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1

2 **A novel glue attachment approach for precise anchoring of hydrophilic EGCG to**
3 **enhance the separation performance and antifouling properties of PVDF**
4 **membranes**

5

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8

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32

33 **Abstract**

34

35 A novel glue attachment approach was proposed to form a durable hydration layer
36 on a hydrophobic PVDF hollow fiber membrane (PVDF HFM) surface to improve its
37 hydrophilicity and antifouling ability during wastewater filtration. The functional glue
38 was synthesized from reclaimed styrene butadiene rubber (SBR) and a hydroxyl group
39 was created with an epoxidation reaction (ESBR). The hydrophilic epigallocatechin-s-
40 gallate (EGCG) was then precisely anchored via hydrogen bonding with multiple
41 phenolic hydroxyl groups in the ESBR without penetrating into the inner matrix of the
42 PVDF to prevent flux decline. The hydrophilicity of the PVDF membrane increased
43 drastically and the water contact angle decreased from 62.7° to 45.1° with only a 25%
44 decline in the pure water flux. Furthermore, due to precise anchoring of the EGCG, the
45 modified EGCG-ESBR/PVDF membrane showed a higher pure water flux (110.6 L m⁻²
46 h⁻¹) and much higher BSA and oil (kerosene) rejection rates (approximately 94.5%
47 and 99.5%, respectively) compared to membranes directly coated with EGCG (EGCG-
48 PVDF). Moreover, the modified membrane also showed higher water flux recovery
49 after multiple filtration cycles. This promising and efficient hydrophilic modification
50 suggests great potential for application of the eco-friendly material in wastewater
51 treatment.

52 *Keywords:* Hydrophilic surface modification; Reclaimed rubber; Epigallocatechin
53 gallate; Hydrogen bond; Anti-fouling performance

54

55 **1. Introduction**

56

57 Membrane filtration is a renewable technique for treating effluent containing small
58 and nondegradable contaminants after biological treatment processes, and it features
59 low energy consumption, an environmentally friendly process, and ease of operation [1,
60 2]. With the development of membrane technology, various functional materials are
61 emerging for wastewater treatment. As a desirable material, polyvinylidene fluoride
62 (PVDF)-based ultrafiltration (UF) membranes have been widely applied in various
63 industries, such as the dairy industry (biomacromolecule filtration) and oil-water
64 suspension applications, because of their excellent mechanical strength, superior
65 thermal stability and good chemical properties [3]. However, the intrinsic hydrophobic
66 property of PVDF membranes is the major barrier. Serious fouling easily occurs on the
67 membrane surface due to the attraction of biometrics, such as proteins and organic
68 species, resulting in a high frequency of membrane cleaning, performance decline, and
69 lifetime shortening [4, 5]. In this regard, many experts are devoted to developing new
70 strategies for achieving high permeability and excellent anti-fouling ability in real

71 applications.

72 Hydrophilic surface modification is a known method for enhancing the
73 performance of PVDF membranes by linking the hydrophilic modifier onto the
74 membrane surface via physical coating/deposition [6-8] and chemical bonding [9-13],
75 such as molecules with functional groups (hydroxyl, amine and sulfonic acid groups:
76 PEG, PVA, PVP, PEGMA, chitosan, catechin, dopamine, trimethoxysilane, etc. [14-23].
77 The modification solution generally treats the membrane via the dip coating or
78 immersion method [24, 25], which have reduced likelihood of defect formation in the
79 membrane matrix compared to conventional methods (such as the blending method [26-
80 29]). Despite these positive developments, a few challenges remain, especially a
81 negative effect on permeability. Sun and Feng [30] prepared a high rejection of amine-
82 modified membranes by doping an amine solution. These membranes exhibited better
83 surface hydrophilicity but significantly lower water permeability due to the blocking of
84 pores in the membrane. Wang et al. [8] developed a strategy for coating functional
85 cellulose molecules on PVDF membranes, which improved the hydrophilicity and
86 antifouling performance of membranes used in protein filtration. However, over-
87 aggregation of the cellulose molecules occurred inside the membrane pores, negatively
88 impacting water permeability due to the narrow pathway. As discussed above, surface
89 hydrophilic modification generally leads to a decline in water permeance due to the

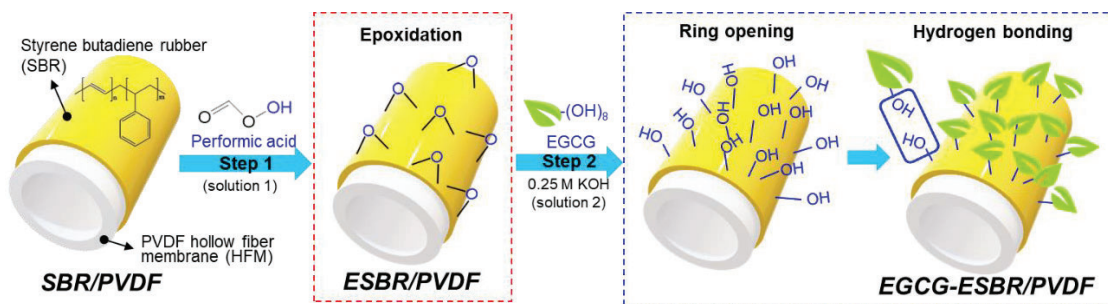
90 denser structure formed by an extra layer on the membrane surface or inside the pores.

91 In addition, the chemical processability of the PVDF membrane should be
92 considered. Several studies have attempted to modify the molecular structures of PVDF
93 membranes directly with grafting methods designed to effectively improve the surface
94 hydrophilicity and separation performance. However, the stable PVDF molecular
95 chains must be activated first by forming hydroxyl groups with, for example, alkali
96 treatment [22, 31] and irradiation treatment (UV, plasma and high-energy electron beam)
97 [3, 32, 33]. The pretreatment process is required for the grafting reaction of the PVDF
98 membrane, which could limit development due to the complicated and expensive
99 process. Therefore, to overcome these shortcomings, it is necessary to develop an
100 environmentally friendly, low-cost, and sustainable hydrophilic modification
101 technology to expand the application potential.

102 To address these issues, we designed a novel, facile and effective surface
103 modification method for fabricating a highly hydrophilic and anti-fouling PVDF HFM
104 using eco-friendly and inexpensive materials and a green glue attachment approach.
105 Plant-derived epigallocatechin-3-gallate (EGCG) is one of the major tea catechins in
106 green tea extracts and is a common polyphenol material, and it exhibits good
107 hydrophilicity due to the abundance of hydroxyl groups [34, 35]. To deposit EGCG
108 onto the PVDF membrane surface controllably without penetrating the inner matrix, we

109 sought to design a glue layer to react with EGCG via hydrogen bonding. Reclaimed
110 styrene butadiene rubber (SBR) derived from waste tires was used as the glue precursor.

111 As shown in Scheme 1, the glue precursor SBR was first coated onto the PVDF
112 HFM surface and went through the epoxidation reaction with peroxyformic acid to form
113 an epoxidized layer ESB. The ESB shows high thermal stability and hydrophilicity
114 due to the oxirane group [36, 37]. The epoxide (oxirane ring) group in this ESB layer
115 is transformed to a hydroxyl group under basic conditions when the EGCG is added,
116 which functions as a precise anchor for the *in situ* reaction with EGCG. Thus, in this
117 reaction, hydrophilic EGCG was effectively reacted with ESB through multiple
118 hydrogen bonds between the hydroxyl groups of ESB and the phenolic hydroxyl
119 groups in EGCG. The ESB acts as a functional glue layer to react precisely with
120 EGCG to form a hydrophilic layer on the membrane surface without clogging its inner
121 pores. Moreover, the strength of this interaction between EGCG and ESB could
122 endow the modified membrane with a durable performance during the filtration process.
123 Thus, in this work, the time required for the epoxidation reaction was optimized, and
124 the effects of the modified membrane on the filtration efficiencies for separation of
125 proteins and oily pollutants from wastewater were evaluated.



126

127 **Scheme 1.** Schematic illustration of the fabrication of the EGCG-ESBR/PVDF

128 membrane via the *in situ* modification approach.

129

130 2. Experimental

131

132 2.1 Material

133

134 A commercial hollow fiber PVDF membrane was obtained from Ray-E Creative
 135 Co., Ltd. (Taipei, Taiwan), and the examined PWF was $\sim 150 \text{ L/m}^2\text{h}$, the BSA rejection
 136 was approximately 90%, and the flux recovery rate was 80.2%. Styrene butadiene
 137 rubber (SBR) was fully regenerated from discarded inner tires [38]. Toluene (purity of
 138 99.8%), methanol (purity of 99.9%), ethanol (EtOH, purity of 99.8%), Tween 60 (purity
 139 of 100%), kerosene (purity of >99%) and oil red O (MW = 408.49 g/mol) were
 140 purchased from Sigma–Aldrich. Reagent grade potassium hydroxide (KOH, purity of
 141 85%) was supplied by Duksan Pure Chemicals Co. Ltd.

142 Bovine serum albumin (BSA) (MW = 67,000 g/mol), which was purchased from
143 Sigma Aldrich, was used as a model protein foulant. The BSA feed was prepared in
144 phosphate-buffered saline (PBS) solution (pH = 7) at a concentration of 1.0 mg/ml.
145 EGCG (purity of >98%) was provided by TCI (Shanghai) Development Co. Ltd.

