








REVIEW

S1P/S1PR signaling pathway advancements in autoimmune diseases

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Sphingosine-1-phosphate (S1P) is a versatile sphingolipid that is generated through the phosphorylation of sphingosine by sphingosine kinase (SPHK). S1P exerts its functional effects by binding to the G protein-coupled S1P receptor (S1PR). This lipid mediator plays a pivotal role in various cellular activities. The S1P/S1PR signaling pathway is implicated in the pathogenesis of immune-mediated diseases, significantly contributing to the functioning of the immune system. It plays a crucial role in diverse physiological and pathophysiological processes, including cell survival, proliferation, migration, immune cell recruitment, synthesis of inflammatory mediators, and the formation of lymphatic and blood vessels. However, the full extent of the involvement of this signaling pathway in the development of autoimmune diseases remains to be fully elucidated. Therefore, this study aims to comprehensively review recent research on the S1P/S1PR axis in diseases related to autoimmunity.

Keywords: Sphingosine-1-phosphate (S1P), sphingosine-1-phosphate receptor (S1PR), S1P/S1PR signaling pathway, autoimmune diseases, research progress.

Introduction

Autoimmune disorders result in damage to organs or the entire body system due to a dysregulation of the immune system and subsequent destruction of cells and tissues [1, 2]. Although autoimmune diseases were rare in the past, recent epidemiological studies indicate a prevalence of 3%–5% in the population, with increasing incidence rates posing significant risks to patients' lives and overall well-being [3]. While the exact causes and mechanisms behind these diseases remain unclear, extensive research has shed light on the crucial role of the sphingosine-1-phosphate (S1P)/S1P receptor (S1PR) signaling pathway in the inflammatory response associated with autoimmune disorders [4]. Consequently, the therapeutic potential of targeting S1PRs for the treatment of autoimmune and inflammatory conditions has garnered considerable attention from the scientific community [5]. This article provides an overview of the current research advancements pertaining to the involvement of the S1P/S1PR axis in autoimmune diseases.

Physiopathological role of S1P and S1PRs

S1P, a member of the sphingolipid family, is a biologically active lipid mediator that undergoes conversion from ceramide to sphingosine by ceramidase catalysis [6]. Subsequently, sphingosine is phosphorylated to S1P by sphingosine kinase 1 (SPHK1) and sphingosine kinase 2 (SPHK2) [7]. SPHKs primarily exist in two distinct isoforms, namely, SPHK1 and SPHK2, which are associated with the synthesis of S1P in extracellular and

intracellular compartments, respectively [8]. S1P functions as a pleiotropic lipid mediator that regulates the activities of numerous cell types. However, in mammalian systems, S1P is found predominantly in blood and lymphatic fluid under normal physiological conditions. Erythrocytes, platelets, and endothelial cells are the primary sources of circulating S1P [9], although mast cells and platelets can also produce excessive amounts of S1P in an inflammatory or pre-thrombotic state [10]. Erythrocytes lack S1P-degrading enzymes but express the specific S1P transporter Mfsd2b [11]. Deficiency of Mfsd2b leads to a 50% reduction in plasma S1P concentration. Conversely, endothelial cells express another specific S1P transporter, Spns2, which contributes to the maintenance of circulating S1P concentrations. Deficiency of Spns2 results in a 40% decrease in S1P concentration in plasma and 80% decrease, indicating that lymphatic endothelial cells predominantly maintain S1P in lymph [12, 13]. Platelets also lack S1P-degrading enzymes but express the S1P transporter Mfsd2b, although their contribution to circulating S1P concentrations is minimal, as evidenced by the absence of changes in plasma S1P concentration in platelet-deficient mice [14]. In tissues, S1P is rapidly degraded by S1P lytic enzymes, resulting in a marked contrast between high intravascular S1P concentrations and low tissue S1P concentrations [15]. The S1P₁ receptor plays a crucial role in regulating the trafficking of T and B cells from secondary lymphoid organs into the lymphatic system and bloodstream by facilitating receptor-mediated chemotaxis toward the highest S1P concentration gradient in

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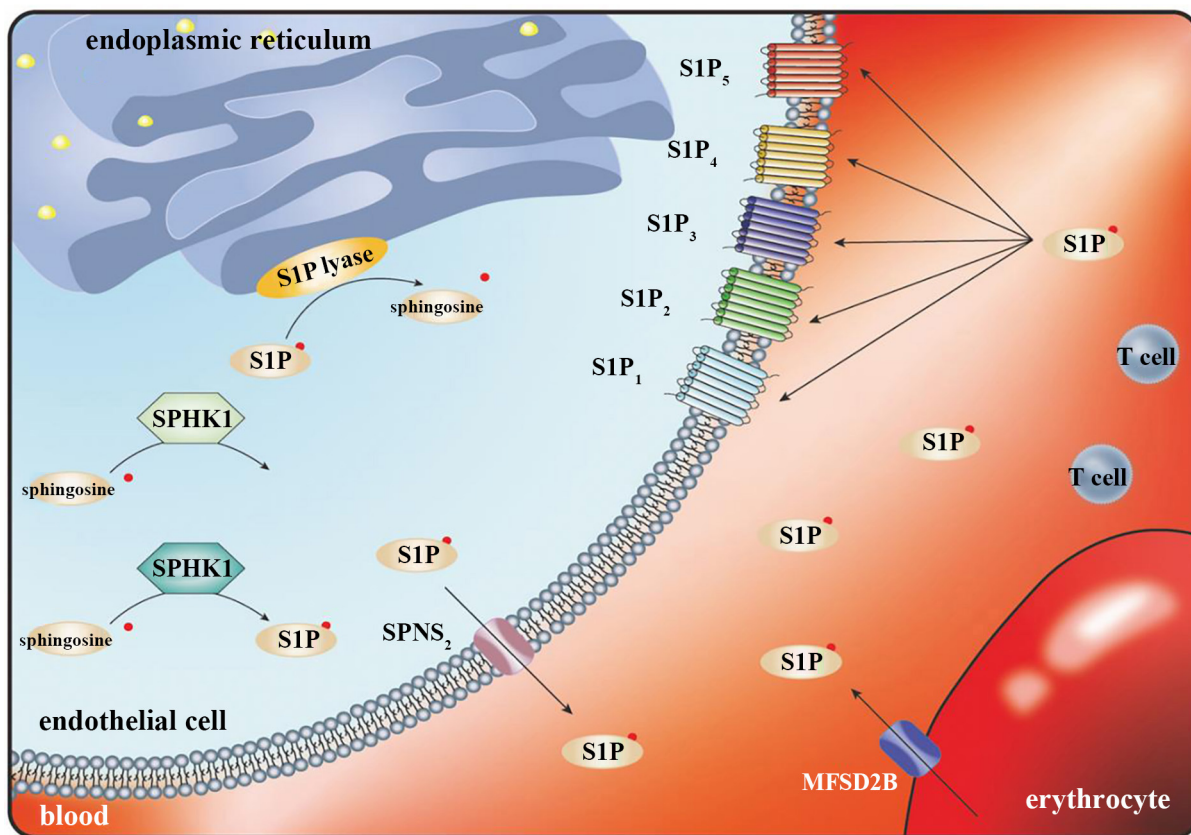


Figure 1. Schematic summary of the current view of S1P signaling pathways. S1P: Sphingosine-1-phosphate; SPHK1: Sphingosine kinase 1.

blood/lymph. This S1P gradient is essential for maintaining physiological tissue homeostasis, by directing cells from the low S1P environment of tissues to the high S1P environment of circulating fluid [16]. Disruptions in the S1P gradient can alter the migratory behavior of immune cells and other cell types. Changes in the local S1P gradient, which can be triggered by local injury and subsequent inflammatory responses, modulate the activity and migratory behavior of immune cells and vascular wall cells. From a pathophysiological perspective, local alterations in S1P gradients support immune system responses and enhance local defense and healing processes. There is speculation that the S1P gradient might play an important role in lymphocyte egress [17]. Initially, S1P was defined as an intercellular second messenger [18, 19] that interacts with intracellular targets, such as inhibin 2 and tumor necrosis factor receptor-associated factor 2. The physiological significance of intracellular S1P signaling was unknown at the time. However, subsequent research has revealed the existence of five high-affinity receptors, known as S1PRs, which are a class of G protein-coupled receptors located on cell membranes. Five S1PR subtypes have been identified, namely, S1P₁, S1P₂, S1P₃, S1P₄, and S1P₅ (Figure 1). S1PRs are characterized by tissue-specific expression and partially synergistic and antagonistic functions. Nearly all cell types in the body appear to express S1PRs. The typical receptor-mediated effects of S1P encompass cell proliferation, migration, and apoptosis [20]. Notably, S1PRs are highly expressed in immune

cells, including neutrophils, dendritic cells, natural killer cells, and macrophages, and may regulate inflammatory diseases and wound healing processes, as well as various cellular trafficking events involving hematopoietic cells and lymphocytes. S1P serves a critical role in immune cell responses, such as B cell migration and their interaction with T cells. Furthermore, it influences the localization of immune cells, such as natural killer cells, within lymph nodes, and modulates their response to interferon-gamma [21]. Consequently, the S1P/S1PR axis is implicated in immune-mediated diseases and has been proposed as a potential target for the prevention or treatment of certain conditions.

S1P₁ is expressed in various tissues and organs, including the brain, lungs, liver, heart, and spleen. It serves as a crucial receptor on the surface of lymphocytes, mediating their homing [22]. Under normal circumstances, S1P₁ expression is upregulated in lymphoid organs due to the influence of lymphocytes, and it is also expressed in oligodendrocytes and neurons in the central nervous system (CNS) [23]. In mesenchymal stem cells, its expression regulates cell migration, proliferation, differentiation, and survival [24]. S1P₁ is involved in vascular system development and is necessary for maintaining endothelial cell barrier integrity and vascular permeability [25]. Consequently, its expression is high in differentiated endothelial cells [26] and is further upregulated in inflammatory environments. S1P₁ exerts anti-inflammatory effects by inhibiting the expression of pro-inflammatory adhesion molecules 2–4 on leukocytes and

limiting cytokine production [4]. Additionally, it plays a role in astrocyte proliferation, dendritic cell migration, heart rate regulation, and ischemia–reperfusion injury [27]. Regulatory T cells suppress the activation and proliferation of CD4+ helper T cells, inhibit the differentiation of cytotoxic CD8+ T cells, and restrict B cell activation. However, S1P₁ inhibits the number and function of regulatory T cells, leading to aggravation of autoimmune diseases [4].

S1P₂ is located in both the plasma membrane and cytoplasm. It is expressed in both the CNS and the immune system and plays a pivotal role in the inhibition of apoptosis, cell proliferation, actin remodeling, and B cell localization in the follicle. Moreover, it is also implicated in the development of the cardiac, auditory, and vestibular systems. Recent studies have revealed that S1P₂ negatively impacts myelin repair by modulating blood–brain barrier permeability, while also exerting a significant influence on demyelination [28]. Interestingly, S1P₂, in striking contrast to S1P₁, promotes the inflammatory response [29].

S1P₃ is located on the plasma membrane and is expressed in certain organs (heart, lungs, kidneys, and spleen), peripheral cells, and fibroblasts, with lower expression in epithelial cells. However, its expression increases in cases of lung injury [30]. S1P₃ plays a crucial role in cell proliferation, differentiation, apoptosis, and migration, although its function remains contentious [31]. Research suggests that S1P₃ plays a vital role in hematopoietic stem cells and leukemia stem cells, exhibiting a protective effect against bacterial sepsis [32, 33]. Furthermore, S1P₃ enhances the Notch signaling pathway, which is associated with retinal astrocyte proliferation. However, its role in the immune system is still unclear. Some studies indicate that S1P₃ participates in regulating the biological functions of certain immune cells, such as dendritic cells, macrophages, and natural killer cells. Currently, most research on S1P₃ is focused on its vascular-related aspects.

S1P₄ receptor is expressed primarily in hematopoietic, lymphoid, and pulmonary tissues. It exerts inhibitory effects on tumor proliferation, controls the movement of immune cells, and facilitates the stimulation and maturation of dendritic cells [34, 35]. Moreover, S1P₄ expressed on dendritic cells plays a pivotal role in enabling regulatory T cells to dampen the activity of cytotoxic T lymphocytes through its involvement in the regulation of helper T cell and interleukin 27 (IL-27) synthesis [36].

