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Title: Integral valorization of *Leucaena diversifolia* by hydrothermal and pulp processing

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Keywords: *Leucaena diversifolia*; Hydrothermal treatment; Autohydrolysis; Gross Heating Value; Ethanol-soda-anthraquinone delignification

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Abstract: Wood from the leguminous tree, *Leucaena diversifolia*, was subjected to hydrothermal treatment (autohydrolysis) at 160 to 180 °C and 0 to 30 min. followed by ethanol-soda-anthraquinone delignification. The liquid phase contained 18.65 g of sugars per liter, and the solid phase had a gross heating value of 19.083 MJ/kg, but could also be uses as a source of cellulose pulp for the production of paper with tear, burst and tensile indexes of 2.4 Nm²/kg, 2.6 MPam²/kg and 40.7 KNm/kg respectively. *Leucaena diversifolia* lends itself readily to valorization for energy production, and also to integral, fractional exploitation by autohydrolysis and ethanol-soda-anthraquinone delignification, which can additionally bring environmental benefits to cropping zones.

Reviewers' comments:

Please make the following modifications to shorten the manuscript and to remove wording and typographical errors:

1. Modify title: Integral valorization of *Leucaena diversifolia* by hydrothermal and pulp processing.

The title has been changed.

2. The current abstract should be condensed:

Wood?? from the leguminous tree, *Leucaena diversifolia*, was subjected to hydrothermal treatment (autohydrolysis) at ...[what conditions?] followed by ethanol-soda-anthraquinone delignification. The liquid phase contained 18.65 g of sugars per liter, and the solid phase had a gross heating value of 19.083 MJ/kg, but could also be used as a source of cellulose pulp for the production of paper with tear, burst and tensile indexes of, respectively. [Add concluding sentence as to the importance/utility of your study.]

The abstract has been modified and summarized

3. Shorten Introduction:

Leucaena diversifolia is a leguminous tree that can grow 6-20 m tall and is adapted to Mediterranean conditions. The tree produces large amounts of biomass around 70 tons/ha/year (ref?) and possesses a high re-sprouting ability. It fixes nitrogen in symbiosis with *Rhizobium* sp. and thus improves soil fertility and has been exploited for soil conservation and consolidation (Goel and Behl, 2002). Moreover, *Leucaena* produces hard and strong wood and high-quality forage. Lignocellulosic biomass such as that obtained from *L. diversifolia* can be converted to useful fractions through "biorefining" (Clark et al., 2011). One refining strategy involves treatments that hydrolyze some of the polysaccharides within the biomass. Hydrothermal treatment causes the total or partial dissolution of hemicelluloses as the hydrolysis of acetyl groups gives rise to acetic acid which in turn catalyzes the hydrolysis of the polysaccharide (reference?) leading to the release of sugar oligomers, xylose and arabinose and other chemicals. These compounds can be used for a variety of applications in the chemical, pharmaceutical and food industries, and also as fermentation media for the production of substances such as ethanol fuel and xylitol (Rivas et al., 2002). The solid fraction from the hydrothermal process contains lignin and cellulose and can be employed as raw material for alcohol production, ruminant feed, raw material for pulp and paper making and for energy production. Removal of lignin from the solid fraction provides cellulose pulp for paper making. "Green" pulping methods involving no sulphur (e.g. the ethanol-soda-anthraquinone process) are now capable of providing pulp in a high yield, with low residual lignin, high brightness and good strength properties (Shatalov and Pereira, 2004). In the current study, a variety of *Leucaena diversifolia* that ...[what is the improvement?] was subjected to integral fractionation by autohydrolysis and by delignification. The liquor from the autohydrolysis treatment was evaluated as a source of oligomers and monosaccharides, and the solid phase was assessed as raw material for production of combustion energy and the manufacturing of cellulose pulp and paper.

The Introduction has been modified and summarized. Several references have been added.

4. 2.1. Source and analysis of biomass

Leucaena diversifolia biomass was obtained from trees grown for seven years on a plantation in Huelva (southwestern Spain). The plants were grown in a nursery in pots, inoculated with *Rhizobium*, and planted at 3 month of age in two plots (plants were placed at 1.8 x 0.6 m -10.800 plants/ha-) with a complete randomized block design with 4 replicates. No fertilizer was added to the plots which were made up of sandy loam with a pH of 6-8 and moderate to substantial depth. *Leucaena diversifolia* sample??? [what specifically? Trunks, branches, leaves, everything?] were milled to pass a 8-cm screen. The chips were reduced again {how?} to pieces 2 to 10 mm long, and fines were removed by sieving through a 0.6-mm mesh. Samples were air-dried, mixed and stored [how?]liquid chromatography (HPLC). Samples ["aliquots" apply to liquids only] with a particle size of < 0.5 mm were subjected to moisture, extractives determination (TAPPI T 264-cm-07) and to quantitative acid hydrolysis with 72% H₂SO₄ at 121 °C and 2 atm for 60 min (TAPPI T 249 cm-09). The solid residue after hydrolysis was recovered by filtration through a 0.45 μm filter and considered as Klason lignin.order to estimate (after corrections for stoichiometry and sugar decomposition) the cellulose (as glucan), hemicelluloses (as xylan), and acetyl group contents (Díaz et al., 2004).

Paragraphs have been changed and included the following phrases:

"*Leucaena diversifolia* wood samples (trunks and branches) were milled"

"The chips were reduced again in a mill laboratory"

"and stored in hermetic plastic bag."

5. 2.2. Biorefining process

Figure 1 shows the flow of processes carried out with the *L. diversifolia* samples. A hydrothermal treatment (autohydrolysis) was carried out in order to obtain a solid phase that was delignified with an ethanol-soda-anthraquinone process to obtain cellulose pulp and a liquid phase with hemicellulosics sugars.

The paragraph has been included.

6. 2.3.1. Operation conditions in hydrothermal treatment

Raw material and water were mixed at a liquid/solid ratio of 8 kg water per kg raw material on a dry basis (the moisture content of the material was considered to be water) and treated in a 2-L stainless steel reactor (Parr Instruments Company, Moline, IL). The reactor was fitted with four blade turbine impellers, heated by An aliquot of the liquor was filtered through 0.45 μm membranes and A second aliquot (25 mL) was subjected to quantitative post-hydrolysis with 4 % H₂SO₄ at 121 °C for 60 min before HPLC analysis. The calorific value of the autohydrolysis solid phase was determined in section 2.1.

Paragraphs have been changed.

7. 2.3.2. Experimental desing for hydrothermal treatment

.... This allowed relating the dependent (yield, soluble lignin, calorific value and glucose, xylose, arabinose, oligomers and acetyl group contents) and independent (temperature and time of process) variables of the autohydrolysis process with a minimum number of experiment. Based on these previous studies with *Leucaena diversifolia* and others lignocellulosic materials, operation conditions were selected (Alfaro et al., 2010, Garrote et al., 2003, Alfaro et al.,

2009). The autohydrolysis temperature and autohydrolysis time were 160, 172 and 184°C, and 0, 15 and 30 min (at operation temperature), respectively. The liquid/solid ratio was 8/1 in all experiments.....

Paragraphs have been changed.

8. 2.4. Ethanol-soda-anthraquinone pulping process

The operation conditions for pulping without prior autohydrolysis were: Chips with a length of 3-6 cm and a thickness of 1 cm, soda concentration in the cooking liquor 17% by weight, ethanol concentration 45% in volume, anthraquinone concentration 0.10% in weight, initial liquor/solid ratio of 8/1 (dry wt. basis), operation time 60 min and operation temperature 185°C. For samples obtained after autohydrolysis, a soda concentration of 13% and a operation temperature of 170 °C were used. Both pulps were obtained using a 8-L bath cylindrical reactor that was heated by means of electrical resistances and linked to a control unit including the required instrument for measurement and control of the pressure and temperature. Preheating was done for 25-30 min to reach the operation temperature. After completion of the treatment, the reactor was cooled, and the pulp was separated from the liquor and washed in a bag of 0.1 mm mesh. The pulp was defibered on a Sprout-Waldron refiner and passed again through a strainer (0.4mm mesh) in order to isolate the uncooked material (<0.5%).

Paragraphs have been changed.