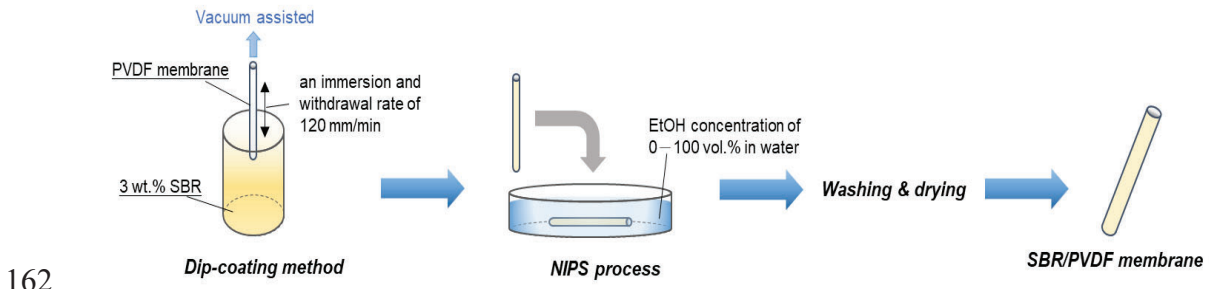
146

147 *2.2 Fabrication of the hollow fiber SBR/PVDF composite membrane*

148

149 The SBR glue layer on the PVDF HFMs was prepared by a vacuum-assisted dip-
150 coating method [39] and a nonsolvent-induced phase separation (NIPS) process. As
151 shown in Fig. 1, 3 wt.% SBR was first introduced into toluene and magnetically stirred
152 overnight until the solutions were homogeneous (the optimal SBR concentration was
153 chosen by the water permeance test and SEM surface image observed, as shown in Fig.
154 S1). Two end sides of the dried PVDF membranes sealed with an epoxy resin were
155 coated with SBR solution by the dip-coating method at an immersion and withdrawal
156 rate of 120 mm/min at a vacuum degree of 1.0 atm for 30 s. Next, the membranes were
157 immersed in a coagulating bath (with the concentration of EtOH in water ranging from
158 0–100 vol%) for phase inversion and then treated with methanol and n-hexane to
159 completely remove the residual solvent by solvent exchange. Finally, the composite
160 membranes were dried under ambient conditions and employed in the hydrophilic

161 modification process.



162
163 **Fig. 1.** Preparation of the SBR/PVDF membrane by nonsolvent-induced phase
164 separation.

165

166 2.3 Hydrophilic modification processes

167

168 The surface of the SBR/PVDF composite membrane was further modified with
169 hydrophilic chemical groups. The modification included a two-step reaction: (1) the
170 SBR skin layer of the composite membrane was epoxidized to form the ESBP/PVDF
171 membrane, and then (2) EGCG was coated onto the membrane surface via *in situ*
172 deposition to prepare the EGCG-ESBR/PVDF membrane. Table S1 summarizes the
173 fabrication conditions of the resulting membranes via the aforementioned modification
174 processes.

175 In the first step, the SBR intermediate layer was epoxidized *in situ* with an aqueous
176 mixture of formic acid (>98 vol%) and hydrogen peroxide (30 vol%) with a mole ratio
177 of 1.5. The dried SBR/PVDF membranes were immersed in the mixture for 0.5-3 h,

178 and the excess solution was drained to obtain the ESBP/PVDF membrane. Next, the
179 EGCG was dissolved in a 0.25 M KOH solution at 85 °C overnight and magnetically
180 stirred to obtain a homogeneous coating solution containing 0.1 mg/ml EGCG. The
181 pristine membrane or the epoxidized membranes were immersed in the EGCG solution,
182 and the deposition reaction was performed for 2 to 12 h. The membrane was then
183 washed with distilled water until a neutral pH was reached. Afterward, the membranes
184 were dried overnight in an oven at 60 °C to obtain the EGCG-PVDF membrane or
185 EGCG-ESBR/PVDF membrane.

186

187 *2.4 Characterization methods*

188

189 The surface morphologies and the elemental maps of the membranes were
190 observed with field emission scanning electron microscopy (FE-SEM; Model JEOL
191 JSM-6700F from Hitachi Co., Japan) and an energy dispersive X-ray (EDX) detector,
192 respectively. The surface hydrophilicity of the membranes was determined by a contact
193 angle (CA) measuring instrument (KRÜSS Instruments, Mobile Drop GH11). All
194 reported CA data were obtained by using the sessile drop technique with deionized
195 water droplets; an average of three drops was applied to each surface. The surface
196 roughnesses of the membrane were analyzed by atomic force microscopy (AFM;

197 Bruker Dimension Icon, Bruker, MA, USA). The chemical structure of the membranes
198 was measured by attenuated total reflectance Fourier transform infrared (ATR-FTIR)
199 spectroscopy using an FTIR-4100 spectrophotometer (JASCO, USA). All transmission
200 spectra were recorded in the range of 4000–600 cm^{-1} at a scanning rate of 10 kHz with
201 a total of 16 scans. The molecular weight cutoffs (MWCOs) of the membranes were
202 assessed with rejection of polyethylene glycol (PEG) solutes with various molecular
203 weights ranging from 1 to 300 kDa in a 500 mg L^{-1} feed solution. A photometer (WTW,
204 photoLab® S6, Germany) was used to determine the PEG concentrations of the feed
205 solutions and the permeate after filtration for 1 h. The rejection rate for each solute was
206 calculated with equation (2) and plotted against the molecular weight. Finally, the
207 MWCO of the membrane was estimated from 90% solute removal.

208

209 2.5 *Assessment of filtration and antifouling performance*

210

211 A lab-scale cross-flow filtration system step-up was used to evaluate the
212 permeation and separation performances of the fabricated membranes. A membrane
213 module consisting of a fiber with an effective length of 10 cm was used. The hollow
214 fiber membranes were initially prepared by using deionized water at 2.5 bar for 30 min
215 in advance of the test. After compaction, the pure water flux (PWF, J_w , $\text{L m}^{-2} \text{h}^{-1}$) was

216 measured under a pressure of 2 bar and calculated with equation (1) [40]:

$$PWF (J_w, L \cdot m^{-2} \cdot h^{-1}) = \frac{V}{A \cdot \Delta t} \quad (1)$$

217

218 where V is the volume of permeate (L) collected in the determined time interval, Δt (h),

219 and A is the effective membrane area (m^2).

220 A filtration test with a model foulant was subsequently performed to measure the

221 anti-fouling properties of the resulting membrane. The foulant feed included both 1

222 mg/ml BSA aqueous solution for protein filtration studies and 1 mg/ml Tween 60-

223 stabilized kerosene-in-water emulsions for the oil/water separation studies. During

224 filtration, the permeate flux (J_f) was measured. The foulant rejection (R) was obtained

225 by using a UV-visible spectrophotometer (U-3900 Spectrophotometer from Hitachi

226 Co., Japan) to analyze the concentrations of foulant in the feed (C_f , mg/L) (wavelength

227 at 280 nm for BSA protein, at 518 nm for oil red O) and the collected permeate (C_p ,

228 mg/L), and R was calculated with equation (2) [41]:

229

$$Rejection (R, \%) = \frac{C_f - C_p}{C_f} \times 100\% \quad (2)$$

230

231 After filtration, the membranes were cleaned by backwashing with pure water for
 232 30 min. The pure water flux (J_w) of the cleaned membrane was then reevaluated. To
 233 determine the antifouling properties of the membranes during the filtration process, the
 234 membrane fouling (R_m), total fouling (R_t), and irreversible and reversible fouling (R_{ir}
 235 and R_r) were calculated with equations (3), (4), (5), and (6), respectively [42]:

236

$$\text{Membrane fouling } (R_m, \text{m}^{-1}) = \frac{\Delta P}{\mu \times J_w} \quad (3)$$

$$\text{Total fouling } (R_t, \text{m}^{-1}) = \frac{\Delta P}{\mu \times J_f} \quad (4)$$

$$\text{Irreversible fouling } (R_{ir}, \text{m}^{-1}) = \frac{\Delta P}{\mu \times J_{w,i}} \quad (5)$$

$$\text{Reversible fouling } (R_r, \text{m}^{-1}) = R_t - R_m - R_{ir} \quad (6)$$

237

238 where ΔP is the transmembrane pressure (Pa); μ is the permeate viscosity (Pa s); $J_{w,i}$ is
 239 the PWF before foulant filtration; and the flux recovery rate (FRR), total flux decline
 240 ratio (FDR) and irreversible flux decline ratio (DR_{ir}) were calculated with equations (7),
 241 (8), and (9), respectively [43]:

242

$$\text{FRR } (\%) = \frac{J_{w,2}}{J_{w,1}} \times 100\% \quad (7)$$

$$\text{FDR } (\%) = \left(1 - \frac{J_f}{J_{w,1}}\right) \times 100\% \quad (8)$$

$$\text{DR}_{ir} (\%) = \left(1 - \frac{J_{w,2}}{J_{w,1}}\right) \times 100\% \quad (9)$$

