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1	On the design of food effect studies in adults for extrapolating oral
2	drug absorption data to infants: An exploratory study highlighting the
3	importance of infant food
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# 28 Abstract

29 In the present investigation, it was explored whether food effect on drug absorption in adults is similar 30 with the food effect after administration of an infant meal with the drug product to adults. After 31 confirming lack of pharmaceutical and pharmacokinetic interaction, a paracetamol suspension and an 32 ibuprofen suspension were co-administered to eight healthy adults on a crossover basis in three 33 different occasions, i.e. in the fasted state (as defined by regulatory agencies, fasted conditions), in the 34 fed state (as defined by regulatory agencies, fed conditions) and under conditions simulating the fed 35 state in infants (infant fed conditions). Unlike under fed conditions, under infant fed conditions early 36 exposure was significantly lower than under fasted conditions for both paracetamol and ibuprofen. 37 For ibuprofen, C<sub>max</sub> values under infant fed conditions were also significantly higher than under fed 38 conditions. These data suggest that, even for drugs with non-problematic absorption administered in 39 simple dosage forms, food effects in infants may not be adequately evaluated if the protocol suggested 40 by regulatory agencies is applied. The usefulness of the methodology employed in the present 41 investigation for simulating the fed state in infants deserves further evaluation. Until then, food effects 42 in infants should be considered cautiously or be evaluated in infants.

43

## 45 Introduction

Oral drug delivery is the route of choice for drug administration from birth to adolescence (1–3).
Therefore, understanding drug and drug formulation performance in relation to the prandial
conditions is essential for ensuring safety and efficacy of products to be administered to paediatric
patients, especially newborns (birth – 27 days) and infants (28 days – 2 years) whose diet is specific
(100 % milk in newborns) (4–6).

51

52 Understanding the impact of prandial conditions on drug/drug product performance in paediatric 53 patients is limited by ethical concerns and the subsequent difficulty to perform such studies. 54 Difficulties in recruitment are reflected by the limited number of food effect studies in children 55 published to date [(25 to the best of our knowledge, (7-27)]. Importantly, most of these studies either 56 do not focus on a specific paediatric subpopulation (9–12,20–28) or focus on school-children (13– 57 15,17). As a result, differences in gastrointestinal physiology across paediatric subpopulations and differences in meals administered to evaluate the impact of prandial conditions increase data 58 59 variability and drastically decrease their usefulness.

60

61 In recent years, there has been a growing interest in investigating whether food effect data collected 62 in adults are useful for paediatric products (2). Based on a recent draft guidance issued by the U.S. 63 Food and Drug Administration (FDA), when the same to-be-marketed formulation that is approved for 64 use in adults is approved for use in a paediatric population, a separate food effect study is not 65 necessary (6) and the same may also apply in case a paediatric formulation is very similar to the adult formulation and has been approved based on in vitro dissolution tests (6). To date, nine food effect 66 67 studies (7 drugs) in infants and young children have been published (McCracken et al. 1978 (8) – age 68 range 2-46 months; Ginsburg et al. 1979 (7) – age range 4-45 months). All studies were performed on 69 a predominantly crossover basis and in all of them the tested product was an antibiotic suspension. 70 Fasting was defined as no food or milk substance for two hours before and after drug ingestion. The 71 fed state was induced with milk or infant formula co-administered with the product, i.e. 4 oz of milk 72 or infant formula administered immediately after drug administration (8) or 4 oz of milk or infant 73 formula (Similac<sup>®</sup> or Infamil<sup>®</sup>) administered with the drug (7). The impact of food on plasma levels 74 based on these studies is summarised and compared with the impact of food on the plasma levels of 75 the same antibiotics in adults in Table I. The adult studies were performed with immediate release 76 products, after overnight fasting (fasting state) and 0-60 min after a solid meal (fed state), on a 77 crossover basis. Based on the data shown in Table I, only erythromycin ethyl-succinate seems to have 78 similar food effect in infants and in adults. It should be noted that most of the data presented in Table I 79 have been collected more than forty years ago.

80

81 Another concern, when food effect data on oral drug absorption in adults are to be extrapolated to 82 paediatric populations, relates to the design of food effect studies in adults. The recent guideline on 83 how to conduct food effect studies for newly developed paediatric formulations issued by the FDA 84 suggests that the food effect for paediatric formulations could be evaluated in adults using foods and 85 quantities of food that are commonly consumed with drugs in paediatric populations with a 86 subsequent extrapolation of the results to the paediatric population (6). Although this may be a 87 practical approach to consider, conceptually, it is different from that applied to date for the evaluation 88 of food effects on adult pharmaceutical products. In adults, relevant studies aim at detecting the 89 maximum effect on bioavailability by employing a high-calorie, high-fat meal, with less emphasis on its 90 exact composition (5,6). Importantly, studies in adults are performed by administering the drug 91 product 30 minutes after the initiation of consumption of the meal in order to maximise the potential 92 effect, whereas in paediatric populations drug are usually administered together with meals (19).

93

The aim of the present study was to explore whether food effect on drug absorption in adults is similar with the food effect after administration of an infant meal with the drug product to adults. Specifically, comparative bioavailability studies of two drugs were performed under three different prandial and dosing conditions, i.e.

98	<ul> <li>fasted state conditions as defined by regulatory agencies (fasted conditions)</li> </ul>
99	• fed state conditions as defined by regulatory agencies (fed conditions), and
100	<ul> <li>simulated infant fed state conditions (infant fed conditions)</li> </ul>
101	Paracetamol (high solubility, weak acid, pka 9.5) and ibuprofen (low solubility, weak acid, pka 4.5) (41–
102	43) were selected as model drugs based on their luminal stability and high intestinal permeability. After
103	confirming the lack of pharmaceutical interaction and pharmacokinetic interaction, based on available
104	literature data (44,45), the drugs were co-administered using commercially available paediatric
105	suspensions, i.e. variations of dosing should impact primarily gastric emptying (paracetamol) or gastric
106	emptying and, perhaps, dissolution (ibuprofen).

# 107 Materials and Methods

108

109 Materials

The commercially available paediatric suspensions Panadol<sup>®</sup> (24 mg/mL, *GlaxoSmithKline Consumer Healthcare (Ireland) Ltd.*) and Nurofen<sup>®</sup> (20 mg/mL, *ReckittBenckiser Healthcare International Ltd.*) were acquired from a local pharmacy. Paracetamol (Ph. Eur.) and ibuprofen (Ph. Eur.) powders were kindly donated by Uni-Pharma SA (Athens, Greece). Acetonitrile and methanol (Merck, Darmstadt, Germany) and water (Fischer Scientific, Schwerte, Germany) were of HPLC grade. All other chemicals were of analytical grade.

116

117 As listed in the patient information leaflet, the Panadol® formulation is composed of the following excipients: malic acid, azorubine, xanthan gum, maltitol syrup, strawberry flavour L10055, sorbitol 118 119 70 % (w/v) (crystallising), sodium methyl parahydroxybenzoate, sodium ethyl parahydroxybenzoate, 120 sodium propyl parahydroxybenzoate, sorbitol, anhydrous citric acid, purified water. According to manufacturer information, the formulation contains 133.3 mg sorbitol (incl. maltitol syrup 121 122 content)/mL (46), that is, 5.6 g of sorbitol in the total volume of formulation (42 mL) administered to 123 the volunteers. This results in a total caloric content of 11.8 kcal for the administered 42 mL Panadol® 124 suspension.

