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Defying the 24-h day: Unexpected diversity in socially synchronized rhythms of shorebirds

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All organisms exhibit alternating phases of different behaviours. These biological rhythms are thought to be under strong selection, influenced by the rhythmicity of the environment. Behavioural rhythms are well studied in isolated individuals under laboratory conditions^{1,2}, but in free-living populations, individuals have to temporally synchronize their activities with those of others. The behavioural rhythms that emerge from such social synchronization and the underlying evolutionary and ecological drivers that shape them are poorly understood²⁻⁵. Here, we address this for a particularly sensitive phase of social synchronization when pair members care for their offspring⁶, and thereby potentially compromise their individual rhythms. Specifically, we use data from 729 nests of 91 populations of 32 biparentally-incubating shorebird species, where parents synchronize to achieve continuous coverage of developing eggs. We found remarkable within- and between-species diversity in incubation rhythms. Between species, the median length of one parent's incubation bout varied from one to 19 hours, while period length – the cycle of female and male probability to incubate – varied from six to 43 hours. The length of incubation bouts was unrelated to variables reflecting energetic demands, but species relying on crypsis had longer incubation bouts than those that are readily visible or actively protect their nest against predators. Rhythms strictly entrainable to the 24-h light-dark

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26 **cycle were less likely at high latitudes and absent in 18 species. Our results indicate that even under similar**
27 **environmental conditions and despite 24-h environmental cues, social synchronization can generate far more**
28 **diverse behavioural rhythms than expected from studies of individuals in captivity. The risk of predation, not**
29 **the risk of starvation, may be a key factor underlying the diversity in these rhythms.**

30 The activity rhythms of organisms, from bacteria to humans, are typically the output of internal
31 'biological clocks' synchronized to the daily environmental cycle¹. These biological clocks and the resulting
32 rhythms are largely genetically determined and conserved across species⁷⁻⁹. Yet, behavioural rhythms of
33 individuals are further shaped by the social environment, including potential mates, allies, competitors, prey, or
34 predators^{3-5,10,11}. As a result of such social interactions, individuals can temporally segregate their daily activities
35 (e.g. prey avoiding predators, subordinates avoiding dominants) or synchronize their activities (e.g. foraging in
36 group, communal defence against predators, pairs reproducing or caring for offspring)^{3-5,12,13}. Although social
37 synchronization of conspecifics has been documented across taxonomic groups²⁻⁵ and although the behavioural
38 rhythms that emerge from social synchronization have occasionally been studied in captive individuals^{2-5,14}, we
39 know little about socially emerged rhythms in nature^{3-5,10,11}, and even less about the factors that influence them
40 such as evolutionary history, energetic demands, predation risk and periodicity in the environment.

41 Here, we report on a large-scale study of socially emerged rhythms and their correlates in the wild,
42 using biparental incubation in shorebirds as a model. Incubation by both parents prevails in almost 80% of non-
43 passerine families¹⁵ and is the most common form of care in shorebirds¹⁶. Biparental shorebirds are typically
44 monogamous¹⁷, mostly lay three or four eggs in an open¹⁷ nest on the ground¹⁷, and cover their eggs almost
45 continuously¹⁵. Pairs achieve this through synchronization of their activities such that one of them is responsible
46 for the nest at a given time (i.e. an incubation bout). Alternating female and male bouts generate an incubation
47 rhythm with a specific period length (cycle of high and low probability for a parent to incubate).

48 We collected data on incubation rhythms from 91 populations of 32 shorebird species belonging to 10
49 genera (Fig. 1), extracted the length of 34,225 incubation bouts from 729 nests, and determined the period
50 length for pairs in 584 nests (see Extended Data Fig. 1).

51 We found vast between- and within- species variation in incubation bout length and in period length
52 (Fig. 2-4 & Extended Data Fig. 2). Different species, but also different pairs of the same species, adopted
53 strikingly different incubation rhythms, even when breeding in the same area (see, e.g. incubation rhythms in
54 Barrow, Alaska, represented by ① in Fig. 2; incubation rhythms for each nest are in Supplementary
55 Actograms¹⁸). Whereas in some pairs parents exchanged incubation duties about 20 times a day (Fig. 3a; e.g.
56 *Charadrius semipalmatus*, Fig. 2a), in others a single parent regularly incubated for 24 hours (Fig. 3a; e.g.
57 *Limnodromus scolopaceus*, Fig. 2a), with exceptional bouts of up to 50 hours (Supplementary Actograms¹⁸).
58 Similarly, whereas incubation rhythms of pairs in 22% of nests followed a strict 24-h period (Fig. 3b; e.g. *Tringa*
59 *flavipes*, Fig. 2a), the rhythms of others dramatically deviated from a 24-h period (Fig. 3b) resulting in ultradian
60 (<20-h in 12% of nests; e.g. *Numenius phaeopus*; Fig. 2a), free-running like (e.g. *Calidris alpina*; Fig. 2a) and
61 infradian rhythms (>28-h in 8% of nests), some with period lengths up to 48-h (e.g. *Limnodromus scolopaceus*;
62 Fig. 2a). This variation in period length partly related to the variation in bout length (Fig. 4): in the suborder
63 Scolopaci period length correlated positively with median bout length, but in the suborder Charadrii species
64 with 24-h periods had various bout lengths, and species with similar bout lengths had different period lengths
65 (Fig. 4).

66 Despite substantial within-species variation, we found a strong evolutionary signal for both bout and
67 period length with a coefficient of phylogenetic signal λ close to 1 (Extended Data Table 1). This is consistent
68 with the notion that biological rhythms are largely genetically determined and conserved among related
69 species⁸⁻¹⁰. However, the phylogenetic effect seems unevenly distributed over taxonomic level. Suborder
70 explained 33% of the phenotypic variance in both bout and period length, with the Scolopaci having longer
71 incubation bouts and periods than the Charadrii (Extended Data Table 2; Fig. 3 & 4). Species explained 41% of
72 the phenotypic variation in bout length and 46% in period length, but genus explained little (<1% in both;
73 Extended Data Table 2), suggesting that despite a strong phylogenetic signal, these traits can rapidly diverge
74 (Fig. 3c).

75 Two ecological factors may explain the observed variation in bout length. First, the 'energetic demands
76 hypothesis' stipulates that the length of an incubation bout depends on a bird's energetic state^{15,19}. This

77 predicts that (1) large species will have longer incubation bouts than smaller species, because they radiate less
78 body heat per unit of mass and (2) incubation bouts will shorten with increasing breeding latitude, because –
79 everything else being equal – energy stores will deplete faster in colder environments (Extended Data Fig. 3a-b
80 shows latitudinal cline in summer temperatures). However, bout length was unrelated to body size (Fig. 5a) and
81 correlated positively (instead of negatively) with latitude (Fig. 5b). These correlational results across populations
82 and species support recent experimental findings within species²⁰ and suggest that in biparentally-incubating
83 shorebirds energetic demands are not an important ecological driver underlying variation in bout length.

84 An alternative explanation for variation in the length of incubation bouts relates to anti-predation
85 strategies. Those species that rely primarily on parental crypsis (Extended Data Fig. 4a) benefit from reduced
86 activity near the nest, because such activity can reveal the nest's location to potential predators^{21,22}. Thus, in
87 these species, selection will favour fewer change-overs at the nest, and hence longer incubation bouts. In
88 contrast, species that are clearly visible when sitting on the nest or that rely on active anti-predation behaviour
89 (Extended Data Fig. 4b), including having a partner on the watch for predators, leaving the nest long before the
90 predator is nearby and mobbing the predator¹⁷, obtain no advantage from minimizing activity. For these
91 species, bout length can shorten, which may be advantageous for other reasons (e.g. reduced need to store fat).
92 We quantified anti-predation strategy as the distance at which the incubating parent left the nest when
93 approached by a human (escape distance), because cryptic species stay on the nest longer (often until nearly
94 stepped upon)¹⁷. Despite the large geographical distribution of the studied species, with related variability in
95 the suite of predators and predation pressure²³, and even when controlling for phylogeny (which captures much
96 of the variation in anti-predation strategy, Extended data Fig. 5), escape distance negatively correlated with the
97 length of incubation bouts (Fig. 5c). This result suggests that bout length co-evolved with the anti-predation
98 strategy.

99 Under natural conditions, most organisms exhibit 24-h rhythmicity, but during summer, when most
100 shorebirds breed, the 24-h variation in light decreases with latitude leading to continuous polar daylight²⁴
101 (Extended Data Fig. 4c-d). Such reduced variation in 24-h light intensity may cause a loss of 24-h rhythmicity^{25,26}.
102 As a consequence, circadian behavioural rhythms should exhibit a latitudinal cline²⁴. As predicted, incubation

103 rhythms with periods that do not follow the 24-h light-dark cycle, such as ‘free running-like patterns’ (left
104 column in Fig. 2), occurred more often in shorebirds breeding at higher latitudes (Fig. 5d). The absolute
105 deviations of periods from 24-h and 24-h harmonics also increased with latitude (Fig. 5e; Extended Data Table
106 3). Although this supports the existence of a latitudinal cline in socially emerged behavioural rhythms²⁴, we
107 found a substantial number of rhythms that defy the 24-h day even at low and mid latitudes (Fig 5d-e).

