

Streicker, D. et al. (2024) Developing transmissible vaccines for animal infectious diseases. Science, 384(6693), pp. 275-277. (doi: 10.1126/science.adn3231)



© The Authors 2024. Reproduced under a <u>Creative Commons Attribution 4.0</u> International License.

https://doi.org/10.1126/science.adn3231

https://eprints.gla.ac.uk/323783/

Deposited on: 3 April 2024

Enlighten – Research publications by members of the University of Glasgow https://eprints.gla.ac.uk 1

# Title: Developing transmissible vaccines for animal infectious diseases

2

Authors: Daniel G. Streicker<sup>1,2\*</sup>, Megan E. Griffiths<sup>1,2</sup>, Rustom Antia<sup>3</sup>, Laura Bergner<sup>1,2</sup>, Peter 3 Bowman<sup>4</sup><sup>+</sup>, Maria Vitoria dos Santos de Moraes<sup>5</sup>, Kevin Esvelt<sup>6</sup>, Mike Famulare<sup>7</sup>, Amy Gilbert<sup>8</sup>, 4 Biao He<sup>9</sup>, Michael A. Jarvis<sup>10,11,12</sup>, David A. Kennedy<sup>13</sup>, Jennifer Kuzma<sup>14</sup>, Carolyne Nasimiyu 5 Wanyonyi<sup>15</sup>, Christopher Remien<sup>16</sup>, Tonie Rocke<sup>17</sup>, Kyle Rosenke<sup>12</sup>, Courtney Schreiner<sup>18</sup>, Justin 6 Sheen<sup>19</sup>, David Simons<sup>20</sup>, Ivet A. Yordanova<sup>21</sup>, James J. Bull<sup>22</sup> and Scott L. Nuismer<sup>22\*</sup> 7 8 **Affiliations:** 9 <sup>1</sup>School of Biodiversity, One Health and Veterinary Medicine, College of Medical, Veterinary 10 11 and Life Sciences, University of Glasgow; Glasgow G12 8QQ, United Kingdom. 12 <sup>2</sup>MRC-University of Glasgow Centre for Virus Research; Glasgow G61 1QH, United Kingdom. <sup>3</sup> Department of Biology, Emory University; Atlanta, GA, 30322 United States of America. 13 <sup>4</sup> School of Veterinary Medicine, University of California-Davis; Davis, CA, 995616, United 14 15 States of America. 16 <sup>5</sup> Faculty of Veterinary Medicine and Animal Sciences, University of São Paulo; São Paulo, 17 05508-270, Brazil. <sup>6</sup>Media Laboratory, Massachusetts Institute of Technology; Cambridge, MA, 02139, United 18 States of America. 19 20 <sup>7</sup> Institute for Disease Modeling, Bill & Melinda Gates Foundation; Seattle, WA, 98109, United 21 States of America. 22 <sup>8</sup> United States Department of Agriculture, Animal and Plant Health Inspection Service, National 23 Wildlife Research Center; Fort Collins, CO, 80521, United States of America. <sup>9</sup> Department of Infectious Diseases, College of Veterinary Medicine, University of Georgia; 24 25 Athens, GA, 30602, United States of America <sup>10</sup> School of Biomedical Sciences, University of Plymouth; Devon, PL4 8AA, United Kingdom 26 27 <sup>11</sup> The Vaccine Group, Ltd.; Devon, PL6 6BU, United Kingdom <sup>12</sup> Laboratory of Virology, National Institute of Allergy and Infectious Diseases, National 28 29 Institutes of Health; Hamilton, MT, 59840, United States of America <sup>13</sup> Department of Biology and Center for Infectious Disease Dynamics, The Pennsylvania State 30 31 University; University Park, PA, 16802, United States of America 1

