

Citation for published version: Zoumpouli, GA, Zhang, Z, Wenk, J & Prasse, C 2021, 'Aqueous ozonation of furans: Kinetics and transformation mechanisms leading to the formation of  $\alpha$ , $\beta$ -unsaturated dicarbonyl compounds', *Water Research*, vol. 203, 117487. https://doi.org/10.1016/j.watres.2021.117487

DOI: 10.1016/j.watres.2021.117487

Publication date: 2021

Document Version Peer reviewed version

Link to publication

Publisher Rights CC BY-NC-ND

**University of Bath** 

## **Alternative formats**

If you require this document in an alternative format, please contact: openaccess@bath.ac.uk

**General rights** 

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy

If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

1	Aqueous Ozonation of Furans: Kinetics and Transformation Mechanisms
2	Leading to the Formation of $\alpha$ , $\beta$ -Unsaturated Dicarbonyl Compounds
3	
4	Garyfalia A. Zoumpouli <sup>a,b,c,d,1</sup> , Zhuoyue Zhang <sup>d</sup> , Jannis Wenk <sup>b,c</sup> , and Carsten Prasse <sup>d,e,*</sup>
5	
6 7	<sup>a</sup> Centre for Doctoral Training, Centre for Sustainable Chemical Technologies, University of Bath, Bath BA2 7AY, UK.
8 9	<sup>b</sup> Department of Chemical Engineering, University of Bath, Bath BA2 7AY, UK. <sup>c</sup> Water Innovation and Research Centre (WIRC), University of Bath, Bath BA2 7AY, UK.
10 11	<sup>d</sup> Department of Environmental Health and Engineering, Johns Hopkins University, Baltimore, MD 21218, USA.
12 13	<sup>e</sup> Risk Sciences and Public Policy Institute, Johns Hopkins Bloomberg School of Public Health, Baltimore, MD, United States
14 15 16	<sup>1</sup> Present address: Cranfield Water Science Institute, Cranfield University, College Road, Cranfield MK43 0AL, UK
17	Corresponding author
18	* To whom correspondence should be addressed:
19	Email: cprasse1@jhu.edu
20	Address: 3400 N Charles Street, Baltimore, MD 21218
21	
22	
23	
24	
25	
26	
27	
28	
29	
30	

#### 31 Abstract

Despite the widespread occurrence of furan moieties in synthetic and natural compounds, their 32 fate in aqueous ozonation has not been investigated in detail. Reaction rate constants of seven 33 commonly used furans with ozone were measured and ranged from  $k_{\rm O3}$  = 8.5  $\times$  10^4 to 3.2  $\times$ 34 10<sup>6</sup> M<sup>-1</sup> s<sup>-1</sup>, depending on the type and position of furan ring substituents. Transformation 35 product analysis of the reaction of furans with ozone focusing on the formation of toxic 36 organic electrophiles using a novel amino acid reactivity assay revealed the formation of  $\alpha$ , $\beta$ -37 unsaturated dicarbonyl compounds, 2-butene-1,4-dial (BDA) and its substituted analogues 38 (BDA-Rs). Their formation can be attributed to ozone attack at the reactive  $\alpha$ -C position 39 leading to furan ring opening. The molar yields of  $\alpha$ ,  $\beta$ -unsaturated dicarbonyl compounds 40 varied with the applied ozone concentration reaching maximum values of 7% for 2-furoic 41 42 acid. The identified  $\alpha$ , $\beta$ -unsaturated dicarbonyls are well-known toxicophores that are also formed by enzymatic oxidation of furans in the human body. In addition to providing data on 43 kinetics, transformation product analysis and proposed reaction mechanisms for the ozonation 44 45 of furans, this study raises concern about the presence of  $\alpha,\beta$ -unsaturated dicarbonyl compounds in water treatment and the resulting effects on human and environmental health. 46

47

## 48 Keywords: organic micropollutants, transformation products, reactivity-directed 49 analysis, dicarbonyls, toxic by-products

50

#### 51 <u>Highlights</u>

• Ozonation kinetics and transformation products of substituted furans were investigated

• High ozone reactivity of furans depending on type and position of substituents

• Identification of toxic  $\alpha$ , $\beta$ -unsaturated dicarbonyls using an amino acid reactivity assay

- Yields of toxic dicarbonyls are compound dependent and reached up to 7%
- 56

#### 57 **1. Introduction**

Furans are heterocyclic aromatics comprising a five-membered ring of four carbons and one 58 oxygen atom. The use of furan derivatives for the production of biomass-derived fuels, 59 polymers and other chemicals has dramatically increased over the last decades (Eldeeb and 60 Akih-Kumgeh, 2018; Gandini et al., 2016; Keay and Dibble, 1996; Tong et al., 2010). 61 Furfural, a commodity chemical and precursor of many other furans, has a global production 62 capacity of 280 kTon per year, with 65% used to produce furfuryl alcohol (Mariscal et al., 63 2016). In addition to their industrial applications, furans are common moieties in a variety of 64 naturally occurring compounds including terpenes and fatty acids, and can be formed 65 abiotically from the oxidation of natural organic matter (Hannemann et al., 1989; Huber et al., 66 67 2010; Wang et al., 2014). The extensive use of furan-containing chemicals and their natural occurrence make them likely contaminants in wastewater and drinking water resources as 68 69 evidenced by the detection of furan-containing compounds, particularly pharmaceuticals, in wastewater effluent and surface water (Jelic et al., 2011; Kasprzyk-Hordern et al., 2008; 70 71 Kostich et al., 2014).

Ozonation is increasingly used for the removal of trace organic contaminants in wastewater 72 73 treatment, wastewater reuse and drinking water production (Gottschalk et al., 2009). Ozone is a selective oxidant that primarily reacts with electron-rich moieties such as double bonds (von 74 Gunten, 2003; von Sonntag and von Gunten, 2012). Transformation products of the ozonation 75 76 of organic compounds include carbonyls formed by cleavage of olefinic bonds or benzene rings, N-oxides and hydroxylamines by oxidation of amines, and sulfoxides by oxidation of 77 thioethers (Hübner et al., 2015; Lee and von Gunten, 2016). The identification of ozone 78 79 transformation products with (eco)toxicological implications is of importance (von Gunten, 2018). For example, the main ozonation product of carboxy-acyclovir inhibits the growth of 80 81 green algae, an effect not observed for the parent compound (Schlüter-Vorberg et al., 2015). Similarly, embryotoxicity in a zebrafish assay was observed for the ozonation products of 82 83 carbamazepine while no effects were observed for carbamazepine itself (Pohl et al., 2020).

Despite extensive research on the reaction of ozone with several classes of organic compounds including olefins, phenols and nitrogen-containing compounds (Zoumpouli et al., 2020), studies focussing on the transformation of furans during aqueous ozonation are limited. The dimethylfuran moiety present in the antacid drug ranitidine has been shown to contribute to the high reactivity of this compound with ozone (Jeon et al., 2016). However, no transformation products that are specific for the reaction of ozone with the furan moiety were reported (Christophoridis et al., 2016; Zou et al., 2018). For the diuretic drug furosemide, two
ozonation products were identified indicating the potential relevance of cleavage and/or
opening of the furan ring by ozone (Aalizadeh et al., 2019).

Studies investigating the reaction of furans with ozone in organic solvents or organic 93 solvent/water mixtures suggest the potential involvement of different reaction mechanisms 94 (Bailey and Colomb, 1957; Bailey et al., 1965; Jibben and Wibaut, 1960; White et al., 1965). 95 Jibben and Wibaut (1960) identified glyoxal (a C<sub>2</sub> dicarbonyl) as the sole ozone 96 transformation product of furan and attributed its formation to the reaction of ozone with the 97 two carbon-carbon double bonds ( $\alpha$ - $\beta$  bonds) of the furan ring, leading to a C<sub>2</sub> dicarbonyl 98 containing both  $\beta$ -C atoms, and/or to  $\beta$ , $\beta$ -addition of ozone, leading to two C<sub>2</sub> dicarbonyls that 99 100 contain one  $\alpha$ - and one  $\beta$ -C atom of the furan ring (see Table 1 for nomenclature). In contrast, Bailey and Colomb (1957) and Bailey et al. (1965) observed the formation of  $\alpha$ , $\beta$ -unsaturated 101 dicarbonyl compounds containing all four carbons of the furan ring in experiments with 102 diarylfurans such as 2,5-diphenylfuran. The results of Bailey et al. indicate the relevance of 103 two distinct reaction pathways: (i) ozonolysis of a carbon-carbon double bond ( $\alpha$ - $\beta$  bond), and 104 (ii) electrophilic ozone attack at the reactive  $\alpha$ -C position in either a bidentate or monodentate 105 manner, followed by ring cleavage to form a C<sub>4</sub> dicarbonyl (Bailey, 1982). These C<sub>4</sub> 106 dicarbonyls then form lower-molecular weight transformation products through further 107 reaction with ozone (Bailey, 1982). 108

109 Given the absence of kinetic and mechanistic information on the ozonation of furans in aqueous solutions, the aim of this study was to determine the ozonation kinetics of various 110 commonly used furans and elucidate the formation of ozonation products in water. The 111 112 specific focus was on the formation of toxic  $\alpha,\beta$ -unsaturated dicarbonyl transformation products which have been recently identified as novel, highly-toxic by-products formed during 113 the oxidation of phenols with various oxidants including hydroxyl radicals and chlorine 114 (Prasse et al., 2018; Prasse et al., 2020). In addition,  $\alpha$ , $\beta$ -unsaturated dicarbonyls are also 115 formed during the enzymatic oxidation of furans in the human body (catalyzed by cytochrome 116 P450) and are responsible for their toxicity (Chen et al., 1995; Peterson, 2013; Ravindranath 117 et al., 1984). The studied furans included two high usage pharmaceuticals (furosemide, 118 ranitidine) that can be frequently found in the effluent of wastewater treatment plants (Kostich 119 120 et al., 2014), and seven high production volume industrial chemicals (furfuryl alcohol, 2-furoic 121 acid, 2,5-dimethylfuran, 2-methyl-3-furoic acid, 3-(2-furyl)propanoic acid, 3,4bis(hydroxymethyl)furan, furan-2,5-dicarboxylic acid) (Eldeeb and Akih-Kumgeh, 2018; 122

Gandini et al., 2016). Transformation product formation was followed using liquid chromatography-high resolution mass spectrometry (LC-HRMS). In addition, an amino acid reactivity assay was used to specifically assess the formation of  $\alpha$ ,β-unsaturated dicarbonyls that cannot be directly analysed with LC-HRMS due to the absence of ionizable groups in these molecules (Prasse et al., 2018; Prasse et al., 2020).

