



THE UNIVERSITY *of* EDINBURGH

Edinburgh Research Explorer

Frailty Index associates with GRIN2B in two representative samples from the United States and the United Kingdom

Citation for published version:

Mekli, K, Stevens, A, Marshall, AD, Arpawong, TE, Phillips, DF, Tampubolon, G, Lee, J, Prescott, CA, Nazroo, JY & Pendleton, N 2018, 'Frailty Index associates with GRIN2B in two representative samples from the United States and the United Kingdom', *PLOS ONE*, vol. 13, no. 11, e0207824, pp. 1-12.
<https://doi.org/10.1371/journal.pone.0207824>

Digital Object Identifier (DOI):

[10.1371/journal.pone.0207824](https://doi.org/10.1371/journal.pone.0207824)

Link:

[Link to publication record in Edinburgh Research Explorer](#)

Document Version:

Publisher's PDF, also known as Version of record

Published In:

PLOS ONE

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



RESEARCH ARTICLE

Frailty Index associates with *GRIN2B* in two representative samples from the United States and the United Kingdom

Krisztina Mekli^{1*}, Adam Stevens², Alan D. Marshall³, Thalida E. Arpawong⁴, Drystan F. Phillips^{5,6}, Gindo Tampubolon⁷, Jinkook Lee^{5,6}, Carol A. Prescott^{4,8}, James Y. Nazroo¹, Neil Pendleton⁹

1 Cathie Marsh Institute for Social Research, The University of Manchester, Manchester, United Kingdom, **2** Division of Developmental Biology and Medicine, The University of Manchester, Manchester, United Kingdom, **3** School of Social and Political Science, The University of Edinburgh, Edinburgh, United Kingdom, **4** Department of Psychology, Dornsife College of Letters, Arts and Sciences, University of Southern California, Los Angeles, CA, United States of America, **5** Dornsife Center for Economic and Social Research, University of Southern California, Los Angeles, CA, United States of America, **6** RAND Corporation, Santa Monica, CA, United States of America, **7** Institute for Social Change, The University of Manchester, Manchester, United Kingdom, **8** Davis School of Gerontology, University of Southern California, Los Angeles, CA, United States of America, **9** Division of Neuroscience and Experimental Psychology, The University of Manchester, Manchester, United Kingdom

☯ These authors contributed equally to this work.

* Krisztina.Mekli@manchester.ac.uk



OPEN ACCESS

Citation: Mekli K, Stevens A, Marshall AD, Arpawong TE, Phillips DF, Tampubolon G, et al. (2018) Frailty Index associates with *GRIN2B* in two representative samples from the United States and the United Kingdom. PLoS ONE 13(11): e0207824. <https://doi.org/10.1371/journal.pone.0207824>

Editor: Pavel I. Ortinski, University of South Carolina School of Medicine, UNITED STATES

Received: July 16, 2018

Accepted: November 5, 2018

Published: November 26, 2018

Copyright: © 2018 Mekli et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All the pheno- and genotype data are publicly available. HRS: https://www.ncbi.nlm.nih.gov/projects/gap/cgi-bin/study.cgi?study_id=phs000428.v2.p2 ELSA: <https://www.ebi.ac.uk/ega/studies/EGAS00001001036>.

Funding: This work was supported by the Medical Research Council (<https://www.mrc.ac.uk/>) [grant number: G1001375] (KM, ADM, GT, JYN, NP) and University of Manchester Research Institute (<http://www.staffnet.manchester.ac.uk/services/rbess/research-strategy/umri/>) (KM, AS) in the United

Abstract

The concept of frailty has been used in the clinical and research field for more than two decades. It is usually described as a clinical state of heightened vulnerability to poor resolution of homeostasis after a stressor event, which thereby increases the risk of adverse outcomes, including falls, delirium, disability and mortality. Here we report the results of the first genome-wide association scan and comparative gene ontology analyses where we aimed to identify genes and pathways associated with the deficit model of frailty. We used a discovery-replication design with two independent, nationally representative samples of older adults. The square-root transformed Frailty Index (FI) was the outcome variable, and age and sex were included as covariates. We report one hit exceeding genome-wide significance: the rs6765037 A allele was significantly associated with a decrease in the square-root transformed FI score in the Discovery sample (beta = -0.01958, p = 2.14E-08), without confirmation in the Replication sample. We also report a nominal replication: the rs7134291 A allele was significantly associated with a decrease in the square-root transformed FI score (Discovery sample: beta = -0.01021, p = 1.85E-06, Replication sample: beta = -0.005013, p = 0.03433). These hits represent the *KBTBD12* and the *GRIN2B* genes, respectively. Comparative gene ontology analysis identified the pathways ‘Neuropathic pain signalling in dorsal horn neurons’ and the ‘GPCR-Mediated Nutrient Sensing in Enteroendocrine Cells’, exceeding the p = 0.01 significance in both samples, although this result does not survive correction for multiple testing. Considering the crucial role of *GRIN2B* in brain development, synaptic plasticity and cognition, this gene appears to be a potential candidate to play a role in frailty. In conclusion, we conducted genome-wide association scan and pathway analyses

Kingdom and the National Institute on Aging (<https://www.nia.nih.gov/>) [grant numbers: R01 AG030153 (JL, DFP) and F32 AG048681 (TEA, CAP)] in the United States. The funding sources had no further role in conducting the research or in publication. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. JL and DFP are affiliated with RAND Corporation, Santa Monica. However, the RAND provided no support in the form of salaries for authors [JL, DFP], and did not have any additional role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript. The specific roles of these authors are articulated in the 'author contributions' section'.

