Comparison of the degradations of diphenamid by homogeneous photolysis and heterogeneous photocatalysis in aqueous solution

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- 13
- 14 Abstract

15 In this work, the homogeneous and heterogeneous degradations of diphenamid (DPA) in aqueous solution were conducted by direct photolysis with UVC (254 nm) and by 16 17 photocatalytsis with TiO₂/UVA (350 nm), and the experimental results were compared. It was found that the homogeneous photolysis by UVC irradiation alone was quite efficient to 18 19 degrade DPA up to 100% after 360 min, but was very inefficient to mineralize its 20 intermediates in terms of dissolved organic carbon reduction of only 8%. In contrast, the 21 heterogeneous photocatalysis with TiO₂/UVA showed relatively a lower degree of DPA 22 degradation (51%), but a higher degree of its mineralization (11%) after 360 min. These results reveal that the photocatalysis process has relatively poor selectivity to degrade 23 24 different compounds including various intermediates from the DPA degradation, which is 25 beneficial to its mineralization. In addition, over 20 intermediates were identified by LC-MS and ¹H-NMR analyses. Based on the identified intermediates, the reaction mechanisms and 26 27 the detailed pathways of the DPA degradation by photolysis and photocatalysis were 28 proposed, and are presented in this paper.

30 *Keywords:* Diphenamid; Herbicide; NMR; Photolysis; Photocatalysis

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33 **1. Introduction**

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Diphenamid (DPA) as a pre-emergent herbicide is widely employed for control of annual grasses and broadleaf weeds in tomato, potato, peanut, and soybean plants (Schultz and Tweedy, 1972; Sirons et al., 1981). This substance does enter the environment under normal use or through inappropriate disposal and is harmful to aquatic organisms. Therefore, the investigation of remediation treatments of polluted waters containing trace amounts of herbicides is of environmental interests.

41 In the past decades, research on the photochemical degradation of DPA in aqueous 42 solution has progressed quickly because of the high efficiency of mineralization under the mild operating conditions (Rosen, 1967; Schultz and Tweedy, 1972; Rahman et al., 2003). 43 44 For example, Rosen (1967) studied the homogeneous photodegradation of DPA by UV and 45 sunlight irradiation, but the mechanism and the major products from DPA degradation were not well identified. Rahman et al. (2003) investigated the photocatalytic degradation of DPA 46 47 in aqueous P25 TiO₂ suspension under the illumination of a medium pressure mercury lamp. 48 Although they identified five intermediates using a GC-MS technique and suggested a brief 49 pathway of the DPA degradation, the five products identified in their study were not 50 sufficient enough to buildup a thorough pathway delineating the photocatalytic degradation 51 of DPA due to the complexity of the DPA molecular structure. Furthermore, the medium 52 pressure mercury lamp used in their experiment offers a broad band of light emission from 53 200 to 600 nm, thus resulting in both types of processes, homogeneous photolysis and 54 heterogeneous photocatalysis, to possibly occur in the DPA degradation. In short, the literatures reported previously did not supply a clear clarification of the photochemical 55

reactions for the degradation of DPA pollutants in water. Therefore, in order to better understand the difference of DPA degradation by homogeneous photolysis and heterogeneous photocatalysis, two sets of experiments for DPA degradation in aqueous solution by direct photolysis with UVC and photocatalysis with TiO_2/UVA were conducted in this study. In which, the reaction mechanisms and pathways of DPA degradation by direct photolysis and photocatalysis were proposed based on over 20 intermediates identified by the combination of ¹H-NMR and LC-MS analyses.

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64 **2. Experimental**

65 2.1. Materials

DPA chemical (99.9%) was purchased from Aldrich Chemical Company. Self-prepared TiO₂ films as reported before (Liang and Li, 2009) were used as photocatalysts. 0.5 M HCl and 0.5 M NaOH solutions were used to adjust the solution pH. Other chemicals such as CDCl₃ and CH₃CN (Aldrich) were used without further purification. Deionized distilled water was used throughout the experiments.

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72 2.2. Experimental setup and procedure

73 A UV-photoreactor system consists of a quartz reactor, an external UV light source, one piece of TiO₂ film with an area of 3 cm², and 25 mL of aqueous DPA solution. The UV lamp 74 75 was placed perpendicularly on the top of the reactor and the distance between the lamp and 76 the surface of solutions was ~6 cm (see Supplemental Material (SM), Fig. SM-1). In order to 77 compare the effect of UV wavelengths on the photochemical behavior of direct photolysis 78 and photocatalysis, DPA solutions were irradiated with different UV lamps: (a) UVC region, 79 using a G8T5 germicidal lamp with a maximum at 254 nm (Sankyo Denki Co. Ltd., Japan); 80 (b) UVA region, using a F8T5/BL-B black light blue lamp with a wavelength range of 31881 400 nm and its main emission at 350 nm (Hitachi, Japan). Both UV lamps had an equivalent 82 power consumption of 8 W and the corresponding light intensity detected in the experiment was 8.02×10^{-4} W cm⁻² for UVC and 1.01×10^{-3} W cm⁻² for UVA, respectively. Prior to 83 photocatalytic reaction, the DPA solution of 20 mg L^{-1} was magnetically stirred in the dark 84 for 60 min in order to achieve adsorption/desorption equilibrium. Then the DPA 85 concentration (~19.2 mg L^{-1}) in the bulk solution at this time was used as an initial value. All 86 87 reactions were performed at room temperature of ~21 °C. During the photocatalytic reaction, the samples were collected from the solution at different intervals. 88

