# Southern Illinois University Carbondale OpenSIUC

#### Articles

Department of Plant, Soil, and Agricultural Systems

2012

## Genetic Analysis of Relative Water Content (RWC) in Two Recombinant Inbred Line Populations of Soybean [Glycine max (L.) Merr.]

Salvador Virginia Fayetteville State University

Megan Pagan Fayetteville State University

Melissa Cooper Fayetteville State University

Stella K. Kantartzi Southern Illinois University Carbondale

David A. Lightfoot Southern Illinois University Carbondale, ga4082@siu.edu

See next page for additional authors

Follow this and additional works at: http://opensiuc.lib.siu.edu/psas\_articles Published in *Journal of Plant Genome Sciences*, Vol. 1 No. 2 (2012) at doi: 10.5147/jpgs.2012.0058

#### **Recommended** Citation

Virginia, Salvador, Pagan, Megan, Cooper, Melissa, Kantartzi, Stella K., Lightfoot, David A., Meksem, Khalid and Kassem, My A. "Genetic Analysis of Relative Water Content (RWC) in Two Recombinant Inbred Line Populations of Soybean [Glycine max (L.) Merr.]." (Jan 2012).

This Article is brought to you for free and open access by the Department of Plant, Soil, and Agricultural Systems at OpenSIUC. It has been accepted for inclusion in Articles by an authorized administrator of OpenSIUC. For more information, please contact opensiuc@lib.siu.edu.

#### Authors

Salvador Virginia, Megan Pagan, Melissa Cooper, Stella K. Kantartzi, David A. Lightfoot, Khalid Meksem, and My Abdelmajid Kassem

Journal of Plant Genome Sciences 1 (2): 46–53, 2012 doi: 10.5147/jpgs.2012.0058

### Genetic Analysis of Relative Water Content (RWC) in Two Recombinant Inbred Line Populations of Soybean [*Glycine max* (L.) Merr.]

Salvador Virginia<sup>1</sup>, Megan Pagan<sup>1</sup>, Melissa Cooper<sup>1</sup>, Stella K. Kantartzi<sup>2</sup>, David A. Lightfoot<sup>2</sup>, Khalid Meksem<sup>2</sup>, and My Abdelmajid Kassem<sup>1\*</sup>

<sup>1</sup> Plant Genomics and Biotechnology Lab, Department of Biological Sciences, Fayetteville State University, Fayetteville, NC 28301, USA; <sup>2</sup> Department of Plant, Soil, and Agricultural Systems, Southern Illinois University, Carbondale, IL, 62901, USA.

Received: May 27, 2011 / Accepted: March 10, 2012

#### Abstract

Drought affects soybean [Glycine max (L.) Merr.] and other crops productivity in the US and other parts of the world. Relative water content (RWC) is an important indicator for plant water deficit tolerance (WDT). The objective of this study is to map quantitative trait loci (QTL) for RWC and several other leaf traits such as leaf dry weight (LDW), leaf fresh weight (LFW), and leaf turgid weight (LTW) in two soybean recombinant inbred line (RIL) populations, one derived from a cross of 'Essex' and 'Forrest' (ExF, n=94) and the other is derived from a cross of 'PI 438489B' and 'Hamilton' (PIxH, n=50). In the PIxH RIL population, eight QTL were identified and mapped on 6 different linkage groups (LGs) of the soybean genome. No QTL for LFW were identified in this population. In the ExF RIL population, 10 QTL were identified and mapped on 5 different LGs of soybean. Chromosome 18 (LG G) contains clusters of QTL for LFW, LTW, and RWC in the ExF RIL population. This same chromosome contains a QTL for RWC in the PIxH RIL population. The QTL found here are important to be included in breeding programs for soybean water deficit tolerance (WDT).

**Keywords:** QTL, soybean, drought, water deficit tolerance (WDT), relative water content (RWC).

#### Introduction

Soybean [*Glycine max* (L.) Merr.] is an important crop cultivated in the US and worldwide. In 2009, the US exported 1.28 billion bushels of soybeans which accounts for 46% of the world's total production and a profit that exceeded \$21 (Soy Stats, 2012).

