

Zurich Open Repository and Archive

University of Zurich University Library Strickhofstrasse 39 CH-8057 Zurich www.zora.uzh.ch

Year: 2024

Belowground plant allocation regulates rice methane emissions from degraded peat soils

Sriskandarajah, Nijanthini ; Wüst-Galley, Chloé ; Heller, Sandra ; Leifeld, Jens ; Määttä, Tiia ; Ouyang, Zutao ; Runkle, Benjamin R K ; Schiedung, Marcus ; Schmidt, Michael W I ; Tumber-Dávila, Shersingh Joseph ; Malhotra, Avni

DOI: https://doi.org/10.1038/s41598-024-64616-1

Posted at the Zurich Open Repository and Archive, University of Zurich ZORA URL: https://doi.org/10.5167/uzh-260761 Journal Article Published Version



The following work is licensed under a Creative Commons: Attribution 4.0 International (CC BY 4.0) License.

Originally published at:

Sriskandarajah, Nijanthini; Wüst-Galley, Chloé; Heller, Sandra; Leifeld, Jens; Määttä, Tiia; Ouyang, Zutao; Runkle, Benjamin R K; Schiedung, Marcus; Schmidt, Michael W I; Tumber-Dávila, Shersingh Joseph; Malhotra, Avni (2024). Belowground plant allocation regulates rice methane emissions from degraded peat soils. Scientific Reports, 14(1):14593.

DOI: https://doi.org/10.1038/s41598-024-64616-1

scientific reports



OPEN

Belowground plant allocation regulates rice methane emissions from degraded peat soils

Nijanthini Sriskandarajah¹, Chloé Wüst-Galley², Sandra Heller², Jens Leifeld⁰², Tiia Määttä¹, Zutao Ouyang³, Benjamin R. K. Runkle¹, Marcus Schiedung¹, Michael W. I. Schmidt¹, Shersingh Joseph Tumber-Dávila^{7,8} & Avni Malhotra¹,

Carbon-rich peat soils have been drained and used extensively for agriculture throughout human history, leading to significant losses of their soil carbon. One solution for rewetting degraded peat is wet crop cultivation. Crops such as rice, which can grow in water-saturated conditions, could enable agricultural production to be maintained whilst reducing CO₂ and N₂O emissions from peat. However, wet rice cultivation can release considerable methane (CH4). Water table and soil management strategies may enhance rice yield and minimize CH4 emissions, but they also influence plant biomass allocation strategies. It remains unclear how water and soil management influences rice allocation strategies and how changing plant allocation and associated traits, particularly belowground, influence CH₄-related processes. We examined belowground biomass (BGB), aboveground biomass (AGB), belowground: aboveground ratio (BGB:ABG), and a range of root traits (root length, root diameter, root volume, root area, and specific root length) under different soil and water treatments; and evaluated plant trait linkages to CH4. Rice (Oryza sativa L.) was grown for six months in field mesocosms under high (saturated) or low water table treatments, and in either degraded peat soil or degraded peat covered with mineral soil. We found that BGB and BGB:AGB were lowest in water saturated conditions where mineral soil had been added to the peat, and highest in low-water table peat soils. Furthermore, CH₄ and BGB were positively related, with BGB explaining 60% of the variation in CH₄ but only under low water table conditions. Our results suggest that a mix of low water table and mineral soil addition could minimize belowground plant allocation in rice, which could further lower CH₄ likely because root-derived carbon is a key substrate for methanogenesis. Minimizing root allocation, in conjunction with water and soil management, could be explored as a strategy for lowering CH₄ emissions from wet rice cultivation in degraded peatlands.

Over the Holocene, peatlands have accumulated 30% of the world's soil organic carbon (SOC) while covering only 3% of the land area $^{1-3}$. However, peatlands have been extensively drained for agricultural uses 4 , leading to considerable CO_2 emissions 5 . One solution for rewetting drained peatlands while maintaining their agricultural utility, is wet crop cultivation, for example wet rice cultivation 6,7 . However, rice cultivation, currently accounting for ~20% of total agricultural CH_4 emissions globally $^{8-10}$, would lead to increased CH_4 emissions. Water table management and the addition of mineral soil are two strategies by which rice CH_4 emissions might be reduced 11 . For example, mid-season drainage for a period of typically 1–2 weeks is known to reduce CH_4 emissions, both during and following the drainage period, whilst maintaining yields 11 . The addition of mineral soils to peat soils is also a strategy increasingly used by farmers to ease the management of poorly-drained soils (e.g., to improve flood response or to be able to operate heavy machinery on the soils) 12 . Previous work has shown that lower water tables and mineral soil addition can decrease CH_4 emissions in rice grown on degraded peat soil 13 . However, it

¹Department of Geography, University of Zurich, 8057 Zurich, Switzerland. ²Climate and Agriculture Group, Agroscope, Zurich, Switzerland. ³College of Forestry, Wildlife and Environment, Auburn University, Auburn, AL 36849, USA. ⁴Biological and Agricultural Engineering, University of Arkansas, Fayetteville, AR 72701, USA. ⁵Department of Environmental Systems Science, ETH Zurich, 8092 Zurich, Switzerland. ⁶Thünen Institute of Climate-Smart Agriculture, Bundesallee 68, 38116 Braunschweig, Germany. ⁷Department of Environmental Studies, Dartmouth College, Hanover, NH 03755, USA. ⁸Harvard Forest, Harvard University, Petersham, MA 01366, USA. ⁹Biological Sciences Division, Pacific Northwest National Laboratory, Richland, WA 99852, USA. [∞]email: avni.malhotra@pnnl.gov

remains unclear how plant allocation strategies and properties (traits) respond to soil and water management¹⁴ and, in turn, how plant (especially root) traits influence CH_4^{15} .

Water and soil treatments could influence plant allocation and traits in a variety of ways that would influence CH₄. For example, water stress could increase plant carbon allocation to roots ^{14,16}. Soil treatments could influence root growth via changes in macro and micronutrients, soil pore structure, and soil water holding capacity. A reduction in plant-available macro or micronutrients could trigger increased below-ground allocation ^{17,18}. In turn, altered allocation belowground as well as changed root traits could have a range of confounding effects on net CH₄ flux¹⁹. First, roots act as conduits through which CH₄ is produced in the deeper layers of soil and can be transported to the atmosphere^{20,21}. Thus, an increase in root allocation could increase CH₄ transport and enhance the CH₄ flux. Second, through these same conduits, oxygen may be transported into the saturated soil layers where CH₄ oxidation may occur thereby reducing the net CH₄ flux^{20,22}. Third, root exudates may fuel heterotrophic microbes, leading to more CH₄ production^{23–25} or, in contrast, consuming more CH₄²⁶. It remains unclear which plant traits are influenced by water and soil management²⁷, and subsequently, how root traits and CH₄ fluxes are related¹⁹.

To address research gaps on plant and particularly root trait response to water and soil amendments, and downstream effects on CH_4 release from rice, we leveraged an existing mesocosm rice experiment in Switzerland. The experimental plots contain two water table conditions (low vs high) and two soil types (degraded peat, and degraded peat covered with mineral soil)¹³. Previous work from this experiment suggests that a lower water table and mineral soil cover can greatly decrease CH_4 emissions from paddy rice¹³, but the role of root traits and biomass allocation in driving this reduction remain unanswered. Thus, here, we address the following research questions: (1) How do water table and soil amendments influence rice below:aboveground allocation strategies and root traits (biomass, length, diameter, root tissue chemistry)? (2) How do plant allocation and trait changes influence CH4 fluxes?

