



University of Dundee

Ten “Cheat Codes” for Measuring Oxidative Stress in Humans

Cobley, James; Margaritelis, Nikos V.; Chatzinikolaou, Panagiotis N.; Nikolaidis, Michalis G.; Davison, Gareth W.

DOI:
[10.20944/preprints202405.0729.v1](https://doi.org/10.20944/preprints202405.0729.v1)

Publication date:
2024

Licence:
CC BY

Document Version
Early version, also known as pre-print

[Link to publication in Discovery Research Portal](#)

Citation for published version (APA):

Cobley, J., Margaritelis, N. V., Chatzinikolaou, P. N., Nikolaidis, M. G., & Davison, G. W. (2024). Ten “Cheat Codes” for Measuring Oxidative Stress in Humans. Preprints.org. <https://doi.org/10.20944/preprints202405.0729.v1>

General rights

Copyright and moral rights for the publications made accessible in Discovery Research Portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

Review

Not peer-reviewed version

Ten “Cheat Codes” for Measuring Oxidative Stress in Humans

[James Cobley](#)^{*}, Nikos Margaritelis, [Panagiotis Chatzinikolaou](#), [Michalis Nikolaidis](#), [Gareth Davison](#)

Posted Date: 12 May 2024

doi: 10.20944/preprints202405.0729.v1

Keywords: Oxidative stress; ROS; oxidative damage; redox regulation; antioxidant; method



Preprints.org is a free multidiscipline platform providing preprint service that is dedicated to making early versions of research outputs permanently available and citable. Preprints posted at Preprints.org appear in Web of Science, Crossref, Google Scholar, Scilit, Europe PMC.

Copyright: This is an open access article distributed under the Creative Commons Attribution License which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Review

Ten “Cheat Codes” for Measuring Oxidative Stress in Humans

James N. Cobley ^{1,*}, Nikos V. Margaritelis ^{2,†}, Panagiotis N. Chatzinikolaou ^{2,†} and Michalis G. Nikolaidis ^{2,†} and Gareth W. Davison ^{1,†}

¹ Ulster University, Newtonabbey, Northern Ireland, UK.

² Aristotle University of Thessaloniki, Serres, Greece.

* Correspondence: J_cobley@yahoo.com

† These authors contributed equally.

Abstract: Formidable and often seemingly insurmountable conceptual, technical, and methodological challenges hamper the measurement of oxidative stress in humans. For instance, fraught and flawed methods, such as the thiobarbituric acid reactive substances assay kits for lipid peroxidation, rate-limit progress. To advance translational redox research, we present ten comprehensive “cheat codes” for measuring oxidative stress in humans. The cheat codes include analytical approaches to assess reactive oxygen species, antioxidants, biomarkers of oxidative damage and redox regulation. They provide essential conceptual, technical, and methodological information inclusive of curated “do” and “don’t” guidelines. Given the biochemical complexity of oxidative stress, we present a research question-grounded decision tree guide for selecting the most appropriate cheat code (s) to implement in a prospective human experiment. Worked examples demonstrate the benefits of the decision tree-based cheat code selection tool. The ten cheat codes define an invaluable resource for measuring oxidative stress in humans.

Keywords: oxidative stress; ROS; oxidative damage; redox regulation; antioxidant; method

1. Introduction

Many researchers investigating human physiology in health and disease, including sports nutrition and exercise metabolism, need to measure oxidative stress. For example, one must measure oxidative stress to determine whether a nutritional intervention, such as vitamin E (Cobley and Marrin, 2012), acted as an antioxidant to regulate exercise adaptations (Cobley et al., 2015b; Forman et al., 2014). However, formidable conceptual, technical, and methodological challenges pose significant barriers to measuring oxidative stress in humans (Cobley et al., 2017; Murphy et al., 2011). These challenges, which include the inapplicability of cutting-edge genetically-encoded reactive oxygen species (ROS) probes in human studies (Belousov et al., 2006; Pak et al., 2020) and the daunting task of choosing among fraught and flawed assays, such as the thiobarbituric acid reactive substances (TBARS) assay (Forman et al., 2015; Halliwell and Whiteman, 2004), rate-limit progress. They even deter many researchers from measuring oxidative stress altogether. This predicament can be likened to a novice navigating a challenging level in a video game, where 'cheat codes' would help solve complex, seemingly intractable, problems. To assist those grappling with persistent methodological challenges, we define key terms, such as ROS, and elaborate ten comprehensive “cheat codes” for measuring oxidative stress in humans. Finally, we present a research question-grounded decision tree guide for selecting the most appropriate cheat code (s) to implement in a prospective human experiment before offering concluding perspectives.

Part 1. A brief guide to oxygen, ROS, antioxidants, and oxidative stress

1.1. Oxygen

Our 37-trillion cells vociferously consume around 3.5 millilitres per minute of a free radical: oxygen. Diatomic oxygen contains two unpaired electrons—it is a diradical (Fridovich, 1978). The lone, unpaired, electrons in oxygen, spin in parallel antibonding orbitals (Cobley et al., 2018) (e.g., $\uparrow\uparrow$, see Figure 1). Aerobic life is possible because oxygen cannot react with the vast majority of electron spin paired molecules in the body due to the reactions being spin forbidden (Fridovich, 1998). To explain spin forbidden, think of the card game “snap”. Our goal is to “snap”, that is react, our cards, but we can only ‘snap’ spin allowed matches. If you play an oxygen card and we play a guanine DNA base card, then we cannot “snap” as a result of:

No appreciable reaction = oxygen: $\uparrow\uparrow$ guanine-DNA $\downarrow\uparrow$ (spin violated)

If you could “flip” the spin, as happens when UV-light excites oxygen to yield the non-radical singlet oxygen (Ogilby, 2010), then the first one to “snap” wins the game owing to:

Appreciable reaction = singlet oxygen $\downarrow\downarrow$ guanine-DNA $\downarrow\uparrow$ (spin allowed) (Sies and Menck, 1992)

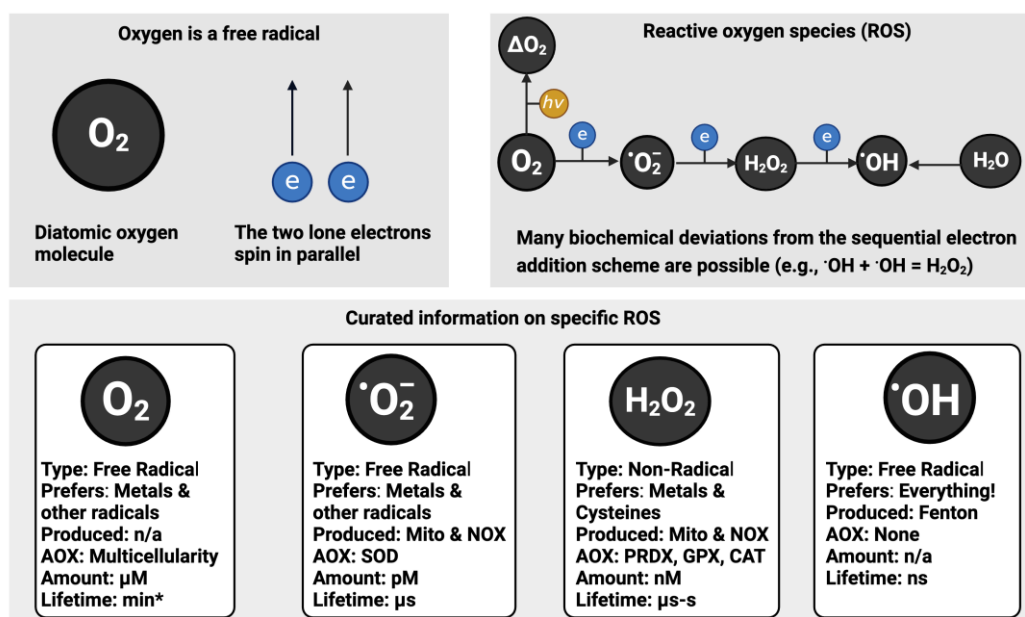


Figure 1. Oxygen (O_2) and reactive oxygen species (ROS). The upper left panel depicts diatomic ground-state molecular dioxygen as a free radical with the two lone electrons (e) spinning in parallel according to the laws of quantum mechanics, such as the Pauli exclusion principle. The upper right panel displays the electronic relationship between different ROS. From O_2 as a starting point, an input of energy can flip the spin whereby the lone electrons spin in parallel in singlet oxygen ($^1\Delta_g\text{O}_2$). Alternatively, univalent reduction of O_2 produces the superoxide ($\text{O}_2^{\cdot-}$). The proton-coupled univalent reduction of $\text{O}_2^{\cdot-}$ produces hydrogen peroxide (H_2O_2). Without the coupling of protons, the extremely unstable peroxide dianion, a non-radical, would be produced. The univalent reduction of H_2O_2 yields a hydroxide anion (omitted for clarity) and the hydroxyl radical ($\cdot\text{OH}$). In some cases, $\cdot\text{OH}$ can be formed from water (H_2O). For example, an input of energy sufficient to split the water in a homolytic fission reaction will produce the hydroxide anion and $\cdot\text{OH}$. The lower panel visually displays curated information on specific ROS. Stating a concentration value for $\cdot\text{OH}$ is difficult: it’s diffusion-controlled reactivity effectively prevents it from accumulating to an appreciable level. Abbreviations: NOX = NADPH oxidase, Mito = mitochondria, PRDX = peroxiredoxin, GPX = glutathione peroxide, CAT = catalase.

1.2. ROS

Oxygen appreciably reacts with molecules able to donate one electron, termed univalent reduction (see equation 1), leading to a zoo-like menagerie of free radical (e.g., superoxide) and non-radical (e.g., hydrogen peroxide) ROS (see Figure 1). As an insightful account remarked (Meo and Venditti, 2020), ROS was first used in the abstract of a paper in 1977 (Weiss et al., 1977). Before “ROS” became de rigor, virtually every paper named the specific species being studied (e.g., (Misra, 1974)).

Many problems stemmed from failing to state the specific species (Flohé, 2020) and certain misconceptions about 'ROS' leading to interpretational errors. Selected instructive points about ROS include:

- Superoxide is not necessarily super. The "superoxide" moniker originated from the odd stoichiometry of a chemical reaction in 1934 (Neuman, 1934). It had nothing to do with any special "super" biochemical reactivity as an oxidant (Sawyer and Valentine, 1981). Sawyer and Valentine commented that the probability of superoxide oxidising a molecule to yield the peroxide dianion is nil. Moreover, McCord and Fridovich discovered superoxide dismutase (SOD) by observing that superoxide reduced ferric cytochrome c (McCord and Fridovich, 1969, 1968).
- Each ROS is biochemically unique (Dickinson and Chang, 2011; Gutteridge, 2015; Winterbourn, 2008). Superoxide appreciably reacts with a small number of targets, such as tryptophan free radicals (Carroll et al., 2018). Conversely, the ferocious hydroxyl radical, rapidly reacts with virtually every organic molecule at a diffusion-controlled rate (Halliwell, 2007).
- There is no set percentage rate of superoxide production from consumed oxygen (Boveris and Chance, 1973; Chance et al., 1979). As studies comparing rest to exercise attest (Goncalves et al., 2015), the dynamic variable rate of mitochondrial superoxide production is context-dependent (Cobley, 2018; Sidlauskaite et al., 2018).
- To quote Sies and Jones "ROS is a term, not a molecule" (Sies and Jones, 2020).

Equation 1: Oxygen + electron → superoxide

1.3. Antioxidants

Even luminaries struggle to classify "antioxidants" (Gutteridge and Halliwell, 2010; Halliwell and Gutteridge, 2015). Unexpected, class defying, antioxidants include multicellularity as a "defence" against oxidative stress by limiting oxygen exposure (Taverne et al., 2018), uncoupling proteins to decrease the probability of mitochondrial superoxide production by lowering the electrochemical proton motive force (James N. Cobley, 2020; Echtaï et al., 2002), and the diverse proteins responsible for repairing oxidative damage (Halliwell, 2023). Despite the perennial search for the best catch-all definition, experts (Murphy et al., 2022) define an antioxidant as:

"any substance that delays, prevents or removes oxidative damage to a target molecule" OR "a substance that reacts with an oxidant to regulate its reactions with other targets, thus influencing redox-dependent biological signalling pathways and/or oxidative damage"

A protein repairing oxidised DNA would fall into the first category whereas catalase (CAT) would fall into the second. Selected instructive points about antioxidants include:

- There is no one "best" antioxidant. Just like ROS, they are all different (see Figure 2).
- A few "frontline" enzymes like SOD do most of the redox "heavy-lifting". Ordinarily, SOD isoforms consume most of the superoxide produced in a cell (Imlay, 2008). So, only picomoles remain for other molecules, such as vitamin C, to "scavenge". For example, 3.01×10^{18} (5 μM) superoxide molecules can be produced per second in *Escherichia coli*, but SOD limits [superoxide] by 4-logs to 6.0×10^{14} molecules corresponding to 0.0009 μM or 900 picomoles (Imlay, 2013).
- An antioxidant is context-dependent. SOD generates hydrogen peroxide and oxygen. So, it is an antioxidant in concert with other enzymes (Winterbourn, 1993). Further, when mis-metalled the mitochondrial SOD isoform produces hydroxyl radical (Ganini et al., 2018).
- Many antioxidants "moonlight". Sticking with SOD, the copper zinc isoform can act as a transcription factor (Tsang et al., 2014) and a cysteine oxidase (Winterbourn et al., 2002). Relatedly, SOD regulates electrochemistry by preventing superoxide, an excellent Brønsted base, consuming protons via spontaneous dismutation (Kettle et al., 2023).
- Some antioxidant enzymes react with many species. Like how superoxide also reacts with (and inactivates) CAT and glutathione peroxidase (GPX) (Blum and Fridovich, 1985; Kono and Fridovich, 1982), emerging evidence demonstrates that SOD metabolises hydrogen sulfide

(Switzer et al., 2023), which complements prior evidence of reactivity, albeit kinetically slow, with hydrogen peroxide (Liochev and Fridovich, 2002).

- No specific antioxidants target the hydroxyl radical. If you had a 100 kg athlete, you'd need to load them with 50 kg of the antioxidant to meaningfully scavenge the hydroxyl radical on mass action and kinetic grounds (Dickinson and Chang, 2011; Winterbourn, 2008).
- Reactive species can be antioxidants. Nitric oxide reacts with lipid peroxyl radicals to terminate a free radical chain reaction (Niki, 2009; Yin et al., 2011). The nitrated lipids so-formed may possess anti-inflammatory properties (Melo et al., 2019).
- As in real estate, location matters. Polyphenols may be antioxidants in the gut (Halliwell et al., 2000) before being metabolised to and secreted in pro-oxidants forms (Owens et al., 2018). Touted 'antioxidants' like polyphenols often exert beneficial effects by acting as pro-oxidants (Halliwell, 2009, 2008).

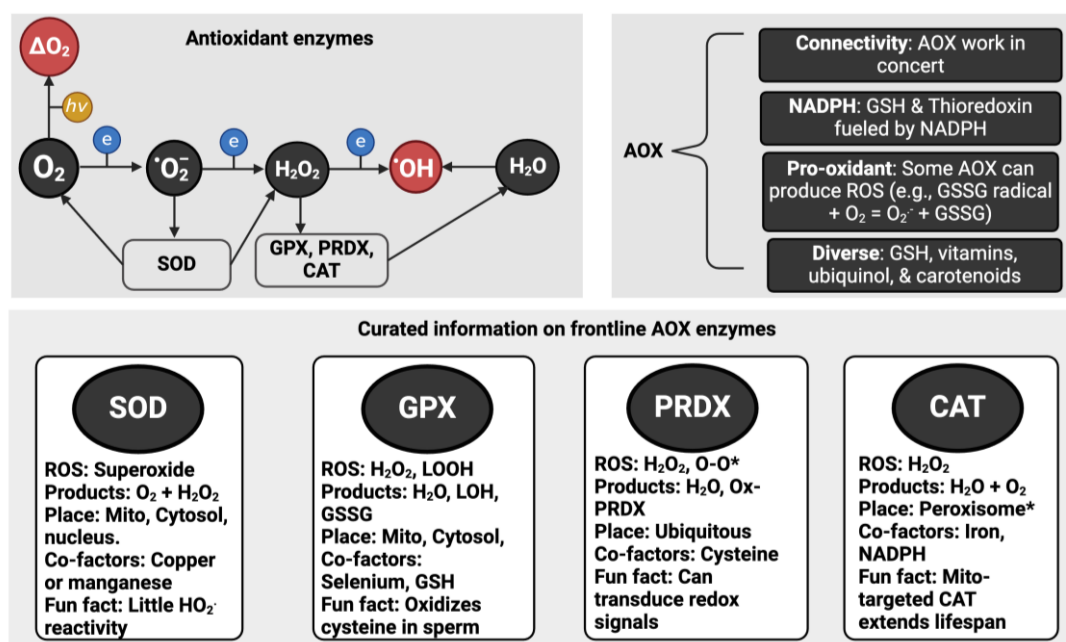


Figure 2. Antioxidants (AOX). The upper left panel visually displays the link between key reactive oxygen species (ROS) and enzymatic antioxidants. Superoxide dismutase (SOD) isoforms metabolise superoxide (O₂⁻) to oxygen (O₂) and hydrogen peroxide (H₂O₂). Peroxiredoxin (PRDX) and glutathione peroxidase (GPX) isoforms can metabolise H₂O₂ to water (H₂O). Catalase (CAT) effectively dismutates H₂O₂ to O₂ and H₂O. There are no known AOX enzymes in humans designed to specifically react with singlet oxygen (1ΔgO₂) and the hydroxyl radical (OH). The upper right panel visually displays interesting features of antioxidants, from the connectivity principle to their biochemical diversity (see the main text for details). The lower panel visually displays curated information for the key AOX enzymes. The PRDX isoforms can react with a range of species with an O-O bond, such as protein and lipid hydroperoxides. Abbreviations: GSH = reduced glutathione, GSSG = oxidised glutathione, GSSG radical = the oxidised glutathione free radical, Mito = Mitochondria.

1.4. Oxidative Stress

Sies (SIES, 1985) defined oxidative stress as an imbalance between ROS and antioxidants (AOX) in favour of the former that leads to oxidative damage (equation 2). Oxidative damage includes chemically diverse oxidised lipid, DNA/RNA, and protein molecules. For example, 2-oxohistidine is an oxidised form of the amino acid histidine (Davies, 2016). While there is always some oxidative damage, persistent and excessive levels of unrepaired molecules are linked to certain diseases (Forman and Zhang, 2021), most notably cancer (Halliwell, 2006, 1987; Hayes et al., 2020). In response to mounting evidence (D'Autréaux and Toledano, 2007; Holmström and Finkel, 2014; Paulsen and

Carroll, 2013), Sies and others redefined oxidative stress to include redox regulation (Jones, 2006; Sies, 2020, 2017, 2015; Sies et al., 2016) (equation 3).

Equation 2: $ROS > AOX = \uparrow$ oxidative damage.

Equation 3: $\uparrow ROS + \sim AOX =$ redox regulation.

Variations of equations 2 and 3 are possible, such as \uparrow oxidative damage = $\sim ROS + \downarrow AOX$. The equations formulate input-output relationships (see Figure 3). The biochemical reactions underlying the output constitute a “process” module (input \rightarrow process \rightarrow output) (Cobley, 2023a). For example, equation 1 specifies a ROS input, such as increased nuclear hydroxyl radical generation, producing an oxidative damage output, such as guanine base oxidation in DNA, via process-specific biochemical reactions (Cobley et al., 2015a). In this case, the AOX module would comprise factors that constrain the Fenton reaction (see equation 4) inclusive of hydrogen peroxide availability as influenced by proximal superoxide production, the nature of the iron ligand, and enzymes, such as CAT. Like many things in redox biology, the identity of the oxidising species formed by the Fenton reaction, first described in 1876, is still contested (Halliwell et al., 2021; Illés et al., 2019; Koppenol, 2022).

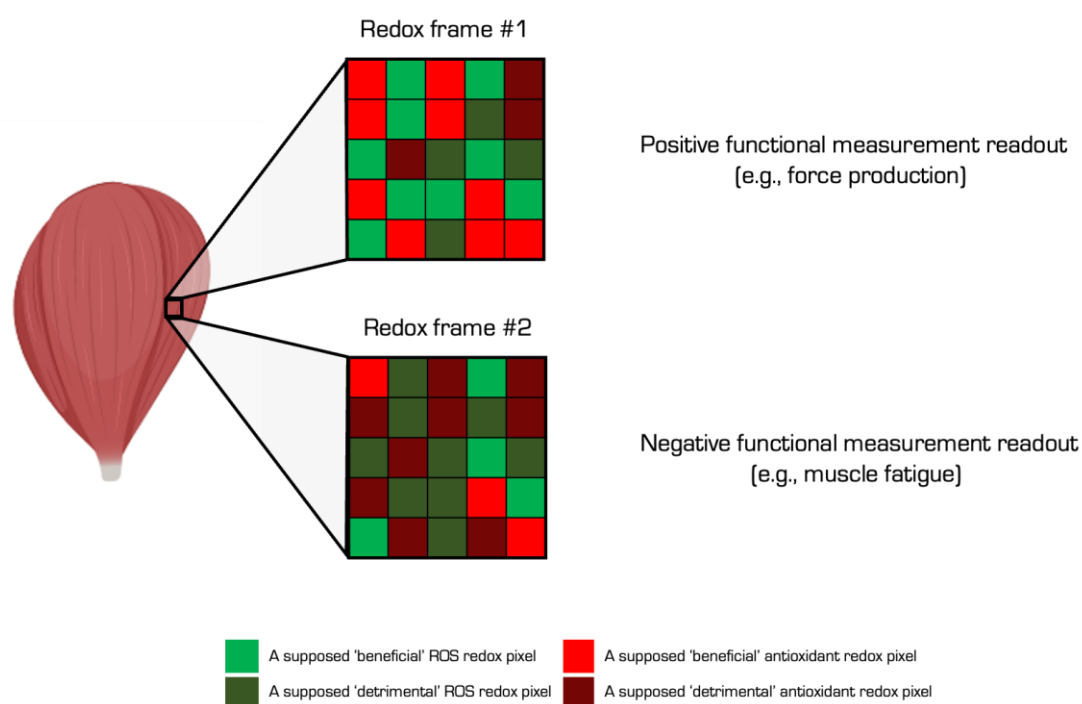


Figure 3. The oxidative stress biochemical pixel concept visualised. In this purely illustrative example, the two redox frames contain different redox pixels corresponding to specific ROS (green) and antioxidant (red) biochemical reactions that we have for the purposes of the figure labelled ‘beneficial’ or ‘detrimental’. How we interpret the pixels depends on the measurement outcome. In this example, the biochemistry displayed in frame 1 corresponds to a positive functional readout: muscle force production. Conversely, the biochemistry displayed in frame 2 corresponds to a negative readout: muscle fatigue. The example also illustrates the dynamic nature of the biochemistry, frame 1 could give way to frame 2 over time, which illustrates how the outcome also depends on when we look.

Equation 4: $H_2O_2 + Fe^{2+} \rightarrow OH \cdot + OH \cdot + Fe^{3+}$ (this equation is an example of the many possible reactions in Fenton systems)

Is oxidative stress good, bad, or neither? The answer and the source of much confusion, is all three. Whether biochemical reactions are ‘good’ or ‘bad’, in so far as anything can be so simply classed, depends on the context. Context-dependent functionality means different interpretations of the same immutable biochemistry can coexist. Like a picture, the same oxidative stress frame can contain ‘good’ and ‘bad’ pixels whereby each pixel defines specific biochemical reactions. The pixels illuminated by a functional measurement spotlight can lead to different, even diametrically opposed, interpretations of the same redox picture (Nikolaidis and Margaritelis, 2018). Measurements

influence interpretations. In humans, whole-body metrics like peak power output, vs. molecular proxies like mRNA levels lead to a neutral vs. negative effect of the same redox picture in the context of antioxidants influencing exercise adaptations (Nikolaidis et al., 2012).

Oxidative stress is evolving to a mobile element fluctuating about a spectrum bookended by “good” oxidative eustress and “bad” oxidative distress (Sies, 2021, 2019; Sies and Ursini, 2021; Ursini et al., 2016). Like the goldilocks homeostatic principles, the modern view that cells need to produce some ROS in light of their beneficial effects implies that insufficient levels of ROS can be harmful, termed reductive stress. Despite misgivings about the term (Gutteridge and Halliwell, 2018), recent evidence unearthed a transcriptional response to insufficient ROS levels (Manford et al., 2021, 2020). To help researchers clearly communicate their work, we recommend that they define key terms using suggested, and in some cases “operational”, definitions (see Table 1).

Table 1. Suggested definitions of selected key terms.

Term	Definition	Example
<i>Free Radical</i>	A molecule with one or more unpaired valence electrons that is capable of independent existence.	Hydroxyl radical
<i>Non-radical</i>	A spin-paired molecule devoid of unpaired electrons.	Hydrogen peroxide
<i>ROS</i>	Free radical and non-radical oxygen-derived reactive species.	Superoxide
<i>Reactive species</i>	A catch-all term for all free radical and non-radical reactive species. Includes oxygen, sulfur, nitrogen, and carbon atom centred species.	Peroxynitrite (sum of the protonated and deprotonated forms)
<i>Reductant</i>	A molecule that donates one or more electrons to a redox reaction partner.	NADPH donating a hydride to reduce oxidised glutathione reductase.
<i>Oxidant</i>	A molecule that takes one or more electrons from a redox reaction partner.	Hydroxyl radical oxidising L-cysteine to a thiyl radical.
<i>Antioxidant</i>	Any substance that delays, prevents or removes oxidative damage to a target molecule.	Vitamin E preventing lipid peroxidation by reducing an alkoxyl radical to a hydroperoxide.
	A substance that reacts with an oxidant to regulate its reactions with other targets, thus influencing redox-dependent biological signalling pathways and/or oxidative damage.	CAT metabolising hydrogen peroxide to prevent reaction with another targets, such as a transition metal ion.
<i>Oxidative stress</i>	An imbalance between ROS and antioxidants leading to disrupted redox-regulation and/or oxidative damage.	See below.

<i>Oxidative eustress</i>	A beneficial imbalance between ROS and antioxidants leading to disrupted redox-regulation and/or oxidative damage.	Specific cysteine oxidation patterns and oxidative macromolecule damage in an acute exercise context.
<i>Oxidative distress</i>	A deleterious imbalance between ROS and antioxidants leading to disrupted redox-regulation and/or oxidative damage.	Aberrant and potentially random patterns of cysteine oxidation and oxidative damage in a musculoskeletal ageing context.