128

#### 129 2. Materials and Methods

#### 130 2.1 Chemicals

Furfuryl alcohol (FFA, CAS no.: 98-00-0), 3,4-bis(hydroxymethyl)furan (BHF, CAS no.: 131 14496-24-3), 2,5-dihydro-2,5-dimethoxyfuran (CAS no.: 332-77-4), 2,5-dimethylfuran 132 (DMF, CAS no.: 625-86-5) in liquid form and 2-furoic acid (FA, CAS no.: 98-00-0), 2-methyl-133 3-furoic acid (MFA, CAS no.: 98-00-0), 3-(2-furyl)propanoic acid (FPA, CAS no.: 935-13-134 7), furan-2,5-dicarboxylic acid (FDCA, CAS no.: 3238-40-2), furosemide (FRS, CAS no.: 54-135 31-9), ranitidine (RAN, CAS no.: 66357-59-3) in powder form were purchased from Sigma-136 Aldrich or Fisher Scientific in high purity ( $\geq 97\%$ ). *N*- $\alpha$ -acetyl-lysine (NAL) was from Sigma 137 Aldrich (>98% purity). *N*-α-acetyl-cysteine (NAC) was from Fisher Scientific (>98% purity). 138 Solvents for analysis, salts for preparation of buffers and tert-butanol were from Fisher 139 Scientific. All experimental and analytical solutions, including stock solutions, were prepared 140 in ultrapure water (resistivity > 18 M $\Omega$  cm<sup>-1</sup>) produced with a Milli-Q (Merck) or ELGA 141 (Veolia) water purification system. 142

143

#### 144 *2.2 Ozonation experiments*

Competition kinetics experiments were performed to determine the second order rate 145 constants for the reaction of furans with ozone in pure water buffered at pH 7 (10 mM 146 147 phosphate buffer, 10 mM tert-butanol). RAN was used as the reference compound, due to its known reaction rate constant with ozone (Jeon et al., 2016) and since initial tests had shown 148 that most of the target furans had an ozone reactivity within approximately one order of 149 magnitude of RAN. For compounds that reacted with ozone with much lower reaction rate 150 constants than RAN, FA was used as the reference compound, after determining its rate 151 constant using RAN. Further details on competition kinetics experiments and calculations are 152 provided in the SI, Supplementary text S1. 153

- Batch ozonation experiments to study the formation of transformation products of furans were
- 155 performed in 20-mL amber glass vials. The reaction solutions (10 mL) contained 15  $\mu$ M of

- the target compound and 10 mM phosphate buffer (pH 7), diluted with ultrapure water. After sampling the initial solution, a volume of concentrated ozone stock solution (see SI, Supplementary text S1) was added to achieve concentrations of 4 to 65  $\mu$ M ozone (0.3 to 4.3  $\mu$ M O<sub>3</sub>/ $\mu$ M target compound). The samples were left uncapped at room temperature for approximately 2 hours to achieve residual ozone depletion. To assess the influence of OH radical scavenging, a subset of experiments (Figures S2 and S3 in the SI) was also performed with addition of 10 mM *tert*-butanol (k<sub>OH</sub>, *tert*-butanol = 6 × 10<sup>8</sup> M<sup>-1</sup> s<sup>-1</sup>) (Buxton et al., 1988).
- Detection of  $\alpha,\beta$ -unsaturated dicarbonyl compounds was accomplished using an amino acid 163 reactivity assay, which is described in detail elsewhere (Prasse, 2021; Prasse et al., 2018; 164 Prasse et al., 2020). The reaction of NAL with  $\alpha,\beta$ -unsaturated dicarbonyls leads to the 165 formation of NAL adducts which can be detected using LC-HRMS (see section 2.3) (Prasse 166 167 et al., 2018). To this end, a small volume of a NAL stock solution was added to the samples (final concentration 150 µM, equivalent to 10 times the initial concentration of the parent 168 compound) followed by incubation at room temperature for 24 hours. Selected experiments 169 were repeated with higher concentration of the target compound (up to 100 µM) to facilitate 170 171 the identification of  $\alpha$ ,  $\beta$ -unsaturated dicarbonyl transformation products. Additionally, for some samples an equimolar mixture of NAL and NAC stock solutions was used instead of the 172 173 NAL stock solution, to enable the detection of dicarbonyls that do not form adducts with NAL alone but do form NAC or NAL+NAC adducts (Prasse et al., 2018). All samples were 174 analyzed within 48 hours. 175
- 176

#### 177 2.3 Analytical approaches

Spectrophotometric measurements to determine ozone concentrations were conducted in 1 cm
quartz glass cuvettes (Hellma) using a Cary 100 UV-VIS spectrophotometer (Agilent
Technologies), or in glass tubes using a DR/2000 Spectrophotometer (Hach).

181 Analysis of furans was performed using high-performance liquid chromatography with UV 182 detection (HPLC-UV). An overview of isocratic elution conditions, retention times and 183 detection wavelengths is provided in Table S1 of the SI. For batch ozonation a Vanquish 184 HPLC system with a DAD detector (Thermo Scientific) and an Acclaim RSLC 120 C18 185 column (5  $\mu$ m, 120 Å, 4.6 × 100 mm) was used. For competition kinetics a Dionex UltiMate 186 3000 system with a DAD detector (Thermo Scientific) and an Acclaim RSLC 120 C18 column 187 (2 - 120 Å 2 + 75 - 1)

187 (3  $\mu m,$  120 Å, 3  $\times$  75 mm) was used.

The formation of ozonation products and NAL, NAC or NAL+NAC adducts was determined 188 via LC-HRMS using an UltiMate 3000 UHPLC system coupled to a Q Exactive HF Orbitrap 189 MS (both Thermo Scientific). For chromatographic separation, a Phenomenex Synergi Hydro-190 RP column (4  $\mu$ m, 80 Å, 1 × 150 mm) was used. External mass calibration was performed 191 every 5 days using a calibration mixture similar to procedures described previously (Prasse et 192 al., 2011). More information on LC-HRMS analysis is provided in the SI, Supplementary text 193 S2. 194 2-butene-1,4-dial (BDA), the  $\alpha,\beta$ -unsaturated dicarbonyl identified in this work, was 195 quantified with standard addition calibration curves based on LC-HRMS analysis of its NAL 196

adduct, similar to a method described previously (Prasse et al., 2018). Stock solutions of BDA 197 198 (1 mM) were prepared through hydrolysis of 2,5-dihydro-2,5-dimethoxyfuran in ultrapure water at room temperature for at least 24 hours. For each experiment, standard addition was 199 applied on one of the samples and the slope of the curve was used for the other samples of that 200 experiment. The limit of detection of BDA in ultrapure water buffered at pH 7 was 1 nM 201 (lowest standard with a signal-to-noise ratio >3) and the limit of quantification was 10 nM 202 (lowest standard with signal-to-noise ratio >10). Ozonation yields of BDA were calculated by 203 dividing the molar concentration of BDA with the molar concentration of the parent 204 compound that reacted (difference between initial and final concentrations). Yields of other 205 BDA analogues (BDA-Rs) without a standard available were estimated using BDA as 206 207 reference standard.

208

#### **3. Results and discussion**

#### 210 *3.1 Kinetics of the reaction of substituted furans with ozone*

Table 1 shows the second order rate constants for the reaction of nine furans with ozone  $(k_{03})$ 211 212 in water at pH 7, including two values that were available in the literature. The competition kinetics plots for seven of the furans are provided in Figure S1. Initial tests indicated that the 213 214 studied furans have a high ozone reactivity, which was expected based on the aromaticity of the furan ring. Competition kinetics experiments showed that the k<sub>03</sub> of FPA, MFA, FRS, FFA 215 and BHF varies only by a factor of 2 (1.7 (± 0.2) to 3.2 (± 0.2) × 10<sup>6</sup> M<sup>-1</sup> s<sup>-1</sup>), while reaction 216 rates for FA and FDCA were lower (5.9 ( $\pm$  0.5)  $\times$  10<sup>5</sup> and 8.5 ( $\pm$  0.7)  $\times$  10<sup>4</sup> M<sup>-1</sup> s<sup>-1</sup>, 217 respectively). The ozone reactivity of most tested furans is comparable to that of phenols and 218 anilines at pH 7 (Lee and von Gunten, 2012). 219

- 221 Table 1. Second order rate constants for the reactions of furans with ozone in buffered water
- at pH 7. The  $\pm$  error of each rate constant was calculated through error propagation from the
- 223 95% confidence interval of the slope of the linear fit and the error of the rate constant of the
- 224 reference compound.

Compound	Structure	koз (M <sup>-1</sup> s <sup>-1</sup> ) at pH 7	Reference
Substituted furan	$\begin{array}{c} \mathbf{R_{5}} \stackrel{\alpha}{\underset{C}{\rightarrow}} \stackrel{\alpha}{\underset{C}{\rightarrow}} \mathbf{R_{2}} \\ \stackrel{\alpha}{\underset{C}{\rightarrow}} \stackrel{\alpha}{\underset{C}{\rightarrow}} \mathbf{R_{2}} \\ \stackrel{\alpha}{\underset{C}{\rightarrow}} \stackrel{\alpha}{\underset{C}{\rightarrow}} \mathbf{R_{3}} \end{array}$		
Ranitidine (RAN)		$2.1 \times 10^{6}$	Jeon et al. (2016)
2,5-Dimethylfuran (DMF)		$2.2 \times 10^5$	Jeon et al. (2016)
3-(2-Furyl)propanoic acid (FPA)	ОН	$(3.2 \pm 0.2)  imes 10^6$	this study
2-Methyl-3-furoic acid (MFA)	Он	$(2.7 \pm 0.1)  imes 10^{6}$	this study
Furosemide (FRS)	Cl O S-NH <sub>2</sub> HO	$(2.2 \pm 0.1)  imes 10^{6}$	this study
Furfuryl alcohol (FFA)	ОН	$(1.7 \pm 0.2) \times 10^{6}$	this study
3,4-Bis(hydroxymethyl)furan (BHF)	О ННО	$(1.7 \pm 0.1)  imes 10^{6}$	this study
2-Furoic acid (FA)	OH OH	$(5.9 \pm 0.5) \times 10^5$	this study
Furan-2,5-dicarboxylic acid (FDCA)	но он	$(8.5 \pm 0.7)  imes 10^4$	this study

Our results suggest that steric effects due to the presence of substituents do not play an important role. A potential explanation for the unsignificant steric hindrance effect has been proposed by Bailey et al. (1965) for the ozonation of diarylfurans. Therefore, the observed differences of reaction kinetics are most likely attributable to the type of substituents (e.g. electron-withdrawing versus electron-donating) and their positions (e.g. located at an  $\alpha$ -carbon versus at a  $\beta$ -carbon) on the furan ring. Electron-donating substituents such as hydroxyl and methyl groups increase the electron density of the furan ring and are therefore expected to

enhance its ozone reactivity, while electron-withdrawing groups such as carboxyl groups have 233 the opposite impact, similar to effects observed for phenols (Lee and von Gunten, 2012). The 234 three acids FPA, MFA and FA have pK<sub>a</sub> values ranging from 3 to 4.4 (Arena et al., 1993), 235 hence they are all dissociated at pH 7. The carboxylate group exerts an electro-donating effect 236 in contrast with the carboxyl (Hansch et al., 1991), as is also evidenced by the similar ozone 237 reactivity of FA and FFA, which has an electron-donating hydroxymethyl substituent. MFA 238 had a higher rate constant than FA, due to the presence of an additional alkyl group and/or the 239 presence of a carboxylate substituent at a  $\beta$ - rather than an  $\alpha$ -carbon. In FPA the carboxylate 240 241 group is separated from the furan ring by two additional carbons (C<sub>2</sub>H<sub>4</sub> group) compared to FA, leading to a 5-fold increase of the rate constant. Comparison of the kinetics of FDCA and 242 FA indicates that the presence of an additional carboxylate group (2,5-substitution of FDCA 243 versus 2-substitution of FA) lowers the ozone reactivity by approximately one order of 244 magnitude. The relatively low rate constant of DMF with ozone that has been reported in the 245 246 literature (Table 1) further indicates slower reaction kinetics for furans containing substituents at both  $\alpha$ -carbons (2,5-substitution). In contrast, BHF (3,4-substitution) had the same rate 247 248 constant as FFA (2-substitution), indicating that substituents located at  $\beta$ -carbons have a lower impact on the reaction rates. Further experiments with a more diverse group of furan 249 250 compounds are necessary to develop Quantitative Structure-Activity Relationships (QSARs) for substituted furans in oxidative water treatment processes similar to those that have been 251 developed for other compound classes such as phenols and amines (Lee et al., 2015; Lee and 252 von Gunten, 2012). 253

