

Heart failure pharmacotherapy and cancer: pathways and pre-clinical/clinical evidence

Nabil V. Sayour ^{1,2,3}, Ágnes M. Paál¹, Pietro Ameri^{4,5}, Wouter C. Meijers⁶, Giorgio Minotti ⁷, Ioanna Andreadou⁸, Antonella Lombardo^{9,10}, Massimiliano Camilli ^{9,10}, Heinz Drexel¹¹, Erik Lerkevang Grove^{12,13}, Gheorghe Andrei Dan^{14,15}, Andreea Ivanescu^{14,15}, Anne Grete Semb¹⁶, Gianluigi Savarese^{17,18}, Dobromir Dobrev ^{19,20,21}, Filippo Crea^{9,10}, Juan-Carlos Kaski ²², Rudolf A. de Boer ⁶, Péter Ferdinandy^{1,23,24†}, and Zoltán V. Varga ^{1,2,3*†}

¹Department of Pharmacology and Pharmacotherapy, Semmelweis University, H-1085 Budapest, Üllői út 26, Hungary; ²HCEMM-SU Cardiometabolic Immunology Research Group, H-1089 Budapest, Nagyvárud tér 4, Hungary; ³MTA-SE Momentum Cardio-Oncology and Cardioimmunology Research Group, H-1089 Budapest, Nagyvárud tér 4, Hungary; ⁴Cardiovascular Disease Unit, IRCCS Ospedale Policlinico San Martino, Italian IRCCS Cardiology Network, Genova, Italy; ⁵Department of Internal Medicine, University of Genova, Genova, Italy; ⁶Department of Cardiology, Thorax Center, Erasmus University Medical Center, Rotterdam, The Netherlands; ⁷University Campus Bio-Medico, Via Álvaro del Portillo, 21, 00128 Rome, Italy; ⁸Laboratory of Pharmacology, School of Pharmacy, National and Kapodistrian University of Athens, Athens, Greece; ⁹Department of Cardiovascular and Pulmonary Sciences, Catholic University of the Sacred Heart, Rome, Italy; ¹⁰Department of Cardiovascular Medicine, Fondazione Policlinico Universitario A. Gemelli IRCCS, Rome, Italy; ¹¹Vorarlberg Institute for Vascular Investigation & Treatment (VIVIT), Carinagasse 47, A-6800 Feldkirch, Austria; ¹²Department of Cardiology, Aarhus University Hospital, Aarhus, Denmark; ¹³Department of Clinical Medicine, Faculty of Health, Aarhus University, Aarhus, Denmark; ¹⁴Carol Davila University of Medicine and Pharmacy, Colentina University Hospital, Bucharest, Romania; ¹⁵Cardiology Department, Colentina Clinical Hospital, Bucharest, Romania; ¹⁶Division of Research and Innovation, REMEDY-Centre for Treatment of Rheumatic and Musculoskeletal Diseases, Diakonhjemmet Hospital, Oslo, Norway; ¹⁷Division of Cardiology, Department of Medicine, Karolinska Institutet, Stockholm, Sweden; ¹⁸Heart and Vascular and Neuro Theme, Karolinska University Hospital, Stockholm, Sweden; ¹⁹Institute of Pharmacology, West German Heart and Vascular Center, University Duisburg-Essen, Essen, Germany; ²⁰Department of Medicine and Research Center, Montreal Heart Institute and Université de Montréal, Montréal, QC, Canada; ²¹Department of Integrative Physiology, Baylor College of Medicine, Houston, TX, USA; ²²Molecular and Clinical Sciences Research Institute, St. George's University of London, London, United Kingdom; ²³Pharmahungary Group, Szeged, Hungary; and ²⁴MTA-SE System Pharmacology Research Group, Department of Pharmacology and Pharmacotherapy, Semmelweis University, Budapest, Hungary

Received 28 July 2023; revised 8 January 2024; accepted 7 February 2024; online publish-ahead-of-print 5 March 2024

Abstract

Heart failure (HF) patients have a significantly higher risk of new-onset cancer and cancer-associated mortality, compared to subjects free of HF. While both the prevention and treatment of new-onset HF in patients with cancer have been investigated extensively, less is known about the prevention and treatment of new-onset cancer in patients with HF, and whether and how guideline-directed medical therapy (GDMT) for HF should be modified when cancer is diagnosed in HF patients. The purpose of this review is to elaborate and discuss the effects of pillar HF pharmacotherapies, as well as digoxin and diuretics on cancer, and to identify areas for further research and novel therapeutic strategies. To this end, in this review, (i) proposed effects and mechanisms of action of guideline-directed HF drugs on cancer derived from pre-clinical data will be described, (ii) the evidence from both observational studies and randomized controlled trials on the effects of guideline-directed medical therapy on cancer incidence and cancer-related outcomes, as synthesized by meta-analyses will be reviewed, and (iii) considerations for future pre-clinical and clinical investigations will be provided.

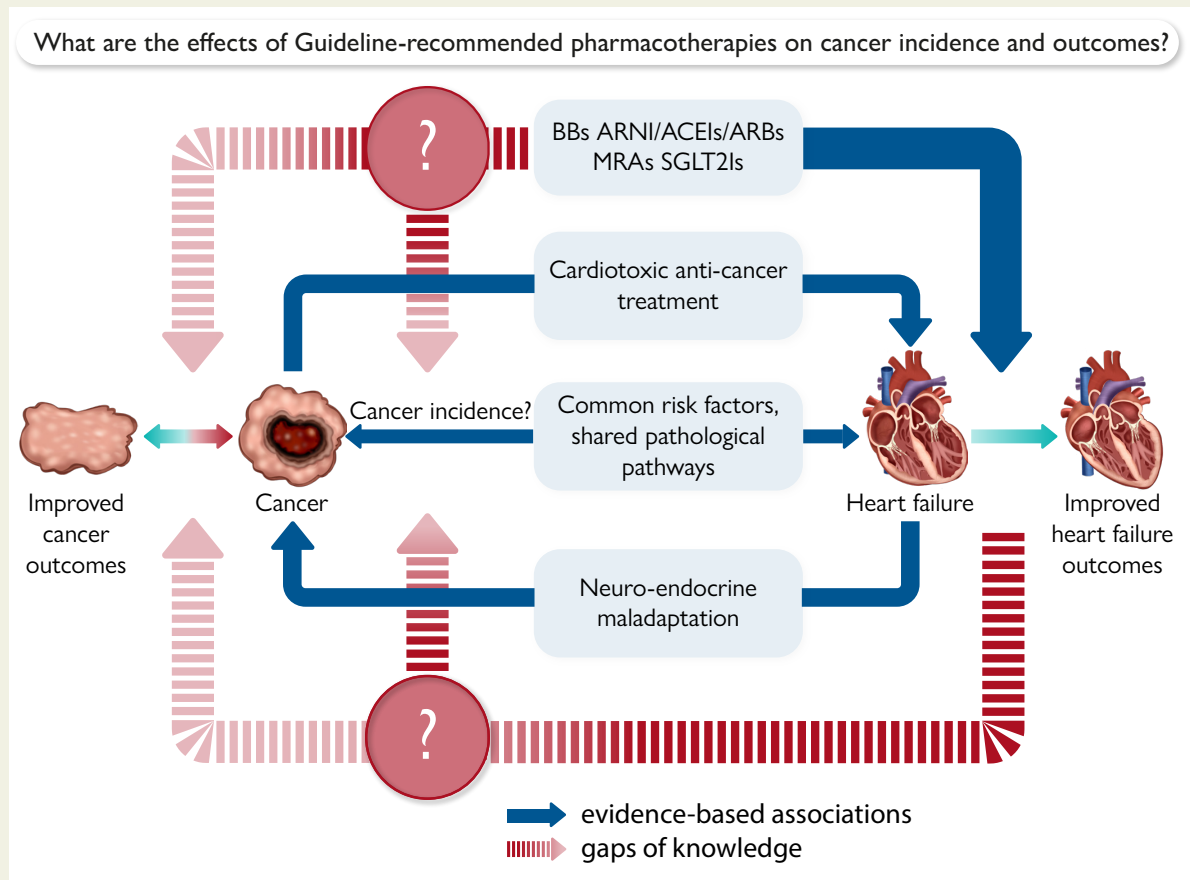
* Corresponding author. Email: varga.zoltan@med.semmelweis-univ.hu

† These authors contributed equally to this work.

© The Author(s) 2024. Published by Oxford University Press on behalf of the European Society of Cardiology.

This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<https://creativecommons.org/licenses/by/4.0/>), which permits unrestricted reuse, distribution, and reproduction in any medium, provided the original work is properly cited.

Graphical Abstract



The risk factors and pathophysiological pathways of both heart failure and cancer are common. The discipline of cardio-oncology investigates how heart failure and cancer progression are connected: on one hand, the cardiac effects of anti-cancer medications or cancer-derived metabolic byproducts are investigated, whereas on the other hand, the possible effects of heart failure on cancer progression are examined, such as those mediated by maladaptive neuroendocrine activation and factors secreted from the failing heart. Nevertheless, there is a lack of systematic knowledge on how heart failure pharmacotherapies affect new-onset cancer incidence or prevalent cancer outcomes, and whether these effects are mediated through the improvement in cardiac functions. ACEI, angiotensin-converting enzyme inhibitor; ARB, angiotensin receptor blocker; BB, beta-blocker; MRA, mineralocorticoid receptor antagonist; SGLT2I, sodium-glucose cotransporter 2 inhibitor.

Keywords

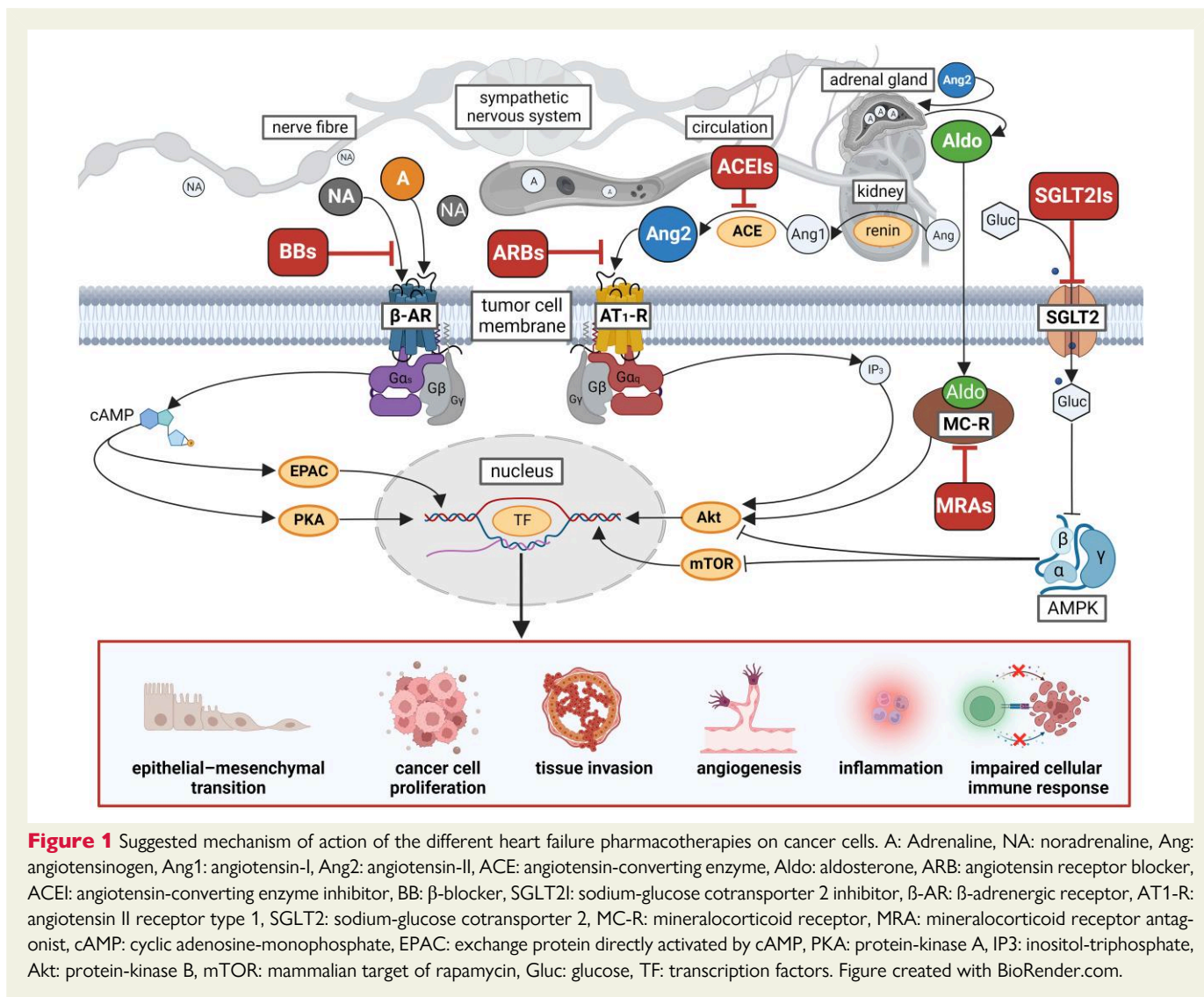
Cardio-oncology • Heart failure • Cancer • Beta-blocker • Mineralocorticoid Receptor antagonist • Sodium-glucose cotransporter 2 inhibitor • Angiotensin receptor blocker • Angiotensin-converting enzyme inhibitor • Angiotensin receptor-neprilysin inhibitor

Introduction

Heart failure (HF) and cancer are leading causes of mortality worldwide.^{1–6} Although HF and cancer are conventionally viewed as two separate disease entities, an implicit bidirectional relationship between them has been identified by recent studies,⁷ as (i) major risk factors and mechanistic pathways overlap in HF and cancer,^{8–11} (ii) in patients with prevalent cancer, cardiovascular diseases are the leading causes of non-cancer mortality,^{12–14} (iii) several cancer pharmacotherapies exert cardiotoxic effects,^{15–18} and (iv) in patients with prevalent HF, the majority of observational evidence reported increased cancer incidence and worse cancer outcomes compared with subjects free of HF, irrespective of patients' age, HF etiology, and cancer type.^{19–25} However, an epidemiological study on men with self-reported HF reported no such associations,²⁶ moreover, a Danish nationwide study reported a significant decrease

in cancer incidence in patients with prevalent HF after adjusting for multiple variables including co-morbidities and medications.²⁷ Interestingly, despite major improvements in HF therapies, cancer incidence in HF patients has remained unchanged for the past 20 years, underscoring the importance of cancer in the setting of HF.^{28,29}

Guideline recommendations exist regarding prevention, screening, monitoring, and treatment of new-onset HF in patients receiving cancer therapies.³ However, no recommendations are available that define if and how HF treatment should be modified (i) to prevent cancer incidence in HF patients, or (ii) when cancer is diagnosed during the course of HF, and no ongoing clinical studies are available addressing these questions. Indeed, based on a systematic search on ClinicalTrials.gov, we found that all of the ongoing clinical studies of cardio-oncology are related to prevention or treatment of cancer therapy-related cardiotoxicity (see [Supplementary data online, Table S1](#)).



The purpose of this review is to provide (i) an overview of the effects and proposed mechanisms of action of GDMT of HF on cancer based on pre-clinical data, and (ii) a balanced interpretation of findings reported in clinical meta-analyses investigating the effects of HF GDMT on cancer incidence and outcomes. Moreover, gaps in knowledge and areas of future pre-clinical and clinical research will also be highlighted.

In this narrative review, we collected evidence from *in vivo* (see [Supplementary data online, Table S2](#)) and *in vitro* (see [Supplementary data online, Table S3](#)) pre-clinical studies, with a special emphasis on drug-type, dosing, cancer type, and endpoints. We also collected information from meta-analyses of clinical studies investigating the effect of HF GDMT on cancer incidence in cancer-free patients, or other cancer-related outcomes (e.g. cancer-specific or recurrence-free survival) in patients with pre-existing cancer at baseline (see [Supplementary data online, Table S4](#)).

Effects of beta-blockers on cancer

Beta-adrenoceptor signalling has been suggested to play a contributory role in cancer biology, as it modulates cancer progression mainly via the activation of protein kinase A and the exchange protein activated by

adenylyl cyclase ([Figure 1](#)).³⁰ Catecholamines regulate beta-adrenoceptors on cancer cells, stromal cells, and tumour-associated macrophages,³¹ resulting in a procarcinogenic microenvironment. Accordingly, beta-blockers (BBs) may have a potential to decrease cancer incidence or improve cancer outcomes.

Pre-clinical studies assessing the effects of beta-blockers on cancer

Effects of BBs on cancer have been extensively investigated in pre-clinical studies, almost unanimously demonstrating potent anti-cancer effects both *in vivo* (see [Supplementary data online, Table S2](#)) and *in vitro* (see [Supplementary data online, Table S3](#)). Most *in vivo* studies tested the non-selective BB propranolol. Propranolol exerted significant anti-cancer effects *in vivo* by inhibiting tumour growth,^{32–35} reducing metastases,^{36,37} influencing tumour immuno-microenvironment,³⁸ and by repressing angiogenesis.³⁹ In contrast, some studies showed that propranolol did not have anti-cancer effects *per se*,⁴⁰ as it could only enhance the effects of other anti-cancer therapies.^{38,41–44} Among *in vivo* studies investigating the anti-cancer effects of non-selective BBs other than propranolol, carvedilol was the most

commonly used agent. Most of these studies demonstrated significant anti-cancer effects of carvedilol when used alone in a variety of cancer types.^{45–52} Other BBs, such as the beta1-selective metoprolol⁴¹ and nebivolol,^{53,54} as well as the non-selective labetalol³³ were also shown to have either anti-cancer effects *per se*, or enhance the anti-cancer effects of other drugs.