32	<sup>14</sup> School of Public and International Affairs and Genetic Engineering and Society Center, North
33	Carolina State University; Raleigh, NC, 27606 United States of America.
34	<sup>15</sup> Global Health Program, Washington State University; Nairobi, Kenya.
35	<sup>16</sup> Department of Mathematics and Statistical Science, University of Idaho; Moscow, ID 83844,
36	United States of America.
37	<sup>17</sup> United States Geological Survey, National Wildlife Health Center; Madison, Wisconsin,
38	53711, United States of America.
39	<sup>18</sup> Department of Ecology and Evolutionary Biology, University of Tennessee Knoxville,
40	Knoxville, TN, 37996 United States of America.
41	<sup>19</sup> Department of Ecology and Evolutionary Biology, Princeton University, Princeton, New
42	Jersey, 08544, United States of America.
43	<sup>20</sup> Centre for Emerging, Endemic and Exotic Diseases, The Royal Veterinary College; London
44	NW1 0TU, United Kingdom.
45	<sup>21</sup> Center for Biological Threats and Special Pathogens, Robert Koch Institute; Berlin, 13353,
46	Germany.
47	<sup>22</sup> Department of Biological Sciences, University of Idaho; Moscow, ID 83844, United States of
48	America.
49	
50	† Present address: Congressional Hunger Center and Land O'Lakes Venture 37, Nakuru, Kenya
51	
52	* Corresponding authors: Daniel G. Streicker ( <u>daniel.streicker@glasgow.ac.uk</u> ); Scott L.
53	Nuismer ( <u>snuismer@uidaho.edu</u> )
54	
55	
56	
57	
58	
59	
60	
61	
62	

### 63 Main text:

Many emerging and re-emerging pathogens originate from wildlife, but nearly all wild species are 64 unreachable using conventional vaccination, which requires capture of and vaccine administration 65 to individual animals. By enabling immunization at scales sufficient to interrupt pathogen 66 transmission, transmissible vaccines (TVs) that spread themselves through wildlife populations by 67 68 infectious processes could potentially transform management of otherwise intractable challenges 69 to public health, wildlife conservation, and animal welfare. However, generating TVs likely 70 requires modifying viruses that would be intended to spread in nature, raising concerns ranging from technical feasibility, to safety and security risks, to regulatory uncertainties (1, 2). We 71 72 propose a series of commitments and strategies for vaccine development, beginning with a priori 73 decisions on vaccine design and continuing through to stakeholder co-development (see the box), that we believe increase the likelihood that the potential risks of vaccine transmission are 74 75 outweighed by benefits to conservation, animal welfare, and zoonosis prevention.

76 The inability to control emerging pathogens at their source translates into mitigation strategies 77 focused on direct protection of humans or domestic animals, an approach that fails to curb the risks 78 and costs of recurring transmission between species (hereafter, spillover). Diseases threatening 79 wildlife health, either through recurrent spillover (e.g., Ebola in great apes) or following host shifts 80 and/or pathogen translocations (e.g., white nose syndrome [WNS] in bats), remain similarly uncontrollable by conventional approaches. Mass distribution of oral vaccines via baits has shown 81 82 that scalable vaccination of wildlife can protect human health and animal welfare; however, bait 83 delivery systems are incompatible with many wild species (3).

84 TVs have been proposed as a scalable, low-cost option to interrupt transmission within and to 85 otherwise unreachable wildlife (4). However, risks of vaccine transmission are well recognized 86 from theory and have been substantiated in conventional vaccines that transmit inadvertently 87 (Figure 1). Most notoriously, sustained transmission of the live attenuated oral polio vaccine 88 enabled reversion to its ancestral polio-causing phenotype. Although deliberate vaccine 89 transmission has only rarely been tested, a vaccine against rabbit hemorrhagic disease (RHD) did 90 explore the possibility using an attenuated myxomavirus-based vaccine (5). Although no ill effects 91 were reported prior to natural vaccine extinction, the myxomavirus used was not host specific and 92 had only a brief co-evolutionary history with the target rabbit species, making its long-term 93 evolutionary trajectory uncertain. Recent interest in TVs has been revitalized by accumulating

evidence that it may be possible to design vaccines that mitigate foreseeable risks while preserving
efficacy. Such TVs are currently being advanced in laboratories, but to our knowledge, none have
been released in any natural population.