First, we hypothesize that under a lower water table, rice plants will allocate a greater fraction of biomass belowground (relative to aboveground) to increase water uptake under water stressed conditions. Simultaneously, root traits related to water/nutrient uptake capacity such as specific root length (SRL) and root surface area, will also increase. Second, we hypothesize that the peat-only soil will generate more above and belowground biomass relative to the mineral-covered peat. We expect this increased productivity because, in this experiment, the degraded peat soil has more plant-available nutrients than peat with mineral cover soil. In mineral-covered peat, plants likely allocate more biomass belowground than aboveground compared to peat-only, to compensate for nitrogen limitation. Under this nutrient limitation, we also expect increased resource acquisition traits such as SRL and root surface area, in the mineral soil. Lastly, we hypothesize that plants with high belowground biomass and larger root diameter will emit more CH₄ due to both high substrate availability from root exudates and greater CH₄ transport through their thicker roots^{19,28}. Our study contributes to a better understanding of how water table and soil management can influence allocation strategies in rice and how these plant traits are related to CH₄ emissions.

Materials and methods Experimental site and design

The Agroscope rice mesocosm experiment (Figs. 1 and 2) is located in Zurich Affoltern, Switzerland (47° 25.8′ N 8° 31.2′ E with an elevation of 466 m.a.s.l.). The local climate is characterized by a mean annual temperature of 9.82 °C and a mean annual precipitation of 1026 mm (from 1990 to 2021)²⁹. This experiment was conducted between March and October 2021 (See Table S1 for dates and details of the experiment). Rice was grown in 24 experimental plots of 1.2 m × 1.2 m size and 1.4 m depth. The 24 plots had different soil (with or without a 30-cm mineral soil layer) and water treatments (targeting high at -6 cm or low at -20 cm) combinations (Fig. 1). Note that water table targets differed in achieved water tables due to the different subsidence of the different soils (detailed below). In the end, the four treatments were: peat and high water table, peat and low water table, peat plus mineral soil cover and high water table, and peat plus mineral soil cover and low water table (hereafter, high-peat, how-peat, high-mineral, low-mineral; Fig. 1).

Soil treatments

The plots (1.4 m deep) were filled with either just degraded peat (16 plots) or degraded peat with a 30 cm mineral soil layer on top (8 plots; Fig. 1). We added a 30 cm mineral layer because this is the typical layer thickness added in Switzerland, where this practice is carried out¹². This resulted in an addition of \sim 388 kg of mineral soil cover treatment (hereafter, 'mineral').

The degraded peat soil was taken from an 80 year old agricultural site in Affoltern am Albis (47° 16′ N, 8° 27′ E). This degraded peat soil had an organic carbon content ($C_{\rm org}$) of 27.4%, C:N ratio by mass of 19.9 and a soil pH of 6.0¹³ and was well mixed before adding to our plots, thus the vertical characteristics of a peat soil are not represented in the experiment. The mineral soil covering the degraded peat soil was taken from a farm in Rüthi, St. Gallen (47° 18′ N, 9° 32′ E) and was a loam containing 41.4% sand, 12.3% clay and 46.3% silt¹³. This soil material is calcareous and has the characteristics: $C_{\rm org}$ 0.6%, C:N ratio of 12.6 and a pH of 7.6¹³. Before application, the mineral soil was mixed with compost (10 kg [dry matter] per plot, pH 8.1, $C_{\rm org}$ 20.5%, C:N = 13.2), a measure also used by Swiss farmers¹². Total C and N content were analyzed with a CHNS–O elemental analyser EuroEA3000 (HEKAtech, Germany). It is worth noting that pH in the degraded peat plus mineral soil is higher than that of a typical unmanaged peatland, where pH is usually lower than 6³0. Thus our experimental mineral soils likely have much higher decomposition rates than a typical unmanaged peatland.



Figure 1. A schematic of the four types of treatments (high-peat, low-peat, high-mineral, low-mineral) evaluated in this study. Experimental plots had high (saturated) or low water treatments and degraded peat-only or degraded peat covered with ~ 30 cm mineral soil treatments. The average growing season water table height and sample sizes are also shown (note the unbalanced design).

Water table treatments

Across the 24 plots, 12 had a 'low' and 12 had a 'high' water table (Fig. 1) but due to differential subsidence, the water table was also dependent on soil treatment. The 'high' water table plots had a growing season water table depth of -6 and -7 cm (below the surface), without and with mineral cover, respectively¹³. The 'low' had growing season water table depths of -11 cm with mineral cover; and -17 cm without mineral cover (Fig. 2). With minor exceptions (see below), the water levels were maintained at these depths throughout the vegetation season and unwanted variations were adjusted (for example after heavy rains; Figure S1 explains the experimental infrastructure). In the high water table, mid-season drainage was applied between 4 and 12th August, where the water level was lowered to -100 cm. Additionally, the water level was lowered (to ca. -10 cm) the day seedlings were planted and hours prior to fertilization. The water level of all plots was reduced to -100 cm the week prior to harvesting. The water level was calculated based on data from water table loggers, adjusted for soil subsidence (Figure S1). The soil subsidence was measured once every month during the vegetation period.

The rationale for the two water table depths is as follows: the high water table roughly corresponds to the -5 cm optimum water table depth for minimizing greenhouse gas emissions³¹. The low water table corresponds to a depth at which conventional management (in the Swiss water management context) can take place and therefore at which farmers might be able to cultivate rice without having to adjust management practices too much.

As a result of the water and soil treatments, resulting growing season average volumetric water contents varied and were as follows for the different treatments: high-peat = $0.65~\rm m^3~m^{-3}$ (excluding the mid-season drainage), low-peat = $0.63~\rm m^3~m^{-3}$, low-mineral = $0.42~\rm m^3~m^{-3}$ high-mineral = no moisture data from these plots (as no CH₄ was measured). Averages are from half-hourly soil moisture measurements from mid-July to October, using Teros-11* (METER Group) soil sensors at 5 cm depth¹³.

Rice cultivation

The experiment used the rice variety, 'Loto' (*Oryza sativa* L.), typically grown as paddy rice in the cool temperate moist climate of the central plateau of Switzerland 32,33 . Rice seeds were sown in seed trays in commercially-available sowing compost. The seed trays were placed for 4 weeks in climate chambers and then for 1 week into a greenhouse. On the 26th May 2021, at the three-leaf stage, the seedlings were transplanted to the experimental site (Fig. 2). Each plot was planted with 34 plants¹³, resulting in a density of 24 plants m⁻². The rice plants were fertilized as seedlings with Wuxal(R) (Syngenta Agro AG), an NPK mineral fertilizer with micronutrients (K, B, Cu, Fe, Mn, Mo, Zn), and after planting out (three times) with an ammonium nitrate fertilizer (NH₄NO₃ with Mg and S), P₂O₅ and K₂O mineral fertilizers (Table S1)¹³. The fertilizer amounts correspond to common greenhouse practice and the Swiss fertilizer recommendations³⁴.

