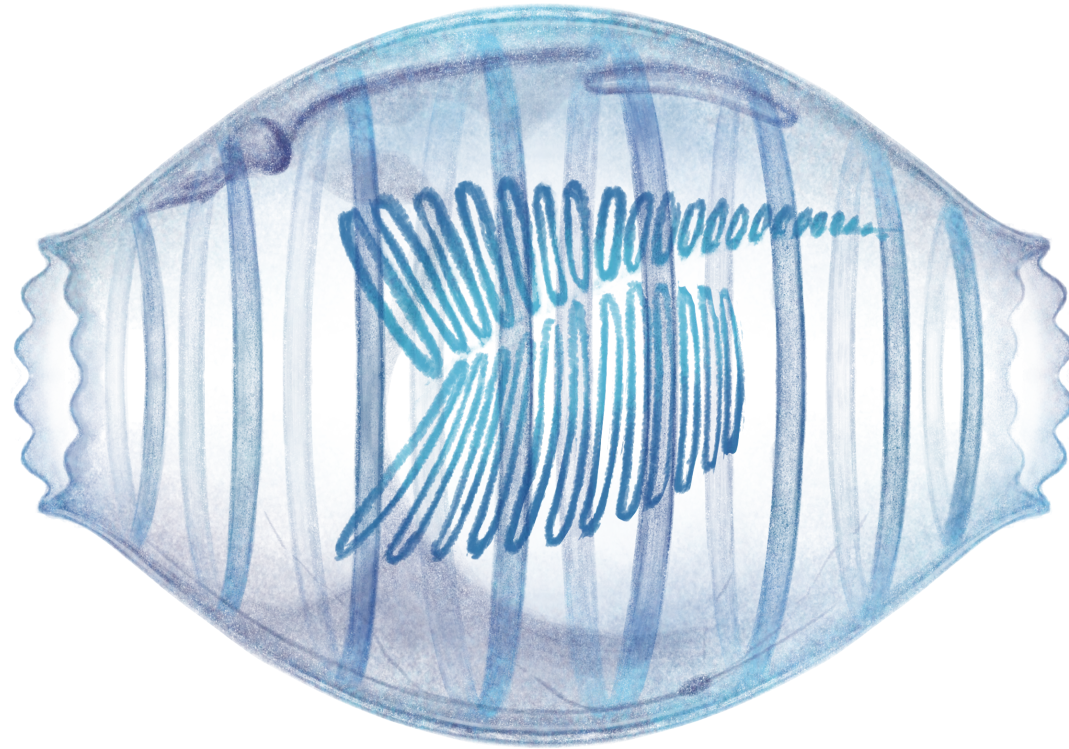
- Meet carbon needs by selective feeding