S1P₅ receptor is present in the epidermis, oligodendrocytes, cytotoxic lymphocytes, as well as myelinated cells residing in the cerebral and splenic regions [37]. S1P₅ exerts a neuroprotective influence through the mediation of oligodendrocyte evasion of demyelination and apoptotic processes [38]. Additionally, it is speculated that S1P₅ may have therapeutic potential in the management of multiple sclerosis (MS) [39].

Role of S1P and S1PRs in multiple sclerosis

MS is an autoimmune disorder characterized by the infiltration of peripheral B and T lymphocytes into the CNS, where they attack the protective myelin sheaths surrounding neuronal

axons. This results in impaired nerve conduction and the development of a disabling neurological condition, typically affecting young adults and showing an increasing incidence globally [40, 41]. Current phenotypic classifications of MS include relapsing–remitting MS, clinically isolated syndrome, radiologically isolated syndrome, primary progressive MS, and secondary progressive MS [42]. The exact pathogenesis of MS remains unclear, but it is believed that self-reactive T cells can breach the blood–brain barrier and initiate inflammatory responses against myelin antigens, leading to demyelination and neurodegeneration. Moreover, research has indicated that the activation of microglia and astrocytes within the CNS releases pro-inflammatory molecules that contribute to neuritis and secondary brain damage in MS and other CNS-related diseases [43, 44]. Of particular interest in MS is the role of S1PRs and their expression within the CNS. S1P, an extracellular signaling molecule, mediates various biological responses, including lymphocyte transport, inflammation, and vascular development, through interactions with S1PRs [45]. Interestingly, S1P is abundantly present in the CNS, and it is believed that S1P signaling modulated by S1PRs plays a crucial role in neuroinflammatory processes, which are major contributors to neurodegeneration [46, 47]. Importantly, many key neuroglial cell types (microglia, astrocytes, oligodendrocytes, and oligodendrocyte precursor cells) express S1PRs [48]. This suggests that modulating S1PRs may have therapeutic potential in the treatment of various neuroinflammatory diseases, as it can influence the activation, proliferation, and migration of glial cells. Currently, S1PR modulators are utilized to enhance systemic immune responses, with therapeutic strategies targeting S1PR initially developed for relapsing–remitting MS. Four S1P modulators (Fingolimod, Siponimod, Ozanimod, and Ponesimod) are currently approved for the treatment of MS (Table 1 and Figure 2) [49]. Notably, S1PRs S1P_{1/3/5} are expressed in neurons, astrocytes, microglia, oligodendrocytes, and oligodendrocyte precursor cells, while S1P₂ is present in neurons, astrocytes, and microglia but not on oligodendrocyte precursor cells [50]. S1PRs are also involved in regulating lymphocyte transport, primarily through the binding of S1P₁ on lymphocytes in MS. When S1P₁ function is blocked, lymphocytes become sequestered in lymph nodes and the thymus, unable to respond to the S1P gradient that facilitates their exit from the lymph nodes. This reduction in circulating lymphocytes can limit the migration of inflammatory cells to the CNS [51]. Furthermore, blocking S1P₁ inhibits the amplification of inflammatory cytokines and the recruitment of immune cells in animal models, emphasizing the goal of MS treatment to hinder immune cell infiltration into the CNS.

Ozanimod (ROCK1063) is an S1P_{1/5} agonist that exhibits 27 times greater selectivity for S1P₁ compared with S1P₅. Its interaction with S1PRs leads to receptor internalization and subsequent degradation via ubiquitin protease-dependent mechanisms, preventing the reassembly of receptors on the cell membrane. This mechanism results in the inhibition of lymphocyte egress from lymph nodes, leading to a decrease in absolute lymphocyte counts. However, this effect is rapidly reversible,

Table 1. S1PR modulators in the treatment of multiple sclerosis

Drug	Receptor selectivity	Disease	Elimination T ^{1/2}	Lymphocyte recovery after drug discontinuation
Fingolimod (FTY720)	S1PR1, 3, 4, 5	Relapsing–remitting multiple sclerosis	7 days	6 weeks
Siponimod (BAF312)	S1PR1, 5	Secondary progressive multiple sclerosis	30 h	1–10 days
Ozanimod (ROC1063)	S1PR1, 5	Relapsing–remitting multiple sclerosis, ulcerative colitis and Crohn's disease	19–20 h	48–72 h

Table lists key protein targets and ligands in this article which are hyperlinked to corresponding entries in <http://www.guidetopharmacology.org>, the common portal for data from the IUPHAR/BPS Guide to PHARMACOLOGY (Southan et al., 2016) and are permanently archived in the Concise Guide to PHARMACOLOGY 2015/16. S1PR: Sphingosine-1-phosphate receptor.

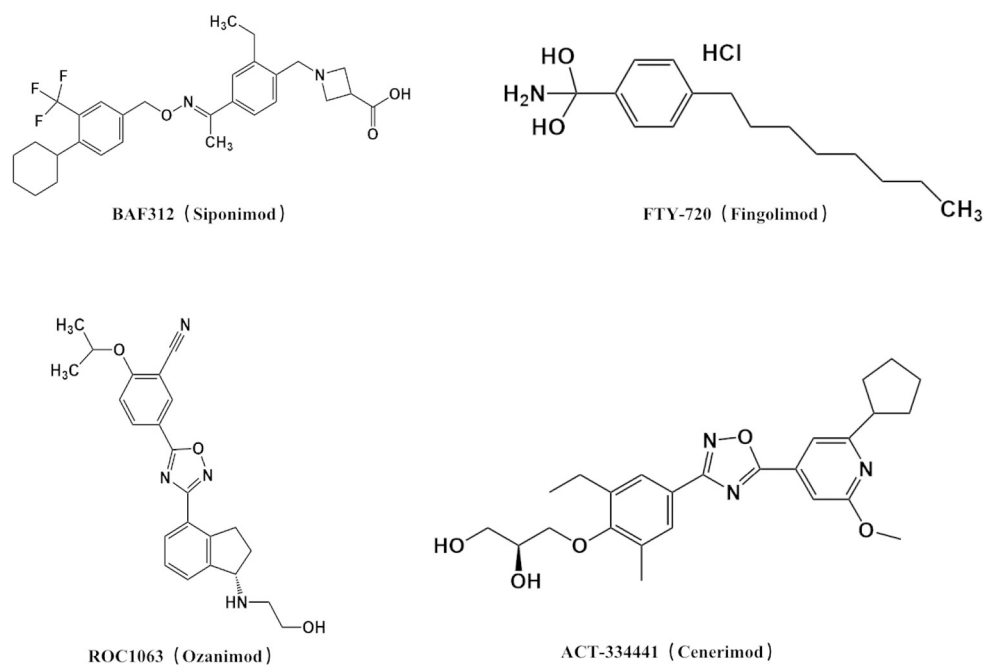


Figure 2. Chemical structure of S1PR modulators. S1PR: Sphingosine-1-phosphate receptor.

with lymphocyte counts returning to baseline within 48–72 h upon discontinuation of the drug [52]. Ozanimod was approved by the FDA in 2020 for the treatment of relapsing MS in adults. It has undergone phase II and two phase III pivotal trials, demonstrating both efficacy and favorable tolerability [53]. In phase II trials, compared to placebo, the use of Ozanimod at doses of 0.5 or 1 mg/day resulted in fewer new or expanded T2 lesions and gadolinium-enhancing lesions on brain magnetic resonance imaging scans, with only three non-treatment-related serious adverse events reported in patients receiving Ozanimod 0.5 mg (optic neuritis, somatoform autonomic dysfunction, and cervical squamous metaplasia related to human papillomavirus). No serious infections or cardiac adverse events were reported, and no cases of macular edema were observed. The most common adverse events in the Ozanimod 1.11 mg and 12 mg groups were nasopharyngitis, headache, and urinary tract infection, relative to the placebo group [54]. Overall, Ozanimod displayed a favorable safety profile, with

no significant events related to cardiovascular, pulmonary, ophthalmic, infectious, or malignant conditions. However, respiratory function should be assessed during treatment [55]. In animal models of relapsing–remitting MS, experimental autoimmune encephalomyelitis (EAE) is the most frequently used model, inducing demyelination in the nervous system. In the EAE mouse model, blocking S1P₁ enhances neuronal survival and inhibits glial cell proliferation and demyelination. Studies have elucidated the anti-inflammatory and neuroprotective effects of Ozanimod, in which it primarily reduces glutamatergic synaptic transmission during EAE. This effect is achieved by attenuating the release of inflammatory cytokines by activated microglia and T cells [56].

Fingolimod (FTY-720), a potent anti-inflammatory agent, gained FDA approval in 2010 as the first oral medication for relapsing–remitting MS. It effectively reduces the annual relapse rate by inhibiting lymphocyte drainage from lymphoid tissues and minimizing neuroinflammation in the

CNS [57]. Phase II trials demonstrated that FTY-720's peak effectiveness against MS occurred at a suboptimal dose that induced lymphocytopenia. Interestingly, S1PRs were found to be expressed in various types of neuronal cells, leading researchers to suggest that FTY-720 might also exert direct neuroprotective effects in the brain [58]. However, phase III trials revealed serious adverse events in some patients, such as seizures, infections, and leukopenia [59]. In addition to its anti-inflammatory properties, FTY-720 reduces astrocyte proliferation and promotes oligodendrocyte differentiation within the CNS, potentially preventing brain atrophy [60]. In mice with EAE, FTY-720 decreased astrocyte proliferation and pro-inflammatory cytokine expression, thereby reducing demyelination, axonal loss, and astrocyte proliferation [61, 62]. Furthermore, the level of inhibition of astrocyte proliferation by FTY-720 correlated with its strength in inhibiting S1P₁ function. This suggests that S1P₁ on astrocytes plays a role in MS-related astrocyte hyperplasia and that FTY-720 requires S1P₁ on astrocytes to exhibit its activity in an EAE mouse model [63]. Moreover, FTY-720 inhibits the release of pro-inflammatory chemokines, cytokines, and neurotoxic factors, such as C-X-C motif chemokine 5, induced by lipopolysaccharide stimulation in astrocytes and microglia [64]. In an EAE mouse model, FTY-720 derivatives ST-1893 and ST-1894 caused severe lymphopenia and alleviated symptoms [65]. Another study focused on novel butterfly derivatives of FTY-720, namely, ST-1505 and ST-1478, both agonists of S1P_{1/3} and S1P_{1/5} receptors, respectively. These derivatives show promise as new drugs for relapsing–remitting MS and other immune diseases. Additionally, they possess histamine H₃ receptor antagonistic properties and may improve cognitive impairment in relapsing–remitting MS. This suggests that S1P₁-selective compounds could potentially be more effective than prodrugs in treating relapsing–remitting MS [66].

Siponimod (BAF312) exhibits strong affinity for S1P₁ and S1P₅ receptors expressed by endothelial cells that constitute the blood–brain barrier and prevent lymphocytes from entering the CNS. By concurrently modulating S1P_{1/5}, Siponimod enhances the properties of the blood–brain barrier, thereby offering additional effectiveness in treating secondary progressive MS [67]. Furthermore, Siponimod induces a balanced state of anti-inflammation and suppression in the immune system by enriching regulatory T cells, transitional regulatory B cells, and B1 subpopulations. These changes potentially contribute to the clinical efficacy of Siponimod in the treatment of secondary progressive MS [68]. In a phase III clinical study spanning over five years, continuous administration of the S1P modulator Siponimod resulted in a significant reduction in disability progression, rate of cognitive deterioration, and maintenance of low annual relapse rates, as well as reductions in total and localized brain atrophy and inflammatory activity in patients with secondary progressive MS. These improvements were particularly prominent in patients who started Siponimod treatment early and maintained its use [69]. Another phase III clinical trial evaluating Siponimod in secondary progressive MS showed that adverse events occurred in 89% of patients (975 of 1099) who received Siponimod treatment. These events included

lymphopenia, elevated hepatic transaminase concentrations, bradycardia and bradyarrhythmia at the start of treatment, macular edema, hypertension, varicella-zoster reactivation, and convulsions, which occurred more frequently in the Siponimod group [70]. Siponimod also exerts an effect on astrocytic hyperplasia and microgliosis. In mice with EAE, administration of Siponimod attenuated the proliferation of astrocytes and microglia, and reduced lymphocyte infiltration, thereby dampening the inflammatory response [71]. It was also observed that Siponimod decreased macrophage infiltration and microglia activation in the spinal cord of EAE mice [38], and in mice with traumatic brain injury, it decreased the activation of CD4+ and CD8+ T cells, as well as the extent of injury [72]. The importance of S1P in the CNS is progressively being recognized, and the multifaceted effects of FTY-720 and other drugs that target SPHK suggest that this approach should be considered for the treatment of other neurodegenerative diseases.