Plant sampling and processing

To characterize root traits, we collected one random rice plant per plot after harvest (18 Oct. 2021). Thus, a total of 24 plants were sampled, i.e. $8 \times$ peat with high water table, $8 \times$ peat with low water table, $4 \times$ mineral cover with high water table, and $4 \times$ mineral cover with low water table (Fig. 1). The rice plant was separated into above-and belowground components. The aboveground part, which included leaves, stem, and rice, was dried at 60 °C for 3–4 days and weighed. The belowground part, including roots and soil, was collected, and immediately



Figure 2. Image of the Agroscope rice experiment, with 1.2×1.2 m growth plots showing grass and rice plots (Taken on October 18, 2021).

brought to the lab. The soil sample including the roots was circa 16 cm $\log \times 16$ cm wide $\times 18$ cm deep. In the lab, the soil was washed using distilled water in an ultrasonic bath for 2–8 h to remove the soil from the roots by ultrasound induced cavitation forces. These partially clean root systems were then stored frozen at – 20 °C to prevent decomposition of the roots until further processing.

Laboratory analysis

Each root system was thawed for 24 h in the fridge for further cleaning and processing. Once thawed, following standard methods³⁵, roots were washed again but this time more thoroughly and until entirely free of soil. This was done using magnifying glasses, distilled water, forceps, and paint brushes to clean the remaining soil from the root surface. After cleaning, the roots were scanned in a scanning tray (29.7 cm \times 42.0 cm) using a Canon Image Runner Advance C5535i with the integrated scanner in grayscale mode at 600 DPI (dots per inch). The scans were then saved as a TIFF-file for further image analysis. After the scanning, the wet weight of the roots was measured, and the roots were dried at 60° for 1–2 days. Subsequently, the roots were weighed again to determine their dry weight. The sampled soils from which the roots were processed had slightly different volumes. Thus, the dry root biomass was normalized to a soil volume of $15 \times 15 \times 15$ cm (3375 cm³) by dividing root biomass weight with the collected soil volume and multiplying by 3375 cm³.

Image processing root scans to quantify root traits

The root scans were analyzed using Rhizovision Explorer v2.0.3³⁶, an open-source software developed for root image processing. For each of the 24 root systems, we were able to obtain the following root traits from Rhizovision: total root length, total root surface area, total root volume, root average diameter and root length for diameter bins from 1 to 6 mm (see Figure S2 and the supplementary section on "Root trait quantification using Rhizovision software"). We also calculated specific root length (SRL) as the ratio of the total root length to BGB, which provides an indication of root length investment per unit mass and is expected to increase when a plant is resource stressed^{37,38}.

Methane and ancillary data

Methane fluxes and aboveground biomass (hereafter, AGB) were measured in the high-peat, low-peat and low-mineral treatments in a previous study¹³. These treatment names correspond to the following treatment names from the previous study¹³: RH, RM, and RM + min, respectively. Briefly, the CH₄ fluxes were measured twice a week during the growing season, using a manual dark static chamber attached to a gas analyzer (cavity ring-down spectrometer; model G2308, Picarro Inc., Santa Clara, CA, USA). The chamber enclosure time for each individual plot measurement was 15 min. The first CH₄ flux measurements were conducted six days prior to transplanting the rice seedlings and continued until the time of harvesting the last rice plants. Additional gas measurements were carried out immediately prior to, and following, fertilization and changes in the water table (e.g. mid-season drainage). For the present study, the average value of the 44 total measurements that were taken for each plot was used to represent CH₄ emissions. In our analysis, CH₄ fluxes were measured from 12 of the 24 root plots ($4 \times \text{high-peat}$, $4 \times \text{low-peat}$, and $4 \times \text{low-mineral plots}$). Following the micrometeorological sign convention, positive CH₄ fluxes in this study are referred to as CH₄ emissions to the atmosphere (source), and negative fluxes as CH₄ uptake by the soil (sink).

Statistical analysis

We used the following variables in our analyses: AGB, BGB, total biomass (AGB+BGB), the ratio of belowground and aboveground biomass (BGB:AGB) and root traits (total root length, total root averaged diameter, total root surface area, and total root volume) from image analysis. We also used growing season average CH₄ fluxes from a previous study¹³. Ancillary variables included mean and standard error of water table depth, and soil carbon, nitrogen, oxygen and carbon:nitrogen ratio¹³. All statistical analyses and data visualizations were performed using JMP*, Version 15.2.0 (SAS Institute Inc., Cary, NC, 1989–2021).

Due to our plant trait data being unbalanced, non-normal and small in sample size, we used a non-parametric Kruskal–Wallis test to evaluate the treatment effects on plant traits and additionally compared treatments using the Steel–Dwass method for pairwise comparisons. The goal of these statistical tests was to compare BGB, AGB, total biomass, and BGB:AGB among the different soil and water treatments (hypothesis I and II). For our third hypothesis, we first used multiple linear regression to establish soil, water and plant trait predictors of CH_4 fluxes. We then also used bivariate linear regressions to further investigate relationships between CH_4 and key plant-trait predictors. We log-transformed CH_4 data to meet the assumption of normality for linear regressions. Lastly, to explore the trait covariation and the relationship between CH_4 and traits, we also used principal components analyses (PCA).

Results

Water table and soil type influence rice biomass

The water table and soil treatments significantly affected rice BGB and AGB and supported our hypothesis that a lower water table increases plant BGB allocation (Fig. 3b). However, our hypothesis that nutrient-poor mineral soils will have a higher belowground allocation than peat soils was not supported. We observed considerable differences in AGB and total biomass between water table treatments in peat soils, and in BGB and BGB:AGB between the high-mineral and low-peat treatments (Fig. 3; Table S2). Low water tables in peat soil approximately halved both above and total biomass relative to high water tables. With lower water tables, AGB decreased from 43 to 19 g (median values in Fig. 3c) and total biomass (AGB+BGB) from 51 to 28 g (Fig. 3a). Meanwhile, BGB halved in the high-mineral, compared to the low-peat treatment (7.5 to 3.6 g shift in median values; Fig. 3d), leading to a 4×reduction in BGB:ABG allocation (from 0.4 to 0.1; Fig. 3b).

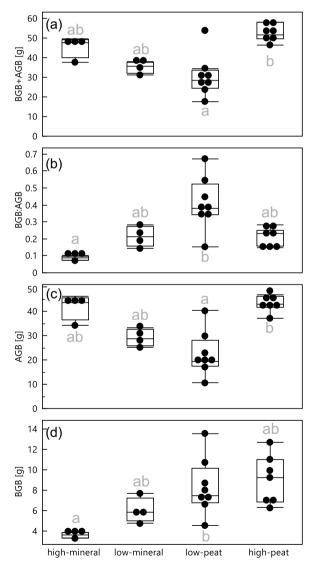


Figure 3. The response of above (AGB) and belowground (BGB) biomass, BGB:AGB and total biomass (AGB+BGB) to high or low water table and peat or peat with mineral soil treatments (referred to as mineral in the figure). For each panel, treatments that do not share a letter code denote pairwise significant differences.

Treatment response of other root traits

Root traits except BGB and BGB:AGB showed very little variation between peat and mineral soil types (Table S3). Across water treatments, no significant difference was seen in the root traits either, but some interesting trends were present (Fig. 4). Notably, roots were slightly but insignificantly longer per unit dry mass (SRL, Fig. 4) under water-stressed conditions. Although root traits varied considerably, the low water table had higher medians and interquartile ranges for all root traits except diameter (Fig. 4; Table S3). We also noted that the low-peat treatment had a much higher total root length when compared with other treatments and especially when compared to the high-mineral treatment (Table S3). Though not significant at p < 0.05, it is worth noting that high-mineral treatment had roughly half the total root length of the low-peat treatment (Kruskal–Wallis test chi square = 6.2, p = 0.0980; Steel–Dwass pairwise comparison Z = 2.17 p = 0.13). While root traits did not show significant treatment effects, we still report trait values and trait covariation as these are important baseline values for rice plants (Table S3).