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90 2.3. Chemical analysis

91 The DPA concentration was determined by HPLC (Finnigan SpectraSYSTEM P4000) consisting of a Pinnacle II C18 reverse-phase column (5 μ m, 4.6 mm \times 250 mm) and a UV 92 93 detector (UV 6000LP) at 254 nm. The mobile phase consisted of acetonitrile/water (v:v = 3:2) with 0.5 vol% acetic acid/phosphoric acid, and flowed at 1.0 mL min⁻¹. In order to identify 94 95 the organic reaction intermediate species, combination of LC-MS and ¹H-NMR analyses were employed. All one-dimensional and two-dimensional (2D) ¹H-NMR spectra were 96 97 recorded at ambient temperature using a NMR spectrometer (Bruker Avance 600 MHz) 98 equipped with a tick carboxypeptidase inhibitor cryoprobe. To avoid missing out any 99 intermediates at a low concentration during the NMR analysis, it is necessary to extract the organic substrates in the final water samples with chloroform (HPLC grade) before the NMR 100 detection. Although the DPA solutions with an initial concentration of 20 mg L⁻¹ were used in 101 102 the experiments to determine the kinetic reaction rate constants (k) of photolysis and photocatalysis reactions, the higher concentration of DPA at 130 mg L⁻¹ was used in the 103 experiments to identify the intermediates by ¹H-NMR analysis. In the meantime, an ion trap 104 mass spectrometer (Finnigan LCQTM DUO) coupled to LC-MS was also used to further verify 105

106 the reaction intermediates through electrospray ionization (ESI). The ESI probe was installed 107 with sheath and auxiliary gasses at 60 and 20 units, respectively. The mass spectrometer was 108 operated in the positive/negative ion mode in the m/z 50-400 range for LC-MS and LC-109 MS/MS. The MS conditions were set at capillary temperature = $250 \,^{\circ}$ C, voltage = $46 \,$ V and 110 spray voltage = $4.5 \,$ kV.

111 Dissolved organic carbon (DOC) concentration was determined by a total organic carbon (TOC) analyzer (TOC-5000A, Shimadzu, Japan) equipped with an auto-sampler (ASI-5000). 112 O_2 as a carrier gas was used in the detecting system. In this analysis, organic carbon = total 113 114 carbon (TC) – inorganic carbon (IC). Under the acidic condition, all inorganic carbon was 115 converted to CO₂ and measured. Prior to the TOC analysis, two standard solutions, potassium hydrogen phthalate (KOOC•C₆H₄•COOH) solutions in the range of 10-100 mg L^{-1} and 116 sodium carbonate (Na₂CO₃) solutions of 1-10 mg L^{-1} , were used to calibrate TC and IC 117 concentrations respectively. The minimum acceptable correlation coefficient (R^2) was 0.9991 118 and the detection limit of the method was 0.1 mg L^{-1} . 119

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121 **3. Results and discussion**

122

123 3.1. Direct photolysis and photocatalysis of DPA in aqueous solutions

The photodecay of DPA by UV light radiation with and without catalyst is thought to follow the pseudo-first-order reaction with respect to the DPA concentration (Rahman et al., 2003). In this study, the semi-log graphs of the DPA degradation by photolysis and photocatalysis versus irradiation time yield straight lines, indicating the pseudo-first-order reaction kinetics. The reaction rate constants (*k*) were evaluated from the experimental data by the linear regression. In all cases R^2 values are higher than 0.97, indicating that the exponential model can well describe the kinetics for DPA degradation in both processes. DOC was also monitored to compare the degree of DPA mineralization in both processes atdifferent pH.

133

134 [Table 1]

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136 The two sets of experiments in aqueous DPA solutions without TiO₂ film at pH 3.5, 7.2 and 9.6 were first conducted under UVC and UVA illumination, respectively for 360 min. 137 138 The experimental results are shown in Table 1. It can be seen that while the direct photolysis 139 with UVA showed almost no reduction of DPA after 360 min at three pH values, the direct 140 photolysis with UVC showed that, DPA was degraded by 100% after 360 min at pH 3.5, 7.2 141 and 9.6, but the corresponding DOC was only reduced by 11, 8 and 9%, respectively. These 142 results indicate that DPA cannot be degraded by direct photolysis under UVA illumination (350 nm) because of its main light absorption at ~260 nm, but it can be excited by UVC light 143 144 (254 nm) and then quickly degraded through direct electron transfer reaction. However, the 145 direct UVC-photolysis had poor mineralization when the DOC removal is considered to 146 reflect the degree of mineralization. The effect of pH on DPA degradation indicated that DPA 147 was degraded from fast to slow in an order of pH 3.5 > 9.6 > 7.2. It can be attributed to the 148 unique ionic states of DPA under acidic and alkaline conditions. Both the acidic and alkaline 149 conditions favored UV absorption by DPA (Fig. SM-2), resulting in faster homolytic or 150 heterolytic breakages in the DPA molecules. With the further photolysis reaction, some acidic 151 products were formed from the DPA degradation.

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To conduct a photocatalysis reaction without direct photolysis, aqueous DPA solution in presence of TiO_2 film was irradiated by the UVA light in the wavelength range of 318-400 nm under different pH conditions and the experimental results are also presented in Table 1. 156 It can be seen that at pH 3.5, DPA was degraded by 63% and DOC was reduced by 13% after 157 360 min whereas at pH 9.6, DPA was degraded by 42% and DOC by 8%. These results 158 indicate that DPA can be degraded by the photocatalysis with TiO₂/UVA, but more slowly 159 than that by the photolysis with UVC. Furthermore, it was found that under acidic and neutral conditions, the DOC removal by the photocatalysis (13 and 11%, respectively) was higher 160 161 than that by photolysis (11 and 8%, respectively). These results might indicate that photocatalysis process has poor selectivity for degrading different intermediates from the 162 163 DPA degradation, which is beneficial to DPA mineralization, especially under the acidic 164 condition, while the direct photolysis showed the relatively lower DOC removals.

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166 3.2. Mechanism of homogeneous and heterogeneous DPA degradation

167 TiO₂-based photocatalysis is well known to produce •OH radicals in water, through the 168 interface oxidation of hydroxide anions or water molecules adsorbed on the semiconductor 169 surface by the holes photogenerated in the valence band of semiconductor. The •OH radicals 170 produced by TiO₂ are the powerful oxidizing species and can initiate the degradation of 171 aromatic compounds by direct attack to the aromatic rings, leading to the formation of 172 hydroxylated intermediate species (as shown in the later discussion). At sufficient contact time and proper operation conditions, it is practically possible to mineralize the target 173 174 pollutant to CO₂ and H₂O. This property results in good mineralization for the TiO₂/UVA 175 photocatalysis. However, it should be noted that the photocatalytic process mainly occurs on 176 the photocatalyst surface, but not in the bulk solution; in other words, the photocatalytic process depends on the adsorption and diffusion of DPA/intermediate molecules on the TiO₂ 177 178 surface, thus leading to a slow rate of DPA degradation.