Role of S1P and S1PRs in systemic lupus erythematosus

Systemic lupus erythematosus (SLE) is a chronic autoimmune disease characterized by various clinical phenotypes. While the exact cause remains unknown, it is believed that a combination of genetic and environmental factors contributes to its development [73]. SLE is characterized by immune system dysfunction and the persistent presence of an inflammatory environment, often resulting in damage to multiple organs and, ultimately, the death of the patient. However, the pathogenesis of SLE remains unclear, and available treatment options are limited. The S1PR, a recognized regulator of lymphocyte transport with potent immunomodulatory functions, is closely associated with SLE and other autoimmune diseases. Elevated levels of S1P have been observed in the serum of SLE patients, which leads to an increase in the number of lymphocytes in the peripheral blood and the infiltration of inflammatory cells into organs [74]. In mouse models of SLE, increased S1P levels have been shown to exacerbate disease activity and organ damage. In patients with SLE, the expression of various pro-inflammatory cytokines is increased, leading to an increase in S1P production. Conversely, elevated levels of S1P further aggravate the inflammatory state by activating inflammatory signaling, recruiting immune cells, and promoting the release of pro-inflammatory mediators [75].

Cenerimod (ACT-334441) is an orally administered modulator of the S1P₁ receptor. Unlike many S1PR modulators, which often induce bronchoconstriction or vasoconstriction, Cenerimod does not cause these side effects in the non-clinical setting. In an animal study involving SLE and Sjögren's syndrome mice, Cenerimod was found to improve both systemic and organ-specific autoimmunity, resulting in reduced inflammation and preserved organ function [76]. A phase II clinical trial investigated the effects of Cenerimod on SLE activity and circulating lymphocytes in patients with mild to moderate SLE. The trial focused on patients with a SLEDAI-2K score of at least two for musculoskeletal or mucosal manifestations, a history of antinuclear or anti-double-stranded DNA antibodies,

or a positive serologic test at screening. The findings from this trial suggest that Cenerimod has the potential to treat SLE in patients, while also ensuring safety [77]. Studies have demonstrated that Cenerimod significantly improves both systemic and organ-specific pathology and inflammation in SLE mice. For example, it reduces levels of plasma anti-double-stranded DNA and immunoglobulin, decreases blood B and T lymphocyte counts, and diminishes circulating and organ-infiltrating lymphocytes. Additionally, Cenerimod decreases proteinuria and improves survival [78]. Similarly, research has shown that Ozanimod, a small molecule modulator of S1P₁ and S1P₅ receptors, attenuates chronic inflammation and renal pathology in SLE mice.

Role of S1P and S1PRs in lupus nephritis

Lupus nephritis is the most prevalent complication of SLE, and its progression indicates a poor prognosis and a high mortality rate [79]. S1P regulates the production of inflammatory mediators, and elevated S1P levels have also been associated with the development of SLE [80]. One study demonstrated that S1P expression was significantly increased in the serum of SLE patients with lupus nephritis activity [81]. Similarly, mice with lupus nephritis had elevated S1P expression in their serum [82]. These findings suggest that S1P and its receptors play a role in the development of lupus nephritis. Moreover, S1P₃ has been observed in the renal cortex, outer medulla, and inner medulla [83]. Activation of the S1P/S1P₃ signaling pathway suppresses the number and function of regulatory T cells. Transplantation of regulatory T cells before clinical disease onset has been shown to reduce kidney injury progression and mortality in mice predisposed to lupus [84]. FTY720 sequesters circulating lymphocytes within lymph nodes, thereby decreasing the infiltration of autoimmune cells (T and B lymphocytes) and inflammatory factors such as IL-6 and TNF- α from the bloodstream and inflammatory tissues into target organs. This leads to the relief of SLE symptoms. Administration of FTY720 to lupus mice has been found to reduce renal injury and alleviate end-stage glomerular inflammation [85]. Additionally, FTY720-treated lupus mice exhibit decreased production of anti-double-stranded DNA in the glomerulus and reduced accumulation of IgG, which consequently inhibits the progression of lupus nephritis [86].

Role of S1P and S1PRs in neuropsychiatric lupus

Neuropsychiatric SLE is a severe complication of SLE, in which patients commonly exhibit nonspecific symptoms like headache and cognitive impairment. However, they may also develop more destructive symptoms, such as memory loss, seizures, and stroke, with a mortality rate second only to that of lupus nephritis [87]. The central and peripheral nervous systems of SLE patients can be targeted, leading to neurological or psychiatric symptoms that contribute to cognitive impairment and depression. Although much remains unknown concerning the pathogenesis of neuropsychiatric SLE [88], one study discovered that the disruption of the intact blood-brain barrier is a primary factor in disease progression [89]. Cenerimod, a selective S1PR modulator, has been found to directly impact brain

endothelial cells, reducing blood-brain barrier permeability and achieving symptom relief [90]. Moreover, Cenerimod also plays a neuroprotective role. In lupus mice, administration of the S1P inhibitor FTY-720 mitigated neurobehavioral deficits, including spatial behavior and depressive behavior, while also reducing cortical, hippocampal, and amygdalar neuronal damage. Additionally, it suppressed inflammatory expression in astrocytes and endothelial cells, although cytokine levels in the cerebral cortex and hippocampus were not significantly decreased. Notably, IgG deposition in the mouse brain was not affected [91]. Similarly, FTY-720 has been shown to significantly attenuate impulsive and depressive-like behavior, reduce levels of inflammatory cytokines, and decrease the infiltration of T cells and neutrophils into the brain parenchyma in lupus mice. Furthermore, FTY720 acts on endothelial cells in the brains of lupus mice by inhibiting the expression of multiple signaling pathways and reducing the permeability of the blood-brain barrier, resulting in decreased central levels of IgG and albumin [90].

Role of S1P and S1PRs in rheumatoid arthritis

Rheumatoid arthritis (RA) is the most prevalent systemic autoimmune rheumatic disease, affecting approximately 1% of the population with a male-to-female ratio of 1:2.5. The disease can manifest at any age but is more commonly observed in middle and old age, with its occurrence increasing with age [92]. The primary pathological changes in RA include synovial inflammation and proliferation, as well as the production of autoantibodies (rheumatoid factor and anti-citrullinated protein antibodies). These autoantibodies contribute to the damage and disability of cartilage and bone [93]. The synovial microenvironment primarily involves fibroblast-like synoviocytes (FLS) and vascular endothelial cells. FLS promotes the release of inflammatory factors and induces vascular endothelial growth factor (VEGF) production, thereby stimulating endothelial progenitor cells and promoting angiogenesis and RA development [94]. The S1P/S1PR axis plays a significant role in RA, as S1PRs were found to be elevated in RA model mice. RAFLS produced IL-6 in response to S1P stimulation, and S1P₃ expression levels increased with pre-activation by tumor necrosis factor alpha (TNF- α). An increase was also observed in IL-6 and matrix metalloproteinase 3 (MMP3) production induced by S1P₃ [95]. Inhibition of the S1P/S1PR (S1P₁/S1P₂/S1P₃/S1P₄) signaling pathway can suppress the proliferation, migration, pro-inflammatory, and apoptosis-promoting activities of RAFLS. Additionally, IL-17 expression can be reduced [96, 97]. Recent studies have demonstrated the expression of S1P and S1P₁ receptors in the synovium in RA. Synovitis is the primary cause of elevated cytokine levels (TNF- α , IL-1, IL-6, IL-1 β), and these inflammatory factors induce SPHK1 in a manner dependent on extracellular signal-regulated kinase signaling [98, 99]. Consequently, high levels of S1P are produced, which partly explains the presence of inflammatory tissue [100]. Furthermore, S1P signaling promotes synovial cell proliferation and induces the production of prostaglandin E2 by upregulating the expression of cyclooxygenase-2 in response to inflammatory

cytokines [101]. Inflammatory cytokines also induce the elevation of MMPs, thereby activating osteoclasts and creating a vicious cycle. Other studies have demonstrated that S1P, through its receptor S1P₁, enhances the expression of receptor activator of NF- κ B ligand (RANKL), leading to the production of inflammatory cytokines and bone erosion [102].

In animal experiments, FTY720 was observed to mitigate the expression of IL-6 and TNF- α in the synovial membrane of mice with RA [103]. Based on animal experiments, gardenia glycosides were discovered to deplete S1P or impede the activation of SPHK1, blocking the interaction between FLS and vascular endothelial cells [104]. Furthermore, recent animal experiments have revealed that gardenia glycosides can inhibit the activation of the S1P/S1PR1 signaling pathway through SPHK1, leading to reduced expression of VEGF, inhibition of neoangiogenesis, diminished production of synovial tissue, alleviation of bone erosion, and effective alleviation of RA symptoms [105]. Additionally, gardenia glycosides can also suppress the SPHK1/S1P pathway to alleviate vascular endothelial cells stimulation caused by FLS, inhibit angiogenesis, and attain therapeutic effects [106]. Recently, S001PR930 and the S1PR1 modulator IMMHI (SYL4) inhibited the progression of arthritis in Sprague-Dawley rats, as evidenced by reduced hind paw swelling and reduced arthritis index, alongside decreased levels of pro-inflammatory cytokines and chemokines in affected joints [107]. Despite these promising outcomes observed in animal studies, no ongoing clinical studies have been conducted on individuals with RA. Therefore, for future RA treatments, in addition to conventional approaches, such as non-steroidal anti-inflammatory drugs, corticosteroids, disease-modifying antirheumatic drugs, and biologics, down-regulation of S1P may be considered as a means to achieve effective RA control.

Role of S1P and S1PRs in systemic sclerosis

Systemic sclerosis (SSc), a rare and intricate systemic autoimmune rheumatic disease, is characterized by immune irregularities, vascular lesions, and fibrosis. Primarily affecting the skin, gastrointestinal tract, lungs, kidneys, and heart [108], SSc presents a mortality risk of up to 20% through its complication of interstitial lung involvement. Following an interstitial lung diagnosis, primary or secondary pulmonary hypertension occurs in 8%–12% of patients, leading to a 25% mortality rate within 3 years [109]. Consequently, urgent exploration into novel therapies is imperative to mitigate the morbidity and mortality associated with SSc. Studies have revealed aberrant activation and differentiation of B cells in SSc patients [110]. Moreover, T cell-derived cytokines exhibit anomalies in SSc patients [111], highlighting the significance of lymphocyte regulation in SSc treatment. The S1P/S1PRs signaling pathway has been implicated in the pathogenesis of SSc. Animal models of fibrosis (lung, liver, and kidney) have demonstrated S1P involvement in fibrotic development, and elevated serum S1P levels have been observed in SSc patients due to increased activation of platelet aggregation that occurs during SSc lesions. This increased S1P binding to its receptors, S1P₂ and S1P₃,

induces vasoconstriction, promotes vascular smooth muscle cell proliferation, and stimulates neointima formation, ultimately causing vasospasm and Raynaud's phenomenon [112]. Skin tissues of SSc patients express S1P_{1/2/3/5}, with reduced expression levels of S1P₁ and S1P₂ receptors and increased levels of S1P₃ receptors due to altered distribution of S1P isoforms in fibroblasts. The impact of S1P₅ on inflammatory and pro-fibrotic processes remains unknown [113, 114]. Furthermore, S1PRs play a role in facilitating the migration of lymphocytes from the thymus or secondary tissues into the bloodstream [115], and obstructing this pathway can impede the expression of inflammatory cytokines in SSc patients, leading to a deceleration of inflammation and fibrosis.