Root biomass and CH₄ emission positively correlated in low water table

Net flux average values of CH_4 ranged from 2.4 g CH_4 m⁻² y⁻¹ in the low water table treatments to 6.4 g CH_4 m⁻² y⁻¹ in the high water treatments¹³. A multiple regression model that included BGB, soil treatment, water table treatment and the interaction between BGB and water table treatment explained 78% of the variation in CH_4 , where only an interaction term between BGB and water table level was significant (model output reported in Table 1; See Figure S4 for non log transformed CH_4 values).

Given this interaction between BGB and low water tables, we analyzed water treatment individually. We found that in the low water table treatments, BGB and BGB:AGB were significantly positively related to CH_4 flux (Fig. 5). In the high water table plots, we did not have enough data (n = 4) to establish a relationship between CH_4 flux and BGB (Figure S4).

Discussion

In this study, we evaluated the response of plant biomass allocation and root traits to different soil and moisture conditions in an experimental rice ($Oryza\ sativa\ L$.) system and investigated root trait linkages to CH_4 flux. We found that plants allocated the least belowground biomass in water-saturated and mineral-covered peat soils and

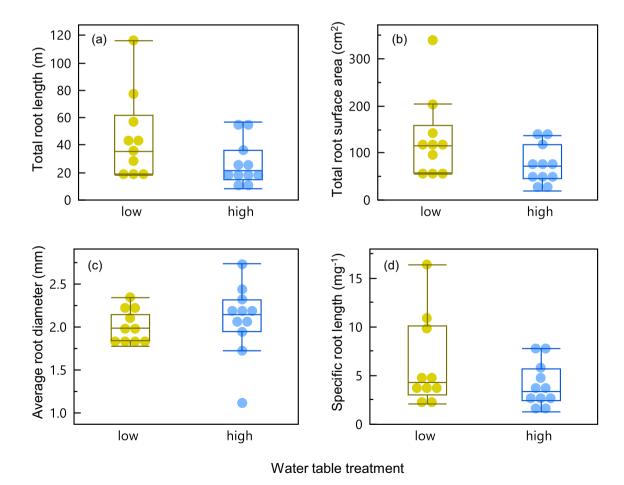


Figure 4. Root traits plotted against water tables. Specific root length (SRL) was computed as root length divided by belowground biomass (BGB). None of the root traits showed significant differences between water or soil treatment.

Model term	Estimate	Std error	t ratio	p value
Intercept	9.02	0.64	14.12	0.0000
BGB	-0.04	0.08	-0.59	0.5756
Soil treatment [mineral]	-0.19	0.19	-1.01	0.3532
Water table treatment [low]	-0.33	0.19	-1.71	0.139
(BGB – 7.9)*Water table treatment [low]	0.25	0.08	3.25	0.0174

Table 1. Best fit model (after removing other root traits and interactions between traits and soil treatment) of CH_4 emission (log mg CH_4 m⁻² growing season⁻¹). Model R^2 = 0.78, p value = 0.0375, n = 12. Note that the model was run on log transformed values of CH_4 flux to resolve issues of non normality.

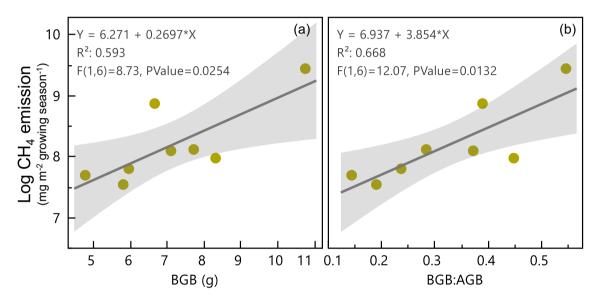


Figure 5. Belowground biomass, (BGB) and the belowground: aboveground (BGB:AGB) as predictors of CH_4 flux. Only the low-peat and low-mineral treatments are considered in this analysis.

the most in water-stressed peat soils. Among our measured plant traits, BGB and BGB:AGB allocation had the strongest link to $\mathrm{CH_4}$ flux but only in low water table conditions. Thus, our results reveal that rice response to different soil–water treatments differs between above and below ground plant organs and that the below ground trait response is a predictor of $\mathrm{CH_4}$ dynamics.

More total biomass with higher water table levels but variable belowground allocation

In our study, as expected for rice, higher water table levels in general increased total plant biomass^{9,39} and lower water tables increased plant allocation to roots⁴⁰. Water-stressed plants in the low water treatments are expected to allocate more BGB to increase their water uptake capacity^{16,41-44}. We also saw some support for this in the other root traits wherein roots from low water table conditions had higher SRL and length allocation (Table S3). We observed a decrease in AGB with lower water tables, which could be due to water limitation for the plant, given the high water demand of rice plants^{45,46}. While the AGB response to water availability is often observed in rice, our study provides evidence that BGB:AGB also responds to the water table changes as seen in other ecosystems and in intact peatlands^{47,48}. A further explanation for lower AGB under low water tables could be due to plant stress from temperature variations⁴⁹. Under high water tables, temperature variations (e.g., low night-time temperatures) would be buffered by the water but low water tables would likely see stronger variations in soil temperatures that could negatively influence plant growth^{50,51}.

Decreased BGB with mineral soil additions

We hypothesized that the mineral soil treatments would have greater BGB than the peat soil due to low nutrients in the mineral soil (Figure S3 and Table S4). We observed the opposite wherein the mineral soil treatment had the lowest BGB and belowground allocation (Fig. 3b and d). One reason for this could have been that the plants in the mineral soil treatment were overall nutrient limited. However, AGB and total biomass data do not support this reason because AGB is not low in mineral soil treatments; rather it is lowest in the peat-only soils (Fig. 3c).

We did not have detailed post-experiment data on macro and micro nutrients to fully assess the role of nutrient limitation in our results. However, we have post-experiment soil carbon (C) and nitrogen content (N), and C:N that suggests that N availability was much higher in the peat soil; Figure S3). Even though C:N has been shown to be a sensitive parameter for peatland degradation⁵², our C:N data could reflect nutrient use that

occurred during the experiment, rather than nutrient status and availability itself. We also have nutrient data from pre-experiment soils, where we find some support for different nutrient availability in the mineral and peat soils (Table S4). Notably, cation exchange capacity was almost 20 times higher in the degraded peat than in the mineral soil suggesting potentially higher nutrient retention in the peat (Table S4). Calcium, magnesium and sodium were also higher in the peat than in the mineral soil. However, potassium was higher in mineral soils compared to peat soils. Potassium is generally considered more important for plant growth than our other reported nutrients and may explain why we do not see high root allocation in the mineral soil treatment. Furthermore, mineral soil mixed with peat has been shown to increase phosphate, potassium, iron and magnesium availability and higher N retention of south retention effects could therefore have influenced the availability of nutrients derived from the fertilization and compost that was added to our mineral soil treatment. Furthermore, the lack of high BGB in the mineral mixture may be because the mineral soil contained more available micronutrients compared to mineral free peat. For example, rice has a high demand of silicon and its limitation influences the overall nutrient uptake and plant biomass so. Thus, detailed nutrient data from after the experiment and nutrient addition experiments would be needed to fully assess the role of nutrients in our observed plant growth response to soil and water treatments.