97 The relative lack of substantive public discourse involving both proponents and critics of TVs has created a scientific landscape with conflicting definitions and immaterial evidence that is 98 99 unhelpful for policymakers, funders, and the organizations charged with oversight of the research 100 and development process. As a group of bioethicists, disease ecologists, evolutionary biologists, 101 immunologists, sociologists, and virologists, including both proponents and critics of TVs, we 102 appraised the potential ecological and societal risks arising from transmission of an engineered 103 viral vaccine (see supplementary materials). The commitments that arose are not intended to 104 establish dogma or legitimize the use of TVs but rather to serve as a conservative starting point which we expect will evolve with societal attitudes, scientific evidence, and technology. 105

106

## 107 INTRINSICALLY SAFE, BIOLOGICALLY COMPELLING VACCINE DESIGNS

Flexible vaccine designs are most easily accommodated using recombinant vaccines that consist of two parts engineered into one genome: a relatively benign animal virus (the vector) and a short genetic segment from the pathogen (the antigenic insert or transgene), which induces an immune response. The goal is to preserve the capacity for transmission between individuals, while adding the ability to immunize, thereby magnifying the vaccination coverage derived from each directly vaccinated individual.

114 As vaccine safety hinges predominately on the properties of the vector, we propose eligibility 115 criteria. First, vaccines derived from cross-species transfer (e.g., myxomavirus-based RHD vaccine) may spread unpredictably causing ecological disruption. New selective environments, 116 117 including the possibility of novel co-infections with recombination-compatible viruses, might also 118 promote evolution towards previously unobserved, harmful phenotypes (5). Vectors would 119 therefore need to be both isolated from and returned to their natural host species. Because 120 competition between TVs and their ancestral (wildtype) or descendant (reversion to non-vaccine strain) viruses may inhibit vaccine spread, vectors that can infect hosts with prior or concurrent 121 122 wildtype infections are desirable. Alternatively, competition with the wildtype may be overcome 123 by repeatedly introducing the vaccine or constructing it using locally rare or absent strains (6, 7).

124 Second, vaccines that cross species boundaries during transmission in nature present similar 125 risks to deliberate cross-species transfer. Vectors would therefore need to be host specific, as 126 demonstrated by representative surveys for cross-species infections in nature, co-evolutionary 127 analyses supporting host-virus co-speciation over host switching, laboratory studies of cellular 128 tropism, and animal inoculation studies. Ecologically plausible exposures in sympatric, non-target species (i.e., those that are not part of the planned vaccination campaign) would need to lead to 129 130 insufficient replication to cause clinical disease or vaccine transmission. Ecological plausibility 131 might be derived from local knowledge, expert opinion, and/or in silico predictions of 132 susceptibility. In cases where multiple host species independently maintain the pathogen and a 133 single viral vector infects these species, safety and efficacy studies should include all relevant 134 hosts.

Third, viruses that would require attenuation (reducing virulence) to align with management goals and stakeholder desires are excluded since perturbing the co-evolved virus-host equilibrium might select for a return to the undesirable ancestral state (fig. S1). Unlike reversion of attenuated vaccines, reversion of TVs to their ancestral phenotype creates no novel health or environmental risks because the ancestral virus naturally circulates in the same host species. This strategy also alleviates the potential concern that TVs could gain pathogenicity by recombining with wildtype strains (*8*).

Misuse of the knowledge acquired during the development of new technology is always a 142 143 concern. Consistent with the core ideology of exploiting natural traits of viruses as built-in safety 144 features, engineering of viral vectors would avoid modifications that increase host range, 145 pathogenicity, or transmissibility. More generally, any technology that could plausibly be harmful 146 if applied to a human-infecting virus should be avoided in TVs designed for animals. For instance, 147 discovering novel molecular mechanisms that augment spread or enhance evolutionary stability 148 might benefit vaccine coverage but could have malicious applications elsewhere. If increased 149 stability is required to reach management objectives, methods could be limited to transgene 150 identity, size, copy number, and placement (9). Alternatively, more intensive or efficient 151 deployment can increase coverage (10).

152

### 153 STAGED DEVELOPMENT WITH ESTABLISHED CHECKPOINTS

154 We believe the criteria described above maximize the safety of TVs without undermining their 155 potential efficacy (10,11). Nevertheless, unforeseeable issues may arise during the vaccine 156 development process which may prompt suspension of a TV's development. A staged 157 development process is needed for early identification and containment of emergent risks. 158 Specifically, TV development would advance from in vitro studies in laboratories, to in vivo animal 159 testing within appropriate biological containment, to limited trials in populations that are naturally 160 (e.g., islands, mountains) or experimentally (e.g., enclosures, semi-field systems) isolated (Figure 161 1). Following an Open Science approach, quantitative benchmarks for safety and efficacy would be defined in advance and transparently shared as checkpoints to continue or not with a given 162 163 vaccine candidate. Instability of recombinant TVs through silencing or purging of the transgene is 164 expected and detrimental to efficacy but acts advantageously as a natural self-limiting mechanism against uncontrolled spread. When technically possible, vaccines themselves should be staged, 165 166 with early experiments using vaccines expected to have a short evolutionary half-life, mitigating 167 risks of prolonged circulation of an undesirable prototype in the event of laboratory escape.