Competing interests: JL and DFP are affiliated with RAND Corporation, Santa Monica. However, this does not alter our adherence to PLOS ONE policies on sharing data and materials.

and have identified genes and pathways with potential roles in frailty. However, frailty is a complex condition. Therefore, further research is required to confirm our results and more thoroughly identify relevant biological mechanisms.

Introduction

Frailty is a state of vulnerability to poor resolution of homeostasis after a stressor event and is a consequence of cumulative decline in many physiological systems over a lifetime [1]. From a clinical perspective, frailty is important because it constitutes a condition of greater risk for adverse health outcomes, such as falls, compromises in mobility, and independence, hospitalization, disability, and death [2].

Research has established that frailty associates with older age, female gender, functional dependence and chronic disease [3]. However, the pathophysiological mechanisms behind frailty are not clear. Studies with a biological focus have identified inflammation [4,5] and hormonal [6,7] dysregulation as important correlates of frailty. Other frailty-associated biomarkers include insulin growth factor, vitamin D, sirtuins, glycoproteins and cystatin C [8]. Hypothesis-driven candidate gene studies have shown associations between frailty and genes involved in inflammatory pathways [9, 10], insulin pathway, cortisol system and apoptosis [8]. These studies highlight the importance of certain genes, but fail to provide a wider picture of the multiple systems involved in frailty. Adequately powered genome-wide association scan studies supplemented with systems biology analyses can fulfil this aim by identifying multiple biological pathways with a potential role in the trait.

Several frailty assessment tools have been proposed, often using different conceptual models of frailty. The deficit model, adds together several impairments and conditions to create a Frailty Index (FI), on the grounds that the more deficits a person has, the more likely that person is to be frail. The FI can easily be constructed using information that is readily available in most health surveys [11] and it performs very well in predicting mortality compared to other frailty measures [12].

Here we report the results of a hypothesis-free genome-wide association scan (GWAS) study of frailty. We used a discovery-replication design with two independent community representative samples of older adults, the Health and Retirement Study (HRS) in the US and the English Longitudinal Study of Ageing (ELSA) in the UK. To ensure maximum replicability we used the similarly constructed FI [11] as outcome variable and a nearly identical genotyping platform in the two samples. We supplemented our study with comparative gene ontology analysis to identify key biological systems in frailty. To our knowledge this is the first study investigating the genetics of frailty in a context of systems biology. We report on genes and pathways with potential roles in frailty. Our results implicate new potential mechanisms to previously reported ones, such as inflammatory pathways and hormonal dysregulations. We expect that our results will contribute towards the better understanding of pathophysiological mechanisms behind frailty and potentiate the search for possible interventions.

Material and methods

Sample

We used the discovery and replication sample design. The Discovery sample was drawn from wave 9 (2008) of the Health and Retirement Study (HRS), a nationally representative sample of

households of older Americans in the United States (<https://hrs.isr.umich.edu/>). We included participants with relatively homologous ancestry, defined by falling within 1 standard deviation of all self-identified Whites for Eigenvectors 1 and 2 for the principal component analysis and within 1 SD for fraction of heterozygotes for autosomal SNPs (<http://hrsonline.isr.umich.edu/>). After excluding participants with sex discrepancy, relatedness and chromosomal anomalies, we had 8539 individuals with genotype data. All participants provided written consent, and ethical approval was granted by the University of Michigan Institutional Review Board.

The Replication sample was the English Longitudinal Study of Ageing in the United Kingdom (ELSA, <http://www.elsa-project.ac.uk/>), a nationally representative cohort of individuals living in England aged 50 and older [13]. In this sample we included 5251 individuals who were interviewed in wave 2 (2004). Participants with non-White ethnicity or sex discrepancy were excluded. All participants provided written consent and ethical approval was granted by the London Multi-Centre Research Ethics Committee.

The investigation was carried out in accordance with the latest version of the Declaration of Helsinki.

Genetic data

The genetic data was obtained by using Illumina's Human Omni2.5-Quad BeadChip (Illumina, San Diego, CA, USA). For the Discovery sample genotyping was performed by the NIH Center for Inherited Disease Research (Johns Hopkins University, Baltimore, MD, USA) using HumanOmni2.5-4v1 platform. For the Replication sample the University College London Genomics (London, UK) performed the genotyping on HumanOmni2.5-8v1. Initial quality control on the genetic data was performed by the data holder.

During the analysis, SNPs with minor allele frequency below 5% in the Discovery sample and below 1% in the Replication sample were also excluded, yielding 1,215,858 and 1,490,612 SNPs entering the analysis for the Discovery and Replication samples, respectively.

Phenotypic measures

A Frailty Index (FI) was created following guidance in the literature [11]. The aim was to use a well-accepted measure which yields sufficient sample size in the genetic association analyses. Briefly, the FI counts health-related problems (deficits) in a range of domains (activities of daily living, cognitive function, falls and fractures, joint replacement, vision, hearing, chronic diseases, cardiovascular diseases, depression). The number of deficits was 45 in the Discovery sample and only individuals with non-missing values for at least 25 of them were included. In the Replication sample the number of deficits was 62, with the minimum requirement of 30 non-missing values.

Details of the development of the FI measures can be found in [S1 Table](#).

Statistical analysis

Phenotypic measures were developed using Stata12 software (Stata Corporation, <http://www.stata.com/>).

To normalise the negatively skewed distribution of the FI we performed square root transformation.

We performed linear regression analysis using the square-root transformed FI with sex and age as covariates. Regression analyses were performed using the PLINK software [14, 15].

To avoid spurious association results arising from unadjusted population substructure we used the first four Eigenvectors as covariates in the Discovery sample. The Replication sample contained only white individuals and literature indicates only modest population stratification

in the British population [16]; therefore we did not include Eigenvectors in this part of the analysis.

Genomic inflation factor was calculated by R (<http://cran.us.r-project.org/>).