FTY720 has been shown to potentially reduce the upregulation of type I interferon in SSc by plasmacytoid dendritic cells [116]. FTY720 has also demonstrated the ability to decrease T helper 17 cell differentiation and functionality, which in turn promotes fibrosis by increasing the abundance of T helper 17 cells and plasmacytoid dendritic cells in a bleomycin-induced scleroderma mouse model [117]. The increased presence of these two cell types has also been observed in SSc patients [118, 119]. Cenerimod, an S1P₁ receptor modulator, reduces blood lymphocytes, and early administration of Cenerimod has been proven effective in reducing skin and lung fibrosis in bleomycin-induced SSc mice. This reduction is accompanied by a decrease in IL-6 and collagen deposition [120]. FTY720 is believed to exert its immunomodulatory effects by reducing blood lymphocytes. However, long-term administration of FTY720 exacerbates bleomycin-induced SSc by causing vascular leakage and intra-alveolar coagulation, ultimately leading to fibrosis and death in SSc mice [121]. Therefore, selective S1PR receptor modulators are more suitable for treating SSc. Studies have shown that Cenerimod administration inhibits the infiltration of CD4+ T cells, CD8+ T cells, and CD11+ B cells into inflamed skin, increases the number of regulatory T cells in the spleen and skin, and reduces the expression of extracellular matrix and fibroblast cytokines in the skin in a study on bleomycin-induced fibrosis models. It also attenuates skin and lung fibrosis [120]. Therefore, Cenerimod holds promise as a treatment option for SSc patients and could potentially slow down the progression of the disease. Multiple S1PRs may play a role in the pathophysiological changes of SSc, affecting various immune, vascular, and cellular aspects. Only through a deeper understanding of these mechanisms can we develop more effective and rational therapeutic approaches.

Role of S1P and S1PRs in bone immunology

The connection and interaction between the skeletal system and the immune system are referred to as osteoimmunology. It has become an indispensable theoretical foundation for understanding the pathophysiological mechanisms of certain diseases, such as RA, ankylosing spondylitis, and osteoarthritis. S1P and S1PRs play a role in both the immune system and bone remodeling. The role of S1P/S1PR1 signaling in bone

remodeling can directly target osteoclastogenesis and osteogenesis, due to the significant role of S1P/S1PR1 signaling in inflammation [122], which is considered a catalyst for inflammation-induced osteoclastogenesis. Moreover, the functionality and polarization of inflammatory cells in the adaptive immune system (T cell subsets) and innate immune cells (macrophages) are also regulated by this signaling axis. This suggests that the S1P/S1PR1 signaling axis can indirectly regulate bone remodeling through the modulation of the immune system [123]. Additionally, S1P controls the migration of osteoclast precursor cells between the bloodstream and the bone marrow, partly mediated by S1PRs expressed on the surface of osteoclast precursor cells (S1P₁ and S1P₂) [124]. Animal studies have indicated that inhibition of S1P₂ function alters the dynamics of osteoclast precursor cell migration, thereby alleviating osteoporosis [125]. However, inflammatory cytokines, such as IL-6, IL-17, and IL-1 β , play important roles in bone loss. IL-6 increases the number of osteoclast precursor cells by upregulating S1P₂, contributing to inflammation-induced bone loss. In a mouse model of RA, elevated IL-6 expression upregulates S1P₂, thereby increasing the number of osteoclast precursor cells. These findings suggest the need to control the expression of inflammatory cytokines in the treatment of RA to prevent systemic bone loss [126]. S1P promotes IL-1 β expression in osteoblasts through the S1P₁ receptor and JAK/STAT3 signaling pathway. Knocking out SPHK1 reduces IL-1 β expression in osteoblasts [101], and IL-17 enhances TNF- α -induced IL-6 synthesis in osteoblasts via activation of the p38 MAP kinase [127]. In addition to IL-17 stimulation, S1P can also promote IL-6 expression in osteoblasts through the phosphoinositide 3-kinases (PI3K), MEK/ERK, and NF- κ B signaling pathways [128]. The elevation of inflammatory cytokines induces VEGF production, exacerbating bone loss. Therefore, IL-6 receptor antibodies alleviate systemic bone loss and reduce the number of osteoclast precursor cells in the tibial bone marrow by downregulating S1P₂ [126]. Similarly, symptom relief can also be achieved by blocking IL-17 expression and reducing IL-6 synthesis. Another study suggests that active vitamin D regulates the migration behavior of osteoclast precursor cells in the circulation, which helps to limit osteoclastogenic bone resorption [129]. Osteoclasts, a special subpopulation of cells with bone-resorbing ability, play a critical role in maintaining normal bone homeostasis, degrading old bone, and promoting the formation of new bone. The expression of SPHK and the production of S1P in osteoclasts stimulate the migration of mesenchymal stem cells by activating kinase signaling pathways [130]. S1P also promotes osteoblastogenesis. Under RANKL stimulation, S1P expression is enhanced in osteoclast precursors, and by binding to S1PRs on the surface of osteoblasts, S1P promotes the migration and survival of osteoblasts. Subsequently, S1P-activated osteoblasts upregulate RANKL expression, which stimulates the migration, fusion, activation, and survival of osteoclasts, leading to increased osteoclastogenesis [131, 132]. Osteoclasts participate in cartilage damage in osteoarthritis via the S1P/S1P₂ pathway. The activity of SPHK1 increases with osteoclast differentiation, and injecting sphingosine analogs into the joint cavity of mice reduces

S1P levels and alleviates bone arthritis [133]. Furthermore, FTY720 alleviates ovariectomy-induced osteoporosis in mice by promoting the recirculation of cell populations containing osteoclast precursors and reducing the number of mature osteoclasts attached to the bone surface [124]. Studies have also shown that SPHK inhibitors such as FTY720 combined with zoledronate can reduce inflammatory bone erosion and inhibit osteoclast differentiation [134]. The levels of S1P generated by SPHK phosphorylation are positively correlated with bone formation markers. By inhibiting S1P lyase to increase S1P levels, there is a significant increase in the quality and strength of bone formation [135]. Regulating the signaling pathways of SPHK1/S1P/S1P_{1/2} holds promise as an effective approach for treating bone metabolic disorders, such as osteoporosis, RA, and osteoarthritis.

The role of S1P and S1PRs in Sjogren's syndrome

Sjogren's syndrome is a systemic autoimmune disease characterized by the gradual destruction of the lacrimal and salivary glands due to an inflammatory process, resulting in dryness of the eyes and mouth [136]. The affected glands are heavily infiltrated by lymphocytes, with T lymphocytes being more prominent in mild cases, while B lymphocytes are most representative in severe cases [137]. Previous studies have demonstrated the involvement of the S1P/S1P₁ axis in the pathogenesis of the disease [138]. Cellular studies have shown the expression of S1P_{1/2/3/4} receptors, as well as SPHK1 and SPHK2, in human submandibular gland cells, and further evidence suggests that S1P triggers the Ca²⁺ signaling pathway and apoptotic pathway in normal submandibular cells, thus affecting the progression of Sjogren's syndrome by interfering with S1P_{1/2/3/4}. Additionally, S1P induces the expression of Fas and IL-6, which are related to the pathology of Sjogren's syndrome [139]. Clinical research has indicated that the S1P signaling pathway may regulate the autoimmune phenotype of Sjogren's syndrome through immune cells and epithelial cells. Lymphocytes in the peripheral blood of primary Sjogren's syndrome patients are significantly higher compared to healthy individuals [138], and S1P plays a major role in lymphocyte migration through interaction with its receptors, S1PRs. Gengerimod can inhibit lymphocyte entry into the circulation, reduce immune infiltration of salivary glands, alleviate salivary gland inflammation, and relieve symptoms in viral-induced salivary gland C57 mouse models, among others. In the chronic sialadenitis mouse model, oral administration of Gengerimod can reduce salivary gland inflammation, as well as T cells and proliferating plasma cells in ectopic lymphoid structures within the salivary gland, leading to a decrease in disease-associated autoantibodies [140]. These findings suggest that future treatments for Sjogren's syndrome may consider the use of S1P_{1/2/3/4} receptor inhibitors to control its progression.

Role of S1P and S1PRs in pulmonary hypertension associated with connective tissue disease

Pulmonary arterial hypertension (PAH) is a prevalent and critical complication of connective tissue diseases, character-

ized by progressive alterations in pulmonary vascular function and structure. The primary pathophysiological manifestations of PAH involve increased vasodilatory tone, impairment in vascular endothelial function, and structural remodeling in the small- and medium-sized pulmonary arteries. These changes give rise to increased pulmonary vascular resistance, right-sided heart hypertrophy, and ultimately, right ventricular failure [141,142]. The development of connective tissue disease-associated PAH is linked to autoimmunity against G protein-coupled receptors, while the involvement of S1P/S1PR signaling in the pathogenesis of PAH has also been reported [143]. S1P is expressed at elevated levels in the human lung [144] and facilitates autophagy activation by accelerating the upregulation of TNF receptor-related factor 2-mediated BECN1 and ubiquitination. Subsequently, cadherin-1 degradation and proliferation of pulmonary artery smooth muscle cells (PASMCs) occur [145]. Another signaling pathway, the Smad2/3/SPHK1/S1P/Notch3 pathway, mediated by tumor growth factor beta 1 (TGF- β 1), also induces PASMC proliferation [146]. Therefore, these aforementioned pathways hold potential as targets for the prevention of PAH. Not only does SPHK/S1P mediates the proliferation of PASMCs and remodeling of the pulmonary artery [141] but S1PRs also regulate the proliferation of PASMCs [147], potentially due to the high expression of S1PRs in these cells. Inflammatory cells and PASMCs represent two major cell populations involved in the pathogenesis of PAH. The processes of inflammation and angiogenesis contribute to vascular remodeling and the excessive proliferation of smooth muscle cells, which are characteristic of severe PAH [148,149]. S1P/S1PR signaling plays a crucial role in maintaining vascular dynamic homeostasis. Impairment in S1P/S1PR signaling leads to vasoconstriction, vascular remodeling, and endothelial dysfunction, thereby increasing pulmonary vascular resistance and mean pulmonary arterial pressure, ultimately culminating in PAH. Early blockade of S1P₂, inhibition of Notch3, silencing of STAT3, miRNA-135b, or YAP all attenuate S1P-induced proliferation and vascular remodeling of PASMCs, thus preventing the development of PAH [150]. Animal experiments have confirmed the finding of increased SPHK1 protein levels and S1P production in the lung tissue of animal models of idiopathic pulmonary hypertension [151], and inhibition of SPHK1, S1P, or S1P₂ slowed down the progression of PAH induced by chronic hypoxia in animals or wild lily pads [152]. However, the treatment of PAH remains challenging. Animal experiments have shown that resveratrol effectively inhibits the progression of PAH by blocking SPHK1/S1P-mediated NF- κ B activation [151]. Similarly, serpentine inhibits highly expressed S1P by regulating the metabolic enzyme SPHK1 in PAH animals. The drug also downregulates miRNA-21, Akt phosphorylation, and mTOR phosphorylation, thereby inhibiting SPHK1 expression. These findings present a potential treatment for connective tissue disease-associated PAH [153]. Stem cell therapy has emerged as a promising therapeutic modality for PAH, with human bone marrow mesenchymal stem cells pretreated with S1P enhancing the therapeutic effect on PAH. Additionally, S1P-induced mesenchymal stem cells enhance the anti-inflammatory and

angiogenic activity of fibroblasts in culture. Furthermore, in wild lily vine-induced PAH animal experiments, cells stimulated with S1P significantly reduce the elevation of right ventricular systolic pressure [154]. By optimizing stem cell therapy in the future and combining it with potential targeted drugs for PAH, novel treatment approaches may be developed for this condition.

Conclusion

The pleiotropic bioactive lipid S1P and its S1PRs play crucial roles in the development and functioning of the immune, cardiovascular, neurological, and skeletal systems. The significance of the S1P/S1PR axis in autoimmune diseases has paved the way for innovative treatments in this domain. Initially, S1P was discovered to regulate the efflux of lymphocytes from lymphoid tissues. By modulating the S1P/S1PR pathway, autoimmune diseases can be attenuated in terms of immune and inflammatory responses. Further research has shed light on the neuroprotective effects of this pathway through its interaction with S1PR on neuronal cells. The advent of S1PR modulators has revolutionized the treatment landscape not only for multiple sclerosis but also for other autoimmune diseases, such as systemic lupus erythematosus, rheumatoid arthritis and systemic sclerosis. Nonetheless, the quest for an optimal treatment for numerous autoimmune diseases persists. As a result, early diagnosis becomes of paramount importance, as it enables clinicians to tailor patient-specific treatments.

