

Simulated Microgravity and Ionizing Radiation changing *Escherichia coli*'s Growth Dynamics

Parker Mann*¹, Mitchell Villafania¹, Collin Topolski², Dr. Hugo Castillo^{3#}

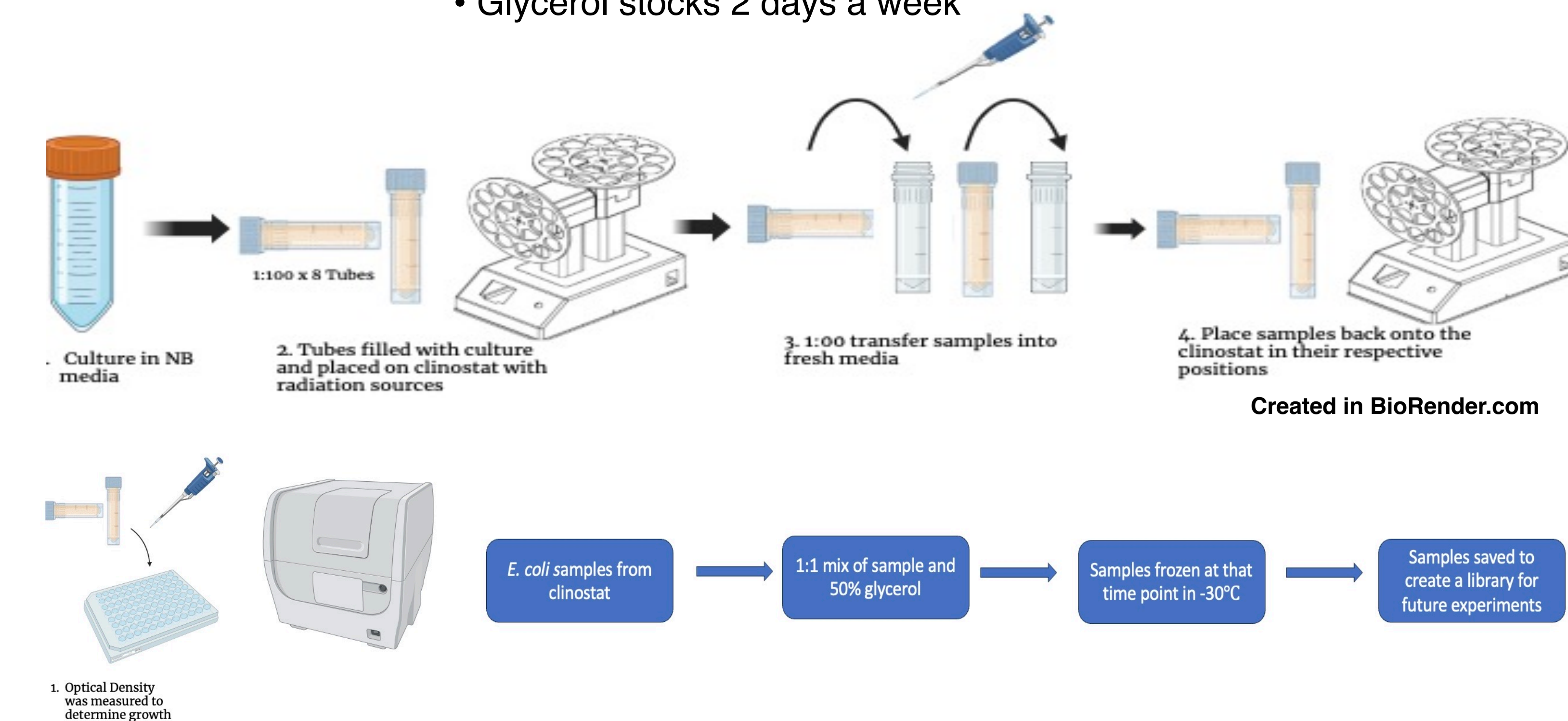
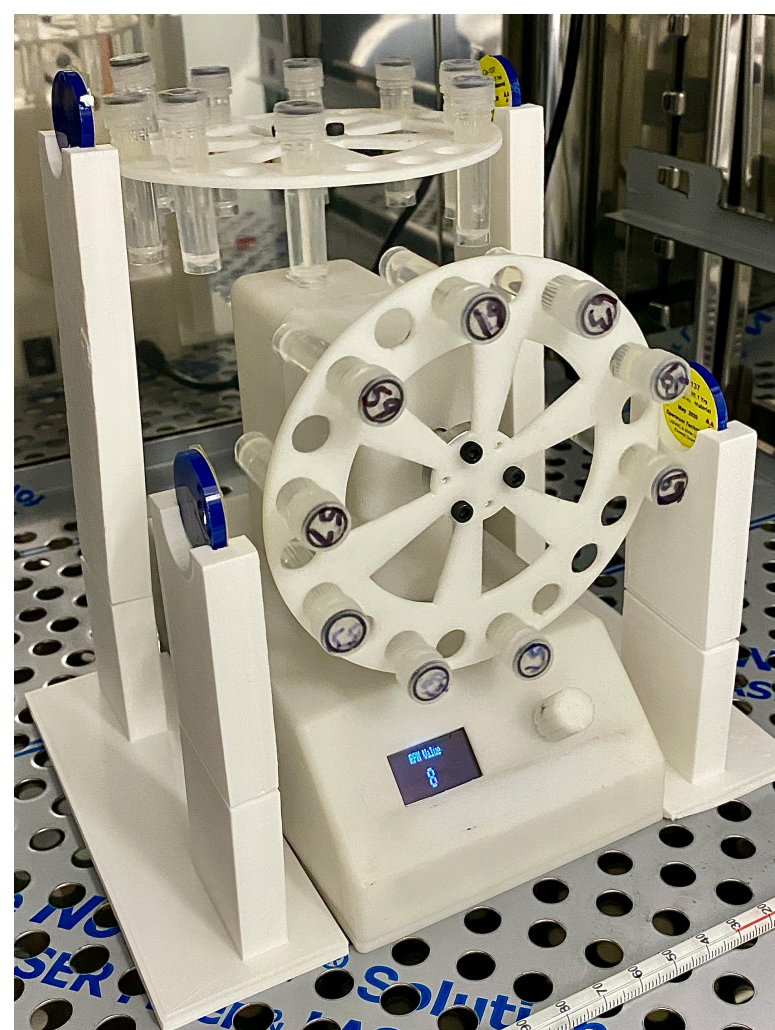
¹College of Arts and Sciences, Embry-Riddle Aeronautical University, Daytona Beach, FL 32114; ²College of Engineering, Embry-Riddle Aeronautical University; ³Department of Human Factors and Behavioral Neurobiology, Embry-Riddle Aeronautical University *Presenting Author # Faculty Advisor