Genetic results annotation was performed by the Ensembl Variant Effect Predictor, Assembly: GRCh38.p5 [17].

We set the nominal replication criteria as $p < 0.05$ and the same direction of effect in the Replication sample.

Comparative gene ontology analysis methods

Biological pathways associated with SNPs were determined using a right-sided Fisher’s exact test (Ingenuity Pathways Analysis [IPA], Qiagen Inc, <https://www.qiagenbioinformatics.com/products/ingenuity-pathway-analysis/>).

Results

Demographic and phenotypic results

Table 1 shows that there were more females than males in both samples. Compared with the Replication sample, the Discovery sample’s mean age and mean FI were significantly higher (t-test $p < 0.0001$). The distribution of the FI measure in the Discovery and Replication sample can be found in S2 Table.

Genetic association results

We report one genome-wide significant hit. The rs6765037 A allele significantly associated with a decrease in the FI score in the Discovery sample ($\beta = -0.01958$, $p = 2.14E-08$), although this was not confirmed in the Replication sample ($\beta = -0.005026$, $p = 0.2041$).

We report 31 other associations below the suggestive ($p < 0.0001$) level, spanning 10 regions in the genome. These SNPs are in high linkage disequilibrium (LD) with each other within one region, therefore there are fewer independent signals. For example, among the 7 SNPs identified on chromosome 14, the lowest LD was 0.917296 between rs73301475 and rs17093546, suggesting only one independent signal in this region. The 32 associations exceeding the suggestive level can be found in S3 Table.

We report one successful nominal replication between the Discovery and Replication samples. The rs7134291 A allele significantly associated with a decrease in the FI score (Discovery sample: $\beta = -0.01021$, $p = 1.85E-06$, Replication sample: $\beta = -0.005013$, $p = 0.03433$).

The genomic inflation factors were 1.033 for the Discovery and 1.001 for the Replication sample. They did not indicate serious population stratification in the samples. We also provide Manhattan and QQ plots to visualise our results, which can be found in S4 Table.

Comparative gene ontology analysis results

Eight pathways were shown to be significantly ($p < 0.05$) represented in the groups of SNPs derived from both analyses. Of the pathways identified, the ‘Neuropathic pain signalling in dorsal horn neurons’ and the ‘GPCR-Mediated Nutrient Sensing in Enteroendocrine Cells’

Table 1. Sample characteristics.

		Discovery sample (HRS)	Replication sample (ELSA)
males (%)females (%)		3546 (41.53) 4993 (58.47)	2397 (45.67) 2851 (54.33)
Frailty Index (FI) measure	available	n = 8232	n = 5248
	mean FI (SE)	0.205 (0.0014)	0.169 (0.0015)
	mean age (SE)	69.4 (0.113)	65.9 (0.130)

<https://doi.org/10.1371/journal.pone.0207824.t001>

achieved $p < 0.01$ in both samples (Fig 1). None of these results survive Bonferroni-correction for multiple testing.

Discussion

To our knowledge, our study is the first genome-wide association study on the deficit model of frailty. Our aim was to use a hypothesis-free approach to identify genes and pathways in order

