



University of
Zurich^{UZH}

Zurich Open Repository and
Archive

University of Zurich
University Library
Strickhofstrasse 39
CH-8057 Zurich
www.zora.uzh.ch

Year: 2023

**The importance of microvascular inflammation in ageing and age-related diseases:
a position paper from the ESH working group on small arteries, section of
microvascular inflammation**

Mengozi, Alessandro ; de Ciuceis, Carolina ; Dell'oro, Raffaella ; Georgiopoulos, Georgios ; Lazaridis, Antonios ; Nosalski, Ryszard ; Pavlidis, George ; Tual-Chalot, Simon ; Agabiti-Rosei, Claudia ; Anyfanti, Panagiota ; Camargo, Livia L ; Dąbrowska, Edyta ; Quarti-Trevano, Fosca ; Hellmann, Marcin ; Masi, Stefano ; Mavraganis, Georgios ; Montezano, Augusto C ; Rios, Francesco J ; Winklewski, Pawel J ; Wolf, Jacek ; Costantino, Sarah ; Gkaliagkousi, Eugenia ; Grassi, Guido ; Guzik, Tomasz J ; Ikonomidis, Ignatios ; Narkiewicz, Krzysztof ; Paneni, Francesco ; Rizzoni, Damiano ; Stamatelopoulos, Kimon ; Stellos, Konstantinos ; et al

DOI: <https://doi.org/10.1097/HJH.0000000000003503>

Posted at the Zurich Open Repository and Archive, University of Zurich

ZORA URL: <https://doi.org/10.5167/uzh-257577>

Journal Article

Published Version



The following work is licensed under a Creative Commons: Attribution 4.0 International (CC BY 4.0) License.

Originally published at:

Mengozi, Alessandro; de Ciuceis, Carolina; Dell'oro, Raffaella; Georgiopoulos, Georgios; Lazaridis, Antonios; Nosalski, Ryszard; Pavlidis, George; Tual-Chalot, Simon; Agabiti-Rosei, Claudia; Anyfanti, Panagiota; Camargo, Livia L; Dąbrowska, Edyta; Quarti-Trevano, Fosca; Hellmann, Marcin; Masi, Stefano; Mavraganis, Georgios; Montezano, Augusto C; Rios, Francesco J; Winklewski, Pawel J; Wolf, Jacek; Costantino, Sarah; Gkaliagkousi, Eugenia; Grassi, Guido; Guzik, Tomasz J; Ikonomidis, Ignatios; Narkiewicz, Krzysztof; Paneni, Francesco; Rizzoni, Damiano; Stamatelopoulos, Kimon; Stellos, Konstantinos; et al (2023). The importance of microvascular inflammation in ageing and age-related diseases: a position paper from the ESH working group on small arteries, section of microvascular inflammation. *Journal of Hypertension*, 41(10):1521-1543.

DOI: <https://doi.org/10.1097/HJH.0000000000003503>

The importance of microvascular inflammation in ageing and age-related diseases: a position paper from the ESH working group on small arteries, section of microvascular inflammation

Alessandro Mengozzi^{a,b,c}, Carolina de Ciuceis^d, Raffaella Dell'oro^e, Georgios Georgiopoulos^f, Antonios Lazaridis^g, Ryszard Nosalski^{h,i,j}, George Pavlidis^{k,l}, Simon Tual-Chalot^m, Claudia Agabiti-Rosei^d, Panagiota Anyfantiⁿ, Livia L. Camargo^{o,p}, Edyta Dabrowska^{q,r}, Fosca Quarti-Trevano^e, Marcin Hellmann^s, Stefano Masi^{a,t}, Georgios Mavraganis^f, Augusto C. Montezano^{o,p}, Francesco J. Rios^{o,p}, Pawel J. Winklewski^u, Jacek Wolf^q, Sarah Costantino^{b,v}, Eugenia Gkaliagkousi^g, Guido Grassi^e, Tomasz J. Guzik^{h,i,j}, Ignatios Ikonomidis^{k,l}, Krzysztof Narkiewicz^q, Francesco Paneni^{b,v,w}, Damiano Rizzoni^{d,x}, Kimon Stamatelopoulos^f, Konstantinos Stellos^{m,y,z,aa}, Stefano Taddei^a, Rhian M. Touyz^{o,p}, Areti Triantafyllou^g, and Agostino Virdis^a

Microcirculation is pervasive and orchestrates a profound regulatory cross-talk with the surrounding tissue and organs. Similarly, it is one of the earliest biological systems targeted by environmental stressors and consequently involved in the development and progression of ageing and age-related disease. Microvascular dysfunction, if not targeted, leads to a steady derangement of the phenotype, which cumulates comorbidities and eventually results in a nonrescuable, very high-cardiovascular risk. Along the broad spectrum of pathologies, both shared and distinct molecular pathways and pathophysiological alteration are involved in the disruption of microvascular homeostasis, all pointing to microvascular inflammation as the putative primary culprit. This position paper explores the presence and the detrimental contribution of microvascular inflammation across the whole spectrum of chronic age-related diseases, which characterise the 21st-century healthcare landscape. The manuscript aims to strongly affirm the centrality of microvascular inflammation by recapitulating the current evidence and providing a clear synoptic view of the whole cardiometabolic derangement. Indeed, there is an urgent need for further mechanistic exploration to identify clear, very early or disease-specific molecular targets to provide an effective therapeutic strategy against the otherwise unstoppable rising prevalence of age-related diseases.

Keywords: ageing, age-related disease, cardiometabolism, inflammation, microcirculation

Abbreviations: AD, Alzheimer's disease; AGEs, advanced glycation end products; AGO-1, argonaute 1; AngII, angiotensin II; ARD, autoimmune rheumatologic diseases; AS, ankylosing spondylitis; AVR, arteriovenous ratio; BBB, blood-brain barrier; BET, bromodomain and extraterminal

Journal of Hypertension 2023, 41:1521–1543

^aDepartment of Clinical and Experimental Medicine, University of Pisa, Pisa, Italy, ^bCenter for Translational and Experimental Cardiology (CTEC), Department of Cardiology, University Hospital Zurich, University of Zurich, Zurich, Switzerland, ^cHealth Science Interdisciplinary Center, Scuola Superiore Sant'Anna, Pisa, ^dDepartment of Clinical and Experimental Sciences, University of Brescia, Brescia, ^eClinica Medica, Department of Medicine and Surgery, University of Milano-Bicocca, Monza, Italy, ^fAngiology and Endothelial Pathophysiology Unit, Department of Clinical Therapeutics, Medical School, National and Kapodistrian University of Athens, Athens, ^gThird Department of Internal Medicine, Aristotle University of Thessaloniki, Papageorgiou Hospital, Thessaloniki, Greece, ^hCentre for Cardiovascular Sciences; Queen's Medical Research Institute; University of Edinburgh, University of Edinburgh, Edinburgh, UK, ⁱDepartment of Internal Medicine, ^jCenter for Medical Genomics OMICRON, Jagiellonian University Medical College, Krakow, Poland, ^kPreventive Cardiology Laboratory and Clinic of Cardiometabolic Diseases, ^l2nd Cardiology Department, Attikon Hospital, Athens, ^mMedical School, National and Kapodistrian University of Athens, Greece, ⁿBiosciences Institute, Vascular Biology and Medicine Theme, Faculty of Medical Sciences, Newcastle University, Newcastle Upon Tyne, UK, ^oSecond Medical Department, Hippokraton Hospital, Aristotle University of Thessaloniki, Thessaloniki, Greece, ^pInstitute of Cardiovascular and Medical Sciences, University of Glasgow, Glasgow, UK, ^qResearch Institute of the McGill University Health Centre (RI-MUHC), McGill University, Montreal, Canada, ^rDepartment of Hypertension and Diabetology, Center of Translational Medicine, ^sCenter of Translational Medicine, ^tDepartment of Cardiac Diagnostics, Medical University, Gdansk, Poland, ^uInstitute of Cardiovascular Science, University College London, London, UK, ^vDepartment of Human Physiology, Medical University of Gdansk, Poland, ^wUniversity Heart Center, Cardiology, University Hospital Zurich, ^xDepartment of Research and Education, University Hospital Zurich, Zurich, Switzerland, ^yDivision of Medicine, Spedali Civili di Brescia, Montichiari, Brescia, Italy, ^zDepartment of Cardiovascular Research, European Center for Angioscience (ECAS), Medical Faculty Mannheim, Heidelberg University, ^{aa}German Centre for Cardiovascular Research (Deutsches Zentrum für Herz-Kreislauf-Forschung, DZHK), Heidelberg/Mannheim Partner Site and ^{ab}Department of Cardiology, University Hospital Mannheim, Heidelberg University, Mannheim, Germany

Correspondence to Agostino Virdis, MD, Department of Clinical and Experimental Medicine, University of Pisa, Via Savi 10, Pisa, Italy. Tel +39 050 992558; fax +39 050 995728; e-mail: agostino.virdis@unipi.it

Received 14 April 2023 **Revised** 13 June 2023 **Accepted** 14 June 2023

J Hypertens 41:1521–1543 Copyright © 2023 The Author(s). Published by Wolters Kluwer Health, Inc. This is an open access article distributed under the Creative Commons Attribution License 4.0 (CCBY), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

DOI:10.1097/HJH.0000000000003503

domain; CADASIL, cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy; CFR, coronary flow reserve; cGAS-STING, DNA-sensing enzyme cyclic GMP-AMP synthase stimulator of interferon genes; CV, cardiovascular; DM, diabetes mellitus; DPP-4i, dipeptidyl peptidase-4 inhibitors; ECs, endothelial cells; EF, endothelial function; Endo-PAT, non-invasive peripheral arterial tonometry; eNOS, endothelial nitric oxide synthase; ET-1, endothelin-1; ETA, endothelin-1 receptor A; FFA, free fatty acids; FMD, flow-mediated dilation; GLP-1RA, glucagon-like peptide-1 receptor agonists; HFpEF, heart failure with preserved ejection fraction; Hb, deoxygenated haemoglobin; HIF-1 α , hypoxia-inducible factor 1 α ; ICAM-1, intercellular adhesion molecule-1; IFN- γ , interferon- γ ; IL, interleukin; JNK, c-Jun N-terminal kinase; LDF, laser Doppler flowmetry; LSCI, laser speckle contrast imaging; M/L, media-to-lumen; MAPK, mitogen-activated protein kinase; MD, microcirculatory dysfunction; MHT, masked hypertension; MTHFR, methylene-tetrahydrofolate reductase; NADPH, nicotinamide adenine dinucleotide phosphate; NF- κ B, nuclear factor kappa-B; NK, natural killer cell; NLRP3, NOD-like receptor family pyrin domain containing 3; NO, nitric oxide; Nrf2, nuclear factor erythroid 2-related factor 2; O₂Hb, oxygenated haemoglobin; PA, psoriatic arthritis; PAT, pulse amplitude tonometry; PCs, pericytes; PI3K, phosphatidylinositol 3-kinase; PKC, protein kinase C; PVAT, perivascular adipose tissue; RA, rheumatoid arthritis; RAGE, receptors for advanced glycation end products; RONS, reactive oxygen and nitrogen species; ROS, reactive oxygen species; SASP, senescent-associated secretory phenotype; SGLT-2i, sodium-glucose cotransporter-2 inhibitors; Sirt1, mammalian silent information regulator 1; SLE, systemic lupus erythematosus; SPECT, single-photon emission tomography; Th, T helper lymphocytes; tHb, total haemoglobin; TLRs, toll-like receptors; TNF- α , tumour necrosis factor- α ; Treg, T regulatory cells; VCAM-1, vascular cell adhesion molecule-1; VEGF, vascular epithelial growth factor; VSMCs, vascular smooth muscle cells; WCH, white coat hypertension; WMH, white matter hyperintensities

INTRODUCTION: MICROCIRCULATION IN CARDIOVASCULAR DISEASE

Microcirculation is the network of terminal vessels of the systemic circulation comprising arterioles, capillaries and venules less than 100 μ m in diameter [1–3]. This network delivers oxygen and other nutrients to tissues while removing carbon dioxide, cellular waste products and toxins [1]. Importantly, microcirculation also regulates fluid homeostasis, temperature control and inflammatory response [1].

Contemporary evidence suggests that microcirculatory impairment may occur in adulthood and further deteriorate across the lifespan. Ageing progressively decreases blood flow and vessel density, ultimately reducing arterial compliance [1]. Beyond ageing, microcirculatory dysfunction

(MD) characterises a multitude of conditions, including diabetes mellitus (DM), hypercholesterolemia, hypertension, peripheral arterial disease, chronic renal failure, menopause, obesity and chronic inflammatory autoimmune disorders [4–8]. Multiple mechanisms may contribute to microcirculatory impairment, including oxidative stress, enhanced leukocyte adhesion, activation of immune cells (both innate and adaptive [9]), endothelial dysfunction, vasoconstriction, attenuated angiogenesis, increased endothelial permeability, microcirculatory plugging and remodelling, lymphatic dysfunction as well as impaired autoregulation [5,10–16].

MD may develop in multiple tissue beds as an underlying systemic process preceding clinical symptoms long before their onset [17,18]. In this context, MD may reflect an early marker of vascular disease and predispose to the development of atherosclerosis [5]. Accordingly, several minimally or noninvasive techniques have been developed to provide useful MD biomarkers in different vascular beds (summarized in Table 1) [19]. However, although circulating biomarkers, including increased triglycerides, C-reactive protein (CRP), cystatin C, homocysteine, nitric oxide (NO), uric acid, interleukin (IL)-6, N-terminal pro-b-type natriuretic peptide, cardiac troponin, thrombomodulin, reninase, neu-regulin-1, von Willebrand factor, serotonin and asymmetric dimethylarginine, are increased in patients with MD, their clinical use for this purpose is not yet validated [18,20–22].

MD is causally associated with the entire spectrum of ageing and age-related diseases, mainly through pro-inflammatory mechanisms (Fig. 1) [23] and may be the substrate for the further development of numerous cardiovascular (CV) diseases, such as coronary artery disease and heart failure with preserved ejection fraction (HFpEF). MD is also found in extra-cardiac tissues (i.e., brain, retina, and lungs) and clinically manifests as dementia, depression, anxiety, vision loss or pulmonary hypertension [14]. In the same context, MD has been implicated in rheumatic (e.g., skin MD) and oncologic diseases [24–32]. With respect to its prognostic value, MD is associated with an increased risk of short- and long-term adverse CV outcomes [33,34]. Both peripheral and coronary MD has been associated with adverse CV events and mortality [35–44]. Furthermore, MD has been linked with progression to kidney failure [18]. Notably, cerebral small vessel disease features are strongly associated with stroke, dementia—especially Alzheimer's disease (AD) and vascular dementias, depression and all-cause mortality [45]. Uterine and placental MD predispose to the onset of preeclampsia [46,47] and to early postnatal microvascular rarefaction and development of MD in offspring [48,49]. Finally, testicular MD and penile skin MD are linked to endocrine disturbances and the future development of hypertension and CV diseases [50,51]. Collectively, an integrative approach to understanding MD is needed to implement effective early diagnosis and treatment strategies.

THE LINK BETWEEN INFLAMMATION AND EARLY MICROVASCULAR AGEING

The microvasculature is pervasive, and its impairment influences every tissue in the human body [52]. Consequently,

TABLE 1. Currently available methods for assessment of microcirculatory function in humans.

| Technique | Tissue | Method of assessment | Advantages | Limitations |
|-------------------------------------------------------|-------------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Peripheral arterial network Finger plethysmography | Arteries of fingers | Reactive hyperemia index in finger blood flow measured by using finger probes | -Safe and noninvasive -No need for specific training -Totally nonoperator-dependent -Prognostic value for adverse CV events | -More expensive -Environmental conditions and the autonomic nervous system may affect measurements |
| Antebrachial plethysmography | Brachial artery | Quantification of forearm blood flow by means of plethysmography during infusion of vasoactive drugs in the brachial artery immediately after cuff deflation and again at 1 and 5 min of reperfusion | -Safe, noninvasive -It allows the simultaneous study of large conduit vessels and small arteries | -Inexpensive -Small errors in the measurement of arterial diameter will result in large errors in the calculation of flow -Room temperature may affect measurements -Measures the local arterial extensibility |
| Transcutaneous oxygen tension | Skin | Quantity of oxygen molecules transferred to the skin microcirculation after heating skin >40°C | -Wide availability | -Time-consuming -Does not assess all ischemic regions |
| Flow mediated skin fluorescence | Skin | Assessment of microcirculation and metabolic regulation based on the measurements of NADH fluorescence intensity in epidermis | -Quick and simple -Good reproducibility -Flowmotion analysis -Correlated with endothelial biomarkers | -Lack of robust evidence on the prognostic value |
| Iontophoresis | Subpapillary plexus, nutritional capillaries, nerve-axon reflex | Delivery of vasodilators (acetylcholine/ SNP) subdermally and measurement of microcirculatory flow with laser Doppler fluxmetry or single-point probes | -Quick and simple -Small coefficients of variation -Correlated with other microvascular beds -Isolation of nerve-axon reflex | -Cannot distinguish subpapillary plexus from nutritional capillaries in glabrous skin |
| Skin pulp blood flow | Nutritional capillaries and AV shunts | Assessment of microcirculation of pulp skin of the toes with laser Doppler fluxmetry or heat and/or radioisotope washout methods | -Simple, can be performed on any area of skin quickly -Distinguishes nutritional capillaries and AV shunts in glabrous skin | -Not correlated with other tissue beds |
| Capillaroscopy | Subpapillary plexus, nutritional capillaries | Evaluation of morphology and blood flow by studying capillary changes with light microscopy or fluorescent dye dynamic capillaroscopy | -Distinguishes microvascular from the interstitial compartments, assesses transcapillary diffusion -Distinguishes subpapillary plexus from nutritional capillaries | -Cost and availability of the equipment -Patients need to be placed in the sitting position -Qualitative evaluation is largely dependent on the operator experience -Need for readily available software among different centers for quantitative evaluation of MD parameters |
| Hand-held vital-microscope device | Sublingual microcirculation, microvascular beds of different types of mucosa and solid organ surfaces | Video observation of the flowing RBCs of the microcirculation (3rd generation device – the newest technology – uses incident darkfield illumination for this purpose) and diffuse capacity | -Noninvasive monitoring of the microcirculation at the bedside -Alterations in sublingual microcirculation are highly sensitive and specific, predicting adverse outcomes -3rd generation device -Computer-controlled image sensor -Better image quality -Evaluation of 30% more capillaries | -Microcirculation can be visualized only if the epithelial layer of the area of interest is thin -Presence of artefacts due to movement -Variability with the use of different generations of HVM |
| Micromyography | Subcutaneous tissues | Measurement of MLR or WLR of small subcutaneous vessels dissected from tissue biopsies by pressure or wire micromyography | -Gold-standard method -Precise and reliable -The most potent predictor of CV events in hypertensive patients | -Locally invasive |
| Laser speckle contrast imaging | Skin and subcutaneous tissues, Retinal and choroidal microcirculation | Measurements of peripheral microcirculatory perfusion on a wide area of tissue LSCI coupled with vascular reactivity tests enables to assess endothelial function Complementary use of a fundus camera with a laser diode or a blue component argon laser for assessment of deep or superficial retinal flow, respectively | -Dynamic, real-time perfusion monitoring -Very good spatial and temporal resolution -Improved spatial and temporal reproducibility as compared to conventional laser Doppler flowmetry -Excellent reproducibility -Safe, noninvasive | -Cost and availability of the equipment -Lack of robust evidence on the prognostic value -Interference by movement artefacts -Limited interpatient comparability -Complexity of quantitative measurements |

TABLE 1 (Continued)

| Technique | Tissue | Method of assessment | Advantages | Limitations |
|------------------------------------------------------------------------------------------------|-----------------------------------|--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|---------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|----------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| Scanning laser Doppler flowmetry | Retinal vascular district | Quantification of the WLR of retinal arterioles using scanning laser Doppler flowmetry | -Easy repeatability and comfortable for patients -Good agreement with wire micromyography | -Lack of robust evidence on the prognostic value -Suboptimal variability in real-life situations due to the indirect nature of the measurement -Lack of prospectives and large epidemiological studies |
| Nears Infrared Spectroscopy | Muscle and brain microcirculation | Noninvasively assessment of (i) microvascular reactivity, (ii) skeletal muscle and brain oxygenation via continuous monitoring of functional changes in oxygenated hemoglobin dissociation | -Easy repeatability -Precise and reliable -Correlated with other microvascular beds | |
| Adaptive optics | Retinal vascular district | Direct measurement of WLR of retinal arterioles by an adaptive optics imaging system using a beam of light | -Better reproducibility than scanning laser Doppler flowmetry | -Lack of robust evidence on the prognostic value |
| Retinal Vessel Diameter | Retinal vascular district | Direct measurement of arteriolar and venular vessels diameter from fundus photographs | -Safe and noninvasive -Low cost and quick -Applicable in large populations -Implicated in large epidemiological studies predicting CV events -correlated with macro- and other microvascular beds | -Lack of normal values -Lack of studies investigating the changes after drug treatment |
| Coronary arterial network Coronary angiography-derived index of microcirculatory resistance | Coronary arteries | Physiological assessment of microvascular disease in coronary circulation from angiographic images | -May predict adverse CV outcome and extensive myocardial injury -Highly reproducible and excellent diagnostic accuracy, not affected by hemodynamic changes - Relatively independent of epicardial coronary disease | -Invasive method -Limited in clinical practice due to required additional procedural time, cost and technical complexity (in non-STEMI patients) |
| Coronary flow reserve | Coronary arteries | The ratio of the maximal or hyperemic flow down a coronary vessel to the resting flow | -Prognostic value for all-cause mortality and CV events -Quantitative and global physiological interrogation of the coronary circulation -Can be measured using noninvasive modalities, including echocardiography, PET and CMR | -Invasive method using a Doppler-tipped coronary guidewire -Difficulty in obtaining a suitable Doppler signal -Suboptimal repeatability of measurements - Lack of a clear cut-off between normal and abnormal CFR |

AV, arteriovenous; CFR, coronary flow reserve; CMR, cardiac magnetic resonance; CV, cardiovascular; EndoPAT, noninvasive peripheral arterial tonometry; HVM, hand-held vital microscope; MLR, media thickness to internal lumen ratio of subcutaneous small resistance arteries; PET, positron emission tomography; RBCs, red blood cells; SNP, sodium nitroprusside; STEMI, ST-elevation myocardial infarction; WLR, wall–lumen ratio.