179 In contrast, the mechanism of the direct UVC-induced photolysis ($\lambda = 254$ nm) is based 180 on the fact that the chemical species undergo the homogeneous reactions in solution, by

181 which molecules are broken down to smaller molecules as a result of photodecomposition of
182 the excited organic compounds. The sequence of events that could occur during a particular
183 DPA photolytic decomposition is summarized below.

 $DPA + hv \rightarrow DPA^* \rightarrow [DPA^{\bullet^+} + DPA^{\bullet^-}]^*$ 184 excitation of DPA (1) $[DPA^{\bullet^+} + DPA^{\bullet^-}]^* \longrightarrow DPA + heat$ 185 thermal deexcitation of DPA (2) $[DPA^{\bullet^+} + DPA^{\bullet^-}]^* \rightarrow I_{uv}$ decomposition of DPA and formation of intermediates (3) 186 $I_{uv} + hv \longrightarrow I_{uv}^*$ excitation of intermediates 187 (4) 188 I_{uv}* \rightarrow products decomposition of intermediates (5)

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Here Eq. 1 involves the excitation of DPA, in which the photo-excited state of DPA^{*} takes 190 191 one-electron from the ground state of DPA molecule to produce radical anion DPA. Subsequent reaction can either undergo thermal deexcitation (Eq. 2) via recombination of 192 radical ions or photodecomposition to produce intermediate I_{uv} (Eq. 3). Some intermediates 193 I_{uv} can also absorb UV light to produce excited state intermediate I_{uv} * as shown in Eq. 4, 194 further undergoing the decomposition towards final products step by step (Eq. 5), but some 195 196 cannot when energy absorded by intermediate molecules directly from UV light is not 197 enough to cause the bond scission. The formation of resistant intermediates just explains the 198 fact that generally direct photolysis has a lower degree of mineralization than that of 199 photocatalysis with TiO₂.

200 201

202 3.3. Identification of intermediates

Both the LC-MS and ¹H-NMR analyses were used to detect the intermediates. It should be noted that the NMR technique can avoid the escaping of some polar intermediates and positional isomers generally occurring in the MS analysis alone. Table 2 and Table SM-1 summarized the analytical results along with the proposed structures for the detected

207 byproducts, in which 21 intermediate products were identified according to the mass ion 208 peaks from MS spectra and the characteristic proton patterns from ¹H-NMR spectra. Three 209 principal groups of intermediate products were identified in both homogeneous and 210 heterogeneous degradation processes: (i) products 1–4 from the reactions relevant to only the 211 N-methyl oxidation or replacement; (ii) products 5-12 from the reactions of the 212 hydroxylation of the aromatic ring; and (iii) products 13–21 from the N-demethylation, 213 oxidation and ring opening. The intermediate products 1-7 were listed in Table 2 and other 214 products 8-21 were shown in Table SM-1.

215

216 [Table 2]

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218 In this paper, both positive and negative ion full-scan mode experiments over the mass 219 range from m/z 50 to 400 were used to determine the m/z of the individual components 220 related to DPA and its degradation products, as shown in Fig. 1. For comparison, the 221 fragmentation pattern obtained from photolysis (Fig. 1a) was analogous to that of TiO₂-222 photocatalysis (Fig. 1b), for the most part, and supports the assigned structure (see Table 2). 223 For example, the major ions with m/z 256 and 272, corresponding to mono- and bi-224 hydroxylation respectively, were both observed in spectra (a) and (b). The major ion with m/z225 333 is due to the couple of fragments of $-CH(C_6H_5)_2$, whereas the minor ion (m/z 196) should 226 be interpreted as arising from cleavage of the bond between the carbonyl group and the N-227 dimethyl group. By analogy, formation of fragment ions at m/z 105 and 123 in MS spectra of 228 byproducts 18 and 19 can be rationalized. The structures of the identified fragments (m/z 287, 229 301, 316, 366) are shown in Table SM-1. It should be pointed out that this type of 230 fragmentation was already observed for iodosulfuron degradation by Sleiman et al. (2006).

231 [Fig. 1]

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233 Some of ¹H-NMR spectra were used to further determine the chemical structures of the 234 degradation products of DPA as shown in Fig. 2. Figure 2a depicts the p-, o- and m- regions 235 (5.35–5.70 ppm) of the 2D-COSY spectra of DPA solutions after the irradiation by direct UVC photolysis for 1 h and by TiO₂/UVA photocatalysis for 24 h. This is the time at which 236 237 the DPA degradation exceeded 50% and the concentrations of its products were sufficient to 238 be observed clearly in the NMR analysis. 2D-COSY analysis was conducted in our study 239 because in a COSY spectrum the coupling interaction is generally allowed to identify which 240 protons couple to each other. From Fig. 2a, it is interesting to find that for the DPA molecule 241 there is the existence of a strong cross peak that corresponds to the chemical shifts of both 242 protons, in which the horizontal line from the spot at 3.00 ppm corresponds to the methyl 243 protons while the vertical line drawn from the spot at 5.58 ppm corresponds to the methine 244 protons. The presence of this cross peak, which correlates the methyl protons and methine 245 protons, confirms that (1) such COSY method can detect the long-range coupling interaction 246 that extends to five bonds (Pavia et al., 2001) and (2) the methyl and methine groups must be 247 present simultaneously in DPA molecules. Besides this, other four types of cross peaks, 248 similar to the nature of DPA molecule, were also observed in this COSY spectrum. They are 249 nonequivalent, probably owing to the nuclear magnetization effect of the methyl neighbor 250 group. Therefore, combining the specifical proton information with corresponding MS data, 251 the four intermediates can be identified as compounds 1–4 arising from oxidation or removal 252 of terminal N-methyl group in DPA molecule, simultaneously appearing in the UVC 253 photolysis and TiO₂/UVA photocatalysis processes.