9. 2.5. Beating process, formation and characterization of paper sheets

The resulting pulps were processed on a Valley beater, according to TAPPI T-200 sp-06, at different process times in order to obtain various drainage levels. The degree of refining was determined in a Shopper-Riegler refinometer (°SR) according to TAPPI T- 227 om-09 Freeness of Pulp. Paper sheets were obtained using an ENJO-F-39.71 sheet former according to TAPPI T 205 sp-06.....The deviations for these parameters from their respective means were less than 5%.

Paragraphs have been changed.

10. 3.1. Chemical and energetic characteristics of raw material

Table 1 shows the chemical and energetic characteristics of *Leucaena* and other lignocellulosic materials. The *Leucaena*-derived material consisted of 32.2% cellulose (analyzed as glucan), (40.2% as β -cellulose), 18.6% hemicelluloses, calculated as the sum of xylan, arabinan and acetyl groups, (32.6% based on holocellulose - β -cellulose), and 21.5% Klason lignin (23.7% after quantitative acid hydrolysis). This composition is similar to that found by other authors for *Leucaena diversifolia* (refs) and other varieties of *Leucaena* (refs) and comparable to hardwoods, such as *Eucalyptus globulus* (refs). The hemicellulose content in *Leucaena* was similar to that of tagasaste (41.4%) (ref.) and herbaceous species (Alfaro et al., 2010, Garrote et al., 2003, Alfaro et al., 2009). The molar ratio of the monomers in *L. diversifolia* was calculated as xylose:acetyl groups:arabinose = 15.5:2.1:1. This composition is typical of acetylglucuronoxylans typically present in hardwoods (Pinto et al., 2005). The gross heating value (GHV) of *L. diversifolia* of 19.1 MJ/kg, is in between those for softwood (20.0 MJ/kg) and hardwood (18.8 MJ/kg) (ref?), similar to that of *Ailanthus* wood (19.0 MJ/kg) (ref?), somewhat higher than that of *Eucalyptus globulus* (Telmo et al., 2010), and much higher than those of wheat straw

(17.0 MJ/kg) or corn stover (17.8 MJ/kg) (Demirbas, 1997). The GHV for the solid residue remaining after *Leucaena* autohydrolysis is shown in Table 2 together with the contents of the autohydrolysis solid phase. The GHV for the solid phase ranged from 19.1 to 19.9 MJ/kg depending on the autohydrolysis conditions. Table 4 shows This further increases the intrinsic value of the compounds extracted into the liquor or reduces the energy cost associated with extraction by autohydrolysis.

Paragraphs have been changed and several references have been added.

11. 3.2. Products of hydrothermal treatment. Modeling and optimization.

Table 2 shows the normalized values of independent variables (temperature and operation time) and the Gross Heating Values, yield, Klason lignin and soluble lignin contents in the solid phase relative to the initial raw material, and also the glucan, xylan, araban contents and acetyl groups in the solid phase, relative to the initial respective polymer content in raw material. Table 3 [it seems Table 4 was mentioned before Table 3] shows the monomeric sugar and oligomer concentrations in the liquid phase in g/L and as percentage relative to respective polymer content in dry raw material. The results were modeled by using the multiple regression methodology described in section ..(Table 4). temperature on yield was considerably smaller at short treatment times (yields of 90-96%) relative to long times (yields of 72-90%).The amount of soluble lignin was, in most cases, lower (2.1% in the raw material and 0.82-2.03% in the autohydrolysis solid fraction). This result suggests substantial dissolution of this fraction during autohydrolysis (Leschinnsky et al., 2009). In a biorefinery process that includes paper production, the amount of sugars in solution should be as high as possible, but degradation of cellulose (as glucan) in the raw material should be avoided as far as possible in order to obtain pulp of acceptable properties (Alfaro et al., 2010). In order to determine the values of the independent variables giving the optimum variable dependent results, the response surfaces for each dependent variable were plotted at maxima and minima levels of the independent variables (Figures 2 and 3). Figure 2 shows the three response surfaces for the glucan derivatives in both autohydrolysis phases. The influence of the variables was essentially linear despite Modeling the results for this hemicellulose fraction was also complicated.response surfaces of Figure 3. The hydrolysis of hemicellulosic xylan in the raw material was substantial over the operational range 0-1 for both operational variables. Based on the results presented in Table 2, xylan was [The following sentence is not clear: In principle, it is advisable to use medium or high temperatures and short or medium treatment times to extract xylose and xylooligomers similarly efficiently as with medium temperatures and long times, albeit with less efficient degradation of glucan (roughly 2 percent points more of glucan in the autohydrolysis solid phase based on the YGLS model of Table 4).] Autohydrolysis extracted araban to a similar extent as xylan derivatives, but time was more influential than temperature on its hydrolysis (see YARS equation in Table 4). 91.6 to 103.7% (data not shown). In regards to acetyl groups, the As a rule, extraction of hemicellulosic derivatives from *L. diversifolia* was more efficient than that from other batches of the same plant grown for 3 years and subjected to non-isothermal treatments (Alfaro et al., 2009); however, it was less efficient than that from *Chamaecytisus proliferus* (Alfaro et al., 2010), another legume. The glucan fraction in *Leucaena* was less markedly Applying a similar thermal treatment to *Paulownia fortunei*, a tree species of similar composition as *Leucaena*

Paragraphs have been changed.

The phrase "In principle, it is advisable to use medium or high temperatures and short or medium treatment times to extract xylose and xylooligomers similarly efficiently as with medium temperatures and long times" has been changed by the sentence "In principle, it is advisable to use medium or high temperatures and short or medium treatment times to extract xylose and xylooligomers similarly as with medium temperatures and long times".

The phrase "albeit with less efficient degradation of glucan" has been changed by the sentence "Also there is less degradation of glucan".

12. 3.3. Characterization of paper sheets from solid phase post-autohydrolysis
Cellulose pulp was obtained from the autohydrolysis solid phase provided by a thermal treatment at 178 °C for 0 min (non-isothermal conditions) [what does this mean? What was the temperature?], and the properties of paper prepared from this pulp were: Kappa number 40.4, viscosity 1119 g cm⁻³ and brightness 29.3 %. The conditions of the hydrothermal treatment preceding delignification were adjusted in such a way as to ensure a high content in cellulose (as glucan) in the solid phase and in potentially valorizable hemicellulosic sugars in the liquid phase. Based on the equations of Table 4, the resulting liquid phase should contain 2.4% xylose and 26.4% xylo-oligomers respect to the initial concentration of xylan in the raw material, whereas the solid phase should consist of 92.2% glucan. The yield of the solid phase would be 81%, its gross heating value 19.6 MJ/kg and the soluble lignin content in the solid phase 1.2% (i.e. 43% of all initial lignin would be dissolved by the autohydrolysis treatment).autohydrolysis solid phase. The Kappa number, viscosity, and brightness of this paper were 27, 1303 g cm⁻³, and 38.5%, respectively. the major strength-related properties ofThe data presented in Figure 4 allows the following conclusions:improved strength-related properties. Similar results were obtained in previous studies on another legume variety, *Chamaecytisus proliferus*, subjectedmarkedly improved by the autohydrolysis treatment; however, there were also substantial ...

All paragraphs have been changed.

13. Table 1 is too confusing and should be separated into two Tables.

Table 1 has been divided into two tables 1a and 1b.

14. Figure 1: Scheme of experimental work. Abbreviations: [soda] w: soda concentration by weight; [ethanol] v: concentration of ethanol by volume; [AQ] w: concentration of anthraquinone by weight; T: process temperature (°C); t: processing time (min). L/S: liquid / solid ratio"

The abbreviations in Figure 1 have been modified.

15. Figure 2: Glucose and gluco-oligomers of the liquor and glucan of the solid as function of temperature and time of the autohydrolysis process

The title of Figure 2 has been changed.

16. Figure 3: Xylose and xylo-oligomers of the liquor and xylan of the solid as function of temperature and time of the autohydrolysis process

The title of Figure 3 has been changed.

17. Figure 4: Properties of the paper sheets vs. Shopper-Riegler degree.[Are the data points averages?]

The title of Figure 4 has been changed.