Anti-cancer effects of various BBs on several cancer types have been assessed by *in vitro* studies, resulting in variable results. For instance, anti-cancer efficacy of propranolol or beta2-adrenoceptor blockade was reported to be higher compared to beta1-selective BBs by the majority of studies.^{34,55–58} Conversely, a study on non-small cell lung cancer cell lines demonstrated no correlation between beta-adrenoceptor selectivity and anti-cancer efficacy of BBs, as both propranolol and the beta1-selective betaxolol significantly decreased colony formation.⁵⁹ Of note, metoprolol, the only beta1-selective BB approved for HF that was investigated in this study, was ineffective in such settings. Moreover, the superiority of beta1-selective BBs over propranolol has also been demonstrated *in vitro*,^{60–62} further complicating the picture.

Of note, BB use emerged not only in prevention of cancer development *per se*, but also in prevention of cancer therapy-related cardiotoxicity, as beta-adrenoceptor signalling was shown to share an intricate conundrum with human epidermal growth factor receptor type 2 (ERBB2) in the cardiovascular system, and also in breast cancer.^{63,64} As a result, the BB carvedilol was shown to prevent ERBB2-blockade-induced cardiotoxicity.⁶⁵

Clinical studies assessing the effects of beta-blockers on cancer

Intriguingly, contrary to pre-clinical studies, meta-analyses on clinical observational studies or randomized controlled trials (RCTs) show disparate results regarding effects of BBs on cancer, both in cancer-free patients, and in cancer patients (Figure 2, see [Supplementary data online, Table S4](#)).

Effects of beta-blockers on risk of cancer in cancer-free patients

In a meta-analysis on nine RCTs, BB use was associated with a non-significant trend towards lower overall risk of any cancer type.⁶⁶ Likewise, a network meta-analysis on 70 RCTs showed that the use of BBs was not associated with any change in risk of any cancer type, or cancer mortality.⁶⁷ In other meta-analyses, BB use was associated with mixed effects on cancer incidence, varied by cancer types. For instance, two meta-analyses showed no association between BB use and risk of new-onset breast, lung, colon, or prostate cancer.^{68,69} Other meta-analyses showed that BB use was associated with a significantly increased risk of melanoma,^{70,71} but of note, these meta-analyses included the same primary studies. In addition, a meta-analysis resulted in a significantly increased risk of kidney or bladder cancer in BB users,⁷² while another meta-analysis demonstrated a significantly reduced risk for hepatocellular carcinoma in patients with liver cirrhosis using non-selective BBs.⁷³ Of interest, none of these meta-analyses used HF patients, or HF as an indication for BB use exclusively, according to their study eligibility criteria.

Effects of beta-blockers on cancer outcomes in patients with prevalent cancer

Meta-analyses on breast cancer showed that BB use was associated with either no effect,^{74–76} or with improved cancer outcomes^{77–79} compared to non-users, even when BBs were started after diagnosis of malignancy.⁷⁹ Most meta-analyses on lung cancer show no

associations of BB use with cancer outcomes.^{74–76,79} Still, one meta-analysis reported that (i) non-selective BB use was associated with significantly worse overall survival of lung cancer patients, and that (ii) BB use (not stratified by selectivity) was associated with improved overall survival in stage III patients and in those without surgical cancer treatment.⁸⁰ With respect to colorectal cancer, no association has been found between BB use and cancer outcomes.^{74–76,79} Regarding malignant melanoma, repeated analyses on the same cohorts hinted towards improved overall survival in patients using BB.^{74–76} Conversely, another meta-analysis that included additional cohort studies showed no association of BB use with beneficial cancer outcomes in patients with malignant melanoma.⁷⁹

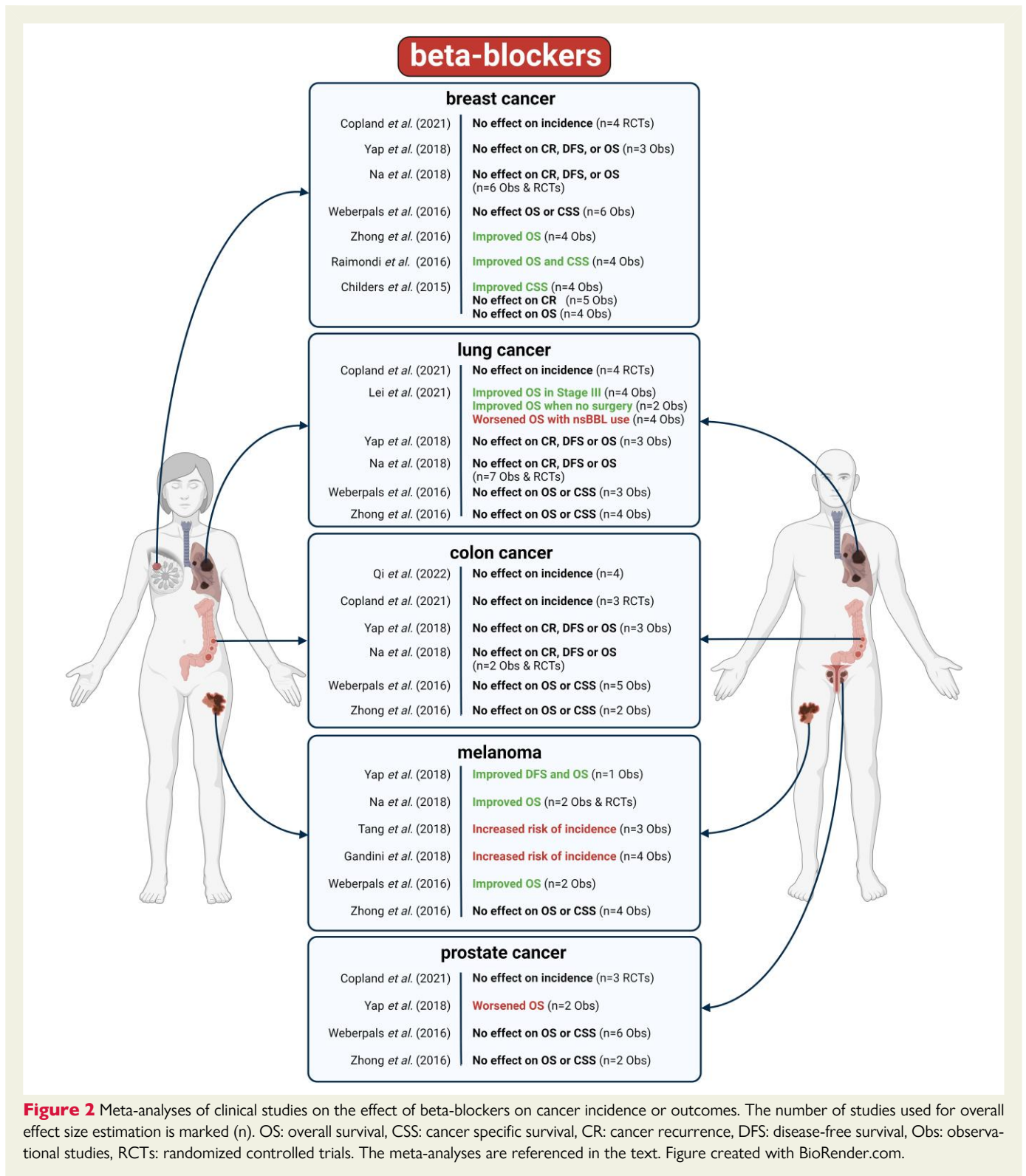
In conclusion, although BBs have been shown to exert significant anti-cancer effects in pre-clinical studies, meta-analyses of clinical studies show inconsistent and sometimes conflicting results regarding the associations of BB use with cancer incidence and outcomes, independently from cancer site and outcome measures. Moreover, there is no consensus on how beta-adrenoceptor selectivity influences the effect of BBs on cancer neither in pre-clinical nor in clinical studies. It should also be stressed that BBs have no proven effects in HF with preserved ejection fraction, and only minor cardioprotective effects in cancer patients receiving chemotherapy.⁸¹ Nevertheless, as the activation of sympathetic nervous system on cancer outcomes has been well-established in both pre-clinical studies and the clinical settings,⁸² there is a strong rationale to further investigate the sympathetic nervous system–cancer relationships.

Effects of renin-angiotensin-aldosterone system inhibitors on cancer

Many studies have shown that dysregulation of the renin-angiotensin-aldosterone system (RAAS) may promote cancer, mainly driven by the AT1R-Akt axis (Figure 1).⁸³ The idea for investigating the effects of RAAS blockade on cancer has first emerged by the retrospective analysis of Lever *et al.*, showing that patients using angiotensin-converting enzyme inhibitors (ACEIs) had a reduced risk for developing cancer, and also suggested the need to assess the effects of angiotensin receptor blockers (ARBs) on cancer as well.⁸⁴ This seminal study gave rise to pre-clinical and clinical studies testing the hypothesis that RAAS blockade entails anti-cancer effects, as well as to investigations demonstrating that components of RAAS are expressed in various human cancers and in their micro-environment,⁸³ which are associated with worse outcomes.^{85,86}

Pre-clinical studies assessing the effects of renin-angiotensin-aldosterone system inhibitors on cancer

The anti-cancer effects of blocking the RAAS with ACEIs were tested in numerous pre-clinical *in vivo* studies, mostly demonstrating benefits, that could be exerted either alone or in combination with other anti-cancer therapies (see [Supplementary data online, Table S2](#)).^{87–91} Nevertheless, contradictory findings were reported in an early study by Wysocki and colleagues, where captopril did not exert significant anti-cancer effects, but was associated with increased mortality in immunocompetent mouse models of renal cancer.⁹² However, in a more recent study using a similar immunocompetent cancer model and the same cancer cell line, captopril significantly reduced primary tumour weight and lung metastases. Of note, captopril treatment was



started 2 days prior to tumour inoculation, and in a lower dose.⁹³ The anti-cancer effects of RAAS blockade by ACEIs are further supported by *in vitro* studies, showing a reduction in cell proliferation, migration, and invasion.^{94–99} Nevertheless, in contrast to these *in vivo* studies, several *in vitro* studies reported no direct anti-cancer effects of

ACEIs,^{91,100–102} or no synergism with other anti-cancer agents (see [Supplementary data online, Table S3](#)).¹⁰³ Whether findings of these pre-clinical studies are a class effect not known, as captopril was assessed almost exclusively. Thus, a comprehensive, systematic research strategy to assess the effects of different types of ACEIs is lacking.

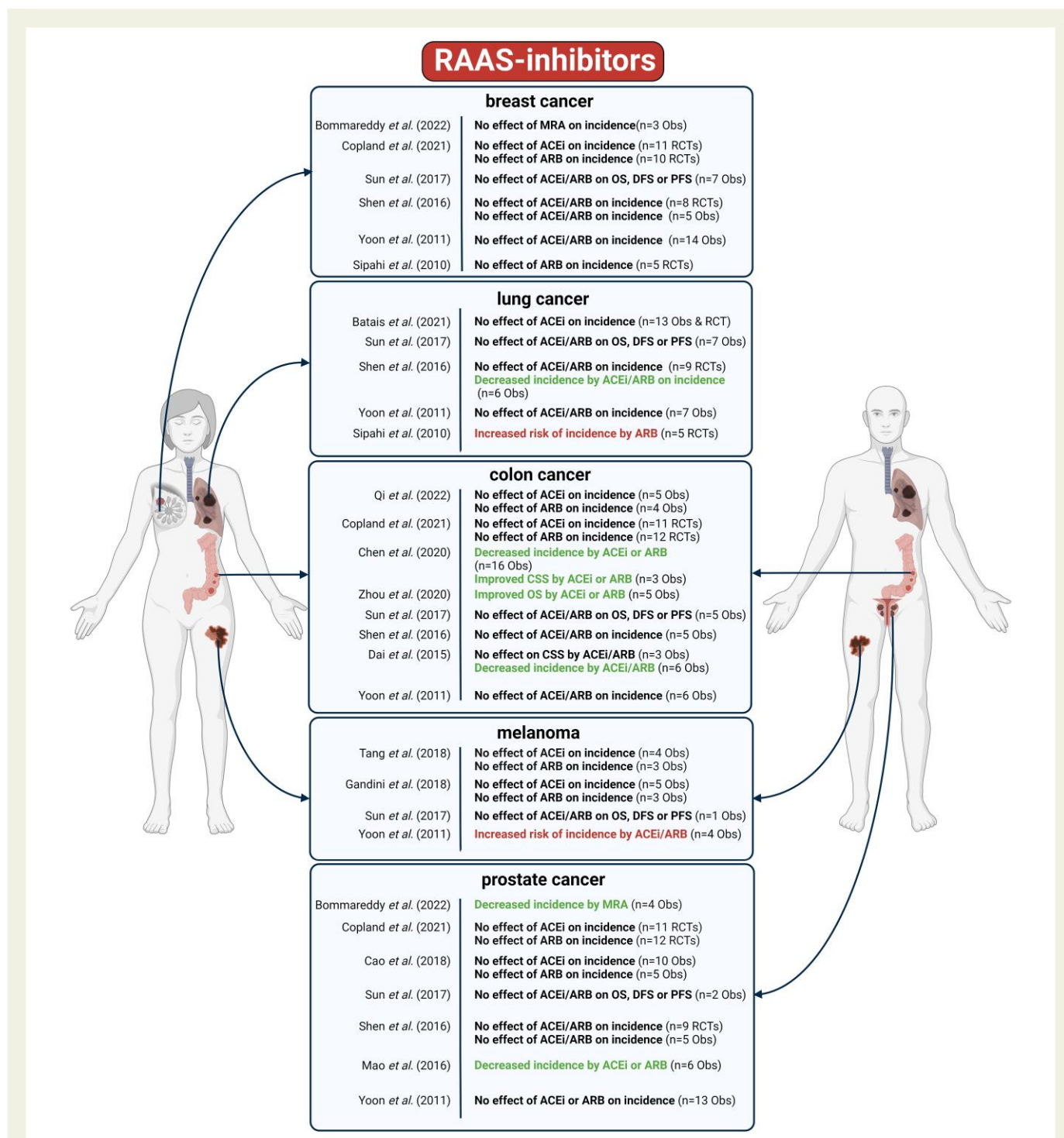


Figure 3 Meta-analyses of clinical studies on the effect of RAAS inhibitors (angiotensin receptor blockers, angiotensin-converting enzyme inhibitors, or mineralocorticoid receptor antagonists) on cancer incidence or outcomes. The number of studies used for overall effect size estimation is marked (n). OS: overall survival, CSS: cancer specific survival, CR: cancer recurrence, DFS: disease-free survival, Obs: observational studies, RCTs: randomized controlled trials. The meta-analyses are referenced in the text. Figure created with BioRender.com.

ARBs inhibit the AT1R, the key target of angiotensin II, which is the major effector peptide of the RAAS. In tumour-bearing mice, ARBs exert significant anti-cancer effects by reducing tumour growth and/or fibrosis,^{104–107} metastases,^{108,109} tumour neo-angiogenesis,^{110–112} and influencing tumour immuno-microenvironment (see [Supplementary](#)

[data online, Table S2](#)).^{108,113} In a seminal study by Rhodes *et al.*, AT1R is overexpressed in 10%–20% of breast cancer cases across multiple independent patient cohorts. The study indicated that marked AT1R-overexpression defines a subpopulation of estrogen receptor-positive, ERBB2-negative breast cancer that may benefit from targeted

therapy with ARBs, most particularly losartan. These findings were obtained in both *in vitro* and *in vivo* models AT1R-overexpressing breast cancer, but not in AT1R^{low} cell line.¹¹⁴ Nevertheless, contradictory results were obtained by a number of *in vitro* and *in vivo* studies demonstrating no or less anti-cancer effects of ARBs, mostly using losartan or irbesartan^{99,115–127} (see [Supplementary data online, Table S3](#)).