254

255 [Fig. 2]

257 The second category of products includes different phenolic compounds resulting from 258 the hydroxylation of benzene ring during the UVC and TiO₂/UVA processes. The different 259 intermediates reflect the difference in the reaction mechanisms of two processes: the 260 homogeneous degradation of DPA by photolysis is dependent upon the bond strength of 261 DPA/intermediate molecules and their energy absorbing ability, while the heterogeneous 262 degradation of DPA by photocatalysis is mainly affected by the attack of •OH radicals. 263 Among these products, compounds 5, 6 and 7 are positional isomers and they were discriminated by ¹H-NMR rather than MS analysis. Since the chemical shift of ortho-proton 264 265 of most phenolic compounds is 6.0–7.0 ppm in many literatures (Limiroli et al., 1996; Es-Safi 266 et al., 2000), the evolution of main products belonging to this group is represented on the 267 NMR spectra in the region of 6.0–7.2 ppm, as shown in Fig. 2b. It can be clearly observed 268 that some strong proton signals occurred in the spectrum (C). According to the mass of 256 269 m/z, compounds 5, 6 and 7 would be recognized with the monosubstitution by OH in the 270 para-, ortho- and meta-position, respectively. While compounds 6 and 7 had the same 271 concentration level, compound 5 showed the higher concentration level of about 4.5 times 272 than that of compounds 6 and 7. This result indicates that compound 5 is a major 273 hydroxylated product from the DPA degradation by TiO₂/UVA due to the regioslective attack 274 of •OH. This can be explained by the electron density of the benzene carbon sites and the 275 steric hindering effects of the benzene ring substituents (-CHCON(CH₃)₂), where the para-276 position is more nucleophilic and accessible than the other two positions at ortho and meta 277 orientation. In contrast, spectrum (B) in Fig. 2b shows very low proton signals of compounds 278 5 and 6, indicating that the hydroxylation of benzene ring in DPA structure is not a 279 dominating process by direct photolysis under UVC irradiation. It should be noted that the 280 different chemical shifts of compounds 5 and 6 in both spectra (B) and (C) resulted from the 281 effect of different final pH, such as pH 6.47 in the direct photolysis but pH 5.56 in the photocatalysis. Water samples were extracted with chloroform to concentrate the organics for further analysis. As a result, it was identified that some multi-hydroxylated products (compounds 8–12, $n_{OH} = 2-5$) appeared in the TiO₂/UVA process but not in the UVC process when corroborating with their mass ion peaks in LC-MS spectra. NMR spectra of organic extracts are not presented in this paper.

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288 3.4. Degradation pathways

289 The results in Table 2 reflect a fact that there are different pathways between the DPA 290 degradations in photolytic and photocatalytic processes due to the evolution of different 291 principal intermediates. Figure 3 illustrates the proposed pathways of DPA degradation by 292 UVC irradiation alone, which may not include every possible reactions but covering the main 293 approaches confirmed by the identified products. The major decay pathways include N-294 methyl oxidation, scission of the N-CO or C-CO bond, dimerization and ring opening. The 295 results have been compared to those from previous research with analogous degradation 296 mechanisms such as tyrosine, phenylthioacetic acid and phenylurea derivatives photolysis 297 (Jin et al., 1995; Filipiak et al., 2006), which are similar to our findings. For specific 298 intermediates, compound 1, arising from N-methyl oxidation, is considered to be the major 299 primary intermediate in the process (see Fig. 2). The oxidation of terminal methyl group 300 involves an intramolecular hydrogen bond between N-methyl and carbonyl groups in the excited state (Jirkovský et al., 1997), and then reacts with dissolved molecular oxygen to 301 302 form formylated product [-N(CH₃)CHO]. This indicates a key role played by the 303 photoexcited oxygen species and the formation of the formylated product by oxidation of the 304 *N*-methyl group was the major reaction observed. The subsequent reduction and further 305 oxidation of compound 1 gave rise to form compounds 2 and 3, in which compound 3 easily 306 underwent the decarboxylation to yield compound 4 via photo-kolbe reactions (Krautler and Bard, 1978). The evolution profiles of compounds 1–4 in Fig. 2 offer useful information in supporting this pathway. On the other hand, no products that obviously come from further oxidation of the second N-methyl group were detected in our experiments. Despite hydroxylated products 5 and 6 were detected, the fact that they only existed at trace levels and implied that hydroxylation of aromatic ring was not a minor pathway in the UVC photolysis process.

313 [Fig. 3]

314

315 Direct irradiation leads to the promotion of a molecule from the ground state to an excited 316 state. Such excited states can undergo homolysis or heterolysis, among other processes. It is 317 known that a 254 nm photon has an equivalence of 4.89 eV, which should be enough 318 energetic to produce homolytic or heterolytic breakages in the molecules (Litter, 2005). 319 Therefore, it can be expected that further photolysis bond scission could take place, followed 320 by radical recommendation or H-/(HO-) abstracting from water solvent. For instance, scission 321 of the N-CO bond leads to yield the diphenylacetic acid (compound 15) while scission of the 322 C-CO bond gives benzhydrol (compound 16) as a major intermediate and a trace amount of 323 compound 13. The cleavage of compound 16 can produce small molecules such as phenol 324 (compound 17) and benzaldehyde (compound 18), where benzaldehyde can easily undergo 325 further oxidation to form secondary intermediate benzoic acid (compound 19) while phenol 326 trends to undergoing the ring opening to form product 20. The observed product 20, 327 belonging to an aliphatic polyene acid, can be successively transformed into product 21 and finally to CO₂ and H₂O. A similar pathway for phenol degradation by photolysis was 328 329 proposed by Jin et al. (1995).