- Diatom
- SAR11
- Syn

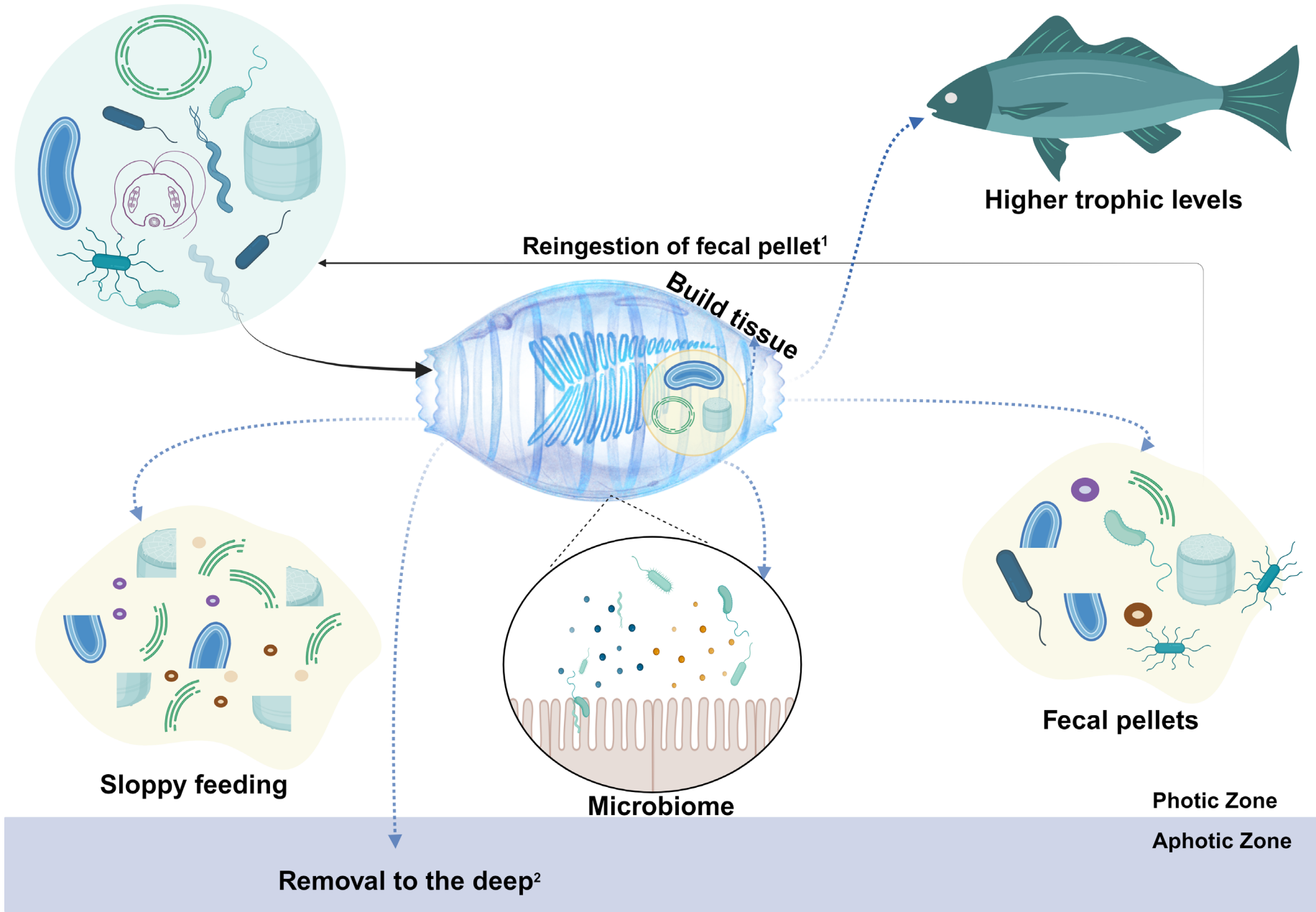
Chapter 4 Conclusions – Quantifying targeted prey

- Doliolids actively feed on SAR11
- Doliolids actively feed on *Synechococcus*
- Doliolid feeding varies between individuals
 - Supporting results from Chapter 3
- Doliolids can meet their carbon requirement through selective feeding on diatoms or *Synechococcus*

Doliolid – microorganism interactions?



Model of doliolid impacts on marine microorganisms



Acknowledgments

Captain and Crew of the R/V Atlantis

Dr. Kevin Johnson

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Dr. Annie Lindgren

Dr. Anne Thompson

Dr. Anna-Louise Reysenbach

Dr. Deborah Duffield

Lab members

Kylee Brevick

Kristin Forgrave

John Gale

Avery Harman

Anvita Kerkar, PhD

Timothy Pettit

Melissa Steinman

Carey Sweeney

Former Lab Members

Andrew Roberts

Collaborators

Gyorgyi Nyerges (Pacific University)

Kelly Sutherland (University of Oregon)

Robert Cowen (Oregon State University)

Su Sponaugle (Oregon State University)

Moritz Schimd (Oregon DFW)

SIM NS

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References

References