Soybean is a great source of proteins and oils (Howell and Carter, 1958; Hartwig, 1973). Soybeans are also used for soy diesel, candles, soy milk, building materials, soy crayons, hand cleaners, and many other products (USB, 2007). Soybean seeds contain numerous nutraceuticals compounds such as isoflavones or phytoestrogens (Eldridge and Kwolek, 1983; Regal et al., 2000). The most studied, in soybean, are daidzein, genistein, and glycitein (Barnes et al., 1990, 1994; Messina and Barnes, 1991; Cassidy et al., 1990, 1994; Huang et al., 1992; Fostis et al., 1993; Jing et al., 1993; Peterson et al., 1993; Anderson et al., 1995; Knight and Eden, 1996; Anthony et al., 1996; Banz et al., 1999; Greer-Baney et al., 1999; Njiti et al., 1999; Regal et al., 2000; Meksem et al., 2001; Munro et al., 2003 ; Kassem et al., 2004). The last decades witnessed the use of large amounts of soy products including isoflavones; however, several studies showed that they might have positive and negative health effects in humans and animals (Barnes et al., 1990, 1994; Njiti et al., 1999; Regal et al., 2000; Meksem et al., 2001).

Corresponding author: mkassem@uncfsu.edu.

Soybeans, like other crops, are subject to numerous diseases and sensitive to drought (Specht et al., 2001). Researchers and plant breeders look to develop soybean cultivars that are drought tolerant or resistant. Drought stress might lower the ability of roots to absorb enough water which leads to wilting. Soybean plants defend themselves against water loss by closing their stomata and avoid dehydration. However, this might reduce the photosynthetic rate as well as the uptake of minerals from the soil. This reduced metabolism, caused by drought, might be reversible when enough water is available. Slow photosynthetic rate has a drastic effect on yield. The stressed plants usually slow or stop growth and flowering, abort grain, drop leaves, and abort pods.

The degree of drought stress can be physiologically estimated by measuring the leaf relative water content (RWC) which can control the plant response to water deficit (Hunt et al., 1987). Mapping QTL for agronomic traits including traits involved in drought tolerance such as water use efficiency (WUE), RWC, and root traits have been identified and mapped in wheat (Dhanda and Sethi, 1998), barley (Teulat et al., 1997, 2003), sunflower (Kiani et al., 2007), corn (Frova et al., 1999), soybean (SoyBase, 2011), and many other crop species. However, the genes controlling these traits and many others are still widely unknown (Keurentjes et al., 2008).

The objective of this study is to genetically map quantitative trait loci (QTL) for leaf fresh weight (LFW), leaf turgid weight (LTW), leaf dry weight (LDW), and leaf relative water content (RWC) in soybean using two recombinant inbred line populations (RIL): One derived from the cross of 'Essex' and 'Forrest' cultivars (ExF, n = 94) and the second is derived from the cross of 'PI 438489B' and 'Hamilton' (PIxH, n = 54) and to compare results from both RIL populations.

#### **Materials and Methods**

#### **Plant Materials**

The 'Essex' by 'Forrest' RIL population (EXF, n = 94) used in this study (Lightfoot et al., 2005) was provided in 2007 (at the F5:16) by Prof. D.A. Lightfoot of SIUC. This ExF RIL population was extensively studied for sudden death syndrome (SDS) re-

sistance, soybean cyst nematode (SCN) resistance, manganese toxicity resistance, seed yield, and several other traits (Kassem et al., 2004a,b, 2006, 2007a,b; Alcivar et al., 2007; Jacobson et al., 2007).

The 'Hartwig' by 'Flyer' RIL population (HxF, n=92) used in this study was also provided in 2007 by Prof. D.A. Lightfoot of SIUC. This HxF RIL population was extensively studied for seed yield, sudden death syndrome (SDS) resistance, maturity, and lodging (Kazi, 2005; Kazi et al., 2007, 2008).

#### **Growth Conditions**

Growth conditions were performed as described in Jacobson et al. (2007). Briefly, one seed per pot, with 3 seed of each RIL, were grown in pots of 30 x 30 cm (diameter x depth) containing approximately 1 Kg of PRO-MIX soil. The PRO-MIX is a ready-made, peat based growing mix containing the Canadian sphagnum peat moss (75-85% by volume), limestone (for pH adjustment), perlite, a wetting agent, and vermiculate. The seeds of the two populations (ExF, n=20; HxF, n=20) were planted in the greenhouse for three months (October – December, 2008). The plants were kept in the greenhouse at  $25\pm10C$  under natural lighting and harvested after 3 months and the relative water contents (RWC) were measured as described below.