RAN contains multiple sites contributing to its high ozone reactivity: the furan ring, a tertiary 254 amine, a thioether and an acetamidine, which is the most reactive moiety (Jeon et al., 2016). 255 Similarly, the high rate constant of FRS can be attributed to both a furan ring and an aniline 256 moiety. Based on QSAR calculations, the  $k_{\rm O3}$  of FRS has been reported as  $6.8\times10^4~M^{-1}~s^{-1}$ 257 which was the sum of the contributions of the secondary amine ( $pK_a = 3.8$ ,  $k_{O3} = 6.2 \times 10^4$ 258  $M^{-1}$  s<sup>-1</sup>) and the benzene ring (partly deactivated,  $k_{03} = 6.5 \times 10^3 M^{-1} s^{-1}$ ) (Lee et al., 2014). 259 This predicted k<sub>03</sub> of FRS is similar to the experimentally determined reactivity of compounds 260 with a p-sulfonylaniline moiety (Dodd et al., 2006). The QSAR model, however, did not 261 consider the reactivity of the furan ring, which explains why the ko3 determined 262 experimentally for FRS in this study ( $k_{03} = 2.2 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$ ) is significantly higher than the 263 264 value predicted by QSAR.

#### 266 *3.2 Transformation of furans by ozone in water*

In addition to determining the ozonation kinetics of furans, the formation of transformation 267 products was investigated. Of particular interest was the formation of  $\alpha$ ,  $\beta$ -unsaturated 268 dicarbonyl compounds due to their potential toxicity and their recent identification in 269 oxidative water treatment processes (Prasse et al., 2018; Prasse et al., 2020). Ozonation 270 products were detected either directly with LC-HRMS, or after derivatization with either NAL 271 or a mixture of NAL and NAC (so called reactivity-directed analysis (RDA) assays) (Prasse, 272 2021). OH radicals were not scavenged in these experiments in order to represent real 273 ozonation conditions where both ozone and OH radicals are present. 274

Transformation of furan-containing pharmaceuticals. For FRS, seven ozonation products 275 were detected (Table S3 and Figure S11). The LC-HRMS results (exact mass, MS<sup>2</sup> spectra) 276 indicate that the benzene ring including the chlorine, sulfonamide and carboxyl moieties 277 remained unmodified in all ozonation products, which is in agreement with the deactivating 278 effect of these three benzene substituents. As such, oxidation of FRS can be attributed to the 279 reaction of ozone with the furfurylamine group, with FRS-278 being the only detected 280 compound that has been previously reported for the reaction with ozone (Aalizadeh et al., 281 2019). Based on the obtained results, the reaction of ozone with the  $\alpha$ -carbon of the furan 282 moiety is indicated to result in the opening of the furan ring (see section 3.3 for more details), 283 leading to the formation of an  $\alpha$ , $\beta$ -unsaturated dicarbonyl transformation product (FRS-347) 284 which has been observed previously in oxidation of FRS in microsomes (Williams et al., 285 2007). The presence of an  $\alpha,\beta$ -unsaturated dicarbonyl group is supported by the MS<sup>2</sup> spectrum 286 including the fragment m/z 262.9885 ( $C_8H_8O_4N_2ClS$ ), which corresponds to cleavage of a 287 288 C<sub>4</sub>H<sub>4</sub>O<sub>2</sub> moiety. Formation of FRS-328, which has been identified in the oxidation of FRS by dimethyldioxirane, can most likely be attributed to the intramolecular reaction between the 289 290 ketoenal group and the amine moiety of FRS-347 (Chen and Burka, 2007). The formation of a ring condensation product is also supported by the absence of MS<sup>2</sup> fragments that indicate 291 cleavage of carbon-containing moieties. FRS-328 is also formed as a product of anodic and 292 electro-Fenton oxidation of FRS (Laurencé et al., 2011; Laurence et al., 2014), and has been 293 294 identified as a human metabolite of FRS with evidence that it is a physio-pathologically relevant neurodegeneration inducer (Laurence et al., 2019). LC-HRMS results obtained for 295 296 FRS-265 show the presence of an additional methyl group compared to saluamine, an FRS hydrolysis product (Cruz et al., 1979; Laurence et al., 2014). The formation of FRS-265 can 297 298 be explained by cleavage of the substituent on the  $\alpha$ -carbon after furan ring opening. The

formation of the other transformation products can be explained by transformation of the furan
and secondary amine moieties, leading to carbonyls (FRS-308, FRS-363) and hydroxylamines
(FRS-266, FRS-308, FRS-363).

The chemical structures of the observed ozonation products suggest the relevance of two 302 reaction pathways involving the opening of the furan ring (Figure 1). The NAL assay was 303 used to assess whether  $\alpha$ , $\beta$ -unsaturated dicarbonyls (other than FRS-347) are formed from the 304 transformation of FRS. BDA was detected as a BDA-NAL adduct, indicating the relevance of 305  $\alpha,\beta$ -unsaturated dicarbonyl formation from the ozone oxidation of furan rings, even though 306 yields were low (< 0.1%). BDA and its substituted analogues have been identified as rat liver 307 microsomal metabolites of furan and furan containing compounds (Chen et al., 1995; 308 309 Peterson, 2013). The ozone dose-dependent formation of BDA and other FRS ozonation products is shown in Figure S12. 310



311

**Figure 1.** Proposed pathways of the ozonation of the furan ring of furosemide (FRS).

313

For RAN, twelve ozonation products were detected with two of them being formed by reaction 314 of ozone with the furan ring (Figures S13, S14, S15 and Table S4). Similar to results obtained 315 by Christophoridis et al. (2016), the LC-HRMS data indicate potential oxidation at different 316 positions of the molecule. However, in contrast to Christophoridis et al. (2016) who observed 317 only one ozonation product containing an additional oxygen atom (C13H22N4O4S) and 318 identified it as RAN-S oxide, two peaks with the same exact mass but distinct MS<sup>2</sup> spectra 319 were detected in the present study (Figure S14a and S14b). For the first peak (retention time: 320 3.7 min), the presence of MS<sup>2</sup> fragments m/z 188.0738 (C<sub>8</sub>H<sub>14</sub>O<sub>2</sub>NS) and 192.0435 321 (C<sub>5</sub>H<sub>10</sub>O<sub>3</sub>N<sub>3</sub>S) indicates the oxidation of the thioether and thus the formation of the RAN-S 322

oxide. In contrast, the absence of both fragments in the MS<sup>2</sup> spectrum of the second peak 323 (retention time: 8.9 min) and the detection of fragment 270.0902 (C<sub>11</sub>H<sub>16</sub>O<sub>3</sub>N<sub>3</sub>S), which is 324 formed via cleavage of a C<sub>2</sub>H<sub>7</sub>ON group, suggests the oxidation of the tertiary amine group 325 and thus the formation of the RAN-N oxide. The formation of both N- and S-oxides is further 326 supported by the detection of RAN-S&N oxide (C13H22N4O5S) and the MS<sup>2</sup> results for this 327 compound (Figure S14c). The formation of other products also indicates the oxidation of the 328 tertiary amine group (Figure S14e and S14h). However, due to the absence of reference 329 standards, no final conclusions can be drawn. Transformation products formed during the 330 331 electrochemical oxidation of RAN have been shown to be more toxic than the parent compound (Olvera-Vargas et al., 2014), emphasizing the need to elucidate the properties of 332 and the risk posed by the ozonation products of RAN. 333

Although all the RAN sub-structures react with ozone with high rates (Jeon et al., 2016), the 334 obtained results primarily demonstrated the formation of ozonation products in which the 335 furan ring remains unmodified. The detection of RAN-252 and RAN-236 also indicated the 336 oxidation of the furan moiety, leading to cleavage of parts of the molecule (Figure S14i and 337 S14j). However, it is possible that more ozonation products resulting from oxidation of the 338 furan ring were formed but could not be detected by LC-HRMS analysis. No  $\alpha$ , $\beta$ -unsaturated 339 dicarbonyl products were detected directly or after derivatization by NAL or a NAL+NAC 340 mixture, therefore dicarbonyls are either not formed from RAN or are degraded further. 341

342

Ozonation products of substituted furans. Based on the results of the furan-containing 343 pharmaceuticals, the formation of  $\alpha$ , $\beta$ -unsaturated dicarbonyls (BDA and BDA-Rs) from 344 simpler substituted furans was investigated to determine how different substituents impact the 345 formation of these toxic by-products. The results for seven tested compounds are summarised 346 347 in Table 2 and details are provided in the SI (Table S2 and Figures S4-S10). Concentrations of BDA were determined using a reference standard. Due to the absence of reference 348 standards, the yields of BDA-Rs were determined by comparing the LC-HRMS peak areas of 349 their NAL adducts with those obtained for BDA. 350

- 351
- 352
- 353
- 354

Compound	_	Substitu		Dicarbonyl	Max.	
Compound	$\mathbf{R}_2$	R <sub>3</sub>	<b>R</b> 4	<b>R</b> 5	formed	yield (%)
DMF	-CH <sub>3</sub>	-H	-H	-CH <sub>3</sub>	$BDA(R_2)(R_5)$	Not quantified
FΡΔ		п	-H	п	BDA	< 0.1
IIA	-C2H4COOH	-П		-П	BDA-R <sub>2</sub>	2.7
MFA	-CH <sub>3</sub>	-COOH	-H	-H	$BDA(R_2)(R_3)$	5.6
FEA		TT	п	TT	BDA	2.4
	-Сп2Оп	-П	-Π	-Π	BDA-R <sub>2</sub>	0.5
BHF	-H	-CH <sub>2</sub> OH	-CH <sub>2</sub> OH	-H	$BDA(R_3)(R_4)$	< 0.1
FA	-COOH	-H	-H	-H	BDA	6.7
FDCA	-COOH	-H	-H	-COOH	_	_

**Table 2.** Maximum yield of  $\alpha$ , $\beta$ -unsaturated dicarbonyls in the aqueous ozonation of substituted furans, based on the detection of NAL, NAC and NAL+NAC adducts.