Part 2. Ten “cheat codes” for measuring oxidative stress in humans

Cheat code 1. Avoid the minefield of measuring ROS directly in humans (at least for now)

Although forward-looking possibilities may permit their measurement in the future (hence the at least for now clause), it is exceptionally difficult to measure ROS in humans because:

- One invariably exposes the sample to 21% oxygen. Mitochondrial superoxide production depends, in part, on [oxygen] (Murphy, 2009). So, raising [oxygen] from 1-10% to 21% (Ast and Mootha, 2019) by aerating the sample would be expected artificially increase superoxide production (Keeley and Mann, 2019).
- The ‘ROS’ in the sample will have inevitably disappeared before one can measure them. They ephemerally flit in and out of existence on nanosecond timescales (10^{-9} of a second). So, what is one really measuring? Potentially, the rate of artificial ROS production in a heavily oxygenated sample (Murphy et al., 2022).
- Although it is possible to minimise the above (e.g., degassing the sample and rapidly adding a probe), it is arduous. Even if artificial generation were minimised, a superoxide probe, for example, must still compete with SOD (Zielonka and Kalyanaraman, 2018), hampering the ability to detect all of the molecule in the sample. There is always the possibility of inadvertent artefacts, such as the release of redox-active iron ions from the haemolysis of erythrocytes in blood samples.
- Many lysis buffers contain ROS, such as hydrogen peroxide and lipid hydroperoxides (LOOH) (Eid et al., 2024)
- Cutting-edge genetically encoded probes cannot be used in humans (Erdogan et al., 2021).

Fortunately, situations, such as the classic studies demonstrating exercised skeletal muscle tissues produced free radicals (Davies et al., 1982), where one needs to measure ROS in humans are relatively rare. Nevertheless, researchers seeking to measure ROS may wish to (1) avoid the fluorescent probe 2',7'-dichlorodihydrofluorescein as it produces ROS (Bonini et al., 2006; Wardman, 2007) and kits claiming to exclusively measure the hydroxyl radical, (2) carefully read assay kits, and (3) implement a valid technique, such as electron paramagnetic resonance-based spin trapping (Bailey et al., 2003). Even amongst experts routinely measuring ROS, unmet methodological needs perennially inspire new approaches (Shchepinova et al., 2017), such as the evolution from pH sensitive to insensitive hydrogen peroxide sensors (Pak et al., 2020). Or boronic to borinic acid-based fluorescent probes for kinetically more efficient hydrogen peroxide sensing (Gatin-Fraudet et al., 2021). Measuring ROS doesn't reveal what they are doing, which is ultimately what we want to discover (Halliwell and Whiteman, 2004).

Cheat code 2. How to infer ROS production in human samples using endogenous reporter molecules

Inspired the Kalyanaraman group (Cheng et al., 2018), one can infer ROS production in humans (see Figure 4) using endogenous reporter molecules of free radicals (aconitase) and non-radicals (peroxiredoxins, PRDX). First, superoxide and other selected species, such as peroxynitrite, react exceptionally fast with aconitase (Hausladen and Fridovich, 1994). Peroxynitrite production depends on superoxide: nitric oxide + superoxide = peroxynitrite (Radi, 2018). By depriving it of an essential

iron atom, they inactivate aconitase (Imlay, 2013). By measuring aconitase activity, as detailed elsewhere (Gardner, 2002; Gardner et al., 1994; Vásquez-Vivar et al., 2000), it is possible to obtain a surrogate readout of the levels of selected ROS. Unlike ectopic approaches whereby adding a probe may perturb the biological system (Roelofs et al., 2015), endogenous reporters preserve the natural state.

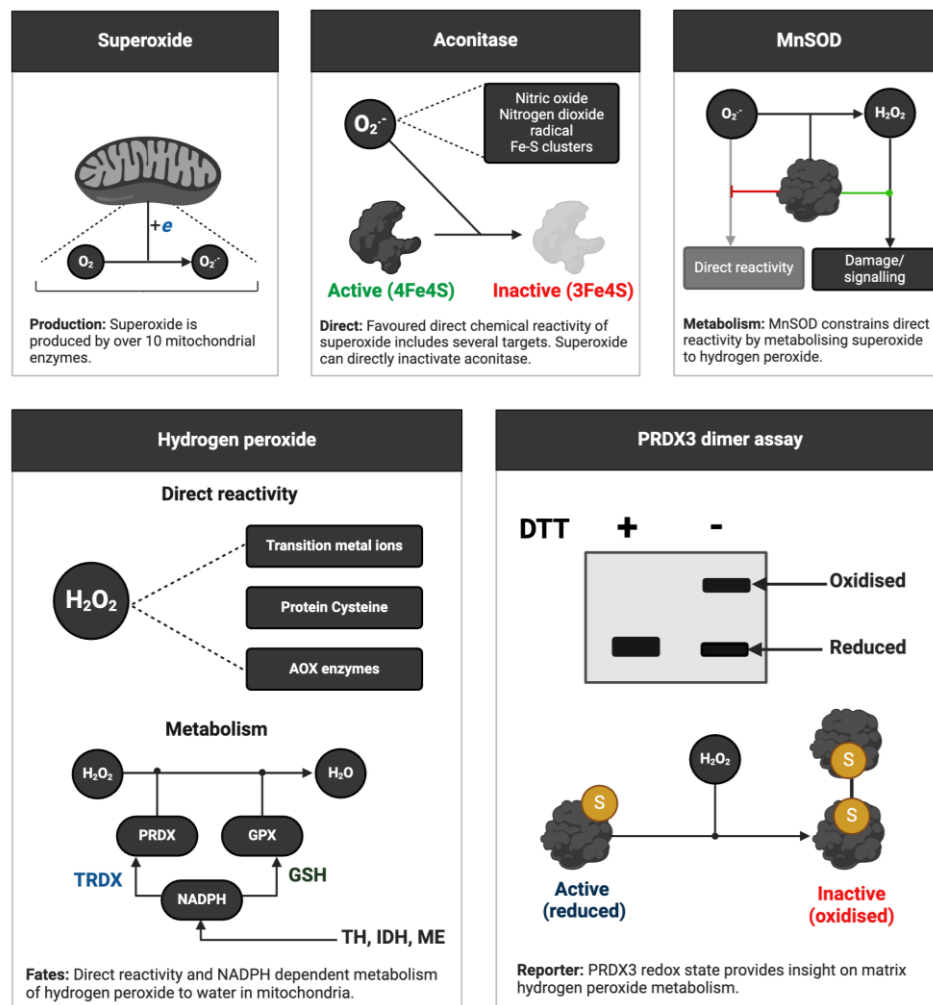


Figure 4. Inferring mitochondrial ROS levels in human samples. Several mitochondrial enzymes can catalyse the univalent reduction of oxygen to superoxide (O_2^-). Superoxide levels in the mitochondrial matrix can be inferred by measuring the activity of the tricarboxylic acid cycle enzyme: aconitase. Superoxide directly and rapidly inactivates aconitase. Manganese superoxide dismutase (MnSOD) activity (see cheat code 4) dictates the concentration of matrix superoxide, playing an important role in controlling its direct reactions with other species, such as aconitase, and the production of hydrogen peroxide (H_2O_2). Hydrogen peroxide primarily reacts with antioxidant enzymes, such as PRDX, transition metal ions, and protein cysteine residues. The concentration of hydrogen peroxide in the mitochondrial matrix is mainly controlled by the activities of the NADPH-dependent glutathione (GSH) and thioredoxin (TRDX) redox systems, which are fuelled by the activity of the transhydrogenase, isocitrate dehydrogenase, and the malic enzyme. To infer matrix hydrogen peroxide homeostasis one can measure PRDX3-specific cysteine oxidation using the dimer assay whereby the oxidised, disulfide bonded homodimeric form, migrates at a higher molecular weight than the reduced, monomeric form, in the absence of the reducing agent, 1-4-dithiothreitol (DTT).

Second, hydrogen peroxide, and species with an O-O bond like peroxynitrite and LOOH (Cardozo et al., 2023), react fast with PRDX isoforms (Armas et al., 2019; Bryk et al., 2000; Karplus, 2015). Their reaction with the catalytic cysteine of most PRDX isoforms produces a sulfenic acid,

which condenses with a neighbouring cysteine, to form a disulfide crosslink (Peskin et al., 2007; Wood et al., 2003). The crosslink covalently staples two monomers together, forming a dimer. Sometimes hydrogen peroxide reacts with the sulfenic acid to form an “over-oxidised” sulfinic acid (Biteau et al., 2003; Peskin et al., 2013), with further reactions yielding sulfonic acids (Akter et al., 2018). One can readily measure PRDX1-3 oxidation by non-reducing immunoblot, the so-called dimer assay, and over-oxidation of multiple isoforms by conventional immunoblotting (Cobley and Husi, 2020).

These techniques have been used to great effect in humans. For example, the Jackson group deployed them to infer exercise-induced ROS production in human skeletal muscle (Pugh et al., 2021; Stretton et al., 2020). These assays provide an integrated readout of the activity of a key antioxidant enzyme and the levels of ROS, especially when combined with similar assays for the thioredoxin redox system (Bersani et al., 2002). The ubiquity of PRDX enzymes means the assays can be applied systemically and in tissues (Low et al., 2006). The amount of PRDX2 (Low et al., 2008) in erythrocytes may support fingertip-based oxidative stress testing.

Cheat code 3. How to hack ‘TAC’ in human samples

As Sies remarked (Sies, 2020), the term total antioxidant capacity (TAC) originated from the ability of plasma to handle an ectopic redox stimulus: the azo-initiator-induced peroxy radical production (Wayner et al., 1985). Given the lack of most antioxidant enzymes (in a redox active form) in plasma (Halliwell and Gutteridge, 2015), the assay can test non-enzymatic antioxidant capacity (NEAC) against specific free radicals in plasma (Bartosz, 2010). If TAC is used judiciously, it is possible to gain some, albeit limited, insights. To hack TAC in humans, consider that:

- It is an artificial redox challenge imposed on ex vivo biological material and may have questionable relevance to the ability of said material to ‘defend’ against other species, such as superoxide.
- It can be useful with aqueous antioxidants, like vitamin C, in so far as confirming nutrient loading, when combined with assays to measure the nutrient content, and potential for redox-activity. The potential is non-equivalent to the actual activity.
- The actual antioxidant activity of blood plasma will be influenced by erythrocytes and surrounding tissues, such as the endothelium.
- There are many commercial ‘kits’ for TAC. Please carefully consider their use and properly report their information. Statements like ‘TAC was measured with X-kit’ without detailing the procedure are discouraged.
- Use other assays to better interpret TAC in plasma (see cheat codes 4-9) and refrain from measuring it in tissues—as a general rule one would be better advised to measure antioxidant enzymes.
- Low-molecular weight ‘antioxidants’ also contribute to TAC activity. For example, peroxy radicals react fast with cysteine, such as the micromole levels of albumin cysteine in plasma. Free radical chain reactions generate other ROS, such as superoxide (Winterbourn, 1993; Winterbourn and Metodiewa, 1994).
- The word total, unless carefully qualified (as in non-enzymatic capacity against peroxy radical), is a misleading misnomer (Sies, 2007).

Cheat code 4. How to measure antioxidants in human samples

As a sizeable exercise literature reviewed elsewhere attests (Powers et al., 2022, 2020, 2016), antioxidants can be measured in human samples using established methods (see Table 2).

Table 2. Methodological approaches for measuring antioxidants in human samples.

<i>n</i>	Name	Description
1	Enzyme activity	The basis of modern SOD assays (Peskin and Winterbourn, 2017)—intercepting xanthine oxidase-induced superoxide before it reduces a reporter molecule like cytochrome C—were in place before SOD was discovered! (McCord and Fridovich, 1968). Many valid microplate and gel-based SOD assays exist. They can be used to quantify SOD

2	Surrogate markers	<p>activity and, with appropriate controls, discern between isoforms (Beauchamp and Fridovich, 1971; Crapo et al., 1978; Misra and Fridovich, 1972). Glutathione peroxidase (GPX) activity and CAT activity can be assessed using established assays (Halliwell and Gutteridge, 2015; Stolwijk et al., 2020). Meanwhile, PRDX and thioredoxin activities can be inferred via non-reducing immunoblotting. Assays to quantify thioredoxin reductase or glutathione reductase activity are available (Arnér, 2017; Dagnell et al., 2017).</p> <p>Antioxidant enzyme activity can be inferred using surrogate markers (Cobley et al., 2017). For example, PRDX1 activity can reflect the phosphorylation state of the enzyme (Woo et al., 2010). Other useful post-translational surrogates include manganese SOD acetylation state, disulfide bond formation in copper zinc SOD, and succinylation of GPX4 (Kerins et al., 2018). Measuring the concentration of a protein by immunoblotting or ELISA, in combination with other markers, can be used to infer a change in antioxidant activity (Gomez-Cabrera et al., 2021).</p>
3	Glutathione	<p>High performance liquid chromatography (HPLC)-based methods are available for measuring reduced (GSH), oxidised (GSSG) and total (GSH + GSSG) glutathione (Giustarini et al., 2003; Murphy et al., 2022). Although HPLC is preferred, plate and kit-based assays are available. Controls (see cheat code 8) can prevent artificial cysteine oxidation (Poole et al., 2020).</p>
4	Low molecular weight molecules	<p>HPLC-based methods can measure the concentration of low molecular weight antioxidants, like vitamin C (Margaritelis et al., 2023). EPR spectrometry can measure the vitamin C and E radicals to provide information on their redox activity in human samples (Bailey et al., 2022).</p>

Curated do and don't guidelines for measuring antioxidants in human samples include:

- Do consider the assay biochemistry. For instance, some SOD assays are prone to artefacts arising from other molecules able to reduce cytochrome c and the complex biochemistry of assay reporter molecules, such as nitro-blue tetrazolium (Beyer and Fridovich, 1987).
- Do quantify the systemic release of antioxidant enzymes by ELISA and immunoblot (Salzano et al., 2014), especially in exosomes (Lisi et al., 2023). Don't measure GSH or antioxidant enzyme activity (e.g., GPX) in plasma/serum (Cobley et al., 2017). The concentration of GSH, glutathione reductase, and NADPH needed to sustain appreciable plasma GPX activity is minimal.
- Do use HPLC, with appropriate controls to block artificial oxidation (e.g., *N*-ethylmaleimide or iodoacetamide (Hansen and Winther, 2009)), to quantify GSH and GSSG. Don't read too much into the thermodynamic reducing potential of the redox couple (GSH/GSSG) as computed using the Nernst equation (Flohé, 2013).
- Do follow best practice for quantitative immunoblotting using validated antibodies (Janes, 2015; Marx, 2013).
- Do consider the possibility that enzyme activities measured *ex vivo* may not reflect what is possible *in vivo*. For example, for thioredoxin reductases much would depend on the continual supply of NADPH (Jones and Sies, 2015).
- Do consider that there is no one true 'best' antioxidant (i.e., there is no one master antioxidant ring to rule them all).

Cheat code 5. How to measure lipid peroxidation in human samples

Pioneering work on lipid peroxidation (see Figure 5), measured by expired pentane gas, is widely credited with founding the exercise redox biochemistry field (Dillard et al., 1978). Decades later how to measure lipid peroxidation, however, remains a vexing question. The vexing nature stems from the biochemical complex lipid peroxidation process generating manifold primary products, which are then metabolised to myriad secondary and tertiary products. While many context-specific factors influence what the 'best' approach is, we suggest approaches to measure lipid peroxidation in human samples (see Table 3).

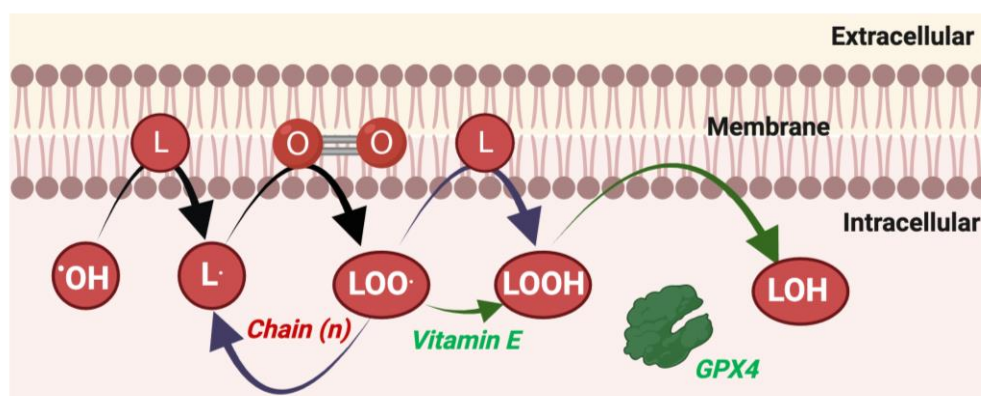


Figure 5. Start: Specific ROS, certain free radicals like hydroxyl radical or protonated superoxide (Bielski et al., 1985), react with unsaturated lipids to generate a lipid radical (L·). Chain: L· rapidly react with oxygen to form peroxy radicals (LOO·). LOO· can react with another lipid to reform L· and produce LOOH. *This is an example of free radicals reacting fast together—oxygen and L· combine to form a non-radical (LOOH) and a free radical (L·).* End: A chain-breaking antioxidant, like ubiquinol, vitamin E or even certain persulfides (Barayeu et al., 2022), break the chain by intercepting LOO·. Enzymes metabolise the resultant LOOH species, most notably glutathione peroxidase 4 (GPX4) (Yang et al., 2014), usually converting them to LOH species.

Table 3. Methodological approaches for measuring lipid peroxidation in human samples.

<i>n</i>	Name	Description
1	Lipidomics	The discovery of ferroptosis—lipid peroxidation and iron dependent cell death (Bayır et al., 2020; Dixon et al., 2012; Jiang et al., 2021)—spurred interest in sophisticated technologies for measuring the myriad of oxidised lipid products at scale using mass spectrometry (MS) (Kagan et al., 2017). One can even tell apart enzymatic from non-enzymatic F ₂ -isoprostanes (Erve et al., 2015, 2017). Commercial lipidomic analytical services are available.
2	HPLC	HPLC can measure lipid peroxidation products (Halliwell and Gutteridge, 2015), such as F ₂ -isoprostanes and MDA. In particular, HPLC analysis of MDA is a valid and highly sensitive biomarker of exercise-induced lipid peroxidation (Spirlandeli et al., 2013).
3	ELISA	ELISA kits can measure some lipid peroxidation products (Halliwell and Lee, 2010), notably F ₂ -isoprostanes. While they can be insufficiently specific in some cases (Nikolaidis et al., 2011), they have illuminated individual responses to exercise in humans (Margaritelis et al., 2014; N.V. Margaritelis et al., 2016; Margaritelis et al., 2018, 2023).
4	LOOH	The ferrous oxidation–xylenol orange (FOX) assay developed by Wolff (Wolff, 1994) can measure lipid hydroperoxides (LOOH). It can be combined with assays for lipid soluble antioxidants, such as vitamin E (Williamson et al., 2020b).

Specificity concerns can plague the FOX assay when certain types of highly oxidised lipids are analysed (Yin and Porter, 2003).

5 **Immunoblot** Some lipid peroxidation products, such as 4-hydroxynonenal (4-HNE), react with DNA and proteins (Zhang and Forman, 2017). Antibodies recognising 4-HNE-conjugated protein epitopes, formed secondary to Michael addition reactions, are available (Waeg et al., 1996). The global immunoblots can infer lipid peroxidation in human samples.

6 **Targeted** In a variant of 5, a specific protein can be immunopurified for targeted analysis of protein-specific lipid peroxidation (James. N. Cobley and Davison, 2022; J. N. Cobley and Davison, 2022). For example, by measuring a mitochondrial protein located near a membrane, this approach can assess organelle-specific lipid peroxidation.

Cheat code 6. How to measure protein oxidation in human samples

Numbers illustrate the formidable challenges of measuring protein oxidation per se let alone in human samples. A typical cell contains about 50% (~10,000) of the proteins encoded by the human genome (Consortium et al., 2001; Slavov, 2022, 2020). The susceptibility of virtually every proteogenic amino acid to oxidation (Kitamura and Galligan, 2023) can generate many, over 10^{10} , redox proteoforms (Aebersold et al., 2018). A proteoform specifies different molecular forms of the protein inclusive of post-translational modifications (Smith et al., 2013; Smith and Kelleher, 2018). Proteoform copy numbers can vary by $\sim 10^6$. Such numbers make it exceedingly difficult to identify and quantify billions of possible redox proteoforms (Brady and Meyer, 2022). While many amino acids can be oxidised (cheat codes 8 & 9 consider cysteine), most studies measure modified forms of tyrosine—3-nitrotyrosine (3-NT)—and protein carbonylation, which affects multiple amino acids, such as lysine residues (Davies, 2016; Hawkins et al., 2009; Hawkins and Davies, 2019). To measure protein oxidation in human samples, we suggest several approaches (see Table 4).

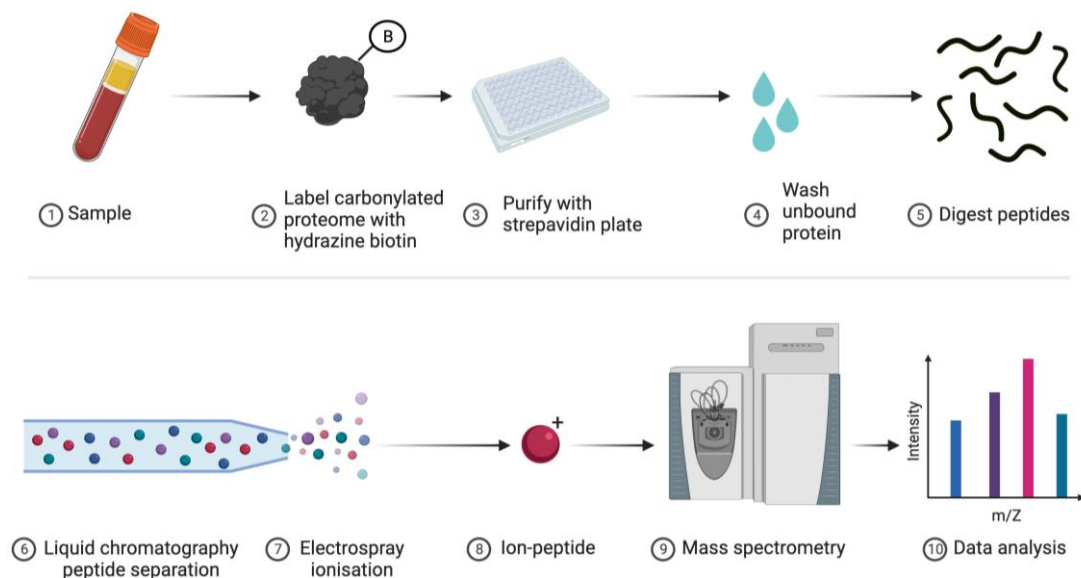


Figure 5. Proteomic-based analysis of carbonylated proteins. 1. A biological sample is obtained. 2. Carbonylated proteins are labelled with reducing agent cleavable hydrazine-conjugated biotin. 3. The carbonylated proteome is separated from the rest of the sample via a streptavidin solid support, such as a 96-well microplate or superparamagnetic beads. 4. Unbound protein is removed by extensive washing. 5. After releasing the purified proteins using a reducing agent, trypsin is used to digest the bound proteins into peptides. 6. HPLC separates the peptides by their hydrophobicity before electrospray ionisation (7-8) and subsequent mass spectrometry analysis (9-10). In principle, this

approach can determine the identity of the oxidised proteins and the amino acid residues that are oxidised.

Table 4. Methodological approaches for measuring protein oxidation in human samples.

<i>n</i>	Name	Description
1	Proteomics	One can collaborate with specialist labs or access services to identify and quantify specific types of oxidised amino acid, such as carbonylated proteins (see Figure 5), on a proteome-wide scale using bottom-up MS (Aebersold and Mann, 2003). Sophisticated modification specific workflows are available (Aldini et al., 2015; Batthyány et al., 2017; Bollineni et al., 2014; Fedorova et al., 2014; Madian et al., 2011; Madian and Regnier, 2010).
2	ELISA	Simple and user-friendly ELISA kits can quantify total protein carbonylation (Buss et al., 1997).
3	Immunoblot	Pan-proteome immunoblots with modification specific antibodies or derivatisation reagents can be performed (Cobley et al., 2014). The OxyBlot™ for protein carbonylation represents an enduringly popular approach (Frijhoff et al., 2015).
4	Fluorescent	Derivatising protein carbonyl groups with fluorophores, such as rhodamine-B hydrazine, allows for quantifying their levels via SDS-PAGE (Georgiou et al., 2018; Weber et al., 2015), especially when protein content can be normalised with a spectrally-distinct amine-reactive probe like AlexaFluor™647-N-hydroxysuccinimide (F-NHS). Novel N-terminal reactive reagents, such as 2-pyridinecarboxyaldehyde, may also be used (Bridge et al., 2023; MacDonald et al., 2015).
5	Targeted	Specific proteins can be analysed by 1-4, such as MS for residue level analysis (Cobley et al., 2019b), when a protein is immunopurified. Targeted approaches can address specific questions (Place et al., 2015; Safdar et al., 2010), especially when the functional impact of the oxidation event is known (see Figure 6). For example, tyrosine 34 nitration impairs manganese SOD activity by electrically repelling superoxide (MacMillan-Crow et al., 1996). Electrostatic repulsion helps explain why the rate of superoxide dismutation via $O_2^{\cdot-} + O_2^{\cdot-}$ colliding to form hydrogen peroxide and oxygen is near zero (Fridovich, 1989). Approach 4 combined with an ELISA assay may support protein-specific oxidative damage analysis in a microplate.

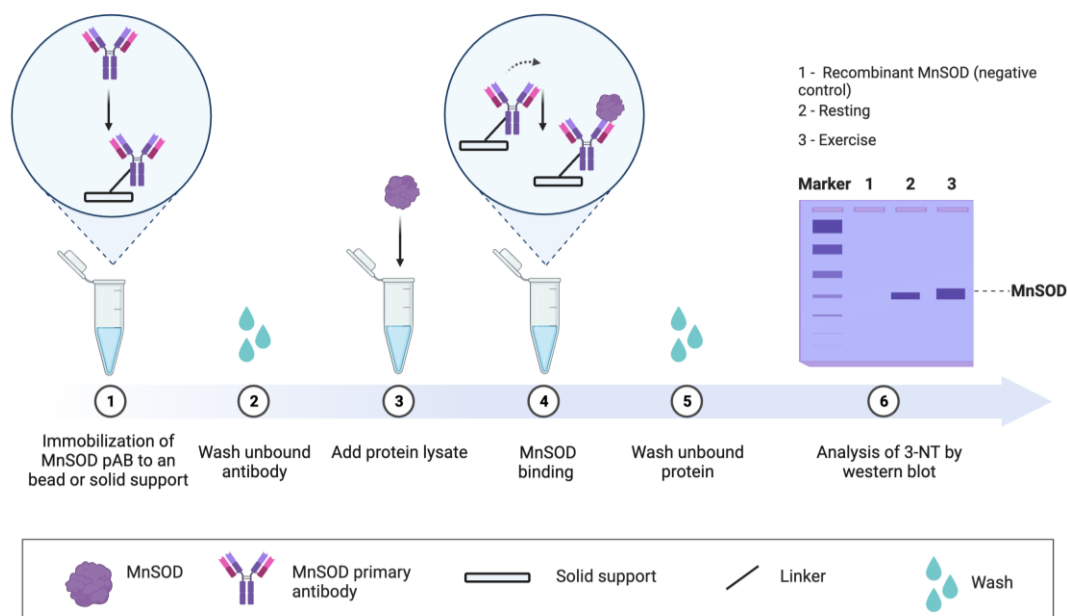


Figure 6. Analysis of MnSOD protein nitration using a targeted immunological approach. 1. An MnSOD capture antibody is bound to protein A or G coated superparamagnetic beads. 2. The unbound antibody is removed by washing. 3. A protein lysate containing MnSOD is added. 4. MnSOD binds to the capture antibody. 5. Unbound proteins are removed by extensive washing. 6. MnSOD nitration is assessed by immunoblotting using a 3-NT antibody. Recombinant SOD can be loaded as a negative control. The basic approach described above can be adapted for other proteins or inverted. In the latter, a 3-NT capture antibody is used as bait to determine whether it binds MnSOD).