Ultimately, both water and soil treatments influenced rice plant biomass and allocation strategy, often with opposite effects below and aboveground. Belowground biomass and allocation were halved when mineral soils were added to water-saturated peat. Meanwhile, AGB and total biomass was halved when peat water tables were lowered. These two results suggest that in these rice plants, AGB was driven by water availability while belowground allocation was driven by soil properties. Similar results have been found in studies evaluating AGB and drought response 45 and root response to soil nutrients 60 but our study illustrates rice allocation responses to moisture and nutrient conditions in one experimental setting.

Rice root trait covariation

While traits other than biomass did not have statistically significant responses to the water table treatments, we did see trends of root traits related to increased resource-acquisition strategies responding to water-stressed conditions (e.g. increased SRL; Table S3). We observed slightly higher SRL in the lower water table treatment than in the high water table providing some evidence of increased soil exploration by roots⁶¹. Root length followed similar trends to BGB (Table S3), but other traits showed no significant responses to water and soil treatments. Some of the lack of responses are also interesting to note. For example, root median diameter was consistently around 2 mm across treatments though the interquartile range was highest in water-saturated mineral soil roots where length and biomass were lowest (Table S3). This suggests a possible trade-off between belowground biomass and allocation to root diameter across our captured trait variation (Figure S5; principal component analysis of all root traits across all treatments). The lack of a response in root diameter to the water treatments is particularly interesting since other studies have reported decreases in root diameter with increases in SRL as a response to drier soil conditions^{27,62,63}, but the direction of the response seems to differ between species and growth forms⁶⁴, and rice genotypes⁶⁵. In order to untangle the variation in root diameter and SRL in different water table levels and soil conditions in future studies, it may be worth including measurements of root stele and cortex fractions to fully evaluate nutrient uptake, and water transport and absorption capacity^{35,66}.

Another trait trade off emerged between aboveground biomass and belowground allocation for soil exploration. AGB is negatively correlated with root traits such as total root surface area, total root volume, and total root length (Figure S5). Interestingly, this tradeoff disappears in a PCA containing only the high water table trait data. Conversely, this above:belowground tradeoff is pronounced in the low water table trait data (Figure S6), suggesting that, as expected, water stressed conditions may exacerbate plant allocation tradeoffs between aboveground carbon fixation and belowground water (or nutrient) uptake.

Our study provides belowground data from rice plants including trait covariation among rice root traits (Table S3). Even though the root traits show no strong significant responses to treatments, these are valuable data to report given that rice root data are sparse⁶⁷. Lastly, it is also possible that other traits such as root system architecture and maximum rooting depth responded to treatments but were not captured by our methods. Nevertheless, our data add to the limited data on rice root trait covariation and support the notion that rice roots adapt quickly (within a growing season) to water/nutrient stress conditions.

Relationships between CH4 flux and root traits under low water table conditions

So far, a limited number of studies have investigated the effect of root traits on $\mathrm{CH_4}$ emissions. and little is known about the interactions between rice root traits, methanogens and methanotrophs, and $\mathrm{CH_4}$ emissions. We found some evidence for rice BGB and BGB:AGB predicting $\mathrm{CH_4}$ emissions, at least in water-stressed conditions (low water table). Our results supported our hypothesis that BGB and $\mathrm{CH_4}$ flux are positively correlated, likely related to increased BGB facilitating $\mathrm{CH_4}$ transport through plants and more rhizodeposition potentially increasing substrates for methanogens as well as plant-mediated $\mathrm{CH_4}$ transport 10,68 . We did not observe the contrary effect of more roots oxygenating the rhizosphere and leading to increased $\mathrm{CH_4}$ oxidation and reduced net $\mathrm{CH_4}$ fluxes as seen in paddy soils²² and salt marsh ecosystems⁶⁹. There may have been a slight increase in $\mathrm{CH_4}$ oxidation with increasing root biomass in our study but it may have been overshadowed by increased substrate provision from rhizodeposits and even from a priming effect of root exudates on $\mathrm{CH_4}$ production⁷⁰, leading to the overall increased $\mathrm{CH_4}$ emissions. These specific hypotheses remain to be further tested using controlled laboratory incubations.

The lack of a strong relationship between CH_4 and root traits other than BGB could be due to the use of proxies instead of more direct measurements of CH_4 -related root traits and processes. For example, we assumed root diameter to be a proxy for aerenchyma volume and, therefore, plant-mediated CH_4 transport from the soil to the

atmosphere¹⁹. However, we found no relationship between root diameter and CH₄ emissions. One reason could be that we did not measure the distinct and opposing processes of CH₄ production and consumption, rather we measured the net flux. Similar to the previous discussion on root biomass, increased diameters could result in the confounding effects of increased CH₄ transport or rhizosphere oxidation⁷¹ via increased soil reduction–oxidation potential^{72,73}, and, ultimately, aerobic CH₄ consumption. Depending on the local soil conditions and microbial composition and abundance, CH₄ can be oxidized before reaching the root⁷⁴ or within the root itself⁷⁵, resulting in a decrease in CH₄ emission. Therefore, it is possible that a lack of relationship between root diameter and net CH₄ emissions, especially in low water table levels in our study may be because of the confounding processes of methanogenesis (increasing due to increased BGB and root exudation) and methanotrophy (increasing due to large diameter) which cannot be reliably separated based on only net CH₄ flux. Therefore, we recommend including measurements of rhizosphere oxidation, such as O₂ concentration from planar optode technology⁷⁶ and redox and O₂ electrodes^{77,78}, in future studies investigating the relationships between BGB and CH₄ emission. In addition, since root diameter and surface area were used as proxies for CH₄ transport and root exudation in our study, respectively, it is possible that other more direct measurements of these processes, such as root porosity²² and root exudation⁷⁰, could have been better predictors of CH₄ flux. BGB is also a general proxy for CH₄ production, consumption and transport and thus overall our weak CH₄-trait relationships could further indicate that we did not measure root traits potentially more relevant for CH₄¹⁹. Nevertheless, since root trait-CH₄ connections have rarely been investigated, this study is one of the first ones to test the use of different proxies for root-mediated CH₄ processes in peat soil, and despite the non-significant relationships, these results should motivate researchers into investigating additional CH₄-relevant root traits.

Another reason for the lack of relationship between root traits and CH_4 flux could be that the changes in soil properties (i.e. mineral vs. peat) and water table level could have overridden the effects of individual root traits on the net CH_4 flux. It has been shown in multiple wetland studies that changes in water table and other abiotic variables can have a stronger effect on net soil CH_4 flux than vegetation 79,80 . However, these relationships have not been adequately investigated in combination with belowground plant traits. Thus, the significant interaction term in the best fit model (Table 1) including low water table and BGB could indicate that root influence on the net CH_4 flux becomes more relevant only when water table level is decreased, further possibly confirming the overriding effect of the water table. The overriding effect of water table on trait- CH_4 relationships is also supported when we visualize trait- CH_4 relationships in the low water table data (PCA in Figure S7). This exploratory analysis suggests that in the tradeoff between plant aboveground and belowground allocation under water stressed conditions (Figure S6a), CH_4 emissions align with the belowground allocation traits (Figure S7), i.e. if water stressed rice plant allocate more belowground, this could mean an increase in CH_4 emissions. Future studies should therefore experimentally test the tradeoffs among rice aboveground allocation, belowground allocation and CH_4 emissions in these systems.