125

The Nurofen<sup>®</sup> formulation is composed of the following excipients: citric acid, sodium citrate, sodium chloride, sodium saccharin, domiphen bromide, purified water, polysorbate 80, maltitol liquid, xanthan gum, strawberry flavor, glycerol. The formulation contains 445.2 mg of maltitol syrup/mL of formulation (47). According to the Ph. Eur. monograph for maltitol syrup, it is composed of 68-85% maltitol (w/v) (48), resulting in a range of 12.1 – 15.1 g maltitol for the formulation volume administered to the volunteers (40 mL). The amount of glycerol in the formulation is 126 mg/mL of formulation (47), resulting in 5.05 g of glycerol for the formulation volume administered to the volunteers. Based on these components, the total caloric content of the 40 mL formulation administered to the volunteers ranges between 45 and 52 kcal.

135

136 Methods

- 137
- 138 Study design

This study was a single-dose, open-label, randomised, crossover, three-phase comparative oral bioavailability study with a washout period of one week. The study was performed in accordance with the ethical standards for studies in humans of the Declaration of Helsinki and its amendments (49) and the International Conference on Harmonization Guideline for Good Clinical Practice (50). The study protocol, informed consent form, and insurance contract received approval by the Executive and Ethics Committee of the Red Cross Hospital of Athens, Greece (Protocol Nr. 4145/14-02-18). The clinical study was conducted at the Gastroenterological Department of the Red Cross Hospital of Athens.

146

147 Subjects

Healthy male adults between the age of 20 and 50 years with Body-Mass-Index (BMI) within 20 % above or below the ideal BMI as determined by the Metropolitan Life Tables were recruited for this study. Ten healthy adult Caucasian males were recruited. A total of eight volunteers completed all three study phases. The participation of one volunteer was discontinued, due to inability of consuming the requested amount of one meal according to the protocol early in the morning. Another volunteer was unable to proceed with his participation after completing one of the study phases for health reasons unrelated to the present study. The mean age of the volunteers who completed the three study phases was 28.4 years (range 21-48 years) and the mean body-mass-index was 23.6 kg/m<sup>2</sup>
(range 20.3-27.7 kg/m<sup>2</sup>). No adverse effects were recorded in the present study.

157

#### 158 Inclusion criteria

The health status of the subjects was confirmed by reviewing their medical history and a general physical examination prior to the study (e.g. blood test to assess electrolyte balance, kidney and liver function, blood morphologic characteristics, glucose and lipid levels, Hepatitis B surface antigen, antibodies against Hepatitis C virus, and HIV combined Ag/Ab test). The volunteers had to be able to abstain from cigarette smoking, alcohol, and over-the-counter and prescription medication(s) for 3 days prior each study phase until the end of the study phase.

165

#### 166 Exclusion criteria

167 Volunteers were excluded based on the existence of a major health problem (cardiovascular, pancreatic, hepatic, thyroid etc.), existence of any condition requiring prescription drug therapy, 168 169 recent history of gastrointestinal disorder symptoms regardless of the severity (e.g. heartburn, 170 constipation etc.), swallowing difficulties, and receipt of an investigational agent (new or generic) 171 within 30 days prior to the initiation of and throughout the study. Further exclusion criteria were the presence of antibodies indicating active acute or chronic HIV, HBV, or HCV infection in the performed 172 173 blood tests. Subjects who could not abstain from use of medication that may affect the gastro-174 intestinal function (including antacids, PPIs, H2-receptor inhibitors, and laxatives) within 30 days of the 175 study were excluded.

#### 177 Experimental protocol

178 The volunteers were required to comply with the fasting period of 12 h before the start of each study 179 day. In the morning of each phase, the subjects arrived at the hospital at 8:00 a.m. and stayed until 180 completion of the study phase. Upon their arrival, the volunteers' health status and compliance with 181 the study protocol was confirmed and water consumption was restricted for the time period of 1h 182 before and 4.5 h after dosing. A standard lunch comprised of a club sandwich and French fries 183 (ca. 1000 kcal) was offered 4.5 h after drugs administration. Blood samples (8 mL) were collected from 184 the forearm vein via peripheral venous catheter prior to drug administration, and 10, 20, 30, 45 min, 185 1, 1.5, 2, 2.5, 3, 4, 5, 6, 8, and 10 h after drugs administration. Upon collection blood was transferred 186 into EDTA-containing Vacutainers<sup>™</sup>, following centrifugation and plasma separation. The plasma 187 samples were divided into two subsamples for separate analysis of ibuprofen and paracetamol to avoid 188 repeated freeze-thaw cycles and were stored at -20° C.

189

190 Subjects were randomised to receive a single dose of 800 mg ibuprofen (40 mL Nurofen® paediatric suspension) and a single dose of 1000 mg paracetamol (42 mL Panadol® paediatric suspension) on 191 192 three different occasions under three different dosing conditions: administration with water - "fasted 193 conditions" according to regulatory guidelines for bioavailability/bioequivalence studies (Phase I), 194 administration with water 30 minutes after the start of a high-fat, high-caloric meal (FDA meal) consumption – "fed conditions" (Phase II) (5,51), and "infant fed conditions" simulating typical 195 196 administration conditions in infants (Phase III). The selected model drugs have shown no relevant 197 pharmacokinetic interactions when co-administered orally and/or intravenously to healthy humans 198 (44,45).

199

In Phase I the formulations were administered with 168 mL of water (the total fluid volume of the
administered formulations and water was 250 mL) in the following manner: 84 mL of water, 20 mL of

Nurofen<sup>®</sup>, and 21 mL of Panadol<sup>®</sup> over 1 minute, followed by 20 mL of Nurofen<sup>®</sup>, 21 mL Panadol<sup>®</sup>, and
84 mL of water over 1 minute. The formulations were administered continuously, without time gaps
in-between. Time zero was set just after the completion of the first minute (Figure 1).

205

In Phase II, the formulations were administered as described for Phase I but 30 minutes after initiation
of ingestion of the FDA meal [two eggs (Golden Eggs<sup>®</sup>, Athens, Greece) fried in 31.3 g of butter
(Lurpak<sup>®</sup>, Danish Dairy Board, Viby, Denmark), two strips of bacon (Nikas<sup>®</sup>, Athens, Greece), two slices
of toast bread (Karamolegos A.E., Koropi, Greece), 56 g of French fries (Everest, Greece) and 240 mL
of whole cow's milk (Delta<sup>®</sup> 3.5% fat, Delta, Athens, Greece)] with a total caloric content of 990 kcal
derived from 25 % carbohydrates, 61 % fats, and 14 % proteins.

212

213 For Phase III, infant formula [Noulac® (Nounou®, Fresland Campina Hellas, Athens, Greece), 214 47 % carbohydrates, 43 % fats, and 10 % proteins], was selected as an age-representative meal in the 215 paediatric subpopulations below the age of 24 months based on its frequent use (2). Breastmilk or 216 infant formula are the exclusive feed until the age of 6 months and remain a main daily feed during 217 infancy (2). Therefore, infant formula can be considered an appropriate meal for testing food effects 218 in infants including infants that are being weaned. The volume of infant formula in the present study 219 was 800 mL (520 kcal) and was based on the recommended infant formula volume for infants, scaled 220 up by a body surface area factor for adults/infants (2). To simulate dosing conditions in infants during 221 feeding, the total volume was split into two portions and 400 mL were consumed at a constant rate 222 over 8 minutes, subsequently 20 mL of Nurofen<sup>®</sup> and 21 mL of Panadol<sup>®</sup> were administered over 223 2 minutes. Upon completion, time zero was set and drugs administration continued by 20 mL of 224 Nurofen<sup>®</sup> and 21 mL of Panadol<sup>®</sup> over 2 minutes, after which the second portion (400 mL) of infant 225 formula was consumed at a constant rate over 8 minutes. The formulations and infant formula were 226 administered continuously, without time gaps in-between.