Mineralocorticoid receptor antagonists (MRAs) represent a third pillar part of HF GDMT. Only a handful of pre-clinical studies investigated effects of MRAs on cancer currently. Leung and colleagues demonstrated that spironolactone decreased the number of intestinal polyps in APJ^{min} mice (a mouse model of spontaneous intestinal adenoma formation), and inhibited metastases in colorectal carcinoma-implantation studies by pathways that are independent of the mineralocorticoid receptor.¹²⁸ Other *in vivo* studies also demonstrated a significant anti-cancer effect of MRAs by reducing tumour volume,^{129–131} and/or by inhibiting metastatic spread¹³² (see [Supplementary data online, Table S2](#)). Accordingly, the majority of *in vitro* studies also show an overall anti-cancer effect of MRAs either given alone or in combination with other therapeutics (see [Supplementary data online, Table S3](#)).^{129,133–137} Of note, Gold and colleagues reported differences in anti-cancer efficacy of different MRAs, showing superiority of spironolactone over eplerenone,¹³⁰ a differential effect that needs further elucidation regarding mechanism. Conversely, lack of anti-cancer effects for spironolactone was also found on liver,¹³¹ and pituitary cancer cell lines.¹³⁸ Moreover, Aldaz and colleagues demonstrated that spironolactone (either alone, or in combination with dexamethasone) protected the glioblastoma cells against radiation-induced damage.¹³⁹

Clinical studies assessing the effects of renin-angiotensin-aldosterone system inhibitors on cancer

The putative effects of RAAS blockade on malignancy in HF patients were investigated in a seminal meta-analysis by Sipahi and colleagues, in which only RCTs of ARBs were analyzed ([Figure 3](#), see [Supplementary data online, Table S4](#)). Here, a significant association between the use of ARBs and overall cancer risk was reported, mostly attributed to new-onset lung cancer.¹⁴⁰ These results raised doubts about the reliability of this meta-analysis, as adjudication of cancer diagnoses was not uniform among the included studies.¹⁴¹

Effects of renin-angiotensin-aldosterone system inhibitors on risk of cancer in cancer-free patients

Other meta-analyses of RCTs reported that ARB or ACEI usage was not associated with cancer risk compared to placebo.^{67,68,142–145} This lack of association between RAAS inhibitor (RAASI) use and incident cancer has also been suggested by meta-analyses of cohort studies across multiple cancer types.^{69–71,146,147} However, meta-analyses of non-randomized investigations demonstrated a significantly decreased incidence of esophagus,¹⁴⁸ colorectal,¹⁴⁹ prostate,¹⁵⁰ and lung cancer,¹⁴³ but an increased risk for renal cancer^{72,148} and melanoma¹⁴⁸ amongst users of ACEI/ARB compared to non-users. Although a large number of meta-analyses have been conducted to investigate the association between ACEI/ARB use and cancer incidence, only a single recent meta-analysis by Bommareddy and colleagues assessed the effect of spironolactone on cancer occurrence.¹⁵¹ This meta-analysis synthesized data from observational studies, showing a significantly decreased risk for prostate cancer, but no effect on other cancer types. Similar to BBs, effects of RAASIs on cancer incidence were mostly assessed in hypertensive, but not in HF populations by the meta-analyses.

Effects of renin-angiotensin-aldosterone system inhibitors on cancer outcomes in patients with prevalent cancer

In contrast to the disparate effects of RAASIs on cancer incidence, meta-analyses of observational studies demonstrated significantly improved cancer outcomes in patients with digestive system malignancies,^{152–154} renal cancer,^{153,155} or all-cause cancer¹⁵⁶ in users of ACEIs or ARBs. Nevertheless, a meta-analysis on RCTs showed the neutral effect of RAASIs in cancer patients irrespective of the cancer type.¹⁵⁷

In conclusion, the effect of RAASIs on new-onset cancer risk is conflicting in the current clinical data, varying mostly by cancer types, and also, by primary study design (i.e. RCT or observational). However, in patients with prevalent cancer, the majority of meta-analyses either show safety, or even improved cancer-related outcomes when RAASIs are used, compared to non-users. Nevertheless, a major factor that complicates the interpretation of these results is the confounding by indication, i.e. most of the clinical data are derived from patients with hypertension, and not with HF, urging for further evidence in the HF populations as well.

Effects of angiotensin receptor-neprilysin inhibitor on cancer

There is a general lack of pre-clinical and clinical evidence regarding the effect of angiotensin receptor-neprilysin inhibitor (ARNI, i.e. sacubitril/valsartan) on cancer, which is another part for HF GDMT,¹ a drug that enhances the beneficial cardiovascular effects of endogenous natriuretic peptides (NP)^{158,159} Of note, in the landmark RCT leading to approval of ARNI for treatment of HF with reduced ejection fraction, proportion of cancer deaths was comparable in the ARNI and ACEI arms.¹⁶⁰ Moreover, in a recent cohort study on patients with HF with mildly reduced ejection fraction, ARNI/ACEI/ARB use significantly increased cancer incidence in the primary outcomes within 3 years, although this association was not significant by falsification analysis.¹⁶¹

NPs have been shown to inhibit tumour growth in several *in vitro* and *in vivo* studies,^{162–165} nevertheless, these associations should be interpreted with caution, as some malignant cells are also able to produce NPs, questioning generalizability of tumour-inhibitory effects of NPs.^{166,167} In addition, it is noteworthy that in principle neprilysin inhibition also increases the availability of factors other than NPs that might influence cancer cell biology.¹⁶⁸ The effects of these substrates should also be considered in future investigations addressing the effects of ARNI on cancer.

Effects of sodium-glucose cotransporter 2 inhibitors on cancer

As glucose is a major substrate required for cancer cell survival and growth, Scafoglio and colleagues hypothesized that the metabolism-shifting effect of sodium-glucose cotransporter 2 inhibitors (SGLT2Is) might be protective against malignancy as well.¹⁶⁹ In this seminal investigation, functional expression of SGLT2 on human pancreatic and prostate cancers was demonstrated. In addition, this was the first study providing evidence on SGLT2Is blocking glucose uptake and reducing

tumour growth in a xenograft model of pancreatic cancer, which led to the conduction of subsequent pre-clinical studies investigating the anti-cancer effects of SGLT2Is.¹⁷⁰

Pre-clinical studies assessing the effects of sodium-glucose cotransporter 2 inhibitors on cancer

Glucose uptake/metabolism-dependent anti-cancer mechanisms of SGLT2Is have been further demonstrated in cancer-bearing mice, which was attributed to activation of adenosine monophosphate-activated protein kinase (AMPK), and thus, to the inhibition of mTOR^{171–173} (Figure 1, Supplementary data online, Table S2). Nevertheless, Kaji *et al.* reported the anti-cancer effect of SGLT2 inhibition to be exerted independently of the systemic glycemic status, although cellular glucose-uptake was not assessed here.¹⁷⁴ Other mechanistic pathways on the anti-cancer effects of SGLT2 blockade were suggested by other studies, showing that SGLT2 inhibition (i) decreases pro-carcinogenic inflammation,¹⁷⁵ (ii) activates AMPK, which leads to inactivation of the protooncogene sonic hedgehog,¹⁷⁶ (iii) suppresses cancer progression by inhibiting the Hippo signalling pathway through downregulating YAP1 expression.¹⁷⁷ In contrast, however, Korfhage and colleagues reported an increased intestinal adenoma burden in female, but not in male APC^{min} mice, when treated with canagliflozin,¹⁷⁸ a result that needs further mechanistic elucidation.

Several *in vitro* studies also indicated that anti-cancer mechanisms of SGLT2Is are mainly attributed to the induction of AMPK,¹⁷⁹ which subsequently leads to the inhibition of the Akt/mTOR pathway (see Supplementary data online, Table S3).¹⁸⁰ In addition, analyses of metabolomics in SGLT2I-treated cancer cell lines showed that besides the glucose-dependent mechanisms, alteration of other metabolic pathways (e.g. fatty acid metabolism) also contributes to the decrease in cancer cell proliferation.^{181,182} Other *in vitro* studies reported that anti-proliferative effects of SGLT2Is are exerted by a significant repression of DNA synthesis,¹⁸³ subsequent cell cycle arrest,¹⁸⁴ and by blocking aberrant activation of β -catenin.¹⁸⁵ In the latter study, dapagliflozin and empagliflozin (the two SGLT2Is that are currently recommended in HF) exerted non-significant effects, questioning the presence of a class effect.

Of note, in addition to tumour growth studies, SGLT2Is were also investigated in cancer therapy-related toxicity studies. For instance, dapagliflozin and empagliflozin were shown to revert ponatinib-induced endothelial cell senescence and dysfunction.¹⁸⁶

Clinical studies assessing the effects of sodium-glucose cotransporter 2 inhibitors on cancer

During the safety trials of SGLT2Is in diabetic patients, no significant increase in overall cancer events was observed. Nevertheless, a nominal increase in bladder cancer incidence in men, and breast cancer incidence in women were noted in the SGLT2I-treated arm.¹⁸⁷ These observations have led to systematic investigations of the association between SGLT2I use and cancer, showing inconsistent results (see Supplementary data online, Table S4 and Figure 4).

For instance, a recent meta-analysis of hyperglycaemic patients reported an overall reduced risk of cancer associated with SGLT2I use, and most particularly with dapagliflozin and ertugliflozin vs. placebo.¹⁸⁸ Of note, in this meta-analysis, two trials with large sample sizes may have shifted the overall effect size towards benefit by SGLT2Is, whereas

the majority of the included studies had large confidence intervals (i.e. small sample sizes) with non-significant effects. Surprisingly, an earlier meta-analysis showed no association of SGLT2I with malignancy, however (i) dapagliflozin significantly increased risk of overall cancer compared to other antidiabetic drugs, and (ii) empagliflozin nominally increased the risk of overall cancer compared to placebo in patients with type 2 diabetes mellitus (T2DM).¹⁸⁹ Another meta-analysis on T2DM patients showed that risk of overall cancer in obese patients was significantly increased in association with SGLT2I use. This meta-analysis also showed a tendency towards increased risk of cancer in studies with a follow-up period of >52 months. Moreover, risk of bladder cancer also significantly increased, mainly associated with the use of empagliflozin.¹⁹⁰ In contrast, Dicembrini and colleagues reported a significant risk reduction in bladder cancer associated with dapagliflozin use, although this result was derived from four RCTs, one of which might have been outweighed, thus, dominating the overall effect size.¹⁹¹

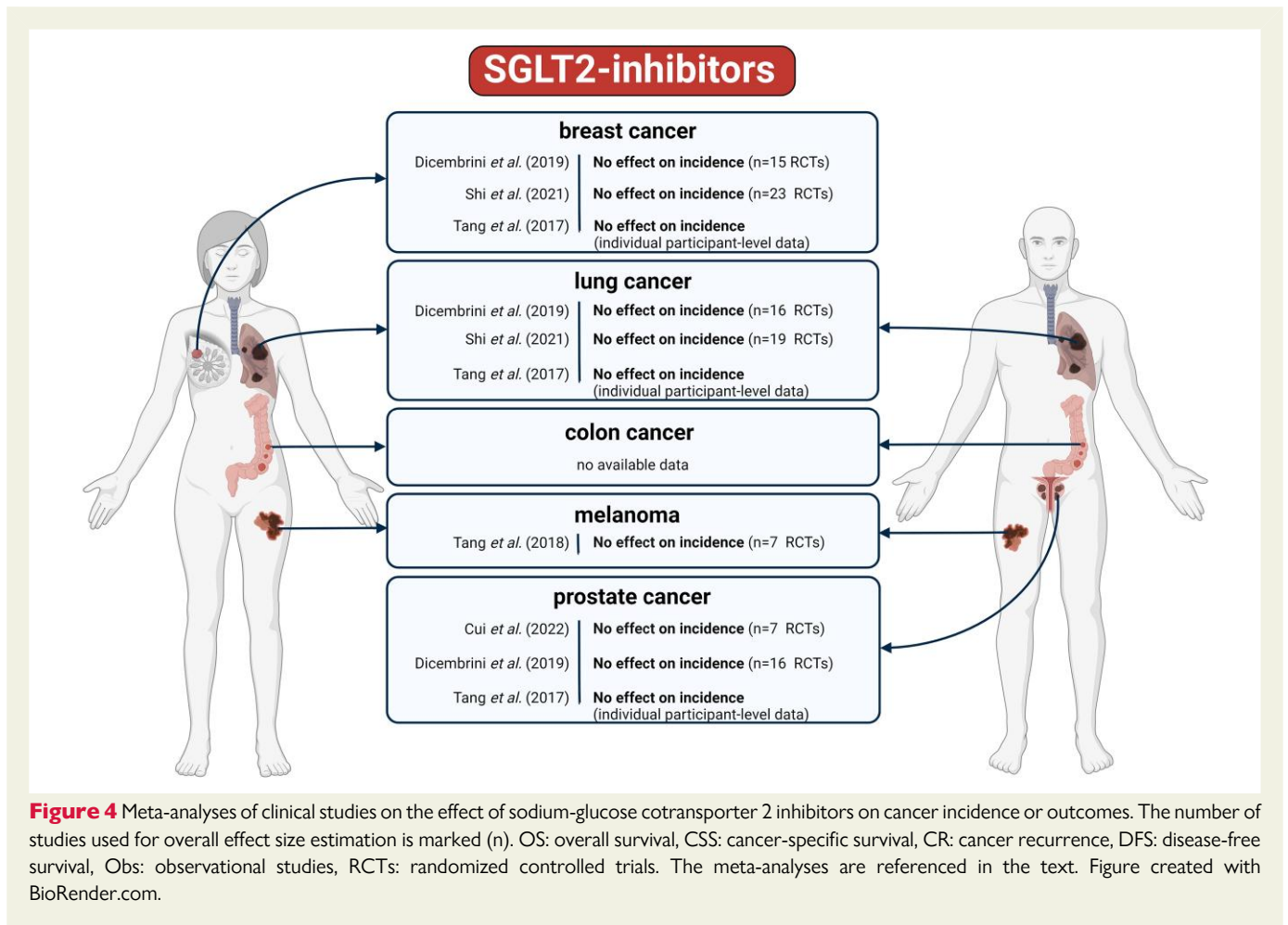
Meta-analyses investigating effects of SGLT2Is on outcomes by cancer types show no significant change in the risk of breast cancer,^{189–191} lung cancer,^{189–191} prostate cancer,^{190–192} or melanoma,¹⁹³ with the latter cancer showing a tendency to increase by SGLT2Is. In addition, similar neutral results were reported regarding renal, pancreatic and hepatocellular cancers as well.¹⁹¹

As the use of SGLT2I with the indication for HF has only recently been introduced, most meta-analyses synthesize data from studies of patients with diabetes. Nevertheless, the putative association between SGLT2I use and cancer outcomes of HF patients—with or without diabetes—requires investigation in future meta-analyses. Moreover, as the above meta-analyses assessed only the risk of cancer, future studies should also assess the outcomes of patients with prevalent cancer.

Effects of digoxin on cancer

Although not considered as a pillar part for HF pharmacotherapy, digoxin is still used in selected HF patients.¹ Effects of digoxin on cancer have been investigated in a variety of *in vitro* studies, mostly showing anti-cancer properties by causing cell-cycle arrest.^{194–200} These findings were supported by *in vivo* studies, showing inhibition of tumour growth,^{200–202} or reducing distant tumour formation.²⁰³ Despite the appealing pre-clinical data, meta-analyses on clinical studies show rather contradictory results. For instance, Ahern and colleagues performed an observational study and a meta-analysis showing a significantly increased risk of breast cancer in digoxin users vs. non-users.²⁰⁴ This finding was further supported by a meta-analysis also reporting significantly increased risk of breast cancer, lung cancer, and colorectal cancer, but not prostate cancer in association with digoxin use.²⁰⁵ In addition, significantly increased all-cause mortality of cancer patients using digoxin was also reported, but no increase in cancer-specific mortality could be detected. It should be emphasized here that these results should be interpreted with caution, as clinical studies may be biased by (i) a higher likelihood of medical contact, and (ii) an intrinsic tendency towards worse outcomes (not only restricted to cancer or cardiovascular outcomes), as patients taking digoxin are on average sicker than those not on this medication.

Overall, there is an apparent discrepancy between the pre-clinical studies (almost unanimously demonstrating anti-cancer effects) and the clinical investigations (showing a tendency towards worse cancer outcomes) regarding effects of digoxin on cancer. This discrepancy highlights the need for increasing the translational value of pre-clinical



research, and the reliability of clinical data that are synthesized by meta-analyses.

Effects of diuretics on cancer

Loop diuretics (e.g. furosemide) are used in HF patients to reduce symptoms and signs of congestion.¹ The target molecule of furosemide, Na-K-2Cl-transporter has been shown to be expressed on cancer cells, playing a key role in cancer cell growth. Pre-clinical studies have demonstrated anti-cancer effects of furosemide, which was attributed to Na-K-2Cl-transporter inhibition,^{206–208} however, no such effects were seen in clinical studies.²⁰⁹ Although thiazides are not the preferred diuretic agents for decongestion purposes in HF, it should be noted that use of hydrochlorothiazide has been brought in association with increased risk of skin cancer,²¹⁰ however, a recent meta-analysis has found no such associations.²¹¹ In summary, the interaction between diuretics and malignancy remains inconclusive, especially in HF populations, and further investigations are required to validate the interaction between diuretics and cancer.

Future directions for decreasing cancer burden of heart failure

Overall, extensive pre-clinical evidence shows significant anti-cancer effects of all HF GDMT drug classes, nevertheless, no such anti-cancer

effects of HF drugs could be confirmed in the clinical reality (Figure 5)—a discrepancy that is not at all restricted to the field of cardio-oncology.^{212,213} These findings emphasize the need to conduct pre-clinical studies of higher translational value, and more robust and reliable clinical studies of higher quality, in order to facilitate the formation of recommendations aiming to decrease cancer burden of HF patients (Figure 6).

Considerations for future pre-clinical studies investigating the effect of heart failure drugs on cancer

Pre-clinical *in vivo* and *in vitro* studies complement each other, as *in vitro* studies might fail in taking into account the complexity of the systematic effects of a drug, while better exploring the direct effects on cancer.

Another limitation for translation stems from the lack of standardized practice for drug dosing and administration, as there is a high variety of doses of the same drug between cancer studies. Also, HF drug doses in cancer studies do not necessarily correspond to doses used in HF studies.

In addition, there is a difference in the interpretation of studies where (i) administration starts prior to tumour inoculation (i.e. tumour growth inhibition study), or where (ii) administration starts after an established tumour nodule has already formed (i.e. tumour growth delay study).