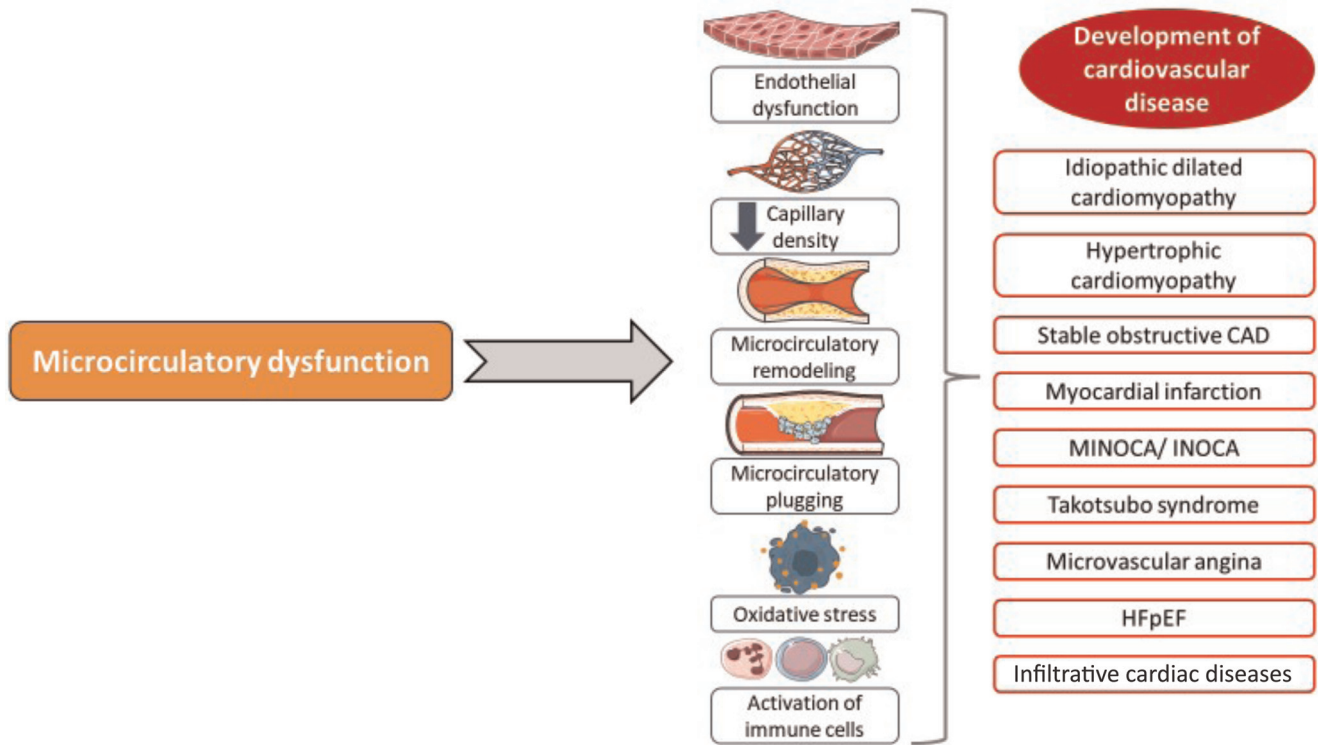


FIGURE 1 Mechanisms linking microcirculatory dysfunction with multiple manifestations of cardiovascular disease. CAD, coronary artery disease; HFpEF, heart failure with preserved ejection fraction; INOCA, ischemia with nonobstructive coronary arteries; MINOCA, myocardial infarction with nonobstructive coronary arteries.

vascular age is a reliable marker of biological age [53]. Microvascular ageing reflects a point at which MD becomes persistent and further deteriorates over time. The onset of MD marks a crucial point in the natural history of ageing.

We need to address lifestyle and environmental stressors to look at the earliest perpetrators of microvascular damage. In addition to the genetic predisposition, each individual is constantly exposed to various noxious *stimuli* that can induce MD. A sedentary lifestyle can increase vascular nicotinamide adenine dinucleotide phosphate (NADPH)-derived reactive oxygen species (ROS) production that affect the endothelial function [54]; similarly, exaggerated exercise training induces mitochondrial dysfunction leading to MD [55]. Unhealthy dietary and eating patterns [56], as well as nutrient overload [57], cause an imbalance in the oxygen supply/demand ratio, activating hypoxia-inducible factor 1 α (HIF-1 α) and promoting impaired angiogenesis [58]. Additional environment determinants, including environment pollutants, temperature, seasonal changes, circadian rhythm and infections have demonstrated significant role in regulating microvascular inflammation and cardiovascular (CV) diseases [59–68].

The final effector through which all these stressors promote microvascular ageing [69–71] is the immune-inflammatory response [60]. Ultimately, they create a low-grade pro-oxidant pro-inflammatory environment [72] that leads to MD [73,74]. Inflammation disrupts microvascular function by increasing ROS generation, reducing NO bioavailability, and leading to vascular wall hyperpermeability and glycocalyx remodelling [75]. In the long term, this promotes the hyperactivation of compensatory pathways such as endothelial and vascular smooth muscle cells

(VSMCs) proliferation, pathological angiogenesis [76–78] and, ultimately, permanent vessel wall remodelling. The lymphatic vasculature also plays a role in this detrimental interplay. While it generally regulates dietary lipid absorption and cholesterol efflux [79], it becomes dysfunctional when exposed to stressors, further compromising local homeostasis. Lymphatic dysfunction results in reduced immune cell clearance, increased insulin resistance [80] and reduced lymphangiogenic potential [79]. These maladaptive changes, which are common in ageing and age-related diseases, ultimately prolong the inflammatory response and microvascular remodelling [81]. The consequence of all these processes is that if the MD is not rapidly targeted and reversed, its alterations become permanent, characterised by epigenetic cues that are not easily targeted by current therapies [82] and predispose to more significant harm when a subsequent exposition to risk factors [83] occurs, even several years ahead. This marks the point when vascular age diverges from chronological age, and ageing diverges from his physiological trajectory.

An aged vasculature is characterised by a low-grade inflammatory state which originates directly from the vessel and the surrounding environment (i.e., perivascular adipose tissue (PVAT) [84], neural terminations [85], abnormal shear stress [86]), even after the removal of the *stimuli*. In this condition, the vessel is not only the target of the damage exerted by the CV risk factors [87] but also becomes the perpetrator by first-hand promoting the low-grade inflammatory response which characterizes chronic time-dependent disease [72]. This, in turn, further dampens microvascular homeostatic control mechanisms and aggravate MD [71]. This vicious cycle is characterised by a

profound cross-talk between nonimmune and immune cells, which is often present in the context of CV disease, for instance, in macrovascular atherosclerotic remodelling [88] and cardiac fibrosis [89]. However, a clear understanding of this dialogue between distinct cell types is still an unmet need, especially in MD setting.

This model accurately reflects cardiometabolic disease. First, environmental stressors significantly influence the natural history of obesity, type 2 diabetes, arterial hypertension, and HFpEF [60]. Second, they all present an early MD [38,90–92] and are characterised by accelerated microvascular ageing [58]. Third, these conditions are tightly connected [52]. Fourth, all of them are characterised by a persistent systemic low-grade inflammation which further deteriorates the cardiometabolic homeostasis and that it has in the microvascular bed one of its primary perpetrators [72,84,93].

Inflammageing [72,94] and immunometabolism [95] are thus fundamental integrated approaches to explore the connections and the cross-talks between environment, metabolic disease, vascular health, and CV risk. Although

this conceptual framework is generally related to cardio-metabolic disease, it might be easily translated to other chronic and time-dependent conditions such as neurodegenerative pathologies, autoimmune diseases, and cancer. As indirect evidence, epigenetic pan-deactivators of vascular inflammation as the inhibitors of bromodomain and extraterminal domain (BET) proteins [96] have recently been proposed for all these disease settings [97–103]. Similarly, anti-inflammatory drugs are attracting substantial attention in the context of CV diseases [104]. But the link between inflammation and microvascular ageing is multi-directional. As environmental stressors link inflammation with microvascular ageing, inflammation also becomes the link between aged vasculature and systemic metabolic diseases, which further promotes microvascular inflammation. The onset of this vicious cycle is at the base of age-related diseases. It is clear that only by an accurate understanding of the pathophysiologic and molecular mechanisms underpinning this connection we will be able to develop therapeutic strategies to challenge the steadily increasing prevalence of chronic diseases [105,106] (Fig. 2).

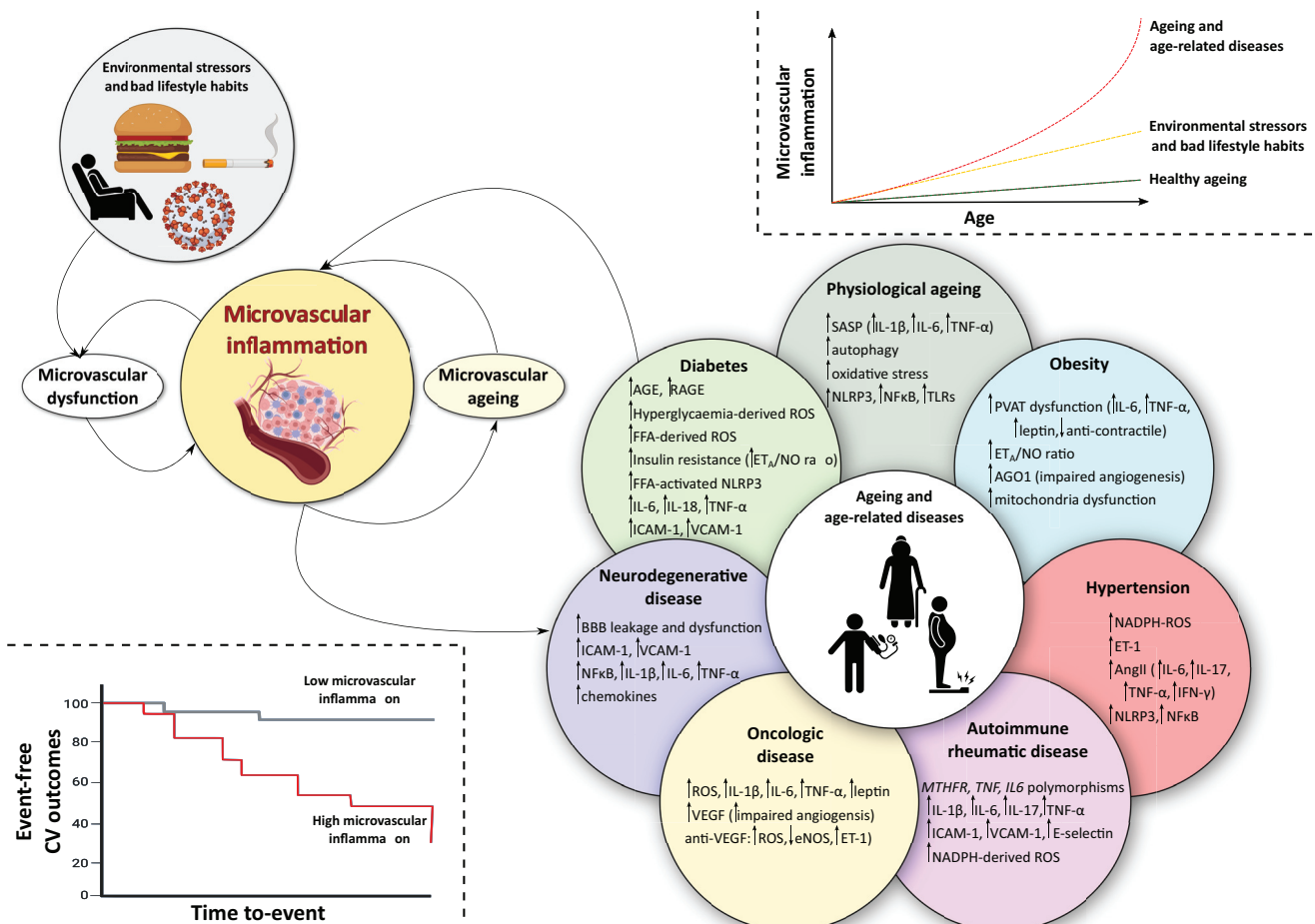


FIGURE 2 Graphical abstract. Microvascular inflammation links environmental stressors to microvascular ageing. Environmental stressors induce microvascular dysfunction, which in turn promotes microvascular inflammation. When microvascular inflammation causes permanent changes in vascular structure and function, microvascular age and biological age diverge. The vessel becomes the architect of microvascular inflammation, exposing the microcirculation to further damage from environmental stressors and thus promoting the onset of cardiometabolic disease, exponentially increasing the degree of microvascular inflammation and the individual cardiovascular risk. The major common and distinct molecular-, cell- and tissue-level mechanisms involved in microvascular inflammation are summarized. AGE, advanced glycation end products; AGO1, argonaute 1; BBB, blood–brain barrier; eNOS, endothelial nitric oxide synthase; ET-1, endothelin-1; ET_A, endothelin-1 receptor A; FFA, free fatty acids; ICAM, intercellular adhesion molecule-1; IFN, interferon-γ; IL, interleukin; MTHFR, methylene-tetrahydrofolate reductase; NADPH, nicotinamide adenine dinucleotide phosphate; NF-κB, nuclear factor kappa-B; NLRP3, NOD-like receptor family pyrin domain containing 3; NO, nitric oxide; PVAT, perivascular adipose tissue; ROS, reactive oxygen species; SASP, senescent-associated secretory phenotype; TLR, toll-like receptors; TNF-α, tumor necrosis factor; VCAM-1, vascular cell adhesion molecule-1; VEGF, vascular epithelial growth factor.

Downloaded from http://journals.lww.com/jhypertension by BHD/MSep/HKav17Eoum1tQIN4a+kLHEZbsIho4XN10 HCW/CX1AW/vYqP/IIQHTID3I3D00dFy71vSFACI3VC1y0abgQZXdG12mWZLeI= on 02/23/2024

Statement: Environmental stressors predispose to unhealthy ageing and age-related disease by promoting MD. If MD is not prevented or rapidly treated, the microvascular environment in turn becomes the perpetrator of microvascular inflammation through detrimental cross-talk between nonimmune and immune elements, leading to the low-grade inflammatory response that characterises unhealthy ageing and age-related disease.

MICROVASCULAR INFLAMMATION ACROSS THE AGE-RELATED DISEASES CONTINUUM

Physiological ageing

Physiological ageing is a natural phenomenon driven by a variety of complex, and yet loosely understood mechanisms that strongly interact with each other. In addition to these, initial emergence of genomic instability, which includes dysregulated DNA damage repair pathways and telomere shortening, other key mechanisms are involved. These include the stimulation of senescence and impairment of autophagy at a cellular level and the consequential development of oxidative stress and microvascular inflammation at the tissue level. Ultimately, these mechanisms contribute to the ageing-related phenotype characterized by endothelial, vascular and consecutively tissue dysfunction (Fig. 3a) [107,108]. Notably, these mechanisms are often bidirectional, ultimately establishing a vicious cycle.

Senescence

Cellular senescence is a stress-induced, durable, cell cycle arrest of previously replication-competent cells and is considered a central hallmark of ageing [109]. Senescence contributes to ageing process through multiple mechanisms, among which the propagation of inflammation prevails. Pertinent to this, it has been shown that senescent cells secrete a plethora of potent pro-inflammatory factors termed the senescent-associated secretory phenotype (SASP), which drive an intense inflammatory response [110,111]. SASP also contributes to the spread of inflammation and oxidative stress from senescent to healthy nonsenescent cells via paracrine fashion, which leads to a pro-inflammatory and pro-oxidant phenotype at a microvascular level [107]. Consistent with its inflammatory potential, several *in vitro* and preclinical data have documented the role of senescence in promoting oxidative stress and endothelial dysfunction [112,113]. In healthy ageing humans, markers of endothelial senescence have been correlated with significantly impaired endothelial function [114].

Autophagy

Autophagy is a highly selective clearance pathway that degrades several defective cellular components through lysosomal activation. Therefore it is tightly associated with the maintenance of cellular and tissue homeostasis and, in the long term, longevity [115]. Altered autophagy has been proposed as a prominent feature of physiological ageing, with increasing evidence suggesting an impaired

autophagic activity across ageing in different organisms [116]. In humans, it has been demonstrated that the expression of autophagy-related genes (i.e., ATG5, BECN1) and the proteolytic function of lysosomes decline with age [117]. Consequently, compromised autophagy leads to cellular and vascular dysfunction and enhanced inflammation, as evidenced by the promotion of oxidative-induced senescence, the production of endothelial reactive oxygen species (ROS) and the development of endothelial dysfunction in both aged mice and human subjects [117–119]. In addition, autophagy has been recognized as a significant inhibitor of inflammasome which is a potent mediator of microvascular inflammation [120].

Oxidative stress

Oxidative stress is a consequence of the imbalance between the production and detoxification of reactive oxygen and nitrogen species (RONS) [121]. Ageing process is associated with reduced activity of the antioxidant transcription factor nuclear factor erythroid 2-related factor 2 (Nrf2) [122]. Hence, accumulation of oxidative damage by RONS produced by NADPH oxidases and mitochondria is considered one of the core pathophysiologic pathways driving physiological ageing and age-associated diseases [123]. Particularly in the context of ageing and age-related diseases, mitochondrial dysfunction and mitochondria-derived ROS are key drivers of the inflammatory response leading to pathogenetic processes [124,125]. The oxidative environment further stresses mitochondrial pathways, leading to the detrimental escape of mitochondrial DNA from organelles and cells [126,127]. Mitochondrial-free mitochondrial DNA and the pro-oxidant environment, in turn, transduce a pro-inflammatory signal within and between cells, leading to the activation of multiple signalling pathways: nucleotide-binding oligomerization domain (NOD)-like receptor family pyrin domain containing 3 (NLRP3) inflammasome activation, DNA-sensing enzyme cyclic GMP-AMP synthase stimulator of interferon genes (cGAS-STING) and toll-like receptor (TLR). It also leads to induction of senescence and SASP production with consequent nuclear factor kappa-B (NF- κ B) activation, as well as hyperactivation of the pro-oxidant mediator p66Shc [82,125], consumption of NAD⁺ and consequent mammalian silent information regulator 1 (Sirt1) dysregulation [52,128]. Furthermore, oxidative stress exerts a detrimental effect on endothelial function by quenching the bioavailable, endothelium-derived NO and reducing both NO availability and endothelial NO synthase (eNOS) expression. Therefore, oxidative stress is strongly linked to the development of endothelial and microvascular dysfunction with ageing in humans [129,130].

