

Citation for published version:

Frederiksen, K, Guy, RH & Petersson, K 2016, 'The potential of polymeric film-forming systems as sustained delivery platforms for topical drugs', *Expert Opinion on Drug Delivery*, vol. 13, no. 3, pp. 349-360. https://doi.org/10.1517/17425247.2016.1124412

DOI: 10.1517/17425247.2016.1124412

Publication date: 2016

Document Version Peer reviewed version

Link to publication

University of Bath

Alternative formats

If you require this document in an alternative format, please contact: openaccess@bath.ac.uk

General rights

Copyright and moral rights for the publications made accessible in the public portal are retained by the authors and/or other copyright owners and it is a condition of accessing publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy If you believe that this document breaches copyright please contact us providing details, and we will remove access to the work immediately and investigate your claim.

The potential of polymeric film-forming systems as sustained delivery platforms for topical drugs

Abstract

Introduction: Dosing regimens requiring multiple daily applications frequently result in poor patient compliance, especially in the treatment of chronic skin diseases. Consequently, development of sustained delivery systems for topical drugs permitting less frequent dosing is of continuing interest for dermatological therapy.

Areas covered: This potential of polymeric film-forming systems (FFS), created *in situ* on the skin, as sustained delivery platforms for topical drug delivery is reviewed. Key formulation parameters that determine delivery efficiency are considered focussing on those that permit a drug reservoir to be established in the upper layers of the skin and/or on the skin surface from which release can be sustained over a prolonged period. The advantageous and superior cosmetic attributes of FFS (compared to conventional semi-solid formulations) that offer significantly improved patient compliance are also addressed.

Expert opinion: The promise of polymeric FFS as convenient and aesthetic platforms for sustained topical drug delivery is clear. Manipulation of the formulation allows the delivery profile to be customised and optimised to take advantage of both a rapid, initial input of drug into the skin (likely due to a transient period of supersaturation) and a slower, controlled release over an extended time from the residual film created thereafter.

Keywords

Dermal drug delivery; polymeric film-forming systems; supersaturation; topical formulations; sustained delivery.

Article highlights

- Optimisation of the formulation of polymeric film-forming systems (FFS) permits sustained drug delivery to be achieved via the formation of a drug reservoir in or on the skin.
- The initial metamorphosis of the formulation, and consequent increase in the degree of drug saturation, enables the establishment of a drug reservoir in the upper skin layers.
- FFS prepared with hydrophobic polymers have greater skin substantivity, facilitating formation of an external drug reservoir from which sustained delivery may be achieved.
- Realisation of the long-term potential of FFS as sustained, topical drug delivery systems requires proof-of-principle to be demonstrated conclusively *in vivo*.
- A key issue is to accomplish the desired therapeutic effect with a FFS formulation that creates an aesthetically acceptable residual polymeric film on the skin.

1 Introduction

The topical treatment of skin diseases is desirable for obvious reasons: the drug product is applied directly at the affected site, achieving levels at the local target that are at least as high as those possible by (e.g.) oral administration, but with very limited systemic exposure and associated side-effects. However, topical delivery can be challenging due to the efficient barrier properties of the stratum corneum (SC). Typically, only a small percentage of the applied drug reaches the target site, the remainder being left in a non-diffusible (i.e., solid) form in the residual film post-application and unavailable for delivery (1, 2).

Patient compliance to the repetitive daily application of conventional, cosmetically suboptimal topical dosage forms (ointments, etc.) is often poor (3-5), compromising the efficacy of the treatment of chronic skin diseases in particular (3). The development of topical, sustained delivery formulations permitting prolonged therapeutic effect and less frequent dosing would offer great benefit, therefore, in dermatological therapy. How can this be achieved? Two strategies that almost certainly overlap in practice are envisaged involving formation of drug "reservoirs" on and in the skin. The former requires creation of a residual film of formulation, in which the drug maintains at least some solubility, with good substantivity and resistance to washing and wear (6). The latter depends on manipulation of the formulation's metamorphosis post-application to relatively rapidly transfer sufficient drug into the outer layers of the SC, the slow diffusion from which can subsequently control delivery of the active to the underlying tissue. This so-called "reservoir" function of the skin (7) has been recognised for many years (8).

Clearly, therefore, the objective of sustained topical delivery will depend on those formulation characteristics that (a) comprise the residual surface film, (b) the volatile excipients, the evap-

orative loss of which drives drug into the SC upon initial application (possibly resulting in transient supersaturation), and (c) the physicochemical properties of the drug that governs its affinity for, and ability to diffuse through, the SC (9).

2 Film-forming systems for topical application

The use of polymeric FFS created *in situ* for topical application is relatively new, although such systems are well-known as controlled-release film coatings of solid oral dosage forms and have been investigated for transdermal delivery as well (10-13). In the latter case, the product Axiron[®] topical solution (Eli Lilly and Co., Indianapolis, IN), a polymeric FFS developed by Acrux, Inc. (Melbourne, Australia), for the transdermal delivery of systemically active testosterone was approved by the U.S. Food & Drug Administration (FDA) in 2010 (14).

The disadvantages of conventional topical dosage forms, especially ointments, include their poor cosmetic attributes (visual appearance and perceived skin feel) and long drying time, making application of medication inconvenient and time-consuming for patients (4, 15) and resulting in poor compliance (16-18). A better and more patient-friendly formulation would be fast drying, aesthetically pleasing, and able to deliver the drug over longer periods of time, (\geq 24 hours, for example). FFS have displayed good tolerance *in vivo* (11, 19), and fulfil many of these requirements supporting the rationale for their study and development. These attributes have been reinforced in a more recent study (20), in which the efficacy of a clobetasol spray was associated with factors beyond simple patient compliance.

The use of FFS technologies for topical application has been examined over the last decade both directly and indirectly. Indeed, some topical gels, already on the market, can be considered as FFS given that the gelling polymers used also display film-forming properties. After their application, a film is eventually formed on the skin, albeit more slowly than from a polymeric solution due to the higher viscosity. For example, DuraPeelTM (Nuvo Research Inc., Mississauga, Ontario, Canada) incorporation into a cream or a gel facilitates formation of clear films on the skin from which the drug substance may then be released for up to 12 hours Page **6** of **38** (21). A FFS based on PharmaDur[®] (Polytherapeutics, Inc., Bridgewater, New Jersey) graft polymers also enables imperceptible and invisible films to be created *in situ* from gelling top-ical solutions (22).