Cheat code 7. How to measure DNA and RNA oxidation in human samples

DNA lesions occur at a rate of 10,000 to 1,000,000 molecular lesions per cell, per day, and if, left unrepaired can block DNA replication and transcription or lead to other serious genome aberrations (Alhmod et al., 2020). Several excellent assays exist with capability of capturing either DNA strand breaks (single or double stranded) or DNA oxidation (nucleoside base damage), and these techniques encompass molecular, fluorescence, chemiluminescence, analytical, and sequencing approaches (see Table 5).

Table 5. Methodological approaches for measuring DNA damage in human samples.

n	Name	Description
1	Fluorescence	Single cell gel electrophoresis (SCGE), or the comet assay, is a technically simple and sensitive method for quantifying single and double-stranded DNA breaks in mononuclear cells. The SCGE assay embeds cells on an agarose gel and if supercoiling is relaxed due to a single or double break, the loop of the DNA is free to migrate during electrophoresis and thereby form a comet tail (Davison, 2016). The SCGE assay can resolve damage up to ~3 breaks per 10 ⁹ Da (Collins and Azqueta, 2012). Several variations of the SCGE assay exist. As oxidation of DNA can occur at a similar rate to strand breaks, base oxidation can be examined via a simple SCGE modification using lesion-specific enzymes (e.g., formamidopyrimidine DNA glycosylase or endonuclease III) to accurately capture oxidised purines (guanine/adenine) and/or pyrimidines (thymine/cytosine). The assay may be combined with fluorescent in situ hybridization (FISH) to detect whole genome, telomeric and centromeric DNA and gene region-specific DNA damage (Davison, 2016). CometChip technology

is now utilised to minimise a high sample-to-sample variation (Sykora et al., 2018).

2 ELISA

ELISA application is a popular immunological method to measure DNA damage, mainly in the form of 8-oxodG. There are several commercially available kit-based options, however caution is paramount regarding assay specificity and reliability.

3 Molecular

Polymerase chain reaction (PCR) or quantitative PCR (qPCR) can map nuclear and mitochondrial DNA damage at nucleotide resolution (Li and Sancar, 2020). In PCR, DNA amplification is stalled at the damaged site via the blocking of the progression of Taq polymerase, which ultimately reduces the number of DNA templates that are devoid of any Taq-blocked lesions (Figuroa-Gonzalez and Perez-Plasencia, 2017). qPCR quantifies DNA damage on both duplex strands. It is possible to quantitatively detect and analyse gene-specific DNA damage (and repair) using qPCR and with only 1-2 ng of total genomic DNA. qPCR has caveats: it depends on high-molecular weight DNA, well defined qPCR conditions, and the intricate calculation of lesion frequencies (Figuroa-González and Pérez-Plasencia, 2017; Santos et al., 2006). Long Amplicon-Quantitative PCR (LA-qPCR) can also provide an overview of total genomic mitochondrial DNA damage (Williamson et al., 2020b). Following double stranded DNA damage, a repair response is usually initiated, where the subsequent phosphorylation of Serine-139 of histone H2AX ensues (Williamson et al., 2020a). The γ H2AX assay is relatively simple to execute and is based on immunofluorescence using a specific Serine-139- γ H2AX antibody to show location in chromatin foci at the sites of DNA damage (Figuroa-Gonzalez and Perez-Plasencia, 2017).

4 Analytical

HPLC coupled with tandem MS is reliable at detecting oxidative DNA damage (e.g., 8-hydroxy-2'-deoxyguanosine) with excellent separation of nucleosides. HPLC-MS yields robust information on location of DNA damage, but high assay cost and required extensive experience can preclude assay use (Figuroa-Gonzalez and Perez-Plasencia, 2017). Gas chromatography coupled with mass spectrometry (GC/MS) is highly sensitive to the detection of several forms of DNA damage including those of the sugar moiety and four hetrocyclic bases (e.g., 8-oxodG, 5-HMUrA, 8-oxoAde, 5-OHUrA, 8-oxoGua; (Ravanat et al., 1999). Although the technique provides impressive structural data in complex samples (such as the detection of a single DNA lesion in DNA with several lesions) the quantification of nucleoside forms of base damage is not as robust compared to liquid chromatography-based methods (Taghizadeh et al., 2008).

5 Sequencing

Innovative next-generation sequencing technology now exists, providing high-throughput and high accuracy DNA sequence data. RADAR-Seq (Zatopek et al., 2019), qDSB-Seq (Zhu et al., 2019), and AP-Seq (Poetsch et al., 2018) are typical examples of sequencing-based technologies designed to quantify and map DNA damage on a genome-wide scale. This approach determines the precise gene locations of DNA damage. Interestingly, the quenching of fluorophores on account of the low redox potential of guanine had to be addressed before next-generation sequencing technologies could be developed (Rodriguez and Krishnan, 2023).

Cellular RNA damage is far more abundant than DNA damage, however, only a few in vivo RNA oxidation indices have been reported. The primary by-product of RNA oxidation, 8-oxoGuo, is chemically similar to 8-oxodG, and this can cause interpretational issues if the correct analysis tool

and sample type are insufficiently considered (Larsen et al., 2022). The principles for analysis of oxidised RNA are based on the following:

- Assay techniques: The validated analytical tools relating to DNA oxidation are applicable to RNA oxidation, as the latter is quantified using ELISA, PCR-based technology, and chromatograph procedures, such as HPLC with electrochemical potential detection (Floyd et al., 1986) and liquid chromatography/mass spectrometry or gas chromatography/mass spectrometry (Dizdaroglu et al., 2001; Larsen et al., 2019). Note, most ELISA kits cannot discriminate between RNA and DNA oxidation products (Larsen et al., 2022).
- Sample type: RNA oxidation can be quantified in urine, blood and/or tissue (cells). Detection of 8-oxoGuo urinary excretion is possible but must be corrected for urine dilution (via urine volume, creatinine or density). Blood plasma is an acceptable material to measure RNA oxidation, although the data should be carefully interpreted with appropriate physiological modelling (Larsen et al., 2019; Poulsen et al., 2019). Tissue quantification has the advantage of tissue-specific interpretation, unlike plasma and urine collection (Larsen et al., 2022).

Cheat code 8. How to measure redox regulation in human samples

While the redox regulation (see box 1) literature dates back to at least 1966 (Jacob and Jandl, 1966) with reviews on the subject published in the 1980's (Saran and Bors, 1989), the turn of the millennium (GUTTERIDGE and HALLIWELL, 2000), marked a seismic shift in how the field interpreted exercise-induced oxidative stress (Richardson et al., 2007). Building on early work (Gómez-Cabrera et al., 2003; Khassaf et al., 2003; Viña et al., 2000), landmark studies demonstrated that nutritional antioxidants impaired beneficial molecular adaptations to exercise in humans (Gomez-Cabrera et al., 2005; Gomez-Cabrera et al., 2008; Ristow et al., 2009). To measure redox regulation in humans (Henriquez-Olguin et al., 2020; Margaritelis et al., 2020; Mason et al., 2020), we suggest multiple approaches (see Table 6).

Box 1. Brief overview of the redox regulation concept.

Redox regulation defines the process whereby ROS-sensitive protein- and cysteine-residue specific redox changes regulate protein function (D'Autréaux and Toledano, 2007; Holmström and Finkel, 2014; Sies and Jones, 2020). A paradigmatic example originating in the 1990's concerns the tyrosine phosphatase PTP1B (Pregel and Storer, 1997). The active site cysteine, Cys215, of PTP1B acts as a nucleophile—a chemical species that forms bonds by donating an electron pair—to dephosphorylate protein tyrosine. To act as a nucleophile, the sulfur atom in Cys215 must be in a reduced, deprotonated state (i.e., RS⁻). ROS can inactivate PTP1B by disabling nucleophilic catalysis (Salmeen et al., 2003). For example, hydrogen peroxide can react with Cys215 to form a sulfenic acid. As an electrophile—a biochemical species, such as a Lewis acid, that seeks out electron pairs for bonding—the sulfenic acid prevents the cysteine from launching a nucleophilic attack on a phosphate bond (Allison, 1976; Paulsen and Carroll, 2013). Reviews on the mechanisms, biological function, and open questions in the redox-regulation field are available elsewhere (Antunes and Brito, 2017; Brigelius-Flohé and Flohé, 2011; D'Autréaux and Toledano, 2007; Holmström and Finkel, 2014; Lennicke and Cochemé, 2021; Marinho et al., 2014; Parvez et al., 2018; Sies and Jones, 2020; Winterbourn and Hampton, 2008).

Table 6. Methodological approaches for measuring redox regulation in humans.

<i>n</i>	Name	Description
1	Proteomics	Sophisticated MS approaches, such as cysteine-reactive phosphate tag technology (Xiao et al., 2020), can identify and quantify cysteine oxidation with residue-resolution on a proteome-wide scale (Day et al., 2021; Kim et al., 2015; Leichert et al., 2008; Li et al., 2023; Yang et al., 2016). Modern methods

allow deep coverage of the cysteine proteome, such as ~34,000 residues across ~9,000 proteins (Xiao et al., 2020). For reference, the full human proteome contains over 200,000 cysteine residues distributed across over 18,000 proteins (Go et al., 2015). New advances provide deep coverage and better quantification (Derks et al., 2022; Sinitcyn et al., 2023). Protein targeted methods are also possible (Held et al., 2010; Malik et al., 2013). Proteomic services, some include data analytical packages, are available.

Approaches (see Figure 7) include immunocapture before streptavidin immunoblotting of biotin-conjugated oxidised cysteines reports target-specific cysteine oxidation (Boivin et al., 2008). Non-reducing immunoblotting quantifies the oxidation of some proteins, such as protein kinase G (Burgoyne et al., 2007). For the many proteins that fail to exhibit endogenous oxidation-induced mobility shifts, cysteines can be derivatised with mobility-shifting polyethylene glycol (PEG)-payloads (Burgoyne et al., 2013; Cobley et al., 2020, 2019a; Lee and Chang, 2019; Leeuwen et al., 2017; Makmura et al., 2001). These assays quantify cysteine redox proteoforms (Cobley, 2023b).

2 Immunological

Some assays can infer the outcomes of redox regulation without measuring cysteine oxidation (Ostrom et al., 2021a, 2021b; Ostrom and Traustadóttir, 2020). Transcriptional approaches, such as qPCR analysis, can infer the activation of redox-sensitive gene expression programmes, notably Nrf2* activity. Immunoblot approaches include (1) degradation of KEAP1* to infer Nrf2 activity, (2) monitoring a reporter, such as the phosphorylation of a signalling protein, and (3) quantifying protein content, such as heat shock proteins (Cobley et al., 2015c). Changes in antioxidant enzyme activity, glutathione, or oxidative damage may also be instructive.

3 Outcomes

*Discovered in 1994 (Moi et al., 1994), Nrf-2 is a transcription factor responsible for inducing the expression of over 300 genes with many of them being linked to antioxidant defence, such as the gene encoding the rate-limiting step in glutathione biosynthesis—gamma glutamate cysteine ligase subunits (Yamamoto et al., 2018). Although the mechanisms are complex, nrf-2 is inhibited in the cytosol by KEAP1. Cysteine oxidation of KEAP1 release nrf-2 to translocate to the nucleus (Dinkova-Kostova et al., 2002).

Selected do and don't guidelines for measuring redox regulation in human samples include:

- Do minimise artificial cysteine oxidation by alkylating reduced cysteines with suitable reagents, such as *N*-ethylmaleimide (NEM). Note these reagents can label other amino acids, react with sulfenic acids, and persulfides (Hansen and Winther, 2009; Reisz et al., 2013; Schilling et al., 2022).
- Do bear in mind that not every protein and every cysteine is yet measurable in one run using MS technology (Timp and Timp, 2020).
- Do consider that different workflows measure different forms of cysteine oxidation, so-called chemotypes. For example, a chemotype-specific proteomic approach demonstrated that fatiguing exercise increased S-glutathionylation, cysteine covalently attached to glutathione via a disulfide bond, in mice (Kramer et al., 2018).
- Do consider supplementing the analysis with global readouts of cysteine oxidation, such as an Ellman's test (Ellman, 1959), or chemotype-specific pan-proteome immunoblots (Saurin et al., 2004).
- Do consider that many techniques don't measure "over-oxidised" chemotypes, such as sulfinic acids.
- Don't assume a technique will necessarily work! Mobility-shift immunoblots usually fail to detect the protein because the bulky PEG-payloads sterically block primary antibody binding.

- Don't assume cysteine oxidation is functional without evidence.
- Don't assume the cysteine oxidation is necessarily oxidative eustress without evidence.
- Don't assume an outcome assay result is caused by cysteine oxidation without evidence.

Immunological techniques

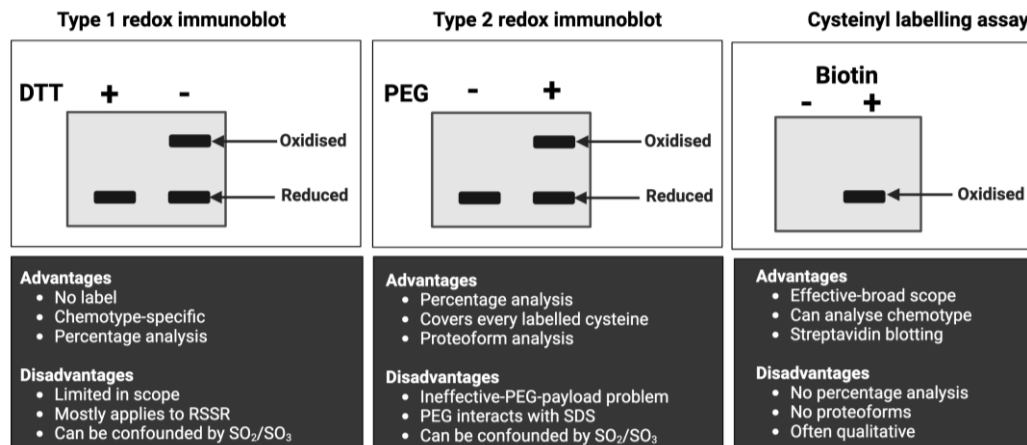


Figure 7. Visual overview of the main macroscale (slab-gel formatted) immunological techniques for measuring protein-specific cysteine redox state in human samples. See Table 6 for specific details. Their main advantages and disadvantages are listed for reference. Abbreviations: DTT = 1,4-dithiothreitol, PEG = polyethylene glycol, RSSR = disulfide bond; SO₂ = cysteine sulfinic acid; SO₃ = cysteine sulfonic acid, SDS = sodium dodecyl sulfate.

Cheat code 9. How to use redox ELISA technology to measure protein cysteine oxidation in humans

Simple and effective ELISA-based fluorescent immunoassays can measure target-specific cysteine oxidation, such as the antibody-linked oxi-state assay (ALISA) and RedoxiFluor (Noble et al., 2021; Tuncay et al., 2022). They use redox state encoded fluorophores, such as AlexaFluor™647-C₂-maleimide (F-MAL), to quantify cysteine oxidation of an immunopurified protein in a microplate (see Figure 8). ALISA/RedoxiFluor bring the benefits of the microplate format, such as high-throughput analysis, to the redox regulation field (see Table 7). They open up new and exciting 'off-the-shelf' opportunities for measuring redox regulation in human samples.

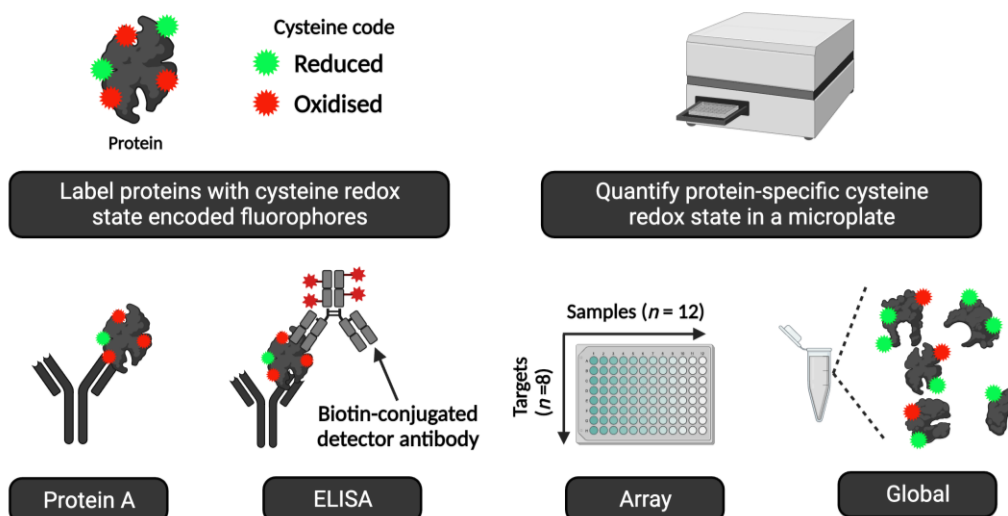


Figure 8. Visual overview of RedoxiFluor. In the upper left panel, reduced and oxidised cysteines in the sample are labelled with spectrally distinct fluorophores. The spectrally distinct fluorophores enable cysteine redox state to be quantified in a number of ways in a microplate (upper right panel).

As depicted in the bottom panel, four different variations of the assay can be used. From left to right. First, a protein-A coated microplate can be used to enrich a protein target of interest using a suitable capture antibody. Second, a conventional ELISA comprising a capture and detector antibody pair can be used to quantify relative target-specific cysteine redox state in percentage and moles. Third, either protein A or an ELISA mode can be deployed to measure the redox state of multiple targets in several samples in an array mode assay. Finally, the global cysteine redox state of the proteome can be determined in a microplate.

ALISA and RedoxiFluor can shed light on exercise- and nutrient-sensitive cysteine oxidation (James Nathan Cobley, 2020). For instance, ALISA revealed how acute maximal exercise decreased cysteine oxidation of the catalytic core subunit of the serine/threonine PP2A protein phosphatase in human erythrocytes (Muggeridge et al., 2022). A follow-up benchmarking study demonstrated excellent assay performance across multiple analytical metrics, from accuracy to reliability (Tuncay et al., 2023). Further, maximal exercise increased the cysteine oxidation of the redox regulated GAPDH protein (Talwar et al., 2023). Visually displayed orthogonal assays, such as gel-based ALISA (Bellissimo et al., 2023), confirmed the validity and specificity of the ‘unseen’ microplate data. Selected points relevant to applying redox ELISA technologies in human samples include:

- Standard operating procedures are available (Tuncay et al., 2022). The cysteine labelling procedures can be adapted to suit specific experimental needs. For example, to omit some costly preparatory steps, reduced cysteines can be labelled with an F-MAL in ALISA. In this case, increased cysteine oxidation would decrease the observed F-MAL signal.
- The assays can operate in different modes, from global (i.e., all proteins/no antibodies) to multiparametric array mode, in microscale and macroscale (e.g., slab-gel format).
- In some cases, the assays provide information on protein function, such as the difference in transcription factor cysteine redox states in the cytosol vs. the nucleus.
- Interpretationally, a change reflects a difference in the rate of ROS-sensitive cysteine oxidation and antioxidant-sensitive reduction across the entire protein. The summed weighted mean of all individual residues. Responding to both ROS and AOX inputs is useful.
- The assays are compatible with chemotype-specific* cysteine labelling (Alcock et al., 2017) and direct-reactivity approaches (Shi and Carroll, 2020). For example, the methods are compatible with reaction-based sulfenic acid fluorophores (Ferreira et al., 2022).
- Unless the target has one cysteine like ND3-Cys39 in complex I (Burger et al., 2022; Chouchani et al., 2013), the assays cannot disclose the identity of the oxidised cysteine residues.
- For some exercise-sensitive proteins, such as PGC-1 α (Cobley et al., 2012; Egan and Sharples, 2023; Egan and Zierath, 2013), the lack of an ELISA kit may make it impossible to run the assay.

*Relevant to cheat code 8-9, the specificity of some chemotype approaches is actively debated (Bischoff et al., 2023; Forman et al., 2017; Stöcker et al., 2018). For example, vitamin C can reduce S-nitrosated cysteine residues but it also reacts with sulfenic acids (Anschau et al., 2020).

Table 7. Type-stratified benefits of the ELISA formatted fluorescent immunoassays: ALISA and RedoxiFluor.

Type	Benefit	Description (useful property as applicable)
ELISA	Throughput	High sample n-plex analysis (adds statistical power)
	Multiplexed	Parallel assessment of multiple 2-10 proteins (enables screening)
	Sensitive	Picomole sensitivity (supports human biomarker studies)
	Rapid	Performed in 1 day with minimal hands-on time (benefits screens)
Redox	Cysteine holistic	Agnostic of any one cysteine residue (adds coverage of the entire molecule)
	Percentages	
	Moles	Quantifies cysteine redox state in percentages (interpretational useful)
	Context	Quantifies cysteine redox state in moles (interpretational useful)

	Chemotype	Provides cysteine proteome context (interpretationally use)
	Process-sensitive	Support chemotype-specific analysis (supports mechanistic studies) Results are sensitive to oxidative and antioxidative processes scaled across every cysteine residue on the target protein (interpretationally useful)
		Draws on highly-principled redox and immunological methods (robust)
Performance	Valid	They work (e.g., compare to Click-PEG) (means to study the specific protein)
	Effective	
	Accurate	Data correspond to ground-truth standards (adds percentage analysis)
	Reliable	High consistency between samples (adds robustness)
	Reproducible	Delivers consistent results (adds robustness)
	Range	Operates across a large dynamic range (useful for human applications)
Microplate	Simple	Simple to understand, interpret, and operate (supports accessibility)
	Easy-to-do	Little technical skill required to deliver actionable results (accessibility)
	Off-the-shelf	Compatible with commercial ELISA kits (accessibility)
	Automated	Delivers rapid and automated data within seconds (time-efficient)

Cheat code 10: How to exploit mathematical modelling and computational analyses in redox biology

So far, we have presented cheat codes for measuring oxidative stress using experimental methods in humans. However, some questions in biology cannot be tackled using experimental methods alone and may require the aid of theoretical approaches. Systems biology investigates the dynamic properties and interactions within a biological object, at cellular and/or organismal level, in a qualitative and quantitative manner by combining experimental and mathematical approaches (Kitano, 2002; Mogilner et al., 2006). The iterative cycle of data-driven modelling and model-driven experimentation, in which hypotheses are formulated and refined until they are validated, can elucidate the emergent properties and mechanisms governing cellular processes and higher-level phenomena (Epstein, 2008; Wolkenhauer, 2014). Such analyses are used in redox biology to quantitatively characterise redox processes and link them with physiological outputs (Margaritelis et al., 2022). For a comprehensive review, the reader is referred elsewhere (Antunes and Brito, 2017; Buettner, 2011; Held, 2020; Pillay et al., 2013; Salvador, 2024; Salvador and Antunes, 2023)

Here, we highlight quantitative studies that have interesting implications for the production, metabolism and signalling properties of specific ROS (Huang and Sikes, 2014; Lancaster, 1997; Langford et al., 2018; Lim et al., 2008, 2016, 2015; Orrico et al., 2018; Travasso et al., 2017; TSOUKIAS, 2008; Winterbourn, 2008). Antunes and Brito formulated a minimal model of hydrogen peroxide (Brito and Antunes, 2014) and provided simple equations that can be used in combination with experimental measurements to estimate the response time and oxidation profile of cysteine-based redox switches, providing insights into the mechanisms of redox signalling. Pillay and colleagues fitted experimental data in computational models and performed a supply-demand analysis for hydrogen peroxide, that is, the production and consumption (Pillay et al., 2016). Their analysis showed that the activities of enzymes responsible for hydrogen peroxide consumption can act synergistically in the face of increasing hydrogen peroxide supply, to limit its steady state concentration. A recent computational study investigated how far superoxide and hydrogen peroxide can travel through capillaries, arterioles, and arteries, what concentrations can be attained under different conditions, and the main determinants of these distances and concentrations (Sousa et al., 2022). A finding relevant to measuring oxidative stress in blood is that 36% and 82% of the plasma hydrogen peroxide is absorbed by the erythrocyte in the capillaries and arterioles, respectively. In another compelling example, it is commonly assumed that neutrophils can directly

oxidise bacteria and other cell structures via a short burst of superoxide and hydrogen peroxide production. However, Winterbourn et al, by modelling the reactions of superoxide and myeloperoxidase, investigated the fate of superoxide and hydrogen peroxide in the neutrophil phagosome, and provided an alternative explanation where superoxide acts as a (i) cofactor for hypochlorous acid production by myeloperoxidase, and/or as a (ii) substrate for the production of other ROS (Kettle et al., 2023; Winterbourn et al., 2006).

Simple back-of-the-envelope calculations can yield important quantitative insights into biological processes/mechanisms and provide experimentally testable predictions (Milo and Phillips, 2006; Phillips et al., 2012). Such calculations may often include simple order-of-magnitude estimates and algebraic equations. A theoretical analysis using only kinetic rates revealed that vitamin C and vitamin E are unlikely to have a major impact on exercise-induced redox signalling by scavenging hydrogen peroxide (Cobley et al., 2015b). Using the kinetics and concentrations of hydrogen peroxide it has also been shown that the frequently reported increases in hydrogen peroxide after exercise are not sufficient to induce redox signalling, except for the case of micro-domains, such as subcellular compartments or organelles (Nikolaidis et al., 2020). Finally, we used simple stoichiometric calculations to show that oxidative stress leading to metabolic reprogramming within erythrocytes controls the concentrations of key molecules, such as ATP, NADPH, and 2,3-bisphosphoglycerate (Chatzinikolaou et al., 2024). In sum, simple numerical calculations can help characterise biological phenomena quantitatively, providing a deeper understanding of a given redox system.

Part 3: Perspectives

Using the cheat codes to measure oxidative stress in humans

Unlike in mathematics, there is no universal ground-truth answer for selecting the most appropriate cheat code stratified oxidative stress assay (s) to implement in human studies (Cobley et al., 2017; Nikos V. Margaritelis et al., 2016). Among the many factors that may be accounted for (e.g., sampling time points (MICHAILIDIS et al., 2007), the following are influential:

- Whether the redox approach is general or targeted, where the general 'catch-all' approach analyses as many distinct oxidative stress processes as possible and targeted ones focus on a specific process, such as lipid peroxidation. In both cases, multiple process-specific analytical indices are preferred. Still, the depth of the analyses depends on whether oxidative stress is a primary, secondary, or tertiary biochemical outcome variable.
- The type of biological material acquired, usually blood and/or tissue samples, and the number of samples dictate what can be measured and how. For example, performing MS-based proteomic analysis on 100 samples is unlikely to be financially viable. Relatedly, the relevant equipment and expertise to undertake the analysis must be available.
- Whether (1) a redox-active molecule, such as antioxidant or pro-oxidant (Jordan et al., 2021) is being studied, (2) oxidative eustress or distress (e.g., harmful age-related DNA damage (Cobley et al., 2013)), is being studied, and (3) oxidative stress is linked to a given outcome variable, such as exercise adaptations.