With the push for increased human space exploration, we must explore the effects of the extreme environment of space to ensure mission success. Research has shown that the increased exposure to space stressors such as microgravity (μG) and increased low-dose ionizing radiation can result in significant physiological and cellular responses. These responses have been observed to include transient and long-term impacts on astronaut microbiomes that alters immune system function. In this study we explored how the antagonistic and synergistic activities of bacteria responded to simulated microgravity (SMG) environments using *Escherichia coli*. We measured growth rates during and post prolonged exposure to radiation and simulated microgravity.

Experimental Design

Initial Exposure Experiment Design

- *Escherichia coli* (Intestinal Microflora)
 - Gram Negative
 - Rod Shaped
- Low-dose ionizing radiation
 - 4 Cs-137 sources
 - $10\mu\text{Ci}$ activity
 - Gamma Radiation
 - Sources placed on 3D printed stands
 - $\sim 1.7\mu\text{SV}$ per hour (unshielded)
 - 3 water tanks were used for shielding inside the incubator
- Simulated Microgravity and Gravity conditions
 - 3D printed EagleStat
- Prolonged Exposure cultures were grown for 28 days
 - Daily transfers 5 days a week
 - Glycerol stocks 2 days a week



Post Exposure Experiment Design

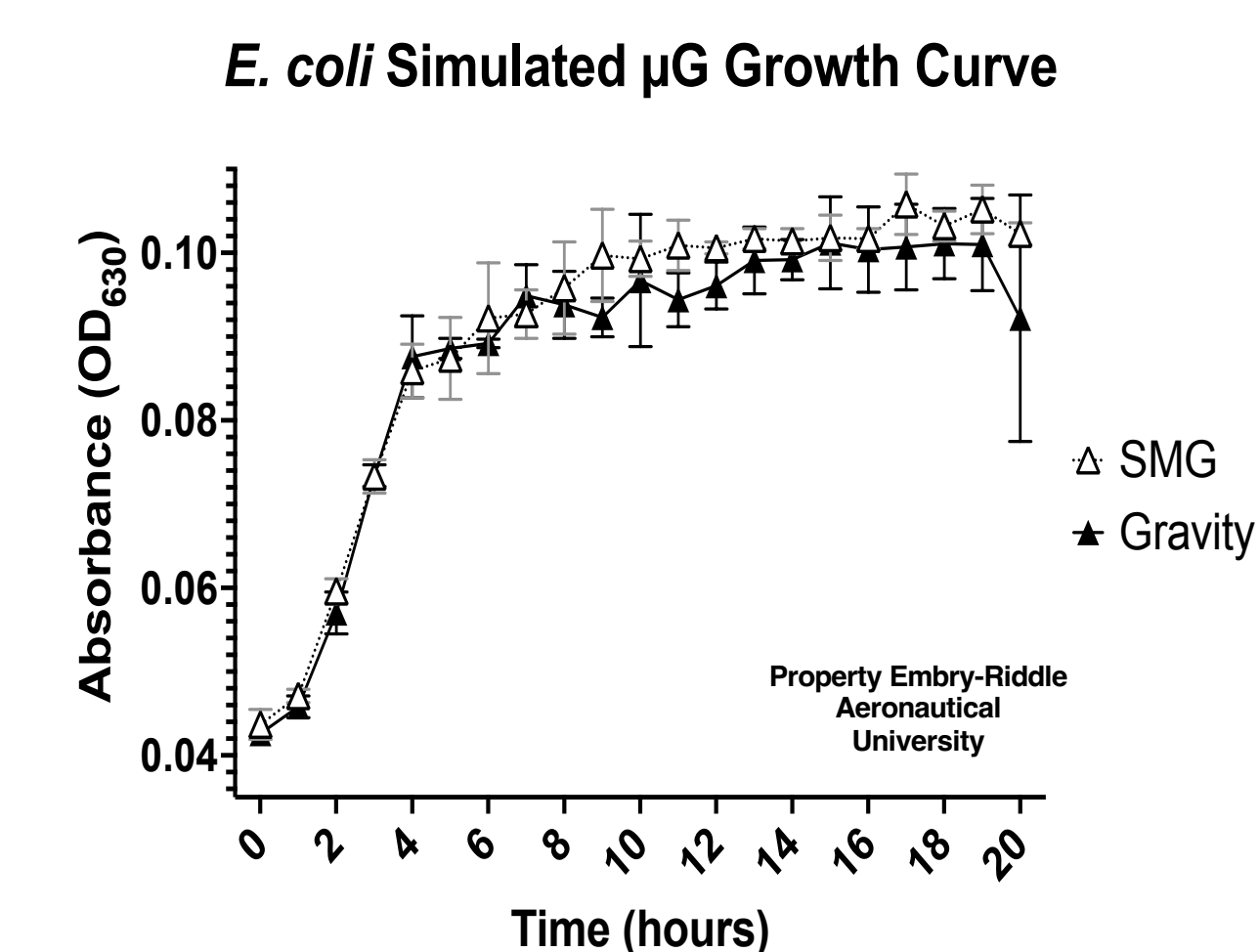
- *Escherichia coli* samples from initial exposure
- Samples grown on 24-well plate with fresh media
- Samples grown in plate reader at 30°C
- Optical Density (OD) measured every hour for 15 hours
- No Radiation or Microgravity