168 Accountable systems to monitor vaccine release, evolution, and spread will be critical throughout 169 the development process. These include re-sequencing of the vaccine to monitor evolutionary changes 170 and periodic *in vitro* monitoring of growth rate or cellular tropism. Since vaccinated animals possess 171 immunity only to pathogen proteins included within the antigenic insert, immunological monitoring 172 could differentiate previously infected and vaccinated animals. The potential for vaccines to create 173 secondary hazards, such as exposure to vehicles used in vaccine deployment (e.g., topical gels, baits, 174 aerosols), also needs to be considered and monitored when appropriate. Researchers should establish 175 contingency plans for foreseeable risks (noting that a contingency plan can include 'no action') and 176 implement appropriate management systems for timely responses to unforeseen events.

177

# 178 EQUITABLE PARTNERSHIPS WITH INTERNATIONAL GOVERNANCE

While the impossibility of individual consent prohibits consideration of TVs for human use, complex ethical issues around consent also arise for TV use in animals. Concerns and requirements around technology development, staged delivery timelines, and identification of any ecological ramifications of reducing pathogen circulation would require reciprocal engagement with relevant stakeholders, including government agencies that regulate vaccine use in animals, wildlife population managers, public health officials, non-government agencies, and affected communities 185 ('co-development'). Initiating this process at project inception and certainly before engineering of 186 vaccine prototypes benefits vaccine developers by identifying technical and community values-187 based constraints that would alter deployment or development targets (12). Communities affected by zoonotic spillover may desire rapid or geographically expanded TV deployment or, due to the 188 189 novelty of TVs, may alternatively focus on potential risks while overlooking benefits. Scientists 190 and communicators with expertise in managing expectations and identifying community 191 champions will play a key role by ensuring that information about vaccine performance or safety is accurately portrayed, thus empowering communities to help make decisions with free, prior, and 192 193 informed consent. Communication and engagement should also raise awareness of the potential 194 for discussions of TVs to reduce acceptance of conventional vaccines, thereby inadvertently 195 harming health.

As with any vaccine, TV development will be subject to existing local, national and international 196 regulations for scientific research, production and testing, environmental impacts, and to funders' 197 198 discretion. One motivation for TVs is to reduce the disproportionate burden of pathogen spillover from 199 wildlife in lower- and middle-income countries. It is therefore unavoidable that some developmental 200 stages for some TVs (e.g., contained field trials) would be undertaken in these countries, while other 201 stages (e.g., vaccine engineering and laboratory-contained animal trials) may be undertaken in 202 countries with more funding and infrastructure. As regulatory requirements also vary across countries, 203 stringent oversight as a shared, international responsibility underpins credibility, for example, requiring 204 ethical and biosafety practices approaching the most conservative standard among partner nations 205 involved. TVs developed to conserve wildlife may avoid the potential geographic mismatches between 206 TV use and development. Greater investment in this area could provide valuable proof of concept for TVs targeting zoonotic spillover. Regardless of management targets, equitable collaborations, wherein 207 208 risks taken and benefits gained are proportionate and undertaken by nationally diverse teams, are 209 warranted across developmental stages.

210

### 211 TOWARDS DEPLOYMENT

In principle, TVs are suited to well-studied host-pathogen systems where spillover from
established reservoir hosts is predictable, recurrent, and costly (e.g., rabies virus, Lassa fever virus,
Nipah virus, Marburg virus) or where low-cost, scalable interventions could reduce pathogen
threats to wildlife (e.g., WNS in bats, Ebola virus disease in non-human primates, retrovirus