358 The yields of BDA and BDA analogues were strongly dependent on the substituents present in different furans. Ozonation of FFA led to the formation of BDA at a maximum molar yield 359 of 2.4 % (Figure 2). This is comparable to the BDA yields formed from UV/H<sub>2</sub>O<sub>2</sub> oxidation 360 of phenol in water (Prasse et al., 2018). Traces of BDA were also detected in the reaction 361 solutions before the addition of ozone. This indicates the potential formation of BDA via 362 hydrolysis of FFA, which aligns with previous reports on the acid-catalyzed hydrolysis of 363 furans (Stamhuis et al., 1964). Besides BDA, a second C4-dicarbonyl compound containing 364 an additional hydroxymethyl group (NAL adduct C<sub>13</sub>H<sub>21</sub>O<sub>5</sub>N<sub>2</sub>, m/z 285.1444) was identified 365 in experiments with FFA (BDA-R in Figure 2). The maximum relative yield of this compound 366 was approximately 0.5%. The MS<sup>2</sup> spectrum of this adduct (Figure S5) contained 367 characteristic masses (m/z 84.0813 and 126.0914) previously observed for NAL adducts of 368 other dicarbonyls (Prasse et al., 2018). Ozonation of BHF did not lead to BDA formation, 369 despite the structural similarity of BHF and FFA. However, the formation of a NAL adduct 370 with m/z 315.1548 was detected in trace amounts, which can be attributed to the formation of 371 a dialdehyde containing two hydroxymethyl substituents (Figure S6). 372



**Figure 2.** A. Chemical structures of furfuryl alcohol (FFA) and its dicarbonyl ozonation products based on the formation of NAL adducts. B. Concentration of FFA and 2-butene-1,4dial (BDA) versus the ozone concentration. C. Molar yield of BDA and hydroxymethyl-BDA (BDA-R) determined by standard addition using a BDA reference standard. Conditions: FFA initial concentration 15  $\mu$ M, in 10 mM phosphate buffer at pH 7.

373

BDA was also identified as an ozonation product of FA, at higher molar yields of 380 approximately 7%, which was the maximum yield observed in this study. In comparison, 381 382 chlorination of phenols has been shown to result in the formation of BDA-Rs at yields ranging from 18% to 46% (Prasse et al., 2020). No other NAL adducts were detected in FA 383 experiments. For MFA, a BDA analogue with a carboxyl and a methyl group attached was 384 detected (Figures 3 and S8), while ozonation of FPA led to formation of both BDA and a 385 dicarbonyl with a propanoic acid group attached (Figures 3 and S7). The BDA molar yield 386 was less than 0.1% in the case of FPA, while the propanoic acid-substituted BDA analogue 387 appeared to be a more important ozonation product with a maximum yield of 2.7%. No NAL 388 or NAC adducts were detected in ozonation of FDCA, in agreement with the results observed 389 390 for RAN, indicating that the presence of two carboxyl substituents impacts both the reaction kinetics and the ozonation pathway. 391



392

**Figure 3.** Molar yields of three  $\alpha$ , $\beta$ -unsaturated dicarbonyls formed in experiments with furancontaining acids at different ozone concentrations. Conditions: furan acid initial concentration 15  $\mu$ M, in 10 mM phosphate buffer at pH 7. Yields were determined by standard addition using a 2-butene-1,4-dial (BDA) reference standard for all three compounds. For MFA, the ionization fragment m/z 269 was used for calculation of yields due to higher intensity.

The absence of a dimethylated BDA analogue in experiments with DMF can most likely be 399 explained by the inability of this compound to react with NAL in the same way as the other 400 dicarbonyl compounds detected, due to the presence of methyl substituents at both  $\alpha$ -carbons. 401 To verify this, additional experiments in the presence of both NAL and NAC were performed 402 and revealed the formation of both NAC and NAL+NAC adducts (Figures S9 and S10). In 403 contrast to NAL which primarily reacts with α,β-unsaturated dicarbonyl compounds via Schiff 404 base formation (i.e. reaction at the carbonyl carbon), reactions of thiols can be attributed to 405 Michael addition (i.e. reaction at the double bond adjacent to the carbonyl group) (LoPachin 406 and Gavin, 2014). The formed thiol adducts can then react in a second step with NAL yielding 407 pyrrole products (Figure S16). 408

The results demonstrate that  $\alpha$ ,β-unsaturated dicarbonyl compounds are relevant ozonation products of furans. The yields are generally low (< 7%), thus indicating the simultaneous formation of other ozonation products. In addition, the results show that BDA and BDA analogues can be transformed further by ozone (Figure S3). In the gas phase, BDA reacts with ozone with a rate constant of  $1.6 \times 10^{-18}$  cm<sup>3</sup> molecule<sup>-1</sup> s<sup>-1</sup> (Liu et al., 1999). Based on gasphase ozonation studies of BDA and other related compounds, the products formed from the further oxidation of BDA include formaldehyde, glyoxal and methylglyoxal (Liu et al., 1999;
Tuazon et al., 1985). These were not analysed in this study, but are the subject of ongoing
investigations.

418

419 3.3 Postulated mechanism for the reaction of furans with ozone leading to α,β-unsaturated
420 dicarbonyls

Even though no information is available about the transformation of furans by ozone in water, 421 previous studies performed in organic solvents have suggested the potential contribution of 422 different reaction mechanisms leading to opening of the furan ring (Bailey and Colomb, 1957; 423 Bailey et al., 1965; Jibben and Wibaut, 1960). Our detection of C4 dicarbonyls (BDA 424 analogues) confirms the importance of electrophilic ozone attack at the reactive  $\alpha$ -C positions 425 of the furan ring, via reaction of ozone with either one or both  $\alpha$ -carbons (Figure 4) (Bailey, 426 1982). The yields of BDA and substituted BDA analogues, however, suggest ozonolysis of 427 furans via reaction with the  $\alpha$ - $\beta$  double bonds as dominant reaction pathway and/or further 428 429 reactions of the C<sub>4</sub> dicarbonyls with ozone.



430

Figure 4. Postulated mechanism for the reaction of furans with ozone leading to formation of
2-butene-1,4-dial (BDA) and its analogues (BDA-Rs).

433

Similar to reaction kinetics, the obtained results further indicate that the yield and type of the formed  $\alpha,\beta$ -unsaturated dicarbonyls strongly depend on the substituents of the parent compound and their position on the furan ring. Two of the tested compounds, MFA and BHF, have substituents on the  $\beta$ -carbon of the furan ring (labelled as R<sub>3</sub> and R<sub>4</sub> in Tables 1 and 2). Both the carboxyl group of MFA and the hydroxymethyl groups of BHF were retained on the formed dicarbonyl compounds after ring opening (Table 2). The results of furans containing substituents on the  $\alpha$ -carbon (labelled as R<sub>2</sub> and R<sub>5</sub> in Tables 1 and 2) are less consistent. For

the ozonation of 2,5-diarylfurans in organic solvents, dicarbonyls containing aryl substituents 441 on both carbonyl carbons have been reported (Bailey et al., 1965; White et al., 1965). As 442 demonstrated in this study, a similar mechanism is also relevant under aqueous conditions for 443 MFA, FPA and DMF, all of which formed dicarbonyls with their  $\alpha$ -C substituents still attached 444 (Table 2). This indicates that the reaction of ozone with furans containing alkyl substituents 445 446 on the  $\alpha$ -carbon also results in the formation of BDA analogues with the substituents retained. In contrast, results obtained for furans containing either hydroxymethyl or carboxylic acid 447 substituents at one of the  $\alpha$ -carbons, indicate the relevance of reactions leading to cleavage of 448 449 the substituent and formation of BDA. This is particularly true for FA for which only the formation of BDA but not BDA-R was observed. The differences in yield of BDA versus 450 BDA-R for FA, FFA and FPA reveal the significant influence of these α-C substituents on the 451 mechanism of BDA formation. FA is predominantly present as a carboxylate anion (pKa 3.0) 452 (Arena et al., 1993) under neutral pH, which is an electron-donating group, while the aldehyde 453 and alkene groups are electron-withdrawing. Thus, one of the potential explanations for BDA 454 formation during aqueous ozonation is through hydrolysis of the BDA-carboxylate yielding 455 456 (E)-4-hydroxybuta-1,3-dien-1-one as intermediate, which is unstable (Lin and Huang, 2018; Yoshimi et al., 2010), and reacts further to BDA (Figure S17). Similar reaction mechanisms 457 458 leading to the removal of the hydroxymethyl and propanoic acid anion group can also be postulated for FFA and FPA, respectively (Figure S17). Furthermore, the descending trend of 459 electron donating capacity from carboxylate anion to hydroxymethyl and propanoic acid 460 groups is in good agreement with the decreasing yield of BDA versus BDA-Rs. Differences 461 in the degradation of FFA and BDA in experiments performed in the presence and absence of 462 tert-butanol as a OH radical scavenger (Figure S3) were minor. However, the presence of tert-463 butanol appeared to have some effect on the formed concentration of BDA and BDA-R 464 (Figure S2). The increased formation in the absence of tert-butanol indicates that BDA 465 analogues can be formed both from reactions with ozone and with OH radicals. 466

467

#### 468 **4.** Conclusions

The selected organic compounds containing furan rings have a high ozone reactivity that is comparable to that of activated benzene rings, and are therefore expected to be efficiently eliminated in water and wastewater ozonation treatment. Further research is required to elucidate the effect of deactivating substituents, such as halogens, on the ozonation rate constant of furans. In complex water matrices containing various furan-bearing compounds, ozonation is likely to result in the formation of a mixture of  $\alpha$ , $\beta$ -unsaturated dicarbonyls. Depending on the applied ozone dose, the dicarbonyls may decompose into smaller aldehydes and carboxylic acids. Future studies will focus on the detection of these further transformation products in real water treatment systems. In addition, it needs to be assessed whether  $\alpha$ , $\beta$ -unsaturated dicarbonyls can be removed during post-treatment steps, for example activated carbon and biofiltration.

The formation of  $\alpha$ , $\beta$ -unsaturated dicarbonyls such as BDA and its analogues, though only representing a small portion of transformation products from ozonation of furans, is a health concern due to their reported toxicity. Furans play an increasing role as 'green chemicals' and are also formed by natural processes in the aquatic environment. The obtained results highlight the necessity to investigate the fate of these compounds in water treatment systems to assess the potential exposures to toxic by-products.