Image Credit: ERAU SML

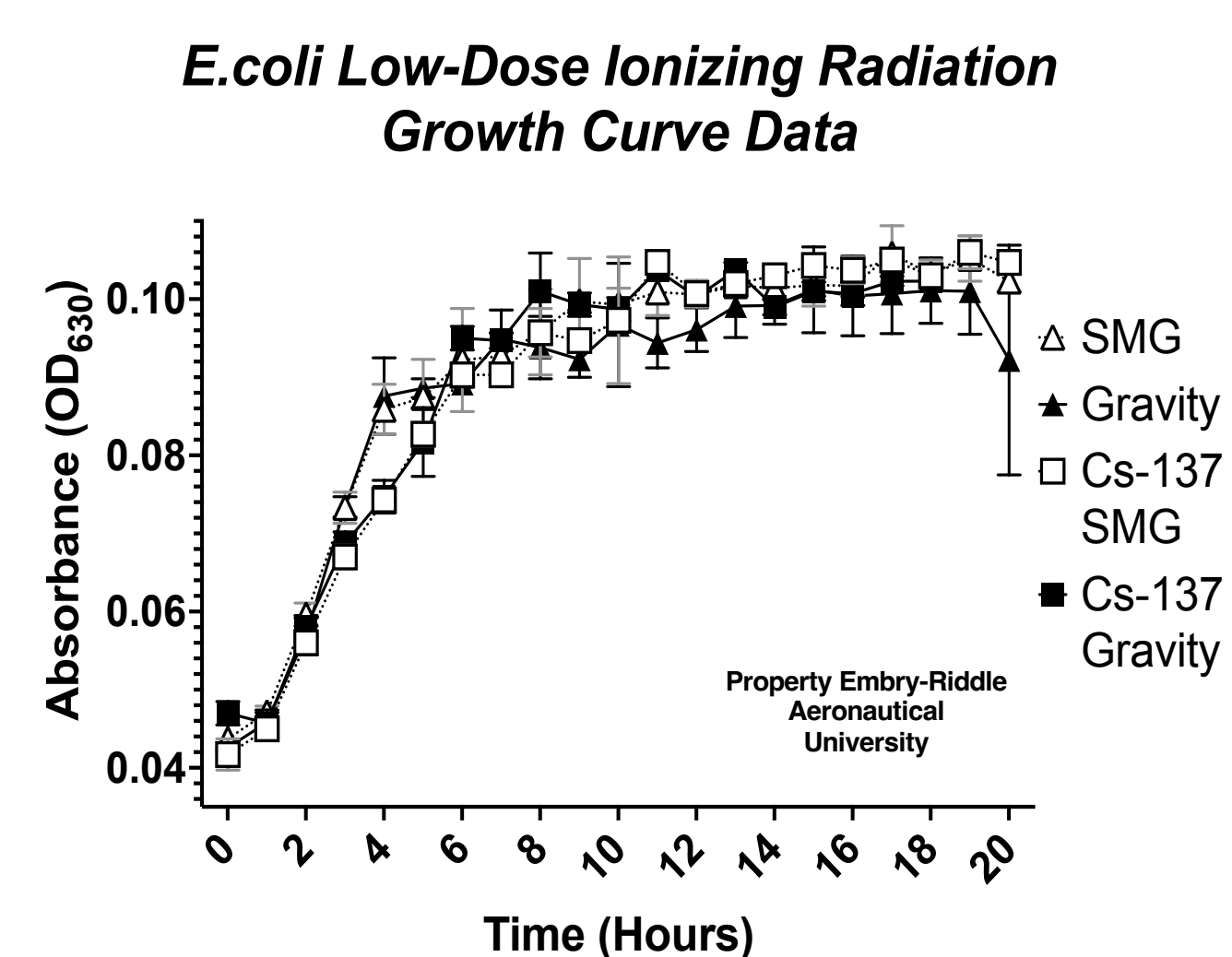
Image Credit: ERAU SML

Growth Results