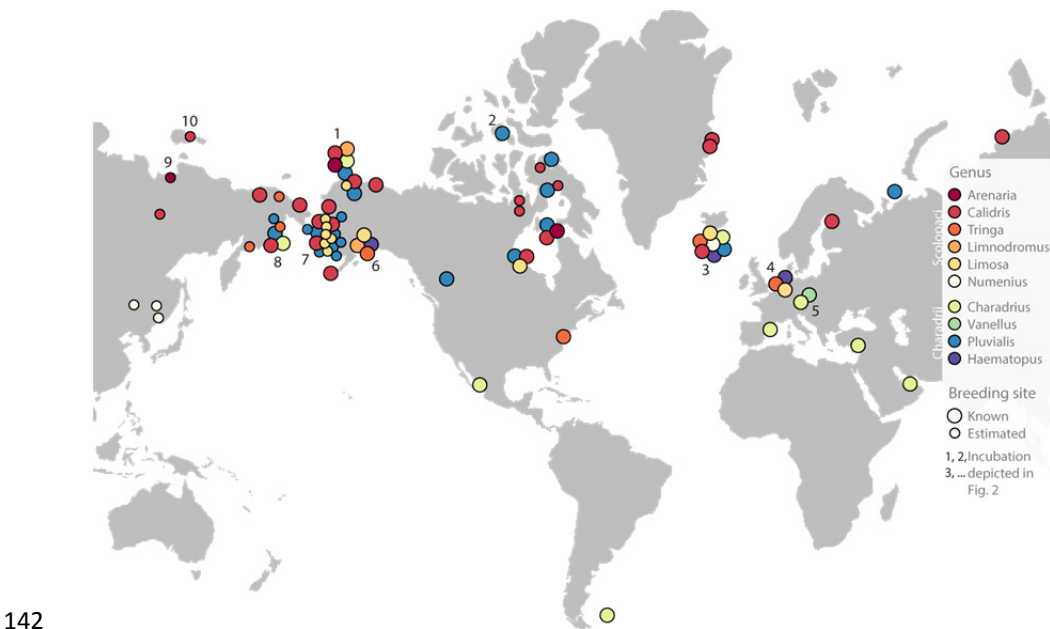
108 Many shorebirds predominantly use tidal habitats, at least away from their breeding ground¹⁷. To
109 anticipate tidal foraging opportunities, these species may have activity patterns with a period length resembling
110 the tidal period. As changing to a different rhythm is costly²⁷, these tidal activity patterns might carry over to
111 incubation. Although half of our species are tidal away from their breeding grounds, and some forage in tidal
112 areas also during breeding (~10% of populations), in only 5% of nests did pairs display a period length that can
113 be entrained by the tide. Moreover, tidal species had similar (not longer) periods than non-tidal ones (Extended
114 Data Table 3). Hence, unlike the 24-h light-dark cycle, tidal life-history seems to play at best a negligible role in
115 determining incubation rhythms.

116 Three main questions arise from our results. First, is variation in incubation bout length in cryptic
117 species related to the actual predation pressure? This can be tested by comparing bout length between
118 populations of a particular species that are exposed to different predator densities, or between years that differ
119 in predation pressure. Second, it remains unclear how the diverse social rhythms emerge. Are these rhythms a
120 consequence of behavioural flexibility, or a ‘fixed’ outcome of synchronization between the circadian clocks of
121 the two individuals involved? An experimental study on ring doves (*Streptopelia risoria*) suggests that parents
122 may even use two timers - circadian oscillation and interval timing - to determine when to incubate²⁸. Parents
123 rapidly adjusted their schedules to phase-shifted photoperiods and their incubation rhythm ‘free-ran’ in
124 constant dim illumination (implying a circadian mechanism), whereas an experimental delay in the onset of an
125 incubation bout did not change the length of the bout because the incubating parent refused to leave the nest
126 until its incubation bout reached the ‘typical’ duration (implying interval timing). Third, what – if any – are the
127 fitness consequences for the parents of having a certain incubation rhythm? For example, the costs of having a
128 particular incubation rhythm may be unevenly distributed between the two parents (e.g. because one parent is

129 on incubation duty when food is more readily available, or because one parent ‘enforces’ its own rhythm at a
130 cost to the other parent).

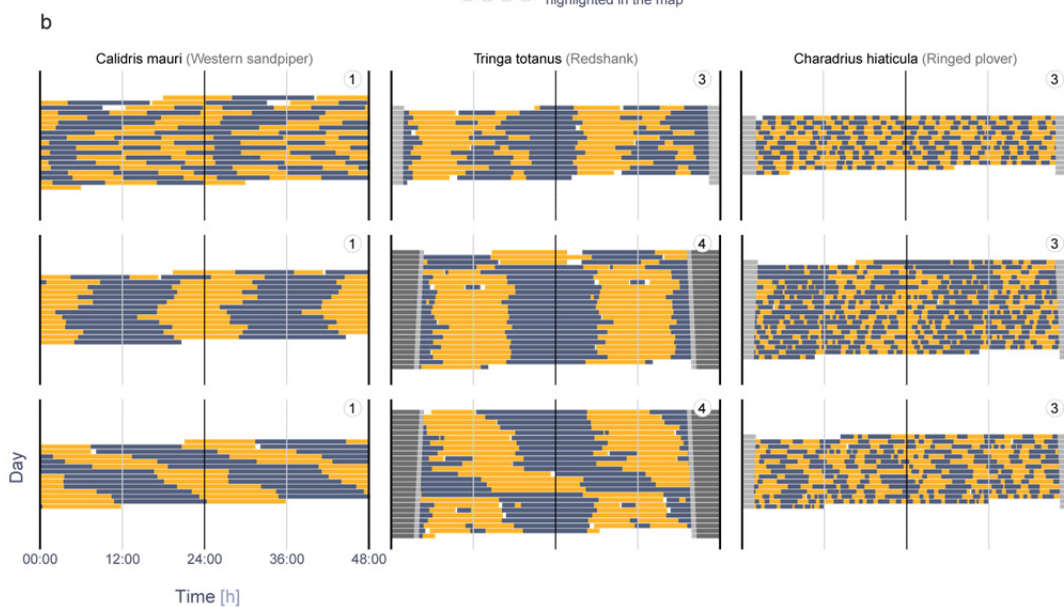
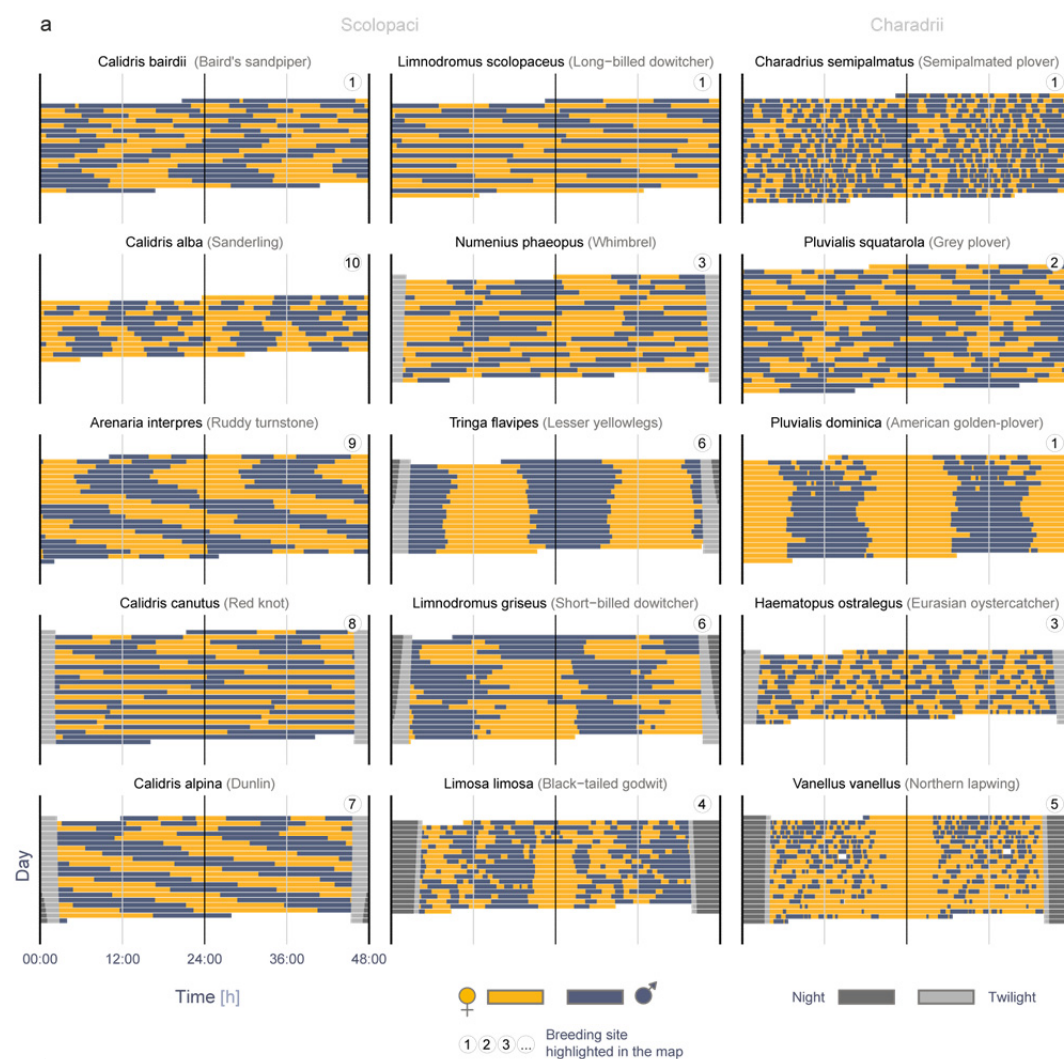
131 In conclusion, our results reveal that under natural conditions social synchronization can generate
132 much more diverse rhythms than expected from previous work, and that these rhythms often defy the
133 assumptions of entrainment to the 24-h day-night cycle. Not risk of starvation, but risk of predation seems to
134 play a key role in determining some of the variation in incubation rhythms. We describe this diversity in the
135 context of biparental incubation, but diverse behavioural rhythms may also arise in many other social settings
136 (e.g. in the context of mating interactions²⁹, vigilance behaviour during group foraging). Essentially, the reported
137 diversity suggests that the expectation that individuals within a pair (or group) should optimize their
138 behavioural rhythms relative to the 24-h day may be too simplistic, opening up a wide field to study the
139 evolutionary ecology of plasticity in circadian clocks.