Leucaena diversifolia lends itself readily to valorization for energy production, and also to integral, fractional exploitation by autohydrolysis and ethanol–soda–anthraquinone delignification, which can additionally bring environmental benefits to cropping zones.

The autohydrolysis solid phase can be used to obtain other chemicals, directly burnt for energy production or processed to obtain cellulose pulp. The liquid phase is rich in monomers and oligomers.

The autohydrolysis process facilitates refining of cellulose pulp allowing a significant improvement in resistance rates of the paper sheets.

1 **Integral valorization of *Leucaena diversifolia* by hydrothermal and pulp processing**

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19
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21 **Abstract**

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24 **Wood from the leguminous tree, *Leucaena diversifolia*, was subjected to**
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26
27 **hydrothermal treatment (autohydrolysis) at 160 to 180 °C and 0 to 30 min.**
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29 **followed by ethanol-soda-anthraquinone delignification. The liquid phase**
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31 **contained 18.65 g of sugars per liter, and the solid phase had a gross heating value**
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33 **of 19.083 MJ/kg, but could also be uses as a source of cellulose pulp for the**
34
35 **production of paper with tear, burst and tensile indexes of 2.4 Nm²/kg, 2.6**
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37 **MPam²/kg and 40.7 KNm/kg respectively. *Leucaena diversifolia* lends itself readily**
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39 **to valorization for energy production, and also to integral, fractional exploitation**
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41 **by autohydrolysis and ethanol–soda–anthraquinone delignification, which can**
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43 **additionally bring environmental benefits to cropping zones.**
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51 **Keywords**

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55 *Leucaena diversifolia*, Hydrothermal treatment, Autohydrolysis, Gross Heating
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57 Value, Ethanol-soda-anthraquinone delignification
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1. Introduction

Environmental prevention plans for industrial processes include provisions for efficient use of resources and raw materials to maximize yields while minimizing waste production and the consumption of energy –which should ideally come from renewable or non-fossil sources. Lignocellulosic biomass is a major renewable source of raw materials and energy integral exploitation of which can provide a number of environmental and economic advantages (Heinimö and Junginger, 2009).

Plant species used to extract energy from industrial crops should be ecologically integrated in a sound manner in order to avoid adverse impacts on the environment and competition between food crops. Using arable land to grow non-food crops can help reduce agricultural surpluses, develop rural economies, reduce aid-dependence and decrease the production of greenhouse gases (Gustavsson et al., 2007).

***Leucaena diversifolia* is a leguminous tree that can grow 6-20 m tall and is adapted to Mediterranean conditions. The tree produces large around 40 tons/ha/year of biomass after three years of growth (Diaz et al., 2007) and 70 tons/ha/year after seven years of growth (this study) and possesses a high re-sprouting ability. It fixes nitrogen in symbiosis with *Rhizobium* sp. and thus improves soil fertility and has been exploited for soil conservation and consolidation (Goel and Behl, 2002). Moreover, *Leucaena* produces hard and strong wood and high-quality forage.**

Lignocellulosic biomass such as that obtained from *L. diversifolia* can be converted to useful fractions through "biorefining" (Clark et al., 2011). One refining strategy involves treatments that hydrolyze some of the polysaccharides within the biomass. Hydrothermal treatment causes the total or partial dissolution

1 of hemicelluloses as the hydrolysis of acetyl groups gives rise to acetic acid which in
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4 turn catalyzes the hydrolysis of the polysaccharide (Garcia et al., 2011)) leading to
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6 the release of sugar oligomers, xylose and arabinose and other chemicals. These
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8 compounds can be used for a variety of applications in the chemical,
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10 pharmaceutical and food industries, and also as fermentation media for the
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12 production of substances such as ethanol fuel and xylitol (Rivas et al., 2002). The
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14 solid fraction from the hydrothermal process contains lignin and cellulose and can
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16 be employed as raw material for alcohol production, ruminant feed, raw material
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18 for pulp and paper making and for energy production.
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23 Removal of lignin from the solid fraction provides cellulose pulp for paper
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25 making. "Green" pulping methods involving no sulphur (e.g. the ethanol-soda-
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27 anthraquinone process) are now capable of providing pulp in a high yield, with low
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29 residual lignin, high brightness and good strength properties (Shatalov and
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31 Pereira, 2004).
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35 In the current study, a variety of *Leucaena diversifolia* (selected greenhouse)
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37 was subjected to integral fractionation by autohydrolysis and by delignification.
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39 The liquor from the autohydrolysis treatment was evaluated as a source of
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41 oligomers and monosaccharides, and the solid phase was assessed as raw material
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43 for production of combustion energy and the manufacturing of cellulose pulp and
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45 paper.
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49 50 51 2. Materials and methods

52 53 54 2.1. Source and analysis of biomass

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57 *Leucaena diversifolia* biomass was obtained from trees grown for seven years
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59 on a plantation in Huelva (southwestern Spain). The plants were grown in a
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nursery in pots, inoculated with Rhizobium, and planted at 3 month of age in two plots (plants were placed at 1.8 x 0.6 m -10.800 plants/ha-) with a complete randomized block design with 4 replicates. No fertilizer was added to the plots which were made up of sandy loam with a pH of 6-8 and moderate to substantial depth.

Leucaena diversifolia wood samples (trunks and branches) were milled to pass a 8-cm screen. The chips were reduced again in a mill laboratory to pieces 2 to 10 mm long, and fines were removed by sieving through a 0.6-mm mesh. Samples were air-dried, mixed and stored in hermetic plastic bag.

The raw material was prepared in accordance with TAPPI T-257 and analyzed for the following parameters: 1% NaOH solubles (TAPPI T 212 om-07), hot water solubles (TAPPI T 207 cm-93), ethanol-benzene extractives (TAPPI T 204 cm-07), Klason lignin (TAPPI T-222 om-98), holocellulose (Wise et al., 1946), α -cellulose (TAPPI T 203 cm-09) and ash (TAPPI T-211) contents. All treatments in this study were in a completely randomized design with four replications (variation coefficient less than 3% and less than 1% for holocellulose and cellulose contents).

In addition, the cellulose and hemicellulose composition of the material, in the form of glucan, xylan, araban and acetyl groups, were determined by high performance liquid chromatography (HPLC). **Samples ("aliquots" apply to liquids only) with a particle size of < 0.5 mm were subjected to moisture, extractives determination (TAPPI T 264-cm-07) and to quantitative acid hydrolysis with 72% H₂SO₄ at 121 °C and 2 atm for 60 min (TAPPI T 249 cm-09). The solid residue after hydrolysis was recovered by filtration through a 0.45 μ m filter and considered as Klason lignin.** Acid soluble lignin was determined following standard method TAPPI T 250 wd-96.

1 The monosaccharides (glucose, xylose and arabinose) and acetic acid contained in
2
3 hydrolysates were determined by HPLC in order to estimate **(after corrections for**
4
5 **stoichiometry and sugar decomposition) the cellulose (as glucan), hemicelluloses**
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7 **(as xylan), and acetyl group contents (Díaz et al., 2004)**. Chromatographic runs were
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9 done with an Agilent 1100 HPLC instrument equipped with an Aminex HPX-87H
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11 column packed with ion-exchange resin under the following conditions: mobile phase,
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13 sulphuric acid at 0.05 mol·L⁻¹; flow rate, 0.6 mL·min⁻¹; column temperature, 30°C;
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15 injected volume, 20 µL. Ashes were determined by calcination (TAPPI T 244-cm-99).
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21 All HPLC determinations were in a completely randomized design with three
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23 replications (variation coefficient less than 4%).
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26 Gross Heating Values of raw material were determined at a constant volume in
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28 accordance with “CEN/TS 14918:2005 (E) Solid biofuels-Method for the determination
29
30 of calorific value” and UNE 164001 EX standards by using a Parr 6300 Automatic
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32 Isoperibol Calorimeter.
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36 **2.2. Biorefining process**

37 **Figure 1 shows the flow of processes carried out with the *L. diversifolia* samples. A**
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39 **hydrothermal treatment (autohydrolysis) was carried out in order to obtain a solid**
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41 **phase that was delignified with an ethanol-soda-anthraquinone process to obtain**
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43 **cellulose pulp and a liquid phase with hemicellulosics sugars.**
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50 **2.3. Hydrothermal treatment**