330 In the heterogeneous photocatalysis process with TiO_2/UVA , hydroxylation reactions of 331 the aromatic ring are the most frequently observed primary reaction due to the attack of •OH

332 (Amine-Khodja et al., 2002). Similarly, these reactions were proposed in Fig. 4, which are 333 significantly different from that of DPA photolytic process. First, isomers 5-7 were identified 334 as the phenolic compounds monosubstituted by OH in para-, ortho- and meta-position, 335 respectively, suggesting that the aromatic ring has an electron-donating effect, due to the electrophilicity of hydroxyl radicals. It is interesting that isomer 5 shows the highest 336 337 concentration level, following by isomers 6 and 7. These results indicate that the 338 monohydroxylation of aromatic ring with corresponding H abstraction by •OH attack not 339 only is a major reaction step in the photocatalytic process but also takes place regioselectively 340 on the aromatic ring. By further attack of •OH, its subsequent reaction with oxidizing species 341 vielded the multihydroxylated intermediates (compounds 8–12), such as di-, tri-, tetra- and 342 quint-substitution phenolics. However, the low concentrations of these products reveal that 343 the multihydroxylation reaction might occur just as a minor route in the photocatalytic 344 degradation of DPA. Further oxidation by •OH attack can result in the benzene ring opening 345 to yield maleic acid (compound 21). The ring opening is possibly due to the formation of a 346 hydroxycyclohexadienyl radical in photocatalysis process (e.g. radical B in Fig. 4), confirmed 347 by Aceituno et al. (2002). Besides this, both oxidation of N-methyl groups and N-348 demethylation on the urea moiety of DPA were also proposed, as shown in Fig. 5. These 349 reactions were conducted through attack by hydroxyl radicals and reaction with oxygen. Although the compounds 1-4 were found in both photolysis and photocatalysis, their 350 351 evolution underwent different reactions by bond scission and •OH attack, respectively.

352 [Fig. 4]

353 [Fig. 5]

354

355 4. Conclusions

356 The experiments demonstrated that DPA degradation by direct photolysis with UVC

proceeded quickly, but its further mineralization was difficult, while the DPA degradation by 357 358 photocatalysis with TiO₂/UVA proceeded more slowly, but achieved a higher extent of mineralization. Using both the LC-MS and ¹H-NMR analyses together is a powerful 359 360 approach to identify more than 20 intermediates from the DPA degradation. The analytical 361 results indicate that two processes undergo some similar reactions such as the oxidation of N-362 methyl group of side chain, but also undergo some different approaches. For example, the 363 hydroxylation of aromatic ring was observed as a major way in the photocatalytic process 364 while it is a minor one in the direct photolytic process. This fundamental research about the 365 mechanism of photolytic and photocatalytic reaction provides essential knowledge with 366 potential to further develop a best process for aqueous DPA degradation in water treatment.

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- 371 Supplemental Material
- 373 Supplementary data associated with this article can be found, in the online version, at xxxxxx.374

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419 List of figure captions

- 420 Fig. 1. The full-scan mass spectra in the m/z 50-400 range for the degradation of DPA by
 421 photolysis (a) and TiO₂-photocatalysis (b).
- 422 Fig. 2. ¹H-NMR analysis of DPA degradation by UVC and TiO₂/UVA treatments: (a) 2D-
- 423 COSY spectra of aqueous DPA solutions, in which the proton signals of methine group
- 424 in DPA and compounds 1–4 were marked; (b) Relevant region of 1D ¹H-NMR spectra
- 425 of DPA and its hydroxylation products 5, 6 and 7.
- 426 Fig. 3. Proposed pathways of the DPA degradation by direct photolysis with UVC.
- 427 Fig. 4. Proposed pathways of the DPA degradation by photocatalysis with TiO₂/UVA. Here,
- R1 is p- or m-phenolic ring; R2 is 3,5-diphenolic ring; and R' includes benzene ring, R1,
 and R2.
- 430 Fig. 5. Mechanism of oxidation of N-methyl group in DPA molecules by photocatalysis

Sustan	DDA dag	ar (1	C/C = 0/	La	10^{-3} (min ⁻¹)/1	\mathbf{p}^2	DOC		
System	DPA decay (1–C/C _o , %)		$k^{a} \times 10^{-3} (\text{min}^{-1})/\text{R}^{2}$			DOC removal (%)			
	pH 3.5	7.2	9.6	рН 3.5	7.2	9.6	pH 3.5	7.2	9.6
UVC	100	100	100	32/0.999	19/0.979	24/0.998	11	8	9
UVA	_b	_	_	_	_	-	_	_	_
TiO ₂ /UVA	63	51	42	1.8/0.979	1.3/0.994	1.0/0.978	13	11	8
137 ^b - no 138 140 141 142 143 144 145 146 147	ot detected	d.		tic constant					
49									
50									

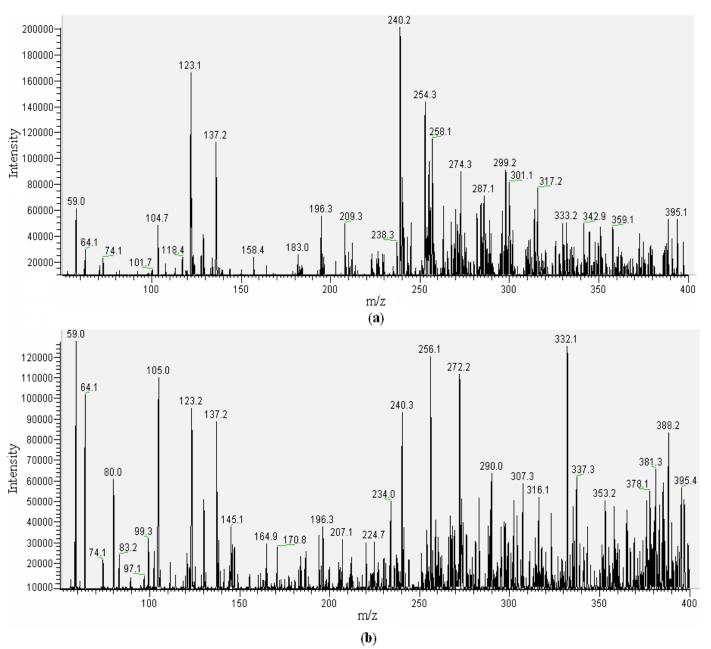
433 Table 1. Effects of solution pH on DPA degradation efficiency, kinetic constants, correlation