140 **Online Content** Methods, along with any additional Extended Data display items and Source Data, are available
141 in the online version of the paper; references unique to these sections appear only in the online paper.



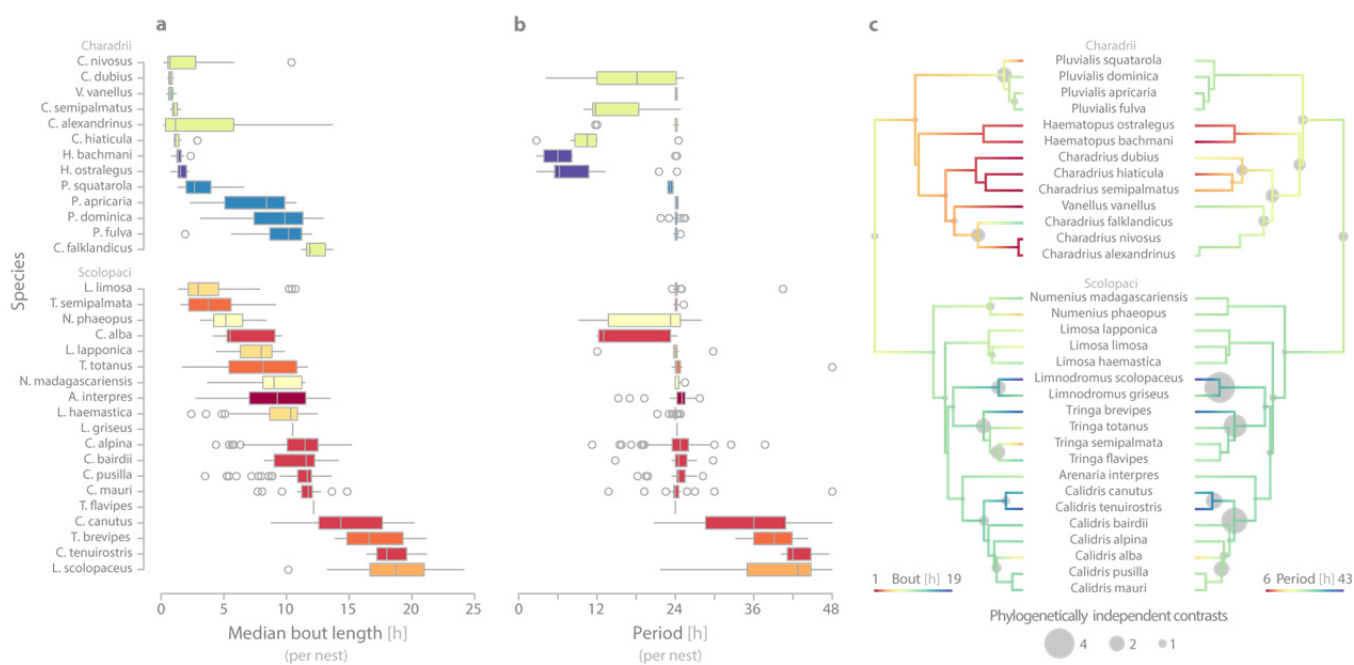
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143 **Figure 1 | Map of shorebird breeding sites with data on incubation rhythms.** The colour of the dots indicates the genus
144 (data from multiple species per genus may be available), the size of the dots refers to data quality (large: exact breeding

145 site known, small: breeding site estimated, see Methods). For nearby or overlapping locations, the dots are scattered to
 146 increase visibility. Numbers refer to those breeding sites for which incubation rhythms (of one or more species) are
 147 depicted in Fig. 2.

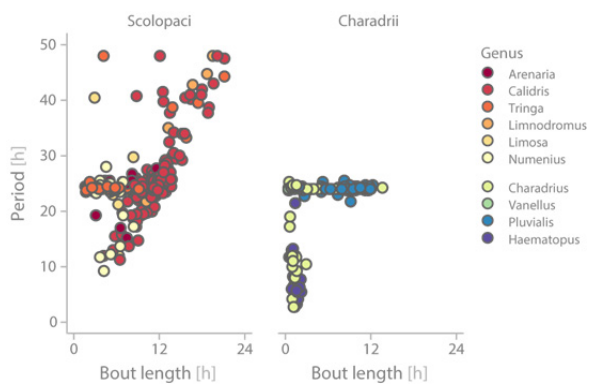


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149 **Figure 2 | Actograms illustrating the diversity of shorebird incubation rhythms. a-b**, Each actogram depicts the bouts of
 150 female (yellow; ♀) and male (blue-grey; ♂) incubation at a single nest over a 24-h period, plotted twice, such that each row
 151 represents two consecutive days. If present, twilight is indicated by light grey bars (▨) and corresponds to the time when
 152 the sun is between 6° and 0° below the horizon, night is indicated by dark grey bars (■) and corresponds to the time
 153 when the sun is < 6° below the horizon. Twilight and night are omitted in the centre of the actogram (24:00) to make the
 154 incubation rhythm visible. The circled numbers (1 2 3 ...) indicate the breeding site of each pair and correspond to the
 155 numbers on the map (Fig. 1). **a**, Between-species diversity. **b**, Within-species diversity. Note that the three rhythms for
 156 Western sandpiper and Ringed plover come from the same breeding location.



157 **Figure 3 | Between- and within-species variation in incubation rhythms and their estimated evolution. a-b**, Box plots are
 158 ordered by species (within suborder) from the shortest to the longest median bout length, and depict the genus (colour as
 159 in Fig. 1), median (vertical line inside the box), the 25th and 75th percentiles (box), 25th percentiles minus 1.5 times
 160 interquartile range and 75th percentile plus 1.5 times interquartile range or minimum/maximum value, whichever is smaller
 161 (bars), and the outliers (circles). $N_{\text{median bout length}} = 729$ and $N_{\text{period}} = 584$ nests. **b**, The red vertical line indicates a 24-h period.
 162 **c**, Observed and reconstructed incubation bout and period length visualised (by colour) on the phylogenetic tree³⁰ using
 163 species' medians (based on population medians) and one of 100 sampled trees (see Methods). The grey circles represent
 164 phylogenetically independent contrasts³¹ and hence emphasize the differences at each tree node.



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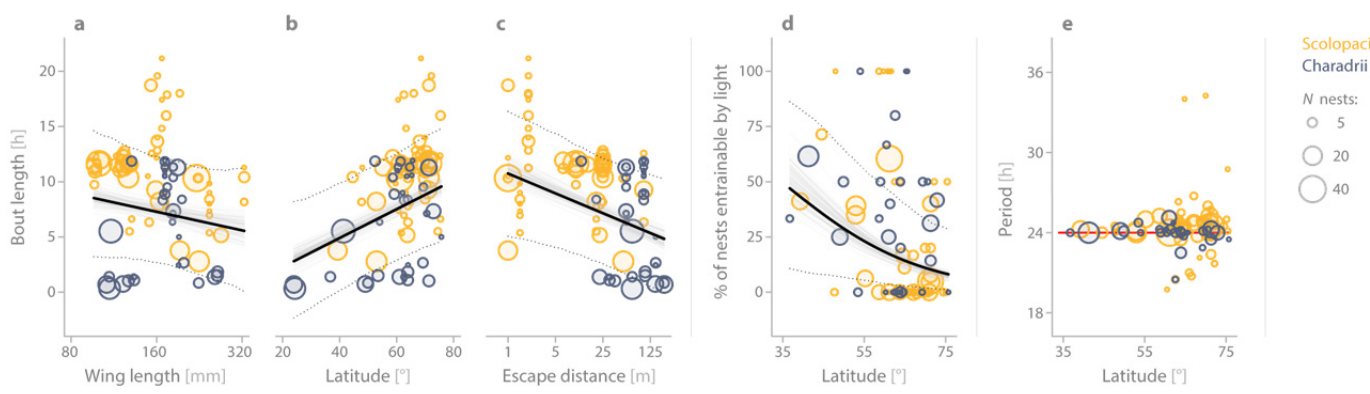
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Figure 4 | Relationship between bout and period length. Each dot represents a single nest ($N = 584$ nests), colours depict the genus. In the suborder Scolopaci the median bout length and period length correlate positively ($r_{Spearman} = 0.56$, $N = 424$ nests); in the suborder Charadrii periods longer than ~ 24 h are absent, and there is no simple relationship between bout and period length ($N = 160$ nests). For species-specific relationships see Extended Data Fig. 2.