51 **2.3.1. Operation conditions in hydrothermal treatment**

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54 **Raw material and water were mixed at a liquid/solid ratio of 8 kg water per**
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56 **kg raw material on a dry basis (the moisture content of the material was**
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1 **considered to be water) and treated in a 2-L stainless steel reactor (Parr**
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3 **Instruments Company, Moline, IL). The reactor was fitted with four blade turbine**
4 **impellers, heated by** an external fabric mantle, and cooled by cool water circulating
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6 through an internal loop. The reaction media was stirred at 150 rpm and heated to reach
7
8 the desired temperature. Time zero was considered the beginning of the isothermal
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10 stage. The temperature of the process was automatically controlled via an internal
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12 cooling coil which was used to cool the reactor after the temperature and operating time
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14 were reached.
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21 At the end of treatment, the solid residue was recovered by filtration and washed
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23 with distilled water for gravimetric yield determination. A fraction was used for pulping
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25 and another fraction was air-dried and milled to a particle size <0.5 mm. Milled samples
26
27 were assayed for lignin (Klason and soluble), glucan, hemicellulose (xylan and araban)
28
29 and acetyl groups using the same methods as for raw material analysis.
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33 **An aliquot of the liquor was filtered through 0.45 µm membranes and used**
34
35 for direct HPLC determination of monosaccharides and acetic acid. **A second aliquot**
36 **(25 mL) was subjected to quantitative post-hydrolysis with 4 % H₂SO₄ at 121 °C**
37 **for 60 min before HPLC analysis. The calorific value of the autohydrolysis solid**
38 **phase was determined in section 2.1.**
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46 *2.3.2. Experimental design for hydrothermal treatment*

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50 A 2ⁿ central composite experimental design that enabled the construction of
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52 second-order polynomial in the independent variables and the identification of statistical
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54 significance in the variables was used. **This allowed relating the dependent (yield,**
55 **soluble lignin, calorific value and glucose, xylose, arabinose, oligomers and acetyl**
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group contents) and independent (temperature and time of process) variables of the autohydrolysis process with a minimum number of experiment.

Independent variables were normalized by using the following equation:

$$X_n = \frac{X - \bar{X}}{(X_{\max} - X_{\min}) / 2}$$

Where X is the absolute value of the independent variable concern, \bar{X} is the average value of the variable, and X_{\max} and X_{\min} are its maximum and minimum values, respectively.

Several experiences from isothermal autohydrolysis were carried out in order to study the effect the severity of the process on the xylooligomer production and cellulose degradation. **Based on these previous studies with *Leucaena diversifolia* and others lignocellulosic materials, operation conditions were selected (Alfaro et al., 2010, Garrote et al., 2003, Alfaro et al., 2009). The autohydrolysis temperature and autohydrolysis time were 160, 172 and 184°C, and 0, 15 and 30 min (at operation temperature), respectively. The liquid/solid ratio was 8/1 in all experiments.**

The number of tests required was calculated as $N = 2^n + 2 \cdot n + n_c$, 2^n being the number of points constituting the factor design, $2n$ that of axial points and n_c that of central points. Under our conditions, $N = 10$.

The experimental results were fitted to the following second-order polynomial:

$$Y = a_o + \sum_{i=1}^n b_i X_{ni} + \sum_{i=1}^n c_i X_{ni}^2 + \sum_{i=1; j=1}^n d_i X_{ni} X_{nj} \quad (i < j)$$

The independent variables used in the equations relating to both types of variables were those having a statistical significant coefficient (**those not exceeding a**

1 significance level of 0.05 in the student 's-test – $t < 2$ - and having a 95%
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3 confidence interval excluding zero)
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5 6 7 *2.4. Ethanol-soda-anthraquinone pulping process* 8 9

10 **The operation conditions for pulping without prior autohydrolysis were:**
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12 **Chips with a length of 3-6 cm and a thickness of 1 cm, soda concentration in the**
13 **cooking liquor 17% by weight, ethanol concentration 45% in volume,**
14 **anthraquinone concentration 0.10% in weight, initial liquor/solid ratio of 8/1 (dry**
15 **wt. basis), operation time 60 min and operation temperature 185°C. For samples**
16 **obtained after autohydrolysis, a soda concentration of 13% and a operation**
17 **temperature of 170 °C were used.** The values of rest of variables were the same. These
18 values of process variables were selected according previous works from the authors
19 (Alfaro et al., 2009). Anthraquinone enhances strength-related properties relative to
20 soda–ethanol paper.
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36 **Both pulps were obtained using a 8-L bath cylindrical reactor that was**
37 **heated by means of electrical resistances and linked to a control unit including the**
38 **required instrument for measurement and control of the pressure and**
39 **temperature. Preheating was done for 25-30 min to reach the operation**
40 **temperature. After completion of the treatment, the reactor was cooled, and the**
41 **pulp was separated from the liquor and washed in a bag of 0.1 mm mesh. The pulp**
42 **was defibered on a Sprout-Waldron refiner and passed again thought a strainer**
43 **(0.4mm mesh) in order to isolate the uncooked material (<0.5%).**
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2.5. Beating process, formation and characterization of paper sheets

The resulting pulps were processed on a Valley beater, according to TAPPI T-200 sp-06, at different process times in order to obtain various drainage levels. The degree of refining was determined in a Shopper-Riegler refinometer (°SR) according to TAPPI T- 227 om-09 Freeness of Pulp. Paper sheets were obtained using an ENJO-F-39.71 sheet former according to TAPPI T 205 sp-06.

The resulting paper sheets were analysed for Tensile Index (TAPPI T-494 om-96), Burst Index (TAPPI T-403 om-97), Tear Index (TAPPI T-414 om-98) and Gurley porosity (TAPPI T-460 om-96) following conditioning in accordance with TAPPI T-220 sp-96. Each value in the experimental results is an average of 12 samples. **The deviations for these parameters from their respective means were less than 5%.**

3. Results and discussion

3.1. Chemical and energetic characteristics of raw material

Table 1 shows the chemical and energetic characteristics of *Leucaena* and other lignocellulosic materials. The *Leucaena*-derived material consisted of 32.2% cellulose (analyzed as glucan), (40.2% as α -cellulose), 18.6% hemicelluloses, calculated as the sum of xylan, araban and acetyl groups, (32.6% based on holocelulose - α -cellulose), and 21.5% Klason lignin (23.7% after quantitative acid hydrolysis). This composition is similar to that found by other authors for *Leucaena diversifolia* (Alfaro et al., 2009) and other varieties of *Leucaena* (Jimenez et al., 2007) and comparable to hardwoods, such as *Eucalyptus globulus* (Garrote