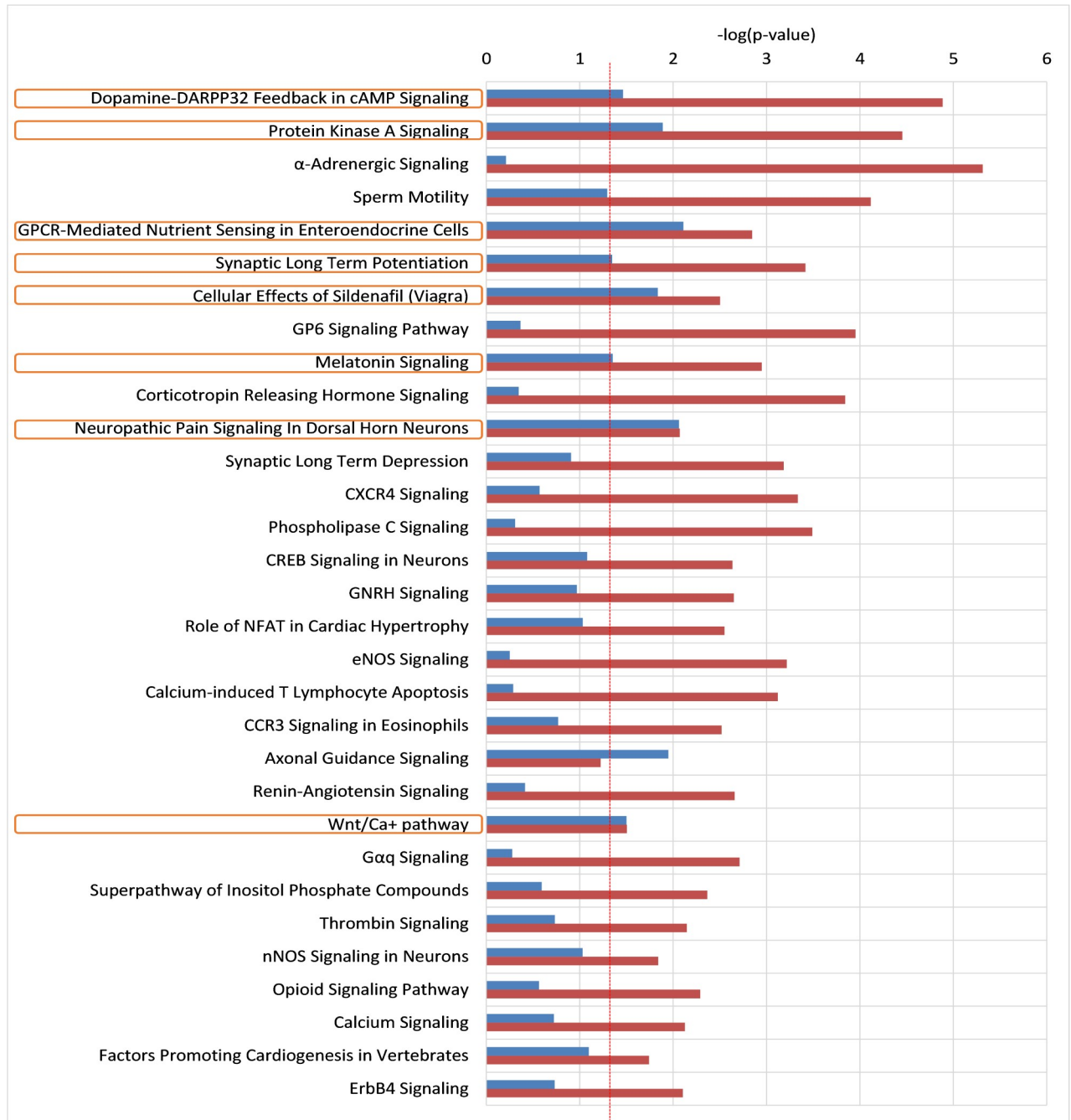


Fig 1. Results of the comparative gene ontology analysis. Eight pathways were shown to be significantly ($p < 0.05$) represented in the groups of SNPs derived from both studies, they are marked with orange boxes. Horizontal red line indicates the $p = 0.05$ significance level. Significance levels on a negative log scale are indicated with blue (Discovery sample) and red (Replication sample) bars.

<https://doi.org/10.1371/journal.pone.0207824.g001>

to understand pathophysiological mechanisms behind frailty. We used two independent, representative samples from the US and the UK. We chose a widely accepted frailty measure, the Frailty Index [11] as the outcome variable.

Our most significant association was between the rs6765037 A allele and a decrease in the FI score. This SNP located in an intergenic region of chromosome 3, close to the 5' region of the *KBTBD12* (Kelch repeat and BTB Domain containing 12) gene. This gene is not well characterised in the literature. According to the Human Protein Atlas, higher RNA expression was found in the heart muscle and in the skeletal muscle [18]. We did not find any past associations in the literature indicating the role of *KBTBD12* or rs6765037 in any phenotype relevant to frailty.

The nominally replicated SNP, rs7134291 is located on chromosome 12, within the first intron of the *GRIN2B* (Glutamate Ionotropic Receptor NMDA Type Subunit 2B) gene. *N*-methyl-D-aspartate receptors (NMDARs) are a family of ionotropic glutamate receptors that mediate a slow Ca^{2+} permeable component of excitatory synaptic transmission in the central nervous system. NMDARs are a tetrameric assembly with the *GRIN2B* encoding the glutamate-binding NR2B subunit [19]. NR2B together with the NR1 and NR2A subunits are expressed in the human cerebral cortex during the second trimester of gestation, a period of intense neurogenesis and synaptogenesis, suggesting involvement of NMDARs in the maturation of human cortical neurons and in early synapse formation [20]. Overexpression of NR2B in the forebrain (cortex and hippocampus) has been shown to lead to increased learning and memory abilities in behavioural tasks in young adult mice [21], as well as in ageing mice [22]. In humans, variants in the *GRIN2B* gene have been associated with impaired cognitive phenotypes such as intellectual disability [23], and developmental delay [24, 25].

NMDA receptors are activated by L-glutamate and glycine. Binding of these co-agonists must take place for the ion channel to open fully. Extracellular Mg^{2+} blocks the ion channel absent membrane depolarization, which requires prior activation of AMPA and kainate receptors. This process is thought to underlie the contribution of NMDARs to synaptic plasticity and long-term potentiation (LTP) [26]. Long-term potentiation in the cortex is a long-lasting highly localised increase in synaptic strength and a molecular substrate for memory and learning [27]. Loss of synaptic plasticity is a characteristic of Alzheimer's disease (AD) and the molecular mechanism possibly involves the NMDARs. Beta-amyloid ($\text{A}\beta$) oligomers, one of the toxic protein species believed to be etiologically related to AD, impair LTP and enhance long-term depression [28]. $\text{A}\beta$ reduces glutamatergic transmission and inhibits synaptic plasticity through regulating the number of NMDARs. [29]. Application of $\text{A}\beta$ has been found to promote endocytosis of NMDARs in cortical neurons, and neurons from a genetic mouse model of AD expressed reduced amount of surface NMDARs [30]. These observations have been supplemented with genetic studies that found associations between *GRIN2B* variants and AD [31, 32]. One of the studies, conducted in a Northern Chinese sample found that the frequency of the rs3764028 C allele was higher in AD cases than in controls, even in APOE ϵ 4-negative cases. This is an upstream SNP, for which a Luciferase reporter assay showed 35–40% lower promoter activity for the rs3764028 C allele compared with the A allele. This indicates that the major C allele might decrease the transcriptional activity of *GRIN2B*. Rs3764028 was not present in our dataset, with the closest SNP being rs12368476, 1077 bp away. The rs12368476 A allele associated with a decrease in the FI in both samples (HRS: beta = -0.003893, $p = 0.045$, ELSA: beta = -0.005127, $p = 0.017$). Our nominally replicated SNP, rs12368476, is 7819 bp away from rs12368476 with an r^2 of 0.4478 between them. It is possible, that one or more SNPs in the upstream region of *GRIN2B* decrease the promoter activity of the gene and hence the association with FI and AD, via decreased synaptic plasticity. The involvement of synaptic plasticity in frailty is further supported by the fact that the 'Synaptic

long-term potentiation' pathway was among the eight pathways exceeding the $p = 0.05$ threshold significance in our comparative gene ontology analysis.