With respect to the treatment of skin disease, an example of a FFS for drug delivery to the SC is Lamisil[®] Once (Novartis Consumer Health SA, Nyon, Switzerland) which contains the anti-fungal agent terbinafine for the treatment of dermatophytoses. The increased residence time of the product on the skin has permitted the daily applications of the conventional cream or gel formulations to be replaced with a single administration (23, 24). Another illustration is provided by the MedSprayTM (Patch in a Can[®]) technology (MedPharm, Ltd., Guildford, U.K.), an aerosol, propellant- and solvent-based polymeric FFS for topical, dermal and transdermal drug delivery (25) that aims to create *in situ* a supersaturated, residual drug film. The performance of the approach has been studied using various corticosteroids (26-29), and a comparative clinical study of MedSprayTM 1% w/w terbinafine and Lamisil[®] Once (topical solution), both dosed only once, has demonstrated comparable anti-fungal activity and positive consumer acceptability (Figure 1) (21).

2.1 Film-formation mechanism

FFS can be either dispersions or solutions of film-forming polymer, depending on the solubility of the polymer in the selected solvent; this difference in starting vehicle will influence the film-formation mechanism (Figure 2) and the mechanical properties of the resulting film (30-32). Films formed from dispersions, especially emulsions, tend to appear cloudy and are less cosmetically acceptable as compared to the generally preferred transparent films formed from solutions. In solution, the polymer chains are highly mobile but come into closer contact as the solvent evaporates, eventually forming a film. From a polymer dispersion, the film is created via the physical process of polymer particle coalescence, the particles deforming as capillary forces increase with solvent evaporation. Incorporation of a plasticiser is often required in this case to lower the minimum film-forming temperature (thereby softening the polymer particles and facilitating their coalescence) (31). For both types of film, the rate of film-formation and the microstructure of the film depend on the rate of solvent evaporation that can, in turn, give rise to differences in drug release profiles (30).

3 FFS vehicle

Drug release from all topical delivery systems, and the rate and extent of the compound's subsequent skin uptake and penetration (i.e., local bioavailability and efficacy) depend sensitively on the composition of the applied formulation (15, 33-36). Interactions between drug, vehicle and skin are complex and determine how the active partitions into, and subsequently diffuses through, the barrier (18). Formulation optimisation is typically directed at maximising the penetration of the drug to its site of action, and this means maximising the thermodynamic activity (37, 38). Classically, the approach has been to develop formulations in which the drug is either at a concentration close to its saturation solubility, or is present as a suspension. For polymeric FFS, of either the solution or dispersion type, there is transformation of the formulation post-application as volatile excipients evaporate to leave a residual film. The delivery profile likely comprises at least two phases, therefore: a rapid uptake of drug as it concentrates in the FFS when solvent(s) evaporate(s), followed by a slower profile governed by release from the residual film. A clear challenge, as a result, is not only to formulate effectively for the initial delivery of drug into the skin, but also to ensure that sufficient drug is left in molecular (as opposed to solid) form in the residual film since only the dissolved compound is capable of diffusing (33). Because of these constraints, it is self-evident that the optimal vehicle will depend on the specific drug and perhaps on its intended concentration too (18, 33, 39).

Table 1 lists properties of the FFS vehicle and the drug substance that may affect release and delivery into the skin. Many factors are interrelated and knowledge of their interactions is essential in understanding the mechanism(s) of drug delivery from FFS and its optimisation.

3.1 Metamorphosis of FFS vehicles

Post-application to the skin, the quantitative composition of a FFS changes significantly, in particular through the loss of volatile excipients (18, 33), resulting ultimately in the creation of the residual polymeric film. In this process, the drug concentration increases, quickly reaching saturation, and with the distinct possibility of (albeit transient) supersaturation on the skin surface and/or in the upper SC (18, 40-42). Enhanced drug flux is therefore anticipated, even greater than the theoretical maximum for the period of supersaturation (25, 27, 37, 41, 43-47). The latter phenomenon also supports the idea that FFS application enables a drug reservoir to be established in the skin from which sustained delivery can be subsequently achieved. Figure 3 schematically outlines the drug delivery consequences of this 'metamorphosis' of a FFS.

Supersaturating formulations (i.e., those which result in the degree of saturation of the drug exceeding 1.0) are inherently thermodynamically unstable, and it is only a matter of time before crystallisation of the drug occurs within the residual film (48). If the solubility of the compound in this film is low, then further delivery is compromised because only drug in molecular form can diffuse (and re-dissolution is likely to be slow or negligible due to the low solubility) (28, 44, 49, 50). To mitigate against this challenge, and to inhibit crystallisation during storage (28, 29), anti-nucleating polymers have been a focus of research in the FFS area, as discussed further below in section 3.2.3.

3.2 Formulation parameters and modulation of release

The drug delivery characteristics of FFS are dependent, at the very least, on the following: solvent (30, 51), polymer type and concentration (10, 11, 52), plasticiser type and concentra-Page **10** of **38** tion (16, 52-55), other incorporated excipients (e.g., penetration enhancer, lipid component) (11, 56-59), and the drug (39, 60-64). Each is now considered in turn.