1 et al., 2003). The hemicellulose content in *Leucaena* was similar to that of tagasaste
2 (41.4%) (Alfaro et al., 2010) and herbaceous species.
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7 The molar ratio of the monomers in *L. diversifolia* was calculated as
8 xylose:acetyl groups:arabinose = 15.5:2.1:1. This composition is typical of
9 acetylglucuronoxylans typically present in hardwoods (Pinto et al., 2005).
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15 The gross heating value (GHV) of *L. diversifolia* of 19.1 MJ/kg, is in between
16 those for softwood (20.0 MJ/kg) and hardwood (18.8 MJ/kg) (Demirbas, 1997),
17 similar to that of *Ailanthus* wood (19.0 MJ/kg) (Demirbas, 1997), somewhat
18 higher than that of *Eucalyptus globulus* (Telmo et al., 2010), and much higher than
19 those of wheat straw (17.0 MJ/kg) or corn stover (17.8 MJ/kg) (Demirbas, 1997).
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29 The GHV for the solid residue remaining after *Leucaena* autohydrolysis is
30 shown in Table 2 together with the contents of the autohydrolysis solid phase. The
31 GHV for the solid phase ranged from 19.1 to 19.9 MJ/kg depending on the
32 autohydrolysis conditions. Table 4 shows the equations modelling the dependent
33 variables of the autohydrolysis process. The equation for Y_{CV} reflects the linear
34 variation of GHV for the solid residue with the operational temperature and treatment
35 time; as can be seen, the highest GHV was obtained with the most drastic hydrothermal
36 conditions.
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48 The autohydrolysis treatment raised the calorific value of the solid phase by up
49 to 4% with respect to the starting raw material. This can be ascribed to the relative
50 increase in the amount of lignin and cellulose present in the solid phase as
51 polysaccharides are extracted and to the increased calorific value of lignin relative to
52 cellulose and hemicelluloses. This further increases the intrinsic value of the
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1 compounds extracted into the liquor or reduces the energy cost associated with
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3 extraction by autohydrolysis.
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9 *3.2. Products of hydrothermal treatment. Modeling and optimization.*
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12 **Table 2 shows the normalized values of independent variables (temperature and**
13 **operation time) and the Gross Heating Values, yield, Klason lignin and soluble**
14 **lignin contents in the solid phase relative to the initial raw material, and also the**
15 **glucan, xylan, araban contents and acetyl groups in the solid phase, relative to the**
16 **initial respective polymer content in raw material.**
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25 **Table 3 shows the monomeric sugar and oligomer concentrations in the liquid**
26 **phase in g/L and as percentage relative to respective polymer content in dry raw**
27 **material. The results were modeled by using the multiple regression methodology**
28 **described in section 2.3.2. (Table 4).**
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35 The solid phase yield obtained under such conditions was 25% lower than that
36 provided by less drastic conditions. The yield exceeded 80% in 70% of the tests. The
37 Y_{YI} model in Table 4 suggests a linear relationship of yield with both operational
38 variables, to which it was inversely proportional. The statistically significant presence
39 of the quadratic term X_7X_7 and the interaction X_7X_t suggests that the influence of
40 **temperature on yield was considerably smaller at short treatment times (yields of**
41 **90-96%) relative to long times (yields of 72-90%).** This confirms the hypothesis that
42 the polysaccharide fraction is hydrolysed from lignocellulosic biomass at fairly high
43 temperatures, but requires an also fairly long time to occur.
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57 The content in Klason lignin of the autohydrolysis solid fraction reflected its
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1 nature as a non-soluble fraction of lignocellulosic biomass. Thus, it ranged from 22.6 to
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3 25.1% of the amount present in the starting raw material, so it changed by a mere 10.2%
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5 at most and had an average value of 23.3%. The errors inherent in the methods used to
6
7 determine lignin preclude more precise conclusions; therefore, insoluble lignin can be
8
9 assumed not to be degraded or solubilized in substantial amounts within the studied
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11 operating ranges. Its average value is close to the lignin content in the raw material as
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13 determined in accordance with TAPPI standard (21.5%) and the HPLC protocol for the
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15 determination of sugars (23.7%).
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21 **The amount of soluble lignin was, in most cases, lower (2.1% in the raw**
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23 **material and 0.82-2.03% in the autohydrolysis solid fraction). This result suggests**
24
25 **substantial dissolution of this fraction during autohydrolysis (Leschinnsky et al.,**
26
27 **2009).** The treatment time was the more influential factor, and its influence and that of
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29 temperature, essentially linear –with a slight contribution of the quadratic term X_7X_7 .
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34 Glucan was the individual polysaccharide affected to the smallest extent by the
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36 autohydrolysis treatment, 90.1% to 98.1% of its content remaining in the autohydrolysis
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38 solid phase. **In a biorefinery process that includes paper production, the amount of**
39
40 **sugars in solution should be as high as possible, but degradation of cellulose (as**
41
42 **glucan) in the raw material should be avoided as far as possible in order to obtain**
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44 **pulp of acceptable properties (Alfaro et al., 2010).** Based on the foregoing, the
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46 glucose and glucooligomer contents in the autohydrolysis liquid phase were relatively
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48 low (0.3–1.7% and 2.2–8.4%, respectively).
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53 The glucose content in the autohydrolysis liquid phase (Y_{GL} equation in Table 4)
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55 was only influenced by linear terms of the operational variables. Clearly, glucan
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57 dissolution as glucose in the liquid phase increased with increasing temperature and
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1 treatment time. The glucooligomer content of the autohydrolysis liquid phase and the
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treatment time. The glucooligomer content of the autohydrolysis liquid phase and the
glucan content of the solid phase were clearly dependent on the quadratic terms $X_T X_T$
and $X_i X_i$, both of which were statistically significant. Although their coefficients were
much lower than those for the linear terms, the presence of these two terms made
interpreting the models more complicated.

In order to determine the values of the independent variables giving the optimum variable dependent results, the response surfaces for each dependent variable were plotted at maxima and minima levels of the independent variables (Figures 2 and 3). Figure 2 shows the three response surfaces for the glucan derivatives in both autohydrolysis phases. The influence of the variables was essentially linear despite the presence of significant quadratic terms. The mass balance for the glucan derivatives was subject to a negligible error (the experimental proportions ranged from 99.5 to 100.7%).

The xylose and xylooligomer contents exhibited substantial variations (0.2–8.8% and 1.1–51.9%, respectively - percentage relative to the content in xylan fraction of the raw material). These results are consistent with the variation of the xylan content of the autohydrolysis solid phase. In fact, the mass balance for xylan derivatives was less precise than that for glucan derivatives (total xylan contents ranged from 99 to 106%).

Modeling the results for this hemicellulose fraction was also complicated. Thus, the Y_{Xy} , Y_{XYOL} and Y_{XYS} models in Table 4 contain statistically significant quadratic or interaction terms and are difficult to interpret in a semi-quantitative manner without the response surfaces of Figure 3. **The hydrolysis of hemicellulosic xylan in the raw material was substantial over the operational range 0-1 for both operational variables. Based on the results presented in Table 2, xylan was hydrolysed to an approximately 20% greater extent (20.76% according to the Y_{XYS} model of Table 4).**

1 Xylan hydrolysis increased markedly in the upper ranges of the independent operational
2 variables (particularly with the treatment time, as can be inferred from the coefficients
3 of the linear terms in the Y_{XYS} model of Table 4 and from Figure 3). **In principle, it is**
4 **advisable to use medium or high temperatures and short or medium treatment**
5 **times to extract xylose and xylooligomers similarly as with medium temperatures**
6 **and long times. Also there is less degradation of glucan (roughly 2 percent points**
7 **more of glucan in the autohydrolysis solid phase based on the Y_{GLS} model of Table**
8 **4).**

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10
11 **Autohydrolysis extracted araban to a similar extent as xylan derivatives, but**
12 **time was more influential than temperature on its hydrolysis (see Y_{ARS} equation in**
13 **Table 4).** Although more than 90% of all araban initially present in the raw material
14 was extracted under the most drastic conditions, the hemicellulose fraction obtained was
15 less useful than that of xylan as it contained hardly 1% of the initial mass of raw
16 material. The cumulative experimental error in the determination of araban in the liquid
17 and solid phases was similar to that for xylan derivatives, with proportions from **91.6 to**
18 **103.7% (data not shown).**

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21 **In regards acetyl groups, the hydrothermal treatment caused the dissolution of**
22 **hemicellulosic polysaccharides. The effect was strengthened by the release of acetyl**
23 **groups from hemicelluloses increasing the acidity of the medium (Önnerud and**
24 **Gellersted, 2003). All acetyl groups were dissolved under the most drastic treatment**
25 **conditions used.**

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27
28 **As a rule, extraction of hemicellulosic derivatives from *L. diversifolia* was**
29 **more efficient than that from other batches of the same plant grown for 3 years**
30 **and subjected to non-isothermal treatments (Alfaro et al., 2009); however, it was**
31 **less efficient than that from *Chamaecytisus proliferus* (Alfaro et al., 2010), another**
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1 **legume. The glucan fraction in Leucaena was less markedly** degraded than that in
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3 Chamaecytisus, which in theory made the solid phase from the former more suitable for
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5 producing cellulose pulp with a biorefining scheme.
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8 A comparison of the components extracted from the hemicellulose fraction of
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10 other plants such as sunflower stalks hydrothermally treated at 165–180 °C revealed
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12 that, although the stalks contained a similar amount of glycan and more xylan (23.9%)
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14 than Leucaena (15.5%), the resulting liquid phases contained similar amounts of
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16 glucose and xylose, and somewhat smaller amounts of oligosaccharides in Leucaena
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18 (16.8 g/L vs 21.0 g/L in sunflower stalks) (Caparrós et al., 2008). **Applying a similar**
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20 **thermal treatment to Paulownia fortunei, a tree species of similar composition as**
21
22 **Leucaena** (34.2% glucan and 18.3% xylan) yielded less glucose (0.84 vs 1.12 g/L) but
23
24 more xylose (2.05 vs 1.22 g/L) and oligomers (16.84 vs 8.08 g/L). Using similar
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26 conditions with eucalyptus provided greater amounts of dissolved sugars (Romani et al.,
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28 2011).
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38 *3.3. Characterization of paper sheets from solid phase post-autohydrolysis*