487

#### 488 Acknowledgements

ZZ and GAZ contributed equally to this study. GAZ was supported by a University of Bath
(UoB) research scholarship and an EPSRC funded Integrated PhD studentship in Sustainable

491 Chemical Technologies: EP/L016354/1. Additional funding for the research visit of GAZ at

492 Johns Hopkins University was provided by the UoB Doctoral College Placement Support

493 Fund. We thank Urs von Gunten for insightful comments on the reaction mechanisms and

494 Nadezda Ojeda for technical assistance with the ozonation experiments.

#### 495 References

- Aalizadeh, R., Nika, M.C. and Thomaidis, N.S. (2019) Development and
  application of retention time prediction models in the suspect and non-target
- screening of emerging contaminants. J Hazard Mater 363, 277-285.
- 499 Arena, G., Cali, R., Maccarone, E. and Passerini, A. (1993) Thermodynamics of
- 500 protonation of some five-membered heteroaryl-carboxylates, -alkanoates and -
- trans-propenoates. Journal of the Chemical Society, Perkin Transactions 2 (10),
  1941-1945.
- Bailey, P.S. (1982) Ozonation in Organic Chemistry. Bailey, P.S. (ed), pp. 111154, Academic Press.
- 505 Bailey, P.S. and Colomb, H.O. (1957) 1,4-ADDITION OF OZONE TO
- 506 FURANS AND PYRROLES. Journal of the American Chemical Society 507 79(15), 4238-4238.
- Bailey, P.S., White, H.M. and Colomb, H.O. (1965) Ozonation of Diarylfurans.
  The Journal of Organic Chemistry 30(2), 487-491.
- Buxton, G.V., Greenstock, C.L., Helman, W.P. and Ross, A.B. (1988) Critical
  Review of rate constants for reactions of hydrated electrons, hydrogen atoms
  and hydroxyl radicals (·OH/·O– in Aqueous Solution. Journal of Physical and
- 513 Chemical Reference Data 17(2), 513-886.
- Chen, L.-J. and Burka, L.T. (2007) Chemical and Enzymatic Oxidation of
  Furosemide: Formation of Pyridinium Salts. Chemical Research in Toxicology
  20(12), 1741-1744.
- 517 Chen, L.-J., Hecht, S.S. and Peterson, L.A. (1995) Identification of cis-2-
- 518 Butene-1,4-dial as a Microsomal Metabolite of Furan. Chemical Research in
- 519 Toxicology 8(7), 903-906.
- 520 Christophoridis, C., Nika, M.C., Aalizadeh, R. and Thomaidis, N.S. (2016)
- 521 Ozonation of ranitidine: Effect of experimental parameters and identification of
- transformation products. Sci Total Environ 557-558, 170-182.
- 523 Cruz, J.E., Maness, D.D. and Yakatan, G.J. (1979) Kinetics and mechanism of
- 524 hydrolysis of furosemide. International Journal of Pharmaceutics 2(5), 275-281.
- 525 Dodd, M.C., Buffle, M.-O. and von Gunten, U. (2006) Oxidation of
- 526 Antibacterial Molecules by Aqueous Ozone: Moiety-Specific Reaction

- 527 Kinetics and Application to Ozone-Based Wastewater Treatment.
  528 Environmental Science & Technology 40(6), 1969-1977.
- Eldeeb, M. and Akih-Kumgeh, B. (2018) Recent Trends in the Production,
  Combustion and Modeling of Furan-Based Fuels. Energies 11(3), 512.
- 531 Gandini, A., Lacerda, T.M., Carvalho, A.J.F. and Trovatti, E. (2016) Progress
- 532 of Polymers from Renewable Resources: Furans, Vegetable Oils, and
- 533 Polysaccharides. Chemical Reviews 116(3), 1637-1669.
- 534 Gottschalk, C., Libra, J.A. and Saupe, A. (2009) Ozonation of water and waste
- water: A practical guide to understanding ozone and its applications, John Wiley& Sons.
- 537 Hannemann, K., Puchta, V., Simon, E., Ziegler, H., Ziegler, G. and Spiteller, G.
- (1989) The common occurrence of furan fatty acids in plants. Lipids 24(4), 296-298.
- Hansch, C., Leo, A. and Taft, R.W. (1991) A survey of Hammett substituent
  constants and resonance and field parameters. Chemical Reviews 91(2), 165195.
- Huber, S.G., Wunderlich, S., Schöler, H.F. and Williams, J. (2010) Natural
  Abiotic Formation of Furans in Soil. Environmental Science & Technology
  44(15), 5799-5804.
- 546 Hübner, U., von Gunten, U. and Jekel, M. (2015) Evaluation of the persistence
- of transformation products from ozonation of trace organic compounds A
  critical review. Water Research 68, 150-170.
- Jelic, A., Gros, M., Ginebreda, A., Cespedes-Sánchez, R., Ventura, F., Petrovic,
  M. and Barcelo, D. (2011) Occurrence, partition and removal of
  pharmaceuticals in sewage water and sludge during wastewater treatment.
  Water Research 45(3), 1165-1176.
- Jeon, D., Kim, J., Shin, J., Hidayat, Z.R., Na, S. and Lee, Y. (2016)
  Transformation of ranitidine during water chlorination and ozonation: Moietyspecific reaction kinetics and elimination efficiency of NDMA formation
- potential. Journal of Hazardous Materials 318, 802-809.

- Jibben, B.P. and Wibaut, J.P. (1960) The ozonisation and the ozonolysis of furanand of some methylated furans in connection with the reactivities of the bonds
- in the ringsystem. Recueil des Travaux Chimiques des Pays-Bas 79(4), 342-360.
- 560 Kasprzyk-Hordern, B., Dinsdale, R.M. and Guwy, A.J. (2008) The occurrence
- of pharmaceuticals, personal care products, endocrine disruptors and illicit drugs in surface water in South Wales, UK. Water Research 42(13), 3498-3518.
- Keay, B.A. and Dibble, P.W. (1996) Comprehensive Heterocyclic Chemistry II.
  Katritzky, A.R., Rees, C.W. and Scriven, E.F.V. (eds), pp. 395-436, Pergamon,
- 564 Katritzky, A.R., Rees, C.W. and Scriven, E.F.V565 Oxford.
- Kostich, M.S., Batt, A.L. and Lazorchak, J.M. (2014) Concentrations of
  prioritized pharmaceuticals in effluents from 50 large wastewater treatment
  plants in the US and implications for risk estimation. Environ Pollut 184, 354359.
- 570 Laurencé, C., Rivard, M., Lachaise, I., Bensemhoun, J. and Martens, T. (2011)
- 571 Preparative access to transformation products (TPs) of furosemide: a versatile 572 application of anodic oxidation. Tetrahedron 67(49), 9518-9521.
- Laurence, C., Rivard, M., Martens, T., Morin, C., Buisson, D., Bourcier, S.,
  Sablier, M. and Oturan, M.A. (2014) Anticipating the fate and impact of organic
  environmental contaminants: a new approach applied to the pharmaceutical
  furosemide. Chemosphere 113, 193-199.
- 577 Laurence, C., Zeghbib, N., Rivard, M., Lehri-Boufala, S., Lachaise, I., Barau,
- C., Le Corvoisier, P., Martens, T., Garrigue-Antar, L. and Morin, C. (2019) A
  new human pyridinium metabolite of furosemide, inhibitor of mitochondrial
  complex I, is a candidate inducer of neurodegeneration. Biochemical
  Pharmacology 160, 14-23.
- Lee, M., Zimmermann-Steffens, S.G., Arey, J.S., Fenner, K. and von Gunten,
- 583 U. (2015) Development of Prediction Models for the Reactivity of Organic
  584 Compounds with Ozone in Aqueous Solution by Quantum Chemical
  585 Calculations: The Role of Delocalized and Localized Molecular Orbitals.
- 586 Environmental Science & Technology 49(16), 9925-9935.
- 587 Lee, Y., Kovalova, L., McArdell, C.S. and von Gunten, U. (2014) Prediction of
- 588 micropollutant elimination during ozonation of a hospital wastewater effluent.
- 589 Water Research 64, 134-148.

- 590 Lee, Y. and von Gunten, U. (2012) Quantitative structure–activity relationships
- 591 (QSARs) for the transformation of organic micropollutants during oxidative
- 592 water treatment. Water Research 46(19), 6177-6195.
- 593 Lee, Y. and von Gunten, U. (2016) Advances in predicting organic contaminant

abatement during ozonation of municipal wastewater effluent: reaction kinetics,

- 595 transformation products, and changes of biological effects. Environmental
- 596 Science: Water Research & Technology 2(3), 421-442.
- Lin, D.-Z. and Huang, J.-M. (2018) Electrochemical N-Formylation of Amines
  via Decarboxylation of Glyoxylic Acid. Organic Letters 20(7), 2112-2115.
- 599 Liu, X., Jeffries, H.E. and Sexton, K.G. (1999) Atmospheric Photochemical
- Degradation of 1,4-Unsaturated Dicarbonyls. Environmental Science &
  Technology 33(23), 4212-4220.
- LoPachin, R.M. and Gavin, T. (2014) Molecular Mechanisms of Aldehyde
  Toxicity: A Chemical Perspective. Chemical Research in Toxicology 27(7),
  1081-1091.
- Mariscal, R., Maireles-Torres, P., Ojeda, M., Sádaba, I. and López Granados,
  M. (2016) Furfural: a renewable and versatile platform molecule for the
  synthesis of chemicals and fuels. Energy & Environmental Science 9(4), 11441189.
- 609 Olvera-Vargas, H., Oturan, N., Brillas, E., Buisson, D., Esposito, G. and Oturan,
- M.A. (2014) Electrochemical advanced oxidation for cold incineration of the
  pharmaceutical ranitidine: Mineralization pathway and toxicity evolution.
  Chemosphere 117, 644-651.
- Peterson, L.A. (2013) Reactive metabolites in the biotransformation of
  molecules containing a furan ring. Chemical Research in Toxicology 26(1), 625.
- Pohl, J., Golovko, O., Carlsson, G., Eriksson, J., Glynn, A., Orn, S. and Weiss,
  J. (2020) Carbamazepine Ozonation Byproducts: Toxicity in Zebrafish (Danio
  rerio) Embryos and Chemical Stability. Environ Sci Technol 54(5), 2913-2921.
- 619 Prasse, C. (2021) Reactivity-directed analysis a novel approach for the
- 620 identification of toxic organic electrophiles in drinking water. Environmental
- 621 Science: Processes & Impacts.

- 622 Prasse, C., Ford, B., Nomura, D.K. and Sedlak, D.L. (2018) Unexpected
- transformation of dissolved phenols to toxic dicarbonyls by hydroxyl radicals
- 624 and UV light. Proc Natl Acad Sci U S A 115(10), 2311-2316.
- 625 Prasse, C., von Gunten, U. and Sedlak, D.L. (2020) Chlorination of Phenols

626 Revisited: Unexpected Formation of  $\alpha,\beta$ -Unsaturated C4-Dicarbonyl Ring

- 627 Cleavage Products. Environmental Science & Technology 54(2), 826-834.
- 628 Prasse, C., Wagner, M., Schulz, R. and Ternes, T.A. (2011) Biotransformation
- 629 of the antiviral drugs acyclovir and penciclovir in activated sludge treatment.
- 630 Environ Sci Technol 45(7), 2761-2769.
- 631 Ravindranath, V., Burka, L.T. and Boyd, M.R. (1984) Reactive metabolites
- from the bioactivation of toxic methylfurans. Science 224(4651), 884.
- 633 Schlüter-Vorberg, L., Prasse, C., Ternes, T.A., Mückter, H. and Coors, A.
- 634 (2015) Toxification by Transformation in Conventional and Advanced
- 635 Wastewater Treatment: The Antiviral Drug Acyclovir. Environmental Science
- 636 & Technology Letters 2(12), 342-346.
- 637 Stamhuis, E.J., Drenth, W. and van den Berg, H. (1964) Mechanism of reactions
- 638 of furans I: A kinetic study of the acid-catalyzed hydrolysis of furan and 2,5-
- dimethylfuran. Recueil des Travaux Chimiques des Pays-Bas 83(2), 167-176.
- Tong, X., Ma, Y. and Li, Y. (2010) Biomass into chemicals: Conversion of
  sugars to furan derivatives by catalytic processes. Applied Catalysis A: General
  385(1), 1-13.
- Tuazon, E.C., Atkinson, R. and Carter, W.P.L. (1985) Atmospheric chemistry
  of cis- and trans-3-hexene-2,5-dione. Environmental Science & Technology
  19(3), 265-269.
- von Gunten, U. (2003) Ozonation of drinking water: Part I. Oxidation kineticsand product formation. Water Research 37(7), 1443-1467.
- von Gunten, U. (2018) Oxidation Processes in Water Treatment: Are We on
  Track? Environ Sci Technol 52(9), 5062-5075.
- von Sonntag, C. and von Gunten, U. (2012) Chemistry of ozone in water and
- 651 wastewater treatment, IWA publishing.
- Wang, B., Wang, L., Li, Y. and Liu, Y. (2014) Heterocyclic terpenes: linear
- furano- and pyrroloterpenoids. RSC Advances 4(24), 12216-12234.