Inflammation

The inflammaging state, a sterile, subclinical, low-grade inflammation increasing with age and promoting the development of age-associated diseases, has been well recognized in the elderly [72]. Indeed, in older adults are frequently reported persistently elevated circulating levels of SASP factors, including IL-1 β , IL-6 and tumour necrosis factor (TNF)- α [131–133]. In elderly, inflammaging is largely considered an aftermath of immunosenescence, a significant immune system dysregulation observed with

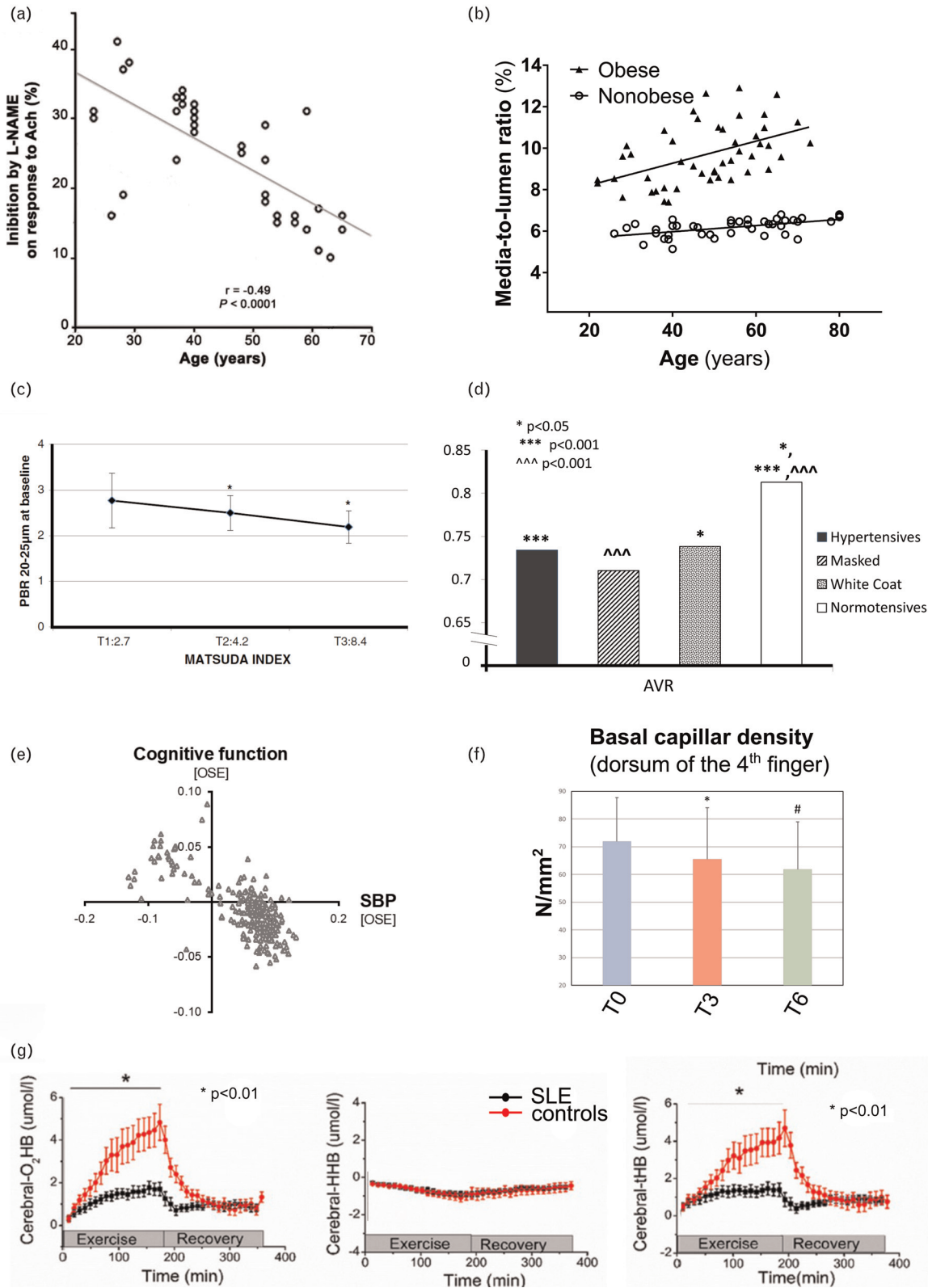


FIGURE 3 Microvascular inflammation and its impact on microvascular dysfunction across ageing and age-related disease. (a) Relationship between age (x-axis) and the inhibition by *N*-nitro-*L*-arginine methylester (L-NAME) on maximal response to acetylcholine (y-axis) in normotensive subjects ($n=41$). Adapted from [108]. (b) Increased media-to-lumen (M/L) ratio per year of age in healthy nonobese (white circle; $n=42$) and obese with no other comorbidities (black triangle; $n=47$) subjects. Regression lines for each group are shown. The M/L ratio is expressed as a percentage (%). Age and M/L are tightly related in both groups (obese: $r=0.487$, $P<0.01$; nonobese: $r=0.555$, $P=0.001$). The slope is five-fold steeper in the obese group. Figure and captions adapted from [125]. (c) Association of tertiles (T1–T2–T3) of Matsuda index with perfused boundary region (PBR) measured in the microvessels ranged from 20 to 25 μm indicating an association between insulin resistance and damaged glycocalyx ($F=4.8$, $P=0.03$) in $n=100$ subjects with different degrees of insulin resistance ($n=40$ first-degree relatives of type-2 diabetes patients, $n=40$ subjects with abnormal oral glucose tolerance test and 20 subjects with normal oral glucose tolerance test without parental history of diabetes). Adapted from [182]. (d) Comparison of the retinal

Downloaded from <http://journals.lww.com/jhypertension> by BHD/MS/EP/HK/AV/17/Eoum/1tQ/N4a+k/LhEZgbsIh04XN10/HCYwCX1AWnYQp/IIQrHID3I3D00dFy71vSFACI3VC1y0abgqQZXdgG12mWZLeI= on 02/23/2024

ageing, which substantially propagates the inflammatory milieu and consists in overall aberrant activation of innate and adaptive immune response [134]. In this context, microvascular inflammation can be exacerbated by an age-associated inappropriate activation of TLRs and the NLRP3 inflammasome complex, both representing crucial activators of the innate immune inflammatory response which leads to increased expression of NF- κ B and the production of several proinflammatory mediators [135–137]. Activation of the ROS-sensitive, proinflammatory effector NF- κ B holds a central role in the ageing-associated inflammatory response. Endothelial cells (ECs) from older humans actively express NF- κ B, which is directly implicated in endothelial dysfunction [132,138] as well as in exacerbating inflammageing and oxidative stress, thus corroborating an intricate relationship between senescence, oxidative stress and inflammation across ageing and extending the vicious cycle [139–141]. Finally, mammalian sirtuins represent another significant ageing-associated mechanism implicated in microvascular inflammation. They are a family of nicotinamide adenine dinucleotide-dependent deacetylases involved in several processes that regulate metabolic homeostasis and modulate the benefits of calorie restriction and exercise. They control mitochondrial function, cell survival, attenuation of inflammatory responses and circadian rhythm. Because of their contribution to many protective pathways and their central involvement in longevity mechanisms, they have attracted increasing attention as potential therapeutic targets [142]. In particular, Sirt1, a deacetylase implicated in many critical physiological responses to altered energy metabolism and stress, has multiple anti-inflammatory, antioxidant and anti-ageing properties [143–145]. Reduced expression of Sirt1 has been observed in ECs and VSMCs obtained from older adults, associated with a senescent phenotype and the development of endothelial dysfunction [125,146,147]. Furthermore, data has shown that persistently reduced levels of Sirt1 lead to upregulation of NF- κ B and NLRP3 inflammasome, hence significantly amplifying the inflammatory response [145,148].

Statement: In physiological ageing, stimulation of senescence, impairment of autophagy, and increased oxidative stress lead to microvascular inflammation at the tissue level. This culminates in the ageing-related phenotype characterized by MD and increased susceptibility to the onset of age-related diseases.

Obesity

Obesity, given its high and steadily increasing prevalence [105], probably represents the closest human model to

exploring the contribution of environmental stressors to microvascular inflammation and accelerated ageing. Its relevance in the global landscape is sadly acknowledged: obesity ranks first in terms of mortality related to metabolic diseases, with no trend towards reduction [149]. The impact of obesity on MD starts very early: in patients with obesity, the slope of the media-to-lumen (M/L) ratio plotted against age diverges from the healthy controls before the age of 20, being five times steeper (Fig. 3b) [125,150]. The increased nutrient supply ultimately overburdens the metabolic pathways, promoting hypoxia with consequent HIF-1 α activation [151]. The resultant pro-oxidant and pro-inflammatory environment promotes the local low-grade inflammatory response, which eventually turns systemic and characterises the generalized MD observed in obesity [91].

Although multiple pathways contribute to its clinical phenotype, obesity is characterised by a prominent role of the PVAT. PVAT is a key member of the microvascular unit [91], with a brown-like and anticontractile phenotype in the healthy [152], which loses its thermogenic capacity and turns pro-contractile in the condition of diseases such as obesity [153]. The deep cross-talk between PVAT and the small vessels is directly responsible for both the inflammatory damage and response characterising obesity [154]. PVAT phenotype shift leads to increased secretion of several adipokines and cytokines, including chemerin, leptin, IL-6 and TNF- α [155]. While experimental studies in mice have reported how leptin leads to MD by first targeting the hypothalamic microvasculature [156], *ex-vivo* observations in humans have demonstrated how PVAT dysfunction, promoted by macrophage activation [157], results in an increase in PVAT-derived cytokines secretion [84,93]. This fuels the vessel-specific inflammatory response [93], induces endothelial dysfunction and further imbalance the homeostatic response from the vasculature by increasing the expression of endothelin-1 (ET-1) and its receptor A (ET_A). The altered ET_A/NO ratio upregulates c-Jun N-terminal kinase (JNK) signalling, increasing NADPH-derived and mitochondria-derived ROS [84]. The bidirectional cross-talk further aggravates PVAT dysfunction, as the endothelium also secretes inflammatory cytokines and angiogenic factors. Indeed, to match the increased nutrient flow, PVAT develops a pro-angiogenic phenotype trying to compensate with an adequate oxygen supply. However, this neo-angiogenic process [76] ultimately proves detrimental and further promotes an overt dysfunctional phenotype for both the PVAT and the ECs. Recently, an elegant exploration in vivo has shown that, in high-fat diet mice, the ECs-specific deletion of argonaute 1 (AGO1), a pivotal

Figure 3 (Continued).

arteriovenous ratio (AVR) in $n=201$ newly diagnosed individuals with hypertension of different phenotypes and normotension. Intergroup comparisons were made with analysis of variance ANOVA with Bonferroni correction after adjustment for age, sex, body mass index (BMI). Individuals with sustained hypertension ($n=103$), masked hypertension (MHT; $n=28$) and white coat hypertension (WCH; $n=20$) had significantly lower AVR than normotensive subjects ($n=50$; $P<0.05$). Adapted from [206]. (e) Observational, standardised coefficients concerning 242 brain imaging-derived phenotypes genetically affected by SBP corresponding to their association with cognitive function or SBP at the imaging visit. Hypertension was used as a model associated with microvascular inflammation. Figure and caption adapted from [257]. (f) Altered capillaroscopy in oncologic disease. Basal capillary density in the dorsum of the fourth finger (Dpre_basal) in patients ($n=20$) with cancer and treated with either a tyrosine kinase inhibitor or a vascular epithelial growth factor inhibitor at the different time points (T0, T3, T6). * T3 vs. T0 $P=0.03$; # T6 vs. T0 $P=0.02$. Data are expressed as mean±standard deviation. Adapted from [338]. (g) Accumulative data of near-infrared-spectroscopy cerebral responses during exercise in systemic lupus erythematosus (SLE; $n=26$) vs. control ($n=27$) group. Oxygenated haemoglobin (O₂Hb), deoxygenated haemoglobin (HHb) and total haemoglobin (tHb) levels were measured. Cerebral O₂Hb continuously increased during exercise in the control group, whereas the SLE group exhibited a plateau in O₂Hb after the first minute of exercise ($P<0.01$). During exercise, the SLE group exhibited significantly lower average-O₂Hb (1.20 ± 0.89 vs. 2.69 ± 2.46 , $P=0.001$), and a lower peak-O₂Hb response (2.89 ± 1.56 vs. 5.83 ± 4.59 , $P=0.004$) compared with the control group. No differences were detected in the average HHb responses between groups. Adapted from [299].

contributor to the ECs response to hypoxia, arrests impaired angiogenesis and reverts the PVAT to a browning phenotype, rescuing the MD and the whole-body metabolic homeostasis [158].

Finally, the overload of the metabolic processes leads to mitochondrial dysfunction in obesity, which in turn promotes microvascular inflammatory response by increasing mitochondria-derived ROS levels [125] and activating NLRP3 [159] and cGAS-STING pathways [160]. Evidence supports sirtuins [125] (main elements in nutrient balance/imbalance signalling [161]) as crucial regulators of this process. In ECs, lower levels of Sirt1 induce MD and are associated with an increase of pro-inflammatory and pro-ageing factors p66shc [82] and Arginase II [150], an increase in mitochondria-derived ROS, and a downregulation of several genes involved in the mitochondria electron transport chain.

The documented MD confirms this experimental evidence in patients with obesity. An increased vascular remodelling in visceral fat arteries [125], an impairment in finger microcirculation detected by dynamic nailfold microcapillaroscopy [162], a thin sublingual microvasculature glycocalyx assessed by sidestream darkfield imaging [163], an increased retinal arteriolar narrowing [164] and a decreased retinal microvasculature response to flicker light [165] all characterise the microvascular damage found in patients with obesity. Remarkably, bariatric surgery, the gold standard treatment for treating severe obesity, showed a remarkable effect in terms of MD rescuing in patients with severe obesity, as shown by an improvement in skin microcirculation [166] and in subcutaneous arteries reactivity, which appears even more robust when including PVAT [167].

Statement: Obesity-related microvascular inflammation is characterised by accelerated ageing starting from adolescence/early adulthood, defined by a derangement in the PVAT phenotype, a hyperactivation of inflammatory pathways (mainly TNF- α and IL-6), an impaired angiogenesis and an early mitochondrial dysfunction.

Diabetes

It is well recognized that low-grade inflammation plays an essential role in the pathogenesis of DM, as well as in the development of diabetic microvascular complications. Studies have demonstrated that inflammatory mediators, such as CRP, TNF- α , IL-6 and IL-18, have elevated expression in DM [168,169]. Hyperglycaemia acutely increases circulating cytokine levels through an oxidative mechanism among subjects both with features of insulin resistance and with clinically overt DM [170,171]. In subjects with type 2 DM, a correlation was observed between high-sensitive CRP and IL-6 with HbA1c independent of the presence of coronary heart disease [172,173]. Furthermore, serum levels of TNF- α were associated with the level of insulin resistance and with HbA1c in diabetic subjects [174].

Diabetic hyperglycaemia increases oxidative stress by excessive intracellular ROS generation, which in turn leads to activation of the NF- κ B pathway resulting in the production of major pro-inflammatory cytokines. Hyperglycaemia-induced oxidative stress increases the formation of advanced

glycation end products (AGEs), which results in increased insulin resistance. Moreover, receptors for advanced glycation end products (RAGE) are involved directly in inflammatory cell recruitment [175,176]. Insulin resistance is associated with endothelial dysfunction. In particular, the endothelial balance between NO-mediated vasodilator actions and ET-1-mediated vasoconstrictor effects of insulin are regulated via phosphatidylinositol 3-kinase (PI3K) and mitogen-activated protein kinase (MAPK) pathways, respectively. In states of insulin resistance, dysregulation of PI3K-dependent signalling may cause an imbalance between the NO production and secretion of ET-1 [177]. Thus, insulin resistance induces vasoconstriction and VSMCs proliferation and plays a significant role in the occurrence of endothelial dysfunction [178]. Indeed, markers of insulin resistance are associated with abnormal arterial elastic properties and impaired coronary microvascular function not only in dysglycaemic subjects but also in first-degree relatives of diabetic subjects before the development of impaired glucose tolerance or DM [179].

Furthermore, oxidative stress is characterized by the production of peroxynitrite that down-regulates NO bioavailability and leads to vasoconstriction. Also, the accumulation of ROS promotes the apoptosis of ECs and augment the expression of intercellular adhesion molecule-1 (ICAM-1), vascular cell adhesion molecule-1 (VCAM-1), and E-selectin resulting in microvascular inflammation and hypercoagulability [180]. Chronic inflammation inhibits the production of endothelial eNOS and promotes the expression of ICAM-1, VCAM-1 and ET-1, further damaging endothelial integrity [181].

Acute and long-term hyperglycaemia have a detrimental effect on endothelial glycocalyx integrity [182–184]. The glycocalyx is a gel-like layer composed of sulphated proteoglycans and glycoproteins that prevents the direct contact of circulating inflammatory cells to the luminal surface of the endothelium [185]. Intriguingly, HbA1c is associated with the impaired perfused boundary region, a marker of the microvascular glycocalyx thickness, while intensified glycaemic control ameliorates glycocalyx integrity in diabetic subjects at the 1-year follow-up [186]. The impaired glycocalyx is an independent predictor of adverse outcomes in subjects without established CV disease [43].

Besides hyperglycaemia, high free fatty acid levels (FFA) may stimulate ROS production via protein kinase C (PKC)-dependent activation of NADPH oxidase in both VSMCs and ECs. This finding may explain the excessive acceleration of atherosclerosis and microcirculation damage in diabetic subjects [187]. In addition, hyperglycaemia and lipotoxicity lead to hyperactivation of NLRP3 inflammasome, which mediates caspase-1 activation and the secretion of pro-inflammatory cytokines IL-1 β and IL-18. Thus, NLRP3 inflammasome activation in DM leads to chronic inflammation and increased vascular permeability [188].

The most common microvascular complication of DM is diabetic retinopathy. Ocular microcirculatory damage on the grounds of hyperglycaemia causes capillary occlusion leading to retinal ischemia and neovascularization [189]. Interestingly, experimental data show that microcirculatory changes, including adherence of neutrophils and leukostasis, in nonocular tissues of diabetic mice appear to be related and reflect retinal microvascular lesions in the

context of diabetic retinopathy [190]. Underlying retinal microvascular dysfunction seems to precede the clinical manifestation of DM-associated CV disease [191]. New advances in retinal vessel analysis provide useful diagnostic tools to improve the prediction and risk stratification of CV disease [192]. However, there is also evidence of nonretinal MD in diabetes: diabetic subjects have impaired dermal microvascular hyperaemia response to local skin heating [193], reduced glycocalyx thickness (Fig. 3C) [194]. Similarly, digital pulse amplitude tonometry (PAT) is impaired in type 2 diabetes and influenced by glucose level fluctuations [195].

Finally, it should be mentioned that several of the novel antidiabetic agents, namely dipeptidyl peptidase-4 inhibitors (DPP-4i), glucagon-like peptide-1 receptor agonists (GLP-1RA) and sodium-glucose cotransporter-2 inhibitors (SGLT-2i), have potential anti-inflammatory properties and improve endothelial function and presumably microcirculation [194,196–200], and this might be – at least partially – responsible of the significant benefit observed in terms of reduction of CV risk [201].

Statement: Hyperglycaemia and FFA-derived ROS produce a substantial increase in oxidative stress, directly affecting microvascular inflammation and promoting an AGE/RAGE and lipotoxic environment, which determines peripheral insulin resistance, further worsening the low-grade inflammatory response.

Arterial hypertension

Arterial hypertension is characterised by diffuse microvascular damage (Fig. 3d, Figure 1, Supplemental Digital Content, <http://links.lww.com/HJH/C223>) [202–206]. Hypertension and inflammation have a bidirectional physiological and pathophysiological background [207,208]. Several human studies have evaluated the relationship between inflammation and essential hypertension. In a meta-analysis ($n = 21\,458$ patients), higher levels of circulating CRP, high-sensitivity CRP (hs-CRP), and IL-6, but not IL-1 β , were associated with the risk of developing hypertension [209]. Studies have also reported correlative links between arterial stiffness and inflammatory markers in essential hypertension [210].

A hallmark of inflammation is the release of inflammatory cytokines such as IL-6, IL-17A, interferon- γ (IFN- γ), and TNF- α by T CD4⁺ cells, and more specifically by subsets of T helper (Th) cells, Th1 and Th17 [9,207,208]. Involvement of IL-6 has been shown in mice, where *Il6* knock-out mice showed reduced hypertension severity in response to angiotensin II (AngII) infusion [211]. Further, in human renal proximal tubular cells, IL-6 increased angiotensinogen expression [212]. IL-17A affects renal sodium handling [213] and inhibits eNOS, causing impaired vasodilatation and increased peripheral vascular resistance [214]. Long-term effects of IL-17A include the promotion of vascular fibrosis, leading to arterial stiffening [214]. In addition, in mice lacking IL-17A, the number of T cells and macrophages in blood vessels was reduced, illustrating the effect many of these cytokines have in attracting more immune cells, further amplifying the immune response

[207]. They induce oxidative stress [215] through enhanced NADPH oxidase subunit expression. Increased oxidative stress affects sodium retention by decreasing the glomerular filtration rate [216]. This, as many of the classical pathophysiological factors in hypertension (ET-1, aldosterone, and AngII), activates the NLRP3 inflammasome through NF- κ B. NLRP3 activation leads to increased levels of proinflammatory cytokines IL-1 β and IL-18, activating immune and vascular cells as T cells (mainly CD4⁺), monocytes, ECs and VSMCs [217].