3.2.1 Drug

Drug penetration across the skin is primarily determined by solubility, molecular structure, and lipophilicity (60, 64). In general, smaller compounds diffuse more readily across the SC than larger ones, and maximum fluxes of drugs with molecular weights greater than 500 Daltons are very low (61, 65). Broadly speaking, skin permeability increases with increasing lipophilicity (the SC being a lipophilic barrier, consistent with its principal role of retarding water loss), at least up to a point (60). For very hydrophobic compounds, however, their very low water solubilities mean that uptake into the viable skin becomes the rate-limiting step (as opposed to diffusion through the SC). Consequently, it appears that a modest level of lipophilicity, corresponding to an octanol-water partition coefficient (P) of 10-1000 (log P = 1-3) (65, 66), coupled with finite oil and water solubilities, are ideal characteristics for good skin penetration. With respect to FFS, while skin permeation remains dependent upon the nature of the drug, the type of polymer used differs also has an impact upon the release and percutaneous absorption of the active thereafter (39). In terms of the skin 'reservoir' effect mentioned earlier, it is evident that drugs, which are more lipophilic than the 'ideal' candidates for rapid penetration, would be more suitable for achieving the sustained delivery profile sought from the FFS residual film (63, 67, 68). It is also worth noting that other molecular structural features have been linked to skin penetration, including the drug's ability to accept or donate hydrogen bonds (factors that impact already, of course, on log P); with respect to FFS, it has been reported that hydrogen-bonding between the drug and the film-forming polymer is a mechanism by which crystallisation of the former can be inhibited (45, 69).

3.2.2 Solvent

Polymeric films formed from solutions are more mechanically resistant than those created from dispersions (30, 51). This difference may be ascribed to differences in film microstructure (Figure 2) with a higher degree of polymer-polymer chain inter-penetration in films formed from solutions (30). With respect to the drug, the casting solvent in FFS can affect the crystalline state of the drug in the residual polymer film (70).

Obviously, the FFS solvent must be volatile to allow formation of the film and, for this reason, organic solvents, such as the lower alcohols (ethanol, isopropanol), are preferred. Polymer solubilities in organic solvents are typically high and, although less environmentally friendly than aqueous-based solvents, for example, the fast evaporation of volatile organic solvents leads to short FFS drying times and better convenience for the patient. Ethanol is the solvent of choice (71), and despite reports of it provoking skin irritation when used at high levels in dermal formulations, the FDA accepts >95% of the dehydrated solvent in topical solutions (72). Ethanol has also been described as a penetration enhancer (58) and there is no question that it facilitates the initial partitioning and uptake of lipophilic drugs into the SC upon application of a FFS. However, on the whole, the literature is limited in terms of the effect of the organic solvent used to create a topical FFS delivery system; indeed, when FFS were tested for transdermal delivery, little or no difference in delivery could be associated with the specific organic solvents used (ethanol, isopropanol and ethyl acetate, including binary mixtures thereof) (39).

3.2.3 Film-forming polymers

Various polymers are potential film-formers, such as cellulose derivatives (11, 31, 73-75), acrylate polymers or copolymers (11, 31, 73, 74), methacrylate polymers or copolymers (11, 12, 31, 59, 73, 74, 76, 77), silicones (11, 73), and vinyl polymers (12, 29, 59, 73-75) (Figure 4). The nature and the concentration of the polymer affect the mechanical properties and cosmetic attributes of the formed film (12, 31, 73, 75-78), as well as its ability to deliver the drug (10-12, 31, 39, 52, 76, 79). Personal experience demonstrates that the film formed on the skin is thick and rigid when the polymer concentration used is too high. Such non-flexible films are uncomfortable to wear, and show clearly that the sensorial, cosmetic attributes of the film should be considered during their development. As far as drug delivery is concerned, while hydroxypropyl cellulose films display long-term, zero-order drug release, for example, those formed with the more hydrophobic polyacrylate and polymethacrylate polymers produce an initial 'burst' release of the compound with the potential to establish a drug reservoir in the SC (Figure 5) (79). Differences in release kinetics can be attributed to factors such as the diffusivity of the drug in the polymeric network and the physical state of the drug (crystalline, amorphous or dissolved) in the film, which in turn depend on the physicochemical properties of the polymer and drug and the interactions between them.

The moisture sorption of polymeric films increases with increasing hydrophilicity of the polymer (53, 76). The plasticising effects of water on the polymer network (80, 81) can increase drug diffusivity and release. The water-solubility of hydrophilic films makes them less resistant to removal (e.g., by washing or in perspiration) and unlikely therefore to have sufficient skin substantivity to function as an external drug reservoir. As with hair care products, substantivity is not only determined by the water solubility of the polymer, but also by its net charge and interaction with skin surface protein and lipids (82). As the skin surface has a net negative charge at physiological pH, films created with cationic polymers display higher substantivity than those formed from neutral or anionic polymers. Furthermore, attractive forces such as hydrogen bonding and van der Waals forces suggest the use of lipophilic and proteinlike polymers (83, 84).

The concentration of the film-forming polymer is crucial. If it is too low, then the film formed can be rather weak (39). On the other hand, a higher polymer content (resulting in a denser film network) may retard drug release (10, 28, 52). A higher polymer concentration also increases the FFS viscosity and increases its drying time, upon application to the skin (28).

Polymer blends have been used to design controlled-release coatings for solid oral dosage forms. Blending hydrophobic and hydrophilic polymers has proven effective, with the hydrophilic polymer component forming pores upon hydration (30, 31, 55, 85). A similar approach has been demonstrated for topical polymeric films as well, to achieve a wide range of release rates and also, in some cases, a different release mechanism (10, 76, 77, 86, 87). However, polymer blend systems are more complex, and phase separation and plasticiser redistribution between the polymers, can occur leading to inconsistent performance (31, 88).

Some film-forming polymers display anti-nucleating properties, i.e., they prevent or inhibit nucleation and crystallisation of the drug (43, 50, 70, 89, 90). Figure 6 illustrates the antinucleating effect of hydroxypropyl cellulose and polymethacrylate copolymer on betamethasone 17-valerate (BMV). Mitigating against drug crystallisation is an obviously desirable feature of film-forming polymers, which can lengthen the period that supersaturation is maintained, and possibly permit a higher degree of supersaturation to be achieved. Because of the importance of maintaining solubilised drug in the topical film, inhibition of crystallisation is Page 14 of 38 key to the establishment of a drug reservoir for sustained delivery. The mechanism of antinucleation involves preferential interaction of the polymer chains with the drug in molecular form thereby hindering solute-solute interactions that otherwise lead to formation of crystals (50, 86). Although other mechanisms of anti-nucleation have been suggested in the literature, the inhibition of crystallisation of the corticosteroid, hydrocortisone acetate, has been explained by its association with the polymer via hydrogen bonding(90). It follows that the antinucleation efficiency of a polymer may depend sensitively upon the manner in which it interacts at a molecular level with the specific drug of interest (Figure 6) (91, 92).