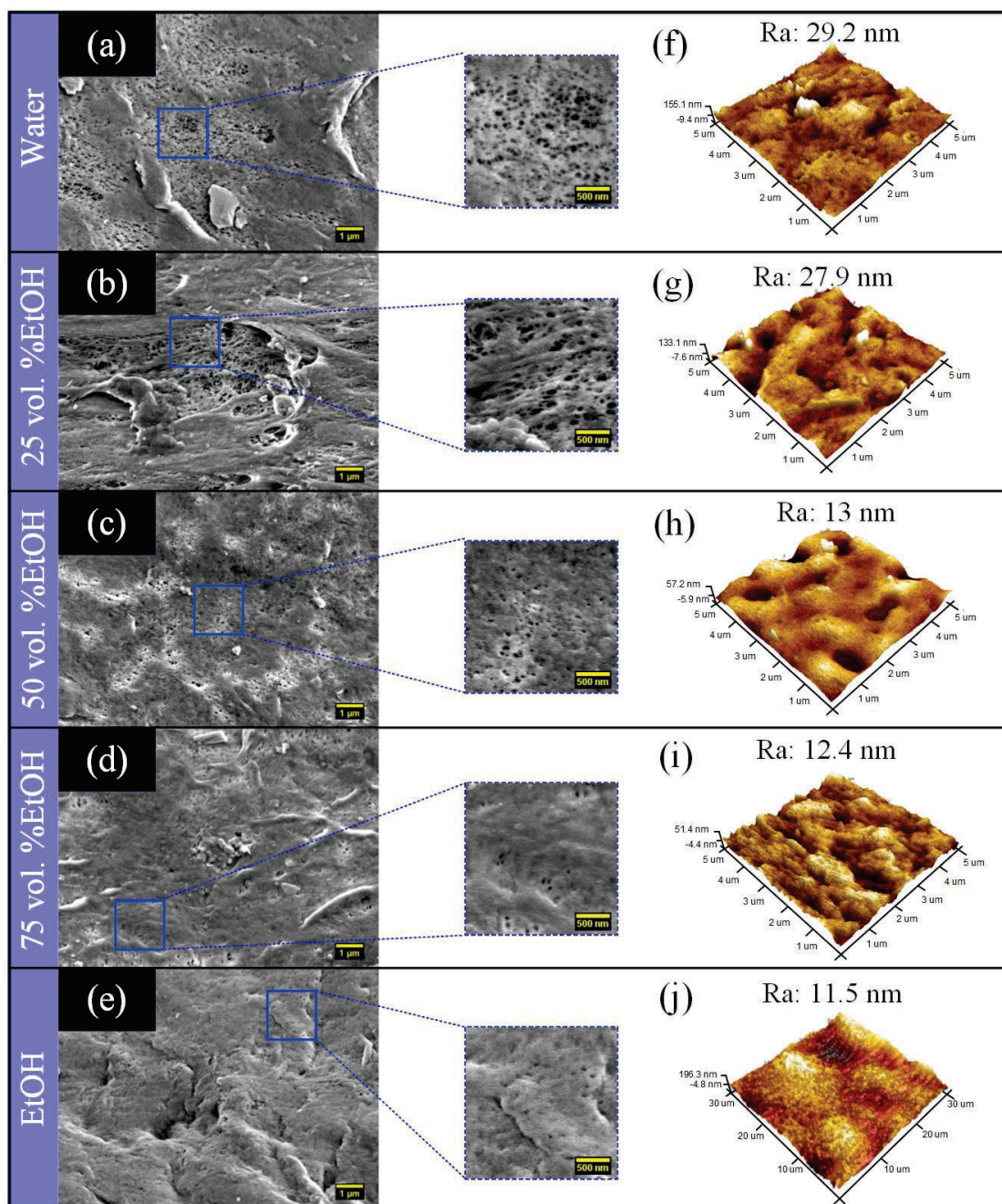
243 **3. Results and discussion**

244

245 *3.1 Characterization and separation performance of the SBR/PVDF composite*
246 *membrane*

247

248 To investigate the influence of nonsolvents on the surface morphology of the
249 membranes in the NIPS process, AFM and FE-SEM were used to evaluate the surface
250 properties of the composite SBR/PVDF membranes (Fig. 2). The SBR glue layer
251 fabricated in the nonsolvent with a low EtOH concentration (a and b) had an irregular
252 surface structure, and macropores were observed. Fig. 2c-e shows the membranes using
253 a 50 vol% and 75 vol% EtOH-water mixture and pure EtOH as nonsolvents. The surface
254 became smoother, and the size and quantity of pores were reduced with increasing
255 EtOH concentration. Similar results were obtained from AFM images of the change in
256 surface roughness (Fig. 2f-j). The average roughness (Ra) of the membrane surface
257 decreased as the EtOH concentration was increased. The presence of EtOH in a
258 nonsolvent can lead to the formation of a relatively smooth membrane surface. The
259 obtained surface morphologies were in good agreement with the results of the protein
260 separation experiment.



261

262 **Fig. 2.** FE-SEM and AFM images of the composite SBR/PVDF membranes coagulated

263 in different nonsolvent baths. (a and f): deionized water; (b and g) 25 vol %EtOH: 75

264 vol % water; (c and h) 50 vol % EtOH: 50 vol % water, (d and i) 75 vol % EtOH: 25

265 vol % water and (e and j) EtOH.

266

267 Solubility in a solvent may affect the liquid–liquid demixing speed during phase
 268 inversion [44]. The miscibility of the solvent and nonsolvent affects membrane
 269 formation, which can be determined by calculating the difference in the solubility
 270 parameters (δ , $\text{MPa}^{1/2}$) of the solvent and nonsolvent. The following equation can be
 271 used to predict the interaction parameters between solvent and nonsolvent [45]:

$$\delta_t = \sqrt{\delta_d^2 + \delta_p^2 + \delta_h^2} \quad (10)$$

$$\delta_d = x_1\delta_{d1} + x_2\delta_{d2} \quad (11)$$

$$\delta_p = x_1\delta_{p1} + x_2\delta_{p2} \quad (12)$$

$$\delta_h = x_1\delta_{h1} + x_2\delta_{h2} \quad (13)$$

272 where δ_t is the total solubility parameter, and the contribution can be divided into three
 273 parts: dispersion force (δ_d), polar force (δ_p) and hydrogen bonding force (δ_h); x_i is the
 274 volume fraction of chemical compound *i*. The solubility parameters of toluene and
 275 nonsolvent are presented in Table 1.

276

277 The difference in solubility parameter, $\Delta\delta_{\text{toluene-NS}}$, between the toluene of the dip-
 278 coating dope and the nonsolvent system of the coagulating bath can be further
 279 calculated:

280

$$\delta_{\text{toluene-NS}} = \sqrt{[(\delta_{d,\text{toluene}} - \delta_{d,\text{NS}})^2 + (\delta_{p,\text{toluene}} - \delta_{p,\text{NS}})^2 + (\delta_{h,\text{toluene}} - \delta_{h,\text{NS}})^2]} \quad (14)$$

281 As shown in Table 1, the solubility difference decreased with increasing EtOH
 282 concentration in the nonsolvent. The results were correlated with the surface structure
 283 of the membranes (shown in Fig. 2), and the higher solubility difference resulted in the
 284 formation of a macroporous structure. In contrast, a lower solubility difference caused
 285 the formation of a denser structure. In other words, the microporous structure can be
 286 inhibited by controlling the EtOH concentration in a nonsolvent. A similar change in
 287 the surface morphologies of the rubber membranes in the EtOH-water nonsolvent
 288 mixture was observed by Kuźmińska et al. [46]. Ren et al. [47] also studied the effect
 289 of EtOH on the membrane microstructure and concluded that the diffusivity in the
 290 solvent–nonsolvent system is a major controlling factor in the formation of the pore
 291 structure.

292 **Table 1** The solubility parameter (δ) in toluene solvent [48] and various nonsolvents
 293 [49]; differences in the solubility parameter in toluene–nonsolvent ($\delta_{toluene-NS}$)

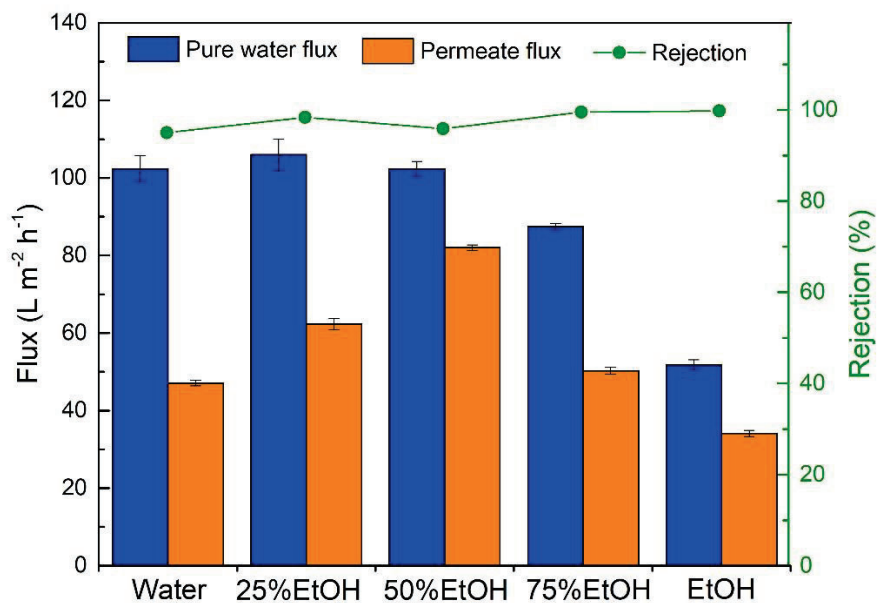
Chemical compound	Solubility parameter (MPa ^{1/2})				$\sqrt{\sum (\Delta\delta_{toluene-NS})^2}$
	δ_t	δ_d	δ_p	δ_h	
Toluene	18.2	18.0	1.4	2.0	-
H ₂ O	47.8	15.6	16.0	42.3	42.9
25% EtOH	42.2	15.7	14.2	36.6	36.9
50% EtOH	36.8	15.7	12.4	30.9	31.0
75% EtOH	31.5	15.8	10.6	25.1	25.0
EtOH	26.5	15.8	8.8	19.4	19.0

294 The pure water flux (PWF) and separation performance of the composite
 295 SBR/PVDF membranes fabricated in the different nonsolvents are shown in Fig. 3. The

296 composite membranes coagulated in the mixtures with low EtOH concentrations (0
297 vol.%, 25 vol.% and 50 vol.%) had a higher pure water flux, approximately 101-105 L
298 $\text{m}^{-2} \text{h}^{-1}$, while the mixtures with higher EtOH concentrations showed a significant
299 decrease in pure water flux. However, the results of protein separation presented a
300 different flux trend. The permeate flux of the membranes increased with the
301 concentration of EtOH, while the permeate flux showed a maximum value of 82 L m^{-2}
302 h^{-1} at 50 vol.% EtOH and then decreased to 50.3 L $\text{m}^{-2} \text{h}^{-1}$ at 75 vol.% EtOH and 34.1 L
303 $\text{m}^{-2} \text{h}^{-1}$ in pure EtOH. However, the BSA rejections of the different composite
304 membranes were nearly the same, approximately 95% or above, possibly because the
305 pore sizes of the membranes were all smaller than the BSA size.