#### Relative Water Content Measurement

Relative water content (RWC) was estimated as described previously (Smart and Bingham, 1974). Briefly, the fresh weight of a mixed sample of five young leaves from each plant from each population is determined. The leaves of the same sample were left floating on distilled water, in Petri dishes, for 4 hours and the turgid weight is then recorded. After that, the leaf tissues were dried in an oven at 650C for 24 hrs and their dry weight was measured. The RWC is calculated according to the following formula (Smart and Bingham, 1974):

Table 1. Means of the leaf traits measured in the 'PI438489B' by 'Hamilton' RIL population and parent lines (PI438489B' and Hamilton).

		RIL	s (n=54)	Parents		
Trait	Mean	St. Dev.	Skewness	P <w*< th=""><th>PI438489B mean</th><th>Hamilton mean</th></w*<>	PI438489B mean	Hamilton mean
Leaf Fresh Weight (g)	0.1924	0.0219	0.8784	0.0023**	0.2636	0.1614
Leaf Turgid Weight (g)	0.3565	0.0299	0.7045	0.0955ns	0.4888	0.4694
Leaf Dry Weight (g)	0.0614	0.0058	1.0375	0.0126*	0.0968	0.0772
Relative Water Content	0.3791	0.0351	0.6530	0.0024**	0.4255	0.2147

\*H0= Data is from Normal distribution. P <0.05 rejects H0

 Table 2. Means of the leaf traits measured in the 'Essex' by 'Forrest' RIL population and parent lines (Essex and Forrest).

	RILs (n=94)				Parents		
Trait	Mean	St Dev.	Skewness	P <w*< th=""><th>Essex mean</th><th>Forrest mean</th></w*<>	Essex mean	Forrest mean	
Leaf Fresh Weight (g)	0.4221	0.1497	0.3599	0.0951ns	0.2478	0.3630	
Leaf Turgid Weight (g)	0.5600	0.1765	0.4603	0.0147*	0.3734	0.5210	
Leaf Dry Weight (g)	0.1666	0.0593	0.6248	0.0281*	0.1124	0.1418	
Relative Water Content	0.6425	0.1195	0.7224	0.0282*	0.5188	0.5833	

\*H0= Data is from Normal distribution. P <0.05 rejects H0

**Table 3.** Pearson correlation coefficients for the leaf traits measured in the 'PI438489B' by 'Hamilton' RIL population and parent lines (PI438489B' and Hamilton).

Traits	Leaf Turgid Weight (g)	Leaf Dry Weight (g)	<b>Relative Water Content</b>
Leaf Fresh Weight (g)	0.7374***	0.7948***	0.7572***
Leaf Turgid Weight (g)		0.9628***	0.3136*
Leaf Dry Weight (g)			0.3913**

\*Significant at P<0.05 probability level. \*\* Significant at P<0.01 probability level. \*\*\* Significant at P<0.001 probability level.

 Table 4. Pearson correlation coefficients for the leaf traits measured in the 'Essex' by 'Forrest' RIL population and parent lines (Essex and Forrest).

Traits	Leaf Turgid Weight (g)	Leaf Dry Weight (g)	Relative Water Content
Leaf Fresh Weight (g)	0.9444***	0.9438***	0.5293***
Leaf Turgid Weight (g)		0.9671***	0.2364*
Leaf Dry Weight (g)			0.3110**

\*Significant at P<0.05 probability level. \*\* Significant at P<0.01 probability level. \*\*\* Significant at P<0.001 probability level.

 Table 5. Comparison of overall means, standard deviations and range of leaf traits measured in the two RIL populations (PI438489B by Hamilton and Essex by Forrest).

Trait	Population	Mean	St Dev	Range
Leaf Fresh Weight (g)	PI438489B x Hamilton	0.1924 <b>b</b> *	0.0219	0.1860-0.7106
	Essex x Forrest	0.4221 <b>a</b>	0.1497	0.0286-0.6030
Leaf Turgid Weight (g)	PI438489B x Hamilton	0.3565 <b>b</b>	0.0299	0.2964-0.8854
	Essex x Forrest	0.5600 <b>a</b>	0.1765	0.0660-0.8780
Leaf Dry Weight (g)	PI438489B x Hamilton	0.0614 <b>b</b>	0.0058	0.0850-0.3084
	Essex x Forrest	0.1666 <b>a</b>	0.0593	0.0092-0.1806
Relative Water Content	PI438489B x Hamilton	0.3791 <b>b</b>	0.0351	0.3668-1.0740
	Essex x Forrest	0.6425 <b>a</b>	0.1195	0.0877-0.8195

\*Levels not connected by the same letter are significantly different at P<0.05 according to Tukey-Kramer test.

