

Commentary

Tryptophan and Membrane Mobility as Conditioners and Brokers of Gut–Brain Axis in Depression

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Abstract: The aim of this brief narrative commentary is to discuss some aspects involved in depression. It is increasingly evident that the phenomenon of mood disorders, despite its unequivocal genetic origin, slips into a multifactorial set of biochemical and molecular events that involve the whole organism. A vast literature has provided evidence that recognizes changes in serotonergic neurotransmission in the pathophysiology of depression. In addition, an increased arachidonic acid/omega-3 fatty acid ratio, which confers to mammalian cell membranes their fluidity, is associated with the depressive state. The combination of the excessive expression of kinurenine and the increased fluidity of the membrane has never been considered in the meaning of a simultaneous effect in the determinism of the depressive condition. Furthermore, various evidence supports the relationship between intestinal microbiota and depression and confirms alterations in the microbiota in depressive pathology.

Keywords: tryptophan; serotonin; depression; membrane fluidity; arachidonic acid; intestinal microbiota

1. Introduction

A paper published in *The Lancet* in 1969 about tryptophan stated, “Psychic depression may result from deficiency of brain serotonin. It is suggested that in depression the production of tryptophane pyrrolase by the liver is stimulated by raised blood-corticosteroid levels. As a result the metabolism of tryptophane is shunted away from serotonin production, and towards kynurenine production” [1].

In fact, this article highlighted how the neurotropic activity of kinurenine, by modifying the bioavailability of tryptophan, favored the reduction of serotonin (5-hydroxytryptamine, 5-HT), opening the door to states of anxiety, psychosis, and cognitive decline—conditions all associated with depression.

In 1973, other authors conducted an experiment in depressed and nondepressed subjects claiming that the data obtained did not support the increase in tryptophan metabolism toward the kinurenine pathway in depression [2]. Despite the contrast of these data, probably attributable to the experimental modalities, the evidence remains that, in the depressive condition, tryptophan triggers a decrease in 5-HT levels.

Over time, up to the present day, the concept of the centrality of tryptophan in the determinism of the reduction of 5-HT, due to the greater transformation of the metabolic pathway of tryptophan into kinurenine, has become increasingly popular [3]. In particular, stress and inflammation, as well as proinflammatory cytokines, can induce the activation of indoleamine 2,3-dioxygenase (IDO) (an enzyme involved in the catabolism of tryptophan to kynurenine), and higher levels of kynurenine have been linked with a depressive condition [4,5] (Figure 1). In addition, genetic variants of 5-HT receptors, as well as tryptophan hydroxylase, have been associated with a higher risk of depression [6].

Furthermore, proinflammatory cytokines increase monoamine reuptake by further reducing 5-HT levels [7–9].