Both the FDA meal (Phase II) and the infant formula (Phase III) were prepared freshly on each clinicalday.

230

## 231 Determination of drug plasma levels

Analysis of each drug was performed separately in duplicate. Sample treatment involved plasma protein precipitation and subsequent centrifugation and drug levels were measured by HPCL-UV based on previously proposed methods (Lalande et al. 1986; Vertzoni et al. 2003). The chromatographic system (SpectraSystem<sup>®</sup>) consisted of a P4000 pump, UV1000 absorbance detector, and an AS3000 autosampler. The above system was controlled by ESIchrome chromatography software package (v. 3.2, Thermo Fisher Scientific, San Jose, CA USA).

238

#### 239 Paracetamol

240 For paracetamol analysis, 300  $\mu$ L trifluoroacetic acid 10 % (v/v) and 150  $\mu$ L plasma sample were mixed 241 vigorously for 1 minute. The sample was centrifuged for 10 minutes at 10° C and 10 000 rpm (52). 242 300  $\mu$ L of the clear supernatant were collected and diluted with 300  $\mu$ L water and injected into the 243 HPLC system. The separation utilised a BDS Hypersil<sup>®</sup> C18 column (250×4.0 mm, 5 µm) equipped with 244 a preceding BDS pre-column (10×4.6 mm, 5  $\mu$ m), with a mobile phase consisting of 10 mM ammonium 245 formate of pH 6.0 and methanol (90:10 v/v). Paracetamol was eluted at an isocratic flowrate of 246 0.8 mL/min and detected at 424 nm. Calibration curves using the peak area of paracetamol in spiked 247 plasma and mobile please showed no significant differences regarding their slope or intercept (t-test, 248 95% confidence interval). Linearity was shown over the working range 7.5 - 4 000 ng/mL, with a 249 regression coefficient ( $R^2$ ) of  $\ge$  0.999. The lower limit of quantification (LLOQ) was 7.5 ng/mL and only

3 out of the 336 samples exhibited drug levels below the LLOQ. Sample quantification was performed
 via calibration curves constructed in spiked individual blank plasma from the corresponding volunteer.

252

#### 253 Ibuprofen

254 For the analysis of ibuprofen, 200  $\mu$ L plasma sample were acidified by addition of 20  $\mu$ L of 5 % (v/v) 255 trifluoroacetic acid, mixed briefly, followed by addition of 380 µL of ice-cold acetonitrile (53). The 256 mixture was vigorously vortexed for 1 minute and subsequently centrifuged (10 minutes, 10° C, 257 10 000 rpm). 300 µL of the clear supernatant were collected, diluted with 300 µL mobile phase and 258 were injected into the HPLC system. Separation was performed with a Fortis® C18 column 259  $(150 \times 3.0 \text{ mm}, 5 \mu\text{m})$  equipped with a preceding BDS pre-column  $(10 \times 4.6 \text{ mm}, 5 \mu\text{m})$ . The mobile phase 260 consisted of acetonitrile and 100 mM sodium acetate of pH 3.5 (60:40 v/v). Ibuprofen was eluted at an 261 isocratic flowrate of 0.5 mL/min and detected at 220 nm. Calibration curves employing the peak area 262 of ibuprofen in spiked plasma and mobile phase showed no significant differences regarding their slope 263 or intercept (t-test, 95% confidence interval). Linearity was shown over the working range 50 - 10 000 ng/mL, with a regression coefficient ( $R^2$ ) of  $\ge$  0.999. The LLOQ was 50 ng/mL and all 336 264 265 plasma samples exhibited drug levels above the LLOQ. Sample quantification for each volunteer was performed via calibration curves in spiked individual blank plasma from the corresponding volunteer. 266

267

268

#### Data analysis

269 Concentrations below the LLOQ were assigned a value of  $0 \mu g/mL$ . The maximum plasma concentration 270 ( $C_{max}$ ) and the time to reach peak plasma levels ( $T_{max}$ ) were read out directly from raw data. The area 271 under the plasma concentration-time curve until the last sampling timepoint (AUC<sub>0-10h</sub>) was calculated 272 applying the linear trapezoidal rule. The area under the plasma concentration-time curve extrapolated 273 to infinity (AUC<sub>0-inf</sub>) was determined with WinNonlin (Version 5.2; Certara USA, Inc., Princeton, USA). 274 Based on a recent draft FDA guidance, for certain classes of drugs (e.g. analgesic drug products) an 275 evaluation of the partial exposure could be required to support the determination of the relative 276 bioavailability of the drug products (FDA, 2019b). In this study, partial AUC values truncated at the 277 median T<sub>max</sub> of each study phase were calculated applying the linear trapezoidal rule, specifically 278 AUC<sub>0-1.5h</sub>, AUC<sub>0-3h</sub>, and AUC<sub>0-4h</sub> for paracetamol and AUC<sub>0-0.75h</sub>, AUC<sub>0-1.5h</sub>, and AUC<sub>0-3h</sub> for ibuprofen corresponding to the median T<sub>max</sub> values in Phases I, II, and III, respectively. Additionally, the partial 279 280 AUC<sub>0-4h</sub> was calculated for ibuprofen, as the absorption phase is assumed to be completed at this 281 timepoint.

282

283 Comparison between study phases was performed via one-way repeated measures Analysis Of 284 Variance (ANOVA) tests with a post-hoc Tukey-test, and statistical significance level was set at p < 0.05 after confirming normality and equal variance for the samples under comparison using SigmaPlot 285 286 (SigmaPlot 11.0, Systat Software Inc., San Jose, USA). The one-way repeated measures ANOVA was 287 conducted for AUC<sub>0-inf</sub>, AUC<sub>0-10h</sub>, and C<sub>max</sub> for both drugs, the partial AUC<sub>0-1.5h</sub>, AUC<sub>0-2.5h</sub>, AUC<sub>0-4h</sub> for 288 paracetamol, and the partial AUC<sub>0-0.75h</sub>, AUC<sub>0-1.5h</sub>, AUC<sub>0-3h</sub>, and AUC<sub>0-4h</sub> for ibuprofen. Friedman repeated 289 measures ANOVA on Ranks was applied for comparison between T<sub>max</sub> values in the three study phases. 290 In all cases significance of difference was considered at 0.05 level.

### 291 Results

292

293 Paracetamol

The mean paracetamol plasma concentration-time profiles and the respective 10<sup>th</sup> and 90<sup>th</sup> percentiles 294 295 are depicted in Figure 2. Under fasted conditions, double peaks in plasma concentration time-profiles 296 were observed in four subjects in the absorption phase with an evident impact on the mean profile 297 (Figure 2A). Similar double peak phenomenon could be observed in three subjects under fed 298 conditions, indicating inconsistent gastric emptying even under fed conditions. Since absorption of 299 paracetamol is controlled by gastric emptying (55–57), these observations indicate discontinuous 300 gastric emptying of suspension in some volunteers both in the fasted conditions and in the fed 301 conditions. The lack of the double-peak phenomenon under infant fed conditions could suggest 302 different gastric emptying mechanism for the formulation administered with infant formula.

303

Paracetamol total exposure (AUC<sub>0-10h</sub> or AUC<sub>0-inf</sub>) and C<sub>max</sub> and T<sub>max</sub> values were not significantly influenced by the prandial and dosing conditions applied in this study (**Table II**). Based on partial AUC values, early exposure under fasted conditions and fed conditions demonstrated no significant difference (**Table II**), in line with C<sub>max</sub> and T<sub>max</sub> data. However, under infant fed conditions, despite the lower total caloric content of infant formula (compared with the meal used to induce fed conditions), absorption of paracetamol was significantly slower than in the fasted state (p<0.05), regardless of the cut-off time point used for estimating the respective partial AUC (**Table II**).