Implications for Swiss peatlands

Switzerland contains about 28,000 ha of peatland⁸¹ which represents 1% of the total country area and 2% of the agricultural land¹². Many of Switzerland's largest degraded peatlands occur in flat valley bottoms where horticultural and staple crop production dominates. In addition to their high greenhouse gas emissions, these surfaces also require drainage systems to be renewed for the continued cultivation of dry crops, a very costly measure. A wet crop system, such as rice, might provide an alternative agricultural use of these degraded peatland systems. A growing network of rice cultivators has been established in the last years³² and increasing temperatures in Switzerland and the associated lengthening of the growing season would further favor rice cultivation. While it is already established that lower water tables and the addition of mineral soils can reduce rice CH₄ emissions¹³, our results suggest that even under low moisture conditions, further managing or genetically modifying root traits to lower root biomass, albeit with caution^{82,83}, could potentially lower CH₄ emissions. Given our small number of samples and experimental system, this notion would require extensive additional testing.

Conclusions

We investigated the effects of soil treatment (peat vs., mineral-covered peat) and water level (high vs low) on rarely-studied root traits (biomass, allocation, diameter, length, volume, and surface area) in rice and established previously-untested links among root traits and $\mathrm{CH_4}$ emission. We found a positive relationship between root biomass and $\mathrm{CH_4}$ emissions in low water conditions, and the lowest belowground allocation in mineral soil-covered peat soils; thus providing preliminary insights into the potential of minimizing root biomass as a $\mathrm{CH_4}$ reducing strategy in these systems. This work opens new research directions to understand whether optimizing (and minimizing) BGB and BGB:AGB could be a viable tool for lowering rice $\mathrm{CH_4}$ emissions from rewetted peatlands.

The results of this study should be understood and concluded with discretion for a few reasons. First, as already discussed above, additional soil nutrient information and measurement of more direct traits would aid in interpreting these results. Secondly, the experiment was short term, representing only one rice species, and using growing season aggregates of both trait and CH_4 flux data. A longer-term study with time-resolved measurements would allow for better delineation of root trait effects on CH_4 .

Nevertheless, this study provided first insights into how soil type and water affect rice biomass allocation and their link to $\mathrm{CH_4}$ emission in degraded peatland soils. Degraded peatlands are a widespread ecosystem where rewetting could have high returns for the global carbon cycle-climate feedback^{5,84}. In the case of wet rice cultivation, high climate benefits may be accompanied by high economic returns and the continued use of this land for agricultural production, two important socio-economic factors for farmers when determining the management of land.

Data availability

All data used in this study are provided as supplementary materials (Supplementary Information 2). Previously published raw methane data are also available at https://data.mendeley.com/datasets/fxmnty8zf8/1 (Wüst, Chloé; Heller, Sandra; Ammann, Christof; Paul, Sonja; Doetterl, Sebastian; Leifeld, Jens (2023), "CH4 and N2O flux data from wet rice grown on organic soil in Switzerland", Mendeley Data, V1, https://doi.org/10.17632/fxmnty8zf8.1). Additional details can also be found in Heller, Sandra. 2021. "Wet rice on organic soils". *Master Thesis, ETH Zürich*.

Received: 27 November 2023; Accepted: 11 June 2024

Published online: 25 June 2024

References

- 1. Gorham, E. Northern peatlands: Role in the carbon cycle and probable responses to climatic warming. *Ecol. Appl.* 1, 182–195 (1991).
- 2. Yu, Z., Loisel, J., Brosseau, D. P., Beilman, D. W. & Hunt, S. J. Global peatland dynamics since the last glacial maximum. *Geophys. Res. Lett.* 37 (2010).
- 3. Joosten, H., Tapio-Biström, M.-L. & Tol, S. Peatlands: Guidance for Climate Change Mitigation through Conservation, Rehabilitation and Sustainable Use (Food and Agriculture Organization of the United Nations Rome, 2012).
- 4. Fluet-Chouinard, E. et al. Extensive global wetland loss over the past three centuries. Nature 614, 281-286 (2023).
- Leifeld, J. & Menichetti, L. The underappreciated potential of peatlands in global climate change mitigation strategies. Nat. Commun. 9, 1071 (2018).
- Zhang, H., Xue, Y., Wang, Z., Yang, J. & Zhang, J. An alternate wetting and moderate soil drying regime improves root and shoot growth in rice. Crop Sci. 49, 2246–2260 (2009).
- 7. Joosten, H. & Clarke, D. Wise use of mires and peatlands. Int. Mire Conserv. Group Int. Peat Soc. 304 (2002).
- 8. van Groenigen, K. J., van Kessel, C. & Hungate, B. A. Increased greenhouse-gas intensity of rice production under future atmospheric conditions. *Nat. Clim. Change* 3, 288–291. https://doi.org/10.1038/nclimate1712 (2013).
- Oliver, V. et al. Effects of water management and cultivar on carbon dynamics, plant productivity and biomass allocation in European rice systems. Sci. Total Environ. 685, 1139–1151 (2019).
- 10. Huang, Y., Sass, R. & Fisher, F. Methane emission from Texas rice paddy soils. 2. Seasonal contribution of rice biomass production to CH₄ emission. *Glob. Change Biol.* **3**, 491–500 (1997).
- Liu, X. et al. Effect of mid-season drainage on CH₄ and N₂O emission and grain yield in rice ecosystem: A meta-analysis. Agric. Water Manage. 213, 1028–1035 (2019).
- 12. Ferré, M. et al. Sustainable management of cultivated peatlands in Switzerland: Insights, challenges, and opportunities. Land Use Policy. 87, 104019. https://doi.org/10.1016/j.landusepol.2019.05.038 (2019).
- 13. Wüst-Galley, C. et al. Methane and nitrous oxide emissions from rice grown on organic soils in the temperate zone. Agric. Ecosyst. Environ. 356, 108641 (2023).
- 14. Kim, Y. et al. Root response to drought stress in rice (Oryza sativa L.). Int. J. Mol. Sci. 21, 1513 (2020).
- Kim, W. J., Bui, L. T., Chun, J.-B., McClung, A. M. & Barnaby, J. Y. Correlation between methane (CH₄) emissions and root aerenchyma of rice varieties. *Plant Breed. Biotechnol.* 6, 381–390 (2018).
- 16. Kano, M., Inukai, Y., Kitano, H. & Yamauchi, A. Root plasticity as the key root trait for adaptation to various intensities of drought stress in rice. *Plant Soil* 342, 117–128 (2011).
- 17. Hinzke, T. et al. Potentially peat-forming biomass of fen sedges increases with increasing nutrient levels. Funct. Ecol. 35, 1579–1595 (2021).
- 18. Klotzbücher, T., Marxen, A., Jahn, R. & Vetterlein, D. Silicon cycle in rice paddy fields: insights provided by relations between silicon forms in topsoils and plant silicon uptake. *Nutr. Cycl. Agroecosyst.* 105, 157–168 (2016).
- 19. Määttä, T. & Malhotra, A. The hidden roots of wetland methane emissions. Glob. Change Biol. 30, e17127 (2024).
- 20. Aulakh, M. S., Wassmann, R., Rennenberg, H. & Fink, A. S. Pattern and amount of aerenchyma relate to variable methane transport capacity of different rice cultivars. *Plant Biol.* 2, 182–194 (2000).
- 21. Wassmann, R. & Aulakh, M. S. The role of rice plants in regulating mechanisms of methane missions. *Biol. Fertil. Soils* 31, 20–29 (2000).
- 22. Jiang, Y. et al. Higher yields and lower methane emissions with new rice cultivars. Glob. Chang. Biol. 23, 4728-4738 (2017).
- 23. Aulakh, M. S., Wassmann, R., Bueno, C. & Rennenberg, H. Impact of root exudates of different cultivars and plant development stages of rice (*Oryza sativa* L.) on methane production in a paddy soil. *Plant Soil* 230, 77–86 (2001).
- 24. Lu, Y., Wassmann, R., Neue, H.-U. & Huang, C. Dynamics of dissolved organic carbon and methane emissions in a flooded rice soil. Soil Sci. Soc. Am. J. 64, 2011–2017 (2000).
- Theint, E. E., Suzuki, S., Ono, E. & Bellingrath-Kimura, S. D. Influence of different rates of gypsum application on methane emission from saline soil related with rice growth and rhizosphere exudation. Catena 133, 467–473 (2015).
- 26. Chen, Y. et al. Rice root morphological and physiological traits interaction with rhizosphere soil and its effect on methane emissions in paddy fields. Soil Biol. Biochem. 129, 191–200 (2019).
- 27. Comas, L. H., Becker, S. R., Cruz, V. M. V., Byrne, P. F. & Dierig, D. A. Root traits contributing to plant productivity under drought. Front. Plant Sci. 4, 442 (2013).
- 28. Bhullar, G. S., Iravani, M., Edwards, P. J. & Olde Venterink, H. Methane transport and emissions from soil as affected by water table and vascular plants. *BMC Ecol.* **13**, 32 (2013).
- 29. MeteoSchweiz. https://www.meteoschweiz.admin.ch/.
- Vitt, D. H. & Chee, W.-L. The relationships of vegetation to surface water chemistry and peat chemistry in fens of Alberta, Canada. Vegetatio 89, 87–106 (1990).
- 31. Freeman, B. W. J. *et al.* Responsible agriculture must adapt to the wetland character of mid-latitude peatlands. *Glob. Change Biol.* **28**, 3795–3811 (2022).
- 32. Gramlich, A., Fabian, Y. & Jacot, K. Faktenblatt Reisanbau-Ökologischer Nassreis-Anbau auf vernässenden Ackerflächen in der Schweiz (Agridea).
- 33. Eggleston, H. S., Buendia, L., Miwa, K., Ngara, T. & Tanabe, K. 2006 IPCC Guidelines for National Greenhouse Gas Inventories. https://www.osti.gov/etdeweb/biblio/20880391 (2006).
- 34. Richner, W., Sinaj, S. & Carlen, C. GRUD 2017: Grundlagen für die Düngung landwirtschaftlicher Kulturen in der Schweiz (Agroscope, 2017).
- 35. Freschet, G. T. et al. A starting guide to root ecology: Strengthening ecological concepts and standardising root classification, sampling, processing and trait measurements. New Phytol. 232, 973–1122 (2021).
- 36. Seethepalli, A. et al. RhizoVision explorer: Open-source software for root image analysis and measurement standardization. AoB Plants 13, lab056 (2021).