#### Statistical Data Analysis

Means, standard errors, skewness and Shapiro-Wilk test for determining the normality of distribution were calculated for PI438489B x Hamilton RIL population. Parental means and standard errors were also computed. Pearson's correlation coef ficients between all trait combinations were determined as well. Results were reported in a Pearson correlation matrix. All analyses were performed by JMP 8.0.2 (SAS Institute Inc., Cary, NC, USA). **Table 6.** The leaf traits QTL identified in the two RIL populations (PI438489B by Hamilton and Essex by Forrest) of soybean. Leaf fresh weight (LFW), leaf turgid weight (LTW), leaf dry weight (LDW), relative water content (RWC). QTL were named according to the Soybean Genetics Committee's recommendations as revised in March 2007. http://soybase.org/resources/QTL.php.

Trait	No.	QTL	Chr./LG	Marker/Interval	Position (cM)	LOD	<b>R</b> <sup>2</sup> (%)
<u>PlxH</u>							
LTW	1	qLTW001	6/C2	ss107912725-ss107925078	39.8–43.3	2.9	0.14
	2	qLTW002	17/D2	ss107913096-ss107916109	7.6–12.8	2.7	0.19
	3	qLTW003	17/D2	ss107929768-ss107913565	15-19.8	4.8	0.25
LDW	1	qLDW001	6/C2	ss107912725-ss107913565	39.4–43.3	2.7	0.15
	2	qLDW002	10/0	ss107930848-ss107917019	56-85	3.3	0.19
RWC	1	qRWC001	2/D1b	ss107912689-ss107929181	39.2–49.5	2.6	0.01
	2	qRWC002	3/N	ss107912938-ss107913615	36.8–38.5	2.7	0.02
	3	qRWC003	18/G	ss107923024-ss107921048	14.8-18.3	2.7	0.01
<u>ExF</u>							
LFW	1	qLFW001	18/G	ATGCGA190-ACCCTC220	92.8–99.8	4.1	0.29
	2	qLFW002	10/0	Satt358–Satt132	0-10.1	3.9	0.38
LTW	1	qLTW001	8/A2	Sat_089—Sat_1 29	3-16.1	2.6	0.24
	2	qLTW002	4/C1	Satt399–Satt195	0–13	2.6	0.13
	3	qLTW003	18/G	Satt324–Satt356	50.3–59.9	4.6	0.40
	4	qLTW004	18/G	Satt570–Satt122	70.8–73.8	5.3	0.48
LDW	1	qLDW001	4/C1	Satt399–Satt195	0–13	2.7	0.14
	2	qLDW002	18/G	Satt324–Satt356	49.5–60.3	4.4	0.37
	3	qLDW003	18/G	Satt570–Satt122	70.8–77.5	3.8	0.36
RWC	1	qRWC001	11/B1	Satt546–Satt604	54.3–73	2.6	0.18

#### **Results and Discussion**

#### **Distribution of Leaf Traits**

The frequency distribution of 'PI438489B' by 'Hamilton' population for 3 out of 4 studied leaf traits (leaf fresh weight, leaf dry weight, and relative water content) departed significantly from normality (P<0.05). Shapiro-Wilk test revealed that only values of leaf turgid weight were normally distributed (Table 1 and Figure 1).

#### **Correlation Coefficients of Leaf Traits**

Correlation coefficients were estimated for each pair-wise leaf trait combination for PI438489Bx Hamilton (Table 2). All traits were found significantly (P<0.05) correlated. The highest correlation was estimated for leaf turgid weight and leaf dry weight ( $r=0.9628^{***}$ ) and the lowest but statistically significant between leaf turgid weight and relative water content ( $r=0.3136^{*}$ ).

#### QTL Analysis of Leaf Traits

Relative water content (RWC) is an important indicator on

49

how plants withstand drought and become tolerant to WD. In this study, we genetically mapped QTL for leaf fresh weight (LFW), leaf turgid weight (LTW), leaf dry weight (LDW), and leaf relative water content (RWC) using the soybean 'Essex' by 'Forrest' RIL population (ExF, n = 94). The phenotypic data for LFW, LDW, LTW, and RWC was compared against the updated ExF genetic linkage map (Kassem et al., 2006) to map QTL for these traits using the CIM of WinQTL Cartographer (Wang et al., 2004).