311

Although there are no published food effect data acquired after administration of paracetamol suspension, data after administration of 1000 mg immediate-release (IR) paracetamol tablets indicate that fed conditions do not affect total exposure, while they decrease C<sub>max</sub> and increase T<sub>max</sub> values 315 (44,58,59). The apparently unaltered  $C_{max}$  and  $T_{max}$  values after administration under fed conditions 316 can be due to the low statistical power (0.049 for  $C_{max}$  comparison), the different gastric disposition of 317 a suspension vs. a tablet, and/or the presence of small amount of calories in the administered 318 suspension.

319

#### 320 Ibuprofen

The mean ibuprofen plasma concentration-time profiles and the respective 10<sup>th</sup> and 90<sup>th</sup> percentiles 321 322 are depicted in Figure 3. Double peaks were observed in the majority of individuals under fasted 323 conditions during the absorption phase, which was reflected in the mean plasma concentration-time 324 profile (Figure 3A). Under fed conditions, double peaks were observed in one subject (for the same 325 volunteer the phenomenon was also evident for paracetamol), while the occurrence during the 326 absorption phase was not clear under infant fed conditions. As for the paracetamol suspension, these 327 observations indicate a discontinuous gastric emptying process of the suspension in some volunteers, 328 primarily under fasted conditions.

329

330 Ibuprofen total exposure (AUC<sub>0-10h</sub> or AUC<sub>0-inf</sub>) appeared not to be significantly influenced by the 331 prandial and dosing conditions applied in this study (Table III). Differences in Cmax and Tmax values 332 between fasted conditions and fed conditions or between fasted conditions and infant fed conditions 333 were not significant. Interestingly, peak exposure (Cmax values) for ibuprofen administration with infant 334 formula was significantly greater than the observed under fed conditions (Table III). These data could 335 be related to initial slow absorption rates and a rapid increase at later times (Figure 3C). Drug dosing 336 under fed conditions significantly reduced early exposure compared to the fasted conditions during 337 the first 45 min after drug administration (Figure 3B). Early exposure was not significantly changed 338 when estimated up to longer times. Under infant fed conditions, all partial AUC values, e.g. AUC<sub>0-0.75h</sub>, 339 AUC<sub>0-1.5h</sub>, AUC<sub>0-3h</sub>, and AUC<sub>0-4h</sub>, were significantly lower compared to the fasted conditions (Table III). This observation is in line with the initial slow absorption rates and the increased absorption rates at later times that could have led to significantly greater C<sub>max</sub> values after infant formula (**Table III**).

342

343 To the best of our knowledge, there are no published data after administration of ibuprofen 344 suspensions under fed conditions. Data acquired for the administration of a 600 mg IR tablet suggest 345 no significant change in total exposure under fed conditions (orange juice included in the meal) (60). 346 However, total exposure (AUC<sub>0-inf</sub>) was decreased when ibuprofen IR tablets were administered at a 347 single dose of 400 mg under fed conditions (orange juice included in the meal) or 800 mg immediately 348 after a liquid test meal (61,62). It should be noted that in the published studies investigating IR tablets, 349 deviations from the fed conditions applied in the present investigation (and recommended by 350 regulators) were evident, e.g. co-administration of orange juice (60,61) and/or drug administration to 351 intubated volunteers 15 min after initiation of liquid meal consumption (62). Moreover, in these 352 studies, decreased C<sub>max</sub> and prolonged T<sub>max</sub> values have been reported after ibuprofen dosing under 353 fed conditions (60–62). As for the paracetamol observations in the present study, the apparently 354 unaltered  $C_{max}$  and  $T_{max}$  values after administration under fed conditions could be caused by the 355 different gastric disposition of suspension vs. the tablet and/or the presence of small amount of 356 calories in administered suspension.

## 357 Discussion

358 Today, oral paediatric formulation development is usually initiated during clinical Phase II stage of the 359 adult drug product timelines (3,63). Throughout the pharmaceutical design process for paediatric 360 formulations paramount emphasis is placed on formulation acceptability and palatability, resulting in 361 the common utilisation of sweeting agents in an attempt to improve the acceptance of paediatric liquid 362 formulations for oral administration (4). The present investigation showed that after administration of 363 paediatric suspension to adults under simulated infant fed conditions, but not under fed conditions, 364 the absorption of paracetamol and ibuprofen is substantially slower compared with the absorption 365 under fasted conditions.

366

In line with the typical excipients found in paediatric liquid formulations, sweetening agents, i.e. maltitol syrup and/or sorbitol, can be found among the excipients listed for the two paediatric suspensions investigated in the present study. Although the polyols included in these formulations exhibit lower caloric content compared to sucrose, and therefore, the total caloric content of the formulations is relatively low (ca. 60 kcal for the two formulations), a certain quantity of calories is inherently and inevitably administered under all studied prandial and dosing conditions.

373

The presence of calories in the formulations could raise concerns whether the subjects are in fasted conditions when these formulations are administered with a glass of water and what might be the possible implications of the caloric content of the formulations on physiological processes in the gastrointestinal tract, particularly regarding the regulation of gastrointestinal motility and gastric emptying. In an investigation performed using a liquid meal containing ca. 400 kcal, the motility phase in which the test meal was introduced, e.g. during quiescence (Phase I) or during late Phase II contractions, were found to be the major determinants for the motility response following meal 381 ingestion and gastric emptying rate (64). Meal administration during late Phase II of the migrating 382 motility complex (MMC) resulted in Phase III-like duodenal activity shortly after meal administration 383 accompanied by a biphasic gastric emptying pattern observed for the gastric emptying marker 384 paracetamol in most of the subjects, whereas meal ingestion during Phase I of the MMC lead to the 385 typical postprandial Phase II-like motility pattern associated with a monophasic pattern of gastric 386 emptying (64). Similar observations were reported when 60 kcal of the same liquid study meal were 387 infused intraduodenally during Phase I or late Phase II, demonstrating that the MMC could influence 388 postprandial responses and it is not entirely interrupted by nutrient simulation (65). In another study, 389 Thompson and colleagues reported that the ingestion of glucose solutions (50 g in 200 mL water) 390 during either MMC Phase I or II did not recognisably alter the appearance of the intestinal motor 391 pattern (66). Briefly, the quiescence phase continued to persist after glucose ingestion during MMC 392 Phase I period, while no apparent change of the duodenal irregular motor pattern or occurrence of 393 MMC Phase III was observed after ingestion of glucose solution during Phase II motor activity (66). The 394 authors concluded that the insignificant differences between MMC Phase III intervals of the two 395 timings of ingestion suggested that glucose ingestion would either produce the same delay in Phase III 396 re-appearance (despite differences in the timing of ingestion) or did not affect the appearance of Phase 397 III contractions, implying no interference of the glucose solution with the MMC (66).

398

399 Based on the insignificant impact of the caloric load of the suspension formulations, the apparently 400 discontinuous pattern of the gastric emptying process under fasted conditions could be related to the 401 variable contractual activity of the gastrointestinal tract and the characteristics of the administered 402 formulations. The double peak phenomenon could be associated with the viscosity enhancing 403 excipients in the formulations administered, e.g. xanthan gum. It could be assumed that the 404 insufficient ability of the suspensions to disperse in the stomach could lead to the emptying of 405 substantial amounts only under intense contractions. Interestingly, the time interval between these 406 double peaks, both after administration of paracetamol and ibuprofen in the fasted state, coincided

with the reported cycle of 1.5-2.5 hours for the peristaltic, phasic contractions of the migrating motility
complex (57,67). This possibility is in line with the wide use of paracetamol as a gastric emptying
marker after administration of rapidly disintegrating tablets or solutions (55) and the rare observation
of the double peak phenomenon in relevant previous works (68).