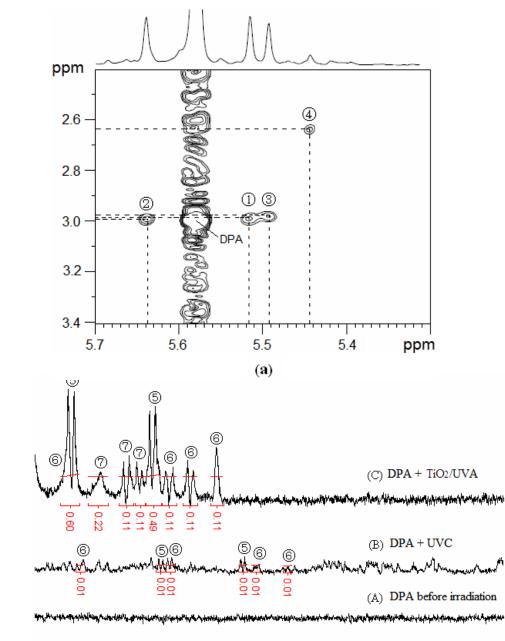
Compoun	d Structures ^(a)	MS data	¹ H-NMR data	Products of	DPA degradation ^(c)
No.		(m/z)	(δ , multiplicity ^(b) , coupling constant)	UVC	TiO ₂ /UVA
DPA co	$ \overset{b}{\longrightarrow} \overset{a}{\longrightarrow} \overset{d}{\overset{O}{\overset{H}{\longrightarrow}}} \overset{CH_3}{\underset{CH_3}{\overset{e}{\underset{H_3}{\overset{H}{\longrightarrow}}}}} $	238 (M-H) ⁻ 240 (M+H) ⁺ 241 (M+2H) ⁺⁺	a: 7.22 (4H, d, <i>J</i> _{<i>ab</i>} = 8.56); b: 7.41 (4H, m); c: 7.35 (2H, t, <i>J</i> _{<i>cb</i>} = 7.18); d: 5.58 (1H, s); e: 3.04 (3H, s); f: 3.00 (3H, s)	+	+
1 _c	$ \begin{array}{c} b & a & d & O \\ \hline & -CH - C - N \\ \hline & CH_{3} & f \\ \hline & OH \end{array} $	251 (M+O-2H) 254 (M+O+H) ⁺ 255 (M+O+2H) ⁺⁺	a: 7.22 (4H, d, <i>J</i> _{ab} = 8.56); b: 7.41 (4H, m); c: 7.35 (2H, t, <i>J</i> _{cb} = 7.18); d: 5.52 (1H, s); e: 8.44 (1H, s); f: 3.11 (3H, s)	+	+
2 _c	$e = CH - CH - C - N \begin{pmatrix} H_2 \\ CH_2 \\ CH_3 \\ F \end{pmatrix}$	255 (M+O) 256 (M+O+H) ⁺ 258 (M+O+3H) ⁺⁺⁺	a: 7.22 (4H, d, <i>J</i> _{<i>ab</i>} = 8.56); b: 7.41 (4H, m); c: 7.35 (2H, t, <i>J</i> _{<i>cb</i>} = 7.18); d: 5.64 (1H, s); e: 5.89 (2H, s); f: 3.04 (3H, s)	+	+
3 c	$ \begin{array}{c} b & a & d & 0 \\ \hline & -CH - C - N \\ \hline & CH_3 & e \end{array} $	268 (M+2O-H) ⁻ 270 (M+2O+H) ⁺ 271 (M+2O+2H) ⁺⁺	a: 7.22 (4H, d, <i>J</i> _{<i>ab</i>} = 8.56); b: 7.41 (4H, m); c: 7.35 (2H, t, <i>J</i> _{<i>cb</i>} = 7.18); d: 5.49 (1H, s); e: 3.08 (3H, s)	+	+
4 c	$ = \underbrace{ \begin{array}{c} b & a \\ \hline \\$	224 (M-CH ₂ -H) ⁻ 226 (M-CH ₂ +H) ⁺ 227 (M-CH ₂ +2H) ⁺⁺	a: 7.22 (4H, d, J_{ab} = 8.56); b: 7.41 (4H, m); c: 7.35 (2H, t, J_{cb} = 7.18); d: 5.43 (1H, s); e: 7.51 (H, m); f: 2.99 (3H, d, J_{fe} = 4.05)	+	+
5	$ \begin{array}{c} C & O \\ -CH - C - N \\ & CH_3 & e \\ & OH \end{array} $	254 (M+O-H) ⁻ 256 (M+O+H) ⁺ 257 (M+O+2H) ⁺⁺	a: 7.10 (2H, d, <i>J</i> _{ab} = 9.11); b: 6.88 (2H, d, <i>J</i> _{ba} = 9.09); c: 5.58 (1H, s); d: 3.04 (3H, s); e: 3.00 (3H,	trace	+
6	$ \underbrace{ \begin{array}{c} & & & \\$	254 (M+O-H) ⁻ 256 (M+O+H) ⁺ 257 (M+O+2H) ⁺⁺	a: 6.91 (1H, d, <i>J</i> _{<i>ab</i>} = 8.79); b: 7.01 (1H, m); c: 7.01 (1H, m); d: 6.95 (1H, d, <i>J</i> _{<i>dc</i>} = 8.58); e: 5.58 (1H, s); f: 3.04 (3H, s); g: 3.00 (3H,	trace	+
7	$ \underbrace{ \begin{array}{c} \begin{array}{c} e & O \\ -CH-C-N \\ cH-C-N \\ \end{array} \\ \begin{array}{c} CH_3 & g \\ \end{array} \\ \begin{array}{c} CH_3 & g \\ \end{array} \\ \begin{array}{c} \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \end{array} \\ \begin{array}{c} \\ \end{array} \\ $	254 (M+O-H) ⁻ 256 (M+O+H) ⁺ 257 (M+O+2H) ⁺⁺	a: 6.71 (1H, s); b: 6.78 (1H, d, $J_{bc} = 9.75$); c: 7.09 (1H, m); d: 6.84 (1H, d, $J_{dc} = 9.50$); e: 5.58 (1H, s); f: 3.04 (3H, s); g: 3.00 (3H,	trace 5)	+

Table 2. Products resulting from the direct photolysed and photocatalysed degradation of DPA by MS and ¹H-NMR analysis.