216 infection and Chlamydiosis in koalas). In practice, whether TVs are pursued over conventional 217 alternatives should be evidence driven. For example, to evaluate whether host behavior or life 218 history may constrain vaccine transmission to impractical levels, the maximum coverage that could 219 be expected from a TV can be estimated from the proportion of individuals in target host 220 populations that are naturally infected with the candidate viral vector. Similarly, the geographic 221 extent of spread can be inferred from vector population genetics (7). Dynamic models derived 222 from these data, and similar data describing the transmission dynamics of the target pathogen 223 (including the potential roles of alternative host species in long-term maintenance), would be expected to support positive benefit-cost ratios of TVs over alternatives, whether through increased 224 225 levels of vaccine coverage or improved immunological protection. When appropriate, models 226 should consider sensitivity to vaccine reversion, reduced vaccine fitness from genetic 227 manipulation, and competition with the wildtype virus (10, 11).

228 Deployment of biological agents that spread in natural populations raises distinct regulatory 229 considerations and may require a broad view of incentives for industrial investment (e.g., philanthropic benefits). When developed and applied carefully, self-spreading agents have 230 231 benefitted human health (e.g., reduction of dengue using Wolbachia endosymbionts in mosquitoes 232 (13)) and agriculture (e.g., control of plant pathogens using phage cocktails and baculoviruses 233 (14)). The TVs we propose add complexity through their requirement for genetic modification. However, other self-spreading interventions harnessing genomic engineering (CRISPR, gene 234 235 drives) are advancing, creating blueprints for how staged co-development can empower evidence-236 based policymaking and find solutions to regulatory, financial, and social challenges (12, 15). 237 Provided that a TV can be safely developed and shows promise for disease control, decisions on 238 real world use would need to consider the balance of knowable harm done by withholding use and 239 knowable harm done by release. The commitments presented here are intended to encourage 240 deliberations characterized by understanding, accountability, and transparency, advancing a 241 collaborative future in which TVs may contribute to the public good.

### 242 REFERENCES AND NOTES

- 243 1. F. Lentzos *et al.*, *Science* **375**, 31 (2022).
- 244 2. J. B. Sandbrink, M. C. Watson, A. M. Hebbeler, K. M. Esvelt, *Nat. Ecol. Evol.* 5, 405 (2021).
- 245 3. J. Maki *et al.*, Vet. Res. 48, 57 (2017).
- 4. S. L. Nuismer, J. J. Bull, Nat. Ecol. Evol. 4, 1168-1173 (2020).

- 247 5. J. M. Torres *et al.*, *Vaccine* **19**, 4536 (2001).
- 248 6. A. J. Basinski *et al.*, *Vaccine* **36** (2018).
- 249 7. M. E. Griffiths *et al.*, *PLOS Biol.* **20**, e3001580 (2022).
- 250 8. R. C. Condit *et al.*, *Vaccine* **34** (2016).
- 251 9. N. C. Layman, B. M. Tuschhoff, S. L. Nuismer, Virus Evol. 7 (2021).
- 252 10. M. E. Griffiths, D. K. Meza, D. T. Haydon, D. G. Streicker, *Proc. Natl. Acad. Sci.* 120,
  253 e2216667120 (2023).
- 254 11. T. J. Varrelman *et al.*, *Proc. Natl. Acad. Sci.* **119**, e2108610119 (2022).
- 255 12. J. Buchthal, S. W. Evans, J. Lunshof, S. R. Telford, K. M. Esvelt, Philos. Trans. R. Soc. B
- 256 Biol. Sci. 374, 1 (2019).
- 257 13. W. A. Nazni *et al.*, *Curr. Biol.* **29**, 4241-4248.e5 (2019).
- 258 14. J. Wagemans *et al.*, *Annu. Rev. Phytopathol.* **60**, 21 (2022).
- 259 15. K. C. Long *et al.*, *Science* **370**, 1417 (2020).
- 260

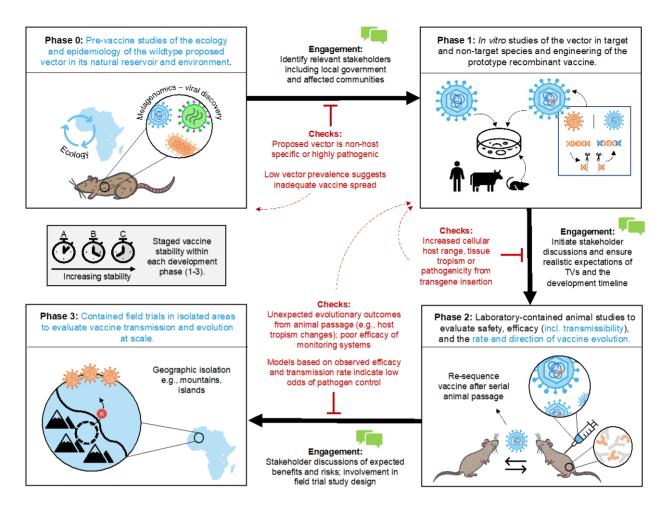
## 261 Acknowledgements:

- 262 We thank Ariel Leon, Daniel Walsh, and members of the Streicker group for helpful comments
- 263 on earlier versions of this manuscript.

264 Funding:

- 265 United States National Science Foundation grant DEB 2216790 (SLN, DGS)
- 266 Wellcome Trust Senior Research Fellowship 217221/Z/19/Z (DGS, MEG, LMB)
- 267 United States National Science Foundation grant DEB 2314616 (SLN)
- 268 United States National Institutes of Health 2R01GM122079-05A1 (SLN)
- 269 Coordenação de Aperfeiçoamento de Pessoal de Nível Superior Brasil (MVSM).
- 270 United Kingdom Biotechnology and Biological Sciences Research Council grant
- 271 BB/M009513/1 (DS)
- German Ministry of Education and Research BMBF grant 01KI2210 (IAY).
- 273 United States National Institutes of Health grant R01GM140459 (DAK).
- 274 Author contributions:
- 275 Conceptualization: DGS, SLN
- 276 Funding acquisition: DGS, SLN

277	Investigation: DGS, MEG, RA, LB, PB, MVSM, KE, MF, AG, BH, MAJ, DAK, JK,
278	CNW, CR, TR, KR, CS, JS, DS, IAY, JJB, SLN
279	Writing - original draft: DGS, MEG, JJB, SLN
280	Writing - review and editing: DGS, MEG, RA, LB, PB, MVSM, KE, MF, AG, BH, MAJ,
281	DAK, JK, CNW, CR, TR, KR, CS, JS, DS, IAY, JJB, SLN
282	Competing interests: MF is an employee of the Institute for Disease Modeling, a
283	research group within, and solely funded by, the Bill and Melinda Gates Foundation; the
284	findings, conclusions, and views expressed herein are those of the authors and do not
285	necessarily represent those of the Bill & Melinda Gates Foundation. KR is supported by
286	the division of intramural research, United States National Institutes of Allergy and
287	Infectious Diseases. The findings and conclusions in this publication should not be
288	construed to represent official USDA determination or policy. Any use of trade, firm, or
289	product names is for descriptive purposes only and does not imply endorsement by the
290	U.S. Government. MAJ, SLN & KR are listed as inventors on a pending patent associated
291	with a betaherpesvirus-vectored vaccine against Lassa fever virus.
292	Data and materials availability: NA
293	
294	
254	
295	
206	
296	
297	
200	
298	
299	
300	
301	
302	



304 Figure 1. Transmissible vaccine development would proceed in discrete phases with

305 established checkpoint criteria (red) necessitating vaccine re-design or an alternative viral

vector. Stakeholder engagement (green dialog boxes), intersectorial meetings of scientists and
 regulators, and fundamental research into the evolution of replicating, engineered organisms

and fundamental research into the evolution of repreating, engineered organismsencompass the full development process. Blue text indicates aspects that are distinct from

- 309 conventional vaccine development.

# **Box 1.** Seven proposed commitments for the responsible development of transmissible

# 322 vaccines for infectious disease control in animals

- 323 1.Vaccines will use naturally occurring, and host specific viruses as vectors, that would be324 isolated from and returned to their natural host species after antigen insertion.
- 325 2.Genetic modifications that increase host range, pathogenicity, or transmissibility, or create326 secondary hazards will not be intentionally pursued.
- 327 3.Technologies that could plausibly be harmful if applied to a human virus should be avoided.
- 4.Development will be staged with defined checkpoints and carried out within appropriatelycontrolled environments.
- 5.Unintended spread and consequences will be monitored throughout development stages, withcontingency plans.
- 332 6.Development will be transparent and community-led.
- 333 7.Safety standards will approach the strictest standards of partner nations involved.