411

Under fed conditions, absorption rates did not change significantly from the ones observed under fasted conditions. This could be attributed either to the power underlying the statistical tests or the fast transfer of the drugs with the administered water into the small intestine, independently from the bulk gastric contents under fed conditions, a phenomenon known as "stomach road" or "Magenstrasse" (69,70). A pathway which may be less easily accessible for IR tablets, possibly due to the tablet disintegration step required prior to drug dissolution and mixing with the administered water that would enable the "Magenstrasse" pathway (71,72).

419

420 Perhaps the most interesting observations can be made from the comparison of infant fed vs. the 421 fasted state data. For both suspensions, unlike to the absorption rates under fed conditions, the 422 absorption rates under infant fed conditions were significantly slower than under fasted conditions. 423 Compared to the inhomogeneous viscous meal used for inducing fed conditions, the homogeneous 424 nature and low viscosity of the infant formula could facilitate mixing between the liquid drug formulation and infant formula and thus led to the emptying of the drug from the stomach with the 425 426 infant meal on a calorie-dependent basis (2). In fact, this slow absorption process led to detection of 427 significant difference in C<sub>max</sub> values for ibuprofen between fed and infant fed conditions (Table III).

428

Finally, from clinical perspective, the onset of pain relief and the timing of peak analgesic effects following paracetamol or ibuprofen intake profit from a faster rate of absorption. Assuming that the food type rather than age is the main determinant of gastric emptying (2,73), data from the present

- 432 study indicate a substantial delay in paracetamol or ibuprofen absorption and probably subsequent
- 433 delayed induction of pharmacodynamic effects when a suspension is administered during feed with
- 434 breastmilk or infant formula in infants.

# 435 Concluding remarks

436 The present exploratory study in healthy adults suggests that even for drugs with non-problematic 437 absorption (no intestinal permeability limitations, highly soluble in the small intestine, no documented 438 intraluminal interactions with food components) administered in simple dosage forms (aqueous 439 suspensions), food effects on drug absorption in infants may not be adequately evaluated by data 440 collected as suggested by regulatory agencies for adult drug products. It would be highly interesting to evaluate the extent to which differences between fasted conditions and infant fed conditions in adults 441 442 reflect differences between fasted state conditions and fed state conditions in infants. Until then, for 443 any drug product, food effects in infants should be considered cautiously or be evaluated in infants.

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# 453 References

455	1.	Ruiz BQ, Desfontaine E, Arenas-López S, Wang S. Pediatric formulation issues identified in
456		Paediatric Investigation Plans. Expert Rev Clin Pharmacol. 2014;7(1):25–30.
457	2.	Guimarães M, Statelova M, Holm R, Reppas C, Symilllides M, Vertzoni M, et al.
458		Biopharmaceutical considerations in paediatrics with a view to the evaluation of orally
459		administered drug products - a PEARRL review. J Pharm Pharmacol. 2019;71(4):603–42.
460	3.	Strickley RG. Pediatric oral formulations: an updated review of commercially available
461		pediatric oral formulations since 2007. J Pharm Sci. 2019;108(4):1335–65.
462	4.	European Medicines Agency (EMA). Guideline on pharmaceutical development of medicines
463		for paediatric use. Guid Doc [Internet]. 2013;44(May):1–23. Available from:
464		https://www.ema.europa.eu/en/documents/scientific-guideline/guideline-pharmaceutical-
465		development-medicines-paediatric-use_en.pdf
466	5.	European Medicines Agency (EMA). Guideline on the investigation of drug interactions. Guid
467		Doc [Internet]. 2012;44(June):1–59. Available from:
468		http://www.ema.europa.eu/docs/en_GB/document_library/Scientific_guideline/2012/07/WC
469		500129606.pdf
470	6.	Food and Drug Administration (FDA). Assessing the effects of food on drugs in INDs and NDAs-
471		clinical pharmacology considerations guidance for industry [Internet]. 2019. Available from:
472		http://www.fda.gov/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/default.ht
473		m
474	7.	Ginsburg CM, Mccracken GH, Thomas ML, Clahsen J, Ginsburg M, Thomas L, et al.
475		Comparative pharmacokinetics of amoxicillin and ampicillin in infants and children. Pediatrics.
476		1979;64(5):627–31.
477	8.	McCracken GH, Ginsburg CM, Clahsen JC, Thomas ML. Pharmacologic evaluation of orally

478 administered antibiotics in infants and children: effect of feeding on bioavailability. Pediatrics.

479 1978;62(5):738–43.

- 480 9. Kearns GL, Abdel-Rahman SM, Jacobs RF, Wells TG, Borin MT. Cefpodoxime pharmacokinetics
  481 in children: Effect of food. Pediatr Infect Dis J. 1998;17(9):799–804.
- 482 10. Tetzlaff TR, McCracken GH, Thomas ML. Bioavailability of cephalexin in children: Relationship
  483 to drug.formulations and meals. J Pediatr. 1978;92(February):292–4.
- 484 11. Finkel Y, Bolme P, Eriksson M. The Effect of Food on the Oral Absorption of Penicillin V
  485 Preparations in Children. 1981;301–4.
- 486 12. Okuno A, Taguchi T, Inyaku F, Yano K, Suzuki Y. Pharmacokinetics of propylthiouracil in
- 487 children and adolescents with Graves disease after a single oral dose. Pediatr Pharmacol.
  488 1983;3(1):43–7.
- Pedersen S, Møller-Petersen J. The Influence of Food on the Bioavailability of a Sustained
  Release Theophylline Formulation. Allergy. 1982;37:531–4.
- 491 14. S. Pedersen. Dealay in the absorption rate of theophylline from a sustained release
- theophylline preparation caused by food. Br J Clin Pharmacol. 1981;(12):904–5.
- 493 15. Pedersen S, Møller-Petersen J. Erratic Absorption of a Slow-Release Theophylline Sprinkle
  494 Product. Pediatrics. 1984;74(4):534–8.
- 495 16. Pedersen S. Absorption of Theo-Dur sprinkle with food: importance of types of meals and
  496 medication times. J Allergy Clin Immunol. 1986;78(4 Part 1):653–60.
- 497 17. Steffensen G, Pedersen S. Food induced changes in theophylline absorption from a once-a-day
  498 theophylline product. Br J Clin Pharmacol. 1986;22(5):571–7.
- 499 18. Lancaster DL, Patel N, Lennard L, Lilleyman JS. 6-Thioguanine in children with acute
- 500 lymphoblastic leukaemia: Influence of food on parent drug pharmacokinetics and 6-
- 501 thioguanine nucleotide concentrations. Br J Clin Pharmacol. 2001;51(6):531–9.
- 502 19. Batchelor H. Influence of food on paediatric gastrointestinal drug absorption following oral