- White, H.M., Colomb, H.O. and Bailey, P.S. (1965) Ozonation of 2,5Diphenylfuran1. The Journal of Organic Chemistry 30(2), 481-486.
- 656 Williams, D.P., Antoine, D.J., Butler, P.J., Jones, R., Randle, L., Payne, A.,
- Howard, M., Gardner, I., Blagg, J. and Park, B.K. (2007) The Metabolism and
- 658 Toxicity of Furosemide in the Wistar Rat and CD-1 Mouse: a Chemical and
- 659 Biochemical Definition of the Toxicophore. Journal of Pharmacology and
- Experimental Therapeutics 322(3), 1208-1220.
- 661 Yoshimi, Y., Kobayashi, K., Kamakura, H., Nishikawa, K., Haga, Y., Maeda,
- 662 K., Morita, T., Itou, T., Okada, Y. and Hatanaka, M. (2010) Addition of alkyl
- radicals, generated from carboxylic acids via photochemical decarboxylation, to
- 664 glyoxylic oxime ether: a mild and efficient route to  $\alpha$ -substituted  $\alpha$ -aminoesters.
- 665 Tetrahedron Letters 51(17), 2332-2334.
- Zou, R., Liao, X., Zhao, L. and Yuan, B. (2018) Reduction of Nnitrosodimethylamine formation from ranitidine by ozonation preceding
  chloramination: influencing factors and mechanisms. Environ Sci Pollut Res Int
  25(14), 13489-13498.
- 670 Zoumpouli, G.A., Siqueira Souza, F., Petrie, B., Féris, L.A., Kasprzyk-Hordern,
- B. and Wenk, J. (2020) Simultaneous ozonation of 90 organic micropollutants
- 672 including illicit drugs and their metabolites in different water matrices.
- 673 Environmental Science: Water Research & Technology 6(9), 2465-2478.





# **Supplementary Information**

# Aqueous Ozonation of Furans: Kinetics and Transformation Mechanisms Leading to the Formation of α,β-Unsaturated Dicarbonyl Compounds

Garyfalia A. Zoumpouli<sup>a,b,c,d,1</sup>, Zhuoyue Zhang<sup>d</sup>, Jannis Wenk<sup>b,c</sup>, and Carsten Prasse<sup>d,\*</sup>

<sup>a</sup> Centre for Doctoral Training, Centre for Sustainable Chemical Technologies, University of Bath, Bath BA2 7AY, UK.

<sup>b</sup> Department of Chemical Engineering, University of Bath, Bath BA2 7AY, UK.

<sup>c</sup> Water Innovation and Research Centre (WIRC), University of Bath, Bath BA2 7AY, UK.

<sup>d</sup> Department of Environmental Health and Engineering, Johns Hopkins University, Baltimore, MD 21218, USA.

\*corresponding author. Email: cprasse1@jhu.edu; Address: 3400 N Charles Street, Baltimore, MD 21218

<sup>1</sup> Present address: Cranfield Water Science Institute, Cranfield University, College Road,

Cranfield MK43 0AL, UK

## Content

Supplementary text S1: ozonation experiments	S3
Supplementary text S2: liquid chromatography-high resolution mass spectrometry	S3
References	S28
Fables	

# Table S1. HPLC-UV parameters for the detection of furans in competition kinetics<br/>experiments. Flow rate 0.5 mL/min, A: ultrapure water with 0.1 % v/v formic or<br/>phosphoric acid, B: acetonitrile.<br/>Table S2. NAL, NAC and NAL+NAC adducts detected in this study with LC-HRMS.<br/>Table S3. Furosemide ozonation products detected with LC-HRMS, including MS2<br/>fragmentation information. Suggested structures are supported by comparison with<br/>literature<sup>4, 5</sup> and/or based on MS2 spectra (Figure S11).<br/>Table S4. Ranitidine ozonation products detected with LC-HRMS, including MS2<br/>fragmentation information. Suggested structures are supported by comparison with<br/>literature<sup>6</sup> and/or based on MS2 spectra (Figure S14).S14

## Figures

Figure S1. Plot of the natural logarithm of the relative concentration of the reference	
compound (RAN or FA) versus the natural logarithm of the relative concentration of	\$5
the target compound (FFA, FA, FRS, FPA, MFA, BHF, FDCA). Linear fit equations	33
are shown including the standard error of the slope.	
Figure S2. Figure S2. Effect of <i>tert</i> -butanol addition on the formation of BDA and	56
BDA-R (hydroxymethyl-BDA) during the ozonation of FFA. FFA initial	30

concentration 15 µM, tert-butanol concentration 10 mM, in 10 mM phosphate buffer at pH 7. Figure S3. Degradation of A) FFA and B) BDA at different ozone concentrations, **S6** with or without addition of 10 mM *tert*-butanol, in 10 mM phosphate buffer at pH 7. Figure S4. Base peak chromatogram and MS<sup>2</sup> spectrum of m/z 255 (NAL adduct of BDA) identified in ozonation experiments with furfuryl alcohol (15 µM initial **S**7 concentration). Figure S5. Base peak chromatogram and MS<sup>2</sup> spectrum of m/z 285 identified in **S**8 ozonation experiments with furfuryl alcohol (15 µM initial concentration). Figure S6. Base peak chromatogram and MS<sup>2</sup> spectrum of m/z 315 identified in ozonation experiments with 3.4-bis(hydroxymethyl)furan (100 µM initial **S**9 concentration). Figure S7. Base peak chromatogram and MS<sup>2</sup> spectrum of m/z 327 identified in S10 ozonation experiments with 3-(2-furyl)propanoic acid (15 µM initial concentration). Figure S8. Base peak chromatogram of m/z 313 (top), base peak chromatogram and MS<sup>2</sup> spectrum of m/z 269 (m/z 313 after loss of CO<sub>2</sub>) identified in ozonation S11 experiments with 2-methyl-3-furoic acid (15 µM initial concentration). Figure S9. Base peak chromatogram and MS<sup>2</sup> spectrum of m/z 276 identified in S12 ozonation experiments with 2,5-dimethylfuran (100 µM initial concentration). Figure S10. Base peak chromatogram and MS<sup>2</sup> spectrum of m/z 428 identified in S13 ozonation experiments with 2,5-dimethylfuran (100 µM initial concentration). Figure S11. MS<sup>2</sup> spectra including fragment structures for the newly detected products a) FRS-308, b) FRS-363 and c) FRS-347 identified in ozonation S15 experiments with furosemide (15 µM initial concentration). Figure S12. Peak area of furosemide (FRS) ozonation products and FRS degradation at different ozone concentrations. BDA is shown as the BDA-NAL adduct. For FRS-265 the peak area of the ionisation fragment m/z 250 is shown. Data points are the S16 average of duplicate experiments (error bars have been omitted). Furosemide initial concentration 15 µM. Figure S13. Proposed reaction pathways for the ozonation of ranitidine. Transformation products are labelled as follows: blue ones are newly detected, black ones are previously reported,<sup>6</sup> while pink ones are those having the same molecular S20 ion m/z as previously reported<sup>6</sup> but different suggested structures based on MS<sup>2</sup> fragment information obtained (Figure S14). Figure S14. (a-j) Base peak chromatograms and MS<sup>2</sup> spectra including fragment structures for ranitidine and its transformation products identified in ozonation S21 experiments (ranitidine initial concentration 50 µM). Figure S15. Peak area of ranitidine (RAN) ozonation products and RAN degradation at different ozone concentrations. Data points are the average of duplicate S26 experiments (error bars have been omitted). Ranitidine initial concentration 15 µM. Figure S16. Reaction of dimethyl-BDA with NAL versus a NAL+NAC mixture, S27 leading to formation of adducts detected with LC-HRMS. Figure S17. Suggested mechanisms for the cleavage of the substituent of BDA-R from FA, FFA and FPA in water, leading to the formation of BDA. S27

Supplementary text S1: ozonation experiments

The competition kinetics experiments were performed in 20-mL amber glass vials. The reaction solutions (10 or 15 mL) contained 7  $\mu$ M of the target compound, 7  $\mu$ M of the reference compound (RAN or FA), 10 mM phosphate buffer (pH 7) and 10 mM *tert*-butanol in ultrapure

water. Ozone stock solution was added to achieve concentrations of 1 to 13  $\mu$ M ozone (0.1 to 0.9  $\mu$ M O<sub>3</sub>/ $\mu$ M target plus reference compound). Samples were magnetically stirred during the addition of the ozone stock solution and then left overnight at room temperature until complete ozone depletion. Residual concentrations of the target and reference compounds were measured within 24 hours. The second order rate constant for the reaction of the target compound with ozone (k<sub>O3,TC</sub>) was calculated from the plot of the natural logarithm of the relative concentration of target compound versus the natural logarithm of the relative concentration of reference compound (see Figure S1), according to the equation (Huber et al., 2003):

$$\ln\left(\frac{[TC]}{[TC]_0}\right) = \frac{k_{O_3,TC}}{k_{O_3,RC}} \ln\left(\frac{[RC]}{[RC]_0}\right)$$

The  $\pm$  error of each rate constant was calculated through error propagation from the 95% confidence interval of the slope of the linear fit and the estimated error of k<sub>O3, RAN</sub> ( $\pm 0.1 \times 10^6 \text{ M}^{-1} \text{ s}^{-1}$ ) (Jeon et al., 2016) or the calculated error of k<sub>O3, FA</sub>.

To prepare the concentrated ozone stock solution for either batch ozonation experiments or competition kinetics, two different systems were used. One was a 500-mL glass reactor that was equipped with a gas diffuser and a water jacket, with the temperature of the recirculating water in the jacket set to 2°C. The other was a 1-L glass bottle placed in an ice bath. Both systems were fed with oxygen containing 50 to 100 mg/L ozone, produced with either a BMT 803N ozone generator (Messtechnik GmbH) or an IOCS integrated ozone system (Pacific Ozone). The dissolved ozone concentration of the ozone stock solution (20 to 30 mg/L) was measured spectrophotometrically both before and after its addition into the reaction solutions, either directly at 258 nm (molar absorptivity of ozone  $\varepsilon$ =2900 M<sup>-1</sup> cm<sup>-1</sup>) or with the indigo method (Bader and Hoigné, 1981).