The contribution of innate and adaptive immune cells to the development of MD leading to hypertension is substantial [218]. Regarding immune cells, in animal models of genetic hypertension, vascular ageing is associated with increased PVAT infiltration of macrophages, neutrophils and natural killer cells (NKs), which promote NADPH oxidase 4-driven microvascular remodelling [9,219,220]. Macrophages, as a major source of ROS, are considered important in this process, although the precise mechanisms by which they are involved remain unclear [9]. Similarly, NKs are found to increase before the development of hypertension in spontaneously hypertensive rats [219]. Neutrophils from the plasma of untreated patients with essential hypertension generate neutrophil extracellular traps that lead to collagen production and consequent microvascular remodelling [221]. In adaptive immunity, T cells are considered to play a predominant role. Following antigen recognition, CD4⁺ T cells are activated and differentiate into T effectors (Th1, Th2 and Th17) or T regulatory cells (T_{reg}), the balance of which influences the inflammatory response [9]. In experimental models of hypertension, as well as in hypertensive patients, the inflammatory response generated by the ratio of T helper lymphocytes (Th) 1/17 (Th1/Th17) is not adequately balanced by the pool of regulatory T lymphocytes (Treg), thus contributing to structural damage of the microcirculation [9]. Recently, it was discovered that the T cell mir214 partially recapitulates and transduces the fibrotic effects of the immune system to the microvasculature, leading to vascular fibrosis, vascular stiffening and remodelling. In particular, cytokines released from PVAT mediate these effects [220]. It is thus clear that the immune system is one of the leading mechanisms supporting the cross-talk between vascular inflammation and hypertension. However, it should be noted that most of the evidence comes from in vivo studies, as further investigations in patients are needed [9].

However, over the last years, evidence has been accumulated showing that this dialogue also involves the sympathetic nervous system [222,223]: an increase in sympathetic activity elicits T-lymphocytes activation and vascular inflammation [224]; significant correlations have been found between circulating plasma norepinephrine, and IL-6 produced by T-lymphocytes as well as TNF- α produced by macrophages and monocytes [225]; chronic sympathetic activation in patients with a peculiar form of high blood pressure desensitizes lymphocyte β_2 -adrenoceptors and thereby alters immune function [225]. On the other hand, inflammation and T-lymphocytes activation, which are both triggered by oxidative stress [226], may favour sympathetic activation, as already shown in other diseases characterized by an adrenergic overdrive, including essential hypertension

[222]. In any case, pro-inflammatory substances and mediators may trigger signals to the central nervous system activating the sympathetic neural component [222].

Statement: arterial hypertension is characterised by increased sodium retention and higher levels of ET-1, aldosterone and AngII, which disrupt the microvascular environment by promoting NAPDH-derived endothelial dysfunction and IL-6, IL-17 and TNF- α -driven inflammatory responses. The innate and adaptive immune systems play a central role. In particular, a balance between effectors (Th1, Th17) and regulators (Treg) T cells orchestrates microvascular inflammation and consequent microvascular remodelling.

Neurodegenerative diseases

The cerebral vasculature is unique in its anatomy and physiology. It constructs a highly specialized blood–brain barrier (BBB) that controls the admission of solvents and ions into the brain and clearance into the blood metabolic end products or endogenous neurotoxin produced by the brain [227,228]. The BBB, therefore, play a critical role in maintaining brain homeostasis. It comprises endothelial cells, basement membrane, pericytes and VSMCs, astrocytes, microglia and neurons [229,230]. Vascular ECs are known to secrete vasoactive substances implicated in regulating cerebral flow, intravascular blood coagulation, and preserving the integrity of the BBB. Their atheroprotective role and homeostasis are controlled by releasing vasoactive factors, especially NO. This leads to cGMP-mediated cerebral vessel relaxation and proper blood supply to the brain tissue and autophagy [231]. Pericytes (PCs) directly encircle endothelial cells and VSMCs and are considered vascular mural cells. ECs and PCs have direct contact through gap junctions and contribute to blood vessel formation and BBB function maintenance by regulating immune cells' entry (CD4⁺ and CD8⁺ T cells, peripheral macrophages and neutrophils) to the central nervous system [232–234].

Similar to the peripheral circulation, impairment in NO production, inflammation and enhanced ROS production [235,236] are vital in promoting ECs dysfunction manifested by increased expression of leukocyte adhesion molecules such as ICAM-1, VCAM-1 and E-selectin. These molecules promote higher immune-endothelial cell interaction and accumulation of inflammatory cells in the vascular and perivascular niches. The higher expression of ICAM-1 and VCAM-1 is observed in cerebral endothelial cells in animal models of cerebral hypoperfusion, while their inhibition protects against cognitive impairment [237,238]. Furthermore, soluble adhesion molecules like sE-selectin, sP-selectin, sICAM-1 and sVCAM-1, considered endothelial dysfunction markers, are elevated in patients with small vessel brain diseases [239].

The chemotactic process is strictly controlled by numerous chemokines secreted by the vascular cells [240], pericytes [233], microglia [241], and astrocytes [242] in a concentration-directed gradient. Recent studies have implicated the importance of CCL2, CCL3, CCL5 and CXCL8 in many vascular and neurodegenerative diseases, including cognitive impairment, stroke and neuroinflammation [243].

Microcerebrovascular endothelial cell activation and BBB leakage promote the migration and accumulation of proinflammatory macrophages [244] and T cells [245,246] in perivascular space (Figure 2, Supplemental Digital Content, <http://links.lww.com/HJH/C223>). The role of various immune cells in the pathogenesis of endothelial dysfunction and vascular inflammation in CV diseases has been well established [220,247]. Activated immune cells release diverse pro-inflammatory mediators, which propagate microvascular inflammation and may provoke microhemorrhages, further escalating the inflammatory process. Co-involvement of IL-1 β , IL-6 and TNF- α , in microvascular brain injury and inflammation has been widely reported [248,249]. IL-1 β is considered the main proinflammatory cytokine that increases the astrocytic production of CCL2, CCL20 and CXCL2 [250]. In addition, IL-1 β impairs microvascular ECs by disturbing tight and adherent junctional proteins and increasing adhesion molecules expression, prompting vascular leakage and the parenchymal infiltration of leukocytes. In contrast, anti-IL-1 β treatment blunts cerebrovascular inflammation and improve outcome in a mouse model of acute ischaemic stroke [251]. IL-6 is a pleiotropic inflammatory cytokine produced by infiltrating leukocytes, ECs, activated microglia and astrocytes. Its expression affects many neuro-inflammatory and neurodegenerative conditions [252–254]. IL-6 mediates the elevation of superoxide production and endothelial impairment by affecting NO–cGMP signalling pathway [255]. In addition, it may enhance CRP released by brain cells [256]. Similarly to IL-6, TNF- α affects proper endothelial function by decreasing eNOS levels by destabilising its mRNA expression. Furthermore, TNF- α activates NF- κ B, a major regulatory transcription factor, playing a pivotal role in regulating various inflammation-related genes, including key inflammatory cytokines (along with IL-1 β and IL-6), chemokines and adhesion molecules.

Cognitive impairment is a hallmark of numerous CV diseases [257]. In hypertension, white matter hyperintensities (WMH) are a critical imaging biomarker linked to this process (Fig. 3E). Indeed, neurovascular inflammation is involved in the aetiology of WMH [258]. Similarly, cerebral small vessel disease has been identified as a key hallmark of a broad range of neurodegenerative conditions. Human neuroimaging and genetic studies show that it is characterised by microvascular endothelial dysfunction impacting cell-cell interactions and leading to brain damage [259]. One broadly studied model of cerebral small vessel disease caused by NOTCH3 mutations, CADASIL (cerebral autosomal dominant arteriopathy with subcortical infarcts and leukoencephalopathy) [260], is characterised by accelerated cognitive decline and dementia, recurrent stroke without vascular risk factors, and mood disturbances. This hereditary disorder provides a unique opportunity to understand some of the molecular mechanisms of small vessel disease [261].

Elevation of proinflammatory mediators such as IL-1 β , IL-6, TNF α and CRP has been linked with cognitive impairment in humans [262,263]. Furthermore, proinflammatory cytokines secreted by immune and vascular cells have direct neurotoxic and apoptotic properties [264], which could perpetuate local neuroinflammation and neurodegradation. Interestingly, neuroinflammatory changes found in the

brains of coronavirus disease 2019 (COVID-19) patients were accompanied by the presence of macrophages and T cells in the perivascular space [265], suggesting higher microvascular inflammation caused by a cytokine storm which might affect the proper BBB function [266]. Furthermore, cerebral microvascular inflammation enhances the prevalence of cognitive impairment even in mild symptomatic COVID-19 subjects [267]. Similarly, experimental and epidemiological studies indicate a relationship between cognitive decline and CV diseases [228,268–270], associated with chronic low-grade inflammation and dysregulation of the immune system [271,272]. In particular AngII is at the crossroad, acting as cardiovascular and immune systems modulator, initiating inflammation by indirect promotion of vascular permeability and the recruitment of peripheral macrophages and CD4⁺ and CD8⁺ T cells [273,274]. In turn, augmented permeability leads to further inflammation and secondary damage to the BBB, with the entry of plasma proteins and neurotoxic substances [275].

The most prevalent form of dementia, AD, is marked by a steady decline in cognitive function and neurodegeneration. The vascular hypothesis suggests that cerebral microvascular alterations are central to the pathogenesis of AD, providing a link with CV disease [276]. Possible mechanisms include neurovascular coupling imbalances and BBB disruption [276]. Impaired removal of beta-amyloid may be a consequence of these neurovascular changes: vascular changes may precede the development of tau pathology [277]. A two-hit hypothesis has been developed in which classical risk factors leading to the development of microvascular dysfunction facilitate AD-specific pathology. This is linked with the development of a vicious cycle between microvascular damage and beta-amyloid aggregates that contribute to AD development. Brain imaging supports these observations, as well as chronic cerebral hypoperfusion, microvascular dysfunction, and perivascular space enlargement – hallmarks of small vessel disease – precede cognitive decline and changes in conventional biomarkers [276]. These mechanisms of small vessel disease are shared between vascular dementia and AD: assessment of retinal microvasculature has shown apparent microvascular dysfunction and remodelling in neurodegenerative diseases [278,279].

Statement: In neurodegenerative disease, MD favours the increased BBB dysfunction and leakage paired with higher NF- κ B activation and consequent microvascular levels of IL-1 β , IL-6, TNF- α , which promotes the onset of a vicious cycle leading to progressive cognitive impairment and increased predisposition to tau pathology.

Autoimmune rheumatic diseases

Autoimmune rheumatic diseases (ARD) are distinct heterogeneous disorders with common immune responses against self-antigens arising from genetic predisposition, dysregulation of the immune system and environmental factors. Among them, chronic inflammatory rheumatic conditions, mainly represented by rheumatoid arthritis (RA), systemic lupus erythematosus (SLE) and spondyloarthritis (ankylosing spondylitis (AS) and psoriatic arthritis (PA)) are

those further characterized by increased and premature CV morbidity and mortality [280]. Atheromatosis is a chronic inflammatory process in which the immune system, blood and vascular cells, and several hormonal systems are primarily involved in the structural and functional damage of the small vessels [281]. Microangiopathy has been used as an important subclinical CV risk indicator, and ARD patients [282,283] have an increased prevalence of CV diseases which cannot be fully explained by the classical CV risk factors [284].

In ARD, a combination of elements is able to contribute specifically to MD and microvascular inflammation: genetic predisposition due to the polymorphism in *MTHFR*, *TNF*, *IL6* loci and the *HLA-DRB1* status [285]; activation of IL-1 β , IL-6, IL-17 and TNF- α pathways [285]; enhanced ECs activation with the expression of ICAM-1, VCAM-1, E-selectin [286]; the increase in NAPDH-derived ROS production, leading to eNOS uncoupling and formation of 3-nitrotyrosine.

RA is the most prevalent autoimmune inflammatory rheumatic disease [287], characterized by a 50% excess in CV mortality compared to the general population [288]. RA has been associated with diffuse microvascular injury (Fig. 3g) [24] documented by decreased myocardial perfusion (Figure 3, Supplemental Digital Content, <http://links.lww.com/HJH/C223>) [289], altered retinal arteriolar diameters [290,291], and dermal capillary density assessed with nailfold capillaroscopy [292], as well as impaired coronary microcirculation evaluated by coronary flow reserve (CFR) [293] and impaired endothelial glycocalyx integrity [294] even in the absence of overt CV disease. This is paired with an attenuated microvascular response to different stimuli, assessed with venous occlusion plethysmography [295,296], and an increased hyperaemic vasodilatory response [297] in RA patients compared to healthy controls. Pronounced impairment of microcirculatory blood flow responses assessed by laser speckle contrast imaging (LSCI) and decreased coronary microvascular perfusion has also been found in RA individuals free from CV disease [298] (Figure 4, Supplemental Digital Content, <http://links.lww.com/HJH/C223>).

In SLE, prevalent in about 0.1% of the general population, apart from the widespread inflammation and tissue damage in the affected organs, the blood vessels, especially the brain and kidneys, could also be severely impaired. Since vascular involvement, presenting as noninflammatory necrotic vasculopathy, thrombotic microangiopathy, and lupus vasculitis, is considered the leading cause of death in patients with SLE, interest has been focused on identifying the presence and role of early, subclinical microcirculation alterations, potentially anticipated before the establishment of CV events [25]. In addition to the classical subclinical structural changes (cortical atrophy and white matter hyperintensities), identified mostly by conventional MRI in patients with SLE regardless of the presence of neuropsychiatric manifestations, functional changes such as a blunted increase in cerebral oxygenation during exercise assessed with near infra-red spectroscopy [299] and hypoperfusion lesions with single-photon emission tomography (SPECT) in comparison to controls [300] are present. They often precede the permanent changes identified by conventional imaging [301] and are also found in sites

different from the classical one targeted by lupus vasculopathy (e.g., SLE nephritis): the fundus [26,302,303], the skin, with capillaroscopic alterations concerning density, dimensions, morphology and haemorrhages, and the myocardium, presenting coronary microvascular dysfunction [304–306]. Functional microcirculation studies with digital PAT [307,308], Portable Oxygen Transmitter [309] and LSCI [310,311] document reduced peripheral perfusion and impaired microvascular reactivity in SLE patients (Figure 4, Supplemental Digital Content, <http://links.lww.com/HJH/C223>).

Subclinical microvascular alterations in psoriatic disease, inflammatory bowel disease, vasculitis and AS have also been studied [312–314]. Psoriatic patients and patients with inflammatory bowel disease present coronary microcirculatory dysfunction, as assessed by CFR, reduced endothelial glycocalyx thickness and microvascular perfusion impairment leading to impaired cardiac function (Figure 4, Supplemental Digital Content, <http://links.lww.com/HJH/C223>) [27,28,315–318].

Remarkably, biological anti-inflammatory therapies and statins in autoimmune diseases improve endothelial glycocalyx and function, as well as coronary and peripheral microcirculation and thus, have beneficial effects on CV function [28,317,319,320], providing indirect evidence of the beneficial impact of targeting microvascular inflammation. Nonetheless, the precise role of MD and microvascular inflammation, in terms of risk prediction and therapeutic target, needs to be addressed appropriately by rigorous prospective studies.

Statement: ARD are a heterogeneous group of diseases characterised by MD and microvascular inflammation, driven by the combination of genetic predisposition and innate immunity hyperactivation via IL-1 β , IL-6, IL-17 and TNF- α pathways. The beneficial effects of anti-inflammatory drugs in terms of CV risk reduction provide indirect evidence of the centrality of microvascular inflammation in ARD.

Oncologic disease

As previously mentioned, systemic and local inflammation have a major role in the development and maintenance of microvascular structural alterations [9]. Oncologic diseases may cause or be associated with systemic inflammation, possibly contributing to the development of hypertension and CV diseases, thus reducing overall survival in these patients [321]. However, due to many clinical reasons, including relatively short follow-up periods, few data are available about microvascular alterations in cancer patients *per se*.

Cancer primarily shares with CV disease several pathophysiological mechanisms and similar risk factors. In this respect, chronic inflammation is a crucial feature in the pathogenesis and progression of both CV disease and cancer. It may be directly involved in the induction of some cancer types (e.g. H-pylori and stomach cancer) or indirectly promote local carcinogenesis and its progression by releasing inflammatory mediators and recruiting immune cells within the tumour microenvironment [29]. Other

mechanisms such as oxidative stress, cytokines, hormones (e.g. leptin), growth and metabolic factors have also been proposed to connect both diseases. However, a clear pathogenetic understanding is still lacking: although T cells appear to be involved, which specific subtype and by which mechanism they induce MD requires further investigation. [29]. The concomitant presence of CV risk factors or conditions such as physical inactivity, smoking, obesity, and diabetes may further induce inflammation worsening the prognosis of cancer and cancer survivor patients [30]. Cancer cells secrete VEGF to stimulate tumour vascularization, which increases vascular permeability and may contribute to microcirculation structural remodelling and perivascular fibrosis [31,32].

Particularly relevant is that several cancer treatments present CV toxicity and may cause MD, microvascular inflammation, hypertension and thus, an increase in CV events [321,322]. While anthracyclines have been mostly related to specific cardiotoxicity [323], VEGF and other tyrosine kinase inhibitors are the most frequently associated anticancer drugs with a dose-dependent increase in blood pressure both in hypertensive patients and in normotensive subjects [322,324,325]. These drugs enormously improve the prognosis for several solid tumours [326], targeting specific pro-angiogenic VEGF signalling involved in the neovascularization of tumours *in vivo* [327]. A consequent increase in blood pressure has been suggested as a pharmacodynamic biomarker and predictor of therapeutic efficacy [328,329]. However, this was not confirmed by other studies [330], and, what is more, poorly controlled hypertension leads to an increase in CV events, causing the discontinuation of anticancer therapy and thus hindering its clinical benefit.

The mechanism underlying vascular toxicity and hypertension induced by VEGF inhibitors is still debated. VEGF-A, the most important isoform of VEGF, may promote the proliferation, differentiation, and migration of endothelial cells by interacting with the VEGF-A receptor, as well as NO production [331]. Accordingly, VEGF inhibition is associated with reduced NO bioavailability because of the inhibition of eNOS and concomitant increase in vascular ROS [324,332], resulting in MD [31]. Activation of the ET-1 system with increased concentrations of ET-1, nephrotoxicity and impaired natriuresis induces hypertension along with the inhibition of other growth factors, including platelet-derived or fibroblast growth factor, c-Kit and FMS-like tyrosine kinase 3 [333]. Recently, a novel molecular mechanism involving the interplay between endothelial microparticles, the endothelin system and endothelial cell pro-inflammatory and redox signalling have been described; such interactions could be important in CV toxicity and hypertension associated with VEGF inhibitors [334]. All these events would favour an increase in peripheral resistance, further increasing MD.

Another consequence of antiangiogenic drugs leading to vascular resistance increase and elevated blood pressure is microvascular rarefaction. A reduction of capillary density during antiangiogenic treatment, reversible with cancer drug discontinuation [335], was observed in some [336–339] but not in other studies [340]. In one of these studies [338], the effect of antiangiogenic drugs on the structure of

retinal arterioles and capillary density was investigated in 20 patients with cancer. No change in systolic or diastolic blood pressure values during treatment was observed [338]; however, during the study, antihypertensive treatment was optimized in most patients. Although no difference was observed in the retinal arteriole wall-to-lumen ratio [19], capillary density was reduced by antiangiogenic drugs after three or six months (Fig. 3f) [338]. These findings might imply that an up-titration antihypertensive treatment is necessary for patients treated with tyrosine kinase inhibitors or a VEGF inhibitor. Indeed, under adequate blood pressure control, microvasculature seems preserved [338]. Since the efficacy of these drugs could be related to the extent of the antiangiogenic effect, the noninvasive evaluation of capillary density should be evaluated by further studies as a predictive parameter of drug efficacy. The better identification of the mechanisms underlying adverse cardiac and vascular effects of anticancer therapies may allow to develop novel vasculoprotective strategies. Only by doing so will patients achieve optimal cancer treatment at the minimum cost to cardiac and vascular health [323].

Statement: In oncological disease, the pan-activation of inflammatory response concurs to induce MD and microvascular inflammation. Even more relevant is that anticancer drugs, particularly anti-VEGF, might cause detrimental derangement in microvascular function and inflammation, thus attenuating their medium/long-term beneficial effects in terms of survival. An adequate increase in treatment to achieve a stronger control of age-related disease (in particular, hypertension) is thus required.

CONCLUSIONS

Ageing and age-related diseases are all characterised by different degrees of MD, leading to high-CV morbidity and mortality. As microvascular inflammation is both the consequence of environmental stressors and the perpetrator of age-related damage, its centrality in CV risk is apparent. However, though damage pathways have been extensively studied over the last decades, a clear understanding of their involvement's temporal and spatial sequence across the age-related disease spectrum is missing. In particular, although the interaction between immune and nonimmune cells is receiving increasing attention, a precise definition of their cross-talk in the context of MD is lacking. Preventing their detrimental dialogue may be crucial to stopping the disease at a very early stage. This gap of knowledge substantially limits the translation in terms of clinical strategies. Targeting microvascular inflammation is still a difficult road to travel: as the microvascular damage leverage epigenetic remodelling [52], early or intensive treatment is required to revert it. However, even as some interventions have demonstrated a potential benefit in terms of rescuing MD (e.g., physical activity, weight loss [166,167], SGLT-2i [200]) and inflammation [104], translational studies addressing microvascular inflammation to identify either early common or disease-specific targets are required. At the same time, we need to clearly understand the strengths and limitations of each technique used to assess MD, as well as the ability to distinguish between microvascular and

macrovascular. Efforts towards standardisation are needed to obtain interpretable results from studies.