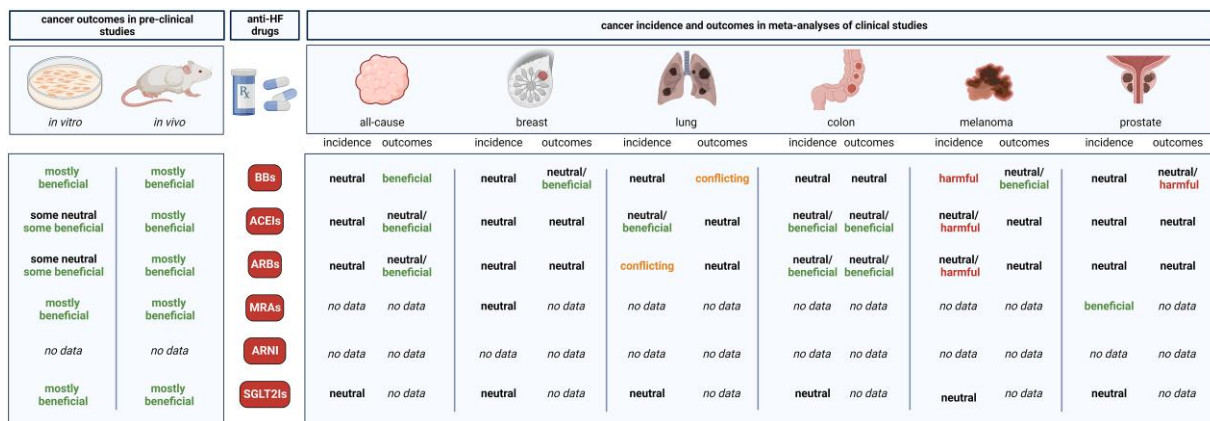


Figure 5 A graphical summary for both the pre-clinical and clinical evidence on the effect of different heart failure pharmacotherapies on cancer. The terms were defined as follows: beneficial: decreases cancer incidence, or improves any patient outcome; neutral: no effect on cancer incidence, or no effect on any patient outcome; harmful: increased incidence or worsening of any patient outcome; conflicting: there are studies showing either benefit or harm on cancer incidence or outcomes. Figure created with BioRender.com.

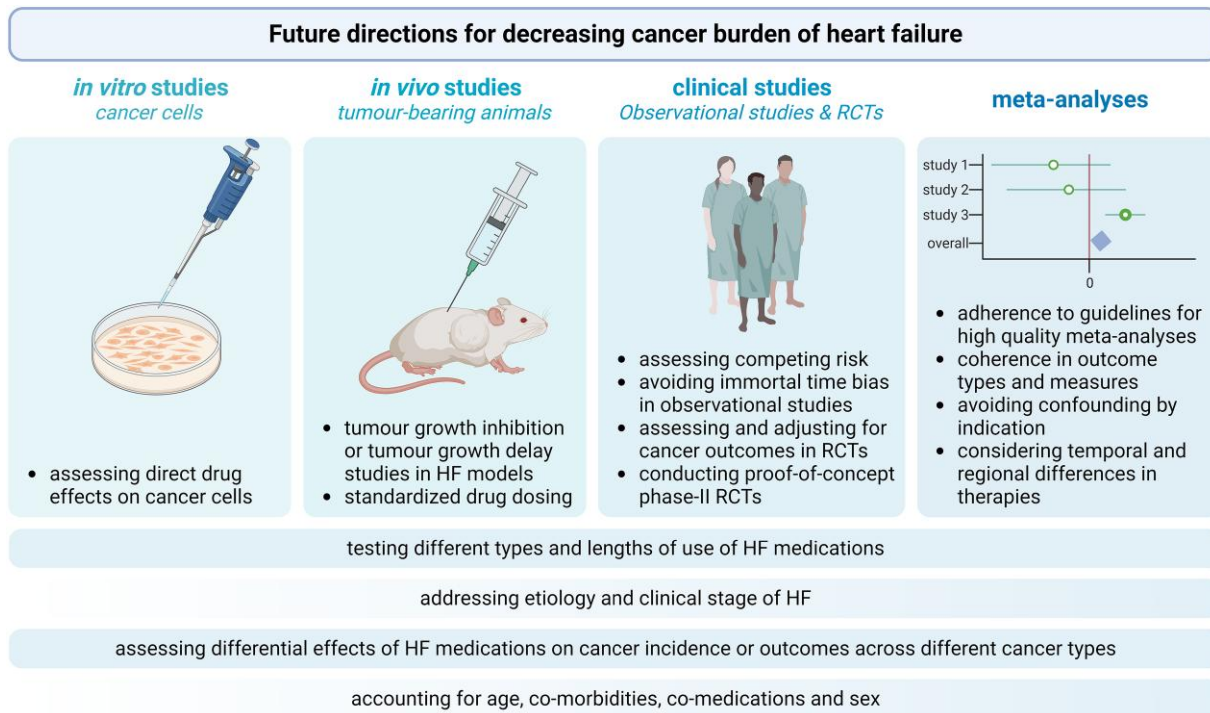


Figure 6 A graphical summary of future directions for decreasing cancer burden of heart failure from pre-clinical studies to clinical investigations and their meta-analyses. Figure created with BioRender.com.

Another important aspect for increasing translational value could be the use of *in vivo* tumour-models where cardio-metabolic diseases are also induced to better mimic the frequent clinical situation of HF with co-morbidities. Although pioneer *in vivo* studies assessing tumour growth in animals with prevalent HF induced by either myocardial infarction or transverse aortic constriction (TAC) have already been published, studies assessing the effect of HF medications either alone or combined in such settings should follow.^{214–217} Of note, a TAC model,

depending on the severity and length of constriction, could mimic cardiac diseases ranging from non-ischemic HF with reduced or preserved ejection fraction, to aortic stenosis,²¹⁸ which mimic important sub-populations of HF patients.

Finally, to further enhance translational value, (i) differential effects of HF medications either combined or alone, (ii) different etiologies and stages of prevalent HF, (iii) different cancer types, (iv) effects of age, co-morbidities—e.g. hypertension, atrial fibrillation, obesity, and

co-medications that are present amongst the majority of HF patients, and (v) sex-based differences should be considered when planning future pre-clinical studies to this field.

Considerations for future clinical investigations on the effect of heart failure drugs on cancer

Several issues are intrinsic to the study of cardiovascular disease and cancer. First, a major obstacle in the synchronous study of cardiovascular disease and cancer is that, inevitably, successful treatment of the one condition will provide an opportunity for the other condition to progress and become a more important cause of death. For instance, a very powerful HF drug might reduce HF-related outcomes and cardiovascular death, e.g. an MRA. But at the same time, this will provide more opportunity for (latent) cancers to progress and become manifest. This *competing risk* by no means is synonymous to a pro-oncogenic effect of MRA. Vice versa, breast cancer survivors have after 10 years a larger cardiovascular risk than cancer risk.¹² Since these women have survived one potentially lethal condition, they have beaten the cancer risk (at least for some time), while their cardiovascular risk continues and likely has risen due to aggressive cancer treatments. This complex interplay complicates the simultaneous study of cardiovascular disease and cancer and is very difficult to adjust.

Second, observational studies are prone to biases, a problem that was touched upon by the meta-analysis of Weberpals and colleagues, where BB use significantly increased overall survival and cancer specific survival of cancer patients. However, when observational studies prone to immortal time bias were excluded from the analysis, no significant effects were found for any investigated outcome.⁷⁴

Third, indication for the use of HF drugs was not always attributed to HF exclusively in current observational studies, but rather to hypertension (for ACEIs/ARBs) or diabetes (for SGLT2Is), causing confounding by indication. Therefore, to generate evidence whether and how HF treatment should be modified to improve (or at least not to worsen) cancer-related outcomes in cohorts of HF patients with prevalent cancer is of paramount importance.

Fourth, crucially, cancer outcomes generally are poorly adjudicated in most cardiovascular RCTs, which intrinsically flaws the outputs of any meta-analysis. Systematic assessment of new-onset cancer risk in future HF RCTs is essential to collect valuable information with potential clinical implications, that may require longer follow-up after termination for cardiovascular end-points.^{219,220} On the other hand, systematic assessment of cardiovascular outcomes in future cancer trials is equally important. For instance, the latest RCT with immune checkpoint inhibitors (ICI) did not systematically collect troponin values, while we know that ICI-mediated myocarditis is a potentially lethal side-effect of immunotherapy, which occurs in 2%–5% of all patients.²²¹

In general, stratifying patients in clinical investigations based on (i) type and length of use of HF pharmacotherapy of different combinations, (ii) etiology and clinical stage of HF, (iii) cancer type, (iv) age, co-morbidities, and co-medications, is essential because individual patients with co-morbidities may require other types of drugs than HF medications, and importantly (v) based on sex, may more clearly show how HF medications affect cancer incidence, progression, and outcomes, being of paramount importance for clinical decision-making. For instance, the differential sex-related effects of HF medications were addressed by Stolfo and colleagues who showed that female HF patients were more likely to receive BBs, diuretics, and digoxin. Of note, digoxin use was associated with an increased risk of death in females,²²² but

females were less likely to receive RAASIs compared to male HF patients.²²³

Overall, definitive answers would be obtained from proof-of-concept phase II RCTs that directly assess the effects of HF drugs on cancer in HF patients; therefore, such investigations are eagerly waiting to be conducted in the future. Of note, although direct effects of HF drugs on cancer, or the effects of successfully reversed HF on cancer may be hard to dissect in future studies, if the outcome is definitive, this question should be addressed by further mechanistic investigations (*Graphical Abstract*).

Besides guideline-directed pharmacotherapies of HF, investigating other therapeutic options would also facilitate solving this issue. For instance, the interleukin-1beta inhibitor canakinumab has been shown to reduce HF-related hospitalization and mortality,^{224,225} and cumulative incidence of lung cancer in atherosclerotic patients,²²⁶ raising the question whether targeting inflammation, a shared pathomechanistic pathway of both HF and cancer, could mean a solution for decreasing cancer burden of HF patients.

In conclusion, translatability of pre-clinical studies, and reliability of clinical investigations should be improved to facilitate decision-making on whether and how HF treatment should be modified to decrease cancer incidence and improve cancer outcomes of HF patients.

Acknowledgements

Figures were in part created with *Biorender.com*.

Supplementary Data

Supplementary data are available at *European Heart Journal* online.

Declarations

Disclosure of Interest

P.F. is the founder and CEO of Pharmahungary Group, a group of R&D companies. G.S. reports grants and personal fees from Vifor, grants from Boehringer Ingelheim, personal fees from Societa' Prodotti Antibiotici, grants and personal fees from AstraZeneca, personal fees from Roche, personal fees from Servier, grants and personal fees from Novartis, personal fees from GENESIS, grants and personal fees from Cytokinetics, personal fees from Medtronic, grants from Boston Scientific, grants and personal fees from PHARMACOSMOS, grants from Merck, grants from Bayer, personal fees from TEVA, outside the submitted work. PA reports speaker and/or advisor fees from Astra Zeneca, Boehringer Ingelheim, Bayer, Novartis, Daiichi Sankyo, Amgen, Janssen, and MSD, all outside the submitted work. ELG has received speaker honoraria or consultancy fees from AstraZeneca, Bayer, Boehringer Ingelheim, Bristol-Myers SquiBB, Pfizer, Novo Nordisk, Lundbeck Pharma, and Organon. He is investigator in clinical studies sponsored by AstraZeneca, Idorsia, or Bayer and has received unrestricted research grants from Boehringer Ingelheim. D.D. obtained honoraria for educational lectures from Novartis, Sanofi, and Daiichi Sankyo, all unrelated to this work. R.A.dB has received research grants and/or fees from AstraZeneca, ABBott, Boehringer Ingelheim, Cardior Pharmaceuticals GmbH, Ionis Pharmaceuticals, Inc., Novo Nordisk, and Roche; and has had speaker engagements with ABBott, AstraZeneca, Bayer, Bristol Myers SquiBB, Novartis, and Roche. AGS has received speakers honoraria from Merck/Schering-Plough, BMS, UCB, Pfizer/

Wyeth, Sanofi, Novartis, Pfizer, Lilly, and Women's College Hospital, Toronto, Canada. Other authors have nothing to declare.

Data Availability

No data were generated or analysed for or in support of this paper.

Funding

The work was supported by the European Union's Horizon 2020 Research and Innovation Programme under grant agreement no. 739593 and by a Momentum Research Grant from the Hungarian Academy of Sciences (LP-2021-38 to Z.V.V.). This project was additionally supported by grants from the National Research, Development and Innovation Office (NKFIH) of Hungary (FK134751 to Z.V.V.). The Ministry for Innovation and Technology in Hungary provided funding to this study under the Thematic Excellence Programme (2020-4.1.1.-TKP2020), the 2020-1.1.5-GYORSÍTÓSÁV call programme (2020-1.1.5-GYORSÍTÓSÁV-2021-00011), the TKP2021- EGA funding scheme (TKP2021-EGA-23), and the Research Excellence Programme (TKP/ITM/NKFIH). This study was also supported by the National Heart Laboratory Project no. RRF-2.3.1-21-2022-00003 implemented with the support provided by the European Union. N.V.S. was supported by the Semmelweis 250 + Excellence PhD Scholarship (EFOP-3.6.3-VEKOP-16-2017-00009) provided by the Semmelweis University, Budapest Hungary, and by the Gedeon Richter Talentum Foundation scholarship. P.A. was supported by the Italian Ministry of Health (GR-2018-12365661—CHANGE Study; Ricerca Corrente 2018-2020 IRCCS Ospedale Policlinico San Martino). This work was carried out with the support of the University of Medicine and Pharmacy "Carol Davila", Bucharest (to A.I.). D.D. was supported by National Institutes of Health grants (R01HL131517, R01HL136389, R01HL089598, R01HL163277, and R01HL160992) and the European Union (large-scale integrative project MAESTRIA, No. 965286). W.C.M. and R.A.dB are supported by the European Research Council (ERC CoG 818715).

Author contributions

Z.V.V. and P.F. conceived the review, and provided overall supervision and funding. N.V.S. wrote the manuscript. Á.M.P. and N.V.S. collected and analysed data regarding *in vivo* and *in vitro* studies. N.V.S. collected and analysed data regarding clinical meta-analyses. N.V.S. made the figures. All authors made substantial contributions to the conception and design, acquisition of data, or analysis and interpretation of data. All authors drafted or revised the manuscript critically for important intellectual content, and agreed to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