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Figure 5 | Predictors of variation in incubation rhythms. **a-c**, Relationships between bout length and body size, measured as female wing length (**a**), breeding latitude (**b**) and anti-predation strategy, quantified as escape distance (**c**) for $N = 729$ nest from 91 populations belonging to 32 species. **d**, The relationship between the proportion of nests with a period length that is entrainable by the 24-h light-dark cycle (i.e. period lengths: 3, 6, 12, 24, or 48h) and breeding latitude ($N = 584$ nests from 88 populations belonging to 30 species). **e**, The distribution of period length over latitude. The period was standardized to 24h so that all 24-h harmonics are depicted as 24h (red line) and respective deviations from each harmonic as deviations from 24h (e.g. period of 12.5h is depicted as 25h). **a-e**, Each circle represents the population median; circle size indicates the number of nests. **a-c**, The solid lines depict the model-predicted relationships, the dotted lines the 95% credible intervals based on the joint posterior distribution of 100 separate MCMCglmm runs each with one of the 100 phylogenetic trees with $\sim 1,100$ independent samples per tree. The grey areas depict the predicted relationships for each of 100 runs (i.e. the full range of regression line estimates across 100 models) and illustrate the uncertainty due to the

184 phylogenetic tree. The predicted relationships stem from a Gaussian phylogenetic mixed-effect models, where the effects
185 of other predictors were kept constant (**a-c**, Extended Data Table 3), or from a binomial phylogenetic mixed-effect model
186 (**d**, Extended Data Table 3).

187 METHODS

188 **Recording incubation.** Incubation data were obtained between 1994 and 2015, for as many shorebird species
189 ($N = 32$) and populations ($N = 91$) as possible, using six methods (for specifications of the equipment see
190 Extended Data Table 4). (1) In 261 nests, a radio frequency identification reader ('RFID') registered presence of
191 tagged parents at the nest. The passive-integrated tag was either embedded in a plastic flag^{32,33}, with which the
192 parents were banded, or glued to the tail feathers³⁴. In 200 nests the RFID was combined with a temperature
193 probe placed between the eggs. The temperature recordings allowed us to identify whether a bird was
194 incubating even in the absence of RFID readings; an abrupt change in temperature demarcated the start or end
195 of incubation³². (2) For 396 nests, light loggers were mounted to the plastic flag or a band that was attached to
196 the bird's leg^{35,36}. The logger recorded maximum light intensity (absolute or relative) for a fixed sampling
197 interval (2-10 min). An abrupt change in light intensity (as opposed to a gradual change caused, e.g. by civil
198 twilight) followed by a period of low or high light intensity demarcated the start or end of the incubation period
199 (Extended Data Fig. 6). (3) For nine nests a GPS tag, mounted on the back of the bird, recorded the position of
200 the bird³⁷. The precision of the position depends on cloud cover and sampling interval³⁷. Hence, to account for
201 the imprecision in GPS positions, we assumed incubation whenever the bird was within 25 m of the nest
202 (Extended Data Fig. 6b). (4) At three nests automated receivers recorded signal strength of a radio-tag attached
203 to the rump of a bird; whenever a bird incubated, the strength of the signal remained constant²⁶
204 (Supplementary Actograms p. 257-9¹⁸). (5) At 53 nests video cameras and (6) for 8 nests continuous
205 observations were used to identify the incubating parents; parent identification was based on plumage, colour
206 rings or radio-tag. In one of the populations, three different methods were used, in seven populations
207 representing seven species two methods. In one nest, two methods were used simultaneously (Extended Data
208 Fig. 6b).

209 **Extraction of incubation bouts.** An incubation bout was defined as the total time allocated to a single parent
210 (i.e. the time between the arrival of a parent at and its departure from the nest followed by incubation of its
211 partner). Bout lengths were only extracted if at least 24h of continuous recording was available for a nest; in
212 such cases, all bout lengths were extracted. For each nest, we transformed the incubation records to local time
213 as (UTC time + $\frac{\text{nest's longitude}}{15}$). Incubation bouts from RFIDs, videos and continuous observations were mostly
214 extracted by an R-script and the results verified by visualizing the extracted and the raw data^{18,32,38,39}; otherwise,
215 MB extracted the bouts manually from plots of raw data^{40,41} (plots of raw data and extracted bouts for all nests
216 are in the Supplementary Actograms¹⁸; the actograms were generated by 'ggplot' and 'xyplot' functions from
217 the 'ggplot2' and 'lattice' R package⁴²⁻⁴⁴). Whenever the start or end of a bout was unclear, we classified these
218 bouts as uncertain (see next paragraph for treatment of unsure bouts). In case of light logger data, the light
219 recordings before and after the breeding period, when the birds were definitely not incubating, helped to
220 distinguish incubation from non-incubation. Whenever an individual tagged with a light logger nested in an
221 environment where the sun was more than 6° below the horizon for part of a day (i.e. night), we assumed an
222 incubation bout when the individual started incubating before the night started and ended incubating after the
223 night ended. When different individuals incubated at the beginning vs. at the end of the night, we either did not
224 quantify these bouts or we indicated the possible time of exchange (based on trend in previous exchanges), but
225 classified these bouts as uncertain (see Supplementary Actograms¹⁸). In total, we extracted 34,225 incubation
226 bouts.

227 The proportion of uncertain bouts within nests had a distribution skewed towards zero (median = 0%,
228 range: 0-100%, $N = 729$ nests), and so did the median proportion of uncertain bouts within populations (median
229 = 2%, range: 0-74%, $N = 91$ populations). Excluding the uncertain bouts did not change our estimates of median
230 bout length (Pearson's correlation coefficient for median bout length based on all bouts and without uncertain
231 bouts: $r = 0.96$, $N = 335$ nests with both certain and uncertain bouts). Hence, in further analyses all bouts were
232 used to estimate median bout length.

233 Note that in some species sexes consistently differ in bout length (Figure 2a, e.g. Northern lapwing). As
234 these differences are small compared to the between-species differences and because in 27 nests (of 8 species)
235 the sex of the parents was unknown, we here use median bout length independent of sex.

236 **Extraction of period length.** The method used for extracting the period length of incubation rhythm for each
237 nest is described in the Extended Data Fig. 1.

238 **Extraction of entrainable periods.** We classified 24-h periods and periods with 24-h harmonics (i.e. 3, 6, 12,
239 48h) as strictly entrainable by 24-h light fluctuations ($N = 142$ nests out of 584). Including also nearest adjacent
240 periods (± 0.25 h) increased the number of nests with entrainable periods ($N = 277$), but results of statistical
241 analyses remained quantitatively similar. We consider periods and harmonics of 12.42h (i.e. 3.1, 6.21, 12.42,
242 24.84h) as strictly entrainable by tide. However, because the periods in our data were extracted in 0.25-h
243 intervals (Extended Data Fig. 1), we classified periods of 3, 6.25, 12.5, 24.75h (i.e. those closest to the strict tide
244 harmonics) as entrainable by tide ($N = 32$ nests out of 584). Including also the second nearest periods (i.e. 3.25,
245 6, 12.25, 25) increased the number of nests entrainable by tide to $N = 55$.

246 **Population or species life-history traits.** For 643 nests, the exact breeding location was known (nests or
247 individuals were monitored at the breeding ground). For the remaining 86 nests (from 27 populations
248 representing 8 species, where individuals were tagged with light loggers on the wintering ground), the breeding
249 location was roughly estimated from the recorded 24-h variation in daylight, estimated migration tracks, and
250 the species' known breeding range⁴⁵⁻⁵². One exact breeding location was in the Southern Hemisphere, so we
251 used absolute latitude in analyses. Analyses without populations with estimated breeding-location or without
252 the Southern Hemisphere population generated quantitatively similar estimates as the analyses on full data.

253 For each population, body size was defined as mean female wing length⁵³, either for individuals
254 measured at the breeding area or at the wintering area. In case no individuals were measured, we used the
255 mean value from the literature (see open access data for specific values and references⁵⁴).

256 Anti-predation strategy was assessed by estimating escape distance of the incubating bird when a
257 human approached the nest, because species that are cryptic typically stay on the nest much longer than non-
258 cryptic species, sometimes until nearly stepped upon^{49,55}. Escape distance was obtained for all species. Forty-

259 four authors of this paper estimated the distance (in m) for one or more species based on their own data or
260 experience. For 10 species, we also obtained estimates from the literature⁴⁹. We then used the median
261 ‘estimated escape distance’ for each species. In addition, for 13 species we obtained ‘true escape distance’.
262 Here, the researcher approached a nest (of known position) and either estimated his distance to the nest or
263 marked his position with GPS when the incubating individual left the nest. For each GPS position, we calculated
264 the Euclidian distance from the nest. In this way we obtained multiple observations per nest and species, and
265 we used the median value per species (weighted by the number of estimates per nest) as the ‘true escape
266 distance’. The species’ median ‘estimated escape distance’ was a good predictor of the ‘true escape distance’
267 (Pearson’s correlation coefficient: $r = 0.89$, $N = 13$ species). For analysis, we defined the escape distance of a
268 species as the median of all available estimates.

269 For each species, we determined whether it predominantly uses a tidal environment outside its
270 breeding ground, i.e. has tidal vs. non-tidal life history (based on^{49,51,52}). For each population with exact breeding
271 location, we scored whether tidal foraging habitats were used by breeding birds for foraging (for three
272 populations this information was unknown)⁵⁴. For all populations with estimated breeding location we assumed,
273 based on the estimated location and known behaviour at the breeding grounds, no use of tidal habitat.

274 **Statistical analyses.** Unless specified otherwise, all analyses were performed on the nest level using median
275 bout length and extracted period length.

276 We used phylogenetically informed comparative analyses to assess how evolutionary history constrains
277 the incubation rhythms (estimated by Pagel’s λ coefficient of phylogenetic signal^{56,57}) and to control for
278 potential non-independence among species due to common ancestry. This method explicitly models how the
279 covariance between species declines as they become more distantly related^{56,58,59}. We used the Hackett⁶⁰
280 backbone phylogenetic trees available at <http://birdtree.org>⁶¹, which included all but one species (*Charadrius*
281 *nivosus*) from our dataset. Following a subsequent taxonomic split⁶², we added *Charadrius nivosus* to these
282 trees as a sister taxon of *Charadrius alexandrinus*. Phylogenetic uncertainty was accounted for by fitting each
283 model with 100 phylogenetic trees randomly sampled from 10,000 phylogenies at <http://birdtree.org>⁶¹.