Given that the mechanism of the S1P/S1PR signaling pathway in autoimmune disorders is still under intensive investigation, the conclusions presented in this review may exhibit a slight bias. Nevertheless, this review has the potential to enhance clinicians' comprehension of the advancements made in the field of S1P/S1PR signaling pathway in autoimmune diseases. Further extensive research into the functionality of S1P signaling in various diseases, combined with the development of more targeted drugs alongside their optimal administration systems, will introduce novel therapeutic alternatives. The pursuit of more specific pharmacological agents and the establishment of refined drug delivery systems will yield innovative therapeutic strategies. The judicious manipulation and utilization of S1PR modulators, coupled with a comprehensive understanding of the crucial role played by the S1P/S1PRs axis in autoimmune diseases, present themselves as promising avenues for future exploration.

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References

- [1] Danieli MG, Antonelli E, Piga MA, Claudi I, Palmeri D, Tonacci A, et al. Alarmins in autoimmune diseases. *Autoimmun Rev* 2022 Sep;21(9):103142. <https://doi.org/10.1016/j.autrev.2022.103142>.

- [2] Xiao ZX, Miller JS, Zheng SG. An updated advance of autoantibodies in autoimmune diseases. *Autoimmun Rev* 2021 Feb;20(2):102743. <https://doi.org/10.1016/j.autrev.2020.102743>.
- [3] Yu C, Gershwin ME, Chang C. Diagnostic criteria for systemic lupus erythematosus: a critical review. *J Autoimmun* 2014 Feb-Mar;48:49:10-3. <https://doi.org/10.1016/j.jaut.2014.01.004>.
- [4] Burg N, Salmon JE, Hla T. Sphingosine 1-phosphate receptor-targeted therapeutics in rheumatic diseases. *Nat Rev Rheumatol* 2022 Jun;18(6):335-51. <https://doi.org/10.1038/s41584-022-00784-6>.
- [5] Wang X, Qiu Z, Dong W, Yang Z, Wang J, Xu H, et al. S1PR1 induces metabolic reprogramming of ceramide in vascular endothelial cells, affecting hepatocellular carcinoma angiogenesis and progression. *Cell Death Dis* 2022 Sep;13(9):768. <https://doi.org/10.1038/s41419-022-05210-z>.
- [6] Fyrst H, Saba JD. An update on sphingosine-1-phosphate and other sphingolipid mediators. *Nat Chem Biol* 2010 Jul;6(7):489-97. <https://doi.org/10.1038/nchembio.392>. Erratum in: *Nat Chem Biol* 2010 Sep;6(9):689. <https://doi.org/10.1038/nchembio0910-689a>.
- [7] Jozefczuk E, Guzik TJ, Siedlinski M. Significance of sphingosine-1-phosphate in cardiovascular physiology and pathology. *Pharmacol Res* 2020 Jun;156:104793. <https://doi.org/10.1016/j.phrs.2020.104793>.
- [8] Maceyka M, Sankala H, Hait NC, Le Stunff H, Liu H, Toman R, et al. SphK1 and SphK2, sphingosine kinase isoenzymes with opposing functions in sphingolipid metabolism. *J Biol Chem* 2005 Nov;280(44):37118-29. <https://doi.org/10.1074/jbc.M502207200>.
- [9] Rivera J, Proia RL, Olivera A. The alliance of sphingosine-1-phosphate and its receptors in immunity. *Nat Rev Immunol* 2008 Oct;8(10):753-63. <https://doi.org/10.1038/nri2400>.
- [10] Tolksdorf C, Moritz E, Wolf R, Meyer U, Marx S, Bien-Möller S, et al. Platelet-derived S1P and its relevance for the communication with immune cells in multiple human diseases. *Int J Mol Sci* 2022 Sep;23(18):10278. <https://doi.org/10.3390/ijms231810278>.
- [11] Vu TM, Ishizu AN, Foo JC, Toh XR, Zhang F, Whee DM, et al. Mfsd2b is essential for the sphingosine-1-phosphate export in erythrocytes and platelets. *Nature* 2017 Oct 26;550(7677):524-8. <https://doi.org/10.1038/nature24053>.
- [12] Hisano Y, Kobayashi N, Yamaguchi A, Nishi T. Mouse SPNS2 functions as a sphingosine-1-phosphate transporter in vascular endothelial cells. *PLoS One* 2012;7(6):e38941. <https://doi.org/10.1371/journal.pone.0038941>.
- [13] Mendoza A, Bréart B, Ramos-Perez WD, Pitt LA, Gobert M, Sunkara M, et al. The transporter Spns2 is required for secretion of lymph but not plasma sphingosine-1-phosphate. *Cell Rep* 2012 Nov;2(5):1104-10. <https://doi.org/10.1016/j.celrep.2012.09.021>.
- [14] Obinata H, Hla T. Sphingosine 1-phosphate and inflammation. *Int Immunol* 2019 Aug;31(9):e17-25. <https://doi.org/10.1093/intimm/dxz037>.
- [15] Nagahashi M, Yamada A, Aoyagi T, Allegood J, Wakai T, et al. Sphingosine-1-phosphate in the lymphatic fluid determined by novel methods. *Heliyon* 2016 Dec;2(12):e00219. <https://doi.org/10.1016/j.heliyon.2016.e00219>.
- [16] Baeyens AAL, Schwab SR. Finding a way out: S1P signaling and immune cell migration. *Annu Rev Immunol* 2020 Apr;38:759-84. <https://doi.org/10.1146/annurev-immunol-081519-083952>.
- [17] Schwab SR, Pereira JP, Matloubian M, Xu Y, Huang Y, Cyster JG. Lymphocyte sequestration through S1P lyase inhibition and disruption of S1P gradients. *Science* 2005 Sep;309(5741):1735-9. <https://doi.org/10.1126/science.1113640>.
- [18] Zhang H, Desai NN, Olivera A, Seki T, Brooker G, Spiegel S. Sphingosine-1-phosphate, a novel lipid, involved in cellular proliferation. *J Cell Biol* 1991 Jul;114(1):155-67. <https://doi.org/10.1083/jcb.114.1.155>.
- [19] Olivera A, Spiegel S. Sphingosine-1-phosphate as second messenger in cell proliferation induced by PDGF and FCS mitogens. *Nature* 1993 Oct;365(6446):557-60. <https://doi.org/10.1038/365557a0>.
- [20] Bryan AM, Del Poeta M. Sphingosine-1-phosphate receptors and innate immunity. *Cell Microbiol* 2018 May;20(5):e12836. <https://doi.org/10.1111/cmi.12836>.
- [21] Patmanathan SN, Wang W, Yap LF, Herr DR, Paterson IC. Mechanisms of sphingosine 1-phosphate receptor signalling in cancer. *Cell Signal* 2017 Jun;34:66-75. <https://doi.org/10.1016/j.cellsig.2017.03.002>.
- [22] Bravo GÁ, Cedeño RR, Casadevall MP, Ramió-Torrentà L. Sphingosine-1-Phosphate (S1P) and S1P signaling pathway modulators, from current insights to future perspectives. *Cells* 2022 Jun;11(13):2058. <https://doi.org/10.3390/cells11132058>.
- [23] Pan S, Mi Y, Pally C, Beerli C, Chen A, Guerini D, et al. A monoselective sphingosine-1-phosphate receptor-1 agonist prevents allograft rejection in a stringent rat heart transplantation model. *Chem Biol* 2006 Nov;13(11):1227-34. <https://doi.org/10.1016/j.chembiol.2006.09.017>.
- [24] Pitson SM, Pébay A. Regulation of stem cell pluripotency and neural differentiation by lysophospholipids. *Neurosignals* 2009;17(4):242-54. <https://doi.org/10.1159/000231891>.
- [25] Camerer E, Regard JB, Cornelissen I, Srinivasan Y, Duong DN, Palmer D, et al. Sphingosine-1-phosphate in the plasma compartment regulates basal and inflammation-induced vascular leak in mice. *J Clin Invest* 2009 Jul;119(7):1871-9. <https://doi.org/10.1172/JCI38575>.
- [26] Liu Y, Wada R, Yamashita T, Mi Y, Deng CX, Hobson JP, et al. Edg-1, the G protein-coupled receptor for sphingosine-1-phosphate, is essential for vascular maturation. *J Clin Invest* 2000 Oct;106(8):951-61. <https://doi.org/10.1172/JCI10905>.
- [27] Karupuchamy T, Behrens EH, González-Cabrera P, Sarkisyan G, Gima L, Boyer JD, et al. Sphingosine-1-phosphate receptor-1 (S1P1) is expressed by lymphocytes, dendritic cells, and endothelium and modulated during inflammatory bowel disease. *Mucosal Immunol* 2017 Jan;10(1):162-71. <https://doi.org/10.1038/mi.2016.35>.
- [28] Seyedsadr MS, Weinmann O, Amorim A, Ineichen BV, Egger M, Mirnajafi-Zadeh J, et al. Inactivation of sphingosine-1-phosphate receptor 2 (S1PR2) decreases demyelination and enhances remyelination in animal models of multiple sclerosis. *Neurobiol Dis* 2019 Apr;124:189-201. <https://doi.org/10.1016/j.nbd.2018.11.018>.
- [29] Blankenbach KV, Schwalm S, Pfeilschifter J, Meyer Zu Heringdorf D. Sphingosine-1-phosphate receptor-2 antagonists: therapeutic potential and potential risks. *Front Pharmacol* 2016 Jun;7:167. <https://doi.org/10.3389/fphar.2016.00167>.
- [30] Sun X, Singleton PA, Letsiou E, Zhao J, Belvitch P, Sammani S, et al. Sphingosine-1-phosphate receptor-3 is a novel biomarker in acute lung injury. *Am J Respir Cell Mol Biol* 2012 Nov;47(5):628-36. <https://doi.org/10.1165/rcmb.2012-0048OC>.
- [31] Li Q, Li Y, Lei C, Tan Y, Yi G. Sphingosine-1-phosphate receptor 3 signaling. *Clin Chim Acta* 2021 Aug;519:32-9. <https://doi.org/10.1016/j.cca.2021.03.025>.
- [32] Shea BS, Opal SM. The role of S1PR3 in protection from bacterial sepsis. *Am J Respir Crit Care Med* 2017 Dec;196(12):1500-2. <https://doi.org/10.1164/rccm.201708-1726ED>.
- [33] Yang C, Yamashita M, Suda T. A novel function of sphingolipid signaling via S1PR3 in hematopoietic and leukemic stem cells. *Blood Cancer Discov* 2020 Dec;2(1):3-5. <https://doi.org/10.1158/2643-3230.BCD-20-0200>.
- [34] Olesch C, Siraït-Fischer E, Berkefeld M, Fink AF, Susen RM, Ritter B, et al. S1PR4 ablation reduces tumor growth and improves chemotherapy via CD8+ T cell expansion. *J Clin Invest* 2020 Oct;130(10):5461-76. <https://doi.org/10.1172/JCI136928>.
- [35] Olesch C, Ringel C, Brüne B, Weigert A. Beyond immune cell migration: the emerging role of the sphingosine-1-phosphate receptor S1PR4 as a modulator of innate immune cell activation. *Mediators Inflamm* 2017;2017:6059203. <https://doi.org/10.1155/2017/6059203>.
- [36] Kunkel GT, Maceyka M, Milstien S, Spiegel S. Targeting the sphingosine-1-phosphate axis in cancer, inflammation and beyond. *Nat Rev Drug Discov* 2013 Sep;12(9):688-702. <https://doi.org/10.1038/nrd4099>.
- [37] Anwar M, Mehta D. Post-translational modifications of S1PR1 and endothelial barrier regulation. *Biochim Biophys Acta Mol Cell Biol Lipids* 2020 Sep;1865(9):158760. <https://doi.org/10.1016/j.bbalip.2020.158760>.
- [38] Jaillard C, Harrison S, Stankoff B, Aigrot MS, Calver AR, Duddy G, et al. Edg8/S1P5: an oligodendroglial receptor with dual function on process retraction and cell survival. *J Neurosci* 2005 Feb;25(6):1459-69. <https://doi.org/10.1523/JNEUROSCI.4645-04.2005>.
- [39] Chun J, Giovannoni G, Hunter SF. Sphingosine 1-phosphate receptor modulator therapy for multiple sclerosis: differential downstream receptor signalling and clinical profile effects. *Drugs* 2021 Feb;81(2):207-31. <https://doi.org/10.1007/s40265-020-01431-8>.