306 There are two main reasons for the difference in the flux trend of pure water and
307 permeate. First, the high permeability of the membranes was in good agreement with
308 their macropores on the surface of the membrane, as confirmed by the SEM images
309 (Fig. 2) and mean pore size (Table 1). In contrast, further increases in the EtOH
310 concentration led to dense surface formation, resulting in a decrease in pure water flux.
311 Second, the higher roughness of the membranes, as noted recently by researchers [50],
312 can easily cause the penetration/adhesion of foulants on the rough surface. Therefore,
313 the resulting membrane at low EtOH concentrations had a lower permeate flux due to
314 fouling formation. According to the above results, a comparison of all membranes

315 showed that 50 vol.% EtOH was the most preferred choice as a nonsolvent because it
316 led to a smoother surface with a relatively higher porosity, so it was used to fabricate
317 subsequent membranes for hydrophilic modification.



318

319 **Fig. 3.** Fluxes and protein filtration efficiencies of the composite SBR/PVDF

320 membranes coagulated in different nonsolvent baths.

321 3.2 Effect of the EGCG coating on the hydrophilic modification of the EGCG-ESBR

322 layer

323

324 As shown in Fig. 4a, variations are apparent in the carbon double bond (C = C)

325 adsorption peak at 971 cm^{-1} . This peak started at a maximum value and then decreased

326 during the epoxidation process until it was completely lost from the reaction. The

327 oxirane groups had relatively weaker adsorption at 1183 cm^{-1} , corresponding to the

328 stretching of C–O–C, which increased with the epoxidation time. The adsorption of the

329 C=O groups at 1717 cm^{-1} increased simultaneously due to the side reaction that took

330 place during the epoxidation reaction [36].

331 We examined the optimal reaction time for modifying the surface hydrophilicities

332 of the membranes and estimated it with the water contact angle (WCA) of the

333 membrane surface, as shown in Fig. 4b. The pristine SBR/PVDF composite membrane

334 had a high WCA of 99%. With the increase in the reaction time, the WCA of the

335 modified membranes (ESBR/PVDF) dramatically decreased to 68.2° until the reaction

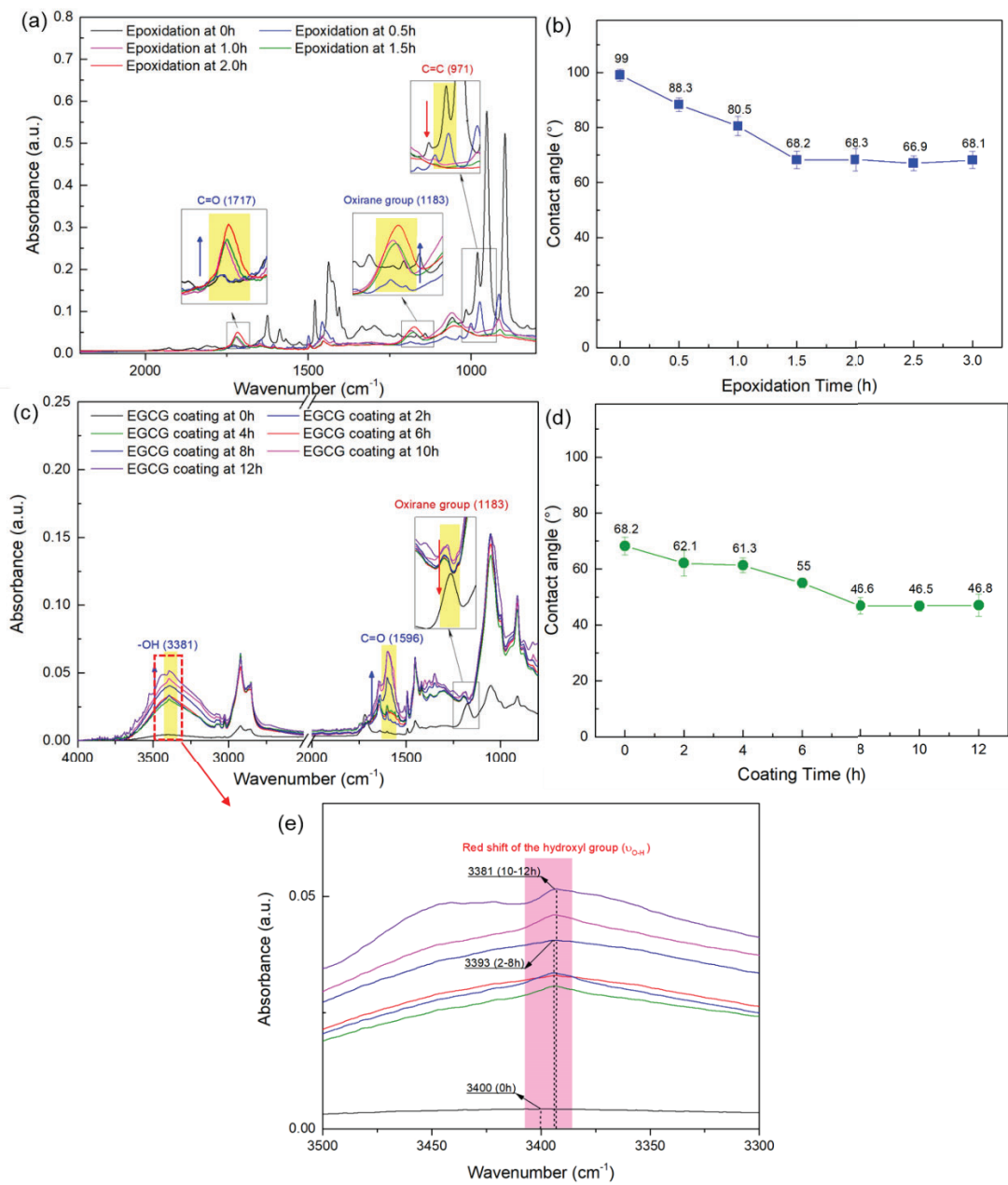
336 time for the epoxidation reaction reached 1.5 h. The decrease in WCA occurred because

337 the hydrophilic oxirane group was formed on the epoxidized membrane surface.

338 Nevertheless, the WCA remained constant when the reaction time was extended to 3 h.

339 Thus, the optimal epoxidation time of the ESBR/PVDF membrane was 1.5 h, and this

340 was used for the next modification step.



341

342 **Fig. 4.** Changes in the (a, c) ATR-FTIR spectra and (b, d) water contact angle (WCA)

343 of the composite membranes undergoing epoxidation and EGCG coating as a function

344 of the reaction time. (e) ATR-FTIR spectra of the EGCG coating on the ESBP/PVDF

345 membrane as a function of reaction time at wavenumbers of hydroxyl groups at 3500–

346 3300 cm⁻¹.

347 In the next reaction step, the FTIR spectra of the EGCG-coated membranes
348 (EGCG-ESBR/PVDF) made with different coating times are shown in Fig. 4c. The
349 EGCG-coated membrane has a relatively distinct absorption peak in the range of 3800-
350 3200 cm^{-1} , representing the stretching vibration of the OH group, and a weaker
351 absorption peak at 1695 and 1596 cm^{-1} due to the C=O functional group [51]. Both of
352 these bands increased in intensity with longer reaction times, verifying the presence of
353 EGCG on the membrane surface.

354 Fig. 4d shows the WCAs of the EGCG-ESBR/PVDF membranes. As expected, a
355 decreasing trend in the WCA was observed with increasing coating time from 2 h to 8
356 h, and the WCA of the modified membranes decreased to approximately 46° and
357 remained constant for 8 to 12 h. Furthermore, the change trend of hydrophilicity can be
358 proven by observing the interaction of hydrogen bonds between EGCG and ESBR in
359 the FTIR spectra. As shown in Fig. 4e, the absorption peak of the hydroxyl group at
360 3400 cm^{-1} in the spectrum of the ESBR/PVDF membrane is redshifted to 3393 cm^{-1}
361 and 3381 cm^{-1} after coating times of 2-8 h and 10-12 h, respectively. It is suggested that
362 the combination of EGCG and ESBR shows a successful adhesive interaction of ESBR
363 and EGCG through an effective hydrogen bond. The results showed that enhanced
364 hydrophilicity was acquired with the increase in EGCG deposition due to the presence
365 of a hydroxyl group on the resultant membrane surface. Therefore, 8 h is the most

366 appropriate coating time for fabricating a highly hydrophilic membrane.