We have devised a research question grounded decision tree for selecting the most appropriate cheat code (s) to implement (see Figure 9). For example, answering the question "*are you interested in oxidative damage or redox regulation?*" with "*oxidative damage*" simplifies the decision-making process by omitting cheat code 8 and 9 at the outset. From this starting point, answering other questions should prove instructive for selecting suitable cheat code stratified assays. For example, selecting indices of mitochondrial lipid peroxidation, such as organelle-specific 4-HNE immunoblotting per cheat code 5, when using a lipophilic mitochondria-targeted antioxidant to modulate exercise-induced oxidative stress (Murphy, 2014; Murphy and Hartley, 2018; Smith et al., 2003).

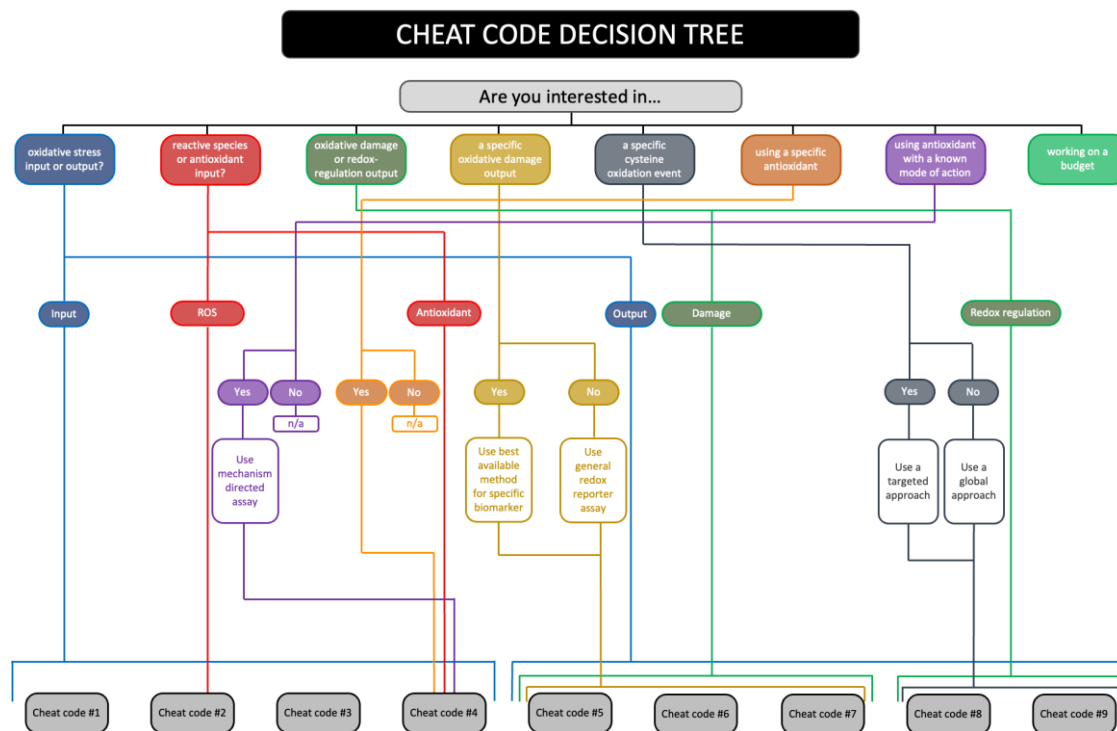


Figure 9. The research question-grounded decision tree for selecting the most appropriate cheat code (s) to implement for experimentally measuring oxidative stress in a prospective experimental human study. For example, if you are interested in a specific antioxidant, such as the mitochondria-targeted quinone (MitoQ), then one should consider implementing cheat code 4. By answering the other questions, one can construct. Note that cheat code 10 can be applied to all the answers and should be considered irrespective of the species of the study (i.e., it is widely applicable). For example, simple kinetic values for the reaction of MitoQ with different ROS would be instrumental for selecting appropriate mechanism-based assays. Please see the main text for a worked example.

Getting the NAC of it: A worked example

To demonstrate how to use the cheat codes, we define a hypothetical prospective example concerning whether NAC acts as an antioxidant to improve exercise performance (Cobley et al., 2011) by reversing ROS-induced fatigue (Reid, 2008). In this example, a ROS input (the exercise-induced redox signature) produces a fatigue output via a process (redox mechanism) involving oxidative damage to contractile proteins, such as myosin chain isoforms. Based on our answers to the questions in the decision tree (see Table 8), we might use:

- Cheat code 4 to verify increased NAC loading via HPLC-based analysis of plasma NAC. And assay GSH and GPX activity in erythrocytes or tissue lysates to infer whether NAC supports the glutathione redox system (Giustarini et al., 2023). This might be expected to alter peroxide metabolism and hence oxidative damage to proteins via lipid peroxidation products, such as 4-HNE (Zhang and Forman, 2017).
- Cheat code 5 to measure lipid peroxidation. In plasma, one might measure LOOH and 4-HNE via the FOX assay and immunoblotting, respectively. In tissue samples, one might measure 4-HNE via immunoblotting. Equally, one might implement a F₂-isoprostanes ELISA in plasma or tissue (Paschalis et al., 2018).
- Cheat code 6 to measure oxidative damage to contractile proteins using targeted analysis. Immunocapture of a specific protein followed by immunoblot analysis for 4-HNE, 3-NT, or protein carbonyls (Place et al., 2015). Like how cutting fingernails yields thiyl radicals in alpha keratin (Chandra and Symons, 1987), mechanical stress produces protein-based free radicals (Zapp et al., 2020). Hence, one could add a spin trap to 'clamp' protein radicals for targeted immunoblot analysis with an anti-trap reagent (Mason and Ganini, 2019). If only circulating

samples were available, the same approaches could be applied in these compartments to test the plausibility of NAC minimising oxidative damage to proteins (albeit non-contractile ones).

- Cheat codes 8-9 with chemotype analysis to determine whether NAC by supporting hydrogen sulfide production elicits beneficial effects by inducing contractile protein-specific persulfidation (Ezeriņa et al., 2018; Kalyanaraman, 2022; Pedre et al., 2021). If fluorescent labels are used, then cheat code 6, 8 and 9 could be implemented simultaneously (Zivanovic et al., 2019). For example, gel-based detection of persulfidation before 4-HNE immunoblotting.

Table 8. Worked example-specific answers to the research question grounded cheat code (s) selection tool.

Are you interested in the	Answer	Refinement	Selection outcomes	Assay (s)
<i>Oxidative stress input or output?</i>	Output	n/a	Consider cheat codes 5-9 Disregard cheat codes 1 & 2	n/a
<i>Reactive species or antioxidant input?</i>	Yes	Interested in an antioxidant (yes)	Consider cheat code 4 Disregard cheat codes 1-3	n/a
<i>Oxidative damage or redox regulation output?</i>	Both	n/a	n/a	n/a
<i>A specific oxidative damage output?</i>	Yes	Yes	Consider cheat code 5-6 Disregard cheat code 7 (based on mechanism)	Global 4-HNE immunoblot Contractile protein immunocapture for targeted 4-HNE analysis
<i>A specific cysteine oxidation event?</i>	Yes	Yes	Consider cheat codes 8 & 9	Gel-based analysis of persulfides using a fluorescent probe
<i>Using a specific antioxidant?</i>	Yes	n/a	Consider cheat code 4	n/a
<i>Using an antioxidant with a known mode of action?</i>	Yes	Use mechanism directed assay	Consider cheat code 4	HPLC of [NAC]

So far, we have elaborated our decisions based on the financial means to investigate two distinct modes of action in a biochemistry-guided catch-all approach. If financial or related concerns like time were limiting, the essential codes, corresponding to a minimal yet mechanistically cohesive approach, would be:

- Cheat code 4: GSH levels (systemic or tissue). Or cheat code 10 (see below).
- Cheat code 5: 4-HNE blot (systemic or tissue).
- Cheat code 6: Myosin-specific 4-HNE levels (tissue).

To appreciate the benefits of the cheat code guide, it is worth contrasting even the minimal analysis to what otherwise be performed. Without the guide, one might easily elect to measure glutathione concentrations in plasma, lipid peroxidation via TBARS, or 4-HNE levels globally as the incorrect conceptual/methodological analogues to cheat codes 4-6. It might also be speciously contended, that NAC scavenges ROS. The kinetic plausibility of this hypothesis for key ROS like superoxide and hydrogen peroxide is questionable (Pedre et al., 2021). Even if it were kinetically plausible, the products need to be considered. For example, the NAC thiyl radical can generate

superoxide! Hence, an alternative analytical selection would confound the mechanistic interpretation of the study, constraining the ability of the researchers to investigate their hypothesis.

Interpretationally, the example investigated the potential existence of a causal relationship between an exercise-induced redox input, effected by specific ROS and AOX, and a whole-body output—exercise performance—via a skeletal muscle fatigue process mediated, in part, by the oxidation of contractile proteins. If we assume the fatigue process mechanism is correct, then for NAC to exert a beneficial effect, in this way, three cheat code checkable points hold:

- NAC enters the circulation before it or a metabolite thereof accumulates in skeletal muscle (checked via cheat code 4: HPLC analysis of NAC).
- NAC indirectly acts as an antioxidant by impacting on a process that influences the oxidation of contractile proteins. The former can be checked via GSH-related lipid peroxidation analysis (cheat code 4-5) and the latter by targeted oxidative damage analysis pursuant to cheat code 6 or hydrogen sulfide donor effect per cheat code 8 or 9.
- By so doing, NAC impacts a whole-body marker of fatigue, such as exercise performance.

How one interprets oxidative stress depends on the selected outcome variable. In this case, interpreting oxidative stress analyses is conditional on a non-redox outcome: exercise performance. The same redox evidence differently could be interpreted differently (Nikolaidis and Margaritelis, 2018) as delaying fatigue to do more work delivers an immediate performance benefit but may sacrifice exercise adaptations in the long run.

Getting a NAC for the numbers

After focusing on NAC and glutathione, we provide a simple quantitative example on glutathione synthesis to showcase how to implement cheat code 10. We used an equation estimating glutathione synthesis (Equation 2), which models the process as a second-order reaction and uses the intracellular concentrations of L-cysteine and L-glutamate, as well as, the rate constant of *de novo* synthesis (Huang et al., 2021). The rate constant and concentrations data were extracted from the literature (Huang et al., 2021; Raftos et al., 2010). We formed the model and performed all calculations using the R Statistical Software and RStudio. Using this model, we calculated glutathione synthesis under two different nutritional scenarios (i) NAC supplementation, and (ii) dietary L-cysteine deficiency. Under physiological conditions, erythrocyte glutathione synthesis was estimated at ≈ 700 μM per day, corroborating previous experimental findings (Lyons et al., 2000). Supplementation with NAC has been shown to lead to a 2-fold increase on average in L-cysteine concentration for about 4 h (Burgunder et al., 1989). Based on this, NAC supplementation was estimated to increase glutathione synthesis by $\approx 16\%$, leading to ≈ 816 μM per day. Dietary L-cysteine deficiency can decrease intracellular cysteine concentration by $\approx 55\%$ (Lyons et al., 2000). Lower L-cysteine availability was estimated to decrease glutathione synthesis by $\approx 40\%$, leading to ≈ 420 μM per day. The lower glutathione synthesis due to L-cysteine deficiency is supported by experimental data (Lyons et al., 2000). This simple and quick quantitative example demonstrates how simple calculations can provide biologically meaningful and experimentally falsifiable/testable estimations. Our example is available on GitHub (<https://github.com/PanosChatzi/Quantitative-redox-biology-calculating-glutathione-synthesis>).

Equation 2: Glutathione synthesis = $k_{\text{synthesis}} \times [\text{L-cysteine}] \times [\text{L-glutamate}]$

On the future of translational human redox research

The failure to measure oxidative stress obscured our understanding of whether nutritional antioxidants altered the onset of disease in humans (Gutteridge and Halliwell, 2010), which is now admittedly a fanciful concept can be likened to appealing to magic (Nikolaidis and Margaritelis, 2023). This pressing problem leached into the sports nutrition and exercise metabolism literature. Many studies investigating whether nutritional antioxidants blunt exercise adaptations failed to measure oxidative stress (Cobley et al., 2015b). Our cheat codes and decision selection tool can prevent these problems from reoccurring by enabling researchers to measure oxidative stress in humans. They bring a range of techniques to bear on the elucidation of the roles ROS play in humans, notably in the progression of diseases and their potential amelioration through diet and exercise. Focusing on diet and exercise themselves, instead of prophylactic antioxidants that often lack the

biochemical capacity to modulate the relevant redox reactions, defines a promising direction for advancing current knowledge. Halliwell counted diet and exercise amongst the best strategies to manipulate redox biology, such as ROS levels, in humans (Halliwell, 2023).

The future of oxidative stress research in sports nutrition and exercise metabolism is bright. The cheat codes can shed light on fundamental redox phenomena. For example, recent data link carbon dioxide/bicarbonate-derived species to the direct protein cysteine oxidation (Dagnell et al., 2019; Radi, 2022; Winterbourn et al., 2023). In effect, carbon dioxide/bicarbonate accelerate the process. The practice of using bicarbonate to modulate performance, may illuminate cysteine oxidation mechanisms in humans. Skeletal muscle studies are ideally placed to pioneer single cell oxidative stress research, especially when clear fibre-type differences (Cobley et al., 2016; Vasileiadou et al., 2023) make such analysis phenotypically meaningful (Rosenberger et al., 2023). They can also address lingering questions in redox signalling (Fuentes-Lemus and Davies, 2023; Glover et al., 2023; Henriquez-Olguin et al., 2023), particularly around the spatial regulation of the process with the potential for phase transition effects (Dai et al., 2023; Kritsiligkou et al., 2023). Finally, the field is set to play a leading role in unravelling the addressing individuality at the dawn of the personalised redox biology era (Margaritelis, 2023). For example, potential differences in exercise-induced oxidative stress manifests in males compared to remain largely unexplored.

Returning to the “at least for now” clause in cheat code 1, sophisticated tools, such as MitoNeoD for measuring mitochondrial superoxide levels (Shchepinova et al., 2017), offer possibilities for eventually measuring ROS levels in vivo using non-invasive technologies, such as positron emission tomography (PET) (Mailloux, 2021). Notable developments relate to PET-tracing of established ROS-reporters like ethidium in animal models (Hou et al., 2018). While there are many challenges to overcome and even then limitations will still apply (e.g., selectivity to one specific ROS (Zielonka and Kalyanaraman, 2018, 2010)), future innovations may enable the non-invasive measurement of ROS levels in humans. More broadly, we expect the cheat codes to evolve in line with technical advances. For example, and with reference to cheat code 5, a promising recent approach leveraged an enzymatic conjugation strategy to identify and quantify novel F₂-isoprostanes in human urine (Milne et al., 2024). Novel analytical approaches, such as the drop blot for single cell western analysis (Hughes et al., 2014; Liu and Herr, 2023), and artificial intelligence breakthroughs allowing for the production of designer proteins, notably antibodies (Hie et al., 2023; Madani et al., 2023), provide fertile ground for further advances. For example, a designer enzyme to selectively reduce sulfenic acids over other chemotypes.

The sheer biochemical complexity of oxidative stress presents multifaceted analytical challenges (Cobley, 2023a; Sies, 1986). Like a redox Laplace demon, there are no means to measure all of the relevant biochemistry in humans: to view every pixel of the picture in every dimension from ROS to oxidative damage. For example, even in comparatively simple cellular systems over 2,000 human proteins have yet to be measured at the peptide let alone proteoform level (Adhikari et al., 2020; Baker et al., 2017; Carbonara et al., 2021). The addressable yet still formidable challenge concerns capturing as much of the oxidative stress picture as possible using “omic” approaches, from state-of-the-art MS approaches to novel technologies, such as nanopore-based protein sequencing (Brinkerhoff et al., 2021; Martin-Baniandres et al., 2023; Yu et al., 2023). We call for a concerted community-wide effort to capture the multi-dimensional oxidative stress space in human samples to elucidate the biochemical nature of the phenomenon. We envisage the resources rationally guiding the validation of a large inventory of single or multiple biomarker panels comprising endogenous process-specific redox reporters, such as a biomarker of mitochondrial complex III-specific superoxide production (Bleier et al., 2015; Murphy, 2009). They would enable one to rationally measure representative pixels as “snapshot” biomarkers of process-specific oxidative stress biochemistry.

2. Conclusion

We tackled the long-standing problem of how to navigate the appreciable complexities of measuring oxidative stress in humans. Our solution defined a cheat code stratified suite of valid analytical approaches for measuring process-specific, such as lipid peroxidation, aspects of oxidative

stress in humans. The cheat codes are poised to advance translational redox research by supporting the measurement of oxidative stress in humans. Much like how they demystify and simplify seemingly impossible video gaming challenges, a set of redox “cheat codes” can hopefully demystify the intricacies of oxidative stress research, breaking down complex concepts into manageable steps for starting and guiding the voyage of discovery that is measuring oxidative stress in humans.

Acknowledgements: Figures 1-2 & 4-8 were created using BioRender (www.biorender.com) and exported with a publication licence.

Conflicts of Interest: The authors have no conflict of interest with regard to the present article and its contents.

References

- Adhikari, S., Nice, E.C., Deutsch, E.W., Lane, L., Omenn, G.S., Pennington, S.R., Paik, Y.-K., Overall, C.M., Corrales, F.J., Cristea, I.M., Eyk, J.E.V., Uhlén, M., Lindskog, C., Chan, D.W., Bairoch, A., Waddington, J.C., Justice, J.L., LaBaer, J., Rodriguez, H., He, F., Kostrzewa, M., Ping, P., Gundry, R.L., Stewart, P., Srivastava, Sanjeeva, Srivastava, Sudhir, Nogueira, F.C.S., Domont, G.B., Vandenbrouck, Y., Lam, M.P.Y., Wennersten, S., Vizcaino, J.A., Wilkins, M., Schwenk, J.M., Lundberg, E., Bandeira, N., Marko-Varga, G., Weintraub, S.T., Pineau, C., Kusebauch, U., Moritz, R.L., Ahn, S.B., Palmblad, M., Snyder, M.P., Aebersold, R., Baker, M.S., 2020. A high-stringency blueprint of the human proteome. *Nat Commun* 11, 5301. <https://doi.org/10.1038/s41467-020-19045-9>
- Aebersold, R., Agar, J.N., Amster, I.J., Baker, M.S., Bertozzi, C.R., Boja, E.S., Costello, C.E., Cravatt, B.F., Fenselau, C., Garcia, B.A., Ge, Y., Gunawardena, J., Hendrickson, R.C., Hergenrother, P.J., Huber, C.G., Ivanov, A.R., Jensen, O.N., Jewett, M.C., Kelleher, N.L., Kiessling, L.L., Krogan, N.J., Larsen, M.R., Loo, J.A., Loo, R.R.O., Lundberg, E., MacCoss, M.J., Mallick, P., Mootha, V.K., Mrksich, M., Muir, T.W., Patrie, S.M., Pesavento, J.J., Pitteri, S.J., Rodriguez, H., Saghatelian, A., Sandoval, W., Schlüter, H., Sechi, S., Slavoff, S.A., Smith, L.M., Snyder, M.P., Thomas, P.M., Uhlén, M., Eyk, J.E.V., Vidal, M., Walt, D.R., White, F.M., Williams, E.R., Wohlschlagler, T., Wysocki, V.H., Yates, N.A., Young, N.L., Zhang, B., 2018. How many human proteoforms are there? *Nat Chem Biol* 14, 206–214. <https://doi.org/10.1038/nchembio.2576>
- Aebersold, R., Mann, M., 2003. Mass spectrometry-based proteomics. *Nature* 422, 198–207. <https://doi.org/10.1038/nature01511>
- Akter, S., Fu, L., Jung, Y., Conte, M.L., Lawson, J.R., Lowther, W.T., Sun, R., Liu, K., Yang, J., Carroll, K.S., 2018. Chemical proteomics reveals new targets of cysteine sulfenic acid reductase. *Nat Chem Biol* 14, 995–1004. <https://doi.org/10.1038/s41589-018-0116-2>
- Alcock, L.J., Perkins, M.V., Chalker, J.M., 2017. Chemical methods for mapping cysteine oxidation. *Chem Soc Rev* 47, 231–268. <https://doi.org/10.1039/c7cs00607a>
- Aldini, G., Domingues, M.R., Spickett, C.M., Domingues, P., Altomare, A., Sánchez-Gómez, F.J., Oeste, C.L., Pérez-Sala, D., 2015. Protein lipoxidation: Detection strategies and challenges. *Redox Biol* 5, 253–266. <https://doi.org/10.1016/j.redox.2015.05.003>
- Alhmod, J.F., Woolley, J.F., Moustafa, A.-E.A., Malki, M.I., 2020. DNA Damage/Repair Management in Cancers. *Cancers* 12, 1050. <https://doi.org/10.3390/cancers12041050>
- Allison, W.S., 1976. Formation and reactions of sulfenic acids in proteins. *Acc. Chem. Res.* 9, 293–299. <https://doi.org/10.1021/ar50104a003>
- Anschau, V., Ferrer-Sueta, G., Aleixo-Silva, R.L., Fernandes, R.B., Tairum, C.A., Tonoli, C.C.C., Murakami, M.T., Oliveira, M.A. de, Netto, L.E.S., 2020. Reduction of sulfenic acids by ascorbate in proteins, connecting thiol-dependent to alternative redox pathways. *Free Radic. Biol. Med.* 156, 207–216. <https://doi.org/10.1016/j.freeradbiomed.2020.06.015>
- Antunes, F., Brito, P.M., 2017. Quantitative biology of hydrogen peroxide signaling. *Redox Biol* 13, 1–7. <https://doi.org/10.1016/j.redox.2017.04.039>
- Armas, M.I.D., Esteves, R., Viera, N., Reyes, A.M., Mastrogiovanni, M., Alegria, T.G.P., Netto, L.E.S., Tórtora, V., Radi, R., Trujillo, M., 2019. Rapid peroxynitrite reduction by human peroxiredoxin 3: Implications for the fate of oxidants in mitochondria. *Free Radical Bio Med* 130, 369–378. <https://doi.org/10.1016/j.freeradbiomed.2018.10.451>
- Arnér, E.S.J., 2017. Selenoproteins, Methods and Protocols. *Methods Mol Biology* 1661, 301–309. https://doi.org/10.1007/978-1-4939-7258-6_21
- Ast, T., Mootha, V.K., 2019. Oxygen and mammalian cell culture: are we repeating the experiment of Dr. Ox? *Nat Metabolism* 1, 858–860. <https://doi.org/10.1038/s42255-019-0105-0>
- Bailey, D.M., Culcasi, M., Filipponi, T., Brugniaux, J.V., Stacey, B.S., Marley, C.J., Soria, R., Rimoldi, S.F., Cerny, D., Rexhaj, E., Pratali, L., Salmòn, C.S., Jáuregui, C.M., Villena, M., Villafuerte, F., Rockenbauer, A., Pietri,

- S., Scherrer, U., Sartori, C., 2022. EPR spectroscopic evidence of iron-catalysed free radical formation in chronic mountain sickness: Dietary causes and vascular consequences. *Free Radic. Biol. Med.* 184, 99–113. <https://doi.org/10.1016/j.freeradbiomed.2022.03.028>
- Bailey, D.M., Davies, B., Young, I.S., Jackson, M.J., Davison, G.W., Isaacson, R., Richardson, R.S., 2003. EPR spectroscopic detection of free radical outflow from an isolated muscle bed in exercising humans. *J. Appl. Physiol.* 94, 1714–1718. <https://doi.org/10.1152/jappphysiol.01024.2002>
- Baker, M.S., Ahn, S.B., Mohamedali, A., Islam, M.T., Cantor, D., Verhaert, P.D., Fanayan, S., Sharma, S., Nice, E.C., Connor, M., Ranganathan, S., 2017. Accelerating the search for the missing proteins in the human proteome. *Nat Commun* 8, 14271. <https://doi.org/10.1038/ncomms14271>
- Barayeu, U., Schilling, D., Eid, M., Silva, T.N.X. da, Schlicker, L., Mitreska, N., Zapp, C., Gräter, F., Miller, A.K., Kappl, R., Schulze, A., Angeli, J.P.F., Dick, T.P., 2022. Hydropersulfides inhibit lipid peroxidation and ferroptosis by scavenging radicals. *Nat Chem Biol* 1–10. <https://doi.org/10.1038/s41589-022-01145-w>
- Bartosz, G., 2010. Non-enzymatic antioxidant capacity assays: Limitations of use in biomedicine. *Free Radic. Res.* 44, 711–720. <https://doi.org/10.3109/10715761003758114>
- Batthyány, C., Bartesaghi, S., Mastrogiovanni, M., Lima, A., Demicheli, V., Radi, R., 2017. Tyrosine-Nitrated Proteins: Proteomic and Bioanalytical Aspects. *Antioxid. Redox Signal.* 26, 313–328. <https://doi.org/10.1089/ars.2016.6787>
- Bayır, H., Anthonymuthu, T.S., Tyurina, Y.Y., Patel, S.J., Amoscato, A.A., Lamade, A.M., Yang, Q., Vladimirov, G.K., Philpott, C.C., Kagan, V.E., 2020. Achieving Life through Death: Redox Biology of Lipid Peroxidation in Ferroptosis. *Cell Chem. Biol.* 27, 387–408. <https://doi.org/10.1016/j.chembiol.2020.03.014>
- Beauchamp, C., Fridovich, I., 1971. Superoxide dismutase: Improved assays and an assay applicable to acrylamide gels. *Anal. Biochem.* 44, 276–287. [https://doi.org/10.1016/0003-2697\(71\)90370-8](https://doi.org/10.1016/0003-2697(71)90370-8)
- Bellissimo, C.A., Delfinis, L.J., Hughes, M.C., Turnbull, P.C., Gandhi, S., DiBenedetto, S.N., Rahman, F.A., Tadi, P., Amaral, C.A., Dehghani, A., Cobley, J., Quadriatero, J., Schlattner, U., Perry, C.G.R., 2023. Mitochondrial creatine sensitivity is lost in the D2.mdx model of Duchenne muscular dystrophy and rescued by the mitochondrial-enhancing compound Olesoxime. *Am J Physiol-cell Ph.* <https://doi.org/10.1152/ajpcell.00377.2022>
- Belousov, V.V., Fradkov, A.F., Lukyanov, K.A., Staroverov, D.B., Shakhbazov, K.S., Terskikh, A.V., Lukyanov, S., 2006. Genetically encoded fluorescent indicator for intracellular hydrogen peroxide. *Nat. Methods* 3, 281–286. <https://doi.org/10.1038/nmeth866>
- Bersani, N.A., Merwin, J.R., Lopez, N.I., Pearson, G.D., Merrill, G.F., 2002. Protein Electrophoretic Mobility Shift Assay to Monitor Redox State of Thioredoxin in Cells. *Methods Enzym.* 347, 317–326. [https://doi.org/10.1016/s0076-6879\(02\)47031-0](https://doi.org/10.1016/s0076-6879(02)47031-0)
- Beyer, W.F., Fridovich, I., 1987. Assaying for superoxide dismutase activity: Some large consequences of minor changes in conditions. *Anal. Biochem.* 161, 559–566. [https://doi.org/10.1016/0003-2697\(87\)90489-1](https://doi.org/10.1016/0003-2697(87)90489-1)
- Bielski, B.H.J., Cabelli, D.E., Arudi, R.L., Ross, A.B., 1985. Reactivity of HO₂/O₂ Radicals in Aqueous Solution. *J. Phys. Chem. Ref. Data* 14, 1041–1100. <https://doi.org/10.1063/1.555739>
- Bischoff, E., Lang, L., Zimmermann, J., Luczak, M., Kiefer, A.M., Niedner-Schatteburg, G., Manolikakes, G., Morgan, B., Deponte, M., 2023. Glutathione kinetically outcompetes reactions between dimedone and a cyclic sulfenamide or physiological sulfenic acids. *Free Radic. Biol. Med.* 208, 165–177. <https://doi.org/10.1016/j.freeradbiomed.2023.08.005>
- Biteau, B., Labarre, J., Toledano, M.B., 2003. ATP-dependent reduction of cysteine–sulphinic acid by *S. cerevisiae* sulphiredoxin. *Nature* 425, 980–984. <https://doi.org/10.1038/nature02075>
- Bleier, L., Wittig, I., Heide, H., Steger, M., Brandt, U., Dröse, S., 2015. Generator-specific targets of mitochondrial reactive oxygen species. *Free Radical Bio Med* 78, 1–10. <https://doi.org/10.1016/j.freeradbiomed.2014.10.511>
- Blum, J., Fridovich, I., 1985. Inactivation of glutathione peroxidase by superoxide radical. *Arch. Biochem. Biophys.* 240, 500–508. [https://doi.org/10.1016/0003-9861\(85\)90056-6](https://doi.org/10.1016/0003-9861(85)90056-6)
- Boivin, B., Zhang, S., Arbiser, J.L., Zhang, Z.-Y., Tonks, N.K., 2008. A modified cysteinyl-labeling assay reveals reversible oxidation of protein tyrosine phosphatases in angiomyolipoma cells. *Proc National Acad Sci* 105, 9959–9964. <https://doi.org/10.1073/pnas.0804336105>
- Bollineni, R.C., Hoffmann, R., Fedorova, M., 2014. Proteome-wide profiling of carbonylated proteins and carbonylation sites in HeLa cells under mild oxidative stress conditions. *Free Radical Bio Med* 68, 186–195. <https://doi.org/10.1016/j.freeradbiomed.2013.11.030>
- Bonini, M.G., Rota, C., Tomasi, A., Mason, R.P., 2006. The oxidation of 2',7'-dichlorofluorescein to reactive oxygen species: A self-fulfilling prophesy? *Free Radical Bio Med* 40, 968–975. <https://doi.org/10.1016/j.freeradbiomed.2005.10.042>
- Boveris, A., Chance, B., 1973. The mitochondrial generation of hydrogen peroxide. General properties and effect of hyperbaric oxygen. *Biochem. J.* 134, 707–716. <https://doi.org/10.1042/bj1340707>
- Brady, M.M., Meyer, A.S., 2022. Cataloguing the proteome: Current developments in single-molecule protein sequencing. *Biophys. Rev.* 3, 011304. <https://doi.org/10.1063/5.0065509>