- [40] Ruiz F, Vigne S, Pot C. Resolution of inflammation during multiple sclerosis. *Semin Immunopathol* 2019 Nov;41(6):711–26. <https://doi.org/10.1007/s00281-019-00765-0>.
- [41] Reich DS, Lucchinetti CF, Calabresi PA. Multiple sclerosis. *N Engl J Med* 2018 Jan;378(2):169–80. <https://doi.org/10.1056/NEJMra1401483>.
- [42] Katz Sand I. Classification, diagnosis, and differential diagnosis of multiple sclerosis. *Curr Opin Neurol* 2015 Jun;28(3):193–205. <https://doi.org/10.1097/WCO.0000000000000206>.
- [43] Gaire BP, Choi JW. Critical roles of lysophospholipid receptors in activation of neuroglia and their neuroinflammatory responses. *Int J Mol Sci* 2021 Jul;22(15):7864. <https://doi.org/10.3390/ijms22157864>.
- [44] Choi JW, Chun J. Lysophospholipids and their receptors in the central nervous system. *Biochim Biophys Acta* 2013 Jan;1831(1):20–32. <https://doi.org/10.1016/j.bbali.2012.07.015>.
- [45] Rivera R, Chun J. Biological effects of lysophospholipids. *Rev Physiol Biochem Pharmacol* 2008;160:25–46. https://doi.org/10.1007/112_0507.
- [46] Moon E, Han JE, Jeon S, Ryu JH, Choi JW, Chun J. Exogenous S1P exposure potentiates ischemic stroke damage that is reduced possibly by inhibiting S1P receptor signaling. *Mediators Inflamm* 2015;2015:492659. <https://doi.org/10.1155/2015/492659>.
- [47] Santos-Nogueira E, López-Serrano C, Hernández J, Lago N, Astudillo AM, Balsinde J, et al. Activation of lysophosphatidic acid receptor type 1 contributes to pathophysiology of spinal cord injury. *J Neurosci* 2015 Jul;35(28):10224–35. <https://doi.org/10.1523/JNEUROSCI.4703-14.2015>.
- [48] Gaire BP, Choi JW. Sphingosine 1-phosphate receptors in cerebral ischemia. *Neuromolecular Med* 2021 Mar;23(1):211–23. <https://doi.org/10.1007/s12017-020-08614-2>.
- [49] Pérez-Jeldres T, Alvarez-Lobos M, Rivera-Nieves J. Targeting sphingosine-1-phosphate signaling in immune-mediated diseases: beyond multiple sclerosis. *Drugs* 2021 Jun;81(9):985–1002. <https://doi.org/10.1007/s40265-021-01528-8>.
- [50] Groves A, Kihara Y, Chun J. Fingolimod: direct CNS effects of sphingosine 1-phosphate (S1P) receptor modulation and implications in multiple sclerosis therapy. *J Neurol Sci* 2013 May;328(1-2):9–18. <https://doi.org/10.1016/j.jns.2013.02.011>.
- [51] McGinley MP, Cohen JA. Sphingosine 1-phosphate receptor modulators in multiple sclerosis and other conditions. *Lancet* 2021 Sep;398(10306):1184–94. [https://doi.org/10.1016/S0140-6736\(21\)00244-0](https://doi.org/10.1016/S0140-6736(21)00244-0).
- [52] Tran JQ, Hartung JP, Peach RJ, Boehm MF, Rosen H, Smith H, et al. Results from the first-in-human study with Ozanimod, a novel, selective sphingosine-1-phosphate receptor modulator. *J Clin Pharmacol* 2017 Aug;57(8):988–96. <https://doi.org/10.1002/jcph.887>.
- [53] Fronza M, Loreface L, Frau J, Cocco E. An overview of the efficacy and safety of ozanimod for the treatment of relapsing multiple sclerosis. *Drug Des Devel Ther* 2021 May;15:1993–2004. <https://doi.org/10.2147/DDDT.S240861>.
- [54] Cohen JA, Arnold DL, Comi G, Bar-Or A, Gujrathi S, Hartung JP, et al. Safety and efficacy of the selective sphingosine 1-phosphate receptor modulator ozanimod in relapsing multiple sclerosis (RADIANCE): a randomised, placebo-controlled, phase 2 trial. *Lancet Neurol* 2016 Apr;15(4):373–81. [https://doi.org/10.1016/S1474-4422\(16\)00018-1](https://doi.org/10.1016/S1474-4422(16)00018-1).
- [55] ClinicalTrials.gov A phase 2/3, Multi-Center, randomized, double-blind, placebo-controlled [Part A] and double-blind, double-dummy, active-controlled [part b], parallel group study to evaluate the efficacy and safety of rpr1063 administered orally to relapsing multiple sclerosis patients 2017. [cited 2017 Aug 6]. Available from: <https://clinicaltrials.gov/ct2/show/NCT02047734?term=NCT02047734&rank=1>.
- [56] Gentile A, De Vito F, Fresegna D, Rizzo FR, Bullitta S, Guadalupi L, et al. Peripherical T cells from multiple sclerosis patients trigger synaptotoxic alterations in central neurons. *Neuropathol Appl Neurobiol* 2020 Feb;46(2):160–70. <https://doi.org/10.1111/nan.12569>.
- [57] La Mantia L, Tramacere I, Firwana B, Pacchetti I, Palumbo R, Filippini G. Fingolimod for relapsing-remitting multiple sclerosis. *Cochrane Database Syst Rev* 2016 Apr;4:CD009371. <https://doi.org/10.1002/14651858.CD009371.pub2>.
- [58] Edmonds Y, Milstien S, Spiegel S. Development of small-molecule inhibitors of sphingosine-1-phosphate signaling. *Pharmacol Ther* 2011 Dec;132(3):352–60. <https://doi.org/10.1016/j.pharmthera.2011.08.004>.
- [59] Chitnis T, Arnold DL, Banwell B, Brück W, Ghezzi A, Giovannoni G, et al. Trial of fingolimod versus interferon beta-1a in pediatric multiple sclerosis. *N Engl J Med* 2018 Sep;379(11):1017–27. <https://doi.org/10.1056/NEJMoa1800149>.
- [60] Tsai HC, Han MH. Sphingosine-1-phosphate (S1P) and S1P signaling pathway: therapeutic targets in autoimmunity and inflammation. *Drugs* 2016 Jul;76(11):1067–79. <https://doi.org/10.1007/s40265-016-0603-2>.
- [61] Rothhammer V, Kenison JE, Tjon E, Takenaka MC, de Lima KA, Borucki DM, et al. Sphingosine 1-phosphate receptor modulation suppresses pathogenic astrocyte activation and chronic progressive CNS inflammation. *Proc Natl Acad Sci U S A* 2017 Feb;114(8):2012–7. <https://doi.org/10.1073/pnas.1615413114>.
- [62] Lucaciu A, Brunkhorst R, Pfeilschifter JM, Pfeilschifter W, Subburayalu J. The S1P-S1PR axis in neurological disorders—insights into current and future therapeutic perspectives. *Cells* 2020 Jun;9(6):1515. <https://doi.org/10.3390/cells9061515>.
- [63] Choi JW, Gardell SE, Herr DR, Rivera R, Lee CW, Noguchi K, et al. FTY720 (fingolimod) efficacy in an animal model of multiple sclerosis requires astrocyte sphingosine 1-phosphate receptor 1 (S1P1) modulation. *Proc Natl Acad Sci U S A* 2011 Jan;108(2):751–6. <https://doi.org/10.1073/pnas.1014154108>.
- [64] O'Sullivan SA, O'Sullivan C, Healy LM, Dev KK, Sheridan GK. Sphingosine 1-phosphate receptors regulate TLR4-induced CXCL5 release from astrocytes and microglia. *J Neurochem* 2018 Mar;144(6):736–47. <https://doi.org/10.1111/jnc.14313>.
- [65] Stepanovska B, Zivkovic A, Enzmann G, Tietz S, Homann T, Kleuser B, et al. Morpholino analogues of fingolimod as novel and selective S1P₁ ligands with in vivo efficacy in a mouse model of experimental antigen-induced encephalomyelitis. *Int J Mol Sci* 2020 Sep;21(18):6463. <https://doi.org/10.3390/ijms21186463>.
- [66] Imeri F, Stepanovska B, Zivkovic A, Enzmann G, Schwalm S, et al. Novel compounds with dual S1P receptor agonist and histamine H₃ receptor antagonist activities act protective in a mouse model of multiple sclerosis. *Neuropharmacology* 2021 Mar;186:108464. <https://doi.org/10.1016/j.neuropharm.2021.108464>.
- [67] Spampinato SF, Merlo S, Sano Y, Kanda T, Sortino MA. Protective effect of the sphingosine-1 phosphate receptor agonist siponimod on disrupted blood brain barrier function. *Biochem Pharmacol* 2021 Apr;186:114465. <https://doi.org/10.1016/j.bcp.2021.114465>.
- [68] Wu Q, Mills EA, Wang Q, Dowling CA, Fisher C, Kirch B, et al. Siponimod enriches regulatory T and B lymphocytes in secondary progressive multiple sclerosis. *JCI Insight* 2020 Feb;5(3):e134251. <https://doi.org/10.1172/jci.insight.134251>.
- [69] Cree BA, Arnold DL, Fox RJ, Gold R, Vermersch P, Benedict RH, et al. Long-term efficacy and safety of siponimod in patients with secondary progressive multiple sclerosis: analysis of EXPAND core and extension data up to >5 years. *Mult Scler* 2022 Sep;28(10):1591–605. <https://doi.org/10.1177/13524585221083194>.
- [70] Kappos L, Bar-Or A, Cree BAC, Fox RJ, Giovannoni G, Gold R, et al. Siponimod versus placebo in secondary progressive multiple sclerosis (EXPAND): a double-blind, randomised, phase 3 study. *Lancet* 2018 Mar;391(10127):1263–73. [https://doi.org/10.1016/S0140-6736\(18\)30475-6](https://doi.org/10.1016/S0140-6736(18)30475-6).
- [71] Gentile A, Musella A, Bullitta S, Fresegna D, De Vito F, Fantozzi R, et al. Siponimod (BAF312) prevents synaptic neurodegeneration in experimental multiple sclerosis. *J Neuroinflammation* 2016 Aug;13(1):207. <https://doi.org/10.1186/s12974-016-0686-4>.
- [72] Cuzzocrea S, Doyle T, Campolo M, Paterniti I, Esposito E, Farr SA, et al. Sphingosine 1-phosphate receptor subtype 1 as a therapeutic target for brain trauma. *J Neurotrauma* 2018 Jul;35(13):1452–66. <https://doi.org/10.1089/neu.2017.5391>.
- [73] Cooper GS, Dooley MA, Treadwell EL, St Clair EW, Parks CG, Gilkeson GS. Hormonal, environmental, and infectious risk factors for developing systemic lupus erythematosus. *Arthritis Rheum* 1998 Oct;41(10):1714–24. [https://doi.org/10.1002/1529-0131\(199810\)41:10<1714::AID-ART3>3.0.CO;2-U](https://doi.org/10.1002/1529-0131(199810)41:10<1714::AID-ART3>3.0.CO;2-U).
- [74] Hla T, Venkataraman K, Michaud J. The vascular S1P gradient—cellular sources and biological significance. *Biochim Biophys Acta* 2008 Sep;1781(9):477–82. <https://doi.org/10.1016/j.bbali.2008.07.003>.
- [75] Aringer M. Inflammatory markers in systemic lupus erythematosus. *J Autoimmun* 2020 Jun;110:102374. <https://doi.org/10.1016/j.jaut.2019.102374>.