3.2.4 Plasticiser

Plasticisers are typically low molecular weight additives that impart flexibility to a polymer. Organic esters, phosphate esters, fatty acid esters, and glycol derivatives are examples of commonly used plasticisers (53, 73, 76, 93, 94). Incorporation of a plasticiser results in a less brittle, stronger, and more flexible film (94, 95). This improvement in mechanical properties is a consequence of the plasticiser increasing the free volume between the polymer chains, and thereby increasing their mobility (95). Plasticisation reduces the polymer glass transition temperature (T_g), above which the chains are mobile (95) and the formed film is flexible. Ideally, this desirable feature is achieved below the skin temperature (\sim 32°C) (96) allowing the film to adapt to the movement of the skin with improved substantivity. Furthermore, incorporation of plasticiser reduces the minimum film-forming temperature and facilitates polymer coalescence and film-formation.

The nature of the plasticiser and its concentration determine its impact on film formation (53-55, 97). Examples of plasticisers of varying lipophilicity are shown in Table 2. Generally, the most efficient plasticisers have structural features in common with the polymer(s) into which they are incorporated. The impact of a plasticiser on drug release and delivery depends on whether a dispersion or solution FFS is being used. In the former case, the plasticiserfacilitated coalescence of polymer particles results in a more complete and dense film, from which slow drug release has been shown (30, 54). In contrast, plasticisation of solution FFS does not affect film formation to the same extent, but generally increases both polymer chain flexibility, and drug diffusivity and release (27, 76, 79, 88, 93). Further, the compatibility of plasticiser and polymer can have an important effect on drug release as well (55); for example, the lipophilic plasticisers, tributyl citrate (TBC) and dibutyl sebacate (DBS), incorporated into a hydrophobic, polyacrylate copolymer-based FFS, enhanced BMV release more than was observed when the polymer was the hydrophilic, hydroxypropyl cellulose (Table 3) (79).

3.2.5 Other excipients

The incorporation of penetration enhancers (58) into FFS can increase drug delivery (7, 11, 98) and establish a larger reservoir in the SC (9). The extent of enhancement depends on a number of factors (including concentration), and binary mixtures of enhancers can induce synergistic effects (11). However, it is recognised that there is always the risk of skin irritation when enhancers are used (99, 100).

The incorporation of a lipid excipient, such as medium chain triglycerides (MCT), into a FFS has been shown to result in a structured, two-phase polymeric film (Figure 7). The softer lipid-enriched inclusions provide an environment in which the solubilised drug is released quickly in an initial phase (59, 77, 92) and then in a sustained fashion thereafter (79, 92).

4 Expert Opinion

Polymeric film-forming systems (FFS) created *in situ* are promising sustained delivery platforms for topical drugs. With an appropriate vehicle composition, FFS can facilitate an initial 'burst' release of drug, establishing a reservoir on and/or in the stratum corneum, from which a sustained, slower delivery to target sites in the lower skin layers can subsequently occur. Evaporation of the volatile solvent component of the FFS causes a 'metamorphosis' of the formulation that results in a residual polymeric film. A transient period of drug supersaturation, perhaps extended by the judicious use of anti-nucleant polymers, during the transformation of the vehicle can be exploited to optimise the delivery profile.

The current literature does not identify a unique "recipe" for an ideal FFS formulation. Rather, it is clear that the selection of polymer, plasticiser, volatile solvent(s) and other excipients must be tailored to the properties of the drug being delivered. Open questions pertaining to the pros and cons of hydrophilic versus hydrophobic polymers, the benefits of plasticis-er/excipient incorporation, and strategies to inhibit drug crystallisation as the organic solvent evaporates, demand further research work at this time.

In terms of the specific aim to use FFS as sustained drug delivery platforms for topical drugs, it is clear that the water solubility of films based on hydrophilic polymers limits their residence time on the skin and undermines their perceived utility. For the moment, hydrophobic polymer-based FFS show greater potential, their water-resistance permitting increased substantivity and a prolonged residence time on the skin, with the consequent formation of a drug reservoir both on and within the outer SC. The metamorphosis of the FFS is crucial for the formulation's ultimate utility as a drug delivery platform: Is a supersaturated state achieved and for how long? Are the anti-nucleant properties of the polymer sufficient to retard/inhibit Page **17** of **38**

drug crystallisation significantly? How can drug and polymer characteristics be matched to maximise favourable interactions to retard/inhibit crystallisation? What are the roles of plasticiser and (for example) lipid excipients in an optimised FFS formulation?

With respect to the selection of appropriate drugs for delivery with the FFS approach, it is logical that more lipophilic compounds have been studied in more detail. Their obvious affinity for the lipophilic SC makes the establishment of a drug reservoir therein, and sustained release therefrom, more likely. The extent to which the delivery approach might be extended to more hydrophilic drugs is unknown.

Although FFS have been shown capable of maintaining drug release *in vitro* over several days, there is a pressing need for long-term *in vivo* investigations, to fully clarify the potential of these formulations to sustain delivery to pharmacological targets in deeper skin layers and to achieve a prolonged therapeutic effect. At the same time, such studies can address optimi-sation of the practical use of FFS, including identification of convenient and fool-proof application methods and dose control (e.g., via the use of aerosols or sprays).

In conclusion, it appears that polymeric film-forming systems do have a positive role to play in the next generation of topical formulations designed to offer sustained drug delivery to and into the skin. The superior cosmetic attributes of FFS, compared to conventional semi-solid topical products, coupled with the potential for less frequent dosing regimens, are attractive features in terms of patient compliance and therapeutic outcome, especially for the treatment of chronic skin diseases.

Declaration of interest

Kit Frederiksen and Richard H. Guy have received research funding from LEO Pharma A/S. Karsten Petersson is employed at LEO Pharma A/S.

Acknowledgements

We thank Dr. Natalie Belsey for generating the SRS images in Figure 6, and Dr. Hazel Garvie-Cook for the AFM image in Figure 7.