- 37. Lucob-Agustin, N. et al. Morpho-physiological and molecular mechanisms of phenotypic root plasticity for rice adaptation to water stress conditions. Breed. Sci. 71, 20–29 (2021).
- 38. Kadam, N. N., Yin, X., Bindraban, P. S., Struik, P. C. & Jagadish, K. S. V. Does morphological and anatomical plasticity during the vegetative stage make wheat more tolerant of water deficit stress than rice?. *Plant Physiol.* **167**, 1389–1401 (2015).
- 39. Thakur, A. K., Mohanty, R. K., Patil, D. U. & Kumar, A. Impact of water management on yield and water productivity with system of rice intensification (SRI) and conventional transplanting system in rice. *Paddy Water Environ.* 12, 413–424 (2014).
- Xu, W. et al. Drought stress condition increases root to shoot ratio via alteration of carbohydrate partitioning and enzymatic activity in rice seedlings. Acta Physiol. Plant 37, 9. https://doi.org/10.1007/s11738-014-1760-0 (2015).
- 41. Hodge, A., Berta, G., Doussan, C., Merchan, F. & Crespi, M. Plant root growth, architecture and function. *Plant Soil* 321, 153–187 (2009).
- 42. Davies, W. J. Root growth response and functioning as an adaptation in water limiting soils.
- 43. Biondini, M. Allometric scaling laws for water uptake by plant roots. J. Theor. Biol. 251, 35-59 (2008).
- 44. Ma, Q. et al. Aboveground and belowground biomass relationships in the Zoige peatland, eastern Qinghai-Tibetan plateau. Wetlands 37, 461–469 (2017).
- 45. Hussain, N., Ahmed, M., Duangpan, S., Hussain, T. & Taweekun, J. Potential impacts of water stress on rice biomass composition and feedstock availability for bioenergy production. https://doi.org/10.3390/su131810449
- Cabangon, R. J. et al. Effect of irrigation method and N-fertilizer management on rice yield, water productivity and nutrient-use
 efficiencies in typical lowland rice conditions in China. Paddy Water Environ. 2, 195–206 (2004).
- 47. Malhotra, A. et al. Peatland warming strongly increases fine-root growth. Proc. Natl. Acad. Sci. U. S. A. 117, 17627-17634 (2020).
- 48. Bardgett, R. D. & van der Putten, W. H. Belowground biodiversity and ecosystem functioning. *Nature* **515**, 505–511 (2014).
- Zhang, Q., Chen, Q., Wang, S., Hong, Y. & Wang, Z. Rice and cold stress: methods for its evaluation and summary of cold tolerancerelated quantitative trait loci. Rice 7, 24 (2014).
- 50. Lakshmi, V., Jackson, T. J. & Zehrfuhs, D. Soil moisture–temperature relationships: Results from two field experiments. *Hydrol. Process.* 17, 3041–3057 (2003).
- 51. Hatfield, J. L. & Prueger, J. H. Temperature extremes: Effect on plant growth and development. Weather Clim. Extremes 10, 4–10 (2015).
- Leifeld, J., Klein, K. & Wüst-Galley, C. Soil organic matter stoichiometry as indicator for peatland degradation. Sci. Rep. 10, 7634 (2020).
- 53. Sardans, J. & Peñuelas, J. Potassium control of plant functions: ecological and agricultural implications. Plants 10, 419 (2021).
- Linquist, B. A., Campbell, J. C. & Southard, R. J. Assessment of potassium soil balances and availability in high yielding rice systems. Nutr. Cycl. Agroecosyst. 122, 255–271 (2022).
- 55. De Datta, S. K. & Mikkelsen, D. S. Potassium nutrition of rice. In *Potassium in Agriculture* 665–699 (American Society of Agronomy, Crop Science Society of America, Soil Science Society of America, 2015).
- 56. Hytönen, J. *et al.* Long term effects of mineral soil addition on the nutrient amounts of peat and on the nutrient status of Scots pine on drained mires. *Suo* **59**, 9–26 (2008).
- 57. Wang, Y., Paul, S. M., Alewell, C. & Leifeld, J. Reduced nitrogen losses from drained temperate agricultural peatland after mineral soil coverage. *Biol. Fertil. Soils* 59, 153–165 (2023).
- 58. Schaller, J. et al. Silicon as a potential limiting factor for phosphorus availability in paddy soils. Sci. Rep. 12, 16329 (2022).
- 59. Ma, J. F. et al. A silicon transporter in rice. Nature 440, 688-691 (2006).
- 60. Xin, W. et al. Adaptation mechanism of roots to low and high nitrogen revealed by proteomic analysis. Rice 14, 5 (2021).
- 61. Ostonen, I. et al. Specific root length as an indicator of environmental change. Plant Biosyst. Int. J. Deal. All Asp. Plant Biol. 141, 426–442 (2007).
- 62. Reid, C. P. P. & Bowen, G. D. Effects of soil moisture on v/a mycorrhiza formation and root development in Medicago. In *The Soil-Root Interface* (eds Harley, J. L. & Russell, R. S.) 211–219 (Academic Press, 1979).
- 63. Bauerle, T. L., Smart, D. R., Bauerle, W. L., Stockert, C. & Eissenstat, D. M. Root foraging in response to heterogeneous soil moisture in two grapevines that differ in potential growth rate. *New Phytol.* 179, 857–866 (2008).
- 64. Larson, J. E. & Funk, J. L. Seedling root responses to soil moisture and the identification of a belowground trait spectrum across three growth forms. *New Phytol.* 210, 827–838 (2016).
- 65. Suralta, R. R. & Yamauchi, A. Root growth, aerenchyma development, and oxygen transport in rice genotypes subjected to drought and waterlogging. *Environ. Exp. Bot.* **64**, 75–82 (2008).
- 66. Henry, A., Cal, A. J., Batoto, T. C., Torres, R. O. & Serraj, R. Root attributes affecting water uptake of rice (*Oryza sativa*) under drought. *J. Exp. Bot.* 63, 4751–4763 (2012).
- 67. Parida, S., Dash, G. K., Samal, K. C., Das, S. & Swain, P. Evaluation of SRL and different root contributing traits of rice (*Oryza sativa* L.) under water stress. *Biol. Forum Int. J.* 13, 810–815 (2021).
- Meijide, A., Gruening, C., Goded, I., Seufert, G. & Cescatti, A. Water management reduces greenhouse gas emissions in a Mediterranean rice paddy field. Agric. Ecosyst. Environ. 238, 168–178 (2017).
- 69. Noyce, G. L., Smith, A. J., Kirwan, M. L., Rich, R. L. & Megonigal, J. P. Oxygen priming induced by elevated CO₂ reduces carbon accumulation and methane emissions in coastal wetlands. *Nat. Geosci.* 16, 63–68 (2023).
- 70. Waldo, N. B., Hunt, B. K., Fadely, E. C., Moran, J. J. & Neumann, R. B. Plant root exudates increase methane emissions through direct and indirect pathways. *Biogeochemistry* 145, 213–234 (2019).
- Armstrong, W., Justin, S. H. F., Beckett, P. M. & Lythe, S. Root adaptation to soil waterlogging. Aquat. Bot. 39, 57–73. https://doi. org/10.1016/0304-3770(91)90022-w (1991).
- 72. Flessa, H. & Fischer, W. R. Plant-induced changes in the redox potentials of rice rhizospheres. *Plant Soil.* 143, 55–60. https://doi.org/10.1007/bf00009128 (1992).
- Faulwetter, J. L. et al. Microbial processes influencing performance of treatment wetlands: A review. Ecol. Eng. 35, 987–1004. https://doi.org/10.1016/j.ecoleng.2008.12.030 (2009).
- 74. Vroom, R. J. E., van den Berg, M., Pangala, S. R., van der Scheer, O. E. & Sorrell, B. K. Physiological processes affecting methane transport by wetland vegetation—A review. *Aquat. Bot.* **182**, 103547 (2022).
- 75. Gerard, G. & Chanton, J. Quantification of methane oxidation in the rhizosphere of emergent aquatic macrophytes: defining upper limits. *Biogeochemistry* 23, 79–97. https://doi.org/10.1007/bf00000444 (1993).
- Larsen, M., Borisov, S. M., Grunwald, B., Klimant, I. & Glud, R. N. A simple and inexpensive high resolution color ratiometric planar optode imaging approach: application to oxygen and pH sensing. *Limnol. Oceanogr. Methods* 9, 348–360. https://doi.org/ 10.4319/lom.2011.9.348 (2011).
- 77. Fritz, C. et al. Zero methane emission bogs: Extreme rhizosphere oxygenation by cushion plants in Patagonia. New Phytol. 190, 398–408 (2011).
- 78. Visser, E. J. W., Bögemann, G. M., van de Steeg, H. M., Pierik, R. & Blom, C. W. P. M. Flooding tolerance of Carex species in relation to field distribution and aerenchyma formation. *New Phytol.* **148**, 93–103 (2000).
- 79. Turetsky, M. R. *et al.* A synthesis of methane emissions from 71 northern, temperate, and subtropical wetlands. *Glob. Change Biol.* **20**, 2183–2197 (2014).
- 80. Moore, T. R. et al. A multi-year record of methane flux at the Mer bleue bog, southern Canada. Ecosystems 14, 646-657 (2011).