#### Supplementary text S2: liquid chromatography-high resolution mass spectrometry

For chromatographic separation the gradient program was at 75  $\mu$ L/min with ultrapure water containing 0.1 % formic acid (A) and methanol (B). The percentage of A was: 0-3 min, 100%; 3-12 min, linear decrease from 100% to 5%; 12-14 min, 5%; 14.1 min, 100%, total run time 20 min. The injection volume was 10  $\mu$ L.

The Electrospray ionization (ESI) source parameters were set as follows. Sheath gas flow rate: 20 arbitrary units (AU); aux gas flow rate: 10 AU; spray voltage: 3.8 kV for positive mode and 2.5 kV for negative mode; capillary temperature: 250°C; S-lens RF level: 60; aux gas

heater temperature: 100°C. Data-dependent acquisition was used to conduct  $MS^2$  experiments. Full scan (50-700 m/z, resolution > 120000) was performed followed by data-dependent  $MS^2$  for the 5 most intense ions with resolution > 60000. Collision induced dissociation (CID) with stepped normalized collision energy of 10%, 30% and 50% was used for fragmentation with an isolation window of 1.0 m/z.

Table S1. HPLC-UV parameters for the detection of furans in competition kinetics experiments. Flow rate 0.5 mL/min, A: ultrapure water with 0.1 % v/v formic or phosphoric acid, B: acetonitrile.

Compound	A/B (%)%)	Retention time (min)	Detection wavelength (nm)
Furfuryl alcohol (FFA)	90/10	2.9	216
2-Furoic acid (FA)	90/10	3.3	252
2-Methyl-3-furoic acid (MFA)	70/30	2.6	245
3-(2-Furyl)propanoic acid (FPA)	70/30	2.9	220
Furosemide (FRS)	60/40	3.0	228
Ranitidine (RAN)	90/10	2.2	320
Furan-2,5-dicarboxylic acid (FDCA)	90/10	2.5	265
3,4-Bis(hydroxymethyl)furan (BHF)	90/10	1.8	215

Table S2. NAL, NAC and NAL+NAC adducts detected in this study with LC-HRMS.

Parent compound	Amino acid added	Adduct m/z (observed)	m/z error (ppm)	Adduct formula [M+H] <sup>+</sup>	Dicarbonyl formula
FFA, FA, FPA, FRS	NAL	255.1338	-0.39	$C_{12}H_{19}O_4N_2$	$C_4H_4O_2$
FFA	NAL	285.1444	-0.35	$C_{13}H_{21}O_5N_2$	$C_5H_6O_3$
FPA	NAL	327.1549	-0.61	$C_{15}H_{23}O_6N_2$	$C_7H_8O_4$
MFA	NAL	313.1391	-0.96	$C_{14}H_{21}O_6N_2$	$C_6H_6O_4$
BHF	NAL	315.1548	-0.95	$C_{14}H_{23}O_6N_2$	$C_6H_8O_4$
DMF	NAC	276.0899	-0.36	$C_{11}H_{18}O_5NS$	$C_6H_8O_2$
DMF	NAL+NAC	428.1852	0.48	$C_{19}H_{30}O_6N_3S$	$C_6H_8O_2$



Figure S1. Plot of the natural logarithm of the relative concentration of the reference compound (RAN or FA) versus the natural logarithm of the relative concentration of the target compound (FFA, FA, FRS, FPA, MFA, BHF, FDCA). Linear fit equations are shown including the standard error of the slope.



Figure S2. Effect of *tert*-butanol addition on the formation of BDA and BDA-R (hydroxymethyl-BDA) during the ozonation of FFA. FFA initial concentration 15  $\mu$ M, *tert*-butanol concentration 10 mM, in 10 mM phosphate buffer at pH 7.



Figure S3. Degradation of A) FFA and B) BDA at different ozone concentrations, with or without addition of 10 mM *tert*-butanol, in 10 mM phosphate buffer at pH 7.



Figure S4. Base peak chromatogram and  $MS^2$  spectrum of m/z 255 (NAL adduct of BDA) identified in ozonation experiments with furfuryl alcohol (15  $\mu$ M initial concentration).



Figure S5. Base peak chromatogram and  $MS^2$  spectrum of m/z 285 identified in ozonation experiments with furfuryl alcohol (15  $\mu$ M initial concentration).



Figure S6. Base peak chromatogram and  $MS^2$  spectrum of m/z 315 identified in ozonation experiments with 3,4-bis(hydroxymethyl)furan (100  $\mu$ M initial concentration).



Figure S7. Base peak chromatogram and  $MS^2$  spectrum of m/z 327 identified in ozonation experiments with 3-(2-furyl)propanoic acid (15  $\mu$ M initial concentration).



Figure S8. Base peak chromatogram of m/z 313 (top), base peak chromatogram and MS<sup>2</sup> spectrum of m/z 269 (m/z 313 after loss of CO<sub>2</sub>) identified in ozonation experiments with 2-methyl-3-furoic acid (15  $\mu$ M initial concentration).



Figure S9. Base peak chromatogram and  $MS^2$  spectrum of m/z 276 identified in ozonation experiments with 2,5-dimethylfuran (100  $\mu$ M initial concentration).



Figure S10. Base peak chromatogram and  $MS^2$  spectrum of m/z 428 identified in ozonation experiments with 2,5-dimethylfuran (100  $\mu$ M initial concentration).

- 1 Table S3. Furosemide ozonation products detected with LC-HRMS, including  $\mathrm{MS}^2$
- 2 fragmentation information. Suggested structures are supported by comparison with literature

3	(Aalizadeh et al	2019; Laurence et	al., 2014), an	nd/or based o	on MS <sup>2</sup> spectra	(Figure S11
3	(Aanzaden et al.,	2019; Laurence et	al., 2014), al	nd/or based o	on MS spectra	(rigure SI

Compound	Retention time (min)	m/z (observed)	m/z error (ppm)	Formula [M+H] <sup>+</sup>	Suggested structure
		331.0143	2.1	C <sub>12</sub> H <sub>12</sub> O <sub>5</sub> N <sub>2</sub> ClS	0 0 H <sub>2</sub> N, //
FRS		250.9885	1.1	$C_7H_8O_4N_2ClS$	- S ОН
	14.1	232.9780	0.9	C7H6O3N2ClS	
		185.9951	0.8	C7H5O3NCl	
		265.0042	0.9	$C_8H_{10}O_4N_2ClS$	
FRS-265	11.1	250.9886	0.7	C7H8O4N2ClS	S OH
	11.1	232.9782	0.1	C7H6O3N2ClS	0
		185.9952	0.3	C7H5O3NCl	
		228 0080	1.4	C. H. O.N.CIS	
		328.9989	2.5	$C_{12}H_{10}O_5N_2CIS$	S 0 <sup>⊖</sup>
FRS-328	8.0	281 0082	1.3	$C_{12}H_8O_4NC1$	0
1 K0-520	0.0	266.0211	1.5	C12H8O5NCI	
		249 0184	13	$C_{12}H_{9}O_{2}NCl$	
		219.0101	1.5	012118031101	он
		308 9939	12	CoH10O6N2ClS	
	12.0	290 9834	1.2	CoHeO5N2CIS	З ОН
FRS-308		262.9886	0.7	$C_8H_8O_4N_2CIS$	
		244.9781	0.5	$C_8H_6O_3N_2ClS$	
					он
		363.0043	1.4	$C_{12}H_{12}O_7N_2ClS$	
FRS-363	9.1, 10.0	335.0095	1.2	$C_{11}H_{12}O_6N_2ClS$	ОН
(two peaks)		316.9990	1.1	$C_{11}H_{10}O_5N_2ClS$	CI
		262.9886	0.7	C <sub>8</sub> H <sub>8</sub> O <sub>4</sub> N <sub>2</sub> ClS	он о
		347.0097	0.6	$C_{12}H_{12}O_6N_2ClS$	
FRS-347	11.8	328.9994	-0.2	$C_{12}H_{10}O_5N_2ClS$	СН
1100011	1110	262.9885	1.1	$C_8H_8O_4N_2ClS$	
		244.9780	0.9	C <sub>8</sub> H <sub>6</sub> O <sub>3</sub> N <sub>2</sub> ClS	
FRS 266	10.6	266.9834	1.1	$C_7H_8O_5N_2ClS$	ССССССССССССССССССССССССССССССССССССССС
FRS-266	10.0	248.9729	0.9	$C_7H_6O_4N_2ClS$	ОН
FRS-278	11.8	278.9834	1.1	C <sub>8</sub> H <sub>8</sub> O <sub>5</sub> N <sub>2</sub> ClS	



4

5 Figure S11. MS<sup>2</sup> spectra including fragment structures for the newly detected products a) FRS-

308, b) FRS-363 and c) FRS-347 identified in ozonation experiments with furosemide (15  $\mu$ M

7 initial concentration).



9

Figure S12. Peak area of furosemide (FRS) ozonation products and FRS degradation at different ozone concentrations. BDA is shown as the BDA-NAL adduct. For FRS-265 the peak area of the ionisation fragment m/z 250 is shown. Data points are the average of duplicate experiments (error bars have been omitted). Furosemide initial concentration 15  $\mu$ M.

- 14
- 15

Table S4. Ranitidine ozonation products detected with LC-HRMS, including MS<sup>2</sup>
 fragmentation information. Suggested structures are supported by comparison with literature