- Bridge, H.N., Leiter, W., Frazier, C.L., Weeks, A.M., 2023. An N terminomics toolbox combining 2-pyridinecarboxaldehyde probes and click chemistry for profiling protease specificity. *Cell Chem. Biol.* <https://doi.org/10.1016/j.chembiol.2023.09.009>
- Brigelius-Flohé, R., Flohé, L., 2011. Basic Principles and Emerging Concepts in the Redox Control of Transcription Factors. *Antioxid Redox Sign* 15, 2335–2381. <https://doi.org/10.1089/ars.2010.3534>
- Brinkerhoff, H., Kang, A.S.W., Liu, J., Aksimentiev, A., Dekker, C., 2021. Multiple rereads of single proteins at single-amino acid resolution using nanopores. *Science* 374, 1509–1513. <https://doi.org/10.1126/science.abl4381>
- Brito, P.M., Antunes, F., 2014. Estimation of kinetic parameters related to biochemical interactions between hydrogen peroxide and signal transduction proteins. *Front. Chem.* 2, 82. <https://doi.org/10.3389/fchem.2014.00082>
- Bryk, R., Griffin, P., Nathan, C., 2000. Peroxynitrite reductase activity of bacterial peroxiredoxins. *Nature* 407, 211–215. <https://doi.org/10.1038/35025109>
- Buettner, G.R., 2011. Superoxide Dismutase in Redox Biology: The Roles of Superoxide and Hydrogen Peroxide. *Anti-cancer Agent Me* 11, 341–346. <https://doi.org/10.2174/187152011795677544>
- Burger, N., James, A.M., Mulvey, J.F., Hoogewijs, K., Ding, S., Fearnley, I.M., Loureiro-López, M., Norman, A.A.I., Arndt, S., Mottahedin, A., Sauchanka, O., Hartley, R.C., Krieg, T., Murphy, M.P., 2022. ND3 Cys39 in complex I is exposed during mitochondrial respiration. *Cell Chem Biol* 29, 636–649.e14. <https://doi.org/10.1016/j.chembiol.2021.10.010>
- Burgoyne, J.R., Madhani, M., Cuello, F., Charles, R.L., Brennan, J.P., Schroder, E., Browning, D.D., Eaton, P., 2007. Cysteine Redox Sensor in PKG α Enables Oxidant-Induced Activation. *Science* 317, 1393–1397. <https://doi.org/10.1126/science.1144318>
- Burgoyne, J.R., Oviusu, O., Eaton, P., 2013. The PEG-switch assay: A fast semi-quantitative method to determine protein reversible cysteine oxidation. *J Pharmacol Toxicol* 68, 297–301. <https://doi.org/10.1016/j.vascn.2013.07.001>
- Burgunder, J.M., Varriale, A., Lauterburg, B.H., 1989. Effect of N-acetylcysteine on plasma cysteine and glutathione following paracetamol administration. *Eur. J. Clin. Pharmacol.* 36, 127–131. <https://doi.org/10.1007/bf00609183>
- Buss, H., Chan, T.P., Sluis, K.B., Domigan, N.M., Winterbourn, C.C., 1997. Protein Carbonyl Measurement by a Sensitive ELISA Method. *Free Radic. Biol. Med.* 23, 361–366. [https://doi.org/10.1016/s0891-5849\(97\)00104-4](https://doi.org/10.1016/s0891-5849(97)00104-4)
- Carbonara, K., Andonovski, M., Coorsen, J.R., 2021. Proteomes Are of Proteoforms: Embracing the Complexity. *Proteomes* 9, 38. <https://doi.org/10.3390/proteomes9030038>
- Cardozo, G., Mastrogiovanni, M., Zeida, A., Viera, N., Radi, R., Reyes, A.M., Trujillo, M., 2023. Mitochondrial Peroxiredoxin 3 Is Rapidly Oxidized and Hyperoxidized by Fatty Acid Hydroperoxides. *Antioxidants* 12, 408. <https://doi.org/10.3390/antiox12020408>
- Carroll, L., Pattison, D.I., Davies, J.B., Anderson, R.F., Lopez-Alarcon, C., Davies, M.J., 2018. Superoxide radicals react with peptide-derived tryptophan radicals with very high rate constants to give hydroperoxides as major products. *Free Radic. Biol. Med.* 118, 126–136. <https://doi.org/10.1016/j.freeradbiomed.2018.02.033>
- Chance, B., Sies, H., Boveris, A., 1979. Hydroperoxide metabolism in mammalian organs. *Physiol Rev* 59, 527–605. <https://doi.org/10.1152/physrev.1979.59.3.527>
- Chandra, H., Symons, M.C.R., 1987. Sulphur radicals formed by cutting α -keratin. *Nature* 328, 833–834. <https://doi.org/10.1038/328833a0>
- Chatzinikolaou, P.N., Margaritelis, N.V., Paschalis, V., Theodorou, A.A., Vrabas, I.S., Kyparos, A., D'Alessandro, A., Nikolaidis, M.G., 2024. Erythrocyte metabolism. *Acta Physiol.* e14081. <https://doi.org/10.1111/apha.14081>
- Cheng, G., Zielonka, M., Dranka, B., Kumar, S.N., Myers, C.R., Bennett, B., Garces, A.M., Machado, L.G.D.D., Thiebaut, D., Ouari, O., Hardy, M., Zielonka, J., Kalyanaraman, B., 2018. Detection of mitochondria-generated reactive oxygen species in cells using multiple probes and methods: Potentials, pitfalls, and the future. *J. Biol. Chem.* 293, 10363–10380. <https://doi.org/10.1074/jbc.ra118.003044>
- Chouchani, E.T., Methner, C., Nadtochiy, S.M., Logan, A., Pell, V.R., Ding, S., James, A.M., Cochemé, H.M., Reinhold, J., Lilley, K.S., Partridge, L., Fearnley, I.M., Robinson, A.J., Hartley, R.C., Smith, R.A.J., Krieg, T., Brookes, P.S., Murphy, M.P., 2013. Cardioprotection by S-nitrosation of a cysteine switch on mitochondrial complex I. *Nat Med* 19, 753–759. <https://doi.org/10.1038/nm.3212>
- Cobley, J., Noble, A., Bessell, R., Guille, M., Husi, H., 2020. Reversible Thiol Oxidation Inhibits the Mitochondrial ATP Synthase in *Xenopus laevis* Oocytes. *Antioxidants* 9, 215. <https://doi.org/10.3390/antiox9030215>
- Cobley, James.N., Davison, G.W., 2022. *Oxidative Eustress in Exercise Physiology*. CRC Press.
- Cobley, J.N., 2023a. 50 shades of oxidative stress: A state-specific cysteine redox pattern hypothesis. *Redox Biol.* 67, 102936. <https://doi.org/10.1016/j.redox.2023.102936>
- Cobley, J.N., 2023b. Oxiforms: Unique cysteine residue- and chemotype-specified chemical combinations can produce functionally-distinct proteoforms. *Bioessays* 45. <https://doi.org/10.1002/bies.202200248>

- Cobley, James N., 2020. Mechanisms of Mitochondrial ROS Production in Assisted Reproduction: The Known, the Unknown, and the Intriguing. *Antioxidants* 9, 933. <https://doi.org/10.3390/antiox9100933>
- Cobley, James Nathan, 2020. Oxidative Stress 447–462. <https://doi.org/10.1016/b978-0-12-818606-0.00023-7>
- Cobley, J.N., 2018. Synapse Pruning: Mitochondrial ROS with Their Hands on the Shears. *Bioessays* 40, 1800031. <https://doi.org/10.1002/bies.201800031>
- Cobley, J.N., Bartlett, J.D., Kayani, A., Murray, S.W., Louhelainen, J., Donovan, T., Waldron, S., Gregson, W., Burniston, J.G., Morton, J.P., Close, G.L., 2012. PGC-1 α transcriptional response and mitochondrial adaptation to acute exercise is maintained in skeletal muscle of sedentary elderly males. *Biogerontology* 13, 621–631. <https://doi.org/10.1007/s10522-012-9408-1>
- Cobley, J.N., Close, G.L., Bailey, D.M., Davison, G.W., 2017. Exercise redox biochemistry: Conceptual, methodological and technical recommendations. *Redox Biol* 12, 540–548. <https://doi.org/10.1016/j.redox.2017.03.022>
- Cobley, J.N., Davison, G.W., 2022. Oxidative Eustress in Exercise Physiology 11–22. <https://doi.org/10.1201/9781003051619-2>
- Cobley, J.N., Fiorello, M.L., Bailey, D.M., 2018. 13 reasons why the brain is susceptible to oxidative stress. *Redox Biol* 15, 490–503. <https://doi.org/10.1016/j.redox.2018.01.008>
- Cobley, J.N., Husi, H., 2020. Immunological Techniques to Assess Protein Thiol Redox State: Opportunities, Challenges and Solutions. *Antioxidants* 9, 315. <https://doi.org/10.3390/antiox9040315>
- Cobley, J.N., Malik, Z.Ab., Morton, J.P., Close, G.L., Edwards, B.J., Burniston, J.G., 2016. Age- and Activity-Related Differences in the Abundance of Myosin Essential and Regulatory Light Chains in Human Muscle. *Proteomes* 4, 15. <https://doi.org/10.3390/proteomes4020015>
- Cobley, J.N., Margaritelis, N.V., Morton, J.P., Close, G.L., Nikolaidis, M.G., Malone, J.K., 2015a. The basic chemistry of exercise-induced DNA oxidation: oxidative damage, redox signaling, and their interplay. *Front Physiol* 6, 182. <https://doi.org/10.3389/fphys.2015.00182>
- Cobley, J.N., Marrin, K., 2012. Vitamin E supplementation does not alter physiological performance at fixed blood lactate concentrations in trained runners. *J. sports Med. Phys. Fit.* 52, 63–70.
- Cobley, J.N., McGlory, C., Morton, J.P., Close, G.L., 2011. N-Acetylcysteine's Attenuation of Fatigue After Repeated Bouts of Intermittent Exercise: Practical Implications for Tournament Situations. *Int J Sport Nutr Exe* 21, 451–461. <https://doi.org/10.1123/ijsnem.21.6.451>
- Cobley, J.N., McHardy, H., Morton, J.P., Nikolaidis, M.G., Close, G.L., 2015b. Influence of vitamin C and vitamin E on redox signaling: Implications for exercise adaptations. *Free Radical Bio Med* 84, 65–76. <https://doi.org/10.1016/j.freeradbiomed.2015.03.018>
- Cobley, J.N., Moulton, P.R., Burniston, J.G., Morton, J.P., Close, G.L., 2015c. Exercise improves mitochondrial and redox-regulated stress responses in the elderly: better late than never! *Biogerontology* 16, 249–264. <https://doi.org/10.1007/s10522-014-9546-8>
- Cobley, J.N., Noble, A., Jimenez-Fernandez, E., Moya, M.-T.V., Guille, M., Husi, H., 2019a. Catalyst-free Click PEGylation reveals substantial mitochondrial ATP synthase sub-unit alpha oxidation before and after fertilisation. *Redox Biol* 26, 101258. <https://doi.org/10.1016/j.redox.2019.101258>
- Cobley, J.N., Sakellariou, G.K., Husi, H., McDonagh, B., 2019b. Proteomic strategies to unravel age-related redox signalling defects in skeletal muscle. *Free Radical Bio Med* 132, 24–32. <https://doi.org/10.1016/j.freeradbiomed.2018.09.012>
- Cobley, J.N., Sakellariou, G.K., Murray, S., Waldron, S., Gregson, W., Burniston, J.G., Morton, J.P., Iwanejko, L.A., Close, G.L., 2013. Lifelong endurance training attenuates age-related genotoxic stress in human skeletal muscle. *Longev Heal* 2, 11. <https://doi.org/10.1186/2046-2395-2-11>
- Cobley, J.N., Sakellariou, G.K., Owens, D.J., Murray, S., Waldron, S., Gregson, W., Fraser, W.D., Burniston, J.G., Iwanejko, L.A., McArdle, A., Morton, J.P., Jackson, M.J., Close, G.L., 2014. Lifelong training preserves some redox-regulated adaptive responses after an acute exercise stimulus in aged human skeletal muscle. *Free Radical Bio Med* 70, 23–32. <https://doi.org/10.1016/j.freeradbiomed.2014.02.004>
- Collins, A.R., Azqueta, A., 2012. DNA repair as a biomarker in human biomonitoring studies; further applications of the comet assay. *Mutat. Res.Fundam. Mol. Mech. Mutagen.* 736, 122–129. <https://doi.org/10.1016/j.mrfmmm.2011.03.005>
- Consortium, I.H.G.S., Research; W.I. for B.R., Center for Genome, Lander, E.S., Linton, L.M., Birren, B., Nusbaum, C., Zody, M.C., Baldwin, J., Devon, K., Dewar, K., Doyle, M., FitzHugh, W., Funke, R., Gage, D., Harris, K., Heaford, A., Howland, J., Kann, L., Lehoczky, J., LeVine, R., McEwan, P., McKernan, K., Meldrim, J., Mesirov, J.P., Miranda, C., Morris, W., Naylor, J., Raymond, Christina, Rosetti, M., Santos, R., Sheridan, A., Sougnez, C., Stange-Thomann, N., Stojanovic, N., Subramanian, A., Wyman, D., Centre; T.S., Rogers, J., Sulston, J., Ainscough, R., Beck, S., Bentley, D., Burton, J., Clee, C., Carter, N., Coulson, A., Deadman, R., Deloukas, P., Dunham, A., Dunham, I., Durbin, R., French, L., Grafham, D., Gregory, S., Hubbard, T., Humphray, S., Hunt, A., Jones, M., Lloyd, C., McMurray, A., Matthews, L., Mercer, S., Milne, S., Mullikin, J.C., Mungall, A., Plumb, R., Ross, M., Shownkeen, R., Sims, S., Center, W.U.G.S., Waterston, R.H., Wilson, R.K., Hillier, L.W., McPherson, J.D., Marra, M.A., Mardis, E.R., Fulton, L.A., Chinwalla, A.T.,

- Pepin, K.H., Gish, W.R., Chissoe, S.L., Wendl, M.C., Delehaunty, K.D., Miner, T.L., Delehaunty, A., Kramer, J.B., Cook, L.L., Fulton, R.S., Johnson, D.L., Minx, P.J., Clifton, S.W., Institute; U.D.J.G., Hawkins, T., Branscomb, E., Predki, P., Richardson, P., Wenning, S., Slezak, T., Doggett, N., Cheng, J.-F., Olsen, A., Lucas, S., Elkin, C., Uberbacher, E., Frazier, M., Center; B.C. of M.H.G.S., Gibbs, R.A., Muzny, D.M., Scherer, S.E., Bouck, J.B., Sodergren, E.J., Worley, K.C., Rives, C.M., Gorrell, J.H., Metzker, M.L., Naylor, S.L., Kucherlapati, R.S., Nelson, D.L., Weinstock, G.M., Center; R.G.S., Sakaki, Y., Fujiyama, A., Hattori, M., Yada, T., Toyoda, A., Itoh, T., Kawagoe, C., Watanabe, H., Totoki, Y., Taylor, T., UMR-8030; G. and C., Weissenbach, J., Heilig, R., Saurin, W., Artiguenave, F., Brottier, P., Bruls, T., Pelletier, E., Robert, C., Wincker, P., Biotechnology; D. of G.A., Institute of Molecular, Rosenthal, A., Platzer, M., Nyakatura, G., Taudien, S., Rump, A., Center; G.S., Smith, D.R., Doucette-Stamm, L., Rubenfield, M., Weinstock, K., Lee, H.M., Dubois, J., Center; B.G.I.G., Yang, H., Yu, J., Wang, J., Huang, G., Gu, J., Biology; M.S.C., The Institute for Systems, Hood, L., Rowen, L., Madan, A., Qin, S., Center; S.G.T., Davis, R.W., Federspiel, N.A., Abola, A.P., Proctor, M.J., Technology; U. of O.A.C. for G., Roe, B.A., Chen, F., Pan, H., Genetics; M.P.I. for M., Ramser, J., Lehrach, H., Reinhardt, R., Center; C.S.H.L., Lita Annenberg Hazen Genome, McCombie, W.R., Bastide, M. de la, Dedhia, N., Biotechnology; G.R.C. for, Blöcker, H., Hornischer, K., Nordtsiek, G., Agarwala, R., Aravind, L., Bailey, J.A., Bateman, A., Batzoglu, S., Birney, E., Bork, P., Brown, D.G., Burge, C.B., Cerutti, L., Chen, H.-C., Church, D., Clamp, M., Copley, R.R., Doerks, T., Eddy, S.R., Eichler, E.E., Furey, T.S., Galagan, J., Gilbert, J.G.R., Harmon, C., Hayashizaki, Y., Haussler, D., Hermjakob, H., Hokamp, K., Jang, W., Johnson, L.S., Jones, T.A., Kasif, S., Kasprzyk, A., Kennedy, S., Kent, W.J., Kitts, P., Koonin, E.V., Korf, I., Kulp, D., Lancet, D., Lowe, T.M., McLysaght, A., Mikkelsen, T., Moran, J.V., Mulder, N., Pollara, V.J., Ponting, C.P., Schuler, G., Schultz, J., Slater, G., Smit, A.F.A., Stupka, E., Szustakowski, J., Thierry-Mieg, D., Thierry-Mieg, J., Wagner, L., Wallis, J., Wheeler, R., Williams, A., Wolf, Y.I., Wolfe, K.H., Yang, S.-P., Yeh, R.-F., Health; S. management: N.H.G.R.I., US National Institutes of, Collins, F., Guyer, M.S., Peterson, J., Felsenfeld, A., Wetterstrand, K.A., Center; S.H.G., Myers, R.M., Schmutz, J., Dickson, M., Grimwood, J., Cox, D.R., Center; U. of W.G., Olson, M.V., Kaul, R., Raymond, Christopher, Medicine; D. of M.B., Keio University School of, Shimizu, N., Kawasaki, K., Minoshima, S., Dallas; U. of T.S.M.C. at, Evans, G.A., Athanasiou, M., Schultz, R., Energy; O. of S., US Department of, Patrinos, A., Trust; T.W., Morgan, M.J., 2001. Initial sequencing and analysis of the human genome. *Nature* 409, 860–921. <https://doi.org/10.1038/35057062>
- Crapo, J.D., McCord, J.M., Fridovich, I., 1978. Preparation and assay of superoxide dismutases. *Methods Enzymol.* 53, 382–393. [https://doi.org/10.1016/s0076-6879\(78\)53044-9](https://doi.org/10.1016/s0076-6879(78)53044-9)
- Dagnell, M., Cheng, Q., Rizvi, S.H.M., Pace, P.E., Boivin, B., Winterbourn, C.C., Arnér, E.S.J., 2019. Bicarbonate is essential for protein-tyrosine phosphatase 1B (PTP1B) oxidation and cellular signaling through EGF-triggered phosphorylation cascades. *J. Biol. Chem.* 294, 12330–12338. <https://doi.org/10.1074/jbc.ra119.009001>
- Dagnell, M., Pace, P.E., Cheng, Q., Friehoff, J., Östman, A., Arnér, E.S.J., Hampton, M.B., Winterbourn, C.C., 2017. Thioredoxin reductase 1 and NADPH directly protect protein tyrosine phosphatase 1B from inactivation during H₂O₂ exposure. *J Biol Chem* 292, 14371–14380. <https://doi.org/10.1074/jbc.m117.793745>
- Dai, Y., Chamberlayne, C.F., Messina, M.S., Chang, C.J., Zare, R.N., You, L., Chilkoti, A., 2023. Interface of biomolecular condensates modulates redox reactions. *Chem.* <https://doi.org/10.1016/j.chempr.2023.04.001>
- D'Autréaux, B., Toledano, M.B., 2007. ROS as signalling molecules: mechanisms that generate specificity in ROS homeostasis. *Nat Rev Mol Cell Bio* 8, 813–824. <https://doi.org/10.1038/nrm2256>
- Davies, K.J.A., Quintanilha, A.T., Brooks, G.A., Packer, L., 1982. Free radicals and tissue damage produced by exercise. *Biochem. Biophys. Res. Commun.* 107, 1198–1205. [https://doi.org/10.1016/s0006-291x\(82\)80124-1](https://doi.org/10.1016/s0006-291x(82)80124-1)
- Davies, M.J., 2016. Protein oxidation and peroxidation. *Biochem J* 473, 805–825. <https://doi.org/10.1042/bj20151227>
- Davison, G.W., 2016. Exercise and Oxidative Damage in Nucleoid DNA Quantified Using Single Cell Gel Electrophoresis: Present and Future Application. *Front. Physiol.* 7, 249. <https://doi.org/10.3389/fphys.2016.00249>
- Day, N.J., Gaffrey, M.J., Qian, W.-J., 2021. Stoichiometric Thiol Redox Proteomics for Quantifying Cellular Responses to Perturbations. *Antioxidants* 10, 499. <https://doi.org/10.3390/antiox10030499>
- Derks, J., Leduc, A., Wallmann, G., Huffman, R.G., Willetts, M., Khan, S., Specht, H., Ralser, M., Demichev, V., Slavov, N., 2022. Increasing the throughput of sensitive proteomics by plexDIA. *Nat Biotechnol* 1–10. <https://doi.org/10.1038/s41587-022-01389-w>
- Dickinson, B.C., Chang, C.J., 2011. Chemistry and biology of reactive oxygen species in signaling or stress responses. *Nat Chem Biol* 7, 504–511. <https://doi.org/10.1038/nchembio.607>
- Dillard, C.J., Litov, R.E., Savin, W.M., Dumelin, E.E., Tappel, A.L., 1978. Effects of exercise, vitamin E, and ozone on pulmonary function and lipid peroxidation. *J. Appl. Physiol.* 45, 927–932. <https://doi.org/10.1152/jappl.1978.45.6.927>
- Dinkova-Kostova, A.T., Holtzclaw, W.D., Cole, R.N., Itoh, K., Wakabayashi, N., Katoh, Y., Yamamoto, M., Talalay, P., 2002. Direct evidence that sulfhydryl groups of Keap1 are the sensors regulating induction of