Considering the crucial role of *GRIN2B* in brain development, synaptic plasticity and cognition, this gene appears to be a plausible candidate to play a role in frailty.

It is possible that the found association is driven by cognitive deficit, which is included in our frailty model. The association may also be indirect, manifesting through other deficits, which are associated with cognition, such as sensory impairment [33] or depression [34]. These deficits are part of the FI, which includes self-reported eyesight and hearing and 8 questions from the Center for Epidemiologic Studies Depression Scale questionnaire (CESD; [35]). However, frailty is defined as a disorder of several inter-related physiological systems [1] and our operationalisation of the FI in both the Discovery and Replication samples included markers of this full range of systems; therefore *GRIN2B*'s role in cognition is unlikely to be a comprehensive explanation for its association with such a complex phenotype.

Indeed, in the GWAS literature, *GRIN2B* has been linked to various other traits. Relevant to our study are IL2 secretion [36], time to major incident event [37] and FSH levels in women [38].

Aging is characterized by raised levels of proinflammatory cytokines such as interleukin-1 (IL-1), interleukin-6 (IL-6) and tumour necrosis factor (TNF) as well as reduced IL-2 levels, reflecting a low-grade chronic systemic proinflammatory state, termed *inflammaging*. As a result, the immune system declines in responsiveness and efficiency resulting in greater susceptibility to age-related diseases and frailty. In this context, the association of a *GRIN2B* variant, rs2268118, with interleukin-2 (IL-2) secretion to vaccinia virus stimulation in smallpox vaccine recipients [36] is noteworthy. Although it appears a plausible candidate, we have not found studies investigating associations between frailty or any related phenotype and IL-2.

Literature suggests that *GRIN2B* may also have a role in human longevity. A GWAS study on time to event (defined by major incident events, such as myocardial infarction, heart failure, stroke, dementia, hip fracture, or cancer) or death, as an alternative phenotype for healthy aging, found that another genetic variant in *GRIN2B*, rs4764043, was associated with increased risk of event [37]. This is relevant to our study, as these items are part of the FI measure (except for death) and they increase the prediction of mortality compared to an FI without these items [39]. The results of the Walter study implies *GRIN2B*'s involvement of the pathophysiology of these diseases through yet unknown mechanisms, and in this way would increase risk of frailty.

A third variant, rs6488619, has been associated with follicle-stimulating hormone (FSH) levels in Caucasian women [38]. Higher levels of FSH were associated with higher FI in a study of 3219 men [7]. In this case the association may be indirect, as higher FSH levels could be markers of testicular dysfunction, which itself is a sensitive marker of homeostatic disruption and poor overall health status in older men.

Of these SNPs, rs4764043 and rs6488619 were included in our GWAS, but neither associated with the FI (results not shown).

Our nominally replicated SNP, rs7134291, has shown some association with triglyceride levels in the literature, although this did not reach genome-wide significance [40]. We found no association between rs7134291 and any lipid biomarker in our study (results not shown).

Taking these results together, *GRIN2B* appears to be a plausible candidate in the pathophysiology of frailty, but these results also indicate the involvement of other mechanisms.

Comparative gene ontology analysis identified two pathways exceeding the $1.0E-02$ threshold in both samples. The first pathway is the 'Neuropathic pain signalling in dorsal horn neurons' with the NMDARs having an important role in it. Neuropathic pain is generally defined as a chronic pain state resulting from peripheral or central nerve injury, or both, and is likely

to be due to long-term plastic changes along the nociceptive pathway. The spinal cord dorsal horn is the first relay station of nociceptive information from periphery to the brain and NMDAs are expressed here abundantly [41, 42]. Activation of postsynaptic NMDA receptors not only participates in glutamatergic sensory synaptic transmission in normal conditions, but more importantly is also involved in the spinal dorsal horn in pathological pain conditions induced by tissue inflammation or nerve injury [43]. All NMDAR subunits are expressed in the spinal dorsal horn; however, NR2B exhibits the largest expression among NR2 subunits [41]. There has been evidence in the literature of an association between pain and the Frailty Index. A study using our Replication sample found that pain status is predictive of incident and worsening frailty [44], as did a prior study of men with chronic widespread pain [45].

The second pathway is the 'GPCR-Mediated Nutrient Sensing in Enteroendocrine Cells'. Food intake is detected by the chemical senses of taste and smell and subsequently by chemosensory cells in the gastrointestinal tract that link the composition of ingested foods to feedback circuits controlling gut motility/secretion, appetite, and peripheral nutrient disposal. A number of G-protein-coupled receptors (GPCRs) have been identified as potential "sensors" of luminal nutrients. These receptors stimulate the release of gut peptides via coupling to $G\alpha_s$, which elevates intracellular cAMP, and $G\alpha_q$, which then results in Ca^{2+} mobilization and Protein Kinase C activation. Inhibitory influences are exerted by signalling through $G\alpha_i$ pathways that decrease cAMP levels [46]. As anorexia in aging, defined as a loss of appetite and/or reduced food intake, affects a significant number of elderly people and is far more prevalent among frail individuals [47], this pathway may play a role in frailty.

Our study has considerable strengths. We used large sample sizes in two independent cohorts. The very similarly constructed phenotypic measures and the nearly identical genotyping platform between the two cohorts ensures maximum replicability. Our samples consist of community dwelling individuals, who are not hospitalised or institutionalised, therefore frailty is not overrepresented. The FI distribution is similar to that previously reported in the literature [11] S2 Table.