284 The analyses were performed with Bayesian phylogenetic mixed-effect models (Fig. 5 and Extended
285 Data Table 1 and 3) and the models were run with the 'MCMCglmm' function from the R package
286 'MCMCglmm'⁶³. In all models, we also accounted for multiple sampling within species and breeding site
287 (included as random effects). In models with a Gaussian response variable, an inverse-gamma prior with shape
288 and scale equal to 0.001 was used for the residual variance (i.e. variance set to one and the degree of belief
289 parameter to 0.002). In models with binary response variables, the residual variance was fixed to one. For all
290 other variance components the parameter-expanded priors were used to give scaled F-distributions with
291 numerator and denominator degrees of freedom set to one and a scale parameter of 1,000. Model outcomes
292 were insensitive to prior parameterization. The MCMC chains ran for 2,753,000 iterations with a burn-in of
293 3,000 and a thinning interval of 2,500. Each model generated ~1,100 independent samples of model parameters
294 (Extended Data Table 1 and 3). Independence of samples in the Markov chain was assessed by tests for
295 autocorrelation between samples and by using graphic diagnostics.

296 First, we used MCMCglmm to estimate Pagel's λ (phylogenetic signal) for bout and period length
297 (Gaussian), and to show that our estimates of these two incubation variables were independent of how often
298 the incubation behaviour was sampled ('sampling' in min, ln-transformed; Extended Data Table 1). Hence, in
299 subsequent models, sampling was not included.

300 Then, we used MCMCglmm to model variation in bout length and period length. Bout length was
301 modelled as a continuous response variable and latitude ($^{\circ}$, absolute), female wing length (mm, ln-transformed)
302 and approach distance (m, ln-transformed) as continuous predictors. Predictors had low collinearity (at nest,
303 population and species level; all Pearson or Spearman correlation coefficients $|r| < 0.28$). To test for potential
304 entrainment to 24-h, period length was modelled as a binary response variable (1 = rhythms with period of 3, 6,
305 12, 24, or 48 h; 0 = rhythms with other periods) and latitude as a continuous predictor. To test how circadian
306 period varies with latitude or life history, period was transformed to deviations from 24-h and 24-h harmonics
307 and scaled by the time span between the closest harmonic and the closest midpoint between two harmonics.
308 For example, a 42h period deviates by -6h from 48h (the closest 24-h harmonic) and hence -6h was divided by
309 12h (the time between 36h – the midpoint of two harmonics - and 48h -the closest harmonic). This way the

310 deviations spanned from -1 to 1 with 0 representing 24-h and its harmonics. The absolute deviations were then
311 modelled as a continuous response variable and latitude as continuous predictor. The deviations were also
312 modelled as a continuous response and species life history (tidal or not) as categorical predictor.

313 In all models the continuous predictor variables were centred and standardized to a mean of zero and a
314 standard deviation of one.

315 We report model estimates for fixed and random effects, as well as for Pagel's λ , by the modes and the
316 uncertainty of the estimates by the highest posterior density intervals (referred to as 95% CI) from the joint
317 posterior distributions of all samples from the 100 separate runs, each with one of the 100 separate
318 phylogenetic trees from <http://birdtree.org>.

319 To help interpret the investigated relationships we assessed whether incubation rhythms evolved
320 within diverged groups of species by plotting the evolutionary tree of the incubation rhythm variables (Fig. 3b),
321 as well as of the predictors (Extended Data Fig. 5).

322 The source of phylogenetic constraint in bout and period length was investigated by estimating the
323 proportion of phenotypic variance explained by suborder, genus and species (Extended Data Table 2). The
324 respective mixed models were also specified with 'MCMCglmm'⁶³ using the same specifications as in the
325 phylogenetic models. Because suborder contained only two levels, we first fitted an intercept mixed model with
326 genus, species, and breeding site as random factors, and used it to estimate the overall phenotypic variance.
327 We then entered suborder as a fixed factor and estimated the variance explained by suborder as the difference
328 between the total variance from the first and the second model. To evaluate the proportion of the variance
329 explained by species, genus and breeding site, we used the estimates from the model that included suborder.

330 R version 3.1.1⁶⁴ was used for all statistical analyses.

331 **Code availability.** All statistical analyses are replicable with the open access data and r-code available from

332 <https://osf.io/wxufm/> (not public yet, but available for peer review:

333 https://osf.io/wxufm/?view_only=d0172c5f0935462f9471d6fdc4e40d86).

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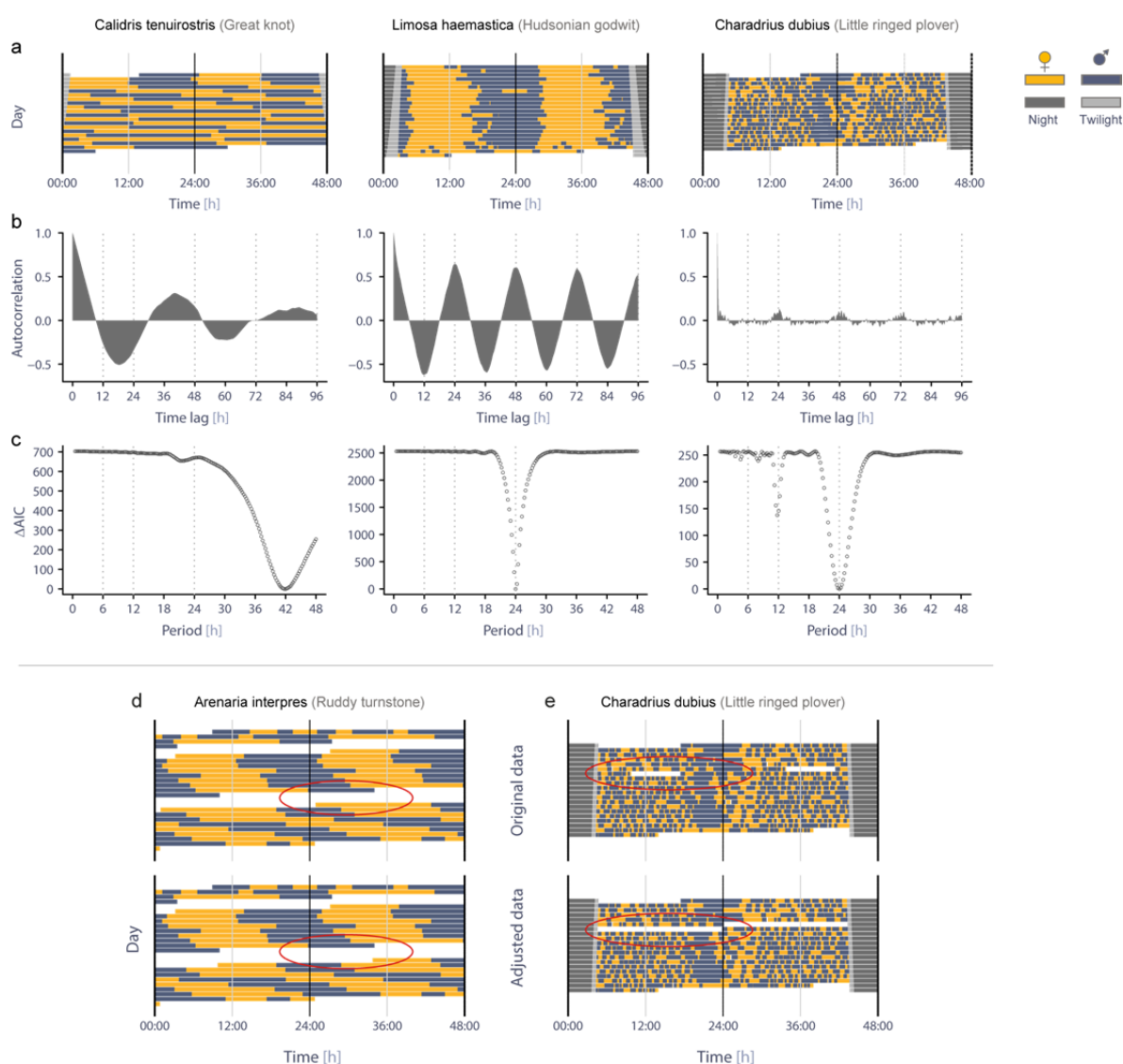
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478 **Author Contributions** M.B. and B.K. conceived the study. All authors except for B.H. collected the primary data. MB
479 managed the data. MB and M.V. developed the methods to extract incubation. M.B. extracted bout lengths and with help
480 from A.R. and M.V. created actograms. M.B. with help from M.V. analysed the data. M.B. and B.K. wrote the paper with
481 input from the other authors. Except for the first, second and last author, the authors are listed alphabetically by their first
482 name.