References

- McDonagh TA, Metra M, Adamo M, Gardner RS, Baumbach A, Böhm M, et al. 2021 ESC guidelines for the diagnosis and treatment of acute and chronic heart failure. *Eur Heart J* 2021;**42**:3599–726. <https://doi.org/10.1093/eurheartj/ehab368>
- Heidenreich PA, Bozkurt B, Aguilar D, Allen LA, Byun JJ, Colvin MM, et al. 2022 AHA/ACC/HFSA guideline for the management of heart failure: a report of the American college of cardiology/American heart association joint committee on clinical practice guidelines. *J Am Coll Cardiol* 2022;**79**:e263–421. [doi:https://doi.org/10.1016/j.jacc.2021.12.012](https://doi.org/10.1016/j.jacc.2021.12.012)
- Lyon AR, López-Fernández T, Couch LS, Asteggiano R, Aznar MC, Bergler-Klein J, et al. 2022 ESC guidelines on cardio-oncology developed in collaboration with the European hematology association (EHA), the European society for therapeutic radiology and oncology (ESTRO) and the international cardio-oncology society (IC-OS): developed by the T. *Eur Heart J* 2022;**43**:4229–361. <https://doi.org/10.1093/eurheartj/ehac244>
- James SL, Abate D, Abate KH, Abay SM, Abbafati C, Abbasi N, et al. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: a systematic analysis for the global burden of disease study 2017. *Lancet* 2018;**392**:1789–858. [https://doi.org/10.1016/S0140-6736\(18\)32279-7](https://doi.org/10.1016/S0140-6736(18)32279-7)
- Savarese G, Becher PM, Lund LH, Seferovic P, Rosano GMC, Coats AJS. Global burden of heart failure: a comprehensive and updated review of epidemiology. *Cardiovasc Res* 2022;**118**:3272–87. <https://doi.org/10.1093/cvr/cvac013>
- Sung H, Ferlay J, Siegel RL, Laversanne M, Soerjomataram I, Jemal A, et al. Global cancer statistics 2020: gLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2021;**71**:209–49. <https://doi.org/10.3322/caac.21660>
- Aboumsallem JP, Moslehi J, de Boer RA. Reverse cardio-oncology: cancer development in patients with cardiovascular disease. *J Am Heart Assoc* 2020;**9**:e013754. <https://doi.org/10.1161/JAHA.119.013754>
- de Boer RA, Hulot J-S, Tocchetti CG, Aboumsallem JP, Ameri P, Anker SD, et al. Common mechanistic pathways in cancer and heart failure. A scientific roadmap on behalf of the translational research committee of the heart failure association (HFA) of the European society of cardiology (ESC). *Eur J Heart Fail* 2020;**22**:2272–89. [doi:https://doi.org/10.1002/ehf.2029](https://doi.org/10.1002/ehf.2029)
- Bertero E, Canepa M, Maack C, Ameri P. Linking heart failure to cancer. *Circulation* 2018;**138**:735–42. <https://doi.org/10.1161/CIRCULATIONAHA.118.033603>
- Lau ES, Paniagua SM, Liu E, Jovani M, Li SX, Takvorian K, et al. Cardiovascular risk factors are associated with future cancer. *JACC Cardio Oncol* 2021;**3**:48–58. [doi:https://doi.org/10.1016/j.jacc.2020.12.003](https://doi.org/10.1016/j.jacc.2020.12.003)
- Narayan V, Thompson EW, Demissei B, Ho JE, Januzzi JL, Ky B. Mechanistic biomarkers informative of both cancer and cardiovascular disease: JACC state-of-the-art review. *J Am Coll Cardiol* 2020;**75**:2726–37. [doi:https://doi.org/10.1016/j.jacc.2020.03.067](https://doi.org/10.1016/j.jacc.2020.03.067)
- Sturgeon KM, Deng L, Bluethmann SM, Zhou S, Trifiletti DM, Jiang C, et al. A population-based study of cardiovascular disease mortality risk in US cancer patients. *Eur Heart J* 2019;**40**:3889–97. <https://doi.org/10.1093/eurheartj/ehz766>
- Zaorsky NG, Churilla TM, Egleston BL, Fisher SG, Ridge JA, Horwitz EM, et al. Causes of death among cancer patients. *Ann Oncol* 2017;**28**:400–7. [doi:https://doi.org/10.1093/annonc/mdw604](https://doi.org/10.1093/annonc/mdw604)
- Mehta LS, Watson KE, Barac A, Beckie TM, Bittner V, Cruz-Flores S, et al. Cardiovascular disease and breast cancer: where these entities intersect: a scientific statement from the American heart association. *Circulation* 2018;**137**:e30–66. <https://doi.org/10.1161/CIR.0000000000000556>
- Zamorano JL, Gottfridsson C, Asteggiano R, Atar D, Badimon L, Bax JJ, et al. The cancer patient and cardiology. *Eur J Heart Fail* 2020;**22**:2290–309. [doi:https://doi.org/10.1002/ehf.1985](https://doi.org/10.1002/ehf.1985)
- Tocchetti CG, Ameri P, de Boer RA, D'Alessandra Y, Russo M, Sorriento D, et al. Cardiac dysfunction in cancer patients: beyond direct cardiomyocyte damage of anticancer drugs: novel cardio-oncology insights from the joint 2019 meeting of the ESC working groups of myocardial function and cellular biology of the heart. *Cardiovasc Res* 2020;**116**:1820–34. <https://doi.org/10.1093/cvr/cvaa222>
- Efentakis P, Andreadou I, Iliodromitis KE, Triposkiadis F, Ferdinandy P, Schulz R, et al. Myocardial protection and current cancer therapy: two opposite targets with inevitable cost. *Int J Mol Sci* 2022;**23**:14121. <https://doi.org/10.3390/ijms232214121>
- Gergely TG, Kucsera D, Tóth VE, Kovács T, Sayour NV, Drobní ZD, et al. Characterization of immune checkpoint inhibitor-induced cardiotoxicity reveals interleukin-17A as a driver of cardiac dysfunction after anti-PD-1 treatment. *Br J Pharmacol* 2023;**180**:740–61. [doi:https://doi.org/10.1111/bph.15984](https://doi.org/10.1111/bph.15984)
- Banke A, Schou M, Videbæk L, Møller JE, Torp-Pedersen C, Gustafsson F, et al. Incidence of cancer in patients with chronic heart failure: a long-term follow-up study. *Eur J Heart Fail* 2016;**18**:260–6. [doi:https://doi.org/10.1002/ehf.472](https://doi.org/10.1002/ehf.472)
- Hasin T, Gerber Y, Weston SA, Jiang R, Killian JM, Manemann SM, et al. Heart failure after myocardial infarction is associated with increased risk of cancer. *J Am Coll Cardiol* 2016;**68**:265–71. [doi:https://doi.org/10.1016/j.jacc.2016.04.053](https://doi.org/10.1016/j.jacc.2016.04.053)
- Battisti NML, Michael S, de Mark B, John D, Clive W, Peake MD, et al. Prevalence of cardiovascular disease in patients with potentially curable malignancies. *JACC Cardio Oncol* 2022;**4**:238–53. <https://doi.org/10.1016/j.jacc.2022.03.004>
- Leedy DJ, Reding KW, Vasbinder AL, Anderson GL, Barac A, Wactawski-Wende J, et al. The association between heart failure and incident cancer in women: an analysis of the women's health initiative. *Eur J Heart Fail* 2021;**23**:1712. [doi:https://doi.org/10.1002/ehf.2207](https://doi.org/10.1002/ehf.2207)
- Roderburg C, Loosen SH, Jahn JK, Gänsbacher J, Luedde T, Kostev K, et al. Heart failure is associated with an increased incidence of cancer diagnoses. *ESC Hear. Fail* 2021;**8**:3628–33. <https://doi.org/10.1002/ehf2.13421>
- Edoardo B, Fabio R, Eliana R, Antonio D, Lucia B, Lidia S, et al. Cancer incidence and mortality according to Pre-existing heart failure in a community-based cohort. *JACC CardioOncol* 2022;**4**:98–109. <https://doi.org/10.1016/j.jacc.2021.11.007>
- Camilli M, Chiabrando JG, Lombardi M, Del Buono MG, Montone RA, Lombardo A, et al. Cancer incidence and mortality in patients diagnosed with heart failure: results from an updated systematic review and meta-analysis. *Cardio-Oncol* 2023;**9**:8. <https://doi.org/10.1186/s40959-023-00158-1>

26. Selvaraj S, Bhatt DL, Claggett B, Djoussé L, Shah SJ, Chen J, et al. Lack of association between heart failure and incident cancer. *J Am Coll Cardiol* 2018;**71**:1501–10. doi: <https://doi.org/10.1016/j.jacc.2018.01.069>
27. Schwartz B, Schou M, Gislason GH, Køber L, Torp-Pedersen C, Andersson C. Prevalence and incidence of Various cancer subtypes in patients with heart failure vs matched controls. *Int J Cardiol* 2020;**316**:209–13. <https://doi.org/10.1016/j.ijcard.2020.05.035>
28. Bruhn J, Malmborg M, Garred CH, Ravn P, Zahir D, Andersson C, et al. Temporal trends in the incidence of malignancy in heart failure: a nationwide danish study. *Eur Heart J* 2023;**44**:1124–32. <https://doi.org/10.1093/eurheartj/ehac797>
29. Ameri P, Bertero E, Meijers WC. Cancer is a comorbidity of heart failure. *Eur Heart J* 2023;**44**:1133–5. <https://doi.org/10.1093/eurheartj/ehac710>
30. Cole SW, Sood AK. Molecular pathways: beta-adrenergic signaling in cancer. *Clin Cancer Res* 2012;**18**:1201–6. <https://doi.org/10.1158/1078-0432.CCR-11-0641>
31. Sloan EK, Priceman SJ, Cox BF, Yu S, Pimentel MA, Tangkanangnu V, et al. The sympathetic nervous system induces a metastatic switch in primary breast cancer. *Cancer Res* 2010;**70**:7042–52. <https://doi.org/10.1158/0008-5472.CAN-10-0522>
32. Zhou C, Chen X, Zeng W, Peng C, Huang G, Li X, et al. Propranolol induced G0/G1/S phase arrest and apoptosis in melanoma cells via AKT/MAPK pathway. *Oncotarget* 2016;**7**:68314–27. <https://doi.org/10.18632/oncotarget.11599>
33. Partecke LI, Speerforck S, Käding A, Seubert F, Kühn S, Lorenz E, et al. Chronic stress increases experimental pancreatic cancer growth, reduces survival and can be antagonised by Beta-adrenergic receptor blockade. *Pancreatology* 2016;**16**:423–33. <https://doi.org/10.1016/j.pan.2016.03.005>
34. Wei W-J, Shen C-T, Song H-J, Qiu Z-L, Luo Q-Y. Propranolol sensitizes thyroid cancer cells to cytotoxic effect of vemurafenib. *Oncol Rep* 2016;**36**:1576–84. <https://doi.org/10.3892/or.2016.4918>
35. Maccari S, Buoncervello M, Rampin A, Spada M, Macchia D, Giordani L, et al. Biphasic effects of propranolol on tumour growth in B16F10 melanoma-bearing mice. *Br J Pharmacol* 2017;**174**:139–49. <https://doi.org/10.1111/bph.13662>
36. Wrobel LJ, Le Gal FA. Inhibition of human melanoma growth by a non-cardioselective β -blocker. *J Invest Dermatol* 2015;**135**:525–31. doi:<https://doi.org/10.1038/jid.2014.373>
37. Sorski L, Melamed R, Matzner P, Lavon H, Shaashua L, Rosenne E, et al. Reducing liver metastases of colon cancer in the context of extensive and Minor surgeries through β -adrenoceptors blockade and COX2 inhibition. *Brain Behav Immun* 2016;**58**:91–8. <https://doi.org/10.1016/j.bbi.2016.05.017>
38. Kuang X, Qi M, Peng C, Zhou C, Su J, Zeng W, et al. Propranolol enhanced the anti-tumor effect of sunitinib by inhibiting proliferation and inducing G0/G1/S phase arrest in malignant melanoma. *Oncotarget* 2018;**9**:802–11. <https://doi.org/10.18632/oncotarget.22696>
39. Pasquier E, Street J, Pouchy C, Carre M, Gifford AJ, Murray J, et al. β -Blockers increase response to chemotherapy via direct antitumour and anti-angiogenic mechanisms in neuroblastoma. *Br J Cancer* 2013;**108**:2485–94. <https://doi.org/10.1038/bjc.2013.205>
40. Palm D, Lang K, Niggemann B, Drell IV TL, Masur K, Zaenker KS, et al. The norepinephrine-driven metastasis development of PC-3 human prostate cancer cells in BALB/c nude mice is inhibited by β -blockers. *Int J Cancer* 2006;**118**:2744–9. doi: <https://doi.org/10.1002/ijc.21723>
41. Kokolus KM, Zhang Y, Sivik JM, Schmeck C, Zhu J, Repasky EA, et al. Beta blocker use correlates with better overall survival in metastatic melanoma patients and improves the efficacy of immunotherapies in mice. *Oncimmunology* 2018;**7**:e1405205. <https://doi.org/10.1080/2162402X.2017.1405205>
42. Fjæstad KY, Rømer AMA, Goitea V, Johansen AZ, Thorseth M-L, Carretta M, et al. Blockade of Beta-adrenergic receptors reduces cancer growth and enhances the response to anti-CTLA4 therapy by modulating the tumor microenvironment. *Oncogene* 2022;**41**:1364–75. <https://doi.org/10.1038/s41388-021-02170-0>
43. Moisan F, Oucherif S, Kaulanjan-Checkmodine P, Prey S, Rousseau B, Bonneu M, et al. Critical role of aquaporin-1 and telocytes in infantile hemangioma response to propranolol Beta blockade. *Proc Natl Acad Sci USA* 2021;**118**:e2018690118. <https://doi.org/10.1073/pnas.2018690118>
44. MacDonald CR, Bucsek MJ, Qiao G, Chen M, Evans L, Greenberg DJ, et al. Adrenergic receptor signaling regulates the response of tumors to ionizing radiation. *Radiat Res* 2019;**191**:585–9. <https://doi.org/10.1667/RR15193.1>
45. Acheampong DO, Baffour IK, Atsu Barku VY, Addo JK, Essuman MA, Boye A. Zanthoxylum Zanthoxyloides alkaloidal extract improves CCL4-induced hepatocellular carcinoma-like phenotypes in rats. *Evid Based Complement Altern Med* 2021;**2021**:3804379. <https://doi.org/10.1155/2021/3804379>
46. Balaha M, Kandeel S, Barakat W. Carvedilol suppresses circulating and hepatic IL-6 responsible for hepatocarcinogenesis of chronically damaged liver in rats. *Toxicol Appl Pharmacol* 2016;**311**:1–11. <https://doi.org/10.1016/j.taap.2016.10.012>
47. Cleveland KH, Yeung S, Huang KM, Liang S, Andresen BT, Huang Y. Phosphoproteome profiling provides insight into the mechanism of action for carvedilol-mediated cancer prevention. *Mol Carcinog* 2018;**57**:997–1007. <https://doi.org/10.1002/mc.22820>
48. Gillis RD, Botteri E, Chang A, Ziegler AI, Chung N-C, Pon CK, et al. Carvedilol blocks neural regulation of breast cancer progression in vivo and is associated with reduced breast cancer mortality in patients. *Eur J Cancer* 2021;**147**:106–16. <https://doi.org/10.1016/j.ejca.2021.01.029>
49. Abdullah Shamim M, Yeung S, Shahid A, Chen M, Wang J, Desai P, et al. Topical carvedilol delivery prevents UV-induced skin cancer with negligible systemic absorption. *Int J Pharm* 2022;**611**:121302. <https://doi.org/10.1016/j.ijpharm.2021.121302>
50. Chang A, Yeung S, Thakkar A, Huang KM, Liu MM, Kanassatega R-S, et al. Prevention of skin carcinogenesis by the β -blocker carvedilol. *Cancer Prev Res* 2015;**8**:27–36. <https://doi.org/10.1158/1940-6207.CAPR-14-0193>
51. Hoare JJ, Osmani B, O'Sullivan EA, Browne A, Campbell N, Metcalf S et al. Carvedilol targets β -arrestins to rewire innate immunity and improve oncolytic adenoviral therapy. *Commun Biol* 2022;**5**:106. doi:10.1038/s42003-022-03041-4
52. Huang KM, Liang S, Yeung S, Oiyemhonlan E, Cleveland KH, Parsa C, et al. Topically applied carvedilol attenuates solar ultraviolet radiation induced skin carcinogenesis. *Cancer Prev Res* 2017;**10**:598–606. <https://doi.org/10.1158/1940-6207.capr-17-0132>
53. Chen Q, Jiang H, Wang Z, Cai L-Y, Jiang Y-C, Xie L, et al. Adrenergic blockade by nebivolol to suppress oral squamous cell carcinoma growth via endoplasmic Reticulum stress and mitochondria dysfunction. *Front Pharmacol* 2021;**12**:691998. <https://doi.org/10.3389/fphar.2021.691998>
54. Niu M, Xu J, Liu Y, Li Y, He T, Ding L, et al. FBXL2 counteracts Grp94 to destabilize EGFR and inhibit EGFR-driven NSCLC growth. *Nat Commun* 2021;**12**:5919. <https://doi.org/10.1038/s41467-021-26222-x>
55. Zhang D, Yong Ma Q, Hu H-T, Zhang M. B2-Adrenergic antagonists suppress pancreatic cancer cell invasion by inhibiting CREB, NF-KB and AP-1. *Cancer Biol Ther* 2010;**10**:19–29. <https://doi.org/10.4161/cbt.10.1.11944>
56. Xie W, He R, Zhang J, He Y, Wan Z, Zhou C, et al. B-blockers inhibit the viability of breast cancer cells by regulating the ERK/COX-2 signaling pathway and the drug response is affected by ADRB2 single-nucleotide polymorphisms. *Oncol Rep* 2019;**41**:341–50. <https://doi.org/10.3892/or.2018.6830>
57. Szweczyk M, Richter C, Briesse V, Richter D-U. A retrospective in vitro study of the impact of anti-diabetics and cardioselective pharmaceuticals on breast cancer. *Anticancer Res* 2012;**32**:2133–8. <https://ar.iiarjournals.org/content/32/5/2133>
58. Koh M, Takahashi T, Kurokawa Y, Kobayashi T, Saito T, Ishida T, et al. Propranolol suppresses gastric cancer cell growth by regulating proliferation and apoptosis. *Gastric Cancer* 2021;**24**:1037–49. <https://doi.org/10.1007/s10120-021-01184-7>
59. Sidorova M, Petrikaitė V. The effect of Beta adrenoceptor blockers on viability and cell colony formation of non-small cell lung cancer cell lines A549 and H1299. *Molecules* 2022;**27**:1938. <https://doi.org/10.3390/molecules27061938>
60. Farhoudand LS, Fiorentzis M, Kraemer MM, Sak A, Stuschke M, Rassaf T, et al. The adrenergic receptor antagonist carvedilol elicits anti-tumor responses in uveal melanoma 3D tumor spheroids and may serve as co-adjuvant therapy with radiation. *Cancers (Basel)* 2022;**14**:3097. <https://doi.org/10.3390/cancers14133097>
61. Duckett MM, Phung SK, Nguyen L, Khammanivong A, Dickerson E, Dusenbery K, et al. The adrenergic receptor antagonists propranolol and carvedilol decrease bone sarcoma cell viability and sustained carvedilol reduces clonogenic survival and increases radiosensitivity in canine osteosarcoma cells. *Vet Comp Oncol* 2020;**18**:128–40. doi: <https://doi.org/10.1111/vco.12560>
62. Zhang D, Ma Q, Shen S, Hu H. Inhibition of pancreatic cancer cell proliferation by propranolol occurs through apoptosis induction: the study of Beta-adrenoceptor antagonist's anticancer effect in pancreatic cancer cell. *Pancreas* 2009;**38**:94–100. <https://doi.org/10.1097/MPA.0b013e318184f50c>
63. Negro A, Brar BK, Gu Y, Peterson KL, Vale W, Lee K-F. ErbB2 is required for G protein-coupled receptor signaling in the heart. *Proc Natl Acad Sci U S A* 2006;**103**:15889–93. <https://doi.org/10.1073/pnas.0607499103>
64. Sysa-Shah P, Tocchetti CG, Gupta M, Rainer PP, Shen X, Kang B-H, et al. Bidirectional cross-regulation between ErbB2 and β -adrenergic signalling pathways. *Cardiovasc Res* 2016;**109**:358–73. <https://doi.org/10.1093/cvr/cvv274>
65. Guglin M, Krischer J, Tamura R, Fink A, Bello-Matricaria L, McCaskill-Stevens W, et al. Randomized trial of lisinopril versus carvedilol to prevent trastuzumab cardiotoxicity in patients with breast cancer. *J Am Coll Cardiol* 2019;**73**:2859–68. <https://doi.org/10.1016/j.jacc.2019.03.495>
66. Monami M, Filippi L, Ungar A, Sgrilli F, Antenore A, Dicembrini I, et al. Further data on Beta-blockers and cancer risk: observational study and meta-analysis of randomized clinical trials. *Curr Med Res Opin* 2013;**29**:369–78. <https://doi.org/10.1185/03007995.2013.772505>
67. Bangalore S, Kumar S, Kjeldsen SE, Makani H, Grossman E, Wetterslev J, et al. Antihypertensive drugs and risk of cancer: network meta-analyses and trial sequential analyses of 324 168 participants from randomised trials. *Lancet Oncol* 2011;**12**:65–82. doi:[https://doi.org/10.1016/S1470-2045\(10\)70260-6](https://doi.org/10.1016/S1470-2045(10)70260-6)
68. Copland E, Canoy D, Nazarzadeh M, Bidel Z, Ramakrishnan R, Woodward M, et al. Antihypertensive treatment and risk of cancer: an individual participant data meta-analysis. *Lancet Oncol* 2021;**22**:558–70. [https://doi.org/10.1016/S1470-2045\(21\)00033-4](https://doi.org/10.1016/S1470-2045(21)00033-4)
69. Qi J, An R, Bhatti P, Spinelli JJ, Murphy RA. Anti-Hypertensive medications and risk of colorectal cancer: a systematic review and meta-analysis. *Cancer Causes Control* 2022;**33**:801–12. <https://doi.org/10.1007/s10552-022-01570-1>
70. Gandini S, Palli D, Spadola G, Bendinelli B, Cocorocchio E, Stanganelli I, et al. Anti-Hypertensive drugs and skin cancer risk: a review of the literature and