ACKNOWLEDGEMENTS

The present work has not been presented previously elsewhere.

Funding: A.M. is supported by an International Grant from the Italian Society of Arterial Hypertension and a Research Grant from the Holcim-Stiftung. S.T.-C. is supported by a Wellcome Trust Institutional Strategic Support grant and a British Heart Foundation grant (PG/23/11093). K. Stellos is supported by the European Research Council (ERC) under the European Union's Horizon 2020 research and innovation programme (MODVASC, grant agreement no. 759248), the Biotechnology and Biological Sciences Research Council (BBSRC) of the UK Research and Innovation (UKRI), and the German Research Foundation DFG (CRC1366 C07, project number 394046768). P.A. has received funding by Greece and the European Union (European Social Fund ESF) through the Operational Programme "Human Resources Development, Education and Lifelong Learning 2014–2020" (MIS 5047870). T.J.G. and R.N. are funded by the European Research Council [ERC and InflammationTENSION; ERC-CoG-726318; to T.J.G.], British Heart Foundation [FS/14/49/30838 and FS/4yPhD/F/20/34127A; RE/18/5/34216], ERA-Net-CVD (ImmuneHyperCog; NCBiR, Poland) and Tenovus Scotland (RN- S22-03).

Conflicts of interest

The authors have no relevant conflict of interest to disclose.

REFERENCES

- Jin K. A microcirculatory theory of aging. *Aging Dis* 2019; 10:676–683.
- Güven G, Hilty MP, Ince C. Microcirculation: physiology, pathophysiology, and clinical application. *Blood Purif* 2020; 49:143–150.
- Reid L, Meyrick B. Microcirculation: definition and organization at tissue level. *Ann N Y Acad Sci* 1982; 384:3–20.
- RG JJ, de Jongh RT, Beijik MA, van Weissenbruch MM, Delemarre-van de Waal HA, Serné EH, *et al.* Individuals at increased coronary heart disease risk are characterized by an impaired microvascular function in skin. *Eur J Clin Invest* 2003; 33:536–542.
- Abularrage CJ, Sidawy AN, Aidinian G, Singh N, Weiswasser JM, Arora S. Evaluation of the microcirculation in vascular disease. *J Vasc Surg* 2005; 42:574–581.
- Arora S, Veves A, Caballero AE, Smakowski P, LoGerfo FW. Estrogen improves endothelial function. *J Vasc Surg* 1998; 27:1141–1146; discussion 1147.
- de Jongh RT, Serné EH, RG. JJ, de Vries G, Stehouwer CD. Impaired microvascular function in obesity: implications for obesity-associated microangiopathy, hypertension, and insulin resistance. *Circulation* 2004; 109:2529–2535.
- Crea F, Camici PG, Bairey Merz CN. Coronary microvascular dysfunction: an update. *Eur Heart J* 2014; 35:1101–1111.
- Rizzoni D, De Ciuceis C, Szczepaniak P, Paradis P, Schiffrin EL, Guzik TJ. Immune system and microvascular remodeling in humans. *Hypertension* 2022; 79:691–705.
- Wiseman S, Marlborough F, Doubal F, Webb DJ, Wardlaw J. Blood markers of coagulation, fibrinolysis, endothelial dysfunction and inflammation in lacunar stroke versus nonlacunar stroke and non-stroke: systematic review and meta-analysis. *Cerebrovasc Dis* 2014; 37:64–75.
- Giwa MO, Williams J, Elderfield K, Jiwa NS, Bridges LR, Kalaria RN, *et al.* Neuropathologic evidence of endothelial changes in cerebral small vessel disease. *Neurology* 2012; 78:167–174.
- Recio-Mayoral A, Rimoldi OE, Camici PG, Kaski JC. Inflammation and microvascular dysfunction in cardiac syndrome X patients without

- conventional risk factors for coronary artery disease. *JACC Cardiovasc Imaging* 2013; 6:660–667.
13. Grochowski C, Litak J, Kamieniak P, Maciejewski R. Oxidative stress in cerebral small vessel disease. Role of reactive species. *Free Radic Res* 2018; 52:1–13.
 14. Feuer DS, Handberg EM, Mehrad B, Wei J, Bairey Merz CN, Pepine CJ, et al. Microvascular dysfunction as a systemic disease: a review of the evidence. *Am J Med* 2022; 135:1059–1068.
 15. Secor D, Li F, Ellis CG, Sharpe MD, Gross PL, Wilson JX, et al. Impaired microvascular perfusion in sepsis requires activated coagulation and P-selectin-mediated platelet adhesion in capillaries. *Intensive Care Med* 2010; 36:1928–1934.
 16. De Backer D, Orbegozo Cortes D, Donadello K, Vincent JL. Pathophysiology of microcirculatory dysfunction and the pathogenesis of septic shock. *Virulence* 2014; 5:73–79.
 17. Thompson CS, Hakim AM. Living beyond our physiological means: small vessel disease of the brain is an expression of a systemic failure in arteriolar function: a unifying hypothesis. *Stroke* 2009; 40:e322–e330.
 18. Nowroozpoor A, Gutterman D, Safdar B. Is microvascular dysfunction a systemic disorder with common biomarkers found in the heart, brain, and kidneys? – a scoping review. *Microvasc Res* 2021; 134:104123.
 19. Rizzoni D, Mengozi A, Masi S, Agabiti Rosei C, De Ciuceis C, Virdis A. New noninvasive methods to evaluate microvascular structure and function. *Hypertension* 2022; 79:874–886.
 20. Odaka Y, Takahashi J, Tsuburaya R, Nishimiya K, Hao K, Matsumoto Y, et al. Plasma concentration of serotonin is a novel biomarker for coronary microvascular dysfunction in patients with suspected angina and unobstructive coronary arteries. *Eur Heart J* 2017; 38:489–496.
 21. Ahmad A, Corban MT, Toya T, Sara JD, Lerman B, Park JY, et al. Coronary microvascular endothelial dysfunction in patients with angina and nonobstructive coronary artery disease is associated with elevated serum homocysteine levels. *J Am Heart Assoc* 2020; 9:e017746.
 22. Rocco E, Grimaldi MC, Maino A, Cappannoli L, Pedicino D, Liuzzo G, et al. Advances and challenges in biomarkers use for coronary microvascular dysfunction: from bench to clinical practice. *J Clin Med* 2022; 11:.
 23. Del Buono MG, Montone RA, Camilli M, Carbone S, Narula J, Lavie CJ, et al. Coronary microvascular dysfunction across the spectrum of cardiovascular diseases: JACC state-of-the-art review. *J Am Coll Cardiol* 2021; 78:1352–1371.
 24. Sandoo A, Veldhuijzen van Zanten JJ, Metsios GS, Carroll D, Kitas GD. Vascular function and morphology in rheumatoid arthritis: a systematic review. *Rheumatology (Oxford)* 2011; 50:2125–2139.
 25. Saygin D, Highland KB, Tonelli AR. Microvascular involvement in systemic sclerosis and systemic lupus erythematosus. *Microcirculation* 2019; 26:e12440.
 26. Aissopou EK, Protogerou AD, Papaioannou TG, Tektonidou M, Tentolouris N, Theodosiadis PG, et al. Retinal vascular calibers in contemporary patients with chronic systemic inflammatory diseases: the Greek REtinal Microcirculation (GREM) study. *Artery Res* 2017; 18:1–6.
 27. Ikonomidis I, Pavlidis G, Lambadiari V, Rafouli-Stergiou P, Makavos G, Thymis J, et al. Endothelial glycocalyx and microvascular perfusion are associated with carotid intima-media thickness and impaired myocardial deformation in psoriatic disease. *J Hum Hypertens* 2022; 36:1113–1120.
 28. van Eijk IC, Peters MJ, Serné EH, van der Horst-Bruinsma IE, Dijkmans BA, Smulders YM, et al. Microvascular function is impaired in ankylosing spondylitis and improves after tumour necrosis factor alpha blockade. *Ann Rheum Dis* 2009; 68:362–336.
 29. Coussens LM, Werb Z. Inflammation and cancer. *Nature* 2002; 420:860–867.
 30. Koene RJ, Prizment AE, Blaes A, Konety SH. Shared risk factors in cardiovascular disease and cancer. *Circulation* 2016; 133:1104–1114.
 31. Terwoord JD, Beyer AM, Gutterman DD. Endothelial dysfunction as a complication of anticancer therapy. *Pharmacol Ther* 2022; 237:108116.
 32. Galán-Arriola C, Vilchez-Tschischke JP, Lobo M, López GJ, de Molina-Iracheta A, Pérez-Martínez C, et al. Coronary microcirculation damage in anthracycline cardiotoxicity. *Cardiovasc Res* 2022; 118:531–541.
 33. Agabiti-Rosei E, Rizzoni D. Microvascular structure as a prognostically relevant endpoint. *J Hypertens* 2017; 35:914–921.
 34. Heagerty AM. Changes in small artery structure in hypertension: ready for prognostic translation? *J Hypertens* 2017; 35:945–946.
 35. Zhang W, Singh S, Liu L, Mohammed AQ, Yin G, Xu S, et al. Prognostic value of coronary microvascular dysfunction assessed by coronary angiography-derived index of microcirculatory resistance in diabetic patients with chronic coronary syndrome. *Cardiovasc Diabetol* 2022; 21:222.
 36. Marks DS, Gudapati S, Prisant LM, Weir B, diDonato-Gonzalez C, Waller JL, et al. Mortality in patients with microvascular disease. *J Clin Hypertens (Greenwich)* 2004; 6:304–309.
 37. Nakanishi K, Fukuda S, Shimada K, Miyazaki C, Otsuka K, Maeda K, et al. Impaired coronary flow reserve as a marker of microvascular dysfunction to predict long-term cardiovascular outcomes, acute coronary syndrome and the development of heart failure. *Circ J* 2012; 76:1958–1964.
 38. Bajaj NS, Osborne MT, Gupta A, Tavakkoli A, Bravo PE, Vita T, et al. Coronary microvascular dysfunction and cardiovascular risk in obese patients. *J Am Coll Cardiol* 2018; 72:707–717.
 39. Gdowski MA, Murthy VL, Doering M, Monroy-Gonzalez AG, Slart R, Brown DL. Association of isolated coronary microvascular dysfunction with mortality and major adverse cardiac events: a systematic review and meta-analysis of aggregate data. *J Am Heart Assoc* 2020; 9:e014954.
 40. Zeiher AM, Drexler H, Wollschläger H, Just H. Endothelial dysfunction of the coronary microvasculature is associated with coronary blood flow regulation in patients with early atherosclerosis. *Circulation* 1991; 84:1984–1992.
 41. Toya T, Sara JD, Ahmad A, Nardi V, Taher R, Lerman LO, et al. Incremental prognostic impact of peripheral microvascular endothelial dysfunction on the development of ischemic stroke. *J Am Heart Assoc* 2020; 9:e015703.
 42. Young A, Garcia M, Sullivan SM, Liu C, Moazzami K, Ko YA, et al. Impaired peripheral microvascular function and risk of major adverse cardiovascular events in patients with coronary artery disease. *Arterioscler Thromb Vasc Biol* 2021; 41:1801–1809.
 43. Ikonomidis I, Thymis J, Simitis P, Koliou GA, Katsanos S, Triantafyllou C, et al. Impaired endothelial glycocalyx predicts adverse outcome in subjects without overt cardiovascular disease: a 6-year follow-up study. *J Cardiovasc Transl Res* 2022; 15:890–902.
 44. Jung C, Fuernau G, Muench P, Desch S, Eitel I, Schuler G, et al. Impairment of the endothelial glycocalyx in cardiogenic shock and its prognostic relevance. *Shock* 2015; 43:450–455.
 45. Rensma SP, van Sloten TT, Launer IJ, Stehouwer CDA. Cerebral small vessel disease and risk of incident stroke, dementia and depression, and all-cause mortality: a systematic review and meta-analysis. *Neurosci Biobehav Rev* 2018; 90:164–173.
 46. Yu W, Gao W, Rong D, Wu Z, Khalil RA. Molecular determinants of microvascular dysfunction in hypertensive pregnancy and preeclampsia. *Microcirculation* 2018; e12508.
 47. Ghosh A, Freestone NS, Anim-Nyame N, Arrignon FIF. Microvascular function in preeclampsia is influenced by insulin resistance and an imbalance of angiogenic mediators. *Physiol Rep* 2017; 5:.
 48. Yu GZ, Aye CY, Lewandowski AJ, Davis EF, Khoo CP, Newton L, et al. Association of maternal antiangiogenic profile at birth with early postnatal loss of microvascular density in offspring of hypertensive pregnancies. *Hypertension* 2016; 68:749–759.
 49. Pan HT, Shi XL, Fang M, Sun XM, Chen PP, Ding JL, et al. Profiling of exosomal microRNAs expression in umbilical cord blood from normal and preeclampsia patients. *BMC Pregnancy Childbirth* 2022; 22:124.
 50. Carlomagno F, Pozza C, Tenuta M, Pofi R, Tarani L, Sesti F, et al. Testicular microvascular flow is altered in Klinefelter syndrome and predicts circulating testosterone. *J Clin Endocrinol Metab* 2022; 107:e236–e245.
 51. Lucas-Herald AK, Montezano AC, Alves-Lopes R, Haddow L, Alimusina M, O'Toole S, et al. Vascular dysfunction and increased cardiovascular risk in hypospadias. *Eur Heart J* 2022; 43:1832–1845.
 52. Ungvari Z, Tarantini S, Donato AJ, Galvan V, Csiszar A. Mechanisms of vascular aging. *Circ Res* 2018; 123:849–867.
 53. Chen G, Yung R. Meta-inflammaging at the crossroad of geroscience. *Aging Med (Milton)* 2019; 2:157–161.
 54. La Favor JD, Dubis GS, Yan H, White JD, Nelson MA, Anderson EJ, et al. Microvascular endothelial dysfunction in sedentary, obese humans is mediated by NADPH Oxidase: influence of exercise training. *Arterioscler Thromb Vasc Biol* 2016; 36:2412–2420.

55. Flockhart M, Nilsson LC, Tais S, Eklblom B, Apró W, Larsen FJ. Excessive exercise training causes mitochondrial functional impairment and decreases glucose tolerance in healthy volunteers. *Cell Metab* 2021; 33:957–970.
56. Neville CE, Montgomery S, Silvestri G, McGowan A, Moore E, Silvestri V, *et al.* Dietary patterns and retinal vessel caliber in the Irish nun eye study. *J Nutr Health Aging* 2018; 22:751–758.
57. van der Velden AIM, van den Berg BM, de Mutsert R, van der Vlag J, Jukema JW, Rosendaal FR, *et al.* Microvascular differences in individuals with obesity at risk of developing cardiovascular disease. *Obesity (Silver Spring)* 2021; 29:1439–1444.
58. Tam BT, Morais JA, Santosa S. Obesity and ageing: two sides of the same coin. *Obes Rev* 2020; 21:e12991.
59. Chen Y, Harris S, Rogers Y, Ahmad T, Asselbergs FW. Nudging within learning health systems: next generation decision support to improve cardiovascular care. *Eur Heart J* 2022; 43:1296–1306.
60. Pillon NJ, Loos RJF, Marshall SM, Zierath JR. Metabolic consequences of obesity and type 2 diabetes: balancing genes and environment for personalized care. *Cell* 2021; 184:1530–1544.
61. Mengozzi A, Carli F, Biancalana E, Della Latta V, Seghieri M, Gastaldelli A, *et al.* Phthalates exposure as determinant of albuminuria in subjects with type 2 diabetes: a cross-sectional study. *J Clin Endocrinol Metab* 2019; 104:1491–1499.
62. De Backer D, Riccottilli F, Ospina-Tascón GA. Septic shock: a microcirculation disease. *Curr Opin Anaesthesiol* 2021; 34:85–91.
63. Nägele MP, Haubner B, Tanner FC, Ruschitzka F, Flammer AJ. Endothelial dysfunction in COVID-19: current findings and therapeutic implications. *Atherosclerosis* 2020; 314:58–62.
64. Tsuzuki K, Shimizu Y, Suzuki J, Pu Z, Yamaguchi S, Fujikawa Y, *et al.* Adverse effect of circadian rhythm disorder on reparative angiogenesis in hind limb ischemia. *J Am Heart Assoc* 2021; 10:e020896.
65. Holmer BJ, Lapierre SS, Jake-Schoffman DE, Christou DD. Effects of sleep deprivation on endothelial function in adult humans: a systematic review. *GeroScience* 2021; 43:137–158.
66. Chaseling GK, Debray A, Gravel H, Ravanelli N, Bartlett AA, Gagnon D. The acute effect of heat exposure on forearm macro- and microvascular function: Impact of measurement timing, heating modality and biological sex. *Exp Physiol* 2022; 108:221–239.
67. Heinonen I, Laukkanen JA. Effects of heat and cold on health, with special reference to Finnish sauna bathing. *Am J Physiol Regul Integr Comp Physiol* 2018; 314:R629–R638.
68. Achebak H, Devolder D, Ballester J. Trends in temperature-related age-specific and sex-specific mortality from cardiovascular diseases in Spain: a national time-series analysis. *Lancet Planet Health* 2019; 3:e297–e306.
69. Taddei S, Virdis A, Mattei P, Ghiadoni L, Gennari A, Fasolo CB, *et al.* Aging and endothelial function in normotensive subjects and patients with essential hypertension. *Circulation* 1995; 91:1981–1987.
70. Taddei S, Virdis A, Mattei P, Ghiadoni L, Fasolo CB, Sudano I, *et al.* Hypertension causes premature aging of endothelial function in humans. *Hypertension* 1997; 29:736–743.
71. Mengozzi A, Pugliese NR, Chiriaco M, Masi S, Virdis A, Taddei S. Microvascular ageing links metabolic disease to age-related disorders: the role of oxidative stress and inflammation in promoting microvascular dysfunction. *J Cardiovasc Pharmacol* 2021; 78:S78–D87.
72. Franceschi C, Garagnani P, Parini P, Giuliani C, Santoro A. Inflammaging: a new immune-metabolic viewpoint for age-related diseases. *Nat Rev Endocrinol* 2018; 14:576–590.
73. Griendling KK, Camargo LL, Rios FJ, Alves-Lopes R, Montezano AC, Touyz RM. Oxidative stress and hypertension. *Circ Res* 2021; 128:993–1020.
74. Masi S, Rizzoni D, Taddei S, Widmer RJ, Montezano AC, Lüscher TF, *et al.* Assessment and pathophysiology of microvascular disease: recent progress and clinical implications. *Eur Heart J* 2020; 00:1–15.
75. Tesaro M, Mauriello A, Rovella V, Annicchiarico-Petruzzelli M, Cardillo C, Melino G, *et al.* Arterial ageing: from endothelial dysfunction to vascular calcification. *J Intern Med* 2017; 281:471–482.
76. Saltiel AR, Olefsky JM. Inflammatory mechanisms linking obesity and metabolic disease. *J Clin Invest* 2017; 127:1–4.
77. Potente M, Gerhardt H, Carmeliet P. Basic and therapeutic aspects of angiogenesis. *Cell* 2011; 146:873–887.
78. Ungvari Z, Tarantini S, Kiss T, Wren JD, Giles CB, Griffin CT, *et al.* Endothelial dysfunction and angiogenesis impairment in the ageing vasculature. *Nat Rev Cardiol* 2018; 15:555–565.
79. Jiang X, Tian W, Nicolls MR, Rockson SG. The lymphatic system in obesity, insulin resistance, and cardiovascular diseases. *Front Physiol* 2019; 10:1402.
80. Chakraborty A, Barajas S, Lammoglia GM, Reyna AJ, Morley TS, Johnson JA, *et al.* Vascular endothelial growth factor-D (VEGF-D) overexpression and lymphatic expansion in murine adipose tissue improves metabolism in obesity. *Am J Pathol* 2019; 189:924–939.
81. Cuijpers I, Simmonds SJ, van Bilsen M, Czarnowska E, González Miqueo A, Heymans S, *et al.* Microvascular and lymphatic dysfunction in HFpEF and its associated comorbidities. *Basic Res Cardiol* 2020; 115:39.
82. Paneni F, Mocharla P, Akhmedov A, Costantino S, Osto E, Volpe M, *et al.* Gene silencing of the mitochondrial adaptor p66(Shc) suppresses vascular hyperglycemic memory in diabetes. *Circ Res* 2012; 111:278–289.
83. Wise IA, Charchar FJ. Epigenetic modifications in essential hypertension. *Int J Mol Sci* 2016; 17:451.
84. Virdis A, Duranti E, Rossi C, Dell'Agnello U, Santini E, Anselmino M, *et al.* Tumour necrosis factor- α participates on the endothelin-1/nitric oxide imbalance in small arteries from obese patients: role of perivascular adipose tissue. *Eur Heart J* 2015; 36:784–794.
85. Marques FZ, Morris BJ. Neurogenic hypertension: revelations from genome-wide gene expression profiling. *Curr Hypertens Rep* 2012; 14:485–491.
86. Ku KH, Subramaniam N, Marsden PA. Epigenetic determinants of flow-mediated vascular endothelial gene expression. *Hypertension* 2019; 74:467–476.
87. Cooke J. Endotheliopathy of obesity. *Circulation* 2020; 142:380–383.
88. Canfrán-Duque A, Rotllan N, Zhang X, Andrés-Blasco I, Thompson BM, Sun J, *et al.* Macrophage-derived 25-hydroxycholesterol promotes vascular inflammation, atherogenesis, and lesion remodeling. *Circulation* 2023; 147:388–408.
89. Brassington K, Kanellakis P, Cao A, Toh BH, Peter K, Bobik A, *et al.* Crosstalk between cytotoxic CD8+ T cells and stressed cardiomyocytes triggers development of interstitial cardiac fibrosis in hypertensive mouse hearts. *Front Immunol* 2022; 13:1040233.
90. Virdis A, Masi S, Colucci R, Chiriaco M, Uliana M, Puxeddu I, *et al.* Microvascular endothelial dysfunction in patients with obesity. *Curr Hypertens Rep* 2019; 21:32.
91. Horton WB, Barrett EJ. Microvascular dysfunction in diabetes mellitus and cardiometabolic disease. *Endocr Rev* 2021; 42:29–55.
92. Petrie JR, Guzik TJ, Touyz RM. Diabetes, hypertension, and cardiovascular disease: clinical insights and vascular mechanisms. *Can J Cardiol* 2018; 34:575–584.
93. Virdis A, Santini F, Colucci R, Duranti E, Salvetti G, Rugani I, *et al.* Vascular generation of tumor necrosis factor- α reduces nitric oxide availability in small arteries from visceral fat of obese patients. *J Am Coll Cardiol* 2011; 58:238–247.
94. Liberale L, Montecucco F, Tardif JC, Libby P, Camici GG. Inflammaging: the role of inflammation in age-dependent cardiovascular disease. *Eur Heart J* 2020; 41:2974–2982.
95. Lee YS, Wollam J, Olefsky JM. An integrated view of immunometabolism. *Cell* 2018; 172:22–40.
96. Tsujikawa LM, Fu L, Das S, Halliday C, Rakai BD, Stotz SC, *et al.* Apabetalone (RVX-208) reduces vascular inflammation in vitro and in CVD patients by a BET-dependent epigenetic mechanism. *Clin Epigenetics* 2019; 11:102.
97. Cummings J, Schwartz GG, Nicholls SJ, Khan A, Halliday C, Toth PP, *et al.* Cognitive effects of the BET protein inhibitor apabetalone: a prespecified montreal cognitive assessment analysis nested in the BETonMACE randomized controlled trial. *J Alzheimers Dis* 2021; 83:1703–1715.
98. Toth PP, Schwartz GG, Nicholls SJ, Khan A, Szarek M, Ginsberg HN, *et al.* Reduction in the risk of major adverse cardiovascular events with the BET protein inhibitor apabetalone in patients with recent acute coronary syndrome, type 2 diabetes, and moderate to high likelihood of nonalcoholic fatty liver disease. *Am J Prev Cardiol* 2022; 11.
99. Cerqueira SR, Benavides S, Lee HE, Ayad NG, Lee JK. BET protein inhibition promotes nonmyeloid cell mediated neuroprotection after rodent spinal cord contusion. *Exp Neurol* 2022;352.
100. Faivre EJ, McDaniel KF, Albert DH, Mantena SR, Plotnik JP, Wilcox D, *et al.* Selective inhibition of the BD2 bromodomain of BET proteins in prostate cancer. *Nature* 2020; 578:306–310.