- [76] Gerossier E, Nayar S, Smith C, Froidevaux S, Barone F, Martinic M. Cenerimod, a potent and selective sphingosine-1-phosphate receptor 1 modulator, controls systemic autoimmunity and organ pathology in mouse models of systemic lupus erythematosus and Sjögren's syndrome [abstract]. *Arthritis Rheumatol* 2019;71. <https://doi.org/10.1002/art.41108>.
- [77] Hermann V, Batalov A, Smakotina S, Juif PE, Cornelisse P. First use of cenerimod, a selective SIP₁ receptor modulator, for the treatment of SLE: a double-blind, randomised, placebo-controlled, proof-of-concept study. *Lupus Sci Med* 2019 Nov;6(1):e000354. <https://doi.org/10.1136/lupus-2019-000354>.
- [78] Strasser DS, Froidevaux S, Sippel V, Gerossier E, Grieder U, Pierlot GM, et al. Preclinical to clinical translation of cenerimod, a novel SIP₁ receptor modulator, in systemic lupus erythematosus. *RMD Open* 2020 Sep;6(2):e001261. <https://doi.org/10.1136/rmdopen-2020-001261>.
- [79] Merscher S, Fornoni A. Podocyte pathology and nephropathy—sphingolipids in glomerular diseases. *Front Endocrinol (Lausanne)* 2014 Jul;5:127. <https://doi.org/10.3389/fendo.2014.00127>.
- [80] Spiegel S, Milstien S. Sphingosine-1-phosphate: an enigmatic signalling lipid. *Nat Rev Mol Cell Biol* 2003 May;4(5):397–407. <https://doi.org/10.1038/nrml103>.
- [81] Watson L, Tullus K, Marks SD, Holt RC, Pilkington C, Beresford MW. Increased serum concentration of sphingosine-1-phosphate in juvenile-onset systemic lupus erythematosus. *J Clin Immunol* 2012 Oct;32(5):1019–25. <https://doi.org/10.1007/s10875-012-9710-3>.
- [82] Merrill AH Jr. De novo sphingolipid biosynthesis: a necessary, but dangerous, pathway. *J Biol Chem* 2002 Jul;277(29):25843–6. <https://doi.org/10.1074/jbc.R200009200>.
- [83] Czyborra C, Bischoff A, Michel MC. Indomethacin differentiates the renal effects of sphingosine-1-phosphate and sphingosylphosphorylcholine. *Naunyn Schmiedebergs Arch Pharmacol* 2006 Apr;373(1):37–44. <https://doi.org/10.1007/s00210-006-0037-6>.
- [84] Scalapino KJ, Tang Q, Bluestone JA, Bonyhadi ML, Daikh DI. Suppression of disease in New Zealand Black/New Zealand White lupus-prone mice by adoptive transfer of ex vivo expanded regulatory T cells. *J Immunol* 2006 Aug;177(3):1451–9. <https://doi.org/10.4049/jimmunol.177.3.1451>.
- [85] Okazaki H, Hirata D, Kamimura T, Sato H, Iwamoto M, Yoshio T, et al. Effects of FTY720 in MRL-lpr/lpr mice: therapeutic potential in systemic lupus erythematosus. *J Rheumatology* Apr 2002, 29 (4) 707–716. Available from: <https://www.jrheum.org/content/29/4/707>
- [86] Gonzalez-Cabrera PJ, Brown S, Studer SM, Rosen H. SIP signaling: new therapies and opportunities. *F1000Prime Rep* 2014 Dec 1;6:109. <https://doi.org/10.12703/P6-109>.
- [87] Schwartz N, Stock AD, Putterman C. Neuropsychiatric lupus: new mechanistic insights and future treatment directions. *Nat Rev Rheumatol* 2019 Mar;15(3):137–52. <https://doi.org/10.1038/s41584-018-0156-8>.
- [88] Zhang Y, Han H, Chu L. Neuropsychiatric lupus erythematosus: future directions and challenges; a systematic review and survey. *Clinics (Sao Paulo)* 2020 Apr;75:e1515. <https://doi.org/10.6061/clinics/2020/e1515>.
- [89] Kivity S, Agmon-Levin N, Zandman-Goddard G, Chapman J, Shoenfeld Y. Neuropsychiatric lupus: a mosaic of clinical presentations. *BMC Med* 2015 Mar;13:43. <https://doi.org/10.1186/s12916-015-0269-8>.
- [90] Shi D, Tian T, Yao S, Cao K, Zhu X, Zhang M, et al. FTY720 attenuates behavioral deficits in a murine model of systemic lupus erythematosus. *Brain Behav Immun* 2018 May;70:293–304. <https://doi.org/10.1016/j.bbi.2018.03.009>.
- [91] Mike EV, Makinde HM, Der E, Stock A, Gulinello M, Gadhvi GT, et al. Neuropsychiatric systemic lupus erythematosus is dependent on sphingosine-1-phosphate signaling. *Front Immunol* 2018 Sep;9:2189. <https://doi.org/10.3389/fimmu.2018.02189>.
- [92] Lee DM, Weinblatt ME. Rheumatoid arthritis. *Lancet* 2001 Sep;358(9285):903–11. [https://doi.org/10.1016/S0140-6736\(01\)06075-5](https://doi.org/10.1016/S0140-6736(01)06075-5).
- [93] McInnes IB, Schett G. The pathogenesis of rheumatoid arthritis. *N Engl J Med* 2011 Dec;365(23):2205–19. <https://doi.org/10.1056/NEJMr1004965>.
- [94] Sun M, Deng R, Wang Y, Wu H, Zhang Z, Bu Y, et al. Sphingosine kinase 1/sphingosine 1-phosphate/sphingosine 1-phosphate receptor 1 pathway: a novel target of geniposide to inhibit angiogenesis. *Life Sci* 2020 Sep;256:117988. <https://doi.org/10.1016/j.lfs.2020.117988>.
- [95] Inoue T, Kohno M, Nagahara H, Murakami K, Sagawa T, Kasahara A, et al. Upregulation of sphingosine-1-phosphate receptor 3 on fibroblast-like synoviocytes is associated with the development of collagen-induced arthritis via increased interleukin-6 production. *PLoS One* 2019 Jun;14(6):e0218090. <https://doi.org/10.1371/journal.pone.0218090>.
- [96] Zhang P, Zhang Q, Shao Z. Silence of SIPR4 represses the activation of fibroblast-like synoviocytes by regulating IL-17/STAT3 signaling pathway. *Inflammation* 2023 Feb;46(1):234–43. <https://doi.org/10.1007/s10753-022-01728-8>.
- [97] Zhao C, Fernandes MJ, Turgeon M, Tancrède S, Di Battista J, Poubelle PE, et al. Specific and overlapping sphingosine-1-phosphate receptor functions in human synoviocytes: impact of TNF- α . *J Lipid Res* 2008 Nov;49(11):2323–37. <https://doi.org/10.1194/jlr.M800143-JLR200>.
- [98] Hait NC, Allegood J, Maceyka M, Strub GM, Harikumar KB, Singh SK, et al. Regulation of histone acetylation in the nucleus by sphingosine-1-phosphate. *Science* 2009 Sep;325(5945):1254–7. <https://doi.org/10.1126/science.1176709>.
- [99] Harikumar KB, Yester JW, Surace MJ, Oyeneran C, Price MM, Huang WC, et al. K63-linked polyubiquitination of transcription factor IRF1 is essential for IL-1-induced production of chemokines CXCL10 and CCL5. *Nat Immunol* 2014 Mar;15(3):231–8. <https://doi.org/10.1038/ni.2810>.
- [100] Ledgerwood LG, Lal G, Zhang N, Garin A, Esses SJ, Ginhoux F, et al. The sphingosine 1-phosphate receptor 1 causes tissue retention by inhibiting the entry of peripheral tissue T lymphocytes into afferent lymphatics. *Nat Immunol* 2008 Jan;9(1):42–53. <https://doi.org/10.1038/ni1534>.
- [101] Hu SL, Huang CC, Tseng TT, Liu SC, Tsai CH, Fong YC, et al. SIP facilitates IL-1 β production in osteoblasts via the JAK and STAT3 signaling pathways. *Environ Toxicol* 2020 Sep;35(9):991–7. <https://doi.org/10.1002/tox.22935>.
- [102] Takeshita H, Kitano M, Iwasaki T, Kitano S, Tsunemi S, Sato C, et al. Sphingosine 1-phosphate (S1P)/S1P receptor 1 signaling regulates receptor activator of NF- κ B ligand (RANKL) expression in rheumatoid arthritis. *Biochem Biophys Res Commun* 2012 Mar;419(2):154–9. <https://doi.org/10.1016/j.bbrc.2012.01.103>.
- [103] Hu PF, Chen Y, Cai PF, Jiang LF, Wu LD. Sphingosine-1-phosphate: a potential therapeutic target for rheumatoid arthritis. *Mol Biol Rep* 2011 Aug;38(6):4225–30. <https://doi.org/10.1007/s11033-010-0545-9>.
- [104] Deng R, Bu Y, Li F, Wu H, Wang Y, Wei W. The interplay between fibroblast-like synovial and vascular endothelial cells leads to angiogenesis via the sphingosine-1-phosphate-induced RhoA-F-Actin and Ras-Erk1/2 pathways and the intervention of geniposide. *Phytother Res* 2021 Sep;35(9):5305–17. <https://doi.org/10.1002/ptr.7211>.
- [105] Wang Y, Wu H, Gui BJ, Liu J, Rong GX, Deng R, et al. Geniposide alleviates VEGF-induced angiogenesis by inhibiting VEGFR2/PKC/ERK1/2-mediated SphK1 translocation. *Phytomedicine* 2022 Jun;100:154068. <https://doi.org/10.1016/j.phymed.2022.154068>.
- [106] Wang Y, Wu H, Deng R, Dai XJ, Bu YH, Sun MH, et al. Geniposide downregulates the VEGF/SphK1/S1P pathway and alleviates angiogenesis in rheumatoid arthritis in vivo and in vitro. *Phytother Res* 2021 Aug;35(8):4347–62. <https://doi.org/10.1002/ptr.7130>.
- [107] Jin J, Ji M, Fu R, Wang M, Xue N, Xiao Q, et al. Sphingosine-1-phosphate receptor subtype 1 (S1P1) modulator IMMHO01 regulates adjuvant- and collagen-induced arthritis. *Front Pharmacol* 2019 Sep;10:1085. <https://doi.org/10.3389/fphar.2019.01085>.
- [108] Varga J, Abraham D. Systemic sclerosis: a prototypic multisystem fibrotic disorder. *J Clin Invest* 2007 Mar;117(3):557–67. <https://doi.org/10.1172/JCI31139>.
- [109] Volkman ER, Fischer A. Update on morbidity and mortality in systemic sclerosis-related interstitial lung disease. *J Scleroderma Relat Disord* 2021 Feb;6(1):11–20. <https://doi.org/10.1177/2397198320915042>.
- [110] Sato S, Fujimoto M, Hasegawa M, Takehara K. Altered blood B lymphocyte homeostasis in systemic sclerosis: expanded naive B cells and diminished but activated memory B cells. *Arthritis Rheum* 2004 Jun;50(6):1918–27. <https://doi.org/10.1002/art.20274>.
- [111] O'Reilly S, Hügler T, van Laar JM. T cells in systemic sclerosis: a reappraisal. *Rheumatology (Oxford)* 2012 Sep;51(9):1540–9. <https://doi.org/10.1093/rheumatology/kes090>.