A total of 8 QTL for LTW, LDW, and RWC were identified and mapped in the 'PI 438489B' by 'Hamilton' RIL population. Three QTL were identified for LTW. The first QTL (qLTW001) was located on chromosome 6 (LG C2), and the second (qLTW002) and third (qLTW003) were located on chromosome 17 (LG D2) (Figure 3, Table 6). These QTL spanned 3.5 cM, 5.2 cM, and 4.8 cM, and have peak LOD scores of 2.9, 2.7, and 4.8, respectively (Figure 3, Table 6). Two QTL were identified for LDW. The first QTL (qLDW001) was located on chromosome 6 (LG C2), and the second (qLTW002) was located on chromosome 10 (LG O). These two QTL spanned 3.9 cM and 29 cM, and have peak LOD scores of 2.7 and 3.3, respectively (Figure 3, Table 6). Three QTL were identified for RWC. The first QTL (qRWC001) was located on chromosome 2 (LG D1b), the second QTL (qRWC002) was located on chromosome 3 (LG N), and the third QTL (qRWC003) was located on chromosome 18 (LG G). these QTL spanned 10.3 cM, 1.7 cM, and 3.5 cM and have peak LOD scores of 2.6, 2.7,



**Figure 1.** Frequency distribution of the leaf traits (leaf fresh weight, leaf turgid, leaf dry weight, and relative water content) in the 'PI438489B' by 'Hamilton' RIL population compared to their parental lines.



**Figure 2.** Frequency distribution of the leaf traits (leaf fresh weight, leaf turgid, leaf dry weight, and relative water content) in the 'Essex' by 'Forrest' RIL population compared to their parental lines.

Chr\_2/LG\_D1b

Chr\_3/LG\_N



Figure 3. The locations of the leaf traits QTL identified in the PIxH RIL population.



**C1** 

G

0

Figure 4. The locations of the leaf traits QTL identified in the ExF RIL population.

and 2.7, respectively (Figure 3, Table 6).

A2

**B1** 

In the 'Essex' by 'Forrest' RIL population, a total of 10 QTL for LFW, LTW, LDW, and RWC were identified and mapped on 5 different LGs. Two QTL were identified for LFW: qLFW001 on chromosome 18 (LG G) and qLFW002 on chromosome 10 (LG O) (Figure 3, Table 6). These QTL spanned 7 cM and 10.1 cM and have peak LOD scores of 4.1 and 3.9, respectively (Figure 3, Table 6). Four QTL were identified for LTW: qLTW001 on chromosome 8 (LG A2), qLTW002 on chromosome 4 (LG C1), and both qLTW003 and qLTW004 on chromosome 18 (LG G). These QTL spanned 13.1 cM, 13 cM, 9.6 cM, 3 cM and had peak LOD scores of 2.6, 4.6, and 5.3, respectively (Figure 3, Table 6). Three QTL were identified for LDW: qLDW001 on chromosome 4 (LG C1) and both qLDW002 and qLDW003 on chromosome 18 (LG G). These QTL spanned 13 cM, 10.8 cM, 6.7 cM and had peak LOD scores of 2.7, 4.4, and 3.8, respectively (Figure 3, Table 6). In this ExF RIL population, one QTL (gRWC001) was identified for RWC on chromosome 11 (LG B1). The QTL spanned 18.7 cM and had a peak LOD score of 2.6 (Figure 3, Table 6).

Clusters of leaf QTL were observed in both populations. In the PIxH RIL population, a cluster of LTW QTL was observed on chromosome 17 (LG D2) in the 7.6 to 19.8 cM interval. Similarly, a cluster of LFW, LTW, and LDW QTL was observed on chromosome 18 (LG G) in an interval spanning approximately 50.3 cM (49.5–99.8 cM) which is in agreement with previous findings of QTL clustering on the soybean genome (Meksem et al., 1999; Ross-Ibarra, 2005; Zhao-Ming et al., 2011). Interestingly, a QTL for leaf area was identified and mapped within this same region using the PI416937 by Young RIL population (Mian et al., 1998). Similarly, 3 other QTL for leaf area were mapped on chromosome 8 (LG A2) using the PI416937 by Young RIL population. The first QTL fits within the same region of qLTW001 reported here, the second and third QTL were mapped 4 cM and 15 cM, respectively from qLTW001 (Mian et al., 1998; SoyBase, 2011). On chromosome 6 (LG C2), several QTL for leaf area were identified using the PI416937 by Young (Mian et al., 1998; Soy-Base, 2011) and Archer by Minsoy (Mansur et al., 1998; Soy-Base, 2011) and Archer by Minsoy (Mansur et al., 1993) RIL populations. These QTL were mapped 53.3 cM to 108 cM from qLTW001 and qLDW001 reported in this study (Mansur et al., 1993; Mian et al., 1998; SoyBase, 2011). The RWC QTL found here maybe important for breeding programs that aim to develop soybean for water deficit tolerance (WDT).