101. Li J, Ma J, Meng G, Lin H, Wu S, Wang J, et al. BET bromodomain inhibition promotes neurogenesis while inhibiting gliogenesis in neural progenitor cells. *Stem Cell Res* 2016; 17:212–221.
102. Gilan O, Rioja I, Knezevic K, Bell MJ, Yeung MM, Harker NR, et al. Selective targeting of BD1 and BD2 of the BET proteins in cancer and immunoinflammation. *Science* 2020; 368:387–394.
103. Klein K. Bromodomain protein inhibition: a novel therapeutic strategy in rheumatic diseases. *RMD open* 2018; 4:e000744.
104. Liberale L, Montecucco F, Schwarz L, Lüscher TF, Camici GG. Inflammation and cardiovascular diseases: lessons from seminal clinical trials. *Cardiovasc Res* 2021; 117:411–422.
105. Ward ZJ, Bleich SN, Craddock AL, Barrett JL, Giles CM, Flax C, et al. Projected U.S. state-level prevalence of adult obesity and severe obesity. *N Engl J Med* 2019; 381:2440–2450.
106. Costantino S, Paneni F, Cosentino F. Ageing, metabolism and cardiovascular disease. *J Physiol* 2016; 594:2061–2073.
107. Donato AJ, Machin DR, Lesniewski LA. Mechanisms of dysfunction in the aging vasculature and role in age-related disease. *Circ Res* 2018; 123:825–848.
108. Bruno RM, Duranti E, Ippolito C, Segnani C, Bernardini N, Di Candio G, et al. Different impact of essential hypertension on structural and functional age-related vascular changes. *Hypertension* 2016; 69:71–78.
109. Gorgoulis V, Adams PD, Alimonti A, Bennett DC, Bischof O, Bishop C, et al. Cellular senescence: defining a path forward. *Cell* 2019; 179:813–827.
110. Di Micco R, Krizhanovsky V, Baker D, d'Adda di Fagnagna F. Cellular senescence in ageing: from mechanisms to therapeutic opportunities. *Nat Rev Mol Cell Biol* 2021; 22:75–95.
111. McHugh D, Gil J. Senescence and aging: causes, consequences, and therapeutic avenues. *J Cell Biol* 2018; 217:65–77.
112. Bhayadia R, Schmidt BM, Melk A, Hömme M. Senescence-induced oxidative stress causes endothelial dysfunction. *J Gerontol A Biol Sci Med Sci* 2016; 71:161–169.
113. Sun X, Feinberg MW. Vascular endothelial senescence: pathobiological insights, emerging long noncoding RNA targets, challenges and therapeutic opportunities. *Front Physiol* 2021; 12:693067.
114. Rossman MJ, Kaplon RE, Hill SD, McNamara MN, Santos-Parker JR, Pierce GL, et al. Endothelial cell senescence with aging in healthy humans: prevention by habitual exercise and relation to vascular endothelial function. *Am J Physiol Heart Circ Physiol* 2017; 313:H890–H895.
115. Dikic I, Elazar Z. Mechanism and medical implications of mammalian autophagy. *Nat Rev Mol Cell Biol* 2018; 19:349–364.
116. Aman Y, Schmauck-Medina T, Hansen M, Morimoto RI, Simon AK, Bjedov I, et al. Autophagy in healthy aging and disease. *Nat Aging* 2021; 1:634–650.
117. Bharath LP, Mueller R, Li Y, Ruan T, Kunz D, Goodrich R, et al. Impairment of autophagy in endothelial cells prevents shear-stress-induced increases in nitric oxide bioavailability. *Can J Physiol Pharmacol* 2014; 92:605–612.
118. Tai H, Wang Z, Gong H, Han X, Zhou J, Wang X, et al. Autophagy impairment with lysosomal and mitochondrial dysfunction is an important characteristic of oxidative stress-induced senescence. *Autophagy* 2017; 13:99–113.
119. LaRocca TJ, Henson GD, Thorburn A, Sindler AL, Pierce GL, Seals DR. Translational evidence that impaired autophagy contributes to arterial ageing. *J Physiol* 2012; 590:3305–3316.
120. Sun Q, Fan J, Billiar TR, Scott MJ. Inflammasome and autophagy regulation – a two-way street. *Mol Med* 2017; 23:188–195.
121. Sies H, Berndt C, Jones DP. Oxidative stress. *Annu Rev Biochem* 2017; 86:715–748.
122. Ungvari Z, Bailey-Downs L, Sosnowska D, Gautam T, Koncz P, Losonczy G, et al. Vascular oxidative stress in aging: a homeostatic failure due to dysregulation of NRF2-mediated antioxidant response. *Am J Physiol Heart Circ Physiol* 2011; 301:363–372.
123. Liguori I, Russo G, Curcio F, Bulli G, Aran L, Della-Morte D, et al. Oxidative stress, aging, and diseases. *Clin Interv Aging* 2018; 13:757–772.
124. Paulus WJ, Tschöpe C. A novel paradigm for heart failure with preserved ejection fraction: comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J Am Coll Cardiol* 2013; 62:263–271.
125. Mengozi A, Costantino S, Paneni F, Duranti E, Nannipieri M, Mancini R, et al. Targeting SIRT1 rescues age- and obesity-induced microvascular dysfunction in ex-vivo human vessels. *Circ Res* 2022; 131:476–491.
126. Xian H, Watari K, Sanchez-Lopez E, Offenberger J, Onyuru J, Sampath H, et al. Oxidized DNA fragments exit mitochondria via mPTP- and VDAC-dependent channels to activate NLRP3 inflammasome and interferon signaling. *Immunity* 2022; 55:1370–1385; e8.
127. Trumpff C, Michelson J, Lagranha CJ, Taleon V, Karan KR, Sturm G, et al. Stress and circulating cell-free mitochondrial DNA: a systematic review of human studies, physiological considerations, and technical recommendations. *Mitochondrion* 2021; 59:225–245.
128. Liu T, Zhang L, Joo D, Sun SC. NF- κ B signaling in inflammation. *Signal Transduct Target Ther* 2017; 2.
129. Jablonski KL, Seals DR, Eskurza I, Monahan KD, Donato AJ. High-dose ascorbic acid infusion abolishes chronic vasoconstriction and restores resting leg blood flow in healthy older men. *J Appl Physiol* 2007; 103:1715–1721.
130. Donato AJ, Eskurza I, Silver AE, Levy AS, Pierce GL, Gates PE, et al. Direct evidence of endothelial oxidative stress with aging in humans: relation to impaired endothelium-dependent dilation and upregulation of nuclear factor-kappaB. *Circ Res* 2007; 100:1659–1966.
131. Schafer MJ, Zhang X, Kumar A, Atkinson EJ, Zhu Y, Jachim S, et al. The senescence-associated secretome as an indicator of age and medical risk. *JCI insight* 2020; 5.
132. Donato AJ, Black AD, Jablonski KL, Gano LB, Seals DR. Aging is associated with greater nuclear NF kappa B, reduced I kappa B alpha, and increased expression of proinflammatory cytokines in vascular endothelial cells of healthy humans. *Ageing cell* 2008; 7:805–812.
133. Singh T, Newman AB. Inflammatory markers in population studies of aging. *Ageing Res Rev* 2011; 10:319–329.
134. Santoro A, Bientinesi E, Monti D. Immunosenescence and inflammation in the aging process: age-related diseases or longevity? *Ageing Res Rev* 2021; 71:101422.
135. Lazaridis A, Gavriilaki E, Douma S, Gkaliagkousi E. Toll-Like receptors in the pathogenesis of essential hypertension. A forthcoming immune-driven theory in full effect. *Int J Mol Sci* 2021; 22.
136. Shaw AC. Effects of aging on human toll-like receptor function.. In: Fulop T, Franceschi C, Hirokawa K, Pawelec G, editors. *Handbook of immunosenescence: basic understanding and clinical implications*. Cham: Springer International Publishing; 2018. pp. 1–12.
137. Latz E, Duweil P. NLRP3 inflammasome activation in inflammaging. *Semin Immunol* 2018; 40:61–73.
138. Pierce GL, Lesniewski LA, Lawson BR, Beske SD, Seals DR. Nuclear factor- κ B activation contributes to vascular endothelial dysfunction via oxidative stress in overweight/obese middle-aged and older humans. *Circulation* 2009; 119:1284–1292.
139. Manea A, Manea SA, Gafencu AV, Raicu M. Regulation of NADPH oxidase subunit p22(phox) by NF- κ B in human aortic smooth muscle cells. *Arch Physiol Biochem* 2007; 113:163–172.
140. Fulop T, Larbi A, Dupuis G, Le Page A, Frost EH, Cohen AA, et al. Immunosenescence and inflamm-aging as two sides of the same coin: friends or foes? *Front Immunol* 2018; 8:1960.
141. Chatterjee S. *Chapter 2 – Oxidative stress, inflammation, and disease*. In: Dziubla T, Butterfield DA, editors. *Oxidative stress and biomaterials*: Academic Press; 2016. pp. 35–58.
142. Kane AE, Sinclair DA. Sirtuins and NAD + in the development and treatment of metabolic and cardiovascular diseases. *Circ Res* 2018; 123:868–885.
143. Kitada M, Ogura Y, Koya D. The protective role of Sirt1 in vascular tissue: its relationship to vascular aging and atherosclerosis. *Ageing (Albany NY)* 2016; 8:2290–2307.
144. Xie J, Zhang X, Zhang L. Negative regulation of inflammation by SIRT1. *Pharmacol Res* 2013; 67:60–67.
145. Zhang HN, Dai Y, Zhang CH, Omondi AM, Ghosh A, Khanra I, et al. Sirtuins family as a target in endothelial cell dysfunction: implications for vascular ageing. *Biogerontology* 2020; 21:495–516.
146. Donato AJ, Magerko KA, Lawson BR, Durrant JR, Lesniewski LA, Seals DR. SIRT-1 and vascular endothelial dysfunction with ageing in mice and humans. *J Physiol* 2011; 589 (Pt 18):4545–4554.
147. Thompson AM, Wagner R, Rzcudlo EM. Age-related loss of Sirt1 expression results in dysregulated human vascular smooth muscle cell function. *Am J Physiol Heart Circ Physiol* 2014; 307:H533–H541.

148. Vachharajani VT, Liu T, Wang X, Hoth JJ, Yoza BK, McCall CE. Sirtuins link inflammation and metabolism. *J Immunol Res* 2016;8167273.
149. Chew NWS, Ng CH, Tan DJH, Kong G, Lin C, Chin YH, *et al.* The global burden of metabolic disease: data from 2000 to 2019. *Cell Metab* 2023; 35:414–428; e3.
150. Masi S, Colucci R, Duranti E, Nannipieri M, Anselmino M, Ippolito C, *et al.* Aging modulates the influence of arginase on endothelial dysfunction in obesity. *Arterioscler Thromb Vasc Biol* 2018; 38:2474–2483.
151. He Q, Gao Z, Yin J, Zhang J, Yun Z, Ye J. Regulation of HIF-1{alpha} activity in adipose tissue by obesity-associated factors: adipogenesis, insulin, and hypoxia. *Am J Physiol Endocrinol Metab* 2011; 300:E877–E885.
152. Antonopoulos AS, Margaritis M, Coutinho P, Shirodaria C, Psarros C, Herdman L, *et al.* Adiponectin as a link between type 2 diabetes and vascular NADPH oxidase activity in the human arterial wall: the regulatory role of perivascular adipose tissue. *Diabetes* 2015; 64:2207–2219.
153. Mengozzi A, Pugliese NR, Taddei S, Masi S, Virdis A. Microvascular inflammation and cardiovascular prevention: the role of microcirculation as earlier determinant of cardiovascular risk. *High Blood Press Cardiovasc Prev* 2021; 29:41–48.
154. Saxton SN, Clark BJ, Withers SB, Eringa EC, Heagerty AM. Mechanistic links between obesity, diabetes, and blood pressure: role of perivascular adipose tissue. *Physiol Rev* 2019; 99:1701–1763.
155. Guzik TJ, Touyz RM. Oxidative stress, inflammation, and vascular aging in hypertension. *Hypertension* 2017; 70:660–667.
156. Gruber T, Pan C, Contreras RE, Wiedemann T, Morgan DA, Skowronski AA, *et al.* Obesity-associated hyperleptinemia alters the gliovascular interface of the hypothalamus to promote hypertension. *Cell Metab* 2021; 33:1155.e10–1170.e10.
157. Withers SB, Agabiti-Rosei C, Livingstone DM, Little MC, Aslan R, Malik RA, *et al.* Macrophage activation is responsible for loss of anticontractile function in inflamed perivascular fat. *Arterioscler Thromb Vasc Biol* 2011; 31:908–913.
158. Tang X, Miao Y, Luo Y, Sriram K, Qi Z, Lin FM, *et al.* Suppression of endothelial AGO1 promotes adipose tissue browning and improves metabolic dysfunction. *Circulation* 2020; 142:35–379.
159. Zhou R, Yazdi AS, Menu P, Tschopp J. A role for mitochondria in NLRP3 inflammasome activation. *Nature* 2011; 469:221–225.
160. Guo Y, Gu R, Gan D, Hu F, Li G, Xu G. Mitochondrial DNA drives noncanonical inflammation activation via cGAS-STING signaling pathway in retinal microvascular endothelial cells. *Cell Commun Signal* 2020; 18:172.
161. Cohen HY, Miller C, Bitterman KJ, Wall NR, Hekking B, Kessler B, *et al.* Calorie restriction promotes mammalian cell survival by inducing the SIRT1 deacetylase. *Science* 2004; 305:390–392.
162. Maranhão PA, de Souza M, Kraemer-Aguar LG, Bouskela E. Dynamic nailfold videocapillaroscopy may be used for early detection of microvascular dysfunction in obesity. *Microvasc Res* 2016; 106:31–35.
163. Lee DH, Dane MJ, van den Berg BM, Boels MG, van Teeffelen JW, de Mutser R, *et al.* Deeper penetration of erythrocytes into the endothelial glycocalyx is associated with impaired microvascular perfusion. *PLoS One* 2014; 9:e96477.
164. Köchli S, Smith W, Lona G, Goikoetxea-Sotelo G, Breet Y, Botha-Le Roux S, *et al.* Obesity, blood pressure and retinal microvascular phenotype in a bi-ethnic cohort of young children. *Atherosclerosis* 2022; 350:51–57.
165. Kotliar KE, Lanzl IM, Schmidt-Trucksäss A, Sitnikova D, Ali M, Blume K, *et al.* Dynamic retinal vessel response to flicker in obesity: a methodological approach. *Microvasc Res* 2011; 81:123–128.
166. Csipo T, Fulop GA, Lipecz A, Tarantini S, Kiss T, Balasubramanian P, *et al.* Short-term weight loss reverses obesity-induced microvascular endothelial dysfunction. *GeroScience* 2018; 40:337–346.
167. Aghamohammadzadeh R, Greenstein AS, Yadav R, Jeziorska M, Hama S, Soltani F, *et al.* Effects of bariatric surgery on human small artery function: evidence for reduction in perivascular adipocyte inflammation, and the restoration of normal anticontractile activity despite persistent obesity. *J Am Coll Cardiol* 2013; 62:128–135.
168. Pickup JC, Chusney GD, Thomas SM, Burt D. Plasma interleukin-6, tumour necrosis factor alpha and blood cytokine production in type 2 diabetes. *Life Sci* 2000; 67:291–300.
169. Nguyen DV, Shaw LC, Grant MB. Inflammation in the pathogenesis of microvascular complications in diabetes. *Front Endocrinol (Lausanne)* 2012; 3:.
170. Esposito K, Nappo F, Marfella R, Giugliano G, Giugliano F, Ciotola M, *et al.* Inflammatory cytokine concentrations are acutely increased by hyperglycemia in humans: role of oxidative stress. *Circulation* 2002; 106:2067–2072.
171. Choi HJ, Jeon SY, Hong WK, Jung SE, Kang HJ, Kim JW, *et al.* Effect of glucose ingestion in plasma markers of inflammation and oxidative stress: analysis of 16 plasma markers from oral glucose tolerance test samples of normal and diabetic patients. *Diabetes Res Clin Pract* 2013; 99:e27–e31.
172. Bahceci M, Tuzcu A, Ogun C, Canoruc N, Iltimur K, Aslan C. Is serum C-reactive protein concentration correlated with HbA1c and insulin resistance in Type 2 diabetic men with or without coronary heart disease? *J Endocrinol Invest* 2005; 28:145–150.
173. Kado S, Nagase T, Nagata N. Circulating levels of interleukin-6, its soluble receptor and interleukin-6/interleukin-6 receptor complexes in patients with type 2 diabetes mellitus. *Acta Diabetol* 1999; 36:67–72.
174. Alzamil H. Elevated serum TNF- α is related to obesity in type 2 diabetes mellitus and is associated with glycemic control and insulin resistance. *J Obes* 2020; 2020:.
175. Nishikawa T, Edelstein D, Du XL, Yamagishi S, Matsumura T, Kaneda Y, *et al.* Normalizing mitochondrial superoxide production blocks three pathways of hyperglycaemic damage. *Nature* 2000; 404:787–790.
176. Chavakis T, Bierhaus A, Nawroth PP. RAGE (receptor for advanced glycation end products): a central player in the inflammatory response. *Microbes Infect* 2004; 6:1219–1225.
177. Muniyappa R, Sowers JR. Role of insulin resistance in endothelial dysfunction. *Rev Endocr Metab Disord* 2013; 14:5–12.
178. Kim JA, Montagnani M, Koh KK, Quon MJ. Reciprocal relationships between insulin resistance and endothelial dysfunction: molecular and pathophysiological mechanisms. *Circulation* 2006; 113:188–1904.
179. Ikonomidis I, Lambadiari V, Pavlidis G, Koukoulis C, Kousathana F, Varoudi M, *et al.* Insulin resistance and acute glucose changes determine arterial elastic properties and coronary flow reserve in dysglycaemic and first-degree relatives of diabetic patients. *Atherosclerosis* 2015; 241:455–462.
180. Lambadiari V, Korakas E, Oikonomou E, Bletsas E, Kountouri A, Goliopoulou A, *et al.* COVID-19, endothelium and the cardiometabolic patient: a possible role for capillary leak syndrome. *Biomedicines* 2022; 10:2379.
181. Min JK, Kim YM, Kim SW, Kwon MC, Kong YY, Hwang IK, *et al.* TNF-related activation-induced cytokine enhances leukocyte adhesiveness: induction of ICAM-1 and VCAM-1 via TNF receptor-associated factor and protein kinase C-dependent NF-kappaB activation in endothelial cells. *J Immunol* 2005; 175:531–540.
182. Ikonomidis I, Pavlidis G, Lambadiari V, Kousathana F, Varoudi M, Spanoudi F, *et al.* Early detection of left ventricular dysfunction in first-degree relatives of diabetic patients by myocardial deformation imaging: the role of endothelial glycocalyx damage. *Int J Cardiol* 2017; 233:105–112.
183. Nieuwdorp M, van Haeften TW, Gouverneur MC, Mooij HL, van Lieshout MH, Levi M, *et al.* Loss of endothelial glycocalyx during acute hyperglycemia coincides with endothelial dysfunction and coagulation activation in vivo. *Diabetes* 2006; 55:480–486.
184. Broekhuizen LN, Lemkes BA, Mooij HL, Meuwese MC, Verberne H, Holleman F, *et al.* Effect of sulodexide on endothelial glycocalyx and vascular permeability in patients with type 2 diabetes mellitus. *Diabetologia* 2010; 53:2646–2655.
185. Lekakis J, Abraham P, Balbarini A, Blann A, Boulanger CM, Cockcroft J, *et al.* Methods for evaluating endothelial function: a position statement from the European Society of Cardiology Working Group on Peripheral Circulation. *Eur J Cardiovasc Prev Rehabil* 2011; 18:775–789.
186. Lambadiari V, Pavlidis G, Kousathana F, Maratou E, Georgiou D, Andreadou I, *et al.* Effects of different antidiabetic medications on endothelial glycocalyx, myocardial function, and vascular function in type 2 diabetic patients: one year follow-Up study. *J Clin Med* 2019; 8:.
187. Inoguchi T, Li P, Umeda F, Yu HY, Kakimoto M, Imamura M, *et al.* High glucose level and free fatty acid stimulate reactive oxygen species production through protein kinase C-dependent activation of NAD(P)H oxidase in cultured vascular cells. *Diabetes* 2000; 49:1939–1945.