483 **Author Information** All information, primary and extracted data, computer code and software necessary to replicate our
484 results, as well as the Supplementary Actograms are open access and archived at Open Science Framework
485 <https://osf.io/wxufm/> (not public yet, but available for peer review: https://osf.io/wxufm/?view_only=d0172c5f0935462f9471d6fdc4e40d86). Reprints and permissions information is available at www.nature.com/reprints.
486
487 The authors declare no competing financial interests. Readers are welcome to comment on the online version of the paper.
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Extended Data Figure 1 | Illustration of the method used for extracting period length of incubation rhythm. a-c, Each column represents an example for a specific nest with long, intermediate and short incubation bouts. **a,** From the extracted bout lengths we created a time series that indicated for each nest, and every 10 min, whether a specific parent (female, if sex was known) incubated or not. Exchange gaps (no parent on the nest) had to be < 6 h to be included (for treatment of exchange gaps > 6 h see **d, e**). **b,** We then estimated the autocorrelation for each 10 min time-lag up to 4 days (R ‘acf’ function⁶⁴). Positive values indicate a high probability that the female was incubating, negative values indicate that it was more likely that the male was incubating. We used only nests that had enough data to estimate the autocorrelation pattern ($N = 584$ nests from 88 populations of 30 species). The visualized autocorrelation time series never resembled white or random noise indicative of an arrhythmic incubation pattern. To determine the period (i.e. cycle of high and low probability for a parent to incubate) that dominated the incubation rhythm, we fitted to the autocorrelation estimates a series of periodic logistic regressions. In each regression, the time lag (in hours) transformed to radians was represented by a sine and cosine function $f(t) = a_0 + b \cdot \left(\cos \frac{2 \cdot \pi \cdot t}{T}\right) + c \cdot \left(\sin \frac{2 \cdot \pi \cdot t}{T}\right) + e$, where $f(t)$ is the autocorrelation at time-lag t , a_0 is the intercept, b is the slope for sine and c the slope for cosine, T represents the length of the fitted period (in hours), and e is an error term. We allowed the period length to vary from 0.5 h to 48 h (in 15 min intervals, giving 191 regressions). **c,** By comparing the Akaike’s Information Criterion⁶⁵ (AIC) of all regressions, we estimated, for each nest, the length of the dominant period in the actual incubation data (best fit). Regressions with ΔAIC ($AIC_{\text{model}} - AIC_{\text{min}}$) close to 0 are considered as having strong empirical support, while models with ΔAIC values ranging from 4-7 have less

507 support⁶⁵. In 73% of all nests, we determined a single best model with $\Delta AIC \leq 3$ (**c**, middle ΔAIC graph), in 20% of nests two best models
508 emerged and in 6% of nests 3 or 4 models had $\Delta AIC \leq 3$ (**c**, left and right ΔAIC graphs). However, in all but three nests, the models with
509 the second, third, etc. best ΔAIC were those with period lengths closest to the period length of the best model (**c**, left and right ΔAIC
510 graphs). This suggests that multiple periodicities are uncommon. **d-e**, The extraction of the period length (described in **a-c**) requires
511 continuous datasets, but some nests had long (>6 h) gaps between two consecutive incubation bouts, for example because of equipment
512 failure or because of unusual parental behaviour. In such cases, we excluded the data from the end of the last bout until the same time
513 the following day, if data were then available again (**d**), or we excluded the entire day (**e**).



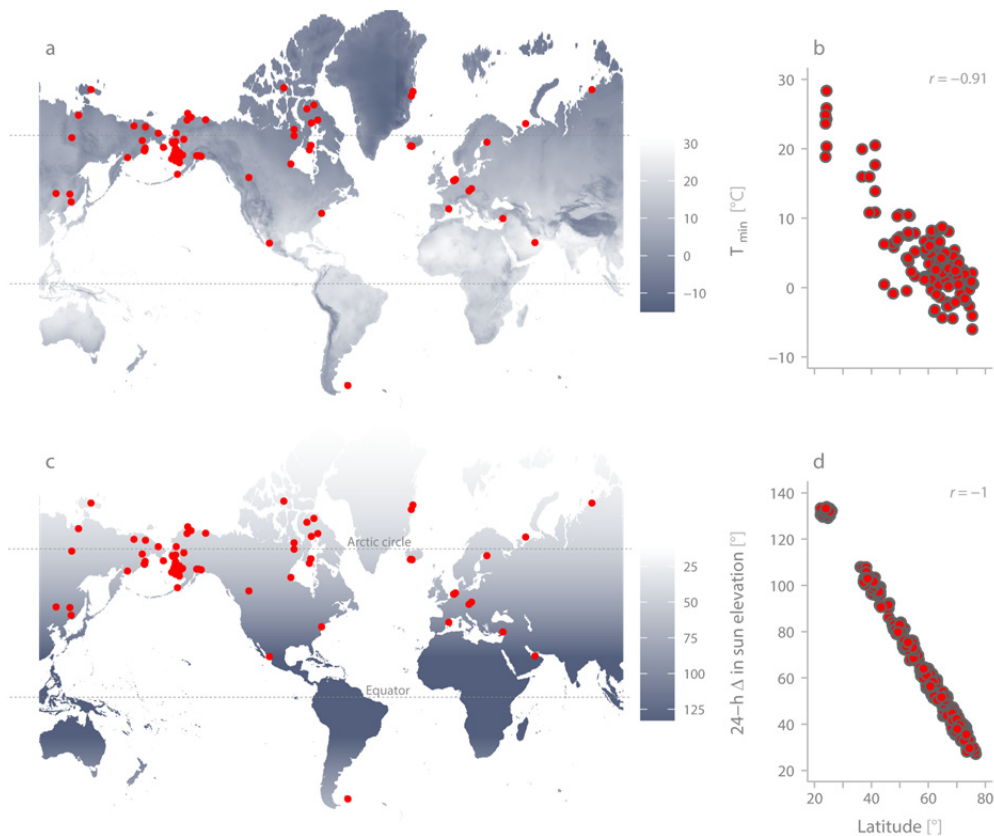
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Extended Data Figure 2 | Relationship between bout and period length for 30 shorebird species. Each dot represents one nest ($N = 584$

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nest), colours indicate the genus.



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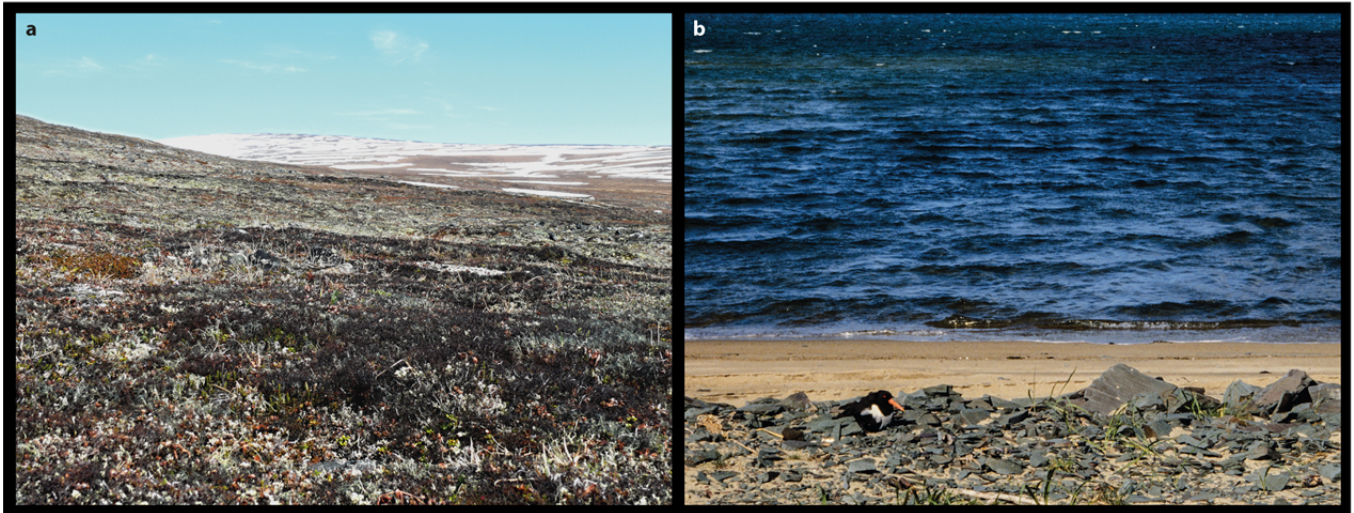
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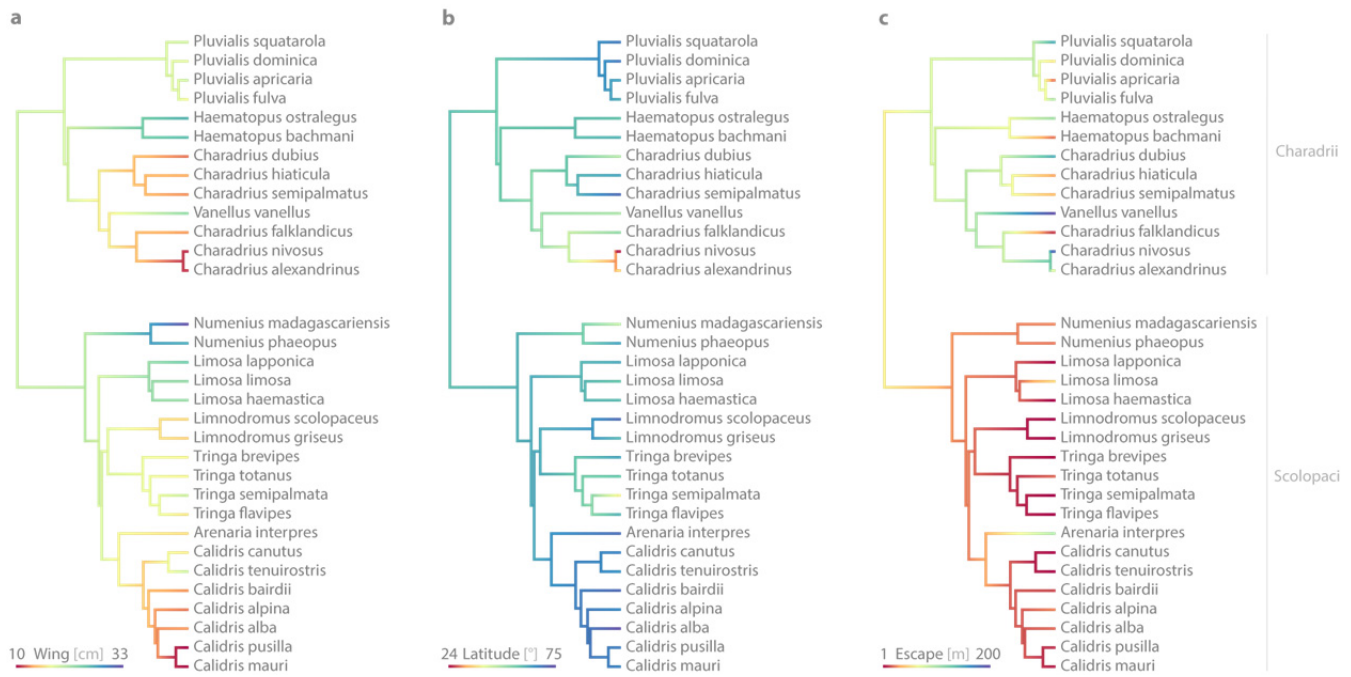
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Extended Data Figure 3 | Ecological correlates of latitude. **a**, Variation in minimum temperature across the globe represented by mean minimum June temperature for the Northern Hemisphere and mean minimum December temperature for the Southern Hemisphere. **b**, Correlation between absolute latitude and the mean minimum temperature based on the month represented by mid-day of incubation data for each nest ($N = 729$). For maximum temperature the correlation was similar ($r = -0.91$, $N = 729$ nests). **c**, Daily variation in sun elevation (i.e. in light conditions) represented as the difference between the noon and midnight sun-elevation for the summer solstice in the Northern Hemisphere and the winter solstice in the Southern Hemisphere. **d**, Correlation between absolute latitude and daily variation in sun elevation for mid-day of incubation data for each nest ($N = 729$ nests). The points are jittered, as else they form a straight line. **a, c**, Red points indicate the breeding sites ($N = 91$). **a-b**, The minimum and maximum monthly temperature data were obtained from www.worldclim.org using the 'raster' R-package⁶⁶. **c-d**, Sun-elevation was obtained by the 'solarpos' function from the 'mapproj' R-package⁶⁷.