#### **Acknowledgements**

The authors would like to thank Mrs. Pam Ratcliff and the crew of undergrad students for taking care of the plants in the greenhouse and the field. We also thank Dr. Silvia Cianzio of Iowa State University for providing the PIxH population.

#### References

Adams NR (1995) Detection of the effects of phytoestrogens on sheep and cattle. J Anim Sci. 73 (5): 1509–1515.

Adlercreutz H (2002) Phytoestrogens and breast cancer. J Steroid Bio-

chem Mol Biol. 83 (1-5): 113-118.

- Anderson JW, Johnstone BM, Cook-Newell ME (1995) Meta-analysis of the effects of soy protein intake on serum lipids. N Engl J Med. 333 (5): 276–282.
- Anthony MS, Clarkson TB, Hughes CL Jr, Morgan TM, Burke GL (1996) Soybean isoflavones improve cardiovascular risk factors without affecting the reproductive system of peripubertal rhesus monkeys. J Nutr. 126 (1): 43–50.
- Banz WJ, Williams MP, Lightfoot DA, Winters TA (1999) The effects of soy protein and soy phytoestrogens on symptoms associated with cardiovascular disease in rats. J Med Food. 2 (3-4): 271–273.
- Barnes S, Grubbs C, Setchell KDR, Carlson J (1990) Soybeans inhibit mammary tumors in rat models of breast cancer. In: Pariza MW, Aeschbacher H-U, Fleton JS, Sato S, eds. Mutagens and Carcinogens in the Diet. New York, NY: Wiley-Liss; pp. 239–253.
- Barnes S, Peterson TG, Grubbs C, Setchell KDR (1994) Potential role of dietary isoflavones in the prevention of cancer. In: Jacobs MM, ed. Diet and Cancer: Markers, Prevention, and Treatment. New York, NY: Plenum Press. pp. 135–147.
- Bringe NA (2001) High beta-conglycinin products and their use. US patent 6 171 640.
- Cassidy A, Bingham S, Setchell KDR (1994) Biological effects of a diet of soy protein rich in isoflavones on the menstrual cycle of premenopausal women. Am J Clin Nutr. 60 (3): 333–340.
- Cassidy JM, Baird WM, Chang CJ (1990) Natural products as a source of potential cancer chemotherapeutic and chemopreventive agents. J Nat Prod. 53 (1): 23–41.
- Eldridge AC, Kwolek WF. Soybean isoflavones: effect of environment and variety on composition. J Agric Food Chem. 1983; 31 (2): 394– 396.
- Fotsis T, Pepper M, Adlercreutz H, et al. Genistein, a dietary-derived inhibitor of in vitro angiogenesis. Proc Natl Acad Sci USA. 1993; 90 (7): 2690–2694.
- Frova C, P. Krajewski, N di Fonzo, M. Villa, and M Sari-Gorla (1999) Genetic analysis of drought tolerance in maize by molecular markers. I. Yield components. Theor Appl Genet 99:280–288.
- Gibson L and G Benson (2005) Origin, History, and Uses of Soybean (Glycine max). Iowa State University, Department of Agronomy.
- Greer-Baney JA, Banz WJ, Lightfoot DA, Winters TA. Dietary soy protein and soy isoflavones: a histological examination of reproductive tissues in rats. J Med Food. 1999; 2 (3-4): 247–249.
- Huang MT, Ferraro T. Phenolic compounds in food and cancer prevention. In: Huang MT, Ho CT, Lee CY, eds. Phenolic Compounds in Food and Their Effects onHealth II.Washington, DC: American Chemical Society Press; 1992: 8–34.
- Hunt ER, BN Rock, and PS Nobel (1987) Measurement of Leaf Relative Water Content by Infrared Reflectance. Remote Sensing of Environment 22: 429–435.
- Jing Y, Nakaya K, Han R. Differentiation of promyelocytic leukemia cells HL-60 induced by daidzein in vitro and in vivo. Anticancer Res. 1993; 13 (4): 1049–1054.
- Kassem MA (2003) Genetic linkage map and QTL mapping in soybean. Ph.D. thesis, SIUC, Carbondale, IL, USA.
- Kassem MA, Meksem K, Iqbal MJ, Njiti VN, Banz WJ, Winters TA, Wood A, Lightfoot DA. Definition of Soybean Genomic Regions That Control Seed Phytoestrogen Amounts. J. Biomed & Biotech 2004, 1 (1):