- 503 administration: a review. Children. 2015;2(2):244–71.
- 504 20. Gan VY, Chu S-Y, Kusmiesz HT, Craft JC. Pharmacokinetics of a clarithromycin suspension in
   505 infants and children. Antimicrob Agents Chemother. 1992;36(11):2478–80.
- 506 21. Stevens RC, Rodman JH, Yong FH, Carey V, Knupp CA, Frenkel LM. Effect of food and
- 507 pharmacokinetic variability on didanosine systemic exposure in HIV-infected children.
- 508 Pediatric AIDS Clinical Trials Group Protocol 144 Study Team. AIDS Res Hum Retroviruses.
- 509 2000;16(5):415-21.
- 510 22. Ginsburg CM, McCracken GH, Petruska M, Olsen K. Effect of feeding on bioavailability of
  511 griseofulvin in children. J Pediatr. 1983;102(2):309–11.
- 512 23. Borrmann S, Sallas WM, Machevo S, González R, Björkman A, Mårtensson A, et al. The effect
- of food consumption on lumefantrine bioavailability in African children receiving artemether-
- 514 Iumefantrine crushed or dispersible tablets (Coartem<sup>®</sup>) for acute uncomplicated Plasmodium
- 515 falciparum malaria. Trop Med Int Heal. 2010;15(4):434–41.
- 516 24. Riccardi R, Balis FM, Ferrara P, Poplack DG, Mastrangelo R. Influence of food intake on
- 517 bioavailability of oral 6-mercaptopurine in children with acute lymphoblastic leukemia.
- 518 Pediatr Hematol Oncol. 1986;3(4):319–24.
- 519 25. Sofianou-Katsoulis A, Khakoo G, Kaczmarski R. Reduction in Bioavailability of 6-
- 520 Mercaptopurine on Simultaneous Administration With Cow'S Milk. Pediatr Hematol Oncol.
- 521 2006;23(6):485–7.
- 522 26. Lonnerholm G, Kreuger A, Lindstrom B, Myrdal U. Oral mercaptopurine in childhood leukemia:
- 523 influence of food intake on bioavailability. Pediatr Hematol Oncol. 1989;6(2):105–12.
- 524 27. Pinkerton CR, Glasgow JFT, Welshman SG, Bridges JM. Can food influence the absorption of
- 525 methotrexate in children with acute lymphoblastic leukaemia? Lancet. 1980;2(8201):944–6.
- 526 28. Pedersen S, Steffensen G. Food and Fasting Absorption of a Single Dose of a Sustained Release
- 527 Theophylline Sprinkle Formulation in Children. Allergy. 1986;41(1):46–50.

- Welling PG, Huang H, Koch PA, Craig WA, Madsen PO. Ampicillin and amoxicillin in fasted and
  nonfasted subjects. J Pharm Sci. 1977;66(4):549–52.
- 530 30. Eshelman FN, Spyker DA. Pharmacokinetics of amoxicillin and ampicillin. Crossover study of
  531 the effect of food. Antimicrob Agents Chemother. 1978;14(4):539–43.
- 532 31. McCarthy CG, Finland M. Absorption and excretion of four penicillins penicillin G, penicillin V,
- 533 phenethicillin and phenylmercaptomethyl penicillin. N Engl J Med. 1960;263(7):315–26.
- 534 32. Cronk GA, Wheatley WB, Fellers GF, Albright H. The relationship of food intake to the
- absorption of potassium alpha-phenoxyethyl penicillin and potassium phenoxymethyl
- penicillin from the gastrointestinal tract. Am J Med Sci. 1960;240(August):219–25.
- 537 33. Welling PG. Influence of food and diet on gastrointestinal drug absorption: a review. J
- 538 Pharmacokinet Biopharm. 1977;5(4).
- 539 34. Khuroo AH, Monif T, Verma PRP, Gurule S. Comparison of effect of fasting and of five different
  540 diets on the bioavailability of single oral dose of amoxicillin 500 mg capsule. Clin Res Regul Aff.
  541 2008;25(2):73–86.
- 542 35. Gower E, Dash CH. Cephalexin : human studies of absorption and excretion of a new
  543 cephalosporin antibiotic. Br J Pharmacol. 1969;37:738–47.
- 54436.Thornhill TS, Levison ME, Johnson WD, Kaye D. In vitro antimicrobial activity and human545pharmacology of cephalexin, a new orally absorbed cephalosporin C antibiotic. Appl
- 546 Microbiol. 1969;17(3):457–61.
- 547 37. Speight TM, Brogden RN, Avery GS. Cephalexin : a review of its antibacterial, pharmacological
  548 and therapeutic properties. Drugs. 1972;3(1):9–78.
- 549 38. Pfeffer M, Jackson A, Ximenes J, Menezes JPDE. Comparative human oral clinical
- 550 pharmacology of cefadroxil, cephalexin, and cephradine. Antimicrob Agents Chemother.
- 551 1977;11(2):331–8.
- 552 39. Lecaillon JB, Hirtz JL, Schoeller IJP, Humbert GUY, Vischer W. Pharmacokinetic comparison of

- 553 cefroxadin (CGP 9000) and cephalexin by simultaneous administration to humans. Antimicrob
  554 Agents Chemother. 1980;18(4):656–60.
- Welling PG, Elliott RL, Pitterle ME, Lyons LL. Plasma levels following single and repeated doses
  of erythromycin estolate and erythromycin stearate. J Pharm Sci. 1979;68(2):150–5.

557 41. Potthast H, Dressman JB, Junginger HE, Midha KK, Oeser H, Shah VP, et al. Biowaiver

558 monographs for immediate release solid oral dosage forms: Ibuprofen. J Pharm Sci.

559 2005;94(10):2121–31.

- Wu CY, Benet LZ. Predicting drug disposition via application of BCS: Transport/absorption/
  elimination interplay and development of a biopharmaceutics drug disposition classification
  system. Pharm Res. 2005;22(1):11–23.
- 43. European Medicines Agency E. Ibuprofen oral use immediate release formulations 200 800
- 564 mg product-specific bioequivalence guidance [Internet]. 2018. p. 1–4. Available from:
- 565 https://www.ema.europa.eu/en/documents/scientific-guideline/ibuprofen-oral-use-
- 566 immediate-release-formulations-200-800-mg-product-specific-bioequivalence\_en.pdf
- 567 44. Atkinson HC, Stanescu I, Frampton C, Salem II, Beasley CPH, Robson R. Pharmacokinetics and
- 568 bioavailability of a fixed-dose combination of ibuprofen and paracetamol after intravenous
- and oral administration. Clin Drug Investig. 2015;35(10):625–32.
- 570 45. Wright CE, Antal EJ, Gillespie WR, Albert KS. Ibuprofen and acetaminophen kinetics when
- 571taken concurrently. Clin Pharmacol Ther. 1983;34(5):707–10.
- 572 46. GlaxoSmithKline Consumer Healthcare Ltd. Summary of product characteristics Panadol baby
  573 suspension [Internet]. 2017. Available from:
- 574 https://www.hpra.ie/HOMEPAGE/medicines/medicines-information/find-a-
- 575 medicine/results/item?change=6301193&pano=PA0678/039/003&t=PANADO...1/2
- 576 47. ReckittBenckiser Healthcare International Ltd. Nurofen Junior Suspension: Summary of
- 577 product characteristics for healthcare professionals [Internet]. Available from:
- 578 https://www.gelbe-liste.de/produkte/Nurofen-Junior-Fiebersaft-Erdbeer-2-Suspension-zum-