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Extended Data Figure 4 | Between-species variation in parental crypsis during incubation. a-b, Shorebirds vary in how visible they are on the nest while incubating. The nearly invisible Great knot (*Calidris tenuirostris*; **a**; central and facing right) sits tight on the nest when approached by a human until nearly stepped upon. In contrast, the conspicuous Eurasian oystercatcher (*Haematopus ostralegus*; **b**) is visible on the nest from afar and when approached by a human leaves the nest about 100 m in advance (Credits: **a**, M. Šálek; **b**, Jan van de Kam).



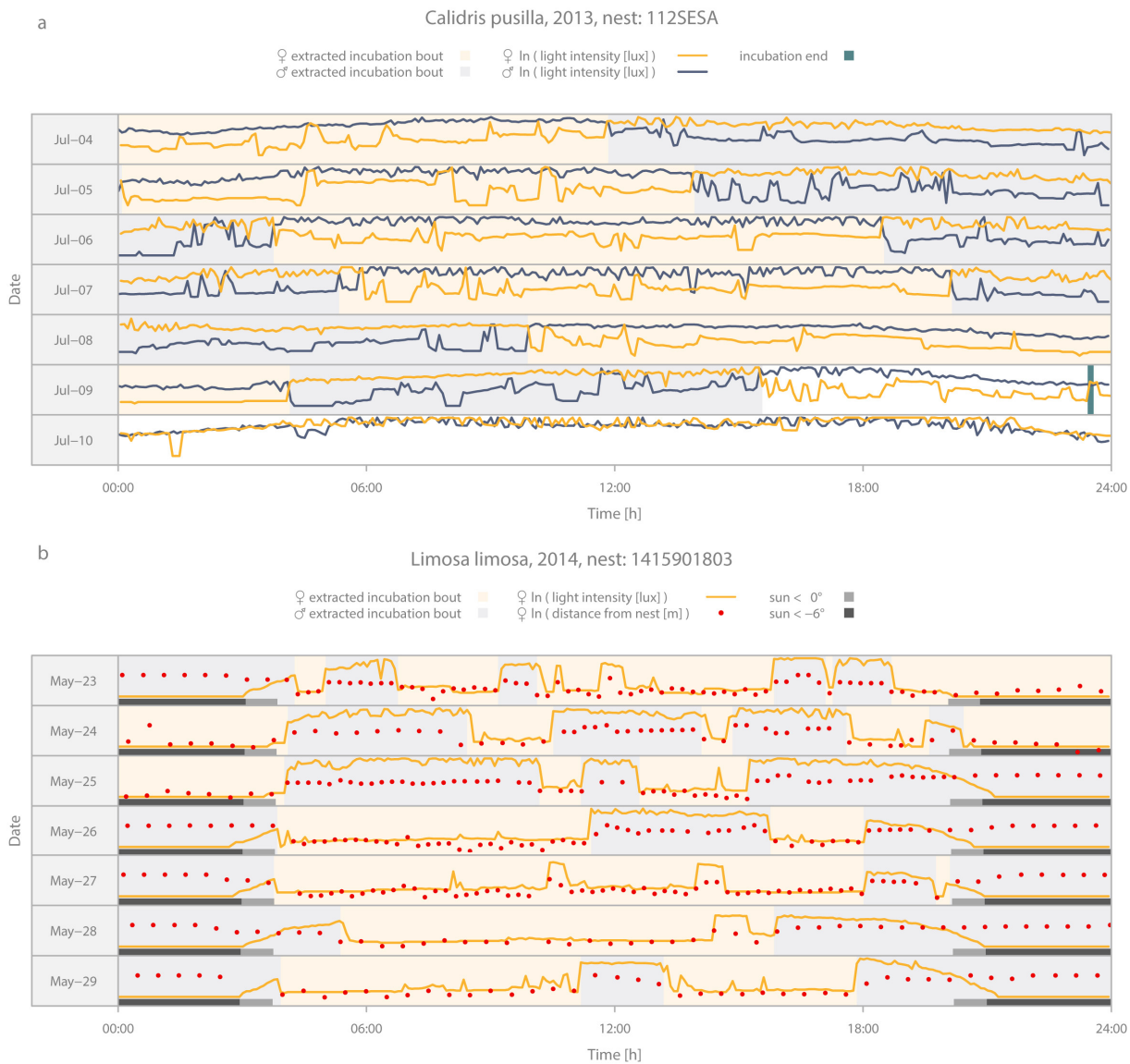
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Extended Data Figure 5 | Phylogenetic relationships for predictors. a, Body size, estimated as female wing length. **b**, Latitude (absolute), **c**, Escape distance. **a-c**, We visualised the evolution of these traits^{30,68} using species' medians (**a-b**; based on population medians), species' estimates of escape distance (**c**) and one of the 100 sampled trees (see Methods).



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Extended Data Figure 6 | Illustration and verification of the method used to extract incubation bouts from light logger data. a, An example of a nest with a light intensity signal from both parents (yellow line — and blue-grey line —). The incubation bouts for a given parent reflect periods dominated by lower light values compared to those of the partner. Note the sharp drop in the light levels at the beginning of each incubation bout and the sharp increase in the light levels at the end. Change-overs between partners occur when the light signal lines cross. Such pronounced changes in light intensity detected by the logger were used to assign incubation even when only a single parent was tagged. Note that after the chicks hatch and leave the nest (July 9, vertical bar), the light intensity signal from both parents remains similar. **b,** An example of a nest where one incubating parent was simultaneously equipped with a light logger and with a GPS tag. The yellow line (—) indicates light levels, red dots (•) indicate the distance of the bird to the nest. As expected, low light levels co-occur with close proximity to the nest, and hence reflect periods of incubation. Although light levels decrease during twilight (light grey horizontal bar; —), the recordings were still sensitive enough to reflect periods of incubation, i.e. the light signal matches the distance (e.g. May-25: female incubated during dawn, but was off the nest during dusk). **a-b,** Rectangles in the background indicate extracted female (light yellow polygon, ■) and male (light blue-grey polygon, ■) incubation bouts.

Extended Data Table 1 | Effects of phylogeny and sampling on bout length and period length.

Response	Effect type	Effect	Posterior mode	95% CI		N (range)
				Lower	Upper	
Median bout [h]	Fixed	Intercept	7.2	1.04	12	1100 (924-2079)
		Sampling	0.16	-0.2	0.61	1100 (809-1644)
	Random (variance)	Phylogeny	25.33	4.6	59.6	1100 (753-1383)
		Species	0.01	0	12.1	1100 (779-1636)
		Breeding site	2.13	0.96	4.28	1100 (808-2242)
		Residual	5.04	4.51	5.61	1100 (838-1444)
	Pagel's λ		1	0.5	1	1100 (814-1316)
Period [h]	Fixed	Intercept	21.94	12.8	30.67	1100 (765-1392)
		Sampling	0.13	-0.41	0.65	1100 (741-1468)
	Random (variance)	Phylogeny	66.22	14.3	153	1100 (729-1638)
		Species	0.06	0	29.36	1100 (729-1435)
		Breeding site	0.01	0	0.88	1100 (814-1378)
		Residual	14.87	13.3	16.84	1100 (884-1460)
	Pagel's λ		1	0.54	1	1100 (740-1523)

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The posterior estimates (modes) of the effect sizes with the highest posterior density intervals (95% CI) and the median and range of the effective sample sizes (N (range)) come from the joint posterior distribution of 100 separate runs each with one of 100 separate phylogenetic trees from <http://birdtree.org>. $N_{\text{bout}} = 729$ nests from 91 populations belonging to 32 species. $N_{\text{period}} = 584$ nests from 88 populations belonging to 30 species.