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40 In addition to determining and modelling the gross heating value for the autohydrolysis
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42 solid phase, we used this material to obtain cellulose pulp and paper with a view to
43
44 assessing its potential valorization by biorefining. **Cellulose pulp was obtained from**
45
46 **the autohydrolysis solid phase (carried out at 178 °C for 0 min. non-isothermal**
47
48 **conditions) and the properties of paper prepared from this pulp were: Kappa**
49
50 **number 40.4, viscosity 1119 g cm⁻³ and brightness 29.3 %. The conditions of the**
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52 **hydrothermal treatment preceding delignification were adjusted in such a way as**
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54 **to ensure a high content in cellulose (as glucan) in the solid phase and in potentially**
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56 **valorizable hemicellulosic sugars in the liquid phase. Based on the equations of**
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1 **Table 4, the resulting liquid phase should contain 2.4% xylose and 26.4% xylo-**
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3 **oligomers respect to the initial concentration of xylan in the raw material, whereas**
4
5 **the solid phase should consist of 92.2% glucan. The yield of the solid phase would**
6
7 **be 81%, its gross heating value 19.6 MJ/kg and the soluble lignin content in the**
8
9 **solid phase 1.2% (i.e. 43% of all initial lignin would be dissolved by the**
10
11 **autohydrolysis treatment).**
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15 In parallel, we obtained unhydrolysed cellulose pulp from *L. diversifolia* under the
16 conditions described in Section 2.4 except that the temperature and soda concentration
17 were 15 °C and 4 percent points higher than those used to obtain pulp from the
18 **autohydrolysis solid phase. The Kappa number, viscosity, and brightness of this**
19 **paper were 27, 1303 g cm⁻³, and 38.5%, respectively.** We hypothesized that the
20 autohydrolysis treatment would destroy the cellulose matrix, thereby facilitating
21 delignification by biorefining and that using the same operational conditions as in the
22 absence of a autohydrolysis treatment would result in excessive degradation of cellulose
23 fibres (Alfaro et al., 2010).
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38 Figure 4 shows the results of the characterization tests for **the major strength-**
39 **related properties of** the resulting paper (tear index, burst index and tensile index), as
40 well as the Gurley porosity of handsheets made by applying variable refining as
41 measured by the Shopper–Riegler index. **The data presented in Figure 4 allows the**
42 **following conclusions:**
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49 (a) The autohydrolysis treatment preceding delignification slightly degraded the
50 strength-related properties and Gurley porosity of the resulting paper.
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54 (b) The treatment substantially facilitated refining of the pulp and increased its strength
55 as a result. This suggests that ethanol–soda pulp from the autohydrolysed material
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1 is amenable to refining and can provide paper with substantially **improved**
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3 **strength-related properties.**
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5 **Similar results were obtained in previous studies on another legume variety,**
6 ***Chamaecytisus proliferus*, subjected** to organosolv delignification with ethanol
7
8 (Alfaro et al., 2010), which is less favoured on a industrial scale.
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11
12 As can also be seen from Figure 4, the improvement in physical properties of the
13 paper sheets was maximal at low Schopper Riegler degree values (up to 20 °SR),
14 whether or not the raw material was subjected to autohydrolysis. The improvement was
15 also substantial between 20 and 38 °SR, but further refining (45–50 °SR) led to worse
16 results. Therefore, pulp from autohydrolysed material should never be refined beyond
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18 40 °SR.
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28 The tear index for paper from autohydrolysed pulp was 18.8% higher at 20–21
29 °SR en even more so (53.7%) at 35–38 °SR. This was the strength-related property most
30 **markedly improved by the autohydrolysis treatment; however, there were also**
31 **substantial** improvements in burst index (22.8–39.0%) and tensile index (10.6–35.7%).
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38 The tensile index and burst index of paper from *L. diversifolia* pulp obtained by
39 non-isothermal autohydrolysis of the raw material, followed by refining at 38 °C, was
40 1.8 and 3.5 times higher, respectively, than that for eucalyptus paper at 181 °C and
41 ethanol delignification (Garrote et al., 2003). Similar results were obtained relative to
42 the legume tagasaste, which exposed the beneficial effect of the thermal treatment and
43 the use of ethanol as delignifying agent on pulp refining (Alfaro et al., 2010) .
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53 The combined use of non-isothermal autohydrolysis and ethanol–soda-
54 anthraquinone delignification to obtain handsheets with improved strength-related
55 properties yielded optimum results under those conditions leading to maximal removal
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1 of lignin fibres while preserving part of hemicellulose. In fact, the hemicellulose
2 fraction plays a central role in fibre swelling and interfibre bonding (García Hortal,
3 2007). It is therefore advisable to preserve a relatively high proportion of xylan –which
4 amounted to 72.43% with the optimum autohydrolysis conditions. The physical
5 properties of paper can be improved by refining the pulp, which removes the primary
6 wall of fibres and facilitates their hydration and swelling, thereby increasing their
7 flexibility and bonding capacity. Refining tends to optimize bonds between fibres at the
8 expense of a loss of individual strength to an extent dependent on the particular raw
9 material and pulping method (Hietanen and Ebeling, 1990).

24 **4. Conclusions**

27 *Leucaena diversifolia* lends itself readily to valorization for energy production,
28 and also to integral, fractional exploitation by autohydrolysis and ethanol–soda–
29 anthraquinone delignification, which can additionally bring environmental benefits to
30 cropping zones. The gross heating value for *Leucaena* is 19.083 MJ/kg. The
31 autohydrolysis solid phase can be used to obtain other chemicals, directly burnt for
32 energy production or processed to obtain cellulose pulp. In addition to providing a
33 liquid phase containing valuable chemicals (18.65 g/L of sugars at 178 °C), the
34 autohydrolysis facilitates refining of cellulose pulp, allows the tear and tensile index to
35 be increased by 53.8 and 22.2%.

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Tables

Table 1a: Chemical composition of *Leucaena Diversifolia*.

Table 1b: Chemical composition of *Leucaena* and others lignocellulosic raw materials

Table 2: Values of independent variables, Gross Heating Value, solid yield, Klason and soluble lignin content (relative to the initial raw material –dry mass-) and composition of the solid fractions (relative to the content in each polymer fraction of the raw material –dry mass-) obtained in the autohydrolysis process using the proposed experimental design

Table 3: Values of independent variables and composition of the liquid fractions obtained in the autohydrolysis process using the proposed experimental design (mg/L and percentage relative to the content in each polymer fraction of the raw material –dry mass-. Oligomer contents are give as monomer equivalents)

Table 4: Equations yielded for each dependent variable.

Figures

Figure 1: Scheme of experimental work

Figure 2: Glucose and gluco-oligomers of the liquor and glucan of the solid as function of temperature and time of the autohydrolysis process.

Figure 3: Xylose and xylo-oligomers of the liquor and xylan of the solid as function of temperature and time of the autohydrolysis process

Figure 4: Properties of the paper sheets variations (deviations from means values were less than 5%) vs. Shopper-Riegler degree.