Compound	Retention time (min)	m/z (observed)	m/z error (ppm)	Formula [M+H] <sup>+</sup>	Suggested structure	
		315.1481	-1.5	$C_{13}H_{23}O_3N_4S$		
		270.0902	-1.8	$C_{11}H_{16}O_3N_3S$		
		224.0974	-1.6	$C_{11}H_{16}ON_2S$		
		176.0487	-0.8	$C_5H_{10}O_2N_3S$		
RAN	8.7	144.0766	-1.2	$C_5H_{10}O_2N_3$	$ \begin{array}{c} -\mathbf{N} & +\mathbf{N}^* & \mathbf{O} \\ \mathbf{S} & \mathbf{S} & \mathbf{S} & \mathbf{N}^* \end{array} $	
		124.0757	0.2	C <sub>7</sub> H <sub>10</sub> ON		
		117.0481	0.2	$C_4H_9N_2S$		
		98.0842	3.1	$C_{5}H_{10}N_{2}$		
		58.0658	11.0	C <sub>3</sub> H <sub>8</sub> N		
		331.1430	-1.3	$C_{13}H_{23}O_4N_4S$		
		313.1330	0.4	$C_{13}H_{21}O_{3}N_{4}S$		
		286.0851	-1.9	$C_{11}H_{16}O_4N_3S$		
		240.0924	-1.3	$C_{11}H_{16}O_2N_2S$		
		222.0818	-1.4	C <sub>11</sub> H <sub>14</sub> ON <sub>2</sub> S		
RAN-S	2.5	192.0435	-1.5	$C_5H_{10}O_3N_3S$		
oxide	3.7	188.0738	-1.1	$C_8H_{14}O_2NS$		
		138.0913	-0.2	C <sub>8</sub> H <sub>12</sub> ON	Н	
		110.0967	2.0	$C_7H_{12}N$		
		94.0417	3.5	C <sub>6</sub> H <sub>6</sub> O		
		82.0656	6.1	C <sub>5</sub> H <sub>8</sub> N		
		58.0658	11.0	$C_3H_8N$		
		331.1429	-1.8	$C_{13}H_{23}O_4N_4S$		
		270.0902	-1.7	$C_{11}H_{16}O_3N_3S$		
	8.9	224.0974	-1.7	$C_{11}H_{16}ON_2S$	0-	
RAN-N		176.0486	-1.0	$C_5H_{10}O_2N_3S$		
oxide		144.0766	-1.2	$C_5H_{10}O_2N_3$		
		130.0559	1.2	$C_5H_{10}N_2S$	Н	
		98.0842	3.2	$C_{5}H_{10}N_{2}$		
		88.0220	4.7	C <sub>3</sub> H <sub>6</sub> NS		
		347.1379	-1.7	$C_{13}H_{23}O_5N_4S$		
		286.0850	-2.0	$C_{11}H_{16}O_4N_3S$		
		240.0920	-2.7	$C_{11}H_{16}O_2N_2S$		
DANGON		193.0513	-1.4	$C_5H_{11}O_3N_3S$		
RAN-S&N	4.2	192.0434	-1.7	$C_5H_{10}O_3N_3S$		
Oxide		146.0506	-1.6	$C_5H_{10}ON_2S$		
		130.0610	-1.1	$C_4H_8O_2N_3$		
		100.0998	2.5	$C_5H_{12}N_2$		
		73.0765	7.0	$C_3H_9N_2$		
		300.1371	-1.6	$C_{13}H_{22}O_3N_3S$		
DAN 2000	2.0	282.1266	-1.6	$C_{13}H_{20}O_2N_3S$		
KAIN-3003	2.0	255.0793	-2.0	$C_{11}H_{15}O_3N_2S$		
			237.0687	-2.0	$C_{11}H_{13}O_{3}N_{2}S$	l Ö

18 (Christophoridis et al., 2016), and/or based on  $MS^2$  spectra (Figure S14).

		188.0737	-1.5	$C_8H_{14}O_2NS$	
		138.0913	-0.3	C <sub>8</sub> H <sub>12</sub> ON	
		110.0966	1.8	$C_7H_{12}N$	
		94.0416	3.5	$C_6H_6O$	
		82.0656	6.1	$C_5H_8N$	
		58.0658	11.0	C <sub>3</sub> H <sub>8</sub> N	
		300.1369	-2.5	$C_{13}H_{22}O_3N_3S$	
		256.1473	-2.1	$C_{12}H_{22}ON_3S$	
		211.0896	-1.9	$C_{10}H_{15}ON_2S$	
		170.0631	-1.9	C <sub>8</sub> H <sub>12</sub> ONS	
DAN 2001	6.0	153.0366	-1.9	C <sub>8</sub> H <sub>9</sub> OS	
KAN-3000	0.0	138.0911	-1.4	C <sub>8</sub> H <sub>12</sub> ON	O S N OH
		125.0055	-0.8	C <sub>6</sub> H <sub>5</sub> OS	0
		124.0757	-0.2	C <sub>7</sub> H <sub>10</sub> ON	
		117.0481	0.4	$C_4H_9N_2S$	
		85.0764	4.8	$C_4H_9N_2$	
		316.1320	-1.8	$C_{13}H_{22}O_4N_3S$	
		272.1423	-1.7	$C_{12}H_{22}O_2N_3S$	
		254.1317	-1.8	$C_{12}H_{20}ON_3S$	
		227.0845	-1.7	$C_{10}H_{15}O_2N_2S$	
DANI 21/C	2.2	209.0741	-0.9	$C_{10}H_{13}ON_2S$	
RAN-316S	2.3	188.0738	-1.1	$C_8H_{14}O_2NS$	S N OH
		138.0912	-0.7	C <sub>8</sub> H <sub>12</sub> ON	0
		110.0966	1.7	C7H12N	
		85.0765	5.4	C <sub>4</sub> H <sub>9</sub> N <sub>2</sub>	
		58.0658	11.0	C <sub>3</sub> H <sub>8</sub> N	
		316.1317	-2.8	C <sub>13</sub> H <sub>22</sub> O <sub>4</sub> N <sub>3</sub> S	
		272.1421	-2.3	$C_{12}H_{22}O_2N_3S$	
		212.0973	-2.2	$C_{10}H_{16}ON_2S$	
RAN-316N	7.2	170.0631	-1.7	C <sub>8</sub> H <sub>12</sub> ONS	S. OH
		153.0365	-2.4	C <sub>8</sub> H <sub>9</sub> OS	
		118.0559	0.1	$C_4H_{10}N_2S$	0
		85.0764	4.7	C <sub>4</sub> H <sub>9</sub> N <sub>2</sub>	
		332.1270	-1.4	$C_{13}H_{22}O_5N_3S$	
		288.1372	-1.5	$C_{12}H_{22}O_3N_3S$	
		243.0795	-1.3	$C_{10}H_{15}O_3N_2S$	
		227.0847	-0.8	$C_{10}H_{15}O_2N_2S$	
D 1 1 2 2 2	4.0	151.0534	-0.9	$C_4H_{11}O_2N_2S$	
RAN-332	4.0	149.0378	-0.9	$C_4H_9O_2N_2S$	S N OH
		138.0912	-0.8	C <sub>8</sub> H <sub>12</sub> ON	ÓH Ö
		134.0508	-0.5	$C_4H_{10}ON_2S$	
		110.0966	1.4	$C_7H_{12}N$	
		<u>85</u> .0765	5.2	$C_4H_9N_2$	
		305.1159	-2.1	$C_{12}H_{21}O_5N_2S$	
		287.1054	-2.0	$C_{12}H_{19}O_4N_2S$	/
DANI 205	2.2	166.0165	-2.0	C <sub>4</sub> H <sub>8</sub> O <sub>4</sub> NS	
KAN-305	2.2	138.0912	-1.1	C <sub>8</sub> H <sub>12</sub> NO	I O <sup>™</sup> S <sup>™</sup> H
		110.0966	1.5	C7H12N	۳ ď
		94.0417	4.3	$C_6H_6O$	

		334.1426	-1.4	$C_{13}H_{24}O_5N_3S$	
		316.1317	-2.8	$C_{13}H_{22}O_4N_3S$	
		255.0790	-3.0	$C_{11}H_{15}O_3N_2S$	_
		180.0558	-1.8	$C_5H_{12}O_3N_2S$	O $N$
RAN-334	2.2	177.0326	-1.1	$C_5H_9O_3N_2S$	S N H
		162.0456	-1.2	$C_5H_{10}O_2N_2S$	
		161.0375	-2.4	$C_5H_9O_2N_2S$	
		113.0710	0.1	C <sub>5</sub> H <sub>9</sub> ON <sub>2</sub>	
		95.0494	2.4	C <sub>6</sub> H <sub>7</sub> O	
		236.0697	-1.2	$C_7H_{14}O_4N_3S$	
	7.5	219.0670	-1.2	$C_7H_{13}O_3N_3S$	
DAN 226		190.0768	-1.6	$C_7H_{14}O_2N_2S$	
KAIN-230		131.0638	0.0	$C_5H_{11}N_2S$	
		119.0163	1.1	$C_4H_7O_2S$	ÔH ''
		73.0112	8.1	C <sub>3</sub> H <sub>5</sub> S	
		252.0645	-1.3	$C_7H_{14}O_5N_3S$	
		234.0540	-1.4	$C_7H_{12}O_4N_3S$	
		206.0719	-0.1	$C_7H_{14}O_3N_2S$	
		193.0518	1.2	$C_5H_{11}O_3N_3S$	
DAN 252	2.4	188.0612	-1.0	$C_7H_{12}O_2N_2S$	$\downarrow$ $\stackrel{\circ}{=}$
KAIN-232	2.4	176.0251	0.5	$C_5H_8O_3N_2S$	
		160.0299	-1.1	$C_5H_8O_2N_2S$	ÓH
		144.0766	-1.0	$C_5H_{10}O_2N_3S$	
		134.0270	-0.5	$C_4H_8O_2NS$	
		98.0842	3.5	$C_{5}H_{10}N_{2}$	



Figure S13. Proposed reaction pathways for the ozonation of ranitidine. Transformation products are labelled as follows: blue ones are newly detected, black ones are previously reported (Christophoridis et al., 2016), while pink ones are those having the same molecular ion m/z as previously reported (Christophoridis et al., 2016), but different suggested structures based on MS<sup>2</sup> fragment information obtained (Figure S14).















Figure S14. (a-j) Base peak chromatograms and  $MS^2$  spectra including fragment structures for ranitidine and its transformation products identified in ozonation experiments (ranitidine initial concentration 50  $\mu$ M).

36



Figure S15. Peak area of ranitidine (RAN) ozonation products and RAN degradation at
different ozone concentrations. Data points are the average of duplicate experiments (error
bars have been omitted). Ranitidine initial concentration 15 μM.





46 Figure S16. Reaction of dimethyl-BDA with NAL versus a NAL+NAC mixture, leading to

47 formation of adducts detected with LC-HRMS.

48



- 50 Figure S17. Suggested mechanisms for the cleavage of the substituent of BDA-R from FA,
- 51 FFA and FPA in water, leading to the formation of BDA.

#### 52 **References**

- 53 Aalizadeh, R., Nika, M.C. and Thomaidis, N.S. (2019) Development and application of
- retention time prediction models in the suspect and non-target screening of emergingcontaminants. J Hazard Mater 363, 277-285.
- 56 Bader, H. and Hoigné, J. (1981) Determination of ozone in water by the indigo method.
- 57 Water Research 15(4), 449-456.
- 58 Christophoridis, C., Nika, M.C., Aalizadeh, R. and Thomaidis, N.S. (2016) Ozonation of
- 59 ranitidine: Effect of experimental parameters and identification of transformation products.
- 60 Sci Total Environ 557-558, 170-182.
- 61 Huber, M.M., Canonica, S., Park, G.-Y. and von Gunten, U. (2003) Oxidation of
- 62 Pharmaceuticals during Ozonation and Advanced Oxidation Processes. Environmental
- 63 Science & Technology 37(5), 1016-1024.
- 64 Jeon, D., Kim, J., Shin, J., Hidayat, Z.R., Na, S. and Lee, Y. (2016) Transformation of
- 65 ranitidine during water chlorination and ozonation: Moiety-specific reaction kinetics and
- elimination efficiency of NDMA formation potential. Journal of Hazardous Materials 318,802-809.
- 68 Laurence, C., Rivard, M., Martens, T., Morin, C., Buisson, D., Bourcier, S., Sablier, M. and
- Oturan, M.A. (2014) Anticipating the fate and impact of organic environmental
  contaminants: a new approach applied to the pharmaceutical furosemide. Chemosphere 113,
  193-199.
- 72
- 73