367

368 3.3 Mechanism of the modification reaction

369

370 The possible mechanism of the proposed two-step modification reaction is

371 illustrated in Fig. 5. In the first step, the epoxidation reaction occurs on the surface of

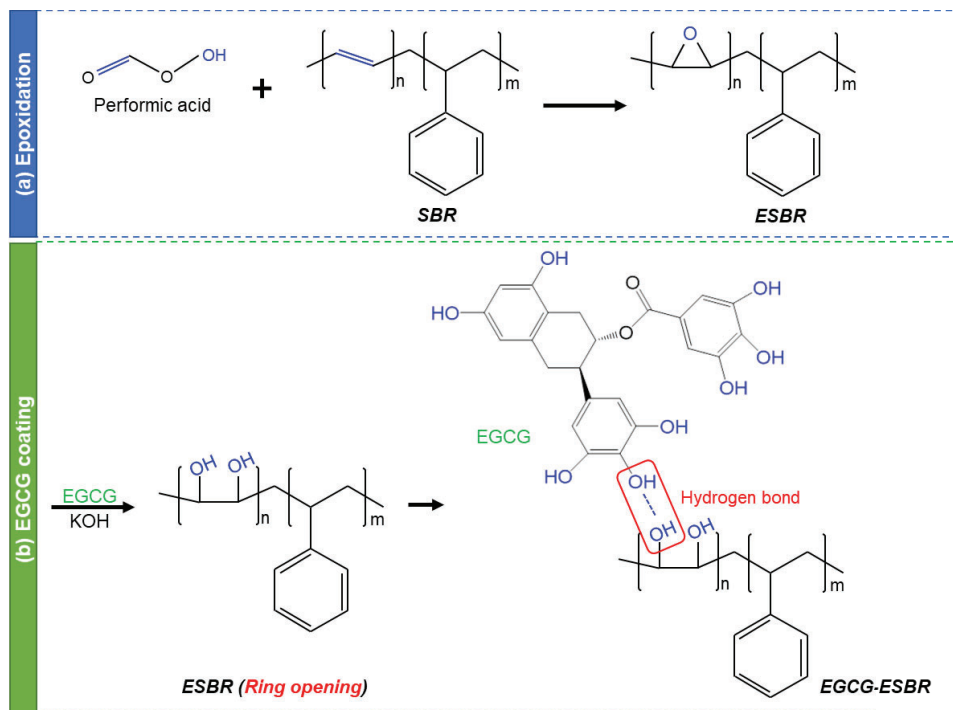
372 the SBR glue layer, resulting in the formation of oxirane groups. In the second step, the

373 basicity can first induce a ring-opening reaction of the oxirane groups of the ESBR glue

374 to form hydroxyl groups, which react with the hydroxyl groups of EGCG during a

375 condensation reaction (click reaction), enabling hydrogen bonding between EGCG and

376 the membrane surfaces.



377

378 **Fig. 5.** Synthetic pathway for (a) the epoxidation of SBR glue and (b) coating of the

379 EGCG.

380 3.4 Effect of hydrophilic modification on surface properties and separation

381 performance

382

383 The pristine PVDF membrane was used as a model to investigate the impact of a

384 straight EGCG coating (EGCG-PVDF) and an ESBR intermediate layer-assisted

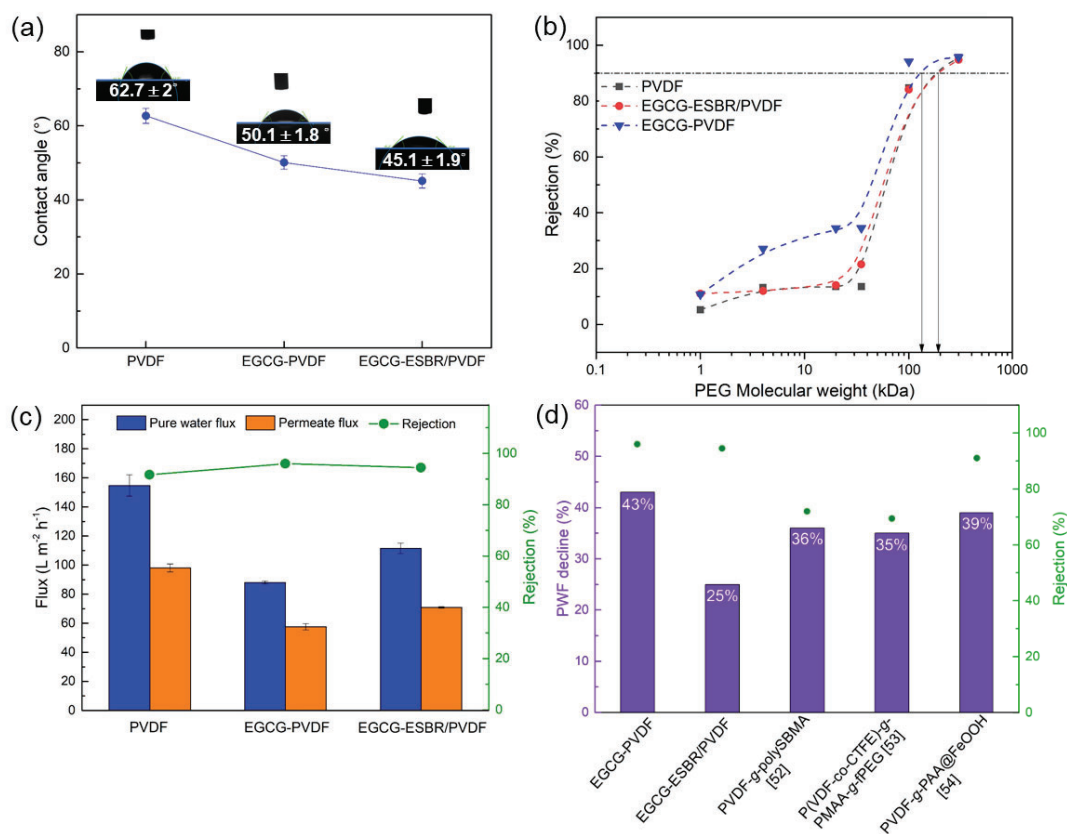
385 EGCG coating (EGCG-ESBR/PVDF) on the surface properties and separation

386 performance. As shown in Fig. 6a, the presence of EGCG on the modified membranes

387 decreased from $62.7 \pm 2^\circ$ for the PVDF membrane to $50.1 \pm 1.8^\circ$ and $45.1 \pm 1.9^\circ$ for the

388 EGCG-PVDF and EGCG-ESBR/PVDF membranes, respectively.

389



390

391 **Fig. 6.** (a) WCAs of the pristine and modified membranes; (b) retention curves of the
392 resulting membranes used to determine the apparent MWCO values; (c) fluxes and BSA
393 rejection rates of the pristine and modified membranes; (d) comparison of recently
394 reported materials with hydrophilic-modified PVDF membranes (PVDF-*g*-polySBMA
395 [52], both P(VDF-co-CTFE)-*g*-PMAA-*g*-fPEG [53] and PVDF-*g*-PAA@FOOH [54]
396 in 1.0 mg/ml BSA solutions).

397

398 To determine the molecular weight cutoff (MWCO) of the pristine and modified
399 membranes, different molecular weights of polyethylene glycols (PEGs) (1, 4, 20, 35,
400 100 and 300 kDa) were used as feed solutions in pressure filtration tests. As shown in
401 [Fig. 6b](#), all of the membranes showed MWCO values of approximately 100 to 300 kDa.
402 The MWCO value of the EGCG-ESBR membrane was similar to that of the pristine
403 membrane. However, the EGCG-PVDF membrane showed a lower MWCO value than
404 the other membranes, consistent with the surface morphology observed by SEM images
405 ([Fig. S2d-f](#)). As expected, the pore size of the membrane surface decreased when
406 EGCG was directly coated onto the pristine membrane ([Fig. S2e](#)). In contrast, for the
407 EGCG-ESBR/PVDF membrane, the surface morphology still maintained the pore
408 structure after EGCG coating ([Fig. S2f](#)). As illustrated in [Fig. S3](#), the chemical
409 elemental compositions of the fabricated membrane were measured by using an energy

410 dispersive spectroscopy (EDS) unit attached to the SEM. This result clearly indicated
411 the presence of higher O signals in the bulk of the EGCG-PVDF membrane than in the
412 EGCG-ESBR/PVDF membrane. This result proved that the ESBR glue layer prevented
413 deposition of the EGCG inside the pores and that an EGCG layer was efficiently
414 deposited onto the membrane surface, which altered the surface properties.

415 The effect of the SBR glue layer on the hydrophilic modifications of the PVDF
416 membrane performance is shown in Fig 6c. The PWF and BSA permeate fluxes of the
417 EGCG-PVDF membrane were significantly decreased by 43% and 41%, respectively,
418 compared to those of the pristine membrane. In marked contrast, the EGCG-
419 ESBR/PVDF membrane showed only slight reductions of 25% and 26% for the PWF
420 and permeate fluxes, respectively. Moreover, the BSA rejection rate of the membrane
421 increased from 91.7% for the PVDF membrane to 95–96% for the EGCG-ESBR/PVDF
422 membrane, which indicated a higher rejection rate with a lower flux decline. As shown
423 by the above surface analyses, the coated ESBR glue layer assisted in the deposition of
424 the EGCG selective layer on the membrane surface. Thus, EGCG effectively improved
425 the separation performance of the PVDF membrane without a negative impact on the
426 flux resulting from pore clogging. The performance of the modified membrane was
427 compared to that for the hydrophilic-modified PVDF membrane, as shown in Fig. 6d.
428 The EGCG-ESBR/PVDF membrane not only showed a lower decline in pure water

429 flux than the other modified membranes but also showed a competitive rejection rate
430 during BSA filtration. More importantly, our modified membrane does not use
431 hydrophilic modifiers, which are expensive or difficult to obtain and require
432 cumbersome and high-energy fabrication processes. Therefore, the coated ESBR glue
433 layer improved the pore structure and hydrophilicity of the membrane surface, which
434 improved the permeability after modification.