- meta-analysis. *Crit Rev Oncol Hematol* 2018;**122**:1–9. doi:<https://doi.org/10.1016/j.critrevonc.2017.12.003>
71. Tang H, Fu S, Zhai S, Song Y, Han J. Use of antihypertensive drugs and risk of malignant melanoma: a meta-analysis of observational studies. *Drug Saf* 2018;**41**:161–9. <https://doi.org/10.1007/s40264-017-0599-x>
 72. Xie Y, Xu P, Wang M, Zheng Y, Tian T, Yang S, et al. Antihypertensive medications are associated with the risk of kidney and bladder cancer: a systematic review and meta-analysis. *Aging (Albany, NY)* 2020;**12**:1545–62. <https://doi.org/10.18632/aging.102699>
 73. Thiele M, Albillos A, Abazi R, Wiest R, Gluud LL, Krag A. Non-Selective Beta-blockers may reduce risk of hepatocellular carcinoma: a meta-analysis of randomized trials. *Liver Int* 2015;**35**:2009–16. doi:<https://doi.org/10.1111/liv.12782>
 74. Weberpals J, Jansen L, Carr PR, Hoffmeister M, Brenner H. Beta blockers and cancer prognosis—the role of immortal time bias: a systematic review and meta-analysis. *Cancer Treat Rev* 2016;**47**:1–11. doi:<https://doi.org/10.1016/j.ctrv.2016.04.004>
 75. Na Z, Qiao X, Hao X, Fan L, Xiao Y, Shao Y, et al. The effects of Beta-blocker use on cancer prognosis: a meta-analysis based on 319,006 patients. *Oncol Targets Ther* 2018;**11**:4913–44. <https://doi.org/10.2147/OTT.S167422>
 76. Yap A, Lopez-Olivo MA, Dubowitz J, Pratt G, Hiller J, Gottumukkala V, et al. Effect of Beta-blockers on cancer recurrence and survival: a meta-analysis of epidemiological and perioperative studies. *Br J Anaesth* 2018;**121**:45–57. doi:<https://doi.org/10.1016/j.bja.2018.03.024>
 77. Childers WK, Hollenbeck CS, Cheriya P. β -Blockers reduce breast cancer recurrence and breast cancer death: a meta-analysis. *Clin Breast Cancer* 2015;**15**:426–31. <https://doi.org/10.1016/j.clbc.2015.07.001>
 78. Raimondi S, Botteri E, Munzone E, Cipolla C, Rotmensz N, DeCensi A, et al. Use of Beta-blockers, angiotensin-converting enzyme inhibitors and angiotensin receptor blockers and breast cancer survival: systematic review and meta-analysis. *Int J Cancer* 2016;**139**:212–9. doi:<https://doi.org/10.1002/ijc.30062>
 79. Zhong S, Yu D, Zhang X, Chen X, Yang S, Tang J, et al. β -Blocker use and mortality in cancer patients: systematic review and meta-analysis of observational studies. *Eur J Cancer Prev* 2016;**25**:440–8. <https://doi.org/10.1097/CEJ.0000000000000192>
 80. Lei Z, Yang W, Zuo Y. Beta-Blocker and survival in patients with lung cancer: a meta-analysis. *PLoS One* 2021;**16**:e0245773. <https://doi.org/10.1371/journal.pone.0245773>
 81. Vaduganathan M, Hirji SA, Qamar A, Bajaj N, Gupta A, Zaha VG, et al. Efficacy of neurohormonal therapies in preventing cardiotoxicity in patients with cancer undergoing chemotherapy. *JACC CardioOncol* 2019;**1**:54–65. doi:<https://doi.org/10.1016/j.jacc.2019.08.006>
 82. Cole SW, Nagaraja AS, Lutgendorf SK, Green PA, Sood AK. Sympathetic nervous system regulation of the tumour microenvironment. *Nat Rev Cancer* 2015;**15**:563–72. <https://doi.org/10.1038/nrc3978>
 83. George AJ, Thomas WG, Hannan RD. The renin–angiotensin system and cancer: old dog, new tricks. *Nat Rev Cancer* 2010;**10**:745–59. <https://doi.org/10.1038/nrc2945>
 84. Lever AF, Hole DJ, Gillis CR, McCallum IR, McInnes GT, MacKinnon PL, et al. Do inhibitors of angiotensin-I-converting enzyme protect against risk of cancer? *Lancet* 1998;**352**:179–84. doi:[https://doi.org/10.1016/S0140-6736\(98\)03228-0](https://doi.org/10.1016/S0140-6736(98)03228-0)
 85. Iino K, Shibata K, Kajiyama H, Yamamoto E, Nagasaka T, Nawa A, et al. Angiotensin II type 1 receptor expression in ovarian cancer and its correlation with tumour angiogenesis and patient survival. *Br J Cancer* 2006;**94**:552–60. <https://doi.org/10.1038/sj.bjc.6602961>
 86. Arrieta O, Villarreal-Garza C, Vizcaíno G, Pineda B, Hernández-Pedro N, Guevara-Salazar P, et al. Association between AT1 and AT2 angiotensin II receptor expression with cell proliferation and angiogenesis in operable breast cancer. *Tumour Biol* 2015;**36**:5627–34. <https://doi.org/10.1007/s13277-015-3235-3>
 87. Nakaya K, Otsuka H, Kondo K, Otani T, Nagata M. Tumor growth-inhibitory effect of an angiotensin-converting enzyme inhibitor (captopril) in a lung cancer Xenograft model analyzed using 18F-FDG-PET/CT. *Nucl Med Commun* 2016;**37**:139–46. <https://doi.org/10.1097/MNM.0000000000000404>
 88. Fujita M, Hayashi I, Yamashina S, Fukamizu A, Itoman M, Majima M. Angiotensin type 1a receptor signaling-dependent induction of vascular endothelial growth factor in stroma is relevant to tumor-associated angiogenesis and tumor growth. *Carcinogenesis* 2005;**26**:271–9. <https://doi.org/10.1093/carcin/bgh324>
 89. Attoub S, Gaben AM, Al-Salam S, Al Sultan MAH, John A, Nicholls MG, et al. Captopril as a potential inhibitor of lung tumor growth and metastasis. *Ann NY Acad Sci* 2008;**1138**:65–72. doi:<https://doi.org/10.1196/annals.1414.011>
 90. Jordan BF, Peeterbroeck J, Karroum O, Diepart C, Magat J, Grégoire V, et al. Captopril and S-nitrosocaptopril as potent radiosensitizers: comparative study and underlying mechanisms. *Cancer Lett* 2010;**293**:213–9. doi:<https://doi.org/10.1016/j.canlet.2010.01.016>
 91. Yang Y, Ma L, Xu Y, Liu Y, Li W, Cai J, et al. Enalapril overcomes chemoresistance and potentiates antitumor efficacy of 5-FU in colorectal cancer by suppressing proliferation, angiogenesis, and NF-KB/STAT3-regulated proteins. *Cell Death Dis* 2020;**11**:477. <https://doi.org/10.1038/s41419-020-2675-x>
 92. Wysocki PJ, Kwiatkowska EP, Kazmierczak U, Suchorska W, Kowalczyk DW, Mackiewicz A. Captopril, an angiotensin-converting enzyme inhibitor, promotes growth of immunogenic tumors in mice. *Clin Cancer Res* 2006;**12**:4095–102. <https://doi.org/10.1158/1078-0432.CCR-05-2489>
 93. Araújo VF, Naves MA, Ravanini JN, Schor N, Teixeira VPC. Renin-Angiotensin system (RAS) blockade attenuates growth and metastatic potential of renal cell carcinoma in mice. *Urol Oncol Semin Orig Invest* 2015;**33**:389.e1–e7. <https://doi.org/10.1016/j.urolonc.2014.11.022>
 94. Brown RE, Lun M, Prichard JW, Blasick TM, Zhang PL. Morphoproteomic and pharmacoproteomic correlates in hormone-receptor-negative breast carcinoma cell lines. *Ann Clin Lab Sci* 2004;**34**:251–62. <http://www.annclinlabsci.org/content/34/3/251.long>
 95. Roprai HK, Kandaneeratchi A, Maidment SL, Christidou M, Trillo-Pazos G, Dexter DT, et al. Evaluation of the effects of swainsonine, captopril, tangeretin and nobilinetin on the biological behaviour of brain tumour cells in vitro. *Neuropathol Appl Neurobiol* 2001;**27**:29–39. <https://doi.org/10.1046/j.0305-1846.2000.00298.x>
 96. Williams RN, Parsons SL, Morris TM, Rowlands BJ, Watson SA. Inhibition of matrix metalloproteinase activity and growth of gastric adenocarcinoma cells by an angiotensin converting enzyme inhibitor in vitro and murine models. *Eur J Surg Oncol* 2005;**31**:1042–50. <https://doi.org/10.1016/j.ejso.2005.04.003>
 97. De la Iglesia Iñigo S, López-Jorge CE, Gómez-Casares MT, Lemes Castellano A, Martín Cabrera P, López Brito J, et al. Induction of apoptosis in leukemic cell lines treated with captopril, trandolapril and losartan: a new role in the treatment of leukaemia for these agents. *Leuk Res* 2009;**33**:810–6. <https://doi.org/10.1016/j.leukres.2008.09.029>
 98. Fendrich V, Lopez CL, Manoharan J, Maschuw K, Wichmann S, Baier A, et al. Enalapril and ASS inhibit tumor growth in a transgenic mouse model of islet cell tumors. *Endocr Relat Cancer* 2014;**21**:813–24. <https://doi.org/10.1530/ERC-14-0175>
 99. Chen Y-H, Huang C-H, Lu H-I, Chen C-H, Huang W-T, Hsieh M-J, et al. Prognostic impact of renin-angiotensin system blockade in esophageal squamous cell carcinoma. *J Renin Angiotensin Aldosterone Syst* 2015;**16**:1185–92. <https://doi.org/10.1177/1470320314535275>
 100. Rasha F, Ramalingam L, Menikdiwela K, Hernandez A, Moussa H, Gollahon L, et al. Renin angiotensin system inhibition attenuates adipocyte-breast cancer cell interactions. *Exp Cell Res* 2020;**394**:112114. <https://doi.org/10.1016/j.yexcr.2020.112114>
 101. Lu Y, Lian S, Ye Y, Yu T, Liang H, Cheng Y, et al. S-Nitrosocaptopril prevents cancer metastasis in vivo by creating the Hostile bloodstream microenvironment against circulating tumor cells. *Pharmacol Res* 2019;**139**:535–49. <https://doi.org/10.1016/j.phrs.2018.10.020>
 102. Carl-McGrath S, Ebert MPA, Lendeckel U, Röcken C. Expression of the local angiotensin II system in gastric cancer may facilitate lymphatic invasion and nodal spread. *Cancer Biol Ther* 2007;**6**:1218–26. <https://doi.org/10.4161/cbt.6.8.4412>
 103. Smith TAD, Phyu SUM, Akabuogu EU. Effects of administered cardioprotective drugs on treatment response of breast cancer cells. *Anticancer Res* 2016;**36**:87–93. <https://ar.iiarjournals.org/content/36/1/87.long>
 104. Okazaki M, Fushida S, Harada S, Tsukada T, Kinoshita J, Oyama K, et al. The angiotensin II type 1 receptor blocker candesartan suppresses proliferation and fibrosis in gastric cancer. *Cancer Lett* 2014;**355**:46–53. <https://doi.org/10.1016/j.canlet.2014.09.019>
 105. Takagi H, Kaji K, Nishimura N, Ishida K, Ogawa H, Takaya H, et al. The angiotensin II receptor blocker losartan sensitizes human liver cancer cells to lenvatinib-mediated cytostatic and angiostatic effects. *Cells* 2021;**10**:575. <https://doi.org/10.3390/cells10030575>
 106. Mainetti LE, Rico MJ, Kaufman CD, Grillo MC, Guercetti J, Baglioni MV, et al. Losartan improves the therapeutic effect of metronomic cyclophosphamide in triple negative mammary cancer models. *Oncotarget* 2020;**11**:3048–60. <https://doi.org/10.18632/oncotarget.27694>
 107. Li W, Li S, Chen IX, Liu Y, Ramjiawan RR, Leung C-H, et al. Combining losartan with radiotherapy increases tumor control and inhibits lung metastases from a HER2/neu-positive orthotopic breast cancer model. *Radiat Oncol* 2021;**16**:48. <https://doi.org/10.1186/s13014-021-01775-9>
 108. Regan DP, Coy JW, Chahal KK, Chow L, Kurihara JN, Guth AM, et al. The angiotensin receptor blocker losartan suppresses growth of pulmonary metastases via AT1R-independent inhibition of CCR2 signaling and monocyte recruitment. *J Immunol* 2019;**202**:3087–102. <https://doi.org/10.4049/jimmunol.1800619>
 109. Cai X-J, Wang Z, Xu Y-Y, Yang G-Y, Zhang R-Y, Wang Y. Candesartan treatment enhances liposome penetration and anti-tumor effect via depletion of tumor stroma and normalization of tumor vessel. *Drug Deliv Transl Res* 2021;**11**:1186–97. <https://doi.org/10.1007/s13346-020-00842-0>
 110. Ruderman S, Eshein A, Valuckaitė V, Dougherty U, Almoghrabi A, Gomes A, et al. Early increase in blood supply (EIBS) is associated with tumor risk in the azoxymethane model of colon cancer. *BMC Cancer* 2018;**18**:814. <https://doi.org/10.1186/s12885-018-4709-7>
 111. Hashemzahi M, Rahmani F, Khoshaklagh M, Avan A, Asgharzadeh F, Barneh F, et al. Angiotensin receptor blocker losartan inhibits tumor growth of colorectal cancer. *EXCLI J* 2021;**20**:506–21. <https://doi.org/10.17179/excli2020-3083>
 112. Oh E, Kim JY, Cho Y, An H, Lee N, Jo H, et al. Overexpression of angiotensin II type 1 receptor in breast cancer cells induces epithelial-mesenchymal transition and promotes tumor growth and angiogenesis. *Biochim Biophys Acta* 2016;**1863**:1071–81. <https://doi.org/10.1016/j.bbmr.2016.03.010>
 113. Saber S, Mahmoud AAA, Goda R, Helal NS, El-Ahwany E, Abdelghany RH. Perindopril, fosinopril and losartan inhibited the progression of diethylnitrosamine-induced