Our study suffers from some limitations. First, the FI measure contained 45 deficits in the Discovery sample and 62 deficits in the Replication sample. However, the missing 17 items in the Discovery sample were distributed across each health domain covered by the Index, so this is unlikely to pose a serious problem. Moreover, an Index with 30–40 variables has been shown to be sufficiently accurate for predicting adverse outcomes [11]. Second, frailty may be influenced by environmental factors, such as nutrition or access to health care, which we did not account for. The latter might be especially important as the health care system is rather different between the US and the UK. Also, the use of samples from different countries may result in cohort effects. FI is a complex phenotype, and therefore is likely to be influenced by many genes with small effects, our study could be underpowered to find associations for less common, but possibly important variants, despite our best efforts. This insufficient power may explain the lack of confirmation of previously reported correlates of frailty, such as members of the inflammatory pathway (*IL-1 β* , *IL-6* and *TNF*) and apoptosis (rs129968 in *CREBBP* and rs3769827 in *CASP8*) [8] in our study. In future studies larger sample sizes will be required to detect these small effects.

Finally, we note that the two pathways exceeding the 1.0E-02 threshold in the gene ontology analysis did not survive adjustment for multiple testing, despite their potential biological relevance. This may be due to insufficient power; therefore additional genetic association and experimental studies are needed to verify these potentially important pathways.

In conclusion, our study's main finding implicates the importance of the *GRIN2B* and a pathway with NR2B playing a potential role using two different analytical techniques in two

independent samples. However, as frailty is a complex condition, further research is required to corroborate our findings and reveal further pathways associated with frailty.

Supporting information

S1 Table. Phenotypes. Lists of deficits for the Frailty Index.
(XLSX)

S2 Table. Graphs. Distribution of the phenotypic variable.
(XLSX)

S3 Table. GenAssoc. The most significant results of the genome-wide association scan.
(XLSX)

S4 Table. Plots. QQ and Manhattan plots of the GWAS.
(XLSX)

Acknowledgments

Mr. John McLoughlin for his technical assistance, programming and Linux administration.
All participants in HRS and ELSA.

Author Contributions

Conceptualization: Krisztina Mekli, Jinkook Lee, James Y. Nazroo, Neil Pendleton.

Data curation: Drystan F. Phillips, Gindo Tampubolon.

Formal analysis: Krisztina Mekli, Adam Stevens, Alan D. Marshall, Thalida E. Arpawong.

Supervision: James Y. Nazroo, Neil Pendleton.

Writing – original draft: Krisztina Mekli, Adam Stevens.

Writing – review & editing: Alan D. Marshall, Thalida E. Arpawong, Drystan F. Phillips, Gindo Tampubolon, Jinkook Lee, Carol A. Prescott, James Y. Nazroo, Neil Pendleton.

References

1. Clegg A, Young J, Iliffe S, Rikkert MO, Rockwood K. Frailty in elderly people. *Lancet*. 2013 381 (9868):752–62. Review. [https://doi.org/10.1016/S0140-6736\(12\)62167-9](https://doi.org/10.1016/S0140-6736(12)62167-9) PMID: 23395245
2. Fried LP, Tangen CM, Walston J, Newman AB, Hirsch C, Gottdiener J, et al. Frailty in older adults: evidence for a phenotype. *J Gerontol A Biol Sci Med Sci*. 2001 56(3):M146–56. PMID: 11253156
3. Walston J, Hadley EC, Ferrucci L, Guralnik JM, Newman AB, Studenski SA, et al. Research agenda for frailty in older adults: toward a better understanding of physiology and etiology: summary from the American Geriatrics Society/National Institute on Aging Research Conference on Frailty in Older Adults. *J Am Geriatr Soc*. 2006 54(6):991–1001. Review. <https://doi.org/10.1111/j.1532-5415.2006.00745.x> PMID: 16776798
4. Collerton J, Martin-Ruiz C, Davies K, Hilken CM, Isaacs J, Kolenda C, et al. Frailty and the role of inflammation, immunosenescence and cellular ageing in the very old: cross-sectional findings from the Newcastle 85+ Study. *Mech Ageing Dev*. 2012 133(6):456–66. <https://doi.org/10.1016/j.mad.2012.05.005> PMID: 22663935
5. Hubbard RE, O'Mahony MS, Savva GM, Calver BL, Woodhouse KW. Inflammation and frailty measures in older people. *J Cell Mol Med*. 2009 13(9B):3103–9. <https://doi.org/10.1111/j.1582-4934.2009.00733.x> PMID: 19438806
6. Cappola AR, Xue QL, Fried LP. Multiple hormonal deficiencies in anabolic hormones are found in frail older women: the Women's Health and Aging studies. *J Gerontol A Biol Sci Med Sci*. 2009 64(2):243–8. <https://doi.org/10.1093/gerona/gln026> PMID: 19182229