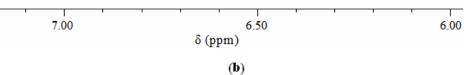
Other products 8-21 were shown in Table SM-1.

[Fig. 1]

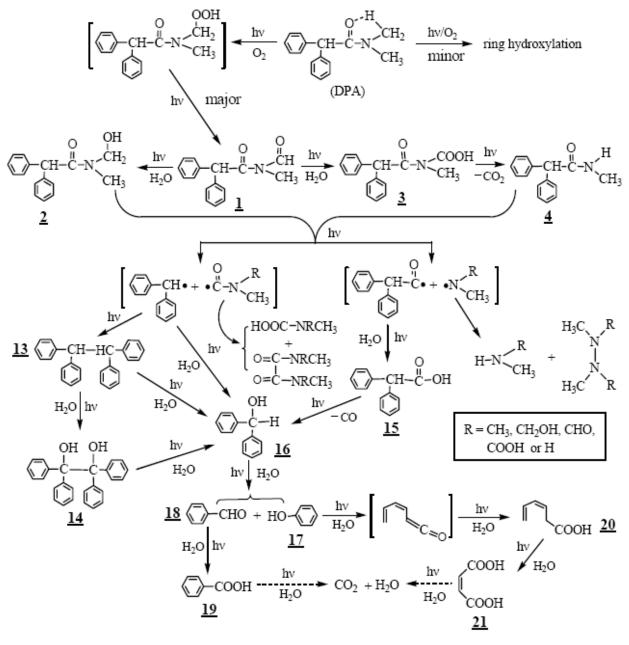




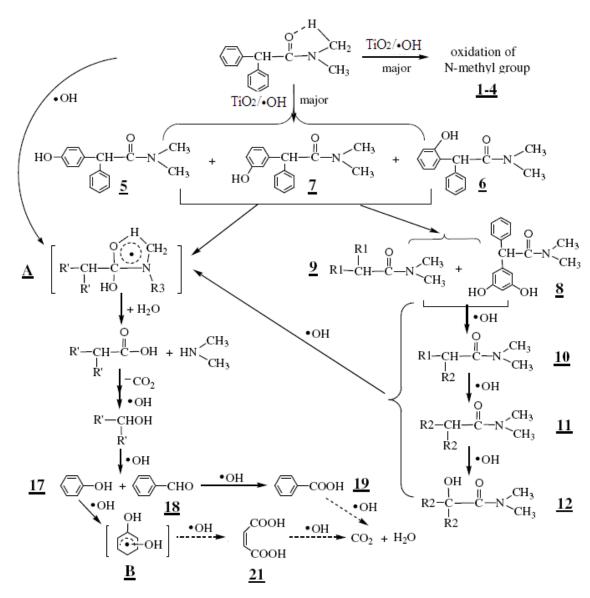
[Fig. 2]



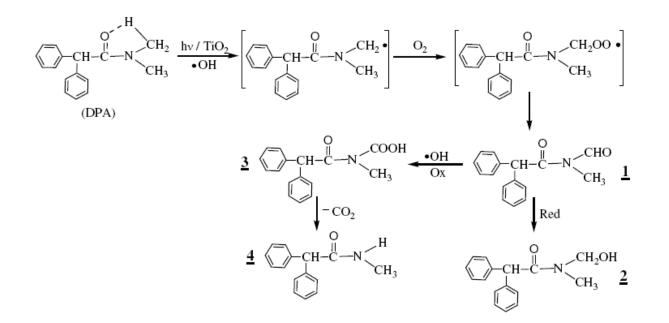
[Fig. 3]



[Fig. 4]

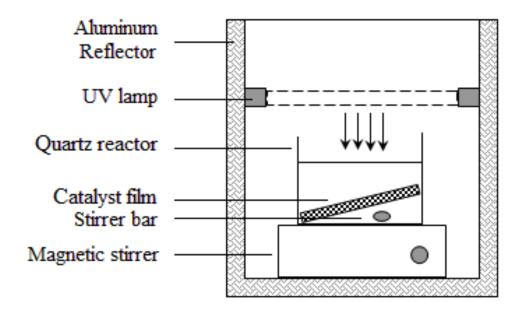


[Fig. 5]



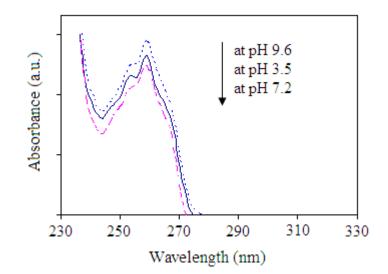
Supplemental Material:

[Fig. SM-1] Diagram of the batch-scale photocatalytic-reactor.



Photocatalytic-reactor

[Fig. SM-2] UV-visible spectra of aqueous DPA solutions at different pH media before irradiation.



(The color figure is intended to be reproduced in black-and-white.)