- hepatocellular carcinoma in mice via the inactivation of nuclear transcription factor kappa-B. *Toxicol Lett* 2018;**295**:32–40. <https://doi.org/10.1016/j.toxlet.2018.05.036>
114. Rhodes DR, Ateeq B, Cao Q, Tomlins SA, Mehra R, Laxman B, et al. AGTR1 overexpression defines a subset of breast cancer and confers sensitivity to losartan, an AGTR1 antagonist. *Proc Natl Acad Sci U S A* 2009;**106**:10284–9. <https://doi.org/10.1073/pnas.09003511106>
 115. Xia T, He Q, Shi K, Wang Y, Yu Q, Zhang L, et al. Losartan loaded liposomes improve the antitumor efficacy of liposomal paclitaxel modified with PH sensitive peptides by inhibition of collagen in breast cancer. *Pharm Dev Technol* 2018;**23**:13–21. <https://doi.org/10.1080/10837450.2016.1265553>
 116. Hu C, Liu X, Ran W, Meng J, Zhai Y, Zhang P, et al. Regulating cancer associated fibroblasts with losartan-loaded injectable peptide hydrogel to potentiate chemotherapy in inhibiting growth and lung metastasis of triple negative breast cancer. *Biomaterials* 2017;**144**:60–72. <https://doi.org/10.1016/j.biomaterials.2017.08.009>
 117. Redondo-Müller MA, Stevanovic-Walker M, Barker S, Puddefoot JR, Vinson GP. Anti-Cancer actions of a recombinant antibody (R6313/G2) against the angiotensin II AT1-Receptor. *Endocr Relat Cancer* 2008;**15**:277–88. <https://doi.org/10.1677/ERC-07-0068>
 118. Kosugi M, Miyajima A, Kikuchi E, Horiguchi Y, Murai M. Angiotensin II type 1 receptor antagonist candesartan as an angiogenic inhibitor in a Xenograft model of bladder cancer. *Clin Cancer Res* 2006;**12**:2888–93. <https://doi.org/10.1158/1078-0432.CCR-05-2213>
 119. Kosugi M, Miyajima A, Kikuchi E, Kosaka T, Horiguchi Y, Murai M. Effect of angiotensin II type 1 receptor antagonist on tumor growth and angiogenesis in a Xenograft model of human bladder cancer. *Hum Cell* 2007;**20**:1–9. <https://doi.org/10.1111/j.1749-0774.2007.00025.x>
 120. Wilisch-Neumann A, Pachow D, Wallesch M, Petermann A, Böhmer FD, Kirches E, et al. Re-Evaluation of cytostatic therapies for meningiomas in vitro. *J Cancer Res Clin Oncol* 2014;**140**:1343–52. <https://doi.org/10.1007/s00432-014-1683-6>
 121. Funao K, Matsuyama M, Kawahito Y, Sano H, Chargui J, Touraine J-L, et al. Telmisartan as a peroxisome proliferator-activated receptor- γ ligand is a new target in the treatment of human renal cell carcinoma. *Mol Med Rep* 2009;**2**:193–8. <https://doi.org/10.3892/mmr.000000083>
 122. Olschewski DN, Hofschneider V, Nielsen N, Seidler DG, Schwab A, Stock C. The angiotensin II type 1 receptor antagonist losartan affects NHE1-dependent melanoma cell behavior. *Cell Physiol Biochem* 2018;**45**:2560–76. <https://doi.org/10.1159/000488274>
 123. Martínez VR, Aguirre MV, Todaro JS, Piro OE, Echeverría GA, Naso LG, et al. Interaction of Zn with losartan. Activation of intrinsic apoptotic signaling pathway in lung cancer cells and effects on alkaline and acid phosphatases. *Biol Trace Elem Res* 2018;**186**:413–29. <https://doi.org/10.1007/s12011-018-1334-x>
 124. Fujihara S, Morishita A, Ogawa K, Tadokoro T, Chiyo T, Kato K, et al. The angiotensin II type 1 receptor antagonist telmisartan inhibits cell proliferation and tumor growth of esophageal adenocarcinoma via the AMPK α /mTOR pathway in vitro and in vivo. *Oncotarget* 2017;**8**:8536–49. <https://doi.org/10.18632/oncotarget.14345>
 125. Oura K, Tadokoro T, Fujihara S, Morishita A, Chiyo T, Samukawa E, et al. Telmisartan inhibits hepatocellular carcinoma cell proliferation in vitro by inducing cell cycle arrest. *Oncol Rep* 2017;**38**:2825–35. <https://doi.org/10.3892/or.2017.5977>
 126. Kobara H, Fujihara S, Iwama H, Matsui T, Fujimori A, Chiyo T, et al. Antihypertensive drug telmisartan inhibits cell proliferation of gastrointestinal stromal tumor cells in vitro. *Mol Med Rep* 2020;**22**:1063–71. <https://doi.org/10.3892/mmr.2020.11144>
 127. Woo Y, Jung Y. Angiotensin II receptor blockers induce autophagy in prostate cancer cells. *Oncol Lett* 2017;**13**:3579–85. <https://doi.org/10.3892/ol.2017.5872>
 128. Leung W-H, Vong QP, Lin W, Janke L, Chen T, Leung W. Modulation of NKG2D ligand expression and metastasis in tumors by spironolactone via RXR γ activation. *J Exp Med* 2013;**210**:2675–92. <https://doi.org/10.1084/jem.20122292>
 129. Shahar OD, Kalousi A, Eini L, Fisher B, Weiss A, Darr J, et al. A high-throughput chemical screen with FDA approved drugs reveals that the antihypertensive drug spironolactone impairs cancer cell survival by inhibiting homology directed repair. *Nucleic Acids Res* 2014;**42**:5689–701. <https://doi.org/10.1093/nar/gku217>
 130. Gold A, Eini L, Nissim-Rafinia M, Viner R, Ezer S, Erez K, et al. Spironolactone inhibits the growth of cancer stem cells by impairing DNA damage response. *Oncogene* 2019;**38**:3103–18. <https://doi.org/10.1038/s41388-018-0654-9>
 131. Kaji K, Yoshiji H, Kitade M, Ikenaka Y, Noguchi R, Shirai Y, et al. Selective aldosterone blocker, eplerenone, attenuates hepatocellular carcinoma growth and angiogenesis in mice. *Hepatal Res* 2010;**40**:540–9. <https://doi.org/10.1111/j.1872-034X.2010.00636.x>
 132. Feldman RD, Ding Q, Hussain Y, Limbird LE, Pickering JG, Gros R. Aldosterone mediates metastatic spread of renal cancer via the G protein-coupled estrogen receptor (GPER). *FASEB J* 2016;**30**:2086–96. <https://doi.org/10.1096/fj.15-275552>
 133. Isoe A, Takeda T, Wakabayashi A, Tsujii K, Li B, Sakata M, et al. Aldosterone stimulates the proliferation of uterine leiomyoma cells. *Gynecol Endocrinol* 2010;**26**:372–7. <https://doi.org/10.3109/09513590903511521>
 134. Lee HJ, Rho J, Gui SR, Kim MK, Lee YK, Lee YS, et al. Effect of aldosterone on the amplification of oncolytic vaccinia virus in human cancer lines. *Korean J Hepatol* 2011;**17**:213–9. <https://doi.org/10.3350/kjhep.2011.17.3.213>
 135. King S, Bray S, Galbraith S, Christie L, Fleming S. Evidence for aldosterone-dependent growth of renal cell carcinoma. *Int J Exp Pathol* 2014;**95**:244–50. <https://doi.org/10.1111/iep.12074>
 136. Fidrus E, Hegedűs C, Janka EA, Paragh G, Emri G, Remenyik É. Inhibitors of nucleotide excision repair decrease UVB-induced mutagenesis-an in vitro study. *Int J Mol Sci* 2021;**22**:1638. <https://doi.org/10.3390/ijms22041638>
 137. Sanomachi T, Suzuki S, Togashi K, Sugai A, Seino S, Okada M, et al. Spironolactone, a classic potassium-sparing diuretic, reduces survivin expression and chemosensitizes cancer cells to non-DNA-damaging anticancer drugs. *Cancers (Basel)* 2019;**11**:1550. <https://doi.org/10.3390/cancers11101550>
 138. Nogami H, Hiraoka Y, Aiso S. Estradiol and corticosterone stimulate the proliferation of a GH cell line, MtT/S: proliferation of growth hormone cells. *Growth Horm IGF Res* 2016;**29**:33–8. <https://doi.org/10.1016/j.ghir.2016.03.006>
 139. Aldaz P, Fernández-Celis A, López-Andrés N, Arozarena I. Novel insights into the role of the mineralocorticoid receptor in human glioblastoma. *Int J Mol Sci* 2021;**22**:11656. <https://doi.org/10.3390/ijms222111656>
 140. Sipahi I, Debanne SM, Rowland DY, Simon DI, Fang JC. Angiotensin-Receptor blockade and risk of cancer: meta-analysis of randomised controlled trials. *Lancet Oncol* 2010;**11**:627–36. [https://doi.org/10.1016/S1470-2045\(10\)70106-6](https://doi.org/10.1016/S1470-2045(10)70106-6)
 141. Pasternak B, Svanström H, Callréus T, Melbye M, Hviid A. Use of angiotensin receptor blockers and the risk of cancer. *Circulation* 2011;**123**:1729–36. <https://doi.org/10.1161/CIRCULATIONAHA.110.007336>
 142. Datzmann T, Fuchs S, Andree D, Hohenstein B, Schmitt J, Schindler C. Systematic review and meta-analysis of randomised controlled clinical trial evidence refutes relationship between pharmacotherapy with angiotensin-receptor blockers and an increased risk of cancer. *Eur J Intern Med* 2019;**64**:1–9. <https://doi.org/10.1016/j.ejim.2019.04.019>
 143. Shen J, Huang Y-M, Wang M, Hong X-Z, Song X-N, Zou X, et al. Renin-Angiotensin system blockade for the risk of cancer and death. *J Renin Angiotensin Aldosterone Syst* 2016;**17**:147032031665667. <https://doi.org/10.1177/1470320316656679>
 144. Sipahi I, Chou J, Mishra P, Debanne SM, Simon DI, Fang JC. Meta-Analysis of randomized controlled trials on effect of angiotensin-converting enzyme inhibitors on cancer risk. *Am J Cardiol* 2011;**108**:294–301. <https://doi.org/10.1016/j.amjcard.2011.03.038>
 145. Coleman CI, Baker WL, Kluger J, White CM. Antihypertensive medication and their impact on cancer incidence: a mixed treatment comparison meta-analysis of randomized controlled trials. *J Hypertens* 2008;**26**:622–9. <https://doi.org/10.1097/HJH.0b013e3282f3ef5e>
 146. Cao L, Zhang S, Jia C-M, He W, Wu L-T, Li Y-Q, et al. Antihypertensive drugs use and the risk of prostate cancer: a meta-analysis of 21 observational studies. *BMC Urol* 2018;**18**:17. <https://doi.org/10.1186/s12894-018-0318-7>
 147. Batais M, Almigbal T, Alotaibi K, Alodhayani A, Alkhashail A, Altheaby A, et al. Angiotensin converting enzyme inhibitors and risk of lung cancer: a systematic review and meta-analysis. *Medicine (Baltimore)* 2021;**100**:e25714. <https://doi.org/10.1097/MD.00000000000025714>
 148. Yoon C, Yang H-S, Jeon I, Chang Y, Park SM. Use of angiotensin-converting-enzyme inhibitors or angiotensin-receptor blockers and cancer risk: a meta-analysis of observational studies. *CMAJ* 2011;**183**:E1073–84. <https://doi.org/10.1503/cmaj.101497>
 149. Dai Y-N, Wang J-H, Zhu J-Z, Lin J-Q, Yu C-H, Li Y-M. Angiotensin-Converting enzyme inhibitors/angiotensin receptor blockers therapy and colorectal cancer: a systematic review and meta-analysis. *Cancer Causes Control* 2015;**26**:1245–55. <https://doi.org/10.1007/s10552-015-0617-1>
 150. Mao Y, Xu X, Wang X, Zheng X, Xie L. Is angiotensin-converting enzyme inhibitors/angiotensin receptor blockers therapy protective against prostate cancer? *Oncotarget* 2016;**7**:6765–73. <https://doi.org/10.18632/oncotarget.6837>
 151. Bommarreddy K, Hamade H, Lopez-Olivo MA, Wehner M, Tosh T, Barbieri JS. Association of spironolactone use with risk of cancer: a systematic review and meta-analysis. *JAMA Dermatol* 2022;**158**:275–82. <https://doi.org/10.1001/jamadermatol.2021.5866>
 152. Zhou Q, Chen D-S, Xin L, Zhou L-Q, Zhang H-T, Liu L, et al. The renin-angiotensin system blockers and survival in digestive system malignancies: a systematic review and meta-analysis. *Medicine (Baltimore)* 2020;**99**:e19075. <https://doi.org/10.1097/MD.00000000000019075>
 153. Sun H, Li T, Zhuang R, Cai W, Zheng Y. Do renin-angiotensin system inhibitors influence the recurrence, metastasis, and survival in cancer patients? *Med* 2017;**96**:e6394. <https://doi.org/10.1097/md.0000000000006394>
 154. Chen X, Yi C-H, Ya K-G. Renin-Angiotensin system inhibitor use and colorectal cancer risk and mortality: a dose-response meta analysis. *J Renin Angiotensin Aldosterone Syst* 2020;**21**:1470320319895646. <https://doi.org/10.1177/1470320319895646>
 155. Asgharzadeh F, Hashemzahi M, Moradi-Marjaneh R, Hassanian SM, Ferns GA, Khazaei M, et al. Angiotensin-Converting enzyme inhibitors and angiotensin receptor blockers as therapeutic options in the treatment of renal cancer: a meta-analysis. *Life Sci* 2020;**242**:117181. <https://doi.org/10.1016/j.lfs.2019.117181>
 156. Song T, Choi CH, Kim MK, Kim M-L, Yun BS, Seong SJ. The effect of angiotensin system inhibitors (angiotensin-converting enzyme inhibitors or angiotensin receptor blockers) on cancer recurrence and survival: a meta-analysis. *Eur J Cancer Prev* 2017;**26**:78–85. <https://doi.org/10.1097/CEJ.0000000000000269>