435

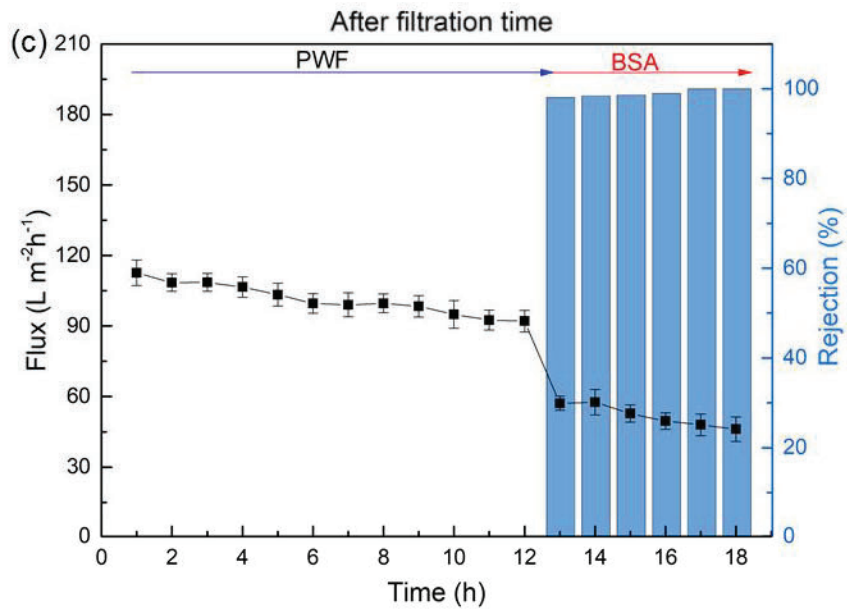
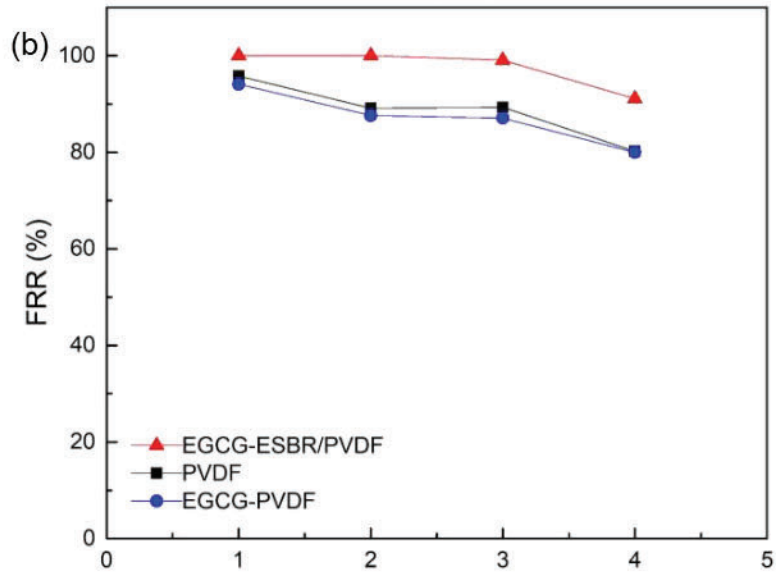
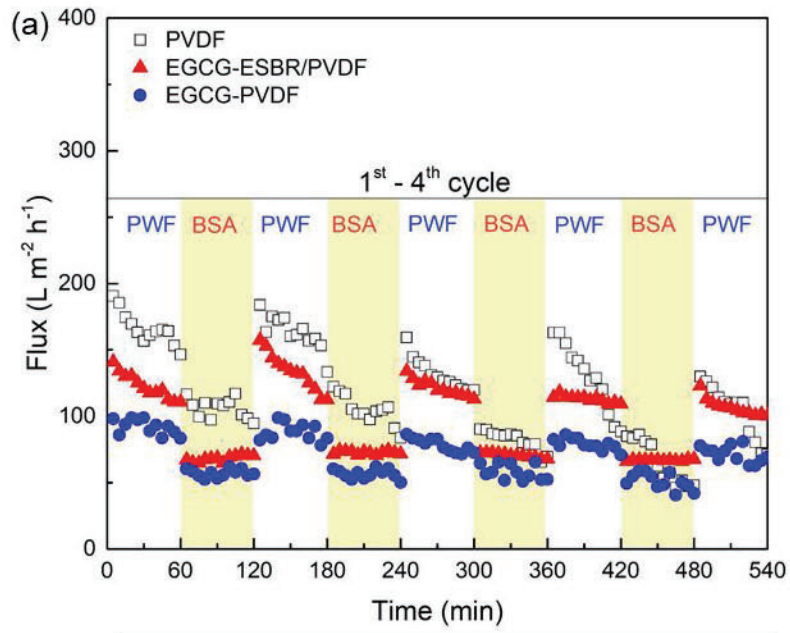
436 *3.5 Anti-fouling performance and the reliability test*

437

438 To further investigate the anti-fouling properties of the membrane, the cyclic
439 filtration performances were evaluated, as shown in Fig. 7a. During each cycle, the BSA
440 permeate flux of the pristine and modified membranes declined rapidly at the start of
441 filtration, especially the pristine membrane, which experienced an obvious decrease in
442 PWF and BSA permeate flux over time during each filtration cycle. Generally, rejected
443 protein molecules were easily deposited on the PVDF membrane surface and even
444 inside the membrane pores. This implied that the membrane underwent severe and
445 irreversible membrane fouling due to adsorption/deposition of the proteins [27, 55].
446 Membrane fouling was aggravated, and the FRR value of the pristine membrane had

447 subsequently decreased from 95.8% to 80.2% after the fourth filtration cycle, as shown
448 in Fig. 7b.

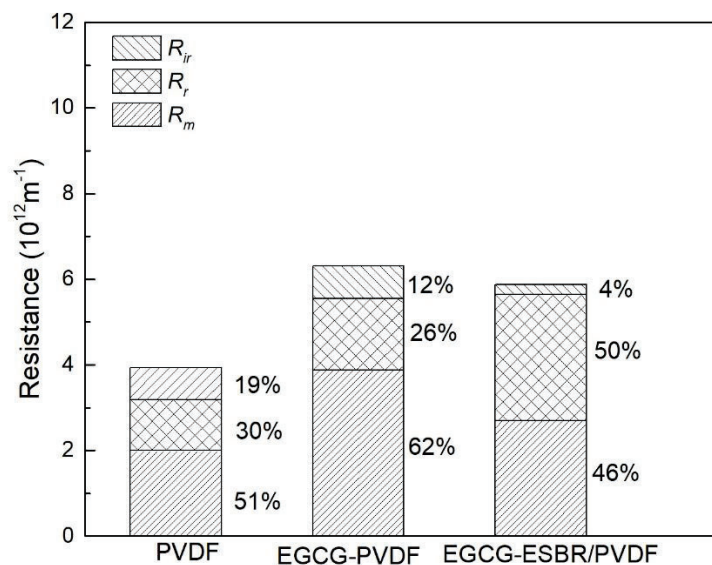
449 Compared to the pristine membrane, the permeate flux of the modified membrane
450 can be maintained at an appropriate value during operation, especially the EGCG-
451 ESBR/PVDF membrane. In marked contrast, the EGCG-ESBR/PVDF membrane
452 showed a higher permeate flux during the fourth filtration cycle than the pristine
453 membrane (Fig. 7a). Moreover, as shown in Fig. 7b, the EGCG-PVDF membrane
454 demonstrated a similar trend in the FRR value to the pristine membrane, while the
455 EGCG-ESBR/PVDF membrane showed only a slight decrease in the FRR value from
456 99.9% to 91.1% after the fourth cycle (Fig. 7b). This was due to the introduction of the
457 ESBR glue layer, which precisely anchored the hydrophilic layer on the membrane
458 surface through hydrophilic modifications with the EGCG coating. Thus, the EGCG-
459 ESBR hydrophilic layer exhibited the benefit of flux recovery and, after BSA filtration,
460 the pure water flux of the membrane was restored to its original value by simple
461 flushing.



463 **Fig. 7.** (a) The fluxes of the pristine PVDF, EGCG-PVDF and EGCG-ESBR/PVDF
464 membranes in the 4-cycle protein filtration experiments; (b) FRR values after different
465 numbers of filtration cycles. (c) The long-term filtration stability evaluation of the
466 EGCG-ESBR/PVDF membrane (after the cyclic filtration test) during a continuous
467 operation for pure water permeation and BSA filtration (The membrane has been stored
468 under atmospheric air over 6 months).