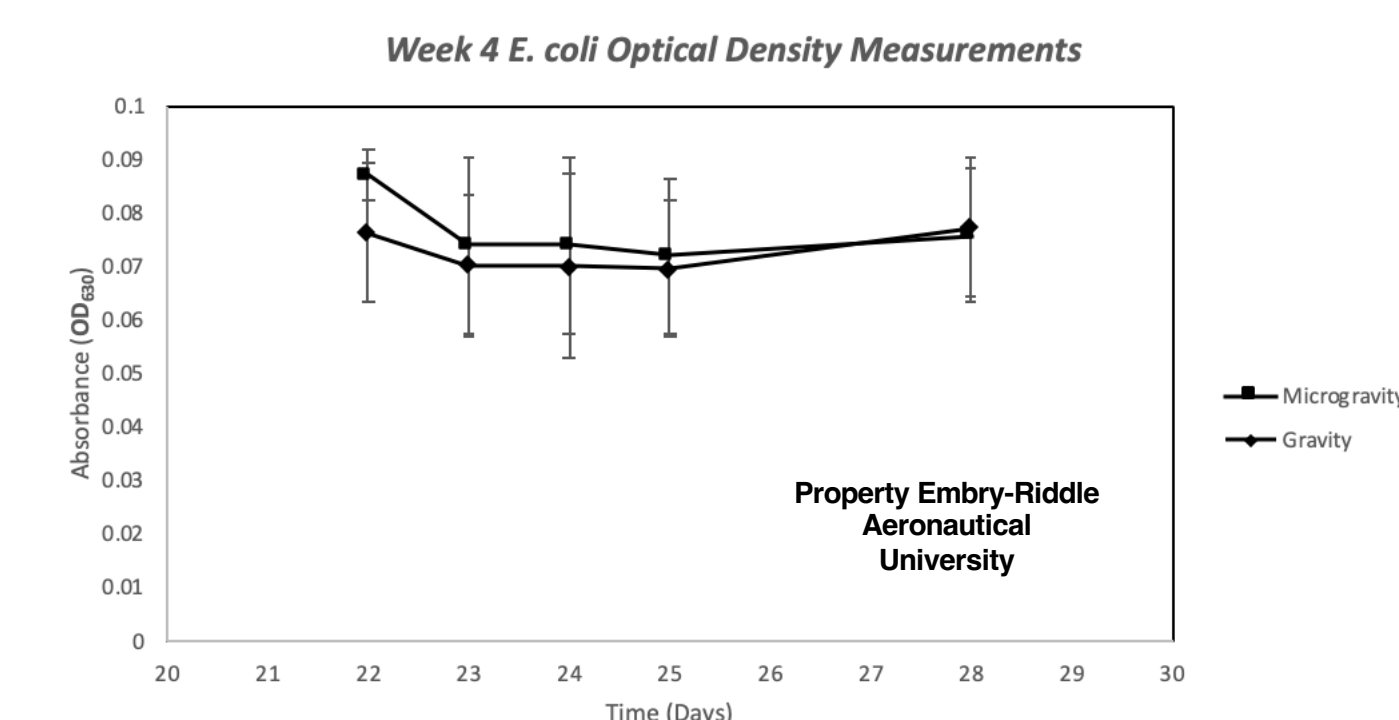
- Slight increase of cells under simulated μG compared to gravity analog