Compound		MS data			DPA degradation ^(c)
No.		(m/z)	$(\delta, multiplicity^{(b)}, coupling constant)$	UVC	TiO ₂ /UVA
8 ^(d)	$ \begin{array}{c} & \bigcirc & & \\ & \bigcirc & -CH - C - N \\ & & & \\ & & & \\ & & & \\ & HO \\ & & & & \\ & & & \\ & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\ & & & & \\$	272 (M+H) ⁺ 273 (M+2H) ⁺⁺ 274 (M+3H) ⁺⁺⁺	a: 6.63 (2H, s); b: 6.57 (1H, s); c: 5.58 (1H, s); d: 3.04 (3H, s); e: 3.00 (3H, s)	-	trace
9 ^(d)	$ \overset{OH}{\overset{\circ}{\underset{HO}{\overset{\bullet}{\underset{HO}{\overset{\bullet}{\underset{HO}{\atopHO}{\overset{\circ}{\underset{HO}{\overset{\circ}{\atopH}{\underset{HO}{\atopHO}{\atopHO}{\atopH}}{\overset{\circ}{\underset{HO}{\atopHO}{\atopHO}{I}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}}$	$\begin{array}{c} 272~{\rm (M+H)}^{+}\\ 273~{\rm (M+2H)}^{++}\\ 274~{\rm (M+3H)}^{+++}\end{array}$	a: 5.58 (1H, s); b: 3.04 (3H, s); c: 3.00 (3H, s)	_	trace
10 ^(d) HO	$ \begin{array}{c} c \\ CH-C-N \\ cH-C-N \\ CH_3 \\ CH_3 \\ e \\ HO \\ b \\ OH \end{array} $	285 (M-3H) 288 (M+H) ⁺ 290 (M+3H) ⁺⁺⁺	a: 6.63 (2H, s); b: 6.57 (1H, s); c: 5.58 (1H, s); d: 3.04 (3H, s); e: 3.00 (3H, s)	_	trace
H0 11 ^(d) H0	\square CH- \square N \square CH ₃ \square	300 (M-3H) 303 (M) 304 (M+H) ⁺ 306 (M+3H) ⁺⁺⁺	a: 6.63 (2H, s); b: 6.57 (1H, s); c: 5.58 (1H, s); d: 3.04 (3H, s); e: 3.00 (3H, s)	_	trace
12 ^(d)	$\begin{array}{c} OH & O\\ CH_{3} & C\\ HO & a & C\\ HO & b & OH \end{array} \xrightarrow{CH_{3} & c} CH_{3} & d\\ CH_{3} & d\\ CH_{3} & d & C\\ CH_{3} & C\\$	316 (M-3H) 318 (M-H) ⁻ 322 (M+3H) ⁺⁺⁺ 323 (M+4H) ⁺⁺⁺⁺	a: 6.63 (2H, s); b: 6.57 (1H, s); c: 5.58 (1H, s); d: 3.04 (3H, s);	_	trace
13 ^(d)	c c CH-CH-C-C	332 (M-2H) 333 (M-H) ⁻ 336 (M+2H) ⁺⁺	a: 7. 51(4H, d, <i>J</i> _{ab} = 8.22); b: 7.53 (4H c: 7.45 (2H, t, <i>J</i> _{cb} = 8.14); d: 4.89 (1H,		_
14 ^(d)	a a OH OH a C C C C	365 (M-H) 366 (M) 367 (M+H) ⁺	a: 7. 49 (5H,s)	+	_

Table SM-1. Products 8-12 resulting from the direct photolysed and TiO₂-photocatalysed degradation of DPA by MS and ¹H-NMR analysis.

Table SM-1 (Continue)							
Compound	Structures ^(a)	MS data	¹ H-NMR data		DPA degradation ^(c)		
No.		(m/z)	(δ , multiplicity ^(b) , coupling constant)	UVC	TiO ₂ /UVA		
15	c	211 (M-H) ⁻ 212 (M) 213 (M+H) ⁺	a: 7. 22 (4H, d, J_{ab} = 8.56); b: 7.41 (4H, m); c: 7.35 (2H, t, J_{cb} = 7.18); d: 5.12 (1H, s);	+	+		
16	c ∕ → − CH−OH	183 (M-H) ⁻ 184 (M) 185 (M+H) ⁺	a: 7.39 (4H, d, <i>J</i> _{<i>ab</i>} = 8.02); b: 7.51 (4H, m); c: 7.47 (2H, t, <i>J</i> _{<i>cb</i>} = 8.81); d: 5.89 (1H, s);	+	+		
17 ^(d)	c ∕ → OH	93 (M-H) [−] 94 (M) 95 (M+H) ⁺	a: 6.85 (2H, d, <i>J</i> _{<i>ab</i>} = 6.54); b: 7.25 (2H, m); c: 6.99 (1H, t, <i>J</i> _{<i>cb</i>} = 7.55)	trace	trace		
18	$c \xrightarrow{b a o}_{CH d}$	105 (M-H) ⁻	a: 7.89 (2H, d, <i>J</i> _{<i>ab</i>} = 9.24); b: 7.51 (2H, m); c: 7.74 (1H, t, <i>J</i> _{<i>cb</i>} = 8.31); d: 9.34 (1H, s);	+	+		
19	c ∕ ⊂ −OH	121 (M-H) ⁻ 122 (M) 123 (M+H) ⁺	a: 8.16 (2H, d, <i>J</i> _{<i>ab</i>} = 9.20); b: 7.67 (2H, m); c: 7.49 (1H, t, <i>J</i> _{<i>cb</i>} = 8.12);	+	+		
20 ^(d)	a COOH	111 (M-H) ⁻ 112 (M) 113 (M+2H) ⁺⁺	a: 5.83 (2H, d, <i>J</i> _{<i>ab</i>} = 8.91); b: 6.15 (1H, m); c: 6.44 (1H, m); d: 5.97 (1H, m); e: 3.38 (2H, d, <i>J</i> _{<i>ed</i>} = 3.85);	+	-		
21 ^(d)	аСООН	115 (M-H) ⁻ 118 (M+2H) ⁺⁺	a: 6.36 (2H, s)	_	+		

(a) Different protons linked on molecular structures are marked as a, b, c, d, e and f, respectively.
 (b) Singlet, doublet, triplet, and multilet are abbreviated as s, d, t and m, respectively; coupling constants (*J*) are given hertz.
 (c) -, absence; +, presence.
 (d) Identified in organic extracts only. The NMR spectra of organic extracts were not showed in the present paper.