469

470 The resistance-series model was used to quantitatively analyze the antifouling
471 properties of the membrane in detail, as shown in Fig. 8. The total resistance (R_t) of the
472 EGCG-PVDF membrane was the highest compared to those of the other membranes
473 due to the larger proportion of membrane resistance (R_m) at 62%. Generally, the
474 membrane resistance to fouling depends on the surface morphologies of the membrane
475 [56]. Thus, this result indicates that the decrease in the pore size of the membrane
476 caused by the direct EGCG coating increases the resistance in the membrane, which
477 could result in a dramatic decrease in the permeance of the membrane.

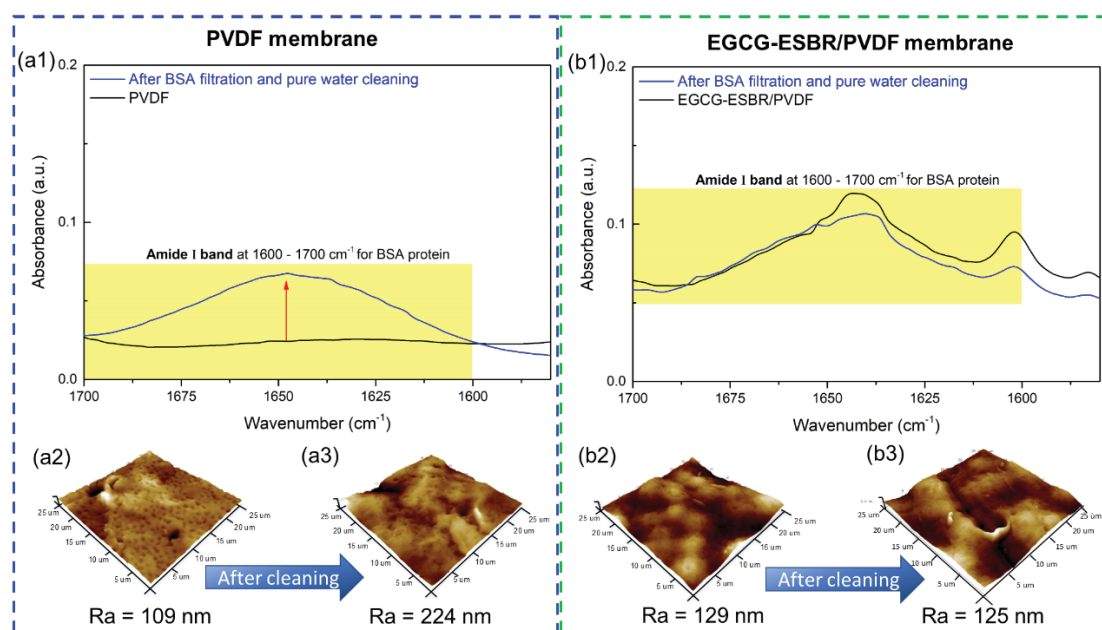


478

479 **Fig. 8.** Resistance analysis of PVDF, EGCG-PVDF and EGCG-ESBR/PVDF for BSA
 480 filtration.

481 The EGCG-ESBR/PVDF membrane exhibited a higher reversible resistance (R_r)
 482 of 50% and a lower membrane resistance (R_m) of 46% despite having an increase in
 483 total resistance compared to the pristine membrane. Moreover, the irreversible
 484 resistance (R_{ir}) significantly decreased to 4%, in contrast to that of the EGCG-PVDF
 485 membrane. The surface morphologies and hydrophilic characterization results
 486 confirmed that EGCG can bind onto only the near-skin layer rather than the inner-pore
 487 surface during the coating process with the help of the ESBR glue on the surface of the
 488 PVDF membrane. The above result was further verified by an observation of membrane
 489 surface properties. The FTIR spectra and AFM images of the PVDF membrane and the
 490 EGCG-ESBR/PVDF membrane are shown in Fig. 9. It can be clearly seen from FTIR
 491 spectra for the fouled membranes (after the cyclic filtration test) cleaned by water, the

492 amide I band at 1600 to 1700 cm^{-1} was still observed on the surface of the cleaned
 493 PVDF membrane, which is related to the BSA protein residues [57]. Moreover, the
 494 surface roughness of the cleaned PVDF membrane was significantly increased to 224
 495 nm from 109 nm of the pristine PVDF membrane. In contrast, the cleaned EGCG-
 496 ESBR/PVDF membrane didn't be observed the visible intensity of BSA protein in the
 497 FTIR spectra as well as an increase in roughness on the membrane surface after long-
 498 term operation. However, it is noteworthy that the membrane roughness has slightly
 499 decreased instead of increasing, which could be attributed to the compacted membrane
 500 during the cyclic filtration rather than the effect of residual solutes. It means that
 501 EGCG-ESBR layer is beneficial to suppressing the residual BSA protein on the
 502 membrane surface.



503

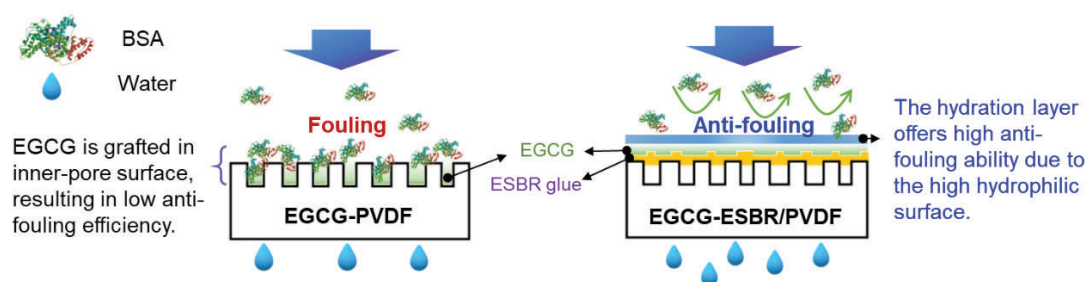
504 **Fig. 9.** FTIR spectra (a1 and b1) and AFM images (a2, 3 and b2, 3) of the PVDF and
505 the EGCG-ESBR/PVDF membranes.

506

507 The long-term stability of the EGCG-ESBR/PVDF membrane was also further
508 evaluated through a continuous operation time at 12 h for pure water permeation and
509 then 6 h for BSA filtration, respectively. This membrane has been tested the cyclic
510 filtration and then stored under atmospheric air over 6 months. As shown in Fig. 7c, the
511 long-term stored membrane maintained a similar PWF and permeate flux after the long-
512 term storage. Moreover, the rejection of membrane during long-term BSA filtration was
513 a constant value of >98%. However, a slight decrease in the flux was observed during
514 long-term operating time, which could be attributed to the transport model in the
515 outside-in system. The membrane is expected to be more compact during the long-term
516 operation, resulting in an increase in transport resistance. This phenomenon has been
517 verified by the above AFM roughness observed (Fig. 7b2 and b3). Overall, although
518 the performance was not as ideal as expected due to the operating condition limitation,
519 the modified membrane showed good stability for long-term continuous operation.

520 A schematic diagram of the fouling process is displayed in Fig 10. The EGCG-
521 PVDF membrane shows the worst surface property because of the inaccurate deposition
522 of EGCG on the surface of the PVDF membrane. EGCG could not effectively increase

523 the hydrophilicity of the membrane surface due to partial permeation into the inter-pore
 524 of the PVDF membrane and formed the hydrophilicity/hydrophobicity of the matrix,
 525 limiting the improvement of the anti-pollution ability. Compared to the EGCG-PVDF
 526 membrane, the hydration layer was formed on the surface of the EGCG-ESBR/PVDF
 527 membrane due to the hydrophilicity of EGCG, which prevented foulants from
 528 becoming trapped or aggregating in the pores and valleys of the membrane surface.
 529 Thus, the EGCG-ESBR/PVDF membrane exhibited good separation performance and
 530 long-term anti-fouling ability due to its excellent surface properties.



531

532 **Fig. 10.** Schematic illustration of the difference in the fouling process between