- phase 2 enzymes that protect against carcinogens and oxidants. *Proc. Natl. Acad. Sci.* 99, 11908–11913. <https://doi.org/10.1073/pnas.172398899>
- Dixon, S.J., Lemberg, K.M., Lamprecht, M.R., Skouta, R., Zaitsev, E.M., Gleason, C.E., Patel, D.N., Bauer, A.J., Cantley, A.M., Yang, W.S., Morrison, B., Stockwell, B.R., 2012. Ferroptosis: An Iron-Dependent Form of Nonapoptotic Cell Death. *Cell* 149, 1060–1072. <https://doi.org/10.1016/j.cell.2012.03.042>
- Dizdaroglu, M., Jaruga, P., Rodriguez, H., 2001. Identification and quantification of 8,5'-cyclo-2'-deoxyadenosine in DNA by liquid chromatography/ mass spectrometry. *Free Radic. Biol. Med.* 30, 774–784. [https://doi.org/10.1016/s0891-5849\(01\)00464-6](https://doi.org/10.1016/s0891-5849(01)00464-6)
- Echtay, K.S., Roussel, D., St-Pierre, J., Jekabsons, M.B., Cadenas, S., Stuart, J.A., Harper, J.A., Roebuck, S.J., Morrison, A., Pickering, S., Clapham, J.C., Brand, M.D., 2002. Superoxide activates mitochondrial uncoupling proteins. *Nature* 415, 96–99. <https://doi.org/10.1038/415096a>
- Egan, B., Sharples, A.P., 2023. Molecular responses to acute exercise and their relevance for adaptations in skeletal muscle to exercise training. *Physiol. Rev.* 103, 2057–2170. <https://doi.org/10.1152/physrev.00054.2021>
- Egan, B., Zierath, J.R., 2013. Exercise Metabolism and the Molecular Regulation of Skeletal Muscle Adaptation. *Cell Metab.* 17, 162–184. <https://doi.org/10.1016/j.cmet.2012.12.012>
- Eid, M., Barayeu, U., Sulková, K., Aranda-Vallejo, C., Dick, T.P., 2024. Using the heme peroxidase APEX2 to probe intracellular H₂O₂ flux and diffusion. *Nat. Commun.* 15, 1239. <https://doi.org/10.1038/s41467-024-45511-9>
- Ellman, G.L., 1959. Tissue sulfhydryl groups. *Arch Biochem Biophys* 82, 70–77. [https://doi.org/10.1016/0003-9861\(59\)90090-6](https://doi.org/10.1016/0003-9861(59)90090-6)
- Epstein, J., 2008. Why model. *Journal of Artificial Societies and Social Simulation*.
- Erdogan, Y.C., Altun, H.Y., Secilmis, M., Ata, B.N., Sevimli, G., Cokluk, Z., Zaki, A.G., Sezen, S., Caglar, T.A., Sevgen, İ., Steinhorn, B., Ai, H., Öztürk, G., Belousov, V.V., Michel, T., Eroglu, E., 2021. Complexities of the chemogenetic toolkit: Differential mDAAO activation by d-amino substrates and subcellular targeting. *Free Radical Bio Med* 177, 132–142. <https://doi.org/10.1016/j.freeradbiomed.2021.10.023>
- Erve, T.J. van 't, Lih, F.B., Kadiiska, M.B., Deterding, L.J., Eling, T.E., Mason, R.P., 2015. Reinterpreting the best biomarker of oxidative stress: The 8-iso-PGF₂α/PGF₂α ratio distinguishes chemical from enzymatic lipid peroxidation. *Free Radical Bio Med* 83, 245–251. <https://doi.org/10.1016/j.freeradbiomed.2015.03.004>
- Erve, T.J. van 't, Kadiiska, M.B., London, S.J., Mason, R.P., 2017. Classifying oxidative stress by F₂-isoprostane levels across human diseases: A meta-analysis. *Redox Biol* 12, 582–599. <https://doi.org/10.1016/j.redox.2017.03.024>
- Ezeriņa, D., Takano, Y., Hanaoka, K., Urano, Y., Dick, T.P., 2018. N-Acetyl Cysteine Functions as a Fast-Acting Antioxidant by Triggering Intracellular H₂S and Sulfane Sulfur Production. *Cell Chem. Biol.* 25, 447–459.e4. <https://doi.org/10.1016/j.chembiol.2018.01.011>
- Fedorova, M., Bollineni, R.C., Hoffmann, R., 2014. Protein carbonylation as a major hallmark of oxidative damage: Update of analytical strategies. *Mass Spectrom Rev* 33, 79–97. <https://doi.org/10.1002/mas.21381>
- Ferreira, R.B., Fu, L., Jung, Y., Yang, J., Carroll, K.S., 2022. Reaction-based fluorogenic probes for detecting protein cysteine oxidation in living cells. *Nat Commun* 13, 5522. <https://doi.org/10.1038/s41467-022-33124-z>
- Figueroa-González, G., Pérez-Plasencia, C., 2017. Strategies for the evaluation of DNA damage and repair mechanisms in cancer. *Oncol. Lett.* 13, 3982–3988. <https://doi.org/10.3892/ol.2017.6002>
- Flohé, L., 2020. Looking Back at the Early Stages of Redox Biology. *Antioxidants* 9, 1254. <https://doi.org/10.3390/antiox9121254>
- Flohé, L., 2013. The fairytale of the GSSG/GSH redox potential. *Biochim. Biophys. Acta (BBA) - Gen. Subj.* 1830, 3139–3142. <https://doi.org/10.1016/j.bbagen.2012.10.020>
- Floyd, R.A., Watson, J.J., Harris, J., West, M., Wong, P.K., 1986. Formation of 8-hydroxydeoxyguanosine, hydroxyl free radical adduct of DNA in granulocytes exposed to the tumor promoter, tetradecanoylphorbolacetate. *Biochem. Biophys. Res. Commun.* 137, 841–846. [https://doi.org/10.1016/0006-291x\(86\)91156-3](https://doi.org/10.1016/0006-291x(86)91156-3)
- Forman, H.J., Augusto, O., Brigelius-Flohe, R., Dennery, P.A., Kalyanaraman, B., Ischiropoulos, H., Mann, G.E., Radi, R., Roberts, L.J., Vina, J., Davies, K.J.A., 2015. Even free radicals should follow some rules: A Guide to free radical research terminology and methodology. *Free Radical Bio Med* 78, 233–235. <https://doi.org/10.1016/j.freeradbiomed.2014.10.504>
- Forman, H.J., Davies, K.J.A., Ursini, F., 2014. How do nutritional antioxidants really work: Nucleophilic tone and para-hormesis versus free radical scavenging in vivo. *Free Radical Bio Med* 66, 24–35. <https://doi.org/10.1016/j.freeradbiomed.2013.05.045>
- Forman, H.J., Davies, M.J., Krämer, A.C., Miotto, G., Zaccarin, M., Zhang, H., Ursini, F., 2017. Protein cysteine oxidation in redox signaling: Caveats on sulfenic acid detection and quantification. *Arch Biochem Biophys* 617, 26–37. <https://doi.org/10.1016/j.abb.2016.09.013>

- Forman, H.J., Zhang, H., 2021. Targeting oxidative stress in disease: promise and limitations of antioxidant therapy. *Nat Rev Drug Discov* 1–21. <https://doi.org/10.1038/s41573-021-00233-1>
- Fridovich, I., 1998. Oxygen Toxicity: A Radical Explanation. *J. Exp. Biol.* 201, 1203–1209. <https://doi.org/10.1242/jeb.201.8.1203>
- Fridovich, I., 1989. Superoxide dismutases An adaptation to a paramagnetic gas. *J. Biol. Chem.* 264, 7761–7764. [https://doi.org/10.1016/s0021-9258\(18\)83102-7](https://doi.org/10.1016/s0021-9258(18)83102-7)
- Fridovich, I., 1978. The Biology of Oxygen Radicals. *Science* 201, 875–880. <https://doi.org/10.1126/science.210504>
- Frijhoff, J., Winyard, P.G., Zarkovic, N., Davies, S.S., Stocker, R., Cheng, D., Knight, A.R., Taylor, E.L., Oettrich, J., Ruskovska, T., Gasparovic, A.C., Cuadrado, A., Weber, D., Poulsen, H.E., Grune, T., Schmidt, H.H.H.W., Ghezzi, P., 2015. Clinical Relevance of Biomarkers of Oxidative Stress. *Antioxid. Redox Signal.* 23, 1144–1170. <https://doi.org/10.1089/ars.2015.6317>
- Fuentes-Lemus, E., Davies, M.J., 2023. Effect of crowding, compartmentalization and nanodomains on protein modification and redox signaling – current state and future challenges. *Free Radical Bio Med* 196, 81–92. <https://doi.org/10.1016/j.freeradbiomed.2023.01.011>
- Ganini, D., Santos, J.H., Bonini, M.G., Mason, R.P., 2018. Switch of Mitochondrial Superoxide Dismutase into a Prooxidant Peroxidase in Manganese-Deficient Cells and Mice. *Cell Chem Biol* 25, 413–425.e6. <https://doi.org/10.1016/j.chembiol.2018.01.007>
- Gardner, P.R., 2002. Aconitase: Sensitive target and measure of superoxide. *Methods Enzym.* 349, 9–23. [https://doi.org/10.1016/s0076-6879\(02\)49317-2](https://doi.org/10.1016/s0076-6879(02)49317-2)
- Gardner, P.R., Nguyen, D.D., White, C.W., 1994. Aconitase is a sensitive and critical target of oxygen poisoning in cultured mammalian cells and in rat lungs. *Proc. Natl. Acad. Sci.* 91, 12248–12252. <https://doi.org/10.1073/pnas.91.25.12248>
- Gatin-Fraudet, B., Ottenwelter, R., Saux, T.L., Norsikian, S., Pucher, M., Lombès, T., Baron, A., Durand, P., Doisneau, G., Bourdreux, Y., Iorga, B.I., Erard, M., Jullien, L., Guianvarc’h, D., Urban, D., Vauzeilles, B., 2021. Evaluation of borinic acids as new, fast hydrogen peroxide-responsive triggers. *Proc. Natl. Acad. Sci.* 118, e2107503118. <https://doi.org/10.1073/pnas.2107503118>
- Georgiou, C.D., Zisimopoulos, D., Argyropoulou, V., Kalaitzopoulou, E., Ioannou, P.V., Salachas, G., Grune, T., 2018. Protein carbonyl determination by a rhodamine B hydrazide-based fluorometric assay. *Redox Biol* 17, 236–245. <https://doi.org/10.1016/j.redox.2018.04.017>
- Giustarini, D., Dalle-Donne, I., Colombo, R., Milzani, A., Rossi, R., 2003. An improved HPLC measurement for GSH and GSSG in human blood. *Free Radic. Biol. Med.* 35, 1365–1372. <https://doi.org/10.1016/j.freeradbiomed.2003.08.013>
- Giustarini, D., Milzani, A., Dalle-Donne, I., Rossi, R., 2023. How to Increase Cellular Glutathione. *Antioxidants* 12, 1094. <https://doi.org/10.3390/antiox12051094>
- Glover, M.R., Davies, M.J., Fuentes-Lemus, E., 2023. Oxidation of the active site cysteine residue of glyceraldehyde-3-phosphate dehydrogenase to the hyper-oxidized sulfonic acid form is favored under crowded conditions. *Free Radic. Biol. Med.* <https://doi.org/10.1016/j.freeradbiomed.2023.12.015>
- Go, Y.-M., Chandler, J.D., Jones, D.P., 2015. The cysteine proteome. *Free Radical Bio Med* 84, 227–245. <https://doi.org/10.1016/j.freeradbiomed.2015.03.022>
- Gomez-Cabrera, M., Borrás, C., Pallardó, F.V., Sastre, J., Ji, L.L., Viña, J., 2005. Decreasing xanthine oxidase-mediated oxidative stress prevents useful cellular adaptations to exercise in rats. *J. Physiol.* 567, 113–120. <https://doi.org/10.1113/jphysiol.2004.080564>
- Gomez-Cabrera, M.C., Carretero, A., Millan-Domingo, F., Garcia-Dominguez, E., Correas, A.G., Olasso-Gonzalez, G., Viña, J., 2021. Redox-related biomarkers in physical exercise. *Redox Biol* 42, 101956. <https://doi.org/10.1016/j.redox.2021.101956>
- Gomez-Cabrera, M.-C., Domenech, E., Romagnoli, M., Arduini, A., Borrás, C., Pallardo, F.V., Sastre, J., Viña, J., 2008. Oral administration of vitamin C decreases muscle mitochondrial biogenesis and hampers training-induced adaptations in endurance performance. *Am J Clin Nutrition* 87, 142–149. <https://doi.org/10.1093/ajcn/87.1.142>
- Gómez-Cabrera, M.-C., Pallardó, F.V., Sastre, J., Viña, J., García-del-Moral, L., 2003. Allopurinol and Markers of Muscle Damage Among Participants in the Tour de France. *JAMA* 289, 2503–2504. <https://doi.org/10.1001/jama.289.19.2503-b>
- Goncalves, R.L.S., Quinlan, C.L., Pervoshchikova, I.V., Hey-Mogensen, M., Brand, M.D., 2015. Sites of Superoxide and Hydrogen Peroxide Production by Muscle Mitochondria Assessed ex Vivo under Conditions Mimicking Rest and Exercise*. *J. Biol. Chem.* 290, 209–227. <https://doi.org/10.1074/jbc.m114.619072>
- Gutteridge, J.M.C., Halliwell, B., 2018. Mini-Review: Oxidative stress, redox stress or redox success? *Biochem Bioph Res Co* 502, 183–186. <https://doi.org/10.1016/j.bbrc.2018.05.045>
- Gutteridge, J.M.C., Halliwell, B., 2010. Antioxidants: Molecules, medicines, and myths. *Biochem Bioph Res Co* 393, 561–564. <https://doi.org/10.1016/j.bbrc.2010.02.071>

- GUTTERIDGE, J.M.C., HALLIWELL, B., 2000. Free Radicals and Antioxidants in the Year 2000: A Historical Look to the Future. *Ann Ny Acad Sci* 899, 136–147. <https://doi.org/10.1111/j.1749-6632.2000.tb06182.x>
- Halliwell, B., 2023. Understanding mechanisms of antioxidant action in health and disease. *Nat. Rev. Mol. Cell Biol.* 1–21. <https://doi.org/10.1038/s41580-023-00645-4>
- Halliwell, B., 2009. The wanderings of a free radical. *Free Radical Bio Med* 46, 531–542. <https://doi.org/10.1016/j.freeradbiomed.2008.11.008>
- Halliwell, B., 2008. Are polyphenols antioxidants or pro-oxidants? What do we learn from cell culture and in vivo studies? *Arch. Biochem. Biophys.* 476, 107–112. <https://doi.org/10.1016/j.abb.2008.01.028>
- Halliwell, B., 2007. Biochemistry of oxidative stress. *Biochemical Society Transactions.*
- Halliwell, B., 2006. Oxidative stress and cancer: have we moved forward? *Biochem J* 401, 1–11. <https://doi.org/10.1042/bj20061131>
- Halliwell, B., 1987. Oxidants and human disease: some new concepts1. *FASEB J.* 1, 358–364. <https://doi.org/10.1096/fasebj.1.5.2824268>
- Halliwell, B., Adhikary, A., Dingfelder, M., Dizdaroglu, M., 2021. Hydroxyl radical is a significant player in oxidative DNA damage in vivo. *Chem. Soc. Rev.* 50, 8355–8360. <https://doi.org/10.1039/d1cs00044f>
- Halliwell, B., Gutteridge, J., 2015. Free Radicals in Biology and Medicine.
- Halliwell, B., Lee, C.Y.J., 2010. Using Isoprostanes as Biomarkers of Oxidative Stress: Some Rarely Considered Issues. *Antioxid Redox Sign* 13, 145–156. <https://doi.org/10.1089/ars.2009.2934>
- Halliwell, B., Whiteman, M., 2004. Measuring reactive species and oxidative damage in vivo and in cell culture: how should you do it and what do the results mean? *Brit J Pharmacol* 142, 231–255. <https://doi.org/10.1038/sj.bjp.0705776>
- Halliwell, B., Zhao, K., Whiteman, M., 2000. The gastrointestinal tract: A major site of antioxidant action? *Free Radic. Res.* 33, 819–830. <https://doi.org/10.1080/10715760000301341>
- Hansen, R.E., Winther, J.R., 2009. An introduction to methods for analyzing thiols and disulfides: Reactions, reagents, and practical considerations. *Anal Biochem* 394, 147–158. <https://doi.org/10.1016/j.ab.2009.07.051>
- Hausladen, A., Fridovich, I., 1994. Superoxide and peroxyxynitrite inactivate aconitases, but nitric oxide does not. *J. Biol. Chem.* 269, 29405–29408. [https://doi.org/10.1016/s0021-9258\(18\)43893-8](https://doi.org/10.1016/s0021-9258(18)43893-8)
- Hawkins, C.L., Davies, M.J., 2019. Detection, identification, and quantification of oxidative protein modifications. *J. Biol. Chem.* 294, 19683–19708. <https://doi.org/10.1074/jbc.rev119.006217>
- Hawkins, C.L., Morgan, P.E., Davies, M.J., 2009. Quantification of protein modification by oxidants. *Free Radic. Biol. Med.* 46, 965–988. <https://doi.org/10.1016/j.freeradbiomed.2009.01.007>
- Hayes, J.D., Dinkova-Kostova, A.T., Tew, K.D., 2020. Oxidative Stress in Cancer. *Cancer Cell* 38, 167–197. <https://doi.org/10.1016/j.ccell.2020.06.001>
- Held, J.M., 2020. Redox Systems Biology: Harnessing the Sentinels of the Cysteine Redoxome. *Antioxid Redox Sign* 32, 659–676. <https://doi.org/10.1089/ars.2019.7725>
- Held, J.M., Danielson, S.R., Behring, J.B., Atsriku, C., Britton, D.J., Puckett, R.L., Schilling, B., Campisi, J., Benz, C.C., Gibson, B.W., 2010. Targeted Quantitation of Site-Specific Cysteine Oxidation in Endogenous Proteins Using a Differential Alkylation and Multiple Reaction Monitoring Mass Spectrometry Approach. *Mol Cell Proteomics* 9, 1400–1410. <https://doi.org/10.1074/mcp.m900643-mcp200>
- Henriquez-Olguin, C., Meneses-Valdes, R., Jensen, T.E., 2020. Compartmentalized muscle redox signals controlling exercise metabolism – Current state, future challenges. *Redox Biol* 35, 101473. <https://doi.org/10.1016/j.redox.2020.101473>
- Henriquez-Olguin, C., Meneses-Valdes, R., Kritsiligkou, P., Fuentes-Lemus, E., 2023. From workout to molecular switches: How does skeletal muscle produce, sense, and transduce subcellular redox signals? *Free Radic. Biol. Med.* 209, 355–365. <https://doi.org/10.1016/j.freeradbiomed.2023.10.404>
- Hie, B.L., Shanker, V.R., Xu, D., Bruun, T.U.J., Weidenbacher, P.A., Tang, S., Wu, W., Pak, J.E., Kim, P.S., 2023. Efficient evolution of human antibodies from general protein language models. *Nat Biotechnol* 1–9. <https://doi.org/10.1038/s41587-023-01763-2>
- Holmström, K.M., Finkel, T., 2014. Cellular mechanisms and physiological consequences of redox-dependent signalling. *Nat Rev Mol Cell Bio* 15, 411–421. <https://doi.org/10.1038/nrm3801>
- Hou, C., Hsieh, C.-J., Li, S., Lee, H., Graham, T.J., Xu, K., Weng, C.-C., Doot, R.K., Chu, W., Chakraborty, S.K., Dugan, L.L., Mintun, M.A., Mach, R.H., 2018. Development of a Positron Emission Tomography Radiotracer for Imaging Elevated Levels of Superoxide in Neuroinflammation. *Acs Chem Neurosci* 9, 578–586. <https://doi.org/10.1021/acscchemneuro.7b00385>
- Huang, B.K., Sikes, H.D., 2014. Quantifying intracellular hydrogen peroxide perturbations in terms of concentration. *Redox Biol* 2, 955–962. <https://doi.org/10.1016/j.redox.2014.08.001>
- Huang, J., Co, H.K., Lee, Y., Wu, C., Chen, S., 2021. Multistability maintains redox homeostasis in human cells. *Mol. Syst. Biol.* 17, e10480. <https://doi.org/10.15252/msb.202110480>
- Hughes, A.J., Spelke, D.P., Xu, Z., Kang, C.-C., Schaffer, D.V., Herr, A.E., 2014. Single-cell western blotting. *Nat Methods* 11, 749–755. <https://doi.org/10.1038/nmeth.2992>

- Illés, E., Mizrahi, A., Marks, V., Meyerstein, D., 2019. Carbonate-radical-anions, and not hydroxyl radicals, are the products of the Fenton reaction in neutral solutions containing bicarbonate. *Free Radic. Biol. Med.* 131, 1–6. <https://doi.org/10.1016/j.freeradbiomed.2018.11.015>
- Imlay, J.A., 2013. The molecular mechanisms and physiological consequences of oxidative stress: lessons from a model bacterium. *Nat Rev Microbiol* 11, 443–454. <https://doi.org/10.1038/nrmicro3032>
- Imlay, J.A., 2008. Cellular Defenses against Superoxide and Hydrogen Peroxide. *Annu. Rev. Biochem.* 77, 755–776. <https://doi.org/10.1146/annurev.biochem.77.061606.161055>
- Jacob, H.S., Jandl, J.H., 1966. Effects of Sulfhydryl Inhibition on Red Blood Cells III. GLUTATHIONE IN THE REGULATION OF THE HEXOSE MONOPHOSPHATE PATHWAY. *J Biol Chem* 241, 4243–4250. [https://doi.org/10.1016/s0021-9258\(18\)99775-9](https://doi.org/10.1016/s0021-9258(18)99775-9)
- Janes, K.A., 2015. An analysis of critical factors for quantitative immunoblotting. *Sci Signal* 8, rs2. <https://doi.org/10.1126/scisignal.2005966>
- Jiang, X., Stockwell, B.R., Conrad, M., 2021. Ferroptosis: mechanisms, biology and role in disease. *Nat Rev Mol Cell Bio* 22, 266–282. <https://doi.org/10.1038/s41580-020-00324-8>
- Jones, D.P., 2006. Redefining Oxidative Stress. *Antioxid Redox Sign* 8, 1865–1879. <https://doi.org/10.1089/ars.2006.8.1865>
- Jones, D.P., Sies, H., 2015. The Redox Code. *Antioxid Redox Sign* 23, 734–746. <https://doi.org/10.1089/ars.2015.6247>
- Jordan, A.C., Perry, C.G.R., Cheng, A.J., 2021. Promoting a pro-oxidant state in skeletal muscle: Potential dietary, environmental, and exercise interventions for enhancing endurance-training adaptations. *Free Radic. Biol. Med.* 176, 189–202. <https://doi.org/10.1016/j.freeradbiomed.2021.09.014>
- Kagan, V.E., Mao, G., Qu, F., Angeli, J.P.F., Doll, S., Croix, C.S., Dar, H.H., Liu, B., Tyurin, V.A., Ritov, V.B., Kapralov, A.A., Amoscato, A.A., Jiang, J., Anthonymuthu, T., Mohammadyani, D., Yang, Q., Proneth, B., Klein-Seetharaman, J., Watkins, S., Bahar, I., Greenberger, J., Mallampalli, R.K., Stockwell, B.R., Tyurina, Y.Y., Conrad, M., Bayır, H., 2017. Oxidized arachidonic and adrenic PEs navigate cells to ferroptosis. *Nat Chem Biol* 13, 81–90. <https://doi.org/10.1038/nchembio.2238>
- Kalyanaraman, B., 2022. NAC, NAC, Knockin' on Heaven's door: Interpreting the mechanism of action of N-acetylcysteine in tumor and immune cells. *Redox Biol.* 57, 102497. <https://doi.org/10.1016/j.redox.2022.102497>
- Karplus, P.A., 2015. A primer on peroxiredoxin biochemistry. *Free Radical Bio Med* 80, 183–190. <https://doi.org/10.1016/j.freeradbiomed.2014.10.009>
- Keeley, T.P., Mann, G.E., 2019. Defining Physiological Normoxia for Improved Translation of Cell Physiology to Animal Models and Humans. *Physiol. Rev.* 99, 161–234. <https://doi.org/10.1152/physrev.00041.2017>
- Kerins, M.J., Milligan, J., Wohlschlegel, J.A., Ooi, A., 2018. Fumarate hydratase inactivation in hereditary leiomyomatosis and renal cell cancer is synthetic lethal with ferroptosis induction. *Cancer Sci.* 109, 2757–2766. <https://doi.org/10.1111/cas.13701>
- Kettle, A.J., Ashby, L.V., Winterbourn, C.C., Dickerhof, N., 2023. Superoxide: The enigmatic chemical chameleon in neutrophil biology. *Immunol. Rev.* 314, 181–196. <https://doi.org/10.1111/imr.13183>
- Khassaf, M., McArdle, A., Esanu, C., Vasilaki, A., McArdle, F., Griffiths, R.D., Brodie, D.A., Jackson, M.J., 2003. Effect of Vitamin C Supplements on Antioxidant Defence and Stress Proteins in Human Lymphocytes and Skeletal Muscle. *J. Physiol.* 549, 645–652. <https://doi.org/10.1113/jphysiol.2003.040303>
- Kim, H., Ha, S., Lee, H.Y., Lee, K., 2015. ROSics: Chemistry and proteomics of cysteine modifications in redox biology. *Mass Spectrom Rev* 34, 184–208. <https://doi.org/10.1002/mas.21430>
- Kitamura, N., Galligan, J.J., 2023. A global view of the human post-translational modification landscape. *Biochem. J.* 480, 1241–1265. <https://doi.org/10.1042/bcj20220251>
- Kitano, H., 2002. Computational systems biology. *Nature* 420, 206–210. <https://doi.org/10.1038/nature01254>
- Kono, Y., Fridovich, I., 1982. Superoxide radical inhibits catalase. *J. Biol. Chem.* 257, 5751–5754. [https://doi.org/10.1016/s0021-9258\(19\)83842-5](https://doi.org/10.1016/s0021-9258(19)83842-5)
- Koppenol, W.H., 2022. Ferryl for real. The Fenton reaction near neutral pH. *Dalton Trans.* 51, 17496–17502. <https://doi.org/10.1039/d2dt03168j>
- Kramer, P.A., Duan, J., Gaffrey, M.J., Shukla, A.K., Wang, L., Bammler, T.K., Qian, W.-J., Marcinek, D.J., 2018. Fatiguing contractions increase protein S-glutathionylation occupancy in mouse skeletal muscle. *Redox Biol.* 17, 367–376. <https://doi.org/10.1016/j.redox.2018.05.011>
- Kritsiligkou, P., Bosch, K., Shen, T.K., Meurer, M., Knop, M., Dick, T.P., 2023. Proteome-wide tagging with an H₂O₂ biosensor reveals highly localized and dynamic redox microenvironments. *Proc. Natl. Acad. Sci.* 120, e2314043120. <https://doi.org/10.1073/pnas.2314043120>
- Lancaster, J.R., 1997. A Tutorial on the Diffusibility and Reactivity of Free Nitric Oxide. *Nitric Oxide* 1, 18–30. <https://doi.org/10.1006/niox.1996.0112>
- Langford, T.F., Deen, W.M., Sikes, H.D., 2018. A mathematical analysis of Prx2-STAT3 disulfide exchange rate constants for a bimolecular reaction mechanism. *Free Radical Bio Med* 120, 239–245. <https://doi.org/10.1016/j.freeradbiomed.2018.03.039>