- [112] Pattanaik D, Brown M, Postlethwaite AE. Vascular involvement in systemic sclerosis (scleroderma). *J Inflamm Res* 2011;4:105–25. <https://doi.org/10.2147/JIR.S18145>.
- [113] Schaper K, Dickhaut J, Japtok L, Kietzmann M, Mischke R, Kleuser B, et al. Sphingosine-1-phosphate exhibits anti-proliferative and anti-inflammatory effects in mouse models of psoriasis. *J Dermatol Sci* 2013 Jul;71(1):29–36. <https://doi.org/10.1016/j.jdermsci.2013.03.006>.
- [114] Wünsche C, Koch A, Goldschmeding R, Schwalm S, Meyer Zu Heringdorf D, Huwiler A, et al. Transforming growth factor β 2 (TGF- β 2)-induced connective tissue growth factor (CTGF) expression requires sphingosine 1-phosphate receptor 5 (SIP5) in human mesangial cells. *Biochim Biophys Acta* 2015 May;1851(5):519–26. <https://doi.org/10.1016/j.bbaliip.2015.01.003>.
- [115] Matloubian M, Lo CG, Cinamon G, Lesneski MJ, Xu Y, Brinkmann V, et al. Lymphocyte egress from thymus and peripheral lymphoid organs is dependent on SIP receptor 1. *Nature* 2004 Jan;427(6972):355–60. <https://doi.org/10.1038/nature02284>.
- [116] Teijaro JR, Studer S, Leaf N, Kiosses WB, Nguyen N, Matsuki K, et al. SIP1-mediated IFNAR1 degradation modulates plasmacytoid dendritic cell interferon- α autoamplification. *Proc Natl Acad Sci U S A* 2016 Feb;113(5):1351–6. <https://doi.org/10.1073/pnas.1525356113>.
- [117] Park MJ, Moon SJ, Lee EJ, Jung KA, Kim EK, Kim DS, et al. IL-1-IL-17 signaling axis contributes to fibrosis and inflammation in two different murine models of systemic sclerosis. *Front Immunol* 2018 Jul;9:1611. <https://doi.org/10.3389/fimmu.2018.01611>.
- [118] Gabsi A, Heim X, Dlala A, Gati A, Sakhri H, Abidi A, et al. TH17 cells expressing CD146 are significantly increased in patients with systemic sclerosis. *Sci Rep* 2019 Nov;9(1):17721. <https://doi.org/10.1038/s41598-019-54132-y>.
- [119] Gonçalves RSG, Pereira MC, Dantas AT, Almeida AR, Marques CDL, Rego MJB, et al. IL-17 and related cytokines involved in systemic sclerosis: perspectives. *Autoimmunity* 2018 Feb;51(1):1–9. <https://doi.org/10.1080/08916934.2017.1416467>.
- [120] Kano M, Kobayashi T, Date M, Tennichi M, Hamaguchi Y, Strasser DS, et al. Attenuation of murine sclerodermatous models by the selective SIP1 receptor modulator cenerimod. *Sci Rep* 2019 Jan;9(1):658. <https://doi.org/10.1038/s41598-018-37074-9>.
- [121] Shea BS, Brooks SF, Fontaine BA, Chun J, Luster AD, Tager AM. Prolonged exposure to sphingosine 1-phosphate receptor-1 agonists exacerbates vascular leak, fibrosis, and mortality after lung injury. *Am J Respir Cell Mol Biol* 2010 Dec;43(6):662–73. <https://doi.org/10.1165/rcmb.2009-0345OC>.
- [122] Maceyka M, Harikumar KB, Milstien S, Spiegel S. Sphingosine-1-phosphate signaling and its role in disease. *Trends Cell Biol* 2012 Jan;22(1):50–60. <https://doi.org/10.1016/j.tcb.2011.09.003>.
- [123] Xiao L, Zhou Y, Friis T, Beagley K, Xiao Y. SIP-SIP1 signaling: the “Sphinx” in osteoimmunology. *Front Immunol* 2019 Jun;10:1409. <https://doi.org/10.3389/fimmu.2019.01409>.
- [124] Ishii M, Egen JG, Klauschen F, Meier-Schellersheim M, Saeki Y, Vacher J, et al. Sphingosine-1-phosphate mobilizes osteoclast precursors and regulates bone homeostasis. *Nature* 2009 Mar;458(7237):524–8. <https://doi.org/10.1038/nature07713>.
- [125] Ishii M, Kikuta J, Shimazu Y, Meier-Schellersheim M, Germain RN. Chemorepulsion by blood SIP regulates osteoclast precursor mobilization and bone remodeling in vivo. *J Exp Med* 2010 Dec;207(13):2793–8. <https://doi.org/10.1084/jem.20101474>.
- [126] Tanaka K, Hashizume M, Mihara M, Yoshida H, Suzuki M, Matsumoto Y. Anti-interleukin-6 receptor antibody prevents systemic bone mass loss via reducing the number of osteoclast precursors in bone marrow in a collagen-induced arthritis model. *Clin Exp Immunol* 2014 Feb;175(2):172–80. <https://doi.org/10.1111/cei.12201>.
- [127] Tokuda H, Kanno Y, Ishisaki A, Takenaka M, Harada A, Kozawa O. Interleukin (IL)-17 enhances tumor necrosis factor- α -stimulated IL-6 synthesis via p38 mitogen-activated protein kinase in osteoblasts. *J Cell Biochem* 2004 Apr;91(5):1053–61. <https://doi.org/10.1002/jcb.20004>.
- [128] Hu SL, Huang CC, Tzeng TT, Liu SC, Tsai CH, Fong YC, et al. SIP promotes IL-6 expression in osteoblasts through the PI3K, MEK/ERK and NF- κ B signaling pathways. *Int J Med Sci* 2020 May;17(9):1207–14. <https://doi.org/10.7150/ijms.44612>.
- [129] Kikuta J, Kawamura S, Okiji F, Shirazaki M, Sakai S, Saito H, et al. Sphingosine-1-phosphate-mediated osteoclast precursor monocyte migration is a critical point of control in antitumor-resorptive action of active vitamin D. *Proc Natl Acad Sci U S A* 2013 Apr;110(17):7009–13. <https://doi.org/10.1073/pnas.1218799110>.
- [130] Quint P, Ruan M, Pederson L, Kassem M, Westendorf JJ, Khosla S, et al. Sphingosine 1-phosphate (SIP) receptors 1 and 2 coordinately induce mesenchymal cell migration through SIP activation of complementary kinase pathways. *J Biol Chem* 2013 Feb;288(8):5398–406. <https://doi.org/10.1074/jbc.M112.413583>.
- [131] Kim JM, Lin C, Stavre Z, Greenblatt MB, Shim JH. Osteoblast-osteoclast communication and bone homeostasis. *Cells* 2020 Sep;9(9):2073. <https://doi.org/10.3390/cells9092073>.
- [132] Takayanagi H. Osteoimmunology and the effects of the immune system on bone. *Nat Rev Rheumatol* 2009 Dec;5(12):667–76. <https://doi.org/10.1038/nrrheum.2009.217>.
- [133] Cherifi C, Latourte A, Vettorazzi S, Tuckermann J, Provot S, Ea HK, et al. Inhibition of sphingosine 1-phosphate protects mice against chondrocyte catabolism and osteoarthritis. *Osteoarthritis Cartilage* 2021 Sep;29(9):1335–45. <https://doi.org/10.1016/j.joca.2021.06.001>.
- [134] Gu M, Pan B, Chen W, Xu H, Wu X, Hu X, et al. SPHK inhibitors and zoledronic acid suppress osteoclastogenesis and wear particle-induced osteolysis. *Front Pharmacol* 2022 Feb;12:794429. <https://doi.org/10.3389/fphar.2021.794429>.
- [135] Weske S, Vaidya M, Reese A, von Wnuck Lipinski K, Keul P, Bayer JK, et al. Targeting sphingosine-1-phosphate lyase as an anabolic therapy for bone loss. *Nat Med* 2018 May;24(5):667–78. <https://doi.org/10.1038/s41591-018-0005-y>.
- [136] Cornec D, Jamin C, Pers JO. Sjögren’s syndrome: where do we stand, and where shall we go? *J Autoimmun* 2014 Jun;51:109–14. <https://doi.org/10.1016/j.jaut.2014.02.006>.
- [137] Christodoulou MI, Kapsogeorgou EK, Moutsopoulos HM. Characteristics of the minor salivary gland infiltrates in Sjögren’s syndrome. *J Autoimmun* 2010 Jun;34(4):400–7. <https://doi.org/10.1016/j.jaut.2009.10.004>.
- [138] Sekiguchi M, Iwasaki T, Kitano M, Kuno H, Hashimoto N, Kawahito Y, et al. Role of sphingosine 1-phosphate in the pathogenesis of Sjögren’s syndrome. *J Immunol* 2008 Feb;180(3):1921–8. <https://doi.org/10.4049/jimmunol.180.3.1921>.
- [139] Seo J, Koo NY, Choi WY, Kang JA, Min JH, Jo SH, et al. Sphingosine-1-phosphate signaling in human submandibular cells. *J Dent Res* 2010 Oct;89(10):1148–53. <https://doi.org/10.1177/0022034510376044>.
- [140] Gerossier E, Nayar S, Froidevaux S, Smith CG, Runser C, Iannizzotto V, et al. Cenerimod, a selective SIP1 receptor modulator, improves organ-specific disease outcomes in animal models of Sjögren’s syndrome. *Arthritis Res Ther* 2021 Nov;23(1):289. <https://doi.org/10.1186/s13075-021-02673-x>.
- [141] Chen J, Tang H, Sysol JR, Moreno-Vinasco L, Shioura KM, Chen T, et al. The sphingosine kinase 1/sphingosine-1-phosphate pathway in pulmonary arterial hypertension. *Am J Respir Crit Care Med* 2014 Nov;190(9):1032–43. <https://doi.org/10.1164/rccm.201401-0121OC>.
- [142] Gluschke H, Siegert E, Minich WB, Hackler J, Riemekasten G, Kuebler WM, et al. Autoimmunity to sphingosine-1-phosphate-receptors in systemic sclerosis and pulmonary arterial hypertension. *Front Immunol* 2022 Jul;13:935787. <https://doi.org/10.3389/fimmu.2022.935787>.
- [143] Cartier A, Hla T. Sphingosine 1-phosphate: lipid signaling in pathology and therapy. *Science* 2019 Oct;366(6463):ear5551. <https://doi.org/10.1126/science.aar5551>.
- [144] Yan X, Wang J, Zhu Y, Feng W, Zhai C, Liu L, et al. SIP induces pulmonary artery smooth muscle cell proliferation by activating calcineurin/NFAT/OPN signaling pathway. *Biochem Biophys Res Commun* 2019 Aug;516(3):921–7. <https://doi.org/10.1016/j.bbrc.2019.06.160>.
- [145] Zhai C, Feng W, Shi W, Wang J, Zhang Q, Yan X, et al. Sphingosine-1-phosphate promotes pulmonary artery smooth muscle cells proliferation by stimulating autophagy-mediated E-cadherin/CDH1 down-regulation. *Eur J Pharmacol* 2020 Oct;884:173302. <https://doi.org/10.1016/j.ejphar.2020.173302>.
- [146] Wang J, Feng W, Li F, Shi W, Zhai C, Li S, et al. SphK1/SIP mediates TGF- β 1-induced proliferation of pulmonary artery smooth muscle cells and its potential mechanisms. *Pulm Circ* 2019 Jan-Mar;9(1):2045894018816977. <https://doi.org/10.1177/2045894018816977>.
- [147] Birker-Robaczewska M, Studer R, Haenig B, Menyhart K, Hofmann S, Nayler O. bFGF induces SIP1 receptor expression and functionality in human pulmonary artery smooth muscle cells. *J Cell Biochem* 2008 Nov;105(4):1139–45. <https://doi.org/10.1002/jcb.21918>.

- [148] Price LC, Wort SJ, Perros F, Dorfmüller P, Huertas A, Montani D, et al. Inflammation in pulmonary arterial hypertension. *Chest* 2012 Jan;141(1):210–21. <https://doi.org/10.1378/chest.11-0793>.
- [149] Groth A, Vrugt B, Brock M, Speich R, Ulrich S, Huber LC. Inflammatory cytokines in pulmonary hypertension. *Respir Res* 2014 Apr;15(1):47. <https://doi.org/10.1186/1465-9921-15-47>.
- [150] Wang J, Yan X, Feng W, Wang Q, Shi W, Chai L, et al. S1P induces proliferation of pulmonary artery smooth muscle cells by promoting YAP-induced Notch3 expression and activation. *J Biol Chem* 2021 Jan-Jun;296:100599. <https://doi.org/10.1016/j.jbc.2021.100599>.
- [151] Shi W, Zhai C, Feng W, Wang J, Zhu Y, Li S, et al. Resveratrol inhibits monocrotaline-induced pulmonary arterial remodeling by suppression of SphK1-mediated NF- κ B activation. *Life Sci* 2018 Oct;210:140–9. <https://doi.org/10.1016/j.lfs.2018.08.071>.
- [152] Shi W, Wang Q, Wang J, Yan X, Feng W, Zhang Q, et al. Activation of yes-associated protein mediates sphingosine-1-phosphate-induced proliferation and migration of pulmonary artery smooth muscle cells and its potential mechanisms. *J Cell Physiol* 2021 Jun;236(6):4694–708. <https://doi.org/10.1002/jcp.30193>.
- [153] Yao L, Yang YX, Cao H, Ren HH, Niu Z, Shi L. Osthole attenuates pulmonary arterial hypertension by the regulation of sphingosine 1-phosphate in rats. *Chin J Nat Med* 2020 Apr;18(4):308–20. [https://doi.org/10.1016/S1875-5364\(20\)30038-8](https://doi.org/10.1016/S1875-5364(20)30038-8).
- [154] Kang H, Kim KH, Lim J, Kim YS, Heo J, Choi J, et al. The therapeutic effects of human mesenchymal stem cells primed with sphingosine-1 phosphate on pulmonary artery hypertension. *Stem Cells Dev* 2015 Jul;24(14):1658–71. <https://doi.org/10.1089/scd.2014.0496>.

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