References

1. Wester RC, Maibach HI. Cutaneous pharmacokinetics: 10 steps to percutaneous absorption. Drug Metabolism Reviews. 1983;14(2):169-205.

2. ****** Surber C, Davis AF. Bioavailability and bioequivalence of dermatological formulations. In: Walter KA, editor. Dermatological and transdermal formulation. Drugs and the pharmaceutical sciences. New York: Marcel Dekker Inc.; 2002. p. 401-98. *Elegant exposition of the metamorphosis of topical formulations upon application to the skin.*

3. Devaux S, Castela A, Archier E, Gallini A, Joly P, Misery L, et al. Adherence to topical treatment in psoriasis: a systematic literature review. Journal of the European Academy of Dermatology and Venereology. 2012;26:61-7.

4. Fouéré S, Adjadj L, Pawin H. How patients experience psoriasis: results from a European survey. Journal of the European Academy of Dermatology and Venereology. 2005;19:2-6.

5. Shergill B, Zokaie S, Carr AJ. Non-adherence to topical treatments for actinic keratosis. Patient PreferAdherence. 2013;8:35-41.

6. Couteau C, Demé A, Cheignon C, et al. Influence of the hydrophilic–lipophilic balance of sunscreen emulsions on their water resistance property. Drug Development and Industrial Pharmacy. 2012;38(11):1405-7.

7. Roberts MS, Cross SE, Anissimov YG. Factors affecting the formation of a skin reservoir for topically applied solutes. Skin Pharmacology and Physiology. 2004;17(1):3-16.

8. Vickers C. Existence of reservoir in the stratum corneum - Experimental proof. ArchDermatol. 1963;88:20-3.

9. Vickers C. Stratum corneum reservoir for drugs. In: Montagna W, van Scott E, Stoughton R, editors. Advances in Biology of Skin Pharmacology and the skin; 19721972. p. 177-89.

10. Misra A, Raghuvanshi RS, Ganga S, et al. Formulation of a transdermal system for biphasic delivery of testosterone. Journal of Controlled Release. 1996;39(1):1-7.

11. ****** Schroeder IZ, Franke P, Schaefer UF, Lehr CM. Delivery of ethinylestradiol from film forming polymeric solutions across human epidermis *in vitro* and *in vivo* in pigs. Journal of Controlled Release. 2007;118(2):196-203. *Important contribution to the field of topical film-forming polymeric formulations.*

12. Ammar HO, Ghorab M, Mahmoud AA, et al. Rapid pain relief using transdermal film forming polymeric solution of ketorolac. Pharmaceutical Development and Technology. 2013;18(5):1005-16.

13. Ammar H, Ghorab M, El-Nahhas S, et al. Polymeric matrix system for prolonged delivery of tramadol hydrochloride, Part II: Biological evaluation. AAPS PharmSciTech. 2009;10(3):1065-70.

14. Wang C, Ilani N, Arver S, et al. Efficacy and safety of the 2% formulation of testosterone topical solution applied to the axillae in androgen-deficient men. Clinical Endocrinology. 2011;75(6):836-43.

15. Wirén K, Frithiof H, Sjöqvist C, et al. Enhancement of bioavailability by lowering of fat content in topical formulations. British Journal of Dermatology. 2009;160(3):552-6.

16. Tang W, Bhushan B. Adhesion, friction and wear characterization of skin and skin cream using atomic force microscope. Colloids and Surfaces B: Biointerfaces. 2010;76(1):1-15.

17. Timm K, Myant C, Nuguid H, Spikes HA, Grunze M. Investigation of friction and perceived skin feel after application of suspensions of various cosmetic powders. International Journal of Cosmetic Science. 2012;34(5):458-65.

18. Surber C, Smith EW. The mystical effects of dermatological vehicles. Dermatology. 2005;210(2):157-68.

19. ***** Brown M, Evans C, Muddle A, Turner R, Lim S, Reed J, et al. Efficacy, tolerability and consumer acceptability of terbinafine topical spray versus terbinafine topical solution: A phase IIa, randomised, observer-blind, comparative study. American Journal of Clinical Dermatology. 2013;14(5):413-9. *Illustration of clinical efficacy of a topical FFS*.

20. Bhutani T, Koo J, Maibach HI. Efficacy of clobetasol spray: Factors beyond patient compliance. Journal of Dermatological Treatment. 2012;23(1):11-5.

21. Inc. NR. DuraPeel. Nuvo Research Inc [Internet]. 2014 4/1/2014. Available from: http://www.nuvoresearch.com/technology/durapeel.

22. Polytherapeutics. Core Technology - PharmaDur(R). Polytherapeutics Inc [Internet]. 2010 4/1/2014. Available from: <u>http://www.polytherapeutics.com/node/3</u>.

23. Kienzler JL, Queille-Roussel C, Mugglestone CJ, et al. Stratum corneum pharmacokinetics of the anti-fungal drug, terbinafine, in a novel topical formulation, for single-dose application in dermatophytoses. Current Medical Research and Opinion. 2007;23(6):1293-302.

24. James I, Loria-Kanza Y, Jones TC. Short–duration topical treatment of tinea pedis using terbinafine emulsion gel: Results of a dose–ranging clinical trial. Journal of Dermatological Treatment. 2007;18(3):163-8.

25. Brown MB, Jones SA, inventorsTopical film-forming monophasic formulations patent WO 2007/031753 A2. 2007.

26. Jones SA, Reid ML, Brown MB. Determining degree of saturation after application of transiently supersaturated metered dose aerosols for topical delivery of corticosteroids. Journal of Pharmaceutical Sciences. 2009;98(2):543-54.

27. Reid ML, Brown MB, Jones SA. Manipulation of corticosteroid release from a transiently supersaturated topical metered dose aerosol using a residual miscible co-solvent. Pharmaceutical Research. 2008;25(11):2573-80.

28. Reid ML, Jones SA, Brown MB. Transient drug supersaturation kinetics of beclomethasone dipropionate in rapidly drying films. International Journal of Pharmaceutics. 2009;371(1–2):114-9.