- 81. Agroscope. A model-based carbon inventory for Switzerland's mineral agricultural soils using RothC. https://doi.org/10.34776/as105e
- 82. Kabouw, P., van Dam, N. M., van der Putten, W. H. & Biere, A. How genetic modification of roots affects rhizosphere processes and plant performance. *J. Exp. Bot.* **63**, 3475–3483 (2012).
- 83. Jiménez, J. D. L. C. & Pedersen, O. Mitigation of greenhouse gas emissions from rice via manipulation of key root traits. *Rice* 16, 1–10 (2023)
- 84. Leifeld, J., Wüst-Galley, C. & Page, S. Intact and managed peatland soils as a source and sink of GHGs from 1850 to 2100. *Nat. Clim. Change* 9, 945–947 (2019).

Acknowledgements

We would like to thank University of Zurich's Soils and Biogeochemistry group for their experimental and lab infrastructure, notably Thomy Keller and Yves Brügger. We acknowledge funding from the University of Zurich Stiftung für Wissenschaftliche Forschung (STWF-22-028) and the Swiss National Science Foundation (project 200021_215214) awarded to AM. AM was also supported by a Laboratory Directed Research and Development Program at the Pacific Northwest National Laboratory. BRKR acknowledges support of the U.S. National Science Foundation under CAREER program award 1752083.

Author contributions

AM, CW-G and NS designed the study with input from BRKR, MWIS and ZO. CW-G and JL designed and maintained the experimental plots. CW-G and SH provided background data. NS collected the samples and conducted the laboratory work. NS and AM conducted the data analysis and interpretation with help from ST-D, TM and MS. NS wrote the first draft with help from AM, and all authors edited and commented on the manuscript. AM supervised the project.

Competing interests

The authors declare no competing interests.

Additional information

Supplementary Information The online version contains supplementary material available at https://doi.org/10.1038/s41598-024-64616-1.

Correspondence and requests for materials should be addressed to A.M.

Reprints and permissions information is available at www.nature.com/reprints.

Publisher's note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by/4.0/.

This is a U.S. Government work and not under copyright protection in the US; foreign copyright protection may apply 2024