52-60.

- Kazi S (2005) Identification of twenty three loci conditioning agronomic traits in recombinant inbred and near isogeneic soybean lines derived from Flyer by Hartwig. MS Thesis SIUC Carbondale, IL, USA.
- Kazi S, Njiti VN, Doubler TW, Yuan J, Iqbal MJ, Cianzio S, Lightfoot DA (2007) Registration of the Flyer by Hartwig recombinant inbred line mapping population. J Plant Regis 1:175–178.
- Keurentjes JJB, M Koornneef M, and D Vreugdenhil (2008) Quantitative genetics in the age of moics. Curr Opinion in Plant Biology 11: 123–128.
- Knight DC and JA Eden (1996) A review of the clinical effects of phytoestrogens. Obstet Gynecol 87 (5): 897–904.
- Mansur LM, KG Lark, H Kross, and A Oliveira (1993) Interval mapping of quantitative trait loci for reproductive, morphological, and seed traits of soybean (Glycine max L.). Theor Appl Genet 86 (8): 907–913.
- Meksem K, TW Doubler, K Chancharoenchai, VN Njiti, SJC Chang, PR Arelli, PE Cregan, LE Gray, PT Gibson, and DA Lightfoot (1999) Clustering among loci underlying soybean resistance to Fusarium solani, SDS and SCN in near-isogenic lines. Theor Appl Genet 99:1131–1142.
- Meksem K, VN Njiti, WJ Banz, MJ Iqbal, MA Kassem, DL Hyten, J Yuang, TA Winters, and DA Lightfoot. Genomic regions that underlie soybean seed isoflavone content. J. Biomed. & Biotech. 2001 1 (1): 38-44.
- Messina M, Barnes S. The role of soy products in reducing risk of cancer. J Natl Cancer Inst. 1991; 83 (8): 541–546.
- Mian MAR, DA Ashley, and HR Boerma (1998) An additional QTL for water use efficiency in soybean. Crop Science 38: 390–393.
- Morrissey C, Watson RW. Phytoestrogens and prostate cancer. Curr Drug Targets. 2003; 4 (3): 231–241.
- Munro IC, Harwood M, Hlywka JJ, et al. Soy isoflavones: a safety review. Nutr Rev. 2003; 61 (1): 1–33.
- Peterson TG, Barnes S. Genistein and biochanin A inhibit the growth of human prostate cancer skin tumor promotion, Ill. Inhibitory effects of isoflavonoids fromWisteria brachybotrys on Epstein-Barr virus activation. J Nat Prod. 1988; 51(6):1266–1270.
- Ross-Ibarra J (2005) Quantitative trait loci and the study of plant domestication. Genetica 123: 197–204.
- Smart R and GE Bingham (1974) Rapid estimates of relative water content. Plant Physiol. 53: 258–260.
- Soy Stats (2010) http://www.soystats.com/2010/Default-frames.htm. Last accessed in December 2010.
- Teulat B, N Zoumarou-Wallis, B Rotter, M Ben Salem, H Bahri, and D This (2003) QTL for relative water content in field-grown barley and their stability across Mediterranean environments. Theor Appl Genet 108: 181–188.
- Teulat B, P Monneveux, J Wery, C Borries, I Souyris, A Charrier, and D This (1997) Relationships between relative water content and growth parameters under water stress in Barley: A QTL study. New Phytologist 137: 99–107.
- United Soybean Board (2007) The Soy Products Guide, USB. www. unitedsoybean.org.
- Zhao-Ming QI, Y Sun, J Wang, D Zhang, C Liu, G Hu, and QS Chen (2011) Agricultural Sciences in China 10(3): 327-334.