29. Reid ML, Benaouda F, Khengar R, et al. Topical corticosteroid delivery into human skin using hydrofluoroalkane metered dose aerosol sprays. International Journal of Pharmaceutics. 2013;452(1–2):157-65.

30. Lecomte F, Siepmann J, Walther M, et al. Polymer blends used for the coating of multiparticulates: comparison of aqueous and organic coating techniques. Pharmaceutical Research. 2004;21(5):882-90.

31. Siepmann F, Siepmann J, Walther M, et al. Polymer blends for controlled release coatings. Journal of Controlled Release. 2008;125(1):1-15.

32. Felton LA. Mechanisms of polymeric film formation. International Journal of Pharmaceutics. 2013;457(2):423-7.

33. Katz M, Poulsen BJ. Corticoid, vehicle and skin interaction in percutaneous absorption. Journal of the Society of Cosmetic Chemists. 1972;23(9):565-90.

34. Micali G, Lacarrubba F, Anurekha B, et al. The skin barrier. In: Freinkel RK, Woodley DT, editors. The Biology of the Skin. New York: Taylor & Francis; 2001. p. 219-31.

35. Wiedersberg S, Naik A, Leopold CS, et al. Pharmacodynamics and dermatopharmacokinetics of betamethasone 17-valerate: assessment of topical bioavailability. British Journal of Dermatology. 2009;160(3):676-86.

36. Ahlstrom LA, Cross SE, Mills PC. The effects of formulation on the penetration and retention of budesonide in canine skin *in vitro*. The Veterinary Journal. 2013;196(3):456-60.

37. Higuchi T. Physical chemical analysis of percutaneous absorption process from creams and ointments. Journal of the Society of Cosmetic Chemists. 1960;11(2):85-97.

38. Ishii H, Todo H, Sugibayashi K. Effect of thermodynamic activity on skin permeation and skin concentration of triamcinolone acetonide. Chemical and Pharmaceutical Bulletin. 2010;58(4):556-61.

39. Schroeder IZ. Film forming polymeric solutions as drug delivery systems for the skin [PhD thesis]: Department of Biopharmaceutics and Pharmaceutical Technology, Saarland University, Saarbrücken, Germany; 2007.

40. Huang X, Tanojo H, Lenn J, Deng CH, Krochmal L. A novel foam vehicle for delivery of topical corticosteroids. Journal of the American Academy of Dermatology. 2005;53(1, Supplement):S26-S38.

41. * Coldman MF, Poulsen BJ, Higuchi T. Enhancement of percutaneous absorption by the use of volatile: Nonvolatile systems as vehicles. Journal of Pharmaceutical Sciences. 1969;58(9):1098-102. *A classic in the field*.

42. Davis AF, Hadgraft J. Effect of supersaturation on membrane transport: 1. Hydrocortisone acetate. International Journal of Pharmaceutics. 1991;76(1-2):1-8.

43. Megrab NA, Williams AC, Barry BW. Oestradiol permeation through human skin and silastic membrane: effects of propylene glycol and supersaturation. Journal of Controlled Release. 1995;36(3):277-94.

44. Chia-Ming C, Flynn GL, Weiner ND, et al. Bioavailability assessment of topical delivery systems: Effect of vehicle evaporation upon *in vitro* delivery of minoxidil from solution formulations. International Journal of Pharmaceutics. 1989;55(2-3):229-36.

45. Kim JH, Choi HK. Effect of additives on the crystallization and the permeation of ketoprofen from adhesive matrix. International Journal of Pharmaceutics. 2002;236(1–2):81-5.

46. Pellett MA, Castellano S, Hadgraft J, et al. The penetration of supersaturated solutions of piroxicam across silicone membranes and human skin *in vitro*. Journal of Controlled Release. 1997;46(3):205-14.

47. Moser K, Kriwet K, Froehlich C, Kalia Y, Guy R. Supersaturation: enhancement of skin penetration and permeation of a lipophilic drug. Pharmaceutical Research. 2001;18(7):1006-11.

48. Brouwers J, Brewster ME, Augustijns P. Supersaturating drug delivery systems: The answer to solubility-limited oral bioavailability? Journal of Pharmaceutical Sciences. 2009;98(8):2549-72.

49. Poulsen BJ. Diffusion of drugs from topical vehicles: an analysis of vehicle effects. Advances in Biology of Skin. 1972;12:495-509.

50. Cilurzo F, Minghetti P, Casiraghi A, et al. Polymethacrylates as crystallization inhibitors in monolayer transdermal patches containing ibuprofen. European Journal of Pharmaceutics and Biopharmaceutics. 2005;60(1):61-6.

51. Bajdik J, Regdon Jr G, Marek T, et al. The effect of the solvent on the film-forming parameters of hydroxypropyl-cellulose. International Journal of Pharmaceutics. 2005;301(1–2):192-8.

52. Padula C, Nicoli S, Colombo P, et al. Single-layer transdermal film containing lidocaine: modulation of drug release. European Journal of Pharmaceutics and Biopharmaceutics. 2007;66(3):422-8.

53. Lin SY, Chen KS, Run-Chu L. Organic esters of plasticizers affecting the water absorption, adhesive property, glass transition temperature and plasticizer permanence of Eudragit acrylic films. Journal of Controlled Release. 2000;68(3):343-50.

54. Fulzele SV, Satturwar PM, Dorle AK. Polymerized rosin: novel film forming polymer for drug delivery. International Journal of Pharmaceutics. 2002;249(1-2):175-84.

55. Lecomte F, Siepmann J, Walther M, et al. Polymer blends used for the aqueous coating of solid dosage forms: importance of the type of plasticizer. Journal of Controlled Release. 2004;99(1):1-13.

56. Iervolino M, Cappello B, Raghavan SL, et al. Penetration enhancement of ibuprofen from supersaturated solutions through human skin. International Journal of Pharmaceutics. 2001;212(1):131-41.

57. Shelke NB, Sairam M, Halligudi SB, et al. Development of transdermal drug-delivery films with castor-oil-based polyurethanes. Journal of Applied Polymer Science. 2007;103(2):779-88.