- Larsen, E.L., Karstoft, K., Poulsen, H.E., 2022. Exercise and RNA oxidation, in: Copley, J.N., Davison, G.W. (Eds.), *Oxidative Eustress in Exercise Physiology*.
- Larsen, E.L., Weimann, A., Poulsen, H.E., 2019. Interventions targeted at oxidatively generated modifications of nucleic acids focused on urine and plasma markers. *Free Radic. Biol. Med.* 145, 256–283. <https://doi.org/10.1016/j.freeradbiomed.2019.09.030>
- Lee, Y., Chang, G., 2019. Quantitative display of the redox status of proteins with maleimide-polyethylene glycol tagging. *Electrophoresis* 40, 491–498. <https://doi.org/10.1002/elps.201800335>
- Leeuwen, L.A.G. van, Hinchey, E.C., Murphy, M.P., Robb, E.L., Cochemé, H.M., 2017. Click-PEGylation – A mobility shift approach to assess the redox state of cysteines in candidate proteins. *Free Radical Bio Med* 108, 374–382. <https://doi.org/10.1016/j.freeradbiomed.2017.03.037>
- Leichert, L.I., Gehrke, F., Gudiseva, H.V., Blackwell, T., Ilbert, M., Walker, A.K., Strahler, J.R., Andrews, P.C., Jakob, U., 2008. Quantifying changes in the thiol redox proteome upon oxidative stress in vivo. *Proc National Acad Sci* 105, 8197–8202. <https://doi.org/10.1073/pnas.0707723105>
- Lennicke, C., Cochemé, H.M., 2021. Redox metabolism: ROS as specific molecular regulators of cell signaling and function. *Mol Cell* 81, 3691–3707. <https://doi.org/10.1016/j.molcel.2021.08.018>
- Li, W., Sancar, A., 2020. Methodologies for detecting environmentally induced DNA damage and repair. *Environ. Mol. Mutagen.* 61, 664–679. <https://doi.org/10.1002/em.22365>
- Li, X., Gluth, A., Zhang, T., Qian, W., 2023. Thiol redox proteomics: Characterization of thiol-based post-translational modifications. *Proteomics* e2200194. <https://doi.org/10.1002/pmic.202200194>
- Lim, C.H., Dedon, P.C., Deen, W.M., 2008. Kinetic Analysis of Intracellular Concentrations of Reactive Nitrogen Species. *Chem. Res. Toxicol.* 21, 2134–2147. <https://doi.org/10.1021/tx800213b>
- Lim, J.B., Huang, B.K., Deen, W.M., Sikes, H.D., 2015. Analysis of the lifetime and spatial localization of hydrogen peroxide generated in the cytosol using a reduced kinetic model. *Free Radical Bio Med* 89, 47–53. <https://doi.org/10.1016/j.freeradbiomed.2015.07.009>
- Lim, J.B., Langford, T.F., Huang, B.K., Deen, W.M., Sikes, H.D., 2016. A reaction-diffusion model of cytosolic hydrogen peroxide. *Free Radical Bio Med* 90, 85–90. <https://doi.org/10.1016/j.freeradbiomed.2015.11.005>
- Liochev, S.I., Fridovich, I., 2002. Copper, Zinc Superoxide Dismutase and H₂O₂ EFFECTS OF BICARBONATE ON INACTIVATION AND OXIDATIONS OF NADPH AND URATE, AND ON CONSUMPTION OF H₂O₂ *. *J. Biol. Chem.* 277, 34674–34678. <https://doi.org/10.1074/jbc.m204726200>
- Lisi, V., Moulton, C., Fantini, C., Grazioli, E., Guidotti, F., Sgrò, P., Dimauro, I., Capranica, L., Parisi, A., Luigi, L.D., Caporossi, D., 2023. Steady-state redox status in circulating extracellular vesicles: A proof-of-principle study on the role of fitness level and short-term aerobic training in healthy young males. *Free Radic. Biol. Med.* 204, 266–275. <https://doi.org/10.1016/j.freeradbiomed.2023.05.007>
- Liu, Y., Herr, A.E., 2023. DropBlot: single-cell western blotting of chemically fixed cancer cells. <https://doi.org/10.1101/2023.09.04.556277>
- Low, F.M., Hampton, M.B., Peskin, A.V., Winterbourn, C.C., 2006. Peroxiredoxin 2 functions as a noncatalytic scavenger of low-level hydrogen peroxide in the erythrocyte. *Blood* 109, 2611–2617. <https://doi.org/10.1182/blood-2006-09-048728>
- Low, F.M., Hampton, M.B., Winterbourn, C.C., 2008. Peroxiredoxin 2 and Peroxide Metabolism in the Erythrocyte. *Antioxid Redox Sign* 10, 1621–1630. <https://doi.org/10.1089/ars.2008.2081>
- Lyons, J., Rauh-Pfeiffer, A., Yu, Y.M., Lu, X.-M., Zurakowski, D., Tompkins, R.G., Ajami, A.M., Young, V.R., Castillo, L., 2000. Blood glutathione synthesis rates in healthy adults receiving a sulfur amino acid-free diet. *Proc. Natl. Acad. Sci.* 97, 5071–5076. <https://doi.org/10.1073/pnas.090083297>
- MacDonald, J.I., Munch, H.K., Moore, T., Francis, M.B., 2015. One-step site-specific modification of native proteins with 2-pyridinecarboxyaldehydes. *Nat Chem Biol* 11, 326–331. <https://doi.org/10.1038/nchembio.1792>
- MacMillan-Crow, L.A., Crow, J.P., Kerby, J.D., Beckman, J.S., Thompson, J.A., 1996. Nitration and inactivation of manganese superoxide dismutase in chronic rejection of human renal allografts. *Proc. Natl. Acad. Sci.* 93, 11853–11858. <https://doi.org/10.1073/pnas.93.21.11853>
- Madani, A., Krause, B., Greene, E.R., Subramanian, S., Mohr, B.P., Holton, J.M., Olmos, J.L., Xiong, C., Sun, Z.Z., Socher, R., Fraser, J.S., Naik, N., 2023. Large language models generate functional protein sequences across diverse families. *Nat Biotechnol* 1–8. <https://doi.org/10.1038/s41587-022-01618-2>
- Madian, A.G., Diaz-Maldonado, N., Gao, Q., Regnier, F.E., 2011. Oxidative stress induced carbonylation in human plasma. *J Proteomics* 74, 2395–2416. <https://doi.org/10.1016/j.jprot.2011.07.014>
- Madian, A.G., Regnier, F.E., 2010. Profiling Carbonylated Proteins in Human Plasma. *J Proteome Res* 9, 1330–1343. <https://doi.org/10.1021/pr900890k>
- Mailloux, R.J., 2021. An update on methods and approaches for interrogating mitochondrial reactive oxygen species production. *Redox Biol* 45, 102044. <https://doi.org/10.1016/j.redox.2021.102044>
- Makmura, L., Hamann, M., Areopagita, A., Furuta, S., Muoz, A., Momand, J., 2001. Development of a Sensitive Assay to Detect Reversibly Oxidized Protein Cysteine Sulfhydryl Groups. *Antioxid Redox Sign* 3, 1105–1118. <https://doi.org/10.1089/152308601317203611>

- Malik, Z.A., Cobley, J.N., Morton, J.P., Close, G.L., Edwards, B.J., Koch, L.G., Britton, S.L., Burniston, J.G., 2013. Label-Free LC-MS Profiling of Skeletal Muscle Reveals Heart-Type Fatty Acid Binding Protein as a Candidate Biomarker of Aerobic Capacity. *Proteomes* 1, 290–308. <https://doi.org/10.3390/proteomes1030290>
- Manford, A.G., Mena, E.L., Shih, K.Y., Gee, C.L., McMinimy, R., Martínez-González, B., Sherriff, R., Lew, B., Zoltek, M., Rodríguez-Pérez, F., Woldesenbet, M., Kuriyan, J., Rape, M., 2021. Structural basis and regulation of the reductive stress response. *Cell*. <https://doi.org/10.1016/j.cell.2021.09.002>
- Manford, A.G., Rodríguez-Pérez, F., Shih, K.Y., Shi, Z., Berdan, C.A., Choe, M., Titov, D.V., Nomura, D.K., Rape, M., 2020. A Cellular Mechanism to Detect and Alleviate Reductive Stress. *Cell* 183, 46–61.e21. <https://doi.org/10.1016/j.cell.2020.08.034>
- Margaritelis, N.V., 2023. Personalized redox biology: Designs and concepts. *Free Radic. Biol. Med.* 208, 112–125. <https://doi.org/10.1016/j.freeradbiomed.2023.08.003>
- Margaritelis, N.V., Chatzinikolaou, P.N., Chatzinikolaou, A.N., Paschalis, V., Theodorou, A.A., Vrabas, I.S., Kyparos, A., Nikolaidis, M.G., 2022. The redox signal: A physiological perspective. *IUBMB Life* 74, 29–40. <https://doi.org/10.1002/iub.2550>
- Margaritelis, N.V., Cobley, J.N., Paschalis, V., Veskoukis, A.S., Theodorou, A.A., Kyparos, A., Nikolaidis, M.G., 2016. Going retro: Oxidative stress biomarkers in modern redox biology. *Free Radical Bio Med* 98, 2–12. <https://doi.org/10.1016/j.freeradbiomed.2016.02.005>
- Margaritelis, Nikos V., Cobley, J.N., Paschalis, V., Veskoukis, A.S., Theodorou, A.A., Kyparos, A., Nikolaidis, M.G., 2016. Principles for integrating reactive species into in vivo biological processes: Examples from exercise physiology. *Cell Signal* 28, 256–271. <https://doi.org/10.1016/j.cellsig.2015.12.011>
- Margaritelis, N.V., Kyparos, A., Paschalis, V., Theodorou, A.A., Panayiotou, G., Zafeiridis, A., Dipla, K., Nikolaidis, M.G., Vrabas, I.S., 2014. Reductive stress after exercise: The issue of redox individuality. *Redox Biol.* 2, 520–528. <https://doi.org/10.1016/j.redox.2014.02.003>
- Margaritelis, N.V., Nastos, G.G., Vasileiadou, O., Chatzinikolaou, P.N., Theodorou, A.A., Paschalis, V., Vrabas, I.S., Kyparos, A., Fatouros, I.G., Nikolaidis, M.G., 2023. Inter-individual variability in redox and performance responses after antioxidant supplementation: A randomized double blind crossover study. *Acta Physiol.* 238, e14017. <https://doi.org/10.1111/apha.14017>
- Margaritelis, N.V., Paschalis, V., Theodorou, A.A., Kyparos, A., Nikolaidis, M.G., 2020. Redox basis of exercise physiology. *Redox Biol* 35, 101499. <https://doi.org/10.1016/j.redox.2020.101499>
- Margaritelis, N.V., Theodorou, A.A., Paschalis, V., Veskoukis, A.S., Dipla, K., Zafeiridis, A., Panayiotou, G., Vrabas, I.S., Kyparos, A., Nikolaidis, M.G., 2018. Adaptations to endurance training depend on exercise-induced oxidative stress: exploiting redox interindividual variability. *Acta Physiol.* 222, e12898. <https://doi.org/10.1111/apha.12898>
- Marinho, H.S., Real, C., Cyrne, L., Soares, H., Antunes, F., 2014. Hydrogen peroxide sensing, signaling and regulation of transcription factors. *Redox Biol* 2, 535–562. <https://doi.org/10.1016/j.redox.2014.02.006>
- Martin-Baniandres, P., Lan, W.-H., Board, S., Romero-Ruiz, M., Garcia-Manyes, S., Qing, Y., Bayley, H., 2023. Enzyme-less nanopore detection of post-translational modifications within long polypeptides. *Nat. Nanotechnol.* 1–6. <https://doi.org/10.1038/s41565-023-01462-8>
- Marx, V., 2013. Finding the right antibody for the job. *Nat Methods* 10, 703–707. <https://doi.org/10.1038/nmeth.2570>
- Mason, R.P., Ganini, D., 2019. Immuno-spin trapping of macromolecules free radicals in vitro and in vivo – One stop shopping for free radical detection. *Free Radical Bio Med* 131, 318–331. <https://doi.org/10.1016/j.freeradbiomed.2018.11.009>
- Mason, S.A., Trewin, A.J., Parker, L., Wadley, G.D., 2020. Antioxidant supplements and endurance exercise: Current evidence and mechanistic insights. *Redox Biol* 35, 101471. <https://doi.org/10.1016/j.redox.2020.101471>
- McCord, J.M., Fridovich, I., 1969. Superoxide Dismutase AN ENZYMIC FUNCTION FOR ERYTHROCUPREIN (HEMOCUPREIN). *J Biol Chem* 244, 6049–6055. [https://doi.org/10.1016/s0021-9258\(18\)63504-5](https://doi.org/10.1016/s0021-9258(18)63504-5)
- McCord, J.M., Fridovich, I., 1968. The Reduction of Cytochrome c by Milk Xanthine Oxidase. *J. Biol. Chem.* 243, 5753–5760. [https://doi.org/10.1016/s0021-9258\(18\)91929-0](https://doi.org/10.1016/s0021-9258(18)91929-0)
- Melo, T., Montero-Bullón, J.-F., Domingues, P., Domingues, M.R., 2019. Discovery of bioactive nitrated lipids and nitro-lipid-protein adducts using mass spectrometry-based approaches. *Redox Biol.* 23, 101106. <https://doi.org/10.1016/j.redox.2019.101106>
- Meo, S.D., Venditti, P., 2020. Evolution of the Knowledge of Free Radicals and Other Oxidants. *Oxidative Med. Cell. Longev.* 2020, 9829176. <https://doi.org/10.1155/2020/9829176>
- MICHAILIDIS, Y., JAMURTAS, A.Z., NIKOLAIDIS, M.G., FATOUIROS, I.G., KOUTEDAKIS, Y., PAPANOTIRIOU, I., KOURETAS, D., 2007. Sampling Time is Crucial for Measurement of Aerobic Exercise-Induced Oxidative Stress. *Med. Sci. Sports Exerc.* 39, 1107–1113. <https://doi.org/10.1249/01.mss.0b013e318053e7ba>

- Milne, G.L., Nogueira, M.S., Gao, B., Sanchez, S.C., Amin, W., Thomas, S., Oger, C., Galano, J.-M., Murff, H.J., Yang, G., Durand, T., 2024. Identification of novel F2-isoprostane metabolites by specific UDP-glucuronosyltransferases. *Redox Biol.* 70, 103020. <https://doi.org/10.1016/j.redox.2023.103020>
- Milo, R., Phillips, R., 2006. *Cell Biology by the numbers*, 2nd Edition. ed. Garland Science, New York.
- Misra, H.P., 1974. Generation of Superoxide Free Radical during the Autoxidation of Thiols. *J. Biol. Chem.* 249, 2151–2155. [https://doi.org/10.1016/s0021-9258\(19\)42810-x](https://doi.org/10.1016/s0021-9258(19)42810-x)
- Misra, H.P., Fridovich, I., 1972. The Role of Superoxide Anion in the Autoxidation of Epinephrine and a Simple Assay for Superoxide Dismutase. *J. Biol. Chem.* 247, 3170–3175. [https://doi.org/10.1016/s0021-9258\(19\)45228-9](https://doi.org/10.1016/s0021-9258(19)45228-9)
- Mogilner, A., Wollman, R., Marshall, W.F., 2006. Quantitative Modeling in Cell Biology: What Is It Good for? *Dev. Cell* 11, 279–287. <https://doi.org/10.1016/j.devcel.2006.08.004>
- Moi, P., Chan, K., Asunis, I., Cao, A., Kan, Y.W., 1994. Isolation of NF-E2-related factor 2 (Nrf2), a NF-E2-like basic leucine zipper transcriptional activator that binds to the tandem NF-E2/AP1 repeat of the beta-globin locus control region. *Proc. Natl. Acad. Sci.* 91, 9926–9930. <https://doi.org/10.1073/pnas.91.21.9926>
- Muggeridge, D.J., Crabtree, D.R., Tuncay, A., Megson, I.L., Davison, G., Cobley, J.N., 2022. Exercise decreases PP2A-specific reversible thiol oxidation in human erythrocytes: Implications for redox biomarkers. *Free Radical Bio Med* 182, 73–78. <https://doi.org/10.1016/j.freeradbiomed.2022.02.019>
- Murphy, M.P., 2014. Antioxidants as therapies: can we improve on nature? *Free Radical Bio Med* 66, 20–23. <https://doi.org/10.1016/j.freeradbiomed.2013.04.010>
- Murphy, M.P., 2009. How mitochondria produce reactive oxygen species. *Biochem J* 417, 1–13. <https://doi.org/10.1042/bj20081386>
- Murphy, M.P., Bayir, H., Belousov, V., Chang, C.J., Davies, K.J.A., Davies, M.J., Dick, T.P., Finkel, T., Forman, H.J., Janssen-Heininger, Y., Gems, D., Kagan, V.E., Kalyanaraman, B., Larsson, N.-G., Milne, G.L., Nyström, T., Poulsen, H.E., Radi, R., Remmen, H.V., Schumacker, P.T., Thornalley, P.J., Toyokuni, S., Winterbourn, C.C., Yin, H., Halliwell, B., 2022. Guidelines for measuring reactive oxygen species and oxidative damage in cells and in vivo. *Nat Metabolism* 4, 651–662. <https://doi.org/10.1038/s42255-022-00591-z>
- Murphy, M.P., Hartley, R.C., 2018. Mitochondria as a therapeutic target for common pathologies. *Nat Rev Drug Discov* 17, 865–886. <https://doi.org/10.1038/nrd.2018.174>
- Murphy, M.P., Holmgren, A., Larsson, N.-G., Halliwell, B., Chang, C.J., Kalyanaraman, B., Rhee, S.G., Thornalley, P.J., Partridge, L., Gems, D., Nyström, T., Belousov, V., Schumacker, P.T., Winterbourn, C.C., 2011. Unraveling the Biological Roles of Reactive Oxygen Species. *Cell Metab* 13, 361–366. <https://doi.org/10.1016/j.cmet.2011.03.010>
- Neuman, E.W., 1934. Potassium Superoxide and the Three-Electron Bond. *J. Chem. Phys.* 2, 31–33. <https://doi.org/10.1063/1.1749353>
- Niki, E., 2009. Lipid peroxidation: Physiological levels and dual biological effects. *Free Radic. Biol. Med.* 47, 469–484. <https://doi.org/10.1016/j.freeradbiomed.2009.05.032>
- Nikolaidis, M., Margaritelis, N., Matsakas, A., 2020. Quantitative Redox Biology of Exercise. *Int. J. Sports Med.* 41, 633–645. <https://doi.org/10.1055/a-1157-9043>
- Nikolaidis, M.G., Kerksick, C.M., Lamprecht, M., McAnulty, S.R., 2012. Does Vitamin C and E Supplementation Impair the Favorable Adaptations of Regular Exercise? *Oxidative Med. Cell. Longev.* 2012, 707941. <https://doi.org/10.1155/2012/707941>
- Nikolaidis, M.G., Kyparos, A., Vrabas, I.S., 2011. F2-isoprostane formation, measurement and interpretation: The role of exercise. *Prog. Lipid Res.* 50, 89–103. <https://doi.org/10.1016/j.plipres.2010.10.002>
- Nikolaidis, M.G., Margaritelis, N.V., 2023. Free radicals and antioxidants: appealing to magic. *Trends Endocrinol. Metab.* <https://doi.org/10.1016/j.tem.2023.06.001>
- Nikolaidis, M.G., Margaritelis, N.V., 2018. Same Redox Evidence But Different Physiological “Stories”: The Rashomon Effect in Biology. *Bioessays* 40, 1800041. <https://doi.org/10.1002/bies.201800041>
- Noble, A., Guille, M., Cobley, J.N., 2021. ALISA: A microplate assay to measure protein thiol redox state. *Free Radical Bio Med* 174, 272–280. <https://doi.org/10.1016/j.freeradbiomed.2021.08.018>
- Ogilby, P.R., 2010. Singlet oxygen : there is indeed something new under the sun. *Chem Soc Rev* 39, 3181–3209. <https://doi.org/10.1039/b926014p>
- Orrico, F., Möller, M.N., Cassina, A., Denicola, A., Thomson, L., 2018. Kinetic and stoichiometric constraints determine the pathway of H2O2 consumption by red blood cells. *Free Radic. Biol. Med.* 121, 231–239. <https://doi.org/10.1016/j.freeradbiomed.2018.05.006>
- Ostrom, E.L., Berry, S.R., Traustadóttir, T., 2021a. Effects of exercise training on redox stress resilience in young and older adults. *Adv. Redox Res.* 2, 100007. <https://doi.org/10.1016/j.arres.2021.100007>
- Ostrom, E.L., Traustadóttir, T., 2020. Aerobic exercise training partially reverses the impairment of Nrf2 activation in older humans. *Free Radic. Biol. Med.* 160, 418–432. <https://doi.org/10.1016/j.freeradbiomed.2020.08.016>

- Ostrom, E.L., Valencia, A.P., Marcinek, D.J., Traustadóttir, T., 2021b. High intensity muscle stimulation activates a systemic Nrf2-mediated redox stress response. *Free Radic. Biol. Med.* 172, 82–89. <https://doi.org/10.1016/j.freeradbiomed.2021.05.039>
- Owens, D.J., Twist, C., Cobley, J.N., Howatson, G., Close, G.L., 2018. Exercise-induced muscle damage: What is it, what causes it and what are the nutritional solutions? *Eur J Sport Sci* 19, 1–15. <https://doi.org/10.1080/17461391.2018.1505957>
- Pak, V.V., Ezeriņa, D., Lyublinskaya, O.G., Pedre, B., Tyurin-Kuzmin, P.A., Mishina, N.M., Thauvin, M., Young, D., Wahni, K., Gache, S.A.M., Demidovich, A.D., Ermakova, Y.G., Maslova, Y.D., Shokhina, A.G., Eroglu, E., Bilan, D.S., Bogeski, I., Michel, T., Vríz, S., Messens, J., Belousov, V.V., 2020. Ultrasensitive Genetically Encoded Indicator for Hydrogen Peroxide Identifies Roles for the Oxidant in Cell Migration and Mitochondrial Function. *Cell Metab* 31, 642–653.e6. <https://doi.org/10.1016/j.cmet.2020.02.003>
- Parvez, S., Long, M.J.C., Poganik, J.R., Aye, Y., 2018. Redox Signaling by Reactive Electrophiles and Oxidants. *Chem Rev* 118, 8798–8888. <https://doi.org/10.1021/acs.chemrev.7b00698>
- Paschalis, V., Theodorou, A.A., Margaritelis, N.V., Kyparos, A., Nikolaidis, M.G., 2018. N-acetylcysteine supplementation increases exercise performance and reduces oxidative stress only in individuals with low levels of glutathione. *Free Radic. Biol. Med.* 115, 288–297. <https://doi.org/10.1016/j.freeradbiomed.2017.12.007>
- Paulsen, C.E., Carroll, K.S., 2013. Cysteine-Mediated Redox Signaling: Chemistry, Biology, and Tools for Discovery. *Chem Rev* 113, 4633–4679. <https://doi.org/10.1021/cr300163e>
- Pedre, B., Barayeu, U., Ezeriņa, D., Dick, T.P., 2021. The mechanism of action of N-acetylcysteine (NAC): The emerging role of H₂S and sulfane sulfur species. *Pharmacol Therapeut* 228, 107916. <https://doi.org/10.1016/j.pharmthera.2021.107916>
- Peskin, A.V., Dickerhof, N., Poynton, R.A., Paton, L.N., Pace, P.E., Hampton, M.B., Winterbourn, C.C., 2013. Hyperoxidation of Peroxiredoxins 2 and 3 RATE CONSTANTS FOR THE REACTIONS OF THE SULFENIC ACID OF THE PEROXIDATIC CYSTEINE*. *J Biol Chem* 288, 14170–14177. <https://doi.org/10.1074/jbc.m113.460881>
- Peskin, A.V., Low, F.M., Paton, L.N., Maghzal, G.J., Hampton, M.B., Winterbourn, C.C., 2007. The High Reactivity of Peroxiredoxin 2 with H₂O₂ Is Not Reflected in Its Reaction with Other Oxidants and Thiol Reagents*. *J. Biol. Chem.* 282, 11885–11892. <https://doi.org/10.1074/jbc.m700339200>
- Peskin, A.V., Winterbourn, C.C., 2017. Assay of superoxide dismutase activity in a plate assay using WST-1. *Free Radical Bio Med* 103, 188–191. <https://doi.org/10.1016/j.freeradbiomed.2016.12.033>
- Phillips, R., Kondev, J., Theriot, J., Garcia, H., 2012. *Physical biology of the cell*, 2nd Edition. ed. Garland Science, New York.
- Pillay, C.S., Eagling, B.D., Driscoll, S.R.E., Rohwer, J.M., 2016. Quantitative measures for redox signaling. *Free Radical Bio Med* 96, 290–303. <https://doi.org/10.1016/j.freeradbiomed.2016.04.199>
- Pillay, C.S., Hofmeyr, J.-H., Mashamaite, L.N., Rohwer, J.M., 2013. From Top-Down to Bottom-Up: Computational Modeling Approaches for Cellular Redoxin Networks. *Antioxid. Redox Signal.* 18, 2075–2086. <https://doi.org/10.1089/ars.2012.4771>
- Place, N., Ivarsson, N., Venckunas, T., Neyroud, D., Brazaitis, M., Cheng, A.J., Ochala, J., Kamandulis, S., Girard, S., Volungevičius, G., Paužas, H., Mekideche, A., Kayser, B., Martinez-Redondo, V., Ruas, J.L., Bruton, J., Truffert, A., Lanner, J.T., Skurvydas, A., Westerblad, H., 2015. Ryanodine receptor fragmentation and sarcoplasmic reticulum Ca²⁺ leak after one session of high-intensity interval exercise. *Proc. Natl. Acad. Sci.* 112, 15492–15497. <https://doi.org/10.1073/pnas.1507176112>
- Poetsch, A.R., Boulton, S.J., Luscombe, N.M., 2018. Genomic landscape of oxidative DNA damage and repair reveals regioselective protection from mutagenesis. *Genome Biol.* 19, 215. <https://doi.org/10.1186/s13059-018-1582-2>
- Poole, L.B., Furdai, C.M., King, S.B., 2020. Introduction to approaches and tools for the evaluation of protein cysteine oxidation. *Essays Biochem* 64, 1–17. <https://doi.org/10.1042/ebc20190050>
- Poulsen, H.E., Weimann, A., Henriksen, T., Kjær, L.K., Larsen, E.L., Carlsson, E.R., Christensen, C.K., Brandslund, I., Fenger, M., 2019. Oxidatively generated modifications to nucleic acids in vivo: Measurement in urine and plasma. *Free Radic. Biol. Med.* 145, 336–341. <https://doi.org/10.1016/j.freeradbiomed.2019.10.001>
- Powers, S.K., Deminice, R., Ozdemir, M., Yoshihara, T., Bomkamp, M.P., Hyatt, H., 2020. Exercise-induced oxidative stress: Friend or foe? *J. Sport Heal. Sci.* 9, 415–425. <https://doi.org/10.1016/j.jshs.2020.04.001>
- Powers, S.K., Goldstein, E., Schrager, M., Ji, L.L., 2022. Exercise Training and Skeletal Muscle Antioxidant Enzymes: An Update. *Antioxidants* 12, 39. <https://doi.org/10.3390/antiox12010039>
- Powers, S.K., Radak, Z., Ji, L.L., 2016. Exercise-induced oxidative stress: past, present and future. *J. Physiol.* 594, 5081–5092. <https://doi.org/10.1113/jp270646>
- Pregel, M.J., Storer, A.C., 1997. Active Site Titration of the Tyrosine Phosphatases SHP-1 and PTP1B Using Aromatic Disulfides REACTION WITH THE ESSENTIAL CYSTEINE RESIDUE IN THE ACTIVE SITE*. *J Biol Chem* 272, 23552–23558. <https://doi.org/10.1074/jbc.272.38.23552>