188. Lambadiari V, Kousathana F, Raptis A, Katogiannis K, Kokkinos A, Ikonomidis I. preexisting cytokine and NLRP3 inflammasome activation and increased vascular permeability in diabetes: a possible fatal link with worst COVID-19 infection outcomes? *Front Immunol* 2020; 11:.
189. Semeraro F, Cancarini A, dell'Omo R, Rezzola S, Romano MR, Costagliola C. Diabetic retinopathy: vascular and inflammatory disease. *J Diabetes Res* 2015;582060.
190. Herdade AS, Silva IM, Calado Â, Saldanha C, Nguyen NH, Hou I, et al. Effects of diabetes on microcirculation and leukostasis in retinal and nonocular tissues: implications for diabetic retinopathy. *Biomolecules* 2020; 10:.
191. Sörensen BM, Houben AJ, Berendschot TT, Schouten JS, Kroon AA, van der Kallen CJ, et al. Prediabetes and type 2 diabetes are associated with generalized microvascular dysfunction: the Maastricht study. *Circulation* 2016; 134:1339–1552.
192. Hanssen H, Streese L, Vilser W. Retinal vessel diameters and function in cardiovascular risk and disease. *Prog Retin Eye Res* 2022; 91:.
193. Fuchs D, Dupon PP, Schaap LA, Draijer R. The association between diabetes and dermal microvascular dysfunction noninvasively assessed by laser Doppler with local thermal hyperemia: a systematic review with meta-analysis. *Cardiovasc Diabetol* 2017; 16:.
194. Ikonomidis I, Pavlidis G, Thymis J, Birba D, Kalogeris A, Kousathana F, et al. Effects of glucagon-like peptide-1 receptor agonists, sodium-glucose cotransporter-2 inhibitors, and their combination on endothelial glycocalyx, arterial function, and myocardial work index in patients with type 2 diabetes mellitus after 12-Month treatment. *J Am Heart Assoc* 2020; 9:e015716.
195. Torimoto K, Okada Y, Mori H, Tanaka Y. Relationship between fluctuations in glucose levels measured by continuous glucose monitoring and vascular endothelial dysfunction in type 2 diabetes mellitus. *Cardiovasc Diabetol* 2013; 12:1.
196. Tomovic K, Lazarevic J, Kocic G, Deljanin-Ilic M, Anderlueh M, Smelcerovic A. Mechanisms and pathways of anti-inflammatory activity of DPP-4 inhibitors in cardiovascular and renal protection. *Med Res Rev* 2019; 39:404–422.
197. Bray JJH, Foster-Davies H, Salem A, Hoole AL, Obaid DR, Halcox JPP, et al. Glucagon-like peptide-1 receptor agonists improve biomarkers of inflammation and oxidative stress: a systematic review and meta-analysis of randomised controlled trials. *Diabetes Obes Metab* 2021; 23:1806–1822.
198. Iannantuoni F, MdM A, Diaz-Morales N, Falcon R, Bañuls C, Abad-Jimenez Z, et al. The SGLT2 inhibitor empagliflozin ameliorates the inflammatory profile in type 2 diabetic patients and promotes an antioxidant response in leukocytes. *J Clin Med* 2019; 8:.
199. Lambadiari V, Thymis J, Kouretas D, Skaperda Z, Tekos F, Kousathana F, et al. Effects of a 12-month treatment with glucagon-like peptide-1 receptor agonists, sodium-glucose cotransporter-2 inhibitors, and their combination on oxidant and antioxidant biomarkers in patients with type 2 diabetes. *Antioxidants (Basel)* 2021; 10:.
200. Ott C, Jumar A, Striepe K, Friedrich S, Karg MV, Bramlage P, et al. A randomised study of the impact of the SGLT2 inhibitor dapagliflozin on microvascular and macrovascular circulation. *Cardiovasc Diabetol* 2017; 16:.
201. Ahmad E, Lim S, Lamptey R, Webb DR, Davies MJ. Type 2 diabetes. *Lancet* 2022; 400:1803–1820.
202. Triantafyllou A, Anyfanti P, Zabulis X, Gavriilaki E, Karamaounas P, Gkaliagkousi E, et al. Accumulation of microvascular target organ damage in newly diagnosed hypertensive patients. *J Am Soc Hypertens* 2014; 8:542–549.
203. Anyfanti P, Gkaliagkousi E, Triantafyllou A, Dipla K, Zarifis H, Arseniou P, et al. Noninvasive assessment of myocardial perfusion in different blood pressure phenotypes and its association with arterial stiffness indices. *Am J Hypertens* 2019; 32:557–563.
204. Lazaridis A, Triantafyllou A, Dipla K, Dolgyras P, Koletsos N, Anyfanti P, et al. Skin microvascular function, as assessed with laser speckle contrast imaging, is impaired in untreated essential and masked hypertension. *Hypertens Res* 2022; 45:445–454.
205. Dipla K, Triantafyllou A, Koletsos N, Papadopoulos S, Sachpekidis V, Vrabas IS, et al. Impaired muscle oxygenation and elevated exercise blood pressure in hypertensive patients: links with vascular stiffness. *Hypertension* 2017; 70:444–451.
206. Triantafyllou A, Doumas M, Anyfanti P, Gkaliagkousi E, Zabulis X, Petidis K, et al. Divergent retinal vascular abnormalities in normotensive persons and patients with never-treated, masked, white coat hypertension. *Am J Hypertens* 2013; 26:318–325.
207. Madhur MS, Lob HE, McCann LA, Iwakura Y, Blinder Y, Guzik TJ, et al. Interleukin 17 promotes angiotensin II-induced hypertension and vascular dysfunction. *Hypertension* 2010; 55:500–507.
208. Brands MW, Banes-Berceli AK, Inscho EW, Al-Azawi H, Allen AJ, Labazi H. Interleukin 6 knockout prevents angiotensin II hypertension: role of renal vasoconstriction and janus kinase 2/signal transducer and activator of transcription 3 activation. *Hypertension* 2010; 56:879–894.
209. Jayedi A, Rahimi K, Bautista LE, Nazarzadeh M, Zargar MS, Shab-Bidar S. Inflammation markers and risk of developing hypertension: a meta-analysis of cohort studies. *Heart* 2019; 105:686–692.
210. Zanolli L, Briet M, Empana JP, Cunha PG, Mäki-Petäjä KM, Protopogerou AD, et al. Vascular consequences of inflammation: a position statement from the ESH Working Group on Vascular Structure and Function and the ARTERY Society. *J Hypertens* 2020; 38:1682–1698.
211. Satou R, Gonzalez-Villalobos RA, Miyata K, Ohashi N, Urushihara M, Acres OW, et al. IL-6 augments angiotensinogen in primary cultured renal proximal tubular cells. *Mol Cell Endocrinol* 2009; 311:24–31.
212. Norlander AE, Saleh MA, Kamat NV, Ko B, Gnecco J, Zhu L, et al. Interleukin-17A regulates renal sodium transporters and renal injury in angiotensin II-induced hypertension. *Hypertension* 2016; 68:167–174.
213. Nguyen H, Chiasson VL, Chatterjee P, Kopriva SE, Young KJ, Mitchell BM. Interleukin-17 causes Rho-kinase-mediated endothelial dysfunction and hypertension. *Cardiovasc Res* 2013; 97:696–704.
214. Wu J, Thabet SR, Kirabo A, Trott DW, Saleh MA, Xiao L, et al. Inflammation and mechanical stretch promote aortic stiffening in hypertension through activation of p38 mitogen-activated protein kinase. *Circ Res* 2014; 114:626–625.
215. Harrison DG, Gongora MC. Oxidative stress and hypertension. *Med Clin North Am* 2009; 93:621–635.
216. Harrison DG, Coffman TM, Wilcox CS. Pathophysiology of hypertension: the mosaic theory and beyond. *Circ Res* 2021; 128:847–863.
217. Krishnan SM, Sobey CG, Latz E, Mansell A, Drummond GR. IL-1 β and IL-18: inflammatory markers or mediators of hypertension? *Br J Pharmacol* 2014; 171:5589–5602.
218. Drummond GR, Vinh A, Guzik TJ, Sobey CG. Immune mechanisms of hypertension. *Nat Rev Immunol* 2019; 19:517–532.
219. Nosalski R, Mikolajczyk T, Siedlinski M, Saju B, Koziol J, Maffia P, et al. Nox1/4 inhibition exacerbates age dependent perivascular inflammation and fibrosis in a model of spontaneous hypertension. *Pharmacol Res* 2020; 161:105235.
220. Nosalski R, Siedlinski M, Denby L, McGinnigle E, Nowak M, Cat AND, et al. T-cell-derived miRNA-214 mediates perivascular fibrosis in hypertension. *Circ Res* 2020; 126:988–1003.
221. Chrysanthopoulou A, Gkaliagkousi E, Lazaridis A, Arelaki S, Pateinakis P, Ntinopoulou M, et al. Angiotensin II triggers release of neutrophil extracellular traps, linking thromboinflammation with essential hypertension. *JCI Insight* 2021; 6:.
222. Carnevale D. Neural control of immunity in hypertension: council on hypertension mid career award for research excellence, 2019. *Hypertension* 2020; 76:622–628.
223. Grassi G. The sympathetic nervous system in hypertension: roadmap update of a long journey. *Am J Hypertens* 2021; 34:1247–1254.
224. Marvar PJ, Thabet SR, Guzik TJ, Lob HE, McCann LA, Weyand C, et al. Central and peripheral mechanisms of T-lymphocyte activation and vascular inflammation produced by angiotensin II-induced hypertension. *Circ Res* 2010; 107:263–270.
225. Werdan K. The activated immune system in congestive heart failure—from dropsy to the cytokine paradigm. *J Intern Med* 1998; 243:87–92.
226. Guzik TJ, Korbut R, Adamek-Guzik T. Nitric oxide and superoxide in inflammation and immune regulation. *J Physiol Pharmacol* 2003; 54:469–487.
227. Banks WA, Reed MJ, Logsdon AF, Rhea EM, Erickson MA. Healthy aging and the blood-brain barrier. *Nat Aging* 2021; 1:243–254.
228. Barisano G, Montagne A, Kisler K, Schneider JA, Wardlaw JM, Zlokovic BV. Blood-brain barrier link to human cognitive impairment and Alzheimer's disease. *Nat Cardiovasc Res* 2022; 1:108–115.
229. Evans LE, Taylor JL, Smith CJ, Pritchard HAT, Greenstein AS, Allan SM. Cardiovascular comorbidities, inflammation, and cerebral small vessel disease. *Cardiovasc Res* 2021; 117:2575–2588.

230. Zlokovic BV. The blood-brain barrier in health and chronic neurodegenerative disorders. *Neuron* 2008; 57:178–201.
231. Marnett E, Martello A, Caporali A. Autophagy at the interface of endothelial cell homeostasis and vascular disease. *FEBS J* 2022; 289:2976–2991.
232. Wardlaw JM, Smith C, Dichgans M. Small vessel disease: mechanisms and clinical implications. *Lancet Neurol* 2019; 18:684–696.
233. Rustenhoven J, Jansson D, Smyth LC, Dragunow M. Brain pericytes as mediators of neuroinflammation. *Trends Pharmacol Sci* 2017; 38:291–304.
234. McQuaid C, Montagne A. SARS-CoV-2 and vascular dysfunction: a growing role for pericytes. *Cardiovasc Res* 2022.
235. Zhang M, Mao Y, Ramirez SH, Tuma RF, Chabrashvili T. Angiotensin II induced cerebral microvascular inflammation and increased blood-brain barrier permeability via oxidative stress. *Neuroscience* 2010; 171:852–858.
236. Adamski MG, Sternak M, Mohaissen T, Kaczor D, Wieronska JM, Malinowska M, et al. Vascular cognitive impairment linked to brain endothelium inflammation in early stages of heart failure in mice. *J Am Heart Assoc* 2018; 7.
237. Khan MB, Hoda MN, Vaibhav K, Giri S, Wang P, Waller JL, et al. Remote ischemic postconditioning: harnessing endogenous protection in a murine model of vascular cognitive impairment. *Transl Stroke Res* 2015; 6:69–77.
238. Won JS, Kim J, Annamalai B, Shunmugavel A, Singh I, Singh AK. Protective role of S-nitrosoglutathione (GSNO) against cognitive impairment in rat model of chronic cerebral hypoperfusion. *J Alzheimers Dis* 2013; 34:621–635.
239. Rouhl RP, Damoiseaux JG, Lodder J, Theunissen RO, Knottnerus IL, Staals J, et al. Vascular inflammation in cerebral small vessel disease. *Neurobiol Aging* 2012; 33:1800–1806.
240. Mikolajczyk TP, Szczepaniak P, Vidler F, Maffia P, Graham GJ, Guzik TJ. Role of inflammatory chemokines in hypertension. *Pharmacol Ther* 2021; 223:107799.
241. Moreno M, Bannerman P, Ma J, Guo F, Miers L, Soulika AM, et al. Conditional ablation of astroglial CCL2 suppresses CNS accumulation of M1 macrophages and preserves axons in mice with MOG peptide EAE. *J Neurosci* 2014; 34:8175–8185.
242. Gimenez MA, Sim J, Archambault AS, Klein RS, Russell JH. A tumor necrosis factor receptor 1-dependent conversation between central nervous system-specific T cells and the central nervous system is required for inflammatory infiltration of the spinal cord. *Am J Pathol* 2006; 168:1200–1209.
243. Stuart MJ, Baune BT. Chemokines and chemokine receptors in mood disorders, schizophrenia, and cognitive impairment: a systematic review of biomarker studies. *Neurosci Biobehav Rev* 2014; 42:93–115.
244. Stranahan MA, Hao S, Dey A, Yu X, Baban B. Blood-brain barrier breakdown promotes macrophage infiltration and cognitive impairment in leptin receptor-deficient mice. *J Cereb Blood Flow Metab* 2016; 36:2108–2121.
245. Reboldi A, Coisne C, Baumjohann D, Benvenuto F, Bottinelli D, Lira S, et al. C-C chemokine receptor 6-regulated entry of TH-17 cells into the CNS through the choroid plexus is required for the initiation of EAE. *Nat Immunol* 2009; 10:514–523.
246. Kebir H, Kreymborg K, Ifergan I, Dodelet-Devillers A, Cayrol R, Bernard M, et al. Human TH17 lymphocytes promote blood-brain barrier disruption and central nervous system inflammation. *Nat Med* 2007; 13:1173–1175.
247. Murray EC, Nosalski R, MacRitchie N, Tomaszewski M, Maffia P, Harrison DG, et al. Therapeutic targeting of inflammation in hypertension: from novel mechanisms to translational perspective. *Cardiovasc Res* 2021; 117:2589–2609.
248. Rochford KD, Collins LE, McLoughlin A, Cummins PM. Tumour necrosis factor- α -mediated disruption of cerebrovascular endothelial barrier integrity in vitro involves the production of proinflammatory interleukin-6. *J Neurochem* 2016; 136:564–572.
249. Khan AO, Reyat JS, Hill H, Bourne JH, Colicchia M, Newby ML, et al. Preferential uptake of SARS-CoV-2 by pericytes potentiates vascular damage and permeability in an organoid model of the microvasculature. *Cardiovasc Res* 2022; 118:3085–3096.
250. Wang Y, Jin S, Sonobe Y, Cheng Y, Horiuchi H, Parajuli B, et al. Interleukin-1 β induces blood-brain barrier disruption by down-regulating Sonic hedgehog in astrocytes. *PLoS One* 2014; 9:e110024.
251. Liberale L, Diaz-Canestro C, Bonetti NR, Paneni F, Akhmedov A, Beer JH, et al. Postischemic administration of the murine Canakinumab-surrogate antibody improves outcome in experimental stroke. *Eur Heart J* 2018; 39:3511–3517.
252. Erta M, Quintana A, Hidalgo J. Interleukin-6, a major cytokine in the central nervous system. *Int J Biol Sci* 2012; 8:1254–1266.
253. Lyra e Silva NM, Gonçalves RA, Pascoal TA, Lima-Filho RAS, Resende EdPF, Vieira ELM, et al. Pro-inflammatory interleukin-6 signaling links cognitive impairments and peripheral metabolic alterations in Alzheimer's disease. *Transl Psychiatry* 2021; 11:251.
254. Hofer MJ, Campbell IL. Immunoinflammatory diseases of the central nervous system – the tale of two cytokines. *Br J Pharmacol* 2016; 173:716–728.
255. Nosalski R, McGinnigle E, Siedlinski M, Guzik TJ. Novel immune mechanisms in hypertension and cardiovascular risk. *Curr Cardiovasc Risk Rep* 2017; 11:12.
256. Querfurth HW, LaFerla FM. Alzheimer's disease. *N Engl J Med* 2010; 362:329–344.
257. Siedlinski M, Carnevale L, Xu X, Carnevale D, Evangelou E, Caulfield MJ, et al. Genetic analyses identify brain structures related to cognitive impairment associated with elevated blood pressure. *Eur Heart J* 2023.
258. Sole-Guardia G, Custers E, de Lange A, Clijncke E, Geenen B, Gutierrez J, et al. Association between hypertension and neurovascular inflammation in both normal-appearing white matter and white matter hyperintensities. *Acta Neuropathol Commun* 2023; 11:2.
259. Wardlaw JM, Benveniste H, Williams A. Cerebral vascular dysfunctions detected in human small vessel disease and implications for preclinical studies. *Annu Rev Physiol* 2022; 84:409–434.
260. Neves KB, Morris HE, Alves-Lopes R, Muir KW, Moreton F, Delles C, et al. Peripheral arteriopathy caused by Notch3 gain-of-function mutation involves ER and oxidative stress and blunting of NO/sGC/cGMP pathway. *Clin Sci (Lond)* 2021; 135:753–773.
261. Mizuno T, Mizuta I, Watanabe-Hosomi A, Mukai M, Koizumi T. Clinical and genetic aspects of CADASIL. *Front Aging Neurosci* 2020; 12:91.
262. Weaver JD, Huang MH, Albert M, Harris T, Rowe JW, Seeman TE. Interleukin-6 and risk of cognitive decline: MacArthur studies of successful aging. *Neurology* 2002; 59:371–378.
263. Kipinoinen T, Toppala S, Rinne JO, Viitanen MH, Jula AM, Ekblad LL. Association of midlife inflammatory markers with cognitive performance at 10-year follow-up. *Neurology* 2022; 99:e2294–e2302.
264. Wood LB, Winslow AR, Proctor EA, McGuone D, Mordes DA, Frosch MP, et al. Identification of neurotoxic cytokines by profiling Alzheimer's disease tissues and neuron culture viability screening. *Sci Rep* 2015; 5:16622.
265. Lee MH, Perl DP, Steiner J, Pasternack N, Li W, Maric D, et al. Neurovascular injury with complement activation and inflammation in COVID-19. *Brain* 2022; 145:2555–2568.
266. Guzik TJ, Mohiddin SA, Dimarco A, Patel V, Savvatis K, Marelli-Berg FM, et al. COVID-19 and the cardiovascular system: implications for risk assessment, diagnosis, and treatment options. *Cardiovasc Res* 2020; 116:1666–1687.
267. Tavares-Junior JW, de Souza ACC, Borges JWP, Oliveira DN, Siqueira-Neto JI, Sobreira-Neto MA, et al. COVID-19 associated cognitive impairment: a systematic review. *Cortex* 2022; 152:77–97.
268. Apaydin DC, Zakarauskas-Seth BI, Carnevale L, Apaydin O, Perrotta M, Carnevale R, et al. Interferon-gamma drives macrophage reprogramming, cerebrovascular remodeling, and cognitive dysfunction in a zebrafish and a mouse model of ion imbalance and pressure overload. *Cardiovasc Res* 2022; 119:1234–1249.
269. Carnovale C, Perrotta C, Baldelli S, Cattaneo D, Montrasio C, Barbieri SS, et al. Antihypertensive drugs and brain function: mechanisms underlying therapeutically beneficial and harmful neuropsychiatric effects. *Cardiovasc Res* 2022; 119:647–667.
270. Lembo G, Perrotta M. The neurology of hypertension: merging academic specialties to connect heart and brain pathophysiology. *Cardiovasc Res* 2021; 117:e70–e72.
271. Morrens M, Overloop C, Coppens V, Loots E, Van Den Noortgate M, Vandenameele S, et al. The relationship between immune and cognitive dysfunction in mood and psychotic disorder: a systematic review and a meta-analysis. *Mol Psychiatry* 2022; 27:3237–3246.
272. Pape K, Tamouza R, Leboyer M, Zipp F. Immunoneuropsychiatry – novel perspectives on brain disorders. *Nat Rev Neurol* 2019; 15:317–328.