157. Zhao Y-T, Li P-Y, Zhang J-Q, Wang L, Yi Z. Angiotensin II receptor blockers and cancer risk: a meta-analysis of randomized controlled trials. *Medicine (Baltimore)* 2016;**95**:e3600. <https://doi.org/10.1097/MD.00000000000003600>
158. Myhre PL, Vaduganathan M, Claggett B, Packer M, Desai AS, Rouleau JL, et al. B-Type natriuretic peptide during treatment with sacubitril/valsartan: the PARADIGM-HF trial. *J Am Coll Cardiol* 2019;**73**:1264–72. doi:<https://doi.org/10.1016/j.jacc.2019.01.018>
159. Murphy SP, Prescott MF, Camacho A, Iyer SR, Maisel AS, Felker GM, et al. Atrial natriuretic peptide and treatment with sacubitril/valsartan in heart failure with reduced ejection fraction. *JACC Heart Fail* 2021;**9**:127–36. <https://doi.org/10.1016/j.jchf.2020.09.013>
160. Desai AS, McMurray JJV, Packer M, Swedberg K, Rouleau JL, Chen F, et al. Effect of the angiotensin-receptor-neprilysin inhibitor LCZ696 compared with enalapril on mode of death in heart failure patients. *Eur Heart J* 2015;**36**:1990–7. <https://doi.org/10.1093/eurheartj/ehv186>
161. Stolfo D, Lund LH, Sinagra G, Lindberg F, Dahlström U, Rosano G, et al. Heart failure pharmacological treatments and outcomes in heart failure with mildly reduced ejection fraction. *Eur Heart J Cardiovasc Pharmacother* 2023;**9**:526–35. <https://doi.org/10.1093/ehjcvp/pvad036>
162. Vesely BA, McAfee Q, Gower WR Jr, Vesely DL. Four peptides decrease the number of human pancreatic adenocarcinoma cells. *Eur J Clin Invest* 2003;**33**:998–1005. <https://doi.org/10.1046/j.1365-2362.2003.01262.x>
163. Vesely BA, Fitz SR, Gower WR Jr, Vesely DL. Vessel dilator: most potent of the atrial natriuretic peptides in decreasing the number and DNA synthesis of human squamous lung cancer cells. *Cancer Lett* 2006;**233**:226–31. <https://doi.org/10.1016/j.canlet.2005.03.024>
164. Vesely BA, Song S, Sanchez-Ramos J, Fitz SR, Alli AA, Solivan SM, et al. Five cardiac hormones decrease the number of human small-cell lung cancer cells. *Eur J Clin Invest* 2005;**35**:388–98. <https://doi.org/10.1111/j.1365-2362.2005.01501.x>
165. Vesely DL, Clark LC, Garces AH, McAfee QW, Soto J, Gower WR Jr. Novel therapeutic approach for cancer using four cardiovascular hormones. *Eur J Clin Invest* 2004;**34**:674–82. doi:<https://doi.org/10.1111/j.1365-2362.2004.01402.x>
166. Ohsaki Y, Gross AJ, Le PT, Oie H, Johnson BE. Human small cell lung cancer cells produce brain natriuretic peptide. *Oncology* 1999;**56**:155–9. <https://doi.org/10.1159/000011957>
167. Wigle DA, Campling BG, Sarda IR, Shin SH, Watson JD, Frater Y, et al. ANP secretion from small cell lung cancer cell lines: a potential model of ANP release. *Am J Physiol Circ Physiol* 1995;**268**:H1869–74. <https://doi.org/10.1152/ajpheart.1995.268.5.H1869>
168. Pavo N, Prausmüller S, Bartko P, Goliash G, Hülsmann M. Neprilysin as a biomarker: challenges and opportunities. *Card Fail Rev* 2020;**6**:e23. <https://doi.org/10.15420/cfr.2019.21>
169. Scafoglio C, Hirayama BA, Kepe V, Liu J, Ghezzi C, Satyamurthy N, et al. Functional expression of sodium-glucose transporters in cancer. *Proc Natl Acad Sci U S A* 2015;**112**:E4111–9. <https://doi.org/10.1073/pnas.1511698112>
170. Dutka M, Bobiński R, Francuz T, Garczorz W, Zimmer K, Ilczak T, et al. SGLT-2 Inhibitors in cancer treatment; mechanisms of action and emerging new perspectives. *Cancers (Basel)* 2022;**14**:5811. <https://doi.org/10.3390/cancers14235811>
171. Zhou J, Zhu J, Yu S-J, Ma H-L, Chen J, Ding X-F, et al. Sodium-Glucose co-transporter-2 (SGLT-2) inhibition reduces glucose uptake to induce breast cancer cell growth arrest through AMPK/MTOR pathway. *Biomed Pharmacother* 2020;**132**:110821. doi:<https://doi.org/10.1016/j.biopha.2020.110821>
172. Nasiri AR, Rodrigues MR, Li Z, Leitner BP, Perry RJ. SGLT2 inhibition slows tumor growth in mice by reversing hyperinsulinemia. *Cancer Metab* 2019;**7**:10. <https://doi.org/10.1186/s40170-019-0203-1>
173. Villani LA, Smith BK, Marcinko K, Ford RJ, Broadfield LA, Green AE, et al. The diabetes medication canagliflozin reduces cancer cell proliferation by inhibiting mitochondrial Complex-I supported respiration. *Mol Metab* 2016;**5**:1048–56. doi:<https://doi.org/10.1016/j.molmet.2016.08.014>
174. Kaji K, Nishimura N, Seki K, Sato S, Saikawa S, Nakanishi K, et al. Sodium glucose co-transporter 2 inhibitor canagliflozin attenuates liver cancer cell growth and angiogenic activity by inhibiting glucose uptake. *Int J Cancer* 2018;**142**:1712–22. doi:<https://doi.org/10.1002/ijc.31193>
175. Kabel AM, Arab HH, Abd Elmaaboud MA. Effect of dapagliflozin and/or L-arginine on solid tumor model in mice: the interaction between nitric oxide, transforming growth factor-Beta 1, autophagy, and apoptosis. *Fundam Clin Pharmacol* 2021;**35**:968–78. <https://doi.org/10.1111/fcp.12661>
176. Xie Z, Wang F, Lin L, Duan S, Liu X, Li X, et al. An SGLT2 inhibitor modulates SHH expression by activating AMPK to inhibit the migration and induce the apoptosis of cervical carcinoma cells. *Cancer Lett* 2020;**495**:200–10. doi:<https://doi.org/10.1016/j.canlet.2020.09.005>
177. Ren D, Sun Y, Zhang D, Li D, Liu Z, Jin X, et al. SGLT2 promotes pancreatic cancer progression by activating the hippo signaling pathway via the HnRNPK-YAP1 axis. *Cancer Lett* 2021;**519**:277–88. doi:<https://doi.org/10.1016/j.canlet.2021.07.035>
178. Korfhage J, Skinner ME, Basu J, Greenson JK, Miller RA, Lombard DB. Canagliflozin increases intestinal adenoma burden in female ApcMin/+ mice. *J Gerontol A Biol Sci Med Sci* 2022;**77**:215–20. <https://doi.org/10.1093/gerona/glab254>
179. Shoda K, Tsuji S, Nakamura S, Egashira Y, Enomoto Y, Nakayama N, et al. Canagliflozin inhibits glioblastoma growth and proliferation by activating AMPK. *Cell Mol Neurobiol* 2022;**43**:879–92. <https://doi.org/10.1007/s10571-022-01221-8>
180. Xu D, Zhou Y, Xie X, He L, Ding J, Pang S, et al. Inhibitory effects of canagliflozin on pancreatic cancer are mediated via the downregulation of glucose transporter-1 and lactate dehydrogenase A. *Int J Oncol* 2020;**57**:1223–33. <https://doi.org/10.3892/ijo.2020.5120>
181. Nakano D, Kawaguchi T, Iwamoto H, Hayakawa M, Koga H, Torimura T. Effects of canagliflozin on growth and metabolic reprogramming in hepatocellular carcinoma cells: multi-omics analysis of metabolomics and absolute quantification proteomics (IMPAQT). *PLoS One* 2020;**15**:e0232283. <https://doi.org/10.1371/journal.pone.0232283>
182. Papadopoli D, Uchenunu O, Palia R, Chekkal N, Hulea L, Topisirovic I, et al. Perturbations of cancer cell metabolism by the antidiabetic drug canagliflozin. *Neoplasia* 2021;**23**:391–9. <https://doi.org/10.1016/j.neo.2021.02.003>
183. Yamamoto L, Yamashita S, Nomiya T, Kawanami T, Hamaguchi Y, Shigeoka T, et al. Sodium-Glucose cotransporter 2 inhibitor canagliflozin attenuates lung cancer cell proliferation in vitro. *Diabetol Int* 2021;**12**:389–98. <https://doi.org/10.1007/s13340-021-00494-6>
184. Wang L, Liu M, Yin F, Wang Y, Li X, Wu Y, et al. Trilobatin, a novel SGLT1/2 inhibitor, selectively induces the proliferation of human hepatoblastoma cells. *Molecules* 2019;**24**:3390. <https://doi.org/10.3390/molecules24183390>
185. Hung M-H, Chen Y-L, Chen L-J, Chu P-Y, Hsieh F-S, Tsai M-H, et al. Canagliflozin inhibits growth of hepatocellular carcinoma via blocking glucose-influx-induced β -catenin activation. *Cell Death Dis* 2019;**10**:420. <https://doi.org/10.1038/s41419-019-1646-6>
186. Madonna R, Barachini S, Moscato S, Ippolito C, Mattii L, Lenzi C, et al. Sodium-Glucose cotransporter type 2 inhibitors prevent ponatinib-induced endothelial senescence and dysfunction: a potential rescue strategy. *Vascul Pharmacol* 2022;**142**:106949. doi:<https://doi.org/10.1016/j.vph.2021.106949>
187. Lin H-W, Tseng C-H. A review on the relationship between SGLT2 inhibitors and cancer. *Int J Endocrinol* 2014;**2014**:719578. <https://doi.org/10.1155/2014/719578>
188. Benedetti R, Benincasa G, Glass K, Chianese U, Vietri MT, Congi R, et al. Effects of novel SGLT2 inhibitors on cancer incidence in hyperglycemic patients: a meta-analysis of randomized clinical trials. *Pharmacol Res* 2022;**175**:106039. <https://doi.org/10.1016/j.phrs.2021.106039>
189. Shi N, Shi Y, Xu J, Si Y, Yang T, Zhang M, et al. SGLT-2i and risk of malignancy in type 2 diabetes: a meta-analysis of randomized controlled trials. *Front Public Heal* 2021;**9**:668368. <https://doi.org/10.3389/fpubh.2021.668368>
190. Tang H, Dai Q, Shi W, Zhai S, Song Y, Han J. SGLT2 inhibitors and risk of cancer in type 2 diabetes: a systematic review and meta-analysis of randomised controlled trials. *Diabetologia* 2017;**60**:1862–72. <https://doi.org/10.1007/s00125-017-4370-8>
191. Dicembrini I, Nreu B, Mannucci E, Monami M. Sodium-Glucose co-transporter-2 (SGLT-2) inhibitors and cancer: a meta-analysis of randomized controlled trials. *Diabetes Obes Metab* 2019;**21**:1871–7. <https://doi.org/10.1111/dom.13745>
192. Cui H, Wang Y, Yang S, He G, Jiang Z, Gang X, et al. Antidiabetic medications and the risk of prostate cancer in patients with diabetes Mellitus: a systematic review and meta-analysis. *Pharmacol Res* 2022;**177**:106094. <https://doi.org/10.1016/j.phrs.2022.106094>
193. Tang H, Yang K, Song Y, Han J. Meta-Analysis of the association between sodium-glucose co-transporter-2 inhibitors and risk of skin cancer among patients with type 2 diabetes. *Diabetes Obes Metab* 2018;**20**:2919–24. <https://doi.org/10.1111/dom.13474>
194. Chou J-C, Li J-H, Chen C-C, Chen C-W, Lin H, Wang PS. Inhibitory effects of digoxin and digitoxin on cell growth in human ovarian cancer cell line SKOV-3. *Integr Cancer Ther* 2021;**20**:15347354211002662. <https://doi.org/10.1177/15347354211002662>
195. Hou Y-Q, Wang Y-Y, Wang X-C, Liu Y, Zhang C-Z, Chen Z-S, et al. Multifaceted anti-colorectal tumor effect of digoxin on HCT8 and SW620 cells in vitro. *Gastroenterol Rep* 2020;**8**:465–75. <https://doi.org/10.1093/gastro/goaa076>
196. Deng K, Shen J, Wang W, Li M, Li H, Chen C, et al. Sodium chloride (NaCl) potentiates digoxin-induced anti-tumor activity in small cell lung cancer. *Cancer Biol Ther* 2019;**20**:52–64. <https://doi.org/10.1080/15384047.2018.1504723>
197. Joseph JV, Conroy S, Pavlov K, Sontakke P, Tomar T, Eggens-Meijer E, et al. Hypoxia enhances migration and invasion in glioblastoma by promoting a mesenchymal shift mediated by the HIF1 α -ZEB1 axis. *Cancer Lett* 2015;**359**:107–16. <https://doi.org/10.1016/j.canlet.2015.01.010>
198. Zhang X-H, Wang X-Y, Zhou Z-W, Bai H, Shi L, Yang Y-X, et al. The combination of digoxin and GSK2606414 exerts synergistic anticancer activity against leukemia in vitro and in vivo. *BioFactors* 2017;**43**:812–20. doi:<https://doi.org/10.1002/biof.1380>
199. Wang Z, Zheng M, Li Z, Li R, Jia L, Xiong X, et al. Cardiac glycosides inhibit P53 synthesis by a mechanism relieved by src or MAPK inhibition. *Cancer Res* 2009;**69**:6556–64. <https://doi.org/10.1158/0008-5472.CAN-09-0891>
200. Wang Y, Ma Q, Zhang S, Liu H, Zhao B, Du B, et al. Digoxin enhances the anticancer effect on non-small cell lung cancer while reducing the cardiotoxicity of Adriamycin. *Front Pharmacol* 2020;**11**:186. <https://doi.org/10.3389/fphar.2020.00186>

201. Glass OK, Bowie M, Fuller J, Darr D, Usary J, Boss K, et al. Differential response to exercise in claudin-low breast cancer. *Oncotarget* 2017;**8**:100989–1004. <https://doi.org/10.18632/oncotarget.21054>
202. Crezee T, Tesselar MH, Nagarajah J, Corver WE, Morreau J, Pritchard C, et al. Digoxin treatment reactivates in vivo radioactive iodide uptake and correlates with favorable clinical outcome in non-medullary thyroid cancer. *Cell Oncol* 2021;**44**:611–25. <https://doi.org/10.1007/s13402-021-00588-y>
203. Beheshti Zavareh R, Lau KS, Hurren R, Datti A, Ashline DJ, Gronda M, et al. Inhibition of the sodium/potassium ATPase impairs N-glycan expression and function. *Cancer Res* 2008;**68**:6688–97. <https://doi.org/10.1158/0008-5472.CAN-07-6833>
204. Ahern TP, Tamimi RM, Rosner BA, Hankinson SE. Digoxin use and risk of invasive breast cancer: evidence from the nurses' health study and meta-analysis. *Breast Cancer Res Treat* 2014;**144**:427–35. <https://doi.org/10.1007/s10549-014-2886-x>
205. Osman MH, Farrag E, Selim M, Osman MS, Hasanine A, Selim A. Cardiac glycosides use and the risk and mortality of cancer; systematic review and meta-analysis of observational studies. *PLoS One* 2017;**12**:e0178611. <https://doi.org/10.1371/journal.pone.0178611>
206. Panet R, Marcus M, Atlan H. Overexpression of the Na⁺/K⁺/Cl⁻ cotransporter gene induces cell proliferation and phenotypic transformation in mouse fibroblasts. *J Cell Physiol* 2000;**182**:109–18. doi:3.0.CO;202-A" >[https://doi.org/10.1002/\(SICI\)1097-4652\(200001\)182:1<109::AID-JCP12>3.0.CO;2-A](https://doi.org/10.1002/(SICI)1097-4652(200001)182:1<109::AID-JCP12>3.0.CO;2-A)
207. Iwamoto LM, Fujiwara N, Nakamura KT, Wada RK. Na-K-2Cl cotransporter inhibition impairs human lung cellular proliferation. *Am J Physiol Cell Mol Physiol* 2004;**287**:L510–4. <https://doi.org/10.1152/ajplung.00021.2004>
208. Shiozaki A, Nako Y, Ichikawa D, Konishi H, Komatsu S, Kubota T, et al. Role of the Na⁺/K⁺/2Cl⁻ cotransporter NKCC1 in cell cycle progression in human esophageal squamous cell carcinoma. *World J Gastroenterol* 2014;**20**:6844–59. <https://doi.org/10.3748/wjg.v20.i22.6844>
209. Liu P, McMenamin ÚC, Spence AD, Johnston BT, Coleman HG, Cardwell CR. Furosemide use and survival in patients with esophageal or gastric cancer: a population-based cohort study. *BMC Cancer* 2019;**19**:1017. <https://doi.org/10.1186/s12885-019-6242-8>
210. Shin D, Lee ES, Kim J, Guerra L, Naik D, Prida X. Association between the use of thiazide diuretics and the risk of skin cancers: a meta-analysis of observational studies. *J Clin Med Res* 2019;**11**:247–55. <https://doi.org/10.14740/jocmr.3744>
211. Heisel AGU, Vuurboom MD, Daams JG, de Rie MA, Vogt L, van den Born B-JH, et al. The use of specific antihypertensive medication and skin cancer risk: a systematic review of the literature and meta-analysis. *Vascul Pharmacol* 2023;**150**:107173. doi: <https://doi.org/10.1016/j.vph.2023.107173>
212. Sayour NV, Brenner GB, Makkos A, Kiss B, Kovácsházi C, Gergely TG, et al. Cardioprotective efficacy of limb remote ischaemic preconditioning in rats: discrepancy between a meta-analysis and a three-centre in vivo study. *Cardiovasc Res* 2023;**119**:1336–51. <https://doi.org/10.1093/cvr/cvad024>
213. Begley CG, Ioannidis JPA. Reproducibility in science. *Circ Res* 2015;**116**:116–26. <https://doi.org/10.1161/CIRCRESAHA.114.303819>
214. Avraham S, Abu-Sharki S, Shofti R, Haas T, Korin B, Kalfon R, et al. Early cardiac remodeling promotes tumor growth and metastasis. *Circulation* 2020;**142**:670–83. <https://doi.org/10.1161/CIRCULATIONAHA.120.046471>
215. Meijers WC, Maglione M, Bakker SJL, Oberhuber R, Kieneker LM, de Jong S, et al. Heart failure stimulates tumor growth by circulating factors. *Circulation* 2018;**138**:678–91. <https://doi.org/10.1161/CIRCULATIONAHA.117.030816>
216. Koelwyn GJ, Newman AAC, Afonso MS, van Solingen C, Corr EM, Brown EJ, et al. Myocardial infarction accelerates breast cancer via innate immune reprogramming. *Nat Med* 2020;**26**:1452–8. <https://doi.org/10.1038/s41591-020-0964-7>
217. de Wit S, Aboumsallem JP, Shi C, Schouten EM, Bracun V, Meijers WC, et al. Pressure overload-induced cardiac hypertrophy stimulates tumor growth in tumor-prone ApcMin mice. *Circ Heart Fail* 2023;**0**:e010740. <https://doi.org/10.1161/CIRCHEARTFAILURE.123.010740>
218. Sayour NV, Tóth VÉ, Nagy RN, Vörös I, Gergely TG, Onódi Z, et al. Droplet digital PCR is a novel screening method identifying potential cardiac G-protein-coupled receptors as candidate pharmacological targets in a rat model of pressure-overload-induced cardiac dysfunction. *Int J Mol Sci* 2023;**24**:13826. <https://doi.org/10.3390/ijms241813826>
219. Dobbin SJH, Shen L, Petrie MC, Packer M, Solomon SD, McMurray JJV, et al. Characteristics and outcomes of patients with a history of cancer recruited to heart failure trials. *Eur J Heart Fail* 2023;**25**:488–96. doi:<https://doi.org/10.1002/ehf.2818>
220. Tini G, Bertero E, Signori A, Sormani MP, Maack C, De Boer RA, et al. Cancer mortality in trials of heart failure with reduced ejection fraction: a systematic review and meta-analysis. *J Am Heart Assoc* 2020;**9**:e016309. <https://doi.org/10.1161/JAHA.119.016309>
221. Lehmann LH, Heckmann MB, Bailly G, Finke D, Procureur A, Power JR, et al. Cardiomuscular biomarkers in the diagnosis and prognostication of immune checkpoint inhibitor myocarditis. *Circulation* 2023;**148**:473–86. <https://doi.org/10.1161/CIRCULATIONAHA.123.062405>
222. Rathore SS, Wang Y, Krumholz HM. Sex-Based differences in the effect of digoxin for the treatment of heart failure. *N Engl J Med* 2002;**347**:1403–11. <https://doi.org/10.1056/NEJMoa021266>
223. Davide S, Alicia U, Ola V, Anna S, Ljung FU, Rosano GMC, et al. Sex-Based differences in heart failure across the ejection fraction Spectrum. *JACC Heart Fail* 2019;**7**:505–15. <https://doi.org/10.1016/j.jchf.2019.03.011>
224. Ridker PM, Everett BM, Thuren T, MacFadyen JG, Chang WH, Ballantyne C, et al. Antiinflammatory therapy with canakinumab for atherosclerotic disease. *N Engl J Med* 2017;**377**:1119–31. <https://doi.org/10.1056/NEJMoa1707914>
225. Everett BM, Cornel JH, Lainscak M, Anker SD, Abbate A, Thuren T, et al. Anti-inflammatory therapy with canakinumab for the prevention of hospitalization for heart failure. *Circulation* 2019;**139**:1289–99. <https://doi.org/10.1161/CIRCULATIONAHA.118.038010>
226. Ridker PM, MacFadyen JG, Thuren T, Everett BM, Libby P, Glynn RJ, et al. Effect of interleukin-1 β inhibition with canakinumab on incident lung cancer in patients with atherosclerosis: exploratory results from a randomised, double-blind, placebo-controlled trial. *Lancet* 2017;**390**:1833–42. [https://doi.org/10.1016/S0140-6736\(17\)32247-X](https://doi.org/10.1016/S0140-6736(17)32247-X)