7. Tajar A, O'Connell MD, Mitnitski AB, O'Neill TW, Searle SD, Huhtaniemi IT, et al. Frailty in relation to variations in hormone levels of the hypothalamic-pituitary-testicular axis in older men: results from the European male aging study. *J Am Geriatr Soc*. 2011 59(5):814–21. <https://doi.org/10.1111/j.1532-5415.2011.03398.x> PMID: 21568952
8. Viña J, Tarazona-Santabalbina FJ, Pérez-Ros P, Martínez-Arnau FM, Borrás C, Olaso-Gonzalez G, et al. Biology of frailty: Modulation of ageing genes and its importance to prevent age-associated loss of function. *Mol Aspects Med*. 2016 50:88–108. <https://doi.org/10.1016/j.mam.2016.04.005> PMID: 27164416
9. Mekli K, Marshall A, Nazroo J, Vanhoutte B, Pendleton N. Genetic variant of Interleukin-18 gene is associated with the Frailty Index in the English Longitudinal Study of Ageing. *Age Ageing*. 2015 44(6):938–42. <https://doi.org/10.1093/ageing/afv122> PMID: 26396182
10. Mekli K, Nazroo JY, Marshall AD, Kumari M, Pendleton N. Proinflammatory genotype is associated with the frailty phenotype in the English Longitudinal Study of Ageing. *Aging Clin Exp Res*. 2016 28(3):413–21. <https://doi.org/10.1007/s40520-015-0419-z> PMID: 26248682
11. Searle SD, Mitnitski A, Gahbauer EA, Gill TM, Rockwood K. A standard procedure for creating a frailty index. *BMC Geriatr*. 2008 8: 24. <https://doi.org/10.1186/1471-2318-8-24> PMID: 18826625
12. Pijpers E, Ferreira I, Stehouwer CD, Nieuwenhuijzen Kruseman AC. The frailty dilemma. Review of the predictive accuracy of major frailty scores. *Eur J Intern Med*. 2012 23(2):118–23. <https://doi.org/10.1016/j.ejim.2011.09.003> PMID: 22284239
13. Steptoe A, Breeze E, Banks J, Nazroo J. Cohort profile: the English Longitudinal Study of Ageing. *Int J Epidemiol*. 2013 42(6):1640–8. <https://doi.org/10.1093/ije/dys168> PMID: 23143611
14. Chang CC, Chow CC, Tellier LC, Vattikuti S, Purcell SM, Lee JJ. Second-generation PLINK: rising to the challenge of larger and richer datasets. *Gigascience*. 2015 4:7. <https://doi.org/10.1186/s13742-015-0047-8> PMID: 25722852
15. Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am. J. Hum. Genet*. 2007 81(3):559–575. <https://doi.org/10.1086/519795> PMID: 17701901
16. The Wellcome Trust Case Control Consortium. Genome-wide association study of 14,000 cases of seven common diseases and 3,000 shared controls. *Nature*. 2007 447(7145):661–78. <https://doi.org/10.1038/nature05911> PMID: 17554300
17. McLaren W, Pritchard B, Rios D, Chen Y, Flicek P, Cunningham F. Deriving the consequences of genomic variants with the Ensembl API and SNP Effect Predictor. *Bioinformatics*. 2010 26(16):2069–2070. <https://doi.org/10.1093/bioinformatics/btq330> PMID: 20562413
18. Uhlén M, Fagerberg L, Hallström BM, Lindskog C, Oksvold P, Mardinoglu A, et al. Proteomics. Tissue-based map of the human proteome. *Science*. 2015 347(6220):1260419. <https://doi.org/10.1126/science.1260419> PMID: 25613900
19. Hu C, Chen W, Myers SJ, Yuan H, Traynelis SF. Human *GRIN2B* variants in neurodevelopmental disorders. *J Pharmacol Sci*. 2016 132(2):115–121. Review. <https://doi.org/10.1016/j.jphs.2016.10.002> PMID: 27818011
20. Bagasrawala I, Memi F, V Radonjić N, Zecevic N. N-Methyl d-Aspartate Receptor Expression Patterns in the Human Fetal Cerebral Cortex. *Cereb Cortex*. 2017 27(11):5041–5053. <https://doi.org/10.1093/cercor/bhw289> PMID: 27664962
21. Tang YP, Shimizu E, Dube GR, Rampon C, Kerchner GA, Zhuo M, et al. Genetic enhancement of learning and memory in mice. *Nature*. 1999 401(6748):63–9. <https://doi.org/10.1038/43432> PMID: 10485705
22. Cao X, Cui Z, Feng R, Tang YP, Qin Z, Mei B, et al. Maintenance of superior learning and memory function in NR2B transgenic mice during ageing. *Eur J Neurosci*. 2007 25(6):1815–22. <https://doi.org/10.1111/j.1460-9568.2007.05431.x> PMID: 17432968
23. Endeley S, Rosenberger G, Geider K, Popp B, Tamer C, Stefanova I, et al. Mutations in *GRIN2A* and *GRIN2B* encoding regulatory subunits of NMDA receptors cause variable neurodevelopmental phenotypes. *Nat Genet*. 2010 42(11):1021–6. <https://doi.org/10.1038/ng.677> PMID: 20890276
24. Morisada N, Ioroi T, Taniguchi-Ikeda M, Juan Ye M, Okamoto N, Yamamoto T, et al. 12p13 *GRIN2B* deletion is associated with developmental delay and macrocephaly. *Hum Genome Var*. 2016 3:16029. eCollection 2016 Sep 15. <https://doi.org/10.1038/hgv.2016.29> PMID: 27656287
25. Dimassi S, Andrieux J, Labalme A, Lesca G, Cordier MP, Boute O, et al. Interstitial 12p13.1 deletion involving *GRIN2B* in three patients with intellectual disability. *Am J Med Genet A*. 2013 161A(10):2564–9. <https://doi.org/10.1002/ajmg.a.36079> PMID: 23918416
26. Walton HS, Dodd PR. Glutamate-glutamine cycling in Alzheimer's disease. *Neurochem Int*. 2007 50(7–8):1052–66. <https://doi.org/10.1016/j.neuint.2006.10.007> PMID: 17141374