273. Winklewski PJ, Radkowski M, Wszedybyl-Winklewska M, Demkow U. Brain inflammation and hypertension: the chicken or the egg? *J Neuroinflamm* 2015; 12:85.
274. Biancardi VC, Stern JE. Compromised blood-brain barrier permeability: novel mechanism by which circulating angiotensin II signals to sympathoexcitatory centres during hypertension. *J Physiol* 2016; 594:1591–1600.
275. Málkiewicz MA, Mállecki A, Toborek M, Szarmach A, Winklewski PJ. Substances of abuse and the blood brain barrier: Interactions with physical exercise. *Neurosci Biobehav Rev* 2020; 119:204–216.
276. Eisenmenger LB, Peret A, Famakin BM, Spahic A, Roberts GS, Bockholt JH, et al. Vascular contributions to Alzheimer's disease. *Transl Res* 2022; 254:41–53.
277. Zlokovic BV. Neurovascular pathways to neurodegeneration in Alzheimer's disease and other disorders. *Nat Rev Neurosci* 2011; 12:723–738.
278. Yoon SP, Grewal DS, Thompson AC, Polascik BW, Dunn C, Burke JR, et al. Retinal microvascular and neurodegenerative changes in Alzheimer's disease and mild cognitive impairment compared with control participants. *Ophthalmol Retina* 2019; 3:489–499.
279. Theuerle JD, Al-Fiadh AH, Amirul Islam FM, Patel SK, Burrell LM, Wong TY, et al. Impaired retinal microvascular function predicts long-term adverse events in patients with cardiovascular disease. *Cardiovasc Res* 2021; 117:1949–1957.
280. Restivo V, Candiloro S, Daidone M, Norrito R, Cataldi M, Minutolo G, et al. Systematic review and meta-analysis of cardiovascular risk in rheumatological disease: Symptomatic and nonsymptomatic events in rheumatoid arthritis and systemic lupus erythematosus. *Autoimmun Rev* 2022; 21:.
281. Hansson GK. Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med* 2005; 352:1685–1695.
282. Van Doornum S, McColl G, Wicks IP. Accelerated atherosclerosis: an extraarticular feature of rheumatoid arthritis? *Arthritis Rheum* 2002; 46:862–873.
283. Fischer LM, Schlienger RG, Matter C, Jick H, Meier CR. Effect of rheumatoid arthritis or systemic lupus erythematosus on the risk of first-time acute myocardial infarction. *Am J Cardiol* 2004; 93:198–200.
284. Suppiah R, Judge A, Batra R, Flossmann O, Harper L, Höglund P, et al. A model to predict cardiovascular events in patients with newly diagnosed Wegener's granulomatosis and microscopic polyangiitis. *Arthritis Care Res (Hoboken)* 2011; 63:588–596.
285. Bordy R, Tototos P, Prati C, Marie C, Wendling D, Demougeot C. Microvascular endothelial dysfunction in rheumatoid arthritis. *Nat Rev Rheumatol* 2018; 14:404–420.
286. Zanatta E, Colombo C, D'Amico G, d'Humières T, Dal Lin C, Tona F. Inflammation and coronary microvascular dysfunction in autoimmune rheumatic diseases. *Int J Mol Sci* 2019; 20:.
287. Alamanos Y, Voulgari PV, Drosos AA. Incidence and prevalence of rheumatoid arthritis, based on the 1987 American College of Rheumatology criteria: a systematic review. *Semin Arthritis Rheum* 2006; 36:182–188.
288. Aviña-Zubieta JA, Choi HK, Sadatsafavi M, Etmnan M, Esdaile JM, Lacaille D. Risk of cardiovascular mortality in patients with rheumatoid arthritis: a meta-analysis of observational studies. *Arthritis Rheum* 2008; 59:1690–1697.
289. Anyfanti P, Triantafyllou A, Gkaliagkousi E, Triantafyllou G, Koletsos N, Chatzimichaïlidou S, et al. Subendocardial viability ratio in patients with rheumatoid arthritis: comparison with healthy controls and identification of prognostic factors. *Clin Rheumatol* 2017; 36:1229–1236.
290. Van Doornum S, Strickland G, Kawasaki R, Xie J, Wicks IP, Hodgson LA, et al. Retinal vascular calibre is altered in patients with rheumatoid arthritis: a biomarker of disease activity and cardiovascular risk? *Rheumatology (Oxford)* 2011; 50:939–943.
291. Anyfanti P, Triantafyllou A, Gkaliagkousi E, Koletsos N, Athanasopoulos G, Zabalus X, et al. Retinal vessel morphology in rheumatoid arthritis: association with systemic inflammation, subclinical atherosclerosis, and cardiovascular risk. *Microcirculation* 2017; 24:.
292. Anyfanti P, Gkaliagkousi E, Triantafyllou A, Zabalus X, Dolgyras P, Galanopoulou V, et al. Dermal capillary rarefaction as a marker of microvascular damage in patients with rheumatoid arthritis: association with inflammation and disorders of the macrocirculation. *Microcirculation* 2018; 25:e12451.
293. Ikonomidis I, Lekakis JP, Nikolaou M, Paraskevaidis I, Andreadou I, Kaplanoglou T, et al. Inhibition of interleukin-1 by anakinra improves vascular and left ventricular function in patients with rheumatoid arthritis. *Circulation* 2008; 117:2662–2669.
294. Ikonomidis I, Pavlidis G, Katsimbri P, Lambadiari V, Parissis J, Andreadou I, et al. Tocilizumab improves oxidative stress and endothelial glycocalyx: a mechanism that may explain the effects of biological treatment on COVID-19. *Food Chem Toxicol* 2020; 145:111694.
295. Yki-Järvinen H, Bergholm R, Leirisalo-Repo M. Increased inflammatory activity parallels increased basal nitric oxide production and blunted response to nitric oxide in vivo in rheumatoid arthritis. *Ann Rheum Dis* 2003; 62:630–634.
296. Mäki-Petäjä KM, Cheriyan J, Booth AD, Hall FC, Brown J, Wallace SM, et al. Inducible nitric oxide synthase activity is increased in patients with rheumatoid arthritis and contributes to endothelial dysfunction. *Int J Cardiol* 2008; 129:399–405.
297. Arosio E, De Marchi S, Rigoni A, Prior M, Delva P, Lechi A. Forearm haemodynamics, arterial stiffness and microcirculatory reactivity in rheumatoid arthritis. *J Hypertens* 2007; 25:1273–1278.
298. Anyfanti P, Gavriilaki E, Dolgyras P, Nikolaidou B, Dimitriadou A, Lazaridis A, et al. Skin microcirculation dynamics are impaired in patients with rheumatoid arthritis and no cardiovascular comorbidities. *Clin Exp Rheumatol* 2023.
299. Koletsos N, Dipla K, Triantafyllou A, Lazaridis A, Papadopoulos NG, Dolgyras P, et al. Blunted cerebral oxygenation during exercise in systemic lupus erythematosus patients. *Clin Exp Rheumatol* 2023; 41:6–14.
300. Chen JJ, Yen RF, Kao A, Lin CC, Lee CC. Abnormal regional cerebral blood flow found by technetium-99m ethyl cysteinate dimer brain single photon emission computed tomography in systemic lupus erythematosus patients with normal brain MRI findings. *Clin Rheumatol* 2002; 21:516–519.
301. Postal M, Lapa AT, Reis F, Rittner L, Appenzeller S. Magnetic resonance imaging in neuropsychiatric systemic lupus erythematosus: current state of the art and novel approaches. *Lupus* 2017; 26:517–521.
302. Babaoğlu H, Baytaroğlu A, Torğutalp M, Erden A, Kadaşçıfçılar S, Kalyoncu U. Abnormal retinal microvasculature found in active rheumatoid arthritis: a different perspective of microvascular health. *Turk J Med Sci* 2019; 49:20–26.
303. Lee JH, Kim SS, Kim GT. Microvascular findings in patients with systemic lupus erythematosus assessed by fundus photography with fluorescein angiography. *Clin Exp Rheumatol* 2013; 31:871–876.
304. Manchanda AS, Kwan AC, Ishimori M, Thomson LEJ, Li D, Berman DS, et al. Coronary microvascular dysfunction in patients with systemic lupus erythematosus and chest pain. *Front Cardiovasc Med* 2022; 9:867155.
305. Weber BN, Stevens E, Barrett L, Bay C, Sinnette C, Brown JM, et al. Coronary microvascular dysfunction in systemic lupus erythematosus. *J Am Heart Assoc* 2021; 10:e018555.
306. Cutolo M, Melsens K, Wijnant S, Ingegnoli F, Thevissen K, De Keyser F, et al. Nailfold capillaroscopy in systemic lupus erythematosus: a systematic review and critical appraisal. *Autoimmun Rev* 2018; 17:344–352.
307. Taraborelli M, Sciatti E, Bonadei I, Terlizzi V, Fredi M, Zani R, et al. Endothelial dysfunction in early systemic lupus erythematosus patients and controls without previous cardiovascular events. *Arthritis Care Res (Hoboken)* 2018; 70:1277–1283.
308. Aizer J, Karlson EW, Chibnik LB, Costenbader KH, Post D, Liang MH, et al. A controlled comparison of brachial artery flow mediated dilation (FMD) and digital pulse amplitude tonometry (PAT) in the assessment of endothelial function in systemic lupus erythematosus. *Lupus* 2009; 18:235–242.
309. Svensson C, Eriksson P, Bjarnegård N, Jonasson H, Strömberg T, Sjöwall C, et al. Impaired microcirculation and vascular hemodynamics in relation to macrocirculation in patients with systemic lupus erythematosus. *Front Med (Lausanne)* 2021; 8:722758.
310. Koletsos N, Gkaliagkousi E, Lazaridis A, Triantafyllou A, Anyfanti P, Dolgyras P, et al. Skin microvascular dysfunction in systemic lupus erythematosus patients with and without cardiovascular risk factors. *Rheumatology (Oxford)* 2021; 60:2834–2841.
311. Ruaro B, Sulli A, Casabella A, Pizzorni C, Paolino S, Smith V, et al. Peripheral blood perfusion in patients with systemic lupus erythematosus and in primary Raynaud's phenomenon. *Eur J Rheumatol* 2021; 8:7–11.

312. Ahlehoff O, Gislason GH, Charlott M, Jørgensen CH, Lindhardsen J, Olesen JB, *et al.* Psoriasis is associated with clinically significant cardiovascular risk: a Danish nationwide cohort study. *J Intern Med* 2011; 270:147–157.
313. Atzeni F, Nucera V, Galloway J, Zoltán S, Nurmohamed M. Cardiovascular risk in ankylosing spondylitis and the effect of anti-TNF drugs: a narrative review. *Expert Opin Biol Ther* 2020; 20:517–524.
314. Dolgyras P, Lazaridis A, Anyfanti P, Gavriilaki E, Koletsos N, Triantafyllou A, *et al.* Microcirculation dynamics in systemic vasculitis: evidence of impaired microvascular response regardless of cardiovascular risk factors. *Rheumatology (Oxford)* 2022.
315. Osto E, Piaserico S, Maddalozzo A, Forchetti G, Montisci R, Famoso G, *et al.* Impaired coronary flow reserve in young patients affected by severe psoriasis. *Atherosclerosis* 2012; 221:113–117.
316. Triantafyllou C, Nikolaou M, Ikonomidis I, Bamias G, Kouretas D, Andreadou I, *et al.* Effects of anti-inflammatory treatment and surgical intervention on endothelial glycocalyx, peripheral and coronary microcirculatory function and myocardial deformation in inflammatory bowel disease patients: a two-arms two-stage clinical trial. *Diagnostics (Basel)* 2021; 11:.
317. Batko B, Maga P, Urbanski K, Ryszawa-Mrozek N, Schramm-Luc A, Koziej M, *et al.* Microvascular dysfunction in ankylosing spondylitis is associated with disease activity and is improved by anti-TNF treatment. *Sci Rep* 2018; 8:.
318. Margouta A, Anyfanti P, Lazaridis A, Nikolaidou B, Mastrogiannis K, Mallioura A, *et al.* Blunted microvascular reactivity in psoriasis patients in the absence of cardiovascular disease, as assessed by laser speckle contrast imaging. *Life (Basel)* 2022; 12:.
319. Ikonomidis I, Pavlidis G, Katsimbri P, Andreadou I, Triantafyllidi H, Tsoumani M, *et al.* Differential effects of inhibition of interleukin 1 and 6 on myocardial, coronary and vascular function. *Clin Res Cardiol* 2019; 108:1093–1101.
320. Ikonomidis I, Papadavid E, Makavos G, Andreadou I, Varoudi M, Gravani K, *et al.* Lowering interleukin-12 activity improves myocardial and vascular function compared with tumor necrosis factor- α antagonism or cyclosporine in psoriasis. *Circ Cardiovasc Imaging* 2017; 10:.
321. Lyon AR, López-Fernández T, Couch LS, Asteggiano R, Aznar MC, Bergler-Klein J, *et al.* 2022 ESC Guidelines on cardio-oncology developed in collaboration with the European Hematology Association (EHA), the European Society for Therapeutic Radiology and Oncology (ESTRO) and the International Cardio-Oncology Society (IC-OS). *Eur Heart J* 2022; 43:4229–4361.
322. Neves KB, Montezano AC, Lang NN, Touyz RM. Vascular toxicity associated with antiangiogenic drugs. *Clin Sci (Lond)* 2020; 134:2503–2520.
323. Lang NN, Touyz RM. Mechanistic science in cardiovascular-oncology: the way forward to maximise anticancer drug effects and minimise cardiovascular toxicity. *Clin Sci (Lond)* 2021; 135:2661–2663.
324. Caletti S, Paini A, Coschignano MA, De Ciuceis C, Nardin M, Zulli R, *et al.* Management of VEGF-targeted therapy-induced hypertension. *Curr Hypertens Rep* 2018; 20:68.
325. Rizzoni D, De Ciuceis C, Porteri E, Agabiti-Rosei C, Agabiti-Rosei E. Use of antihypertensive drugs in neoplastic patients. *High Blood Press Cardiovasc Prev* 2017; 24:127–132.
326. DeSantis CE, Lin CC, Mariotto AB, Siegel RL, Stein KD, Kramer JL, *et al.* Cancer treatment and survivorship statistics, 2014. *CA Cancer J Clin* 2014; 64:252–271.
327. Folkman J. Role of angiogenesis in tumor growth and metastasis. *Semin Oncol* 2002; 29 (Suppl 16):15–18.
328. Fukuda N, Takahari D, Wakatsuki T, Osumi H, Nakayama I, Matsushima T, *et al.* Early hypertension is associated with better clinical outcomes in gastric cancer patients treated with ramucirumab plus paclitaxel. *Oncotarget* 2018; 9:15219–15227.
329. Osumi H, Shinozaki E, Ooki A, Wakatsuki T, Kamiimabeppu D, Sato T, *et al.* Early hypertension and neutropenia are predictors of treatment efficacy in metastatic colorectal cancer patients administered FOLFIRI and vascular endothelial growth factor inhibitors as second-line chemotherapy. *Cancer Med* 2021; 10:615–625.
330. Hurwitz HI, Douglas PS, Middleton JP, Sledge GW, Johnson DH, Reardon DA, *et al.* Analysis of early hypertension and clinical outcome with bevacizumab: results from seven phase III studies. *Oncologist* 2013; 18:273–280.
331. Takahashi S. Vascular endothelial growth factor (VEGF), VEGF receptors and their inhibitors for antiangiogenic tumor therapy. *Biol Pharm Bull* 2011; 34:1785–1788.
332. Neves KB, Rios FJ, van der Mey L, Alves-Lopes R, Cameron AC, Volpe M, *et al.* VEGFR (vascular endothelial growth factor receptor) inhibition induces cardiovascular damage via redox-sensitive processes. *Hypertension* 2018; 71:638–647.
333. van Dorst DCH, Dobbin SJH, Neves KB, Herrmann J, Herrmann SM, Vermissen J, *et al.* Hypertension and prohypertensive antineoplastic therapies in cancer patients. *Circ Res* 2021; 128:1040–1061.
334. Neves KB, Rios FJ, Jones R, Evans TRJ, Montezano AC, Touyz RM. Microparticles from vascular endothelial growth factor pathway inhibitor-treated cancer patients mediate endothelial cell injury. *Cardiovasc Res* 2019; 115:978–988.
335. Steeghs N, Rabelink TJ, op 't Roodt J, Batman E, Cluitman FH, Weijl NI, *et al.* Reversibility of capillary density after discontinuation of bevacizumab treatment. *Ann Oncol* 2010; 21:1100–1105.
336. Mourad JJ, des Guetz G, Debbabi H, Levy BI. Blood pressure rise following angiogenesis inhibition by bevacizumab. A crucial role for microcirculation. *Ann Oncol* 2008; 19:927–934.
337. Steeghs N, Gelderblom H, Roodt JO, Christensen O, Rajagopalan P, Hovens M, *et al.* Hypertension and rarefaction during treatment with telatinib, a small molecule angiogenesis inhibitor. *Clin Cancer Res* 2008; 14:3470–3476.
338. Coschignano MA, De Ciuceis C, Agabiti-Rosei C, Brami V, Rossini C, Chiarini G, *et al.* Microvascular structural alterations in cancer patients treated with antiangiogenic drugs. *Front Cardiovasc Med* 2021; 8:651594.
339. van der Veldt AA, de Boer MP, Boven E, Eringa EC, van den Eertwegh AJ, van Hinsbergh VW, *et al.* Reduction in skin microvascular density and changes in vessel morphology in patients treated with sunitinib. *Anticancer Drugs* 2010; 21:439–446.
340. Dalbeni A, Ciccarese C, Bevilacqua M, Benati M, Caimmi C, Cerrito L, *et al.* Effects of antiangiogenic drugs on microcirculation and macrocirculation in patients with advanced-stage renal cancer. *Cancers (Basel)* 2018; 11.