58. Williams AC, Barry BW. Penetration enhancers. AdvDrugDeliverRev. 2012;64, Supplement(0):128-37.

59. Lunter D, Daniels R. *In vitro* skin permeation and penetration of nonivamide from novel film-forming emulsions. Skin Pharmacology and Physiology. 2013;26(3):139-46.

60. Potts R, Guy R. Predicting skin permeability. Pharmaceutical Research. 1992;9(5):663-9.

61. Bos JD, Meinardi MMHM. The 500 Dalton rule for the skin penetration of chemical compounds and drugs. Experimental Dermatology. 2000;9(3):165-9.

62. Cross SE, Magnusson BM, Winckle G, et al. Determination of the effect of lipophilicity on the *in vitro* permeability and tissue reservoir characteristics of topically applied solutes in human skin layers. Journal of Investigative Dermatology. 2003;120(5):759-64.

63. Jensen LB, Magnussson E, Gunnarsson L, et al. Corticosteroid solubility and lipid polarity control release from solid lipid nanoparticles. International Journal of Pharmaceutics. 2010;390(1):53-60.

64. Ito Y, Yoshimura M, Tanaka T, et al. Effect of lipophilicity on the bioavailability of drugs after percutaneous administration by dissolving microneedles. Journal of Pharmaceutical Sciences. 2012;101(3):1145-56.

65. Williams AC. Transdermal and topical drug delivery - From theory to clinical practice. London: Pharmaceutical Press; 2003 2003.

66. Brown MB, Martin GP, Jones SA, et al. Dermal and transdermal drug delivery systems: Current and future prospects. Drug Delivery. 2006;13(3):175-87.

67. Leichtnam ML, Rolland H, Wüthrich P, et al. Identification of penetration enhancers for testosterone transdermal delivery from spray formulations. Journal of Controlled Release. 2006;113(1):57-62.

68. Yano T, Nakagawa A, Tsuji M, et al. Skin permeability of various non-steroidal antiinflammatory drugs in man. LifeSci. 1986;39(12):1043-50.

69. Wegiel L, Mauer L, Edgar K, et al. Crystallization of amorphous solid dispersions of resveratrol during preparation and storage – Impact of different polymers. Journal of Pharmaceutical Sciences. 2013;102(1):171-84.

70. Pattnaik S, Swain K, Mallick S, et al. Effect of casting solvent on crystallinity of ondansetron in transdermal films. IntJ Pharm. 2011;406(1-2):106-10.

71. Leichtnam ML, Rolland H, Wüthrich P, et al. Enhancement of transdermal testosterone delivery by supersaturation. Journal of Pharmaceutical Sciences. 2006;95(11):2373-9.

72. Administration USFaD. Inactive ingredient search for approved drug products. FDA Drug Databases [Internet]. 2013 8/23/2013. Available from: http://www.accessdata.fda.gov/scripts/cder/iig/index.cfm.

73. Schroeder IZ, Franke P, Schaefer UF, et al. Development and characterization of film forming polymeric solutions for skin drug delivery. European Journal of Pharmaceutics and Biopharmaceutics. 2007;65(1):111-21.

74. Wicks ZW, Jones FN, Peppas SP, et al. Organic coatings - Science and technology. 3 ed. New Jersey: Wiley & Sons; 2006 2006.

75. Pattnaik S, Swain K, Choudhury P, et al. Alfuzosin hydrochloride transdermal films: evaluation of physicochemical, *in vitro* human cadaver skin permeation and thermodynamic parameters. Int Braz J Urol. 2009;35(6):716-29.

76. Ammar H, Ghorab M, El-Nahhas S, et al. Polymeric matrix system for prolonged delivery of tramadol hydrochloride, Part I: Physicochemical evaluation. AAPS PharmSciTech. 2009;10(1):7-20.

77. Lunter DJ, Daniels R. New film forming emulsions containing Eudragit NE and/or RS 30D for sustained dermal delivery of nonivamide. European Journal of Pharmaceutics and Biopharmaceutics. 2012;82(2):291-8.

78. Garvie-Cook H, Frederiksen K, Petersson K, et al. Characterization of Topical Film-Forming Systems Using Atomic Force Microscopy and Raman Microspectroscopy. Molecular Pharmaceutics. 2015;12(3):751-7.

79. * Frederiksen K, Guy RH, Petersson K. Formulation considerations in the design of topical, polymeric film-forming systems for sustained drug delivery to the skin. European Journal of Pharmaceutics and Biopharmaceutics. 2015;91(0):9-15. *A practical evaluation of the rational development of topical FFS.*

80. Zhang Y, Han JH. Sorption isotherm and plasticization effect of moisture and plasticizers in pea starch film. Journal of Food Science. 2008;73(7):E313-E24.

81. Bley O, Siepmann J, Bodmeier R. Characterization of moisture-protective polymer coatings using differential scanning calorimetry and dynamic vapor sorption. Journal of Pharmaceutical Sciences. 2009;98(2):651-64.

82. Robbins CR. Polymers in hair products. Chemical and Physical Behavior of Human Hair: Springer Berlin-Heidelberg; 2012. p. 489-535.

83. Burnette RR, Ongpipattanakul B. Characterization of the permselective properties of excised human skin during iontophoresis. Journal of Pharmaceutical Sciences. 1987;76(10):765-73.

84. Barel AO, Paye M, Maibach HI. Handbook of cosmetic science and technology. New York: Marcel Dekker Inc.; 2001 2001.

85. Marucci M, Hjärtstam J, Ragnarsson G, et al. Coated formulations: New insights into the release mechanism and changes in the film properties with a novel release cell. Journal of Controlled Release. 2009;136(3):206-12.

86. Misra A, Pal R, Majumdar S, et al. Biphasic testosterone delivery profile observed with two different transdermal formulations. Pharmaceutical Research. 1997;14(9):1264-8.

87. Patel DP, Setty CM, Mistry GN, Patel SL, Patel TJ, Mistry PC, et al. Development and evaluation of ethyl cellulose-based transdermal films of furosemide for improved *in vitro* skin permeation. AAPS PharmSciTech. 2009;10(2):437-42.