- 579 Einnehmen 508519/fachinformation
- European Pharmacopoeia PE. Maltitol, Liquid Maltitolum liquidum. 2008. 2332–2333 p. 580 48.
- 581 49. World Medical Association (WMA). WMA Declaration of Helsinki 1975 – ethical principles for 582 scientic requirements and research protocols. 2013. p. 29–32.
- 583 50. ICH GCP E6. Guildeline for Good Clinical Practice E6(R1) [Internet]. Vol. 1996, ICH harmonised
- 584 tripartite guideline. 1996. Available from: http://academy.gmp-
- 585 compliance.org/guidemgr/files/E6\_R1\_GUIDELINE.PDF
- 586 51. Food and Drug Administration (FDA). Guidance for industry Food-effect bioavailability and fed bioequivalence studies. 2002; (December). Available from: 587
- 588 https://www.fda.gov/downloads/drugs/guidancecomplianceregulatoryinformation/guidances /ucm070241.pdf 589
- 590 52. Vertzoni M V., Archontaki HA, Galanopoulou P. Development and optimization of a reversed-
- 591 phase high-performance liquid chromatographic method for the determination of
- 592 acetaminophen and its major metabolites in rabbit plasma and urine after a toxic dose. J
- 593 Pharm Biomed Anal. 2003;32(3):487–93.
- 594 Lalande M, Wilson DL, Mcgilveray IJ. Rapid high-performance in human plasma. J Chromatogr 53.
- 595 B. 1986;377:410-4.
- 596 Food and Drug Administration (FDA). Bioavailability studies submitted in NDAs or INDs-54.

general considerations guidance for industry [Internet]. 2019. Available from: 597

- 598 http://www.fda.gov/Drugs/GuidanceComplianceRegulatoryInformation/Guidances/default.ht m
- 599
- 600 55. Willems M, Quartero AO, Numans ME. How useful is paracetamol absorption as a marker of 601 gastric emptying? A systematic literature study. Dig Dis Sci. 2001;46(10):2256–62.
- 602 56. Wilson CG, Clarke CP, Starkey YYL, Clarke GD, Clarke CP, Starkey YYL, et al. Comparison of a
- 603 novel fast-dissolving acetaminophen tablet formulation (FD-APAP) and standard
- 604 acetaminophen tablets using gamma scintigraphy and pharmacokinetic studies. Drug Dev Ind

605 Pharm. 2011;37(7):747–53.

- 57. Van Den Abeele J, Rubbens J, Brouwers J, Augustijns P. The dynamic gastric environment and
  its impact on drug and formulation behaviour. Eur J Pharm Sci. 2017;96:207–31.
- 58. Stillings M, Havlik I, Chetty M, Clinton C, Schall R, Moodley I, et al. Comparison of the
- 609 pharmacokinetic profiles of soluble aspirin and solid paracetamol tablets in fed and fasted
- 610 volunteers. Curr Med Res Opin. 2000;16(2):115–24.
- 59. Rostami-Hodjegan A, Shiran MR, Ayesh R, Grattan TJ, Burnett I, Darby-Dowman A, et al. A new
- 612 rapidly absorbed paracetamol tablet containing sodium bicarbonate. I. A four-way crossover
- 613 study to compare the concentration-time profile of paracetamol from the new
- 614 paracetamol/sodium bicarbonate tablet and a conventional paracetamol tablet in fed. Drug
- 615 Dev Ind Pharm. 2002;28(5):523–31.
- 616 60. Levine M, Walker S, Paton T. The effect of food or Sucralfate on the bioavailability of S(+) and
  617 R(-) enantiomers of ibuprofen. J Clin Pharmacol. 1992;32:1110–4.
- 618 61. Klueglich M, Ring A, Scheuerer S, Trommeshauser D. Ibuprofen extrudate, a novel, rapidly
- dissolving ibuprofen formulation: relative bioavailability compared to ibuprofen lysinate and
- regular ibuprofen, and food effect on all formulations. J Clin Pharmacol. 2005;(45):1055–61.
- 621 62. Koenigsknecht M, Sun D, Baker JR, Wen B, Frances A, Zhang H, et al. In vivo dissolution and
- systemic absorption of immediate release ibuprofen in human gastrointestinal tract under fed
  and fasted conditions. Mol Pharm. 2017;14(12):4295–304.
- 624 63. Batchelor H, Kaukonen AM, Klein S, Davit B, Ju R, Ternik R, et al. Food effects in paediatric
- medicines development for products co-administered with food. Int J Pharm. 2018
  Feb;536(2):530–5.
- 627 64. Medhus A, O. Sandstad, Brede J, Husebye E. The migrating motor complex modulates
  628 intestinal motility response and rate of gastric emptying of caloric meals. Neurogastroenterol
- 629 Motil. 1995;7(1):1–8.
- 630 65. Medhus A, Sandstad O, Brede J, Husebye E. Stimulation of the Small Intestine by Nutrients in

- Relation to Phase of the Migrating Motor Complex. Scand J Gastroenterol. 2000;35(5):494–
  500.
- 633 66. Thompson DG, Wingate DL, Thomas M, Harrison D. Gastric emptying as a determinant of the
  634 oral glucose tolerance test. Gastroenterolog. 1982;82(1):51–5.
- 635 67. Hens B, Corsetti M, Spiller R, Marciani L, Vanuytsel T, Tack J, et al. Exploring gastrointestinal
- 636 variables affecting drug and formulation behavior : Methodologies , challenges and
- 637 opportunities. Int J Pharm. 2017;519(1–2):79–97.
- 638 68. Clements J, Heading R, Nimmo W, Prescott L. Kinetics of acetaminophen absorption and
  639 gastric emptying in man. Clin Pharmacol Ther. 1978;24(4):420–31.
- 640 69. Koziolek M, Grimm M, Garbacz G, Weitschies W. Intragastric volume changes after intake of a
- 641 high-caloric, high-fat standard breakfast in healthy human subjects investigated by MRI. Mol
- 642 Pharm. 2014;11(5):1632–9.
- 643 70. Grimm M, Koziolek M, Kühn J, Weitschies W. Interindividual and intraindividual variability of
  644 fasted state gastric fl uid volume and gastric emptying of water. Eur J Pharm Biopharm.
- 645 2018;127(February):309–17.
- 646 71. Kalantzi L, Polentarutti B, Albery T, Laitmer D, Abrahamsson B, Dressman J, et al. The delayed
- 647 dissolution of paracetamol products in the canine fed stomach can be predicted in vitro but it
- 648 does not affect the onset of plasma levels. Int J Pharm. 2005;296(1–2):87–93.
- Abrahamsson B, Albery T, Eriksson A, Gustafsson I, Sjöberg M. Food effects on tablet
  disintegration. Eur J Pharm Sci. 2004 Jun;22(2–3):165–72.
- 651 73. Bonner JJ, Vajjah P, Abduljalil K, Jamei M, Rostami-Hodjegan A, Tucker GT, et al. Does age
- affect gastric emptying time? A model-based meta-analysis of data from premature neonates
- through to adults. Biopharm Drug Dispos. 2015;36(4):245–57.
- 654
- 655

# **Table I** Published food effect data for seven antibiotic suspensions.

	Food effects in infants and pre-school children						Food effects in adults				
	Food				AUC <sub>0-6h</sub> <sup>a</sup>		a x	Refere	Food	Effect on	
Drug	effect	(μg/ι		(µg/m		(h		nce	effect	C <sub>max</sub> , AUC,	Refere
	S	Fast ed	Fe d	Fast ed	Fe d	Fast ed	Fe d		S	and $T_{\text{max}}$	nce
A		6.4	6. 1	18	25	1.0	2.	(8)		C <sub>max</sub> and AUC <sub>0-t</sub> significantly lower; T <sub>max</sub> prolonged on average	(29)
Ampicilli n	Unlik ely	5.0	4. 1	12	12	1.0	1. 0	(7)	Negat ive	C <sub>max</sub> lower on average; AUC <sub>0-t</sub> significantly lower T <sub>max</sub> significantly delayed	(30)
Penicillin G	Likely negat ive	0.98	0. 61	1.7	1. 0	0.5	0. 5	(8)	Uncle ar	C <sub>max</sub> 22% lower on average; AUC <sub>0-t</sub> unchanged ("long- acting" tablet); T <sub>max</sub> prolonged on average	(31)
										AUC <sub>0-2h</sub> significantly lower C <sub>max</sub> 20% and AUC <sub>0-t</sub> 35% higher on average; T <sub>max</sub> prolonged	(32)
Penicillin V	Likely negat ive	2.1	1. 1	3.0	1. 9	0.5	0. 5	(8)	Uncle ar	on average C <sub>max</sub> significantly lower; T <sub>max</sub> prolonged on average urine recovery 10% lower	(33)