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Sampling (how often the incubation behaviour was sampled) was ln-transformed and then mean-centred and scaled (divided by SD). For procedures and specifications related to phylogenetic Bayesian mixed models see Methods. Estimating Pagel's λ on the species level ($N_{\text{bout}} = 32$ species, $N_{\text{period}} = 30$ species) with phylogenetic generalized least-squares using the function 'pgls' from the R-package 'caper'⁶⁹ gave similar results (median (range) $\lambda_{\text{bout}} = 0.73$ (0.63-1) and $\lambda_{\text{period}} = 0.95$ (0.64-1), based on 100 estimates each for one of the 100 trees).

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Extended Data Table 2 | Source of phylogenetic signal

Suborder included	Response	Effect type	Fixed effects	Posterior mode	95% CI		N
					Lower	Upper	
No	Bout [h]	Fixed	Intercept	7.69	4.11	10.76	1100
			Random (variance)	Genus	9.54	0.02	46.6
		Species	11	4.84	25.77	1100	
		Breeding site	2.25	1.06	4.5	1100	
		Residual	5.03	4.55	5.61	1100	
Yes	Bout [h]	Fixed	Intercept (Charadrii)	4.48	1.05	7.66	1100
			Suborder (Scolopaci)	6.07	1.52	10.44	1100
		Random (variance)	Genus	0.09	0	21.35	1100
			Species	11.29	5.53	23.99	1275
			Breeding site	2.31	0.98	4.28	1100
			Residual	5.04	4.52	5.58	1100
No	Period [h]	Fixed	Intercept	23.46	18.22	27.39	1100
			Random (variance)	Genus	19.05	0	97.79
		Species	41.02	17.04	72.66	1100	
		Breeding site	0.01	0	0.9	1100	
		Residual	14.94	13.35	16.76	1100	
Yes	Period [h]	Fixed	Intercept (Charadrii)	18.61	13.12	23.98	940
			Suborder (Scolopaci)	8.92	0.6	15.4	990
		Random (variance)	Genus	0.33	0	59.09	769
			Species	34.41	16.91	68.14	1142
			Breeding site	0.01	0	0.84	1100
			Residual	15.01	13.34	16.86	891

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The posterior estimates (modes) of the effect sizes with the highest posterior density intervals (95% CI) and the effective sample sizes (N) come from a posterior distribution of 1,100 simulated values generated by 'MCMCglmm' in R⁶³. $N_{\text{bout}} = 729$ nests from 91 populations belonging to 32 species. $N_{\text{period}} = 584$ nest from 88 populations belonging to 30 species.

Extended Data Table 3 | Effect of latitude, body size, escape distance and life history on biparental incubation rhythms in shorebirds

Response	Effect type	Fixed effects	Posterior mode	95% CI		N (range)
				Lower	Upper	
Bout [h]	Fixed	Intercept	7.45	2.65	12	1100 (804-1496)
		Wing length	-0.78	-2.5	1.05	1100 (839-1638)
		Latitude	1.72	0.63	2.65	1100 (850-1642)
		Escape distance	-1.68	-3.3	-0.25	1100 (634-2046)
	Random (variance)	Phylogeny	0.19	0	45	1100 (803-1875)
		Species	0.07	0	14.4	1100 (695-1580)
		Breeding site	1.4	0.59	3.02	1100 (833-1480)
	Residual	5.02	4.53	5.64	1100 (516-1916)	
Pagelet's λ	0.72	0.13	1	1100 (731-1407)		
Light entrainable rhythm [1,0] on binomial scale	Fixed	Intercept	-1.62	-3.19	-0.13	1100 (731-1633)
		Latitude	-0.56	-1.15	-0.07	1100 (765-1575)
	Random (variance)	Phylogeny	0.05	0	5.54	1100 (883-1371)
		Species	0.02	0	2.68	1100 (965-2246)
		Breeding site	0	0	0.63	1100 (605-1304)
	Pagelet's λ	0.74	0.02	1	1100 (932-1498)	
Absolute deviations from 24-h	Fixed	Intercept	0.17	-0	0.35	1100 (459-1501)
		Latitude	0.03	-0	0.07	1100 (777-1488)
	Random (variance)	Phylogeny	0	0	0.07	1100 (786-1393)
		Species	0	0	0.03	1100 (861-1412)
		Breeding site	0	0	0	1100 (826-1860)
	Residual	0.03	0.03	0.04	1100 (948-2039)	
Pagelet's λ	0.74	0.02	1	1100 (843-1471)		
Deviations from 24-h	Fixed	Intercept (non-tidal)	0.02	-0.04	0.09	1100 (851-1742)
		Life history (tidal)	-0.02	-0.1	0.04	1100 (702-2257)
	Random (variance)	Phylogeny	0	0	0.01	1100 (806-1692)
		Species	0	0	0	1100 (692-1601)
		Breeding site	0	0	0.01	1100 (656-1490)
	Residual	0.07	0.06	0.08	1100 (760-1563)	
Pagelet's λ	0.77	0.01	1	1100 (864-1451)		

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The posterior estimates (modes) of the effect sizes with the highest posterior density intervals (95% CI) and the median and range of the effective sample sizes (N (range)) come from the joint posterior distribution of 100 separate runs each with one of the 100 separate phylogenetic trees from <http://birdtree.org>. $N_{\text{bout}} = 729$ nests from 91 populations belonging to 32 species. For models on light entrainable rhythm, absolute deviations and deviations from 24-h: $N = 584$ nests from 88 populations belonging to 30 species. Latitude (in bout model: absolute value), wing length (ln-transformed), and escape distance (ln-transformed) were mean-centred and scaled (divided by SD). The estimates for the light-entrainable rhythm are on a binomial scale. For procedures and specifications related to phylogenetic Bayesian mixed models see Methods.

Extended Data Table 4 | Incubation monitoring methods and systems.

Method	Model	Company	www	Year	Sampling interval (min)	N populations	N nests	N both parents tagged
RFID + tag embedded in the flag ^{32,33}	tag: 9.0 × 2.1 mm, 0.087 g, 134.2 kHz	Biomark	http://www.biomark.com/	2011-2014	0.08	19	200	157
	RFID: custom made	Calima Engineering, & Max Planck Institute for Ornithology	http://www.calima.de http://www.orn.mpg.de/en					
	temperature probe: TinyTag Talk PB-5005-0M6 (ø 2.5 mm) with Talk 2 logger TK-4023 or	Gemini Data Loggers Ltd	www.tinytag.info				197	
	MSR® (12 × 6 mm) with MSR® 145 logger	MSR® Electronics GmbH	http://www.msr.ch/					
RFID + tag glued to the tail ³⁴	tag: 11.5 × 2.12 mm, 0.1 g, 125 kHz	Trovan®	http://www.trovan.com	2007	5	1	3	3
	RFID: custom made	EID Aalten BV	http://www.dorset.nu/en/					
	temperature probe: TinyTag Talk PB-5005-0M6 (ø 2.5 mm) with Talk 2 logger TK-4023	Gemini Data Loggers Ltd	www.tinytag.info					
	tag: 11.5 × 2.12 mm, 0.09g, 128 kHz	Trovan®	http://www.trovan.com	2005-2008	1.7-5.5	1	34	34
Light logger ^{35,36,70}	RFID: LID650 with extended memory, driver TM613, Antenna ANT614							
	tag: TIRIS 12 × 6 × 3 mm, 0.4 g, 134.2 kHz	Texas Instruments	http://www.ti.com	1997	1/3	1	6	6
	RFID: TIRIS Micro-reader							
	TIRIS tag: 12 × 6 × 3 mm, 0.4 g, 134.2 kHz	Texas Instruments	http://www.ti.com	2006	1/3	1	18	18
GPS-tracker ³⁷	RFID: custom made	Francis Scientific Instruments Limited	https://www.duedil.com/comp-any/01964877/francis-scientific-instruments-limited					
	MK10, 12, 14 18, 20: 0.7-1.4g, light scale 0-64 (64 ≥ 100 lux)	British Antarctic Survey, currently Biotrack	http://www.biomark.com/	2008-2013	2-10	48	261*	55
Radio-transmitter ²⁶	Intigeo W65A9RK: 0.7g, absolute light levels (lux)	Migrate Technology Ltd	http://www.migratetech.co.uk	2012-2014	5-10	23	136*	11
	UvA-BiTS 4C: 62 × 30 × 12, 14 g	University of Amsterdam	http://www.uva-bits.nl/	2010	10-30	1	8	8
Video ⁷¹⁻⁷⁴	UvA-BiTS 2CDse: 52 × 22 × 9, 7.5 g			2014	20	1	1**	0
	iTag: 26 × 15 × 9 mm, 4 g	e-obs GmbH	http://www.e-obs.de/	2009	0.07	2	3	3
Observations				2005-2006	1/3	1	28	-
				2005-2011	constant	3	25	-
Observations				1994	30	1	5	-
				2011	constant	1	3	-

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For details about methods used in each populations, see Supplementary Data⁵⁴.

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*At one nest a bird with MK logger was recaptured and the logger exchanged for Intigeo logger. This nest appears in N for both logger types.

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**Simultaneously equipped with light logger (Intigeo). This nest appears in N for both GPS-tracer and Intigeo.