Table 1a

<i>Leucaena diversifolia</i>		Chemical and energetic characteristics	
TAPPI characterization		Quantitative acid hydrolysis and HPLC characterization	
1% NaOH soluble.....	14.9 %	Glucan	32.2 %
Hot water solubles	4.8 %	Xylan.....	15.5 %
Alcohol-benzene extractives	3.9 %	Araban	1.0 %
Lignin (after 72% sulphuric hydrolysis)	23.7 %	Acetyl groups	2.1 %
Hot water solubles (after organic extraction)	3.9 %	Klason lignin	21.5%
Holocellulose.....	72.8 %	Soluble lignin	2.1 %
α -cellulose	40.2 %	Ash	1.4 %
Calorific value	19.083 MJ/kg		

Table 1b

Raw Material	Cellulose (%)	Hemicellulose (%)	Lignin (%)	References
Arundo donax (reed)	34.8	29.7	23.0	Caparrós et al., 2007
<i>Chamaecytisus proliferus</i> (tagasaste)	38.9	41.4	19.8	Alfaro et al., 2010
<i>Eucalyptus globulus</i>	43.0, 46.8, 53.4	13.2, n.d., n.d.	25.3, 22.9, nd.	Garrote et al., 2003
<i>Eucalyptus gumifera</i> and <i>saligna</i>	38.0, 45.0	16.0, 15.0	37.0, 25.0	Mok and Antal, 1992
<i>Helianthus annuus</i>	33.8	40.4	19.9	Caparrós et al., 2008
Leucaena	43, 40.8	17, 15.0	25, 26.9	Mok and Antal, 1992
<i>Leucaena leucocephala</i> (Honduras)	39.4, 41.0, 41.2,	28.9, 26.0, 32.9,	18.4, 26.0, 19.4,	Mok and Antal, 1992
<i>L. leucocephala</i> (India)	44.4	31.5	21.4	Jiménez et al., 2007
<i>Leucaena diversifolia</i>	38, 40.10	27.8, 37.8	24.8, 19.09	Alfaro et al., 2009
<i>Leucaena colinsi</i>	43.8	37.0	17.0	Jiménez et al., 2007
<i>Paulownia</i>	48.3, 44.0, 34.2	20.5, 21.2, 22.7	22.1, 27.8, 27.2	Feria et al., 2011, García et al., 2011

Table 2

Normalized values of Temperature (X_T) and Operation Time (X_t)		Gross Heating Value (MJ/Kg)	Yield (%)	Klason Lignin (%)	Soluble Lignin (%)	Glucan (%)	Xylan (%)	Araban (%)	Acetyl Groups (%)
-1	-1	19.143	96	23.3	2.03	98.1	98.7	95.4	81.3
-1	0	19.316	91	23.4	1.63	95.0	94.7	75.4	74.4
-1	1	19.634	87	23.1	1.36	92.6	84.1	58.7	60.0
0	-1	19.342	95	25.1	1.86	95.7	97.5	78.7	71.1
0	0	19.532	84	23.3	1.29	92.7	80.5	54.8	47.9
0	0	19.489	84	23.7	1.31	92.9	79.9	55.5	48.3
0	1	19.723	78	23.1	1.00	90.1	62.5	32.1	26.1
1	-1	19.386	92	22.6	1.57	95.6	90.8	58.7	55.6
1	0	19.666	77	22.6	1.15	91.9	63.4	32.9	24.2
1	1	19.870	72	22.7	0.82	90.1	41.4	8.3	0.0

Table 3

Normalized values of Temperature (X_T) and Operation Time (X_i)		Glucose (%/g/l)	Xylose (%/g/l)	Arabinose (%/g/l)	Acetyl Groups (%/g/l)	Gluco-oligomers (%/g/l)	Xylo-oligomers (%/g/l)	Araban-oligomers (%/g/l)
-1	-1	0.3/0.15	0.2/0.05	1.3/0.02	1.0/0.03	2.2/1.14	1.1/0.27	4.2/0.04
-1	0	0.8/0.41	0.4/0.09	6.2/0.10	5.0/0.15	4.8/2.45	5.0/1.19	18.8/0.18
-1	1	1.6/0.79	2.5/0.59	14.3/0.21	18.9/0.58	6.2/3.15	13.5/3.20	21.2/0.20
0	-1	0.6/0.31	0.2/0.05	6.0/0.09	2.0/0.06	3.7/1.91	2.3/0.56	9.5/0.09
0	0	1.2/0.59	1.7/0.40	12.8/0.19	13.3/0.41	6.7/3.37	21.7/5.15	27.7/0.26
0	0	1.1/0.53	1.6/0.38	13.3/0.19	12.8/0.4	6.8/3.46	24.4/5.80	29.1/0.28
0	1	1.5/0.77	5.0/1.18	18.2/0.26	35.0/1.07	8.4/4.21	38.7/9.12	35.1/0.33
1	-1	0.6/0.33	0.9/0.21	6.6/0.10	7.2/0.22	3.8/1.94	8.2/1.97	13.5/0.13
1	0	1.2/0.61	2.9/0.67	13.6/0.19	20.1/0.61	7.0/3.50	34.7/8.16	37.5/0.36
1	1	1.7/0.84	8.9/2.05	20.2/0.29	58.4/1.77	7.7/3.86	51.9/12.15	50.5/0.48

Table 4

Equation	R ²	F-Snedecor
Solid phase		
$Y_{YI} = 84 - 5.50 X_T - 7.67 X_t + 2.67 X_t X_t - 2.75 X_T X_t$	0.98	82
$Y_{CV} = 19510.1 + 138.2 X_T + 226 X_t$	0.98	140
$Y_{SL} = 1.35 - 0.25 X_T - 0.38 X_t + 0.10 X_t X_t$	0.99	137
$Y_{GLS} = 92.60 - 1.35 X_T - 2.77 X_t + 0.98 X_T X_T + 0.45 X_t X_t$	0.99	233
$Y_{XYS} = 79.24 - 13.67 X_T - 16.02 X_t - 9.28 X_T X_t$	0.99	354
$Y_{ARS} = 61.39 - 13.12 X_T - 20.35 X_t$	0.99	403
$Y_{AGS} = 29.28 - 19.60 X_T - 26.01 X_t + 24.44 X_T X_T - 9.47 X_T X_t$	0.99	373
Liquid phase		
$Y_{GL} = 1.05 + 0.148 X_T + 0.538 X_t$	0.97	116
$Y_{XY} = 1.63 + 1.57 X_T + 2.49 X_t + 1.29 X_t X_t + 1.40 X_T X_t$	0.99	123
$Y_{AR} = 28.28 + 9.55 X_T + 13.26 X_t - 5.96 X_t X_t + 5.04 X_T X_t$	0.99	1230
$Y_{AG} = 12.79 + 10.16 X_T + 17.03 X_t + 7.63 X_t X_t + 8.53 X_T X_t$	0.99	107
$Y_{GLOL} = 6.8 + 0.88 X_T + 2.095 X_t - 0.98 X_T X_T - 0.809 X_t X_t$	0.99	135
$Y_{XYOL} = 20.14 + 12.55 X_T + 15.41 X_t + 7.84 X_T X_t$	0.97	76
$Y_{AROL} = 12.58 + 2.98 X_T + 6.47 X_t - 2.11 X_T X_T$	0.99	289

Where:

Solid phase:

Y_{YI} , Y_{CV} and Y_{SL} denotes the solid yield, Gross Heating Value and soluble lignin contents in solid phase after autohydrolysis respect initial raw material (dry basis).

Y_{GLS} , Y_{XYS} , Y_{ARS} and Y_{AGS} denotes glucan, araban, xylan and acetyl groups contents in solid phase after autohydrolysis respect initial glucan, xylan, araban and acetyl groups contents in raw material (dry basis). X_T and X_t denotes normalized autohydrolysis temperature and time, respectively.