- Pugh, J.N., Stretton, C., McDonagh, B., Brownridge, P., McArdle, A., Jackson, M.J., Close, G.L., 2021. Exercise stress leads to an acute loss of mitochondrial proteins and disruption of redox control in skeletal muscle of older subjects: An underlying decrease in resilience with aging? *Free Radical Bio Med* 177, 88–99. <https://doi.org/10.1016/j.freeradbiomed.2021.10.003>
- Radi, R., 2022. Interplay of carbon dioxide and peroxide metabolism in mammalian cells. *J Biol Chem* 102358. <https://doi.org/10.1016/j.jbc.2022.102358>
- Radi, R., 2018. Oxygen radicals, nitric oxide, and peroxynitrite: Redox pathways in molecular medicine. *Proc National Acad Sci* 115, 5839–5848. <https://doi.org/10.1073/pnas.1804932115>
- Raftos, J.E., Whillier, S., Kuchel, P.W., 2010. Glutathione Synthesis and Turnover in the Human Erythrocyte ALIGNMENT OF A MODEL BASED ON DETAILED ENZYME KINETICS WITH EXPERIMENTAL DATA. *J. Biol. Chem.* 285, 23557–23567. <https://doi.org/10.1074/jbc.m109.067017>
- Ravanat, J.-L., Guicherd, P., Tuce, Z., Cadet, J., 1999. Simultaneous Determination of Five Oxidative DNA Lesions in Human Urine. *Chem. Res. Toxicol.* 12, 802–808. <https://doi.org/10.1021/tx980194k>
- Reid, M.B., 2008. Free radicals and muscle fatigue: Of ROS, canaries, and the IOC. *Free Radic. Biol. Med.* 44, 169–179. <https://doi.org/10.1016/j.freeradbiomed.2007.03.002>
- Reisz, J.A., Bechtold, E., King, S.B., Poole, L.B., Furdul, C.M., 2013. Thiol-blocking electrophiles interfere with labeling and detection of protein sulfenic acids. *Febs J* 280, 6150–6161. <https://doi.org/10.1111/febs.12535>
- Richardson, R.S., Donato, A.J., Uberoi, A., Wray, D.W., Lawrenson, L., Nishiyama, S., Bailey, D.M., 2007. Exercise-induced brachial artery vasodilation: role of free radicals. *Am. J. Physiol.-Hear. Circ. Physiol.* 292, H1516–H1522. <https://doi.org/10.1152/ajpheart.01045.2006>
- Ristow, M., Zarse, K., Oberbach, A., Klötting, N., Birringer, M., Kiehnopf, M., Stumvoll, M., Kahn, C.R., Blüher, M., 2009. Antioxidants prevent health-promoting effects of physical exercise in humans. *Proc National Acad Sci* 106, 8665–8670. <https://doi.org/10.1073/pnas.0903485106>
- Rodriguez, R., Krishnan, Y., 2023. The chemistry of next-generation sequencing. *Nat. Biotechnol.* 41, 1709–1715. <https://doi.org/10.1038/s41587-023-01986-3>
- Roelofs, B.A., Ge, S.X., Studlack, P.E., Polster, B.M., 2015. Low micromolar concentrations of the superoxide probe MitoSOX uncouple neural mitochondria and inhibit complex IV. *Free Radic. Biol. Med.* 86, 250–258. <https://doi.org/10.1016/j.freeradbiomed.2015.05.032>
- Rosenberger, F.A., Thielert, M., Mann, M., 2023. Making single-cell proteomics biologically relevant. *Nat Methods* 20, 320–323. <https://doi.org/10.1038/s41592-023-01771-9>
- Safdar, A., Hamadeh, M.J., Kaczor, J.J., Raha, S., deBeer, J., Tarnopolsky, M.A., 2010. Aberrant Mitochondrial Homeostasis in the Skeletal Muscle of Sedentary Older Adults. *PLoS ONE* 5, e10778. <https://doi.org/10.1371/journal.pone.0010778>
- Salmeen, A., Andersen, J.N., Myers, M.P., Meng, T.-C., Hinks, J.A., Tonks, N.K., Barford, D., 2003. Redox regulation of protein tyrosine phosphatase 1B involves a sulphenyl-amide intermediate. *Nature* 423, 769–773. <https://doi.org/10.1038/nature01680>
- Salvador, A., 2024. Pillars of theoretical biology: “Biochemical systems analysis, I, II and III.” *J. Theor. Biol.* 576, 111655. <https://doi.org/10.1016/j.jtbi.2023.111655>
- Salvador, A., Antunes, A., 2023. How gradients and microdomains determine H₂O₂ redox signalling, in: Medrano-Fren, I., Bienert, G.P., Sitia, R. (Eds.), *Peroxisporins: Redox Signal Mediators In and Between Cells*. CRC Press.
- Salzano, S., Checconi, P., Hanschmann, E.-M., Lillig, C.H., Bowler, L.D., Chan, P., Vaudry, D., Mengozzi, M., Coppo, L., Sacre, S., Atkuri, K.R., Sahaf, B., Herzenberg, Leonard A., Herzenberg, Leonore A., Mullen, L., Ghezzi, P., 2014. Linkage of inflammation and oxidative stress via release of glutathionylated peroxiredoxin-2, which acts as a danger signal. *Proc National Acad Sci* 111, 12157–12162. <https://doi.org/10.1073/pnas.1401712111>
- Santos, J.H., Meyer, J.N., Mandavilli, B.S., Houten, B.V., 2006. DNA Repair Protocols, Mammalian Systems. *Methods Mol. Biol.* 314, 183–199. <https://doi.org/10.1385/1-59259-973-7:183>
- Saran, M., Bors, W., 1989. Oxygen Radicals Acting as Chemical Messengers: A Hypothesis. *Free Radical Res Com* 7, 213–220. <https://doi.org/10.3109/10715768909087944>
- Saurin, A.T., Neubert, H., Brennan, J.P., Eaton, P., 2004. Widespread sulfenic acid formation in tissues in response to hydrogen peroxide. *Proc National Acad Sci* 101, 17982–17987. <https://doi.org/10.1073/pnas.0404762101>
- Sawyer, D.T., Valentine, J.S., 1981. How super is superoxide? *Accounts Chem Res* 14, 393–400. <https://doi.org/10.1021/ar00072a005>
- Schilling, D., Barayeu, U., Steimbach, R.R., Talwar, D., Miller, A.K., Dick, T.P., 2022. Commonly Used Alkylating Agents Limit Persulfide Detection by Converting Protein Persulfides into Thioethers. *Angewandte Chemie Int Ed* 61, e202203684. <https://doi.org/10.1002/anie.202203684>
- Shchepinova, M.M., Cairns, A.G., Prime, T.A., Logan, A., James, A.M., Hall, A.R., Vidoni, S., Arndt, S., Caldwell, S.T., Prag, H.A., Pell, V.R., Krieg, T., Mulvey, J.F., Yadav, P., Copley, J.N., Bright, T.P., Senn, H.M., Anderson, R.F., Murphy, M.P., Hartley, R.C., 2017. MitoNeoD: A Mitochondria-Targeted Superoxide Probe. *Cell Chem Biol* 24, 1285–1298.e12. <https://doi.org/10.1016/j.chembiol.2017.08.003>

- Shi, Y., Carroll, K.S., 2020. Activity-Based Sensing for Site-Specific Proteomic Analysis of Cysteine Oxidation. *Accounts Chem Res* 53, 20–31. <https://doi.org/10.1021/acs.accounts.9b00562>
- Sidlauskaitė, E., Gibson, J.W., Megson, I.L., Whitfield, P.D., Tovmasyan, A., Batinic-Haberle, I., Murphy, M.P., Moulton, P.R., Cobley, J.N., 2018. Mitochondrial ROS cause motor deficits induced by synaptic inactivity: Implications for synapse pruning. *Redox Biol* 16, 344–351. <https://doi.org/10.1016/j.redox.2018.03.012>
- Sies, H., 2021. Oxidative Eustress: On Constant Alert for Redox Homeostasis. *Redox Biol* 41, 101867. <https://doi.org/10.1016/j.redox.2021.101867>
- Sies, H., 2020. Oxidative Stress: Concept and Some Practical Aspects. *Antioxidants* 9, 852. <https://doi.org/10.3390/antiox9090852>
- Sies, H., 2019. *Oxidative Stress*. Academic Press.
- Sies, H., 2017. Hydrogen peroxide as a central redox signaling molecule in physiological oxidative stress: Oxidative eustress. *Redox Biol* 11, 613–619. <https://doi.org/10.1016/j.redox.2016.12.035>
- Sies, H., 2015. Oxidative stress: a concept in redox biology and medicine. *Redox Biol* 4, 180–183. <https://doi.org/10.1016/j.redox.2015.01.002>
- Sies, H., 2007. Total Antioxidant Capacity: Appraisal of a Concept 1, 2. *J. Nutr.* 137, 1493–1495. <https://doi.org/10.1093/jn/137.6.1493>
- Sies, H., 1986. Biochemistry of Oxidative Stress. *Angew. Chem. Int. Ed. Engl.* 25, 1058–1071. <https://doi.org/10.1002/anie.198610581>
- SIES, H., 1985. Oxidative Stress 1–8. <https://doi.org/10.1016/b978-0-12-642760-8.50005-3>
- Sies, H., Berndt, C., Jones, D.P., 2016. Oxidative Stress. *Annu Rev Biochem* 86, 1–34. <https://doi.org/10.1146/annurev-biochem-061516-045037>
- Sies, H., Jones, D.P., 2020. Reactive oxygen species (ROS) as pleiotropic physiological signalling agents. *Nat Rev Mol Cell Bio* 21, 363–383. <https://doi.org/10.1038/s41580-020-0230-3>
- Sies, H., Menck, C.F.M., 1992. Singlet oxygen induced DNA damage. *Mutat. Res.DNAging* 275, 367–375. [https://doi.org/10.1016/0921-8734\(92\)90039-r](https://doi.org/10.1016/0921-8734(92)90039-r)
- Sies, H., Ursini, F., 2021. Homeostatic control of redox status and health. *Iubmb Life*. <https://doi.org/10.1002/iub.2519>
- Sinitcyn, P., Richards, A.L., Weatheritt, R.J., Brademan, D.R., Marx, H., Shishkova, E., Meyer, J.G., Hebert, A.S., Westphall, M.S., Blencowe, B.J., Cox, J., Coon, J.J., 2023. Global detection of human variants and isoforms by deep proteome sequencing. *Nat Biotechnol* 1–11. <https://doi.org/10.1038/s41587-023-01714-x>
- Slavov, N., 2022. Counting protein molecules for single-cell proteomics. *Cell* 185, 232–234. <https://doi.org/10.1016/j.cell.2021.12.013>
- Slavov, N., 2020. Unpicking the proteome in single cells. *Science* 367, 512–513. <https://doi.org/10.1126/science.aaz6695>
- Smith, L.M., Kelleher, N.L., 2018. Proteoforms as the next proteomics currency. *Science* 359, 1106–1107. <https://doi.org/10.1126/science.aat1884>
- Smith, L.M., Kelleher, N.L., Linial, M., Goodlett, D., Langridge-Smith, P., Goo, Y.A., Safford, G., Bonilla*, L., Kruppa, G., Zubarev, R., Rontree, J., Chamot-Rooke, J., Garavelli, J., Heck, A., Loo, J., Penque, D., Hornshaw, M., Hendrickson, C., Pasa-Tolic, L., Borchers, C., Chan, D., Young*, N., Agar, J., Masselon, C., Gross*, M., McLafferty, F., Tsybin, Y., Ge, Y., Sanders*, I., Langridge, J., Whitelegge*, J., Marshall, A., 2013. Proteoform: a single term describing protein complexity. *Nat Methods* 10, 186–187. <https://doi.org/10.1038/nmeth.2369>
- Smith, R.A.J., Porteous, C.M., Gane, A.M., Murphy, M.P., 2003. Delivery of bioactive molecules to mitochondria in vivo. *Proc. Natl. Acad. Sci.* 100, 5407–5412. <https://doi.org/10.1073/pnas.0931245100>
- Sousa, T., Gouveia, M., Travasso, R.D.M., Salvador, A., 2022. How abundant are superoxide and hydrogen peroxide in the vasculature lumen, how far can they reach? *Redox Biol* 58, 102527. <https://doi.org/10.1016/j.redox.2022.102527>
- Spirlandeli, A., Deminice, R., Jordao, A., 2013. Plasma Malondialdehyde as Biomarker of Lipid Peroxidation: Effects of Acute Exercise. *Int. J. Sports Med.* 35, 14–18. <https://doi.org/10.1055/s-0033-1345132>
- Stöcker, S., Laer, K.V., Mijuskovic, A., Dick, T.P., 2018. The Conundrum of Hydrogen Peroxide Signaling and the Emerging Role of Peroxiredoxins as Redox Relay Hubs. *Antioxid Redox Sign* 28, 558–573. <https://doi.org/10.1089/ars.2017.7162>
- Stolwijk, J.M., Falls-Hubert, K.C., Searby, C.C., Wagner, B.A., Buettner, G.R., 2020. Simultaneous detection of the enzyme activities of GPx1 and GPx4 guide optimization of selenium in cell biological experiments. *Redox Biol* 32, 101518. <https://doi.org/10.1016/j.redox.2020.101518>
- Stretton, C., Pugh, J.N., McDonagh, B., McArdle, A., Close, G.L., Jackson, M.J., 2020. 2-Cys peroxiredoxin oxidation in response to hydrogen peroxide and contractile activity in skeletal muscle: A novel insight into exercise-induced redox signalling? *Free Radical Bio Med* 160, 199–207. <https://doi.org/10.1016/j.freeradbiomed.2020.06.020>
- Switzer, C.H., Kasamatsu, S., Ihara, H., Eaton, P., 2023. SOD1 is an essential H₂S detoxifying enzyme. *Proc. Natl. Acad. Sci.* 120, e2205044120. <https://doi.org/10.1073/pnas.2205044120>

- Sykora, P., Witt, K.L., Revanna, P., Smith-Roe, S.L., Dismukes, J., Lloyd, D.G., Engelward, B.P., Sobol, R.W., 2018. Next generation high throughput DNA damage detection platform for genotoxic compound screening. *Sci. Rep.* 8, 2771. <https://doi.org/10.1038/s41598-018-20995-w>
- Taghizadeh, K., McFaline, J.L., Pang, B., Sullivan, M., Dong, M., Plummer, E., Dedon, P.C., 2008. Quantification of DNA damage products resulting from deamination, oxidation and reaction with products of lipid peroxidation by liquid chromatography isotope dilution tandem mass spectrometry. *Nat. Protoc.* 3, 1287–1298. <https://doi.org/10.1038/nprot.2008.119>
- Talwar, D., Miller, C.G., Grossmann, J., Szyrwiell, L., Schwecke, T., Demichev, V., Drazic, A.-M.M., Mayakonda, A., Lutsik, P., Veith, C., Milsom, M.D., Müller-Decker, K., Müllleder, M., Ralser, M., Dick, T.P., 2023. The GAPDH redox switch safeguards reductive capacity and enables survival of stressed tumour cells. *Nat Metabolism* 1–17. <https://doi.org/10.1038/s42255-023-00781-3>
- Taverne, Y.J., Merkus, D., Bogers, A.J., Halliwell, B., Duncker, D.J., Lyons, T.W., 2018. Reactive Oxygen Species: Radical Factors in the Evolution of Animal Life. *Bioessays* 40, 1700158. <https://doi.org/10.1002/bies.201700158>
- Timp, W., Timp, G., 2020. Beyond mass spectrometry, the next step in proteomics. *Sci Adv* 6, eaax8978. <https://doi.org/10.1126/sciadv.aax8978>
- Travasso, R.D.M., Aidos, F.S. dos, Bayani, A., Abranches, P., Salvador, A., 2017. Localized redox relays as a privileged mode of cytoplasmic hydrogen peroxide signaling. *Redox Biol* 12, 233–245. <https://doi.org/10.1016/j.redox.2017.01.003>
- Tsang, C.K., Liu, Y., Thomas, J., Zhang, Y., Zheng, X.F.S., 2014. Superoxide dismutase 1 acts as a nuclear transcription factor to regulate oxidative stress resistance. *Nat Commun* 5, 3446. <https://doi.org/10.1038/ncomms4446>
- TSOUKIAS, N.M., 2008. Nitric Oxide Bioavailability in the Microcirculation: Insights from Mathematical Models. *Microcirculation* 15, 813–834. <https://doi.org/10.1080/10739680802010070>
- Tuncay, A., Crabtree, D.R., Muggeridge, D.J., Husi, H., Cobley, J.N., 2023. Performance benchmarking microplate-immunoassays for quantifying target-specific cysteine oxidation reveals their potential for understanding redox-regulation and oxidative stress. *Free Radical Bio Med* 204, 252–265. <https://doi.org/10.1016/j.freeradbiomed.2023.05.006>
- Tuncay, A., Noble, A., Guille, M., Cobley, J.N., 2022. RedoxiFluor: A microplate technique to quantify target-specific protein thiol redox state in relative percentage and molar terms. *Free Radical Bio Med* 181, 118–129. <https://doi.org/10.1016/j.freeradbiomed.2022.01.023>
- Ursini, F., Maiorino, M., Forman, H.J., 2016. Redox homeostasis: The Golden Mean of healthy living. *Redox Biol* 8, 205–215. <https://doi.org/10.1016/j.redox.2016.01.010>
- Vasileiadou, O., Nastos, G.G., Chatzinikolaou, P.N., Papoutsis, D., Vrampa, D.I., Methenitis, S., Margaritelis, N.V., 2023. Redox Profile of Skeletal Muscles: Implications for Research Design and Interpretation. *Antioxidants* 12, 1738. <https://doi.org/10.3390/antiox12091738>
- Vásquez-Vivar, J., Kalyanaraman, B., Kennedy, M.C., 2000. Mitochondrial Aconitase Is a Source of Hydroxyl Radical AN ELECTRON SPIN RESONANCE INVESTIGATION*. *J Biol Chem* 275, 14064–14069. <https://doi.org/10.1074/jbc.275.19.14064>
- Viña, J., Gimeno, A., Sastre, J., Desco, C., Asensi, M., Pallardó, F.V., Cuesta, A., Ferrero, J.A., Terada, L.S., Repine, J.E., 2000. Mechanism of Free Radical Production in Exhaustive Exercise in Humans and Rats; Role of Xanthine Oxidase and Protection by Allopurinol. *IUBMB Life* 49, 539–544. <https://doi.org/10.1080/15216540050167098>
- Waeg, G., Dimsity, G., Esterbauer, H., 1996. Monoclonal Antibodies for Detection of 4-Hydroxynonenal Modified Proteins. *Free Radic. Res.* 25, 149–159. <https://doi.org/10.3109/10715769609149920>
- Wardman, P., 2007. Fluorescent and luminescent probes for measurement of oxidative and nitrosative species in cells and tissues: Progress, pitfalls, and prospects. *Free Radic. Biol. Med.* 43, 995–1022. <https://doi.org/10.1016/j.freeradbiomed.2007.06.026>
- Wayner, D.D.M., Burton, G.W., Ingold, K.U., Locke, S., 1985. Quantitative measurement of the total, peroxy radical-trapping antioxidant capability of human blood plasma by controlled peroxidation. *FEBS Lett.* 187, 33–37. [https://doi.org/10.1016/0014-5793\(85\)81208-4](https://doi.org/10.1016/0014-5793(85)81208-4)
- Weber, D., Davies, M.J., Grune, T., 2015. Determination of protein carbonyls in plasma, cell extracts, tissue homogenates, isolated proteins: Focus on sample preparation and derivatization conditions. *Redox Biol* 5, 367–380. <https://doi.org/10.1016/j.redox.2015.06.005>
- Weiss, S.J., King, G.W., LoBuglio, A.F., 1977. Evidence for hydroxyl radical generation by human Monocytes. *J. Clin. Investig.* 60, 370–373. <https://doi.org/10.1172/jci108785>
- Williamson, J., Hughes, C.M., Burke, G., Davison, G.W., 2020a. A combined γ -H2AX and 53BP1 approach to determine the DNA damage-repair response to exercise in hypoxia. *Free Radic. Biol. Med.* 154, 9–17. <https://doi.org/10.1016/j.freeradbiomed.2020.04.026>

- Williamson, J., Hughes, C.M., Cobley, J.N., Davison, G.W., 2020b. The mitochondria-targeted antioxidant MitoQ, attenuates exercise-induced mitochondrial DNA damage. *Redox Biol* 36, 101673. <https://doi.org/10.1016/j.redox.2020.101673>
- Winterbourn, C.C., 2008. Reconciling the chemistry and biology of reactive oxygen species. *Nat Chem Biol* 4, 278–286. <https://doi.org/10.1038/nchembio.85>
- Winterbourn, C.C., 1993. Superoxide as an intracellular radical sink. *Free Radical Bio Med* 14, 85–90. [https://doi.org/10.1016/0891-5849\(93\)90512-s](https://doi.org/10.1016/0891-5849(93)90512-s)
- Winterbourn, C.C., Hampton, M.B., 2008. Thiol chemistry and specificity in redox signaling. *Free Radical Bio Med* 45, 549–561. <https://doi.org/10.1016/j.freeradbiomed.2008.05.004>
- Winterbourn, C.C., Hampton, M.B., Livesey, J.H., Kettle, A.J., 2006. Modeling the Reactions of Superoxide and Myeloperoxidase in the Neutrophil Phagosome IMPLICATIONS FOR MICROBIAL KILLING *. *J. Biol. Chem.* 281, 39860–39869. <https://doi.org/10.1074/jbc.m605898200>
- Winterbourn, C.C., Metodiewa, D., 1994. The Reaction of Superoxide with Reduced Glutathione. *Arch Biochem Biophys* 314, 284–290. <https://doi.org/10.1006/abbi.1994.1444>
- Winterbourn, C.C., Peskin, A.V., Kleffmann, T., Radi, R., Pace, P.E., 2023. Carbon dioxide/bicarbonate is required for sensitive inactivation of mammalian glyceraldehyde-3-phosphate dehydrogenase by hydrogen peroxide. *Proc National Acad Sci* 120, e2221047120. <https://doi.org/10.1073/pnas.2221047120>
- Winterbourn, C.C., Peskin, A.V., Parsons-Mair, H.N., 2002. Thiol Oxidase Activity of Copper,Zinc Superoxide Dismutase*. *J Biol Chem* 277, 1906–1911. <https://doi.org/10.1074/jbc.m107256200>
- Wolff, S.P., 1994. Ferrous ion oxidation in presence of ferric ion indicator xylenol orange for measurement of hydroperoxides. *Methods Enzym.* 233, 182–189. [https://doi.org/10.1016/s0076-6879\(94\)33021-2](https://doi.org/10.1016/s0076-6879(94)33021-2)
- Wolkenhauer, O., 2014. Why model? *Front. Physiol.* 5, 21. <https://doi.org/10.3389/fphys.2014.00021>
- Woo, H.A., Yim, S.H., Shin, D.H., Kang, D., Yu, D.-Y., Rhee, S.G., 2010. Inactivation of Peroxiredoxin I by Phosphorylation Allows Localized H₂O₂ Accumulation for Cell Signaling. *Cell* 140, 517–528. <https://doi.org/10.1016/j.cell.2010.01.009>
- Wood, Z.A., Poole, L.B., Karplus, P.A., 2003. Peroxiredoxin Evolution and the Regulation of Hydrogen Peroxide Signaling. *Science* 300, 650–653. <https://doi.org/10.1126/science.1080405>
- Xiao, H., Jedrychowski, M.P., Schweppe, D.K., Huttlin, E.L., Yu, Q., Heppner, D.E., Li, J., Long, J., Mills, E.L., Szpyt, J., He, Z., Du, G., Garrity, R., Reddy, A., Vaites, L.P., Paulo, J.A., Zhang, T., Gray, N.S., Gygi, S.P., Chouchani, E.T., 2020. A Quantitative Tissue-Specific Landscape of Protein Redox Regulation during Aging. *Cell* 180, 968–983.e24. <https://doi.org/10.1016/j.cell.2020.02.012>
- Yamamoto, M., Kensler, T.W., Motohashi, H., 2018. The KEAP1-NRF2 System: a Thiol-Based Sensor-Effector Apparatus for Maintaining Redox Homeostasis. *Physiol Rev* 98, 1169–1203. <https://doi.org/10.1152/physrev.00023.2017>
- Yang, J., Carroll, K.S., Liebler, D.C., 2016. The Expanding Landscape of the Thiol Redox Proteome*. *Mol Cell Proteomics* 15, 1–11. <https://doi.org/10.1074/mcp.o115.056051>
- Yang, W.S., SriRamaratnam, R., Welsch, M.E., Shimada, K., Skouta, R., Viswanathan, V.S., Cheah, J.H., Clemons, P.A., Shamji, A.F., Clish, C.B., Brown, L.M., Girotti, A.W., Cornish, V.W., Schreiber, S.L., Stockwell, B.R., 2014. Regulation of Ferroptotic Cancer Cell Death by GPX4. *Cell* 156, 317–331. <https://doi.org/10.1016/j.cell.2013.12.010>
- Yin, H., Porter, N.A., 2003. Specificity of the ferrous oxidation of xylenol orange assay: analysis of autoxidation products of cholesteryl arachidonate. *Anal. Biochem.* 313, 319–326. [https://doi.org/10.1016/s0003-2697\(02\)00621-8](https://doi.org/10.1016/s0003-2697(02)00621-8)
- Yin, H., Xu, L., Porter, N.A., 2011. Free Radical Lipid Peroxidation: Mechanisms and Analysis. *Chem. Rev.* 111, 5944–5972. <https://doi.org/10.1021/cr200084z>
- Yu, L., Kang, X., Li, F., Mehrafrooz, B., Makhmreh, A., Fallahi, A., Foster, J.C., Aksimentiev, A., Chen, M., Wanunu, M., 2023. Unidirectional single-file transport of full-length proteins through a nanopore. *Nat Biotechnol* 1–10. <https://doi.org/10.1038/s41587-022-01598-3>
- Zapp, C., Obarska-Kosinska, A., Rennekamp, B., Kurth, M., Hudson, D.M., Mercadante, D., Barayeu, U., Dick, T.P., Denysenkov, V., Prisner, T., Bennati, M., Daday, C., Kappl, R., Gräter, F., 2020. Mechanoradicals in tensed tendon collagen as a source of oxidative stress. *Nat Commun* 11, 2315. <https://doi.org/10.1038/s41467-020-15567-4>
- Zatopek, K.M., Potapov, V., Maduzia, L.L., Alpaslan, E., Chen, L., Evans, T.C., Ong, J.L., Ettwiller, L.M., Gardner, A.F., 2019. RADAR-seq: A RARE DAMAGE and REPAIR sequencing method for detecting DNA damage on a genome-wide scale. *DNA Repair* 80, 36–44. <https://doi.org/10.1016/j.dnarep.2019.06.007>
- Zhang, H., Forman, H.J., 2017. 4-hydroxynonenal-mediated signaling and aging. *Free Radical Bio Med* 111, 219–225. <https://doi.org/10.1016/j.freeradbiomed.2016.11.032>
- Zhu, Y., Biernacka, A., Pardo, B., Dojer, N., Forey, R., Skrzypczak, M., Fongang, B., Nde, J., Yousefi, R., Pasero, P., Ginalski, K., Rowicka, M., 2019. qDSB-Seq is a general method for genome-wide quantification of DNA double-strand breaks using sequencing. *Nat. Commun.* 10, 2313. <https://doi.org/10.1038/s41467-019-10332-8>

- Zielonka, J., Kalyanaraman, B., 2018. Small-molecule luminescent probes for the detection of cellular oxidizing and nitrating species. *Free Radical Bio Med* 128, 3–22. <https://doi.org/10.1016/j.freeradbiomed.2018.03.032>
- Zielonka, J., Kalyanaraman, B., 2010. Hydroethidine- and MitoSOX-derived red fluorescence is not a reliable indicator of intracellular superoxide formation: Another inconvenient truth. *Free Radical Bio Med* 48, 983–1001. <https://doi.org/10.1016/j.freeradbiomed.2010.01.028>
- Zivanovic, J., Kouroussis, E., Kohl, J.B., Adhikari, B., Bursac, B., Schott-Roux, S., Petrovic, D., Miljkovic, J.Lj., Thomas-Lopez, D., Jung, Y., Miler, M., Mitchell, S., Milosevic, V., Gomes, J.E., Benhar, M., Gonzalez-Zorn, B., Ivanovic-Burmazovic, I., Torregrossa, R., Mitchell, J.R., Whiteman, M., Schwarz, G., Snyder, S.H., Paul, B.D., Carroll, K.S., Filipovic, M.R., 2019. Selective Persulfide Detection Reveals Evolutionarily Conserved Antiaging Effects of S-Sulfhydration. *Cell Metab* 30, 1152-1170.e13. <https://doi.org/10.1016/j.cmet.2019.10.007>

Disclaimer/Publisher's Note: The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.