88. Frohoff-Hülsmann MA, Schmitz A, Lippold BC. Aqueous ethyl cellulose dispersions containing plasticizers of different water solubility and hydroxypropyl methylcellulose as coating material for diffusion pellets: I. Drug release rates from coated pellets. International Journal of Pharmaceutics. 1999;177(1):69-82.

89. Hadgraft J. Skin deep. European Journal of Pharmaceutics and Biopharmaceutics. 2004;58(2):291-9.

90. Raghavan SL, Trividic A, Davis AF, et al. Crystallization of hydrocortisone acetate: Influence of polymers. International Journal of Pharmaceutics. 2001;212(2):213-21.

91. Jain P, Banga AK. Inhibition of crystallization in drug-in-adhesive-type transdermal patches. International Journal of Pharmaceutics. 2010;394(1-2):68-74.

92. * Garvie-Cook H, Frederiksen K, Petersson K, et al. Biophysical elucidation of the mechanism of enhanced drug release and topical delivery from polymeric film-forming systems. Journal of Controlled Release. 2015;212(0):103-12. *Recent and basic research into the potential of topical FFS*.

93. Rao PR, Diwan PV. Permeability studies of cellulose acetate free films for transdermal use: Influence of plasticizers. Pharmaceutica Acta Helvetiae. 1997;72(1):47-51.

94. Rowe RC. Materials used in the film coating of oral dosage forms. In: Florence AT, editor. Materials used in pharmaceutical formulation. Oxford: Blackwell Scientific Publications; 1984. p. 1-36.

95. Sears JK, Darby JR. The technology of plasticizers. New York: John Wily & Sons; 1982 1982.

96. Development OfEC-oa. No 28 - Guidance document for the conduct of skin absorption studies. Paris: OECD Publishing; 2004 3/5/2004.

97. Shao Z, Morales L, Diaz S, et al. Drug release from kollicoat SR 30D-coated nonpareil beads: Evaluation of coating level, plasticizer type, and curing condition. AAPS PharmSciTech. 2002;3(2):87-96.

98. Luppi B, Bigucci F, Baldini M, Abruzzo A, Cerchiara T, Corace G, et al. Hydroxypropylmethylcellulose films for prolonged delivery of the antipsychotic drug chlorpromazine. Journal of Pharmacy and Pharmacology. 2010;62(3):305-9.

99. Mohammed D, Hirata K, Hadgraft J, et al. Influence of skin penetration enhancers on skin barrier function and skin protease activity. European Journal of Pharmaceutical Sciences. 2014;51(0):118-22.

100. Ting WW, Vest CD, Sontheimer RD. Review of traditional and novel modalities that enhance the permeability of local therapeutics across the stratum corneum. International Journal of Dermatology. 2004;43(7):538-47.

Figures and Tables, plus legends



Figure 1. Treatment success rates after a single application of either MedSprayTM 1% w/w terbinafine or Lamisil[®] Once (topical solution). Redrawn from data in reference (19).



Figure 2. Schematic illustration of the mechanism of polymeric film-formation from a polymer FFS solution (via interaction between mobile polymer chains) and dispersion (deformation and coalescence of polymer particles).

 Table 1. Key properties that influence drug delivery from polymeric film-forming systems.

FFS	Drug	
Solvent	Solubility	
Co-solvent	Physical state of drug in the formed film	
Drying time	Molecular weight	
Film-forming polymer	Chemical structure	
Film-formation mechanism	Hydrogen bond donating/accepting groups	
Supersaturation	Log P	
Anti-nucleation	Diffusivity	
Plasticiser	Drug load	
Penetration enhancer	Drug-vehicle-skin interactions	
Water-resistance		
Skin substantivity		
Residence time		
Film flexibility		
Contact surface area		
Surface energy of film		
Film diffusivity		
Moisture sorption		
Swelling		



Figure 3. Schematic representation of drug thermodynamic activity (upper panel) and concentration in solution (lower panel) during the 'metamorphosis' of a FFS. Initially, both

thermodynamic activity and solubilised concentration increase as volatile excipients evaporate. Drug then either reaches its limiting solubility (activity maximises, solubilised concentration peaks and then begins to fall precipitately), or transient supersaturation occurs creating a thermodynamically metastable state of relatively high solubility. Ultimately, however, this situation cannot be sustained and the system evolves to a residual film in which drug solubility is markedly reduced (but optimally sufficiently finite to continue to provide continued input of the compound into the skin).



Figure 4. Chemical structures of the repeating units of selected film-forming polymers: (a) simple polyvinyl polymers, (b) polyacrylates, (c) polymethacrylates, (d) silicones, and (e) cellulose derivatives.



Figure 5. In vitro release of betamethasone-17-valerate from a hydroxypropyl cellulose and polyacrylate copolymer film (mean \pm standard deviation; n=3). The inset is a zoom of the initial burst release from the film. Redrawn from data in reference (79).



Figure 6. <u>Upper panels</u>: Stimulated Raman scattering (SRS) images of deuterated BMV crystals formed in/on the skin within 30 minutes post-application in either ethanol (left image) or in a FFS based on hydroxypropyl cellulose (right image). SRS contrast is obtained at 2120 cm⁻¹ corresponding to the $-CD_2$ stretching vibration. <u>Lower panels</u>: Micrographs illustrating the differential anti-nucleation efficiency of FFS prepared with BMV and either hydroxypropyl cellulose (left image) or polymethacrylate copolymer (right image).

Table 2. Examples of plasticisers used in polymeric film-forming systems.

Plasticisers
Acetyltributyl citrate, acetyltriethyl citrate
Dibutyl phthalate, diethyl phthalate, dimethyl phthalate
Dibutyl sebacate, diethyl sebacate
Triacetin
Tributyl citrate, triethyl citrate

Table 3. Enhancement ratios of the cumulative release of BMV in 72 hours from hydrophilic and hydrophobic polymer-based FFS when plasticised with either TBC or DBS (data taken from (79)).

	Plasticiser	
FFS polymer	TBC	DBS
Hydroxypropyl cellulose	2.1	2.8
Polyacrylate copolymer	2.6	3.4



Figure 7. Atomic force microscopy image showing the two-phase structure of a hydroxylpropyl cellulose film incorporating MCT.