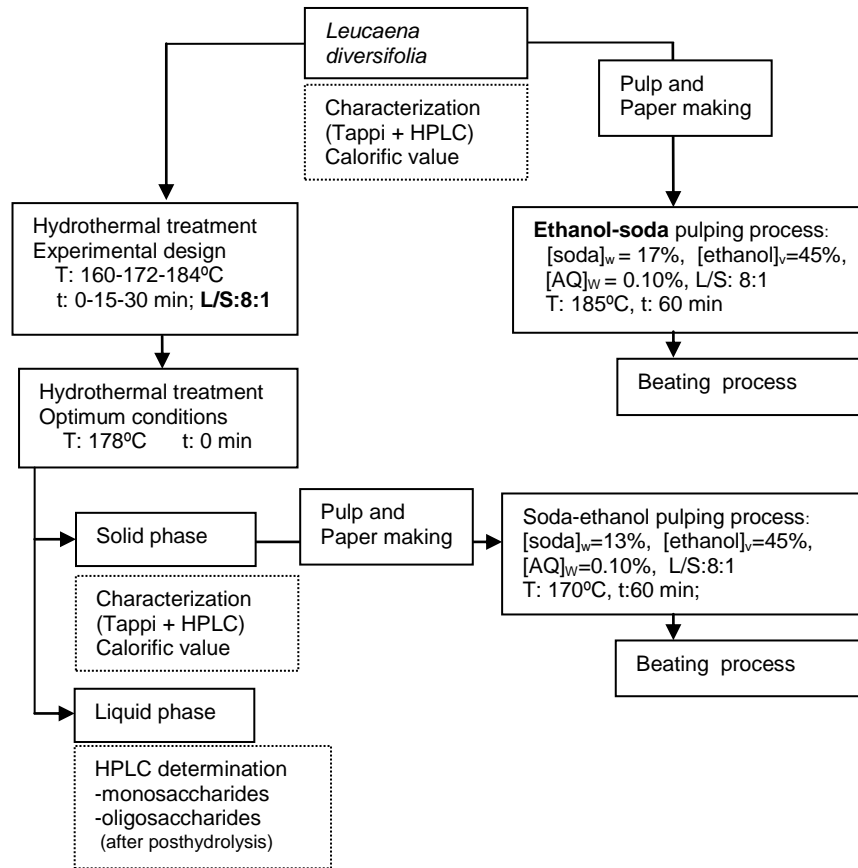
Liquid phase:

Y_{GL} , Y_{XY} , Y_{AR} and Y_{AG} denotes glucose, xylose, arabinose and acetic acid contents in liquid phase after autohydrolysis respect initial glucan, xylan, araban and acetyl groups contents in raw material (dry basis).

Y_{GLOL} , Y_{XYOL} and Y_{AROL} , denotes gluco-oligomers, xylo-oligomers and araban-oligomers contents in liquid phase after autohydrolysis respect initial glucan, xylan and araban contents in raw material (dry basis) respectively. Oligomer contents are give as monomer equivalents.

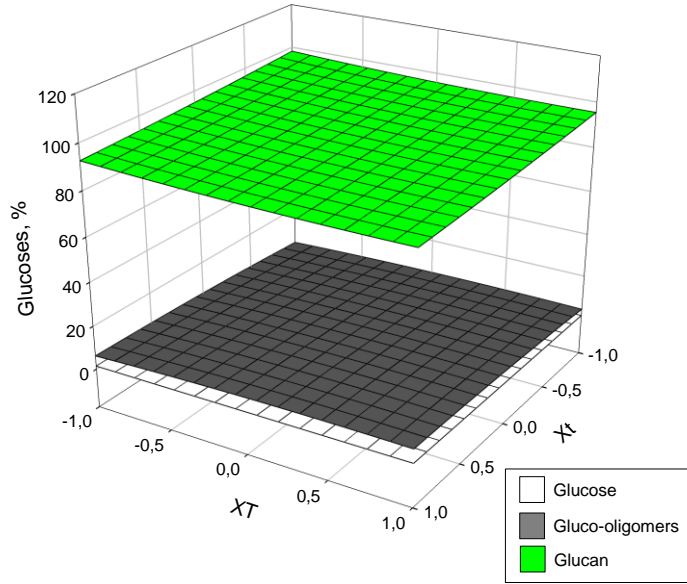
The differences between the experimental values and those estimated by using the previous equations never exceeded 5% of the former (10% for Y_{YI} , Y_{SL} , Y_{GL} , Y_{GLOL} , Y_{GLS}).

Figure 1



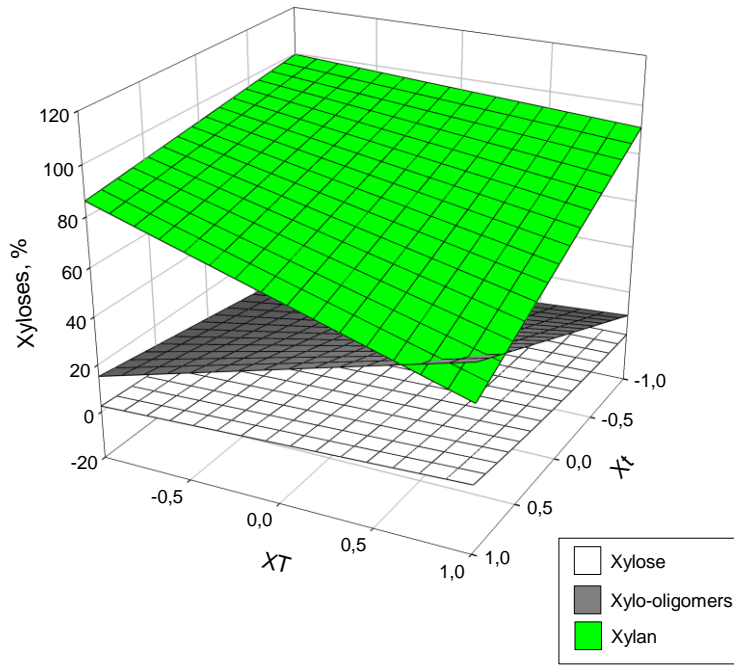
Abbreviations: [soda] w: soda concentration by weight; [ethanol] v: concentration of ethanol by volume; [AQ] w: concentration of anthraquinone by weight; T: process temperature (°C); t: processing time (min). L/S: liquid / solid ratio.

Figure 2



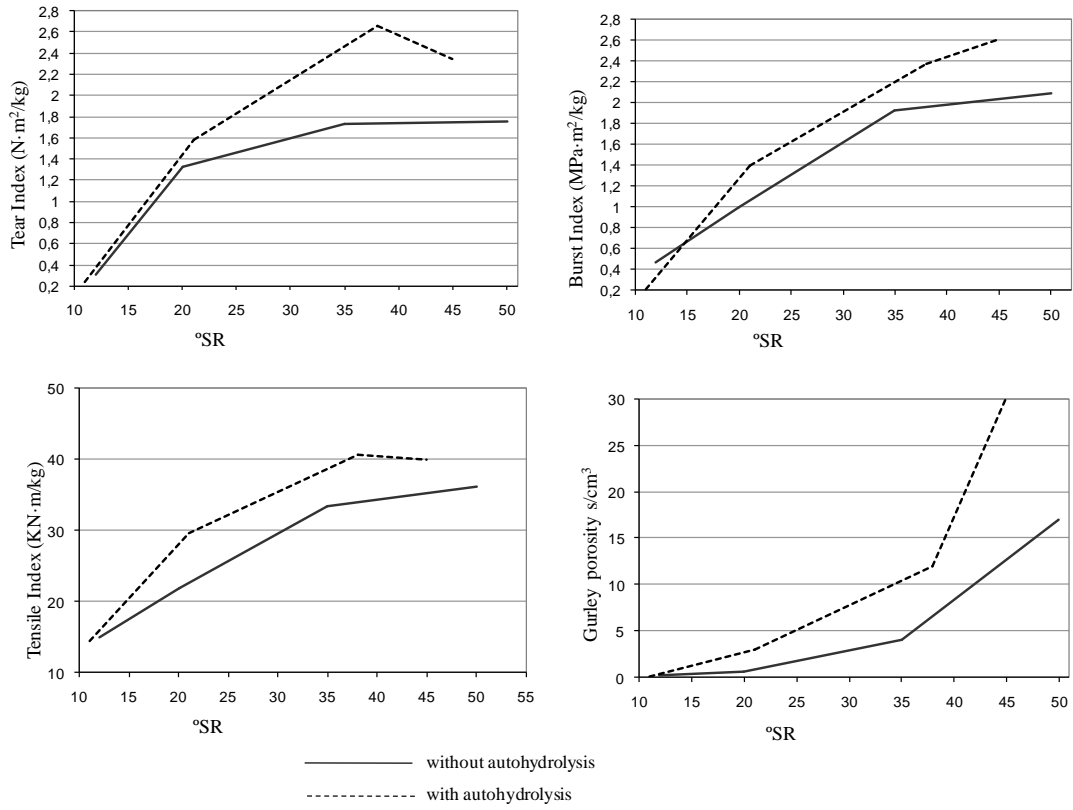
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Figure 3



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Figure 4



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