27. Ji RR, Kohno T, Moore KA, Woolf CJ. Central sensitization and LTP: do pain and memory share similar mechanisms? *Trends Neurosci.* 2003 26(12):696–705. Review. <https://doi.org/10.1016/j.tins.2003.09.017> PMID: 14624855
28. Palop JJ, Mucke L. Amyloid-beta-induced neuronal dysfunction in Alzheimer's disease: from synapses toward neural networks. *Nat Neurosci.* 2010 13(7):812–8. Review. <https://doi.org/10.1038/nn.2583> PMID: 20581818
29. Zhang Y, Li P, Feng J, Wu M. Dysfunction of NMDA receptors in Alzheimer's disease. *Neurol Sci.* 2016 37(7):1039–47. Review. <https://doi.org/10.1007/s10072-016-2546-5> PMID: 26971324
30. Snyder EM, Nong Y, Almeida CG, Paul S, Moran T, Choi EY, et al. Regulation of NMDA receptor trafficking by amyloid-beta. *Nat Neurosci.* 2005 8(8):1051–8. <https://doi.org/10.1038/nn1503> PMID: 16025111
31. Andreoli V, De Marco EV, Trecroci F, Cittadella R, Di Palma G, Gambardella A. Potential involvement of GRIN2B encoding the NMDA receptor subunit NR2B in the spectrum of Alzheimer's disease. *J Neural Transm (Vienna).* 2014 121(5):533–42.
32. Jiang H, Jia J. Association between NR2B subunit gene (*GRIN2B*) promoter polymorphisms and sporadic Alzheimer's disease in the North Chinese population. *Neurosci Lett.* 2009 450(3):356–60. <https://doi.org/10.1016/j.neulet.2008.10.075> PMID: 18983893
33. Lin MY, Gutierrez PR, Stone KL, Yaffe K, Ensrud KE, Fink HA, et al. Vision impairment and combined vision and hearing impairment predict cognitive and functional decline in older women. *J Am Geriatr Soc.* 2004 52(12):1996–2002. <https://doi.org/10.1111/j.1532-5415.2004.52554.x> PMID: 15571533
34. Gotlib IH, Joormann J. Cognition and depression: current status and future directions. *Annu Rev Clin Psychol.* 2010 6:285–312. Review. <https://doi.org/10.1146/annurev.clinpsy.121208.131305> PMID: 20192795
35. Radloff LS. The CES-D Scale: A self-report depression scale for research in the general population. *Applied Psychological Measurement.* 1977 1(3):385–401.
36. Kennedy RB, Ovsyannikova IG, Pankratz VS, Haralambieva IH, Vierkant RA, Poland GA. Genome-wide analysis of polymorphisms associated with cytokine responses in smallpox vaccine recipients. *Hum Genet.* 2012 131(9):1403–21. <https://doi.org/10.1007/s00439-012-1174-2> PMID: 22610502
37. Walter S, Atzmon G, Demerath EW, Garcia ME, Kaplan RC, Kumari M, et al. A genome-wide association study of aging. *Neurobiol Aging.* 2011 32(11):2109.e15–2109.e28.
38. Schuh-Huerta SM, Johnson NA, Rosen MP, Sternfeld B, Cedars MI, Reijo Pera RA. Genetic variants and environmental factors associated with hormonal markers of ovarian reserve in Caucasian and African American women. *Hum Reprod.* 2012 27(2):594–608. <https://doi.org/10.1093/humrep/der391> PMID: 22116950
39. Theou O, Rockwood MR, Mitnitski A, Rockwood K. Disability and co-morbidity in relation to frailty: how much do they overlap? *Arch Gerontol Geriatr.* 2012 55(2):e1–8. <https://doi.org/10.1016/j.archger.2012.03.001> PMID: 22459318
40. Teslovich TM, Musunuru K, Smith AV, Edmondson AC, Stylianou IM, Koseki M, et al. Biological, clinical and population relevance of 95 loci for blood lipids. *Nature.* 2010 466(7307):707–713. <https://doi.org/10.1038/nature09270> PMID: 20686565
41. Wu LJ, Zhuo M. Targeting the NMDA receptor subunit NR2B for the treatment of neuropathic pain. *Neurotherapeutics.* 2009 6(4):693–702. Review. <https://doi.org/10.1016/j.nurt.2009.07.008> PMID: 19789073
42. Liu XJ, Salter MW. Glutamate receptor phosphorylation and trafficking in pain plasticity in spinal cord dorsal horn. *Eur J Neurosci.* 2010 32(2):278–89. <https://doi.org/10.1111/j.1460-9568.2010.07351.x> PMID: 20629726
43. Yan X, Jiang E, Gao M, Weng HR. Endogenous activation of presynaptic NMDA receptors enhances glutamate release from the primary afferents in the spinal dorsal horn in a rat model of neuropathic pain. *J Physiol.* 2013 591(7):2001–19. <https://doi.org/10.1113/jphysiol.2012.250522> PMID: 23359671
44. Wade KF, Marshall A, Vanhoutte B, Wu FC, O'Neill TW, Lee DM. Does Pain Predict Frailty in Older Men and Women? Findings From the English Longitudinal Study of Ageing (ELSA). *J Gerontol A Biol Sci Med Sci.* 2017 72(3):403–409. <https://doi.org/10.1093/gerona/glw226> PMID: 27836906
45. Wade KF, Lee DM, McBeth J, Ravindrarajah R, Gielen E, Pye SR, et al. Chronic widespread pain is associated with worsening frailty in European men. *Age Ageing.* 2016 45(2):268–74. <https://doi.org/10.1093/ageing/afv170> PMID: 26679698
46. Reimann F, Tolhurst G, Gribble FM. G-protein-coupled receptors in intestinal chemosensation. *Cell Metab.* 2012 15(4):421–31. <https://doi.org/10.1016/j.cmet.2011.12.019> PMID: 22482725

47. Martone AM, Onder G, Vetrano DL, Ortolani E, Tosato M, Marzetti E, Landi F. Anorexia of aging: a modifiable risk factor for frailty. *Nutrients*. 2013 5(10):4126–33. <https://doi.org/10.3390/nu5104126> PMID: [24128975](https://pubmed.ncbi.nlm.nih.gov/24128975/)