533 EGCG-PVDF and EGCG-ESBR/PVDF membranes.

534

535

536 3.6 Oil/water emulsion separation performance

537

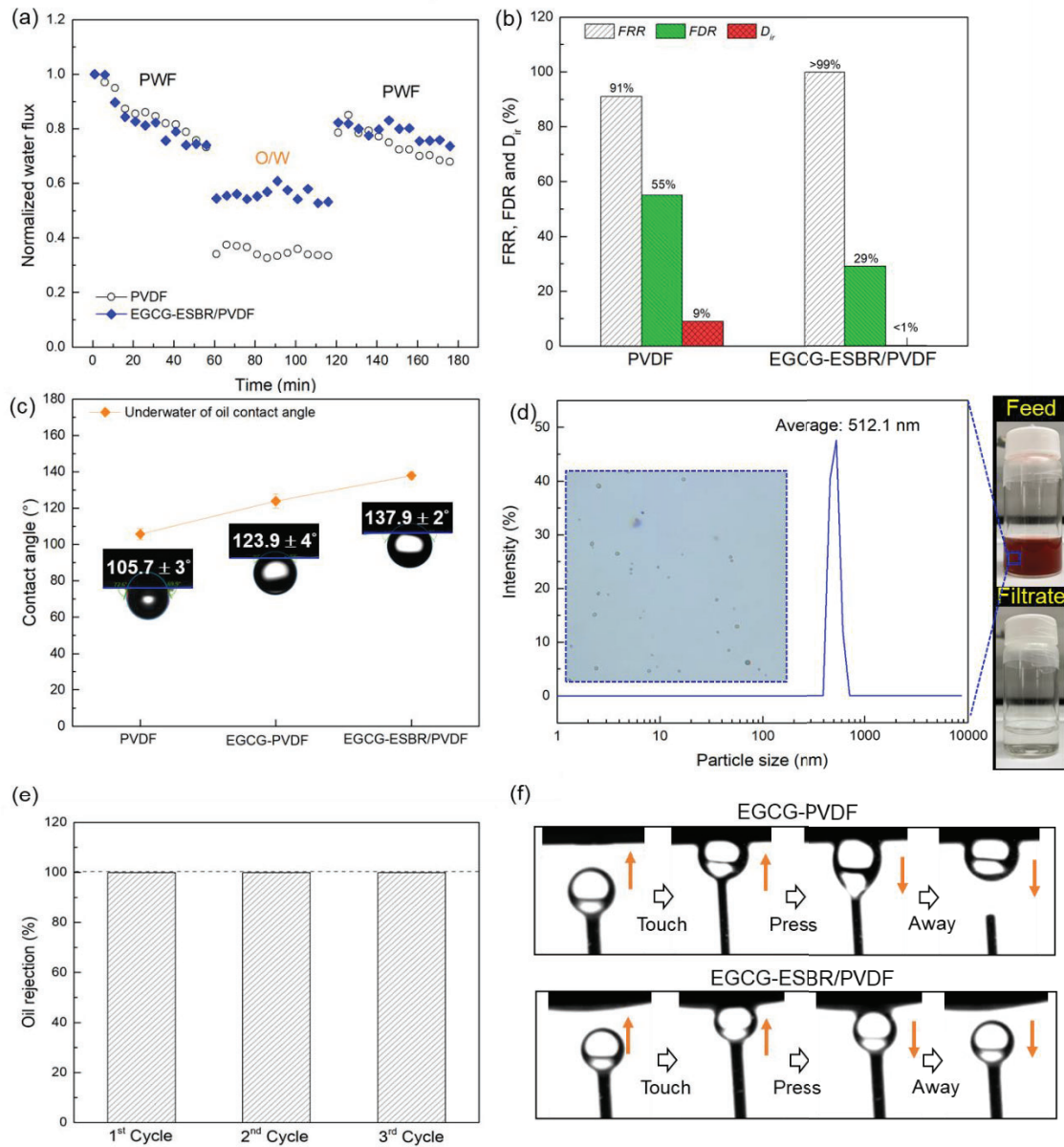
538 In addition, the surface hydrophilicity of the modified PVDF membrane allowed
 539 for oil-in-water emulsion separation. As shown in Fig. 11a, a similar initial pure water
 540 flux decays between the pristine membrane and the modified membrane. However, the

541 permeation flux of the pristine membrane dropped quickly (from 94.5 to 43.1 L m⁻² h⁻
542 ¹) during the 60 min filtration of the oil/water emulsion. The FDR of the pristine
543 membrane reached 55%, and the FRR recovered to 91% after cleaning Fig. 11b). In
544 contrast, the modified membrane showed a lower decline in the permeation flux (from
545 65 to 47 L m⁻² h⁻¹), the Drir was lower than 1%, and the FRR reached 99%. According
546 to the surface wetting properties of the membrane before and after modification (Fig.
547 6a and Fig. 11c), greater improvements in hydrophilicity and oleophobicity were
548 observed. This improvement occurred because of the introduction of the EGCG-ESBR
549 selective layer on the membrane surface, which helped improve the membrane
550 hydrophilicity.

551 As shown in Fig. 11d, the feeds and filtrates are displayed in the optical
552 photographs, while many microscale droplets can be seen in the original emulsions.
553 After filtration, no droplets could be detected in the filtered pollutants. Moreover, as
554 shown in Fig. 11e, the modified membrane exhibited stable and high oil rejection during
555 three-time cycle filtration. The anti-fouling performance was further proven by an
556 underwater-oil-adhesion experiment, as shown in Fig. 11f. Compared to the EGCG-
557 PVDF membrane, the oil droplet can remain in shape when touched and left the EGCG-
558 ESBR/PVDF membrane rather than deform and stay on the membrane surface. It was
559 clear that the highly hydrophilic surface of the modified membrane showed an anti-

560 fouling ability to avoid oil droplets adhering to the membrane (see the Supplementary
561 Video). This result indicated that the EGCG-ESBR/PVDF membrane successfully
562 realized oil-in-water emulsion separation and demonstrated the excellent reusability of
563 the membrane, indicating the excellent surface hydrophilicity and underwater
564 oleophobicity of the EGCG-ESBR deposited on the PVDF membrane.

565 In addition, a comparison of the comprehensive performance for newly developed
566 anti-fouling PVDF membranes reported in the literature and the fabricated membrane
567 in this work is shown in Table 2, showing that the EGCG-ESBR/PVDF membrane
568 presented promising anti-fouling performance. This result indicated that the novel
569 EGCG-ESBR layer is beneficial for enhancing the anti-fouling ability of the membrane,
570 which exhibited good reusability after recycling. In addition, this modification process
571 with the use of reuse and natural materials is not only environmentally friendly but also
572 cost-effective. As discussed above, the novel method of hydrophilic modification is
573 competitive in the separation membrane for organic wastewater treatment.



574

575 **Fig. 11.** (a) Time-dependent normalized fluxes and (b) anti-fouling properties of the

576 pristine membrane and modified membrane in the oil/water emulsion. (c) Underwater

577 oil contact angle (OCA) of the membranes. (d) Optical microscopy images before and

578 after filtration of kerosene in water. (e) The oil rejection of the modified membrane

579 after every cycle during emulsion separation. (f) Real-time recorded underwater-oil-

580 adhesion experiment on EGCG-PVDF and EGCG-ESBR/PVDF membranes.

Table 2 Comparison of antifouling properties of the PVDF membranes for oil-in-water emulsions

Membrane	Oil-in-water emulsions	Environmentally Friendly	Recycle time	Recycle changing rate (R, %)	Rejection properties (%)	Ref.
Ag/EGCG-PVDF	SDS emulsified 0.1% diesel	Middle	5	48	>95.4	[58]
TA-Ti@PVDF	Tween 80 emulsified 1.0% petroleum ether	Middle	10	28	99.3	[59]
TA/DEDAPS PVDF	SDS emulsified 0.1% petroleum ether	Middle	6	3.7	>99.2	[60]
Cellulose-TA-PVA-PVDF	CTAB emulsified 0.2% sunflower	High	30	3.1	99.9	[61]
EGCG-ESBR/PVDF	Tween 60 emulsified 0.1% kerosene	High	3	<1	>99.5	This work

583 **Conclusion**

584 Using a green and facile approach, we designed a green EGCG-ESBR hydration
585 layer to improve the hydrophilic surface of the PVDF membrane. To the best of our
586 knowledge, this is the first work that reports a strategy for fabricating high anti-fouling
587 performance membranes through glue attachment. This approach effectively prevented
588 closing or shrinking of the pores and maintained the surface morphology while
589 improving the membrane performance. The EGCG-ESBR/PVDF demonstrated
590 superior protein separation performance to that of the pristine PVDF membrane. The
591 SBR glue layer coated on the membrane surface not only reduced the surface roughness
592 but also enhanced the precise anchoring of the hydrophilic EGCG coating. Under the
593 optimized conditions, EGCG-ESBR/PVDF exhibited the best antifouling performance
594 during BSA rejection (94.5%) and oil rejection (>99.5%). Moreover, the resulting
595 membrane showed remarkable stability during multiple filtration cycles and long-term
596 test. If desired, even higher water fluxes should be achievable by further optimizing the
597 thickness of the glue layer on the membrane surface while using another highly
598 hydrophilic modifier as the selective layer. In the future, reliability estimates for use
599 with industrial wastewater and longer operating times will confirm the application
600 potential in practical wastewater. In summary, this work provides new insights into the
601 hydrophilic modifications of membrane surfaces for improving separation performance

602 and fouling resistance.

603

604 **Declaration of interests**

605 The authors declare that they have no known competing financial interests or
606 personal relationships that could have appeared to influence the work reported in this
607 paper.

608

609 **Acknowledgment**

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612 005-083-MY3) is gratefully acknowledged.

613

614 **Nomenclature**

ΔP	filtration pressure (Pa)
A	effective membrane area (m ²)
BSA	bovine serum albumin
CA	contact angle
DR _{ir}	irreversible flux decline ratio
EGCG	epigallocatechin gallate
ESBR	epoxidized styrene–butadiene–styrene block copolymer membrane

FDR	flux decline ratio
FRR	flux recovery rate
J_f	permeation flux ($\text{L m}^{-2} \text{h}^{-1}$)
J_w	water flux ($\text{L m}^{-2} \text{h}^{-1}$)
J_w'	water flux after cleaning ($\text{L m}^{-2} \text{h}^{-1}$)
MWCO	molecular weight cutoff
NIPS	nonsolvent-induced phase separation
NS	nonsolvent
PET	polyethylene terephthalate
PS	polystyrene
PSF	polysulfone
PVDF	polyvinylidene fluoride
PWF	pure water flux
R	foulant rejection (%)
R_{ir}	irreversible fouling (m^{-1})
R_m	membrane fouling (m^{-1})
R_r	reversible fouling (m^{-1})
R_t	total fouling (m^{-1})
SA	sulfonic acid group
SBR	styrene–butadiene–styrene block copolymer
t	filtration time (h)
V	volume of permeate (L)
WCA	water contact angle
OCA	underwater oil contact angle
x	volume fraction

δ_d	solubility parameter of dispersion force
δ_h	solubility parameter of hydrogen bonding force
δ_p	solubility parameter of polar force
$\Delta\delta$	difference in solubility parameter
μ	permeate viscosity (Pa s)

615

616 **References**

617

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