	-							-			1
		5.4	3. 2	16	14	1.0	1. 5	(7) <sup>b</sup>		C <sub>max</sub> and AUC <sub>0-t</sub> unchanged T <sub>max</sub> significantly delayed	(30)
Amoxicill in	Unlik ely		7.	24	24	1.0	1.	(7) <sup>c</sup>	Likely negat ive	C <sub>max</sub> and AUC <sub>0-t</sub> significantly lower; T <sub>max</sub> prolonged on average	(29)
		0.5	9	27	2-	1.0	0			C <sub>max</sub> and AUC <sub>0-t</sub> significantly lower; T <sub>max</sub> not significantly prolonged	(34)
Cephalex	Likely	Likely			22				Unlik	C <sub>max</sub> unchanged; AUC <sub>0-t</sub> unchanged; T <sub>max</sub> unchanged/s lightly prolonged	(35–38)
in	negat ive	23.4	9. 0	40.0	23 .0	0.5	1.	(8)	ely	C <sub>max</sub> 40% lower on average; AUC <sub>0-t</sub> 10% lower on average; T <sub>max</sub> prolonged on average	(39)
Erythrom ycin Estolate	Unlik ely	4.7	4. 8	45	40	2.0	2. 0	(8)	Positi ve	C <sub>max</sub> and AUC <sub>0-t</sub> significantly increased; T <sub>max</sub> significantly delayed	(40)
Erythrom ycin Ethyl- succinate	Likely positi ve	0.82	1. 4	2.4	4. 8	1.0	1. 0	(8)	Likely positi ve	Serum levels to 12 hr post-dosing increased on average	(33)

 $^{a}$  C<sub>max</sub>, AUC<sub>0-6</sub> (µg/mL·h), and T<sub>max</sub> values from the mean plasma profiles were published in studies in infants

659 <sup>b</sup> Amoxicillin dose 15 mg/kg; <sup>c</sup> Amoxicillin dose 25 mg/kg

660 **Table II** Mean ± SD values of pharmacokinetic parameters for paracetamol in each phase of the clinical

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study.

Parameter	Phase I Fasted conditions	Phase II Fed conditions	Phase III Infant fed conditions		
AUC <sub>0-inf</sub> (µg/mL×h)	39.4 ± 9.7	$40.4 \pm 11.0$	39.2 ± 10.1		
AUC <sub>0-10h</sub> (µg/mL×h)	35.8 ± 7.9	35.5 ± 8.9	34.0 ± 8.0		
C <sub>max</sub> (µg/mL)	7.85 ± 1.54	6.96 ± 2.42	7.24 ± 1.32		
T <sub>max</sub> (h)	1.5 (0.33 - 4) <sup>a</sup>	2.5 (1.0 - 5) <sup>a</sup>	4 (1.5 - 5) <sup>a</sup>		
AUC <sub>0-1.5h</sub> (µg/mL×h)	6.78 ± 3.14	5.27 ± 2.99	2.12 ± 1.37 <sup>b</sup>		
AUC <sub>0-2.5h</sub> (μg/mL×h)	12.7 ± 4.4	10.5 ± 4.8	5.81 ± 2.72 <sup>b</sup>		
AUC <sub>0-4h</sub> (µg/mL×h)	21.4 ± 5.2	18.5 ± 5.9	13.7 ± 4.3 <sup>b</sup>		

662 <sup>a</sup> median value (range)

663 <sup>b</sup> significantly different from Phase I

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Table III Mean ± SD values of pharmacokinetic parameters for ibuprofen in each phase of the clinical
 study.

Parameter	Phase I Fasted conditions	Phase II Fed conditions	Phase III Infant fed conditions		
AUC <sub>0-inf</sub> (µg/mL×h)	205 ± 60	203 ± 47	213 ± 54		
AUC <sub>0-10h</sub> (µg/mL×h)	192 ± 50	185 ± 40	194 ± 44		
C <sub>max</sub> (μg/mL)	45.0 ± 7.4	41.3 ± 10.6	49.6 ± 9.0 <sup>c</sup>		
T <sub>max</sub> (h)	0.75 (0.33 – 4) <sup>a</sup>	1.5 (1.0 – 3) <sup>a</sup>	3.3 (0.33 – 5) <sup>a</sup>		
AUC <sub>0-0.75h</sub> (µg/mL×h)	19.4 ± 8.2	10.8 ± 6.5 <sup>b</sup>	7.7 ± 9.0 <sup>b</sup>		
AUC <sub>0-1.5h</sub> (µg/mL×h)	46.7 ± 15.6	32.6 ± 19.6	18.6 ± 17.4 <sup>b</sup>		
AUC <sub>0-3h</sub> (µg/mL×h)	96.9 ± 21.0	80.5 ± 34.4	52.6 ± 29.2 <sup>b</sup>		
AUC <sub>0-4h</sub> (µg/mL×h)	126 ± 25	109 ± 36	85.2 ± 29.4 <sup>b</sup>		

- 668 <sup>a</sup> median value (range)
- 669 <sup>b</sup> significantly different from Phase I
- 670 <sup>c</sup> significantly different from Phase II
- 671
- 672

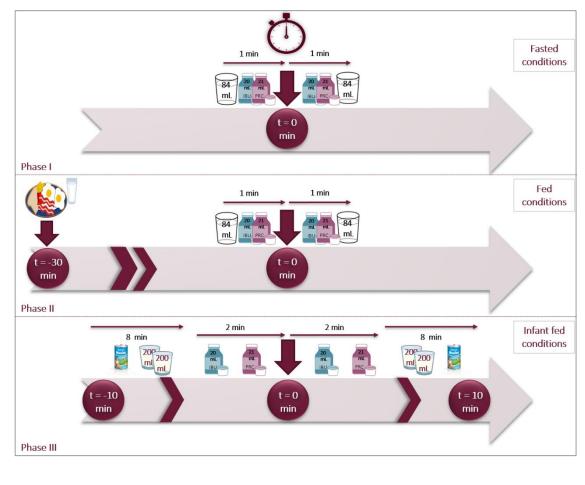
### 674 Figure Captions

Figure 1 Graphical depiction of the times of meals vs. drug products administrations in the present
 clinical study: Phase I, fasted conditions; Phase II, fed conditions; Phase III, infant fed conditions.

**Figure 2** Mean plasma paracetamol concentration-time profiles following co-administration of 1000 mg paracetamol suspension and 800 mg ibuprofen suspension to healthy male adults (n=8) under different prandial and dosing conditions: (A) fasted conditions, (B) fed conditions, (C) infant fed conditions. The shaded area represents the 10<sup>th</sup> and 90<sup>th</sup> percentiles estimated from the experimental data points.

**Figure 3** Mean plasma ibuprofen concentration-time profiles following co-administration of 1000 mg paracetamol suspension and 800 mg ibuprofen suspension to healthy male adults (n=8) under different prandial and dosing conditions: (A) fasted conditions, (B) fed conditions, (C) infant fed conditions. The shaded area represents the 10<sup>th</sup> and 90<sup>th</sup> percentiles estimated from the experimental data points.

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- 690 Figure 1