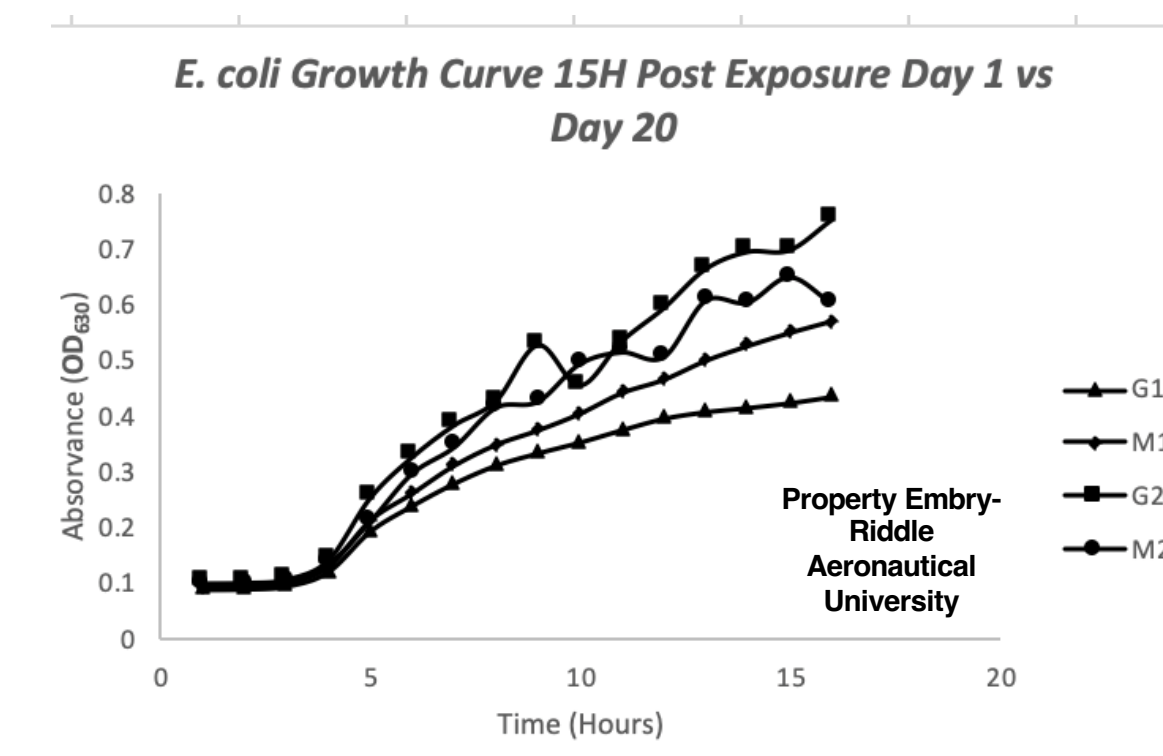
- Initial Low-dose ionizing radiation growth curve test
- $\sim 0.5 \text{ mSv/Hr}$



- After 4 weeks of exposure no significant difference between μG and gravity analog



- Increase of cells on day 20 compared to day 1 in both μG and gravity analog



References

- Kuehnast, T., Abbott, C., Pausan, M. R., Pearce, D. A., Moissl-Eichinger, C., & Mahner, A. (2022). The crewed journey to Mars and its implications for the human microbiome. *Microbiome*, 10(1), 1–14. <https://doi.org/10.1186/s40168-021-01222-7>
- Tesei, D., Jewczynko, A., Lynch, A. M., & Urbaniak, C. (2022). Understanding the Complexities and Changes of the Astronaut Microbiome for Successful Long-Duration Space Missions. *Life*, 12(4), 1–47. <https://doi.org/10.3390/life12040495>
- Voorhies, A. A., Mark Ott, C., Mehta, S., Pierson, D. L., Crucian, B. E., Feiveson, A., Oubre, C. M., Torralba, M., Moncera, K., Zhang, Y., Zurek, E., & Lorenzi, H. A. (2019). Study of the impact of long-duration space missions at the International Space Station on the astronaut microbiome. *Scientific Reports*, 9(1), 1–17. <https://doi.org/10.1038/s41598-019-46303-8>

Ongoing Research

- Long Duration SMG and Low-Dose Ionizing Radiation study on other models
- Further evaluation of response to long duration with DNA and RNA sequencing and biofilm assays.

Acknowledgements

Thank you to Dr. Castillo and the Space Microbiology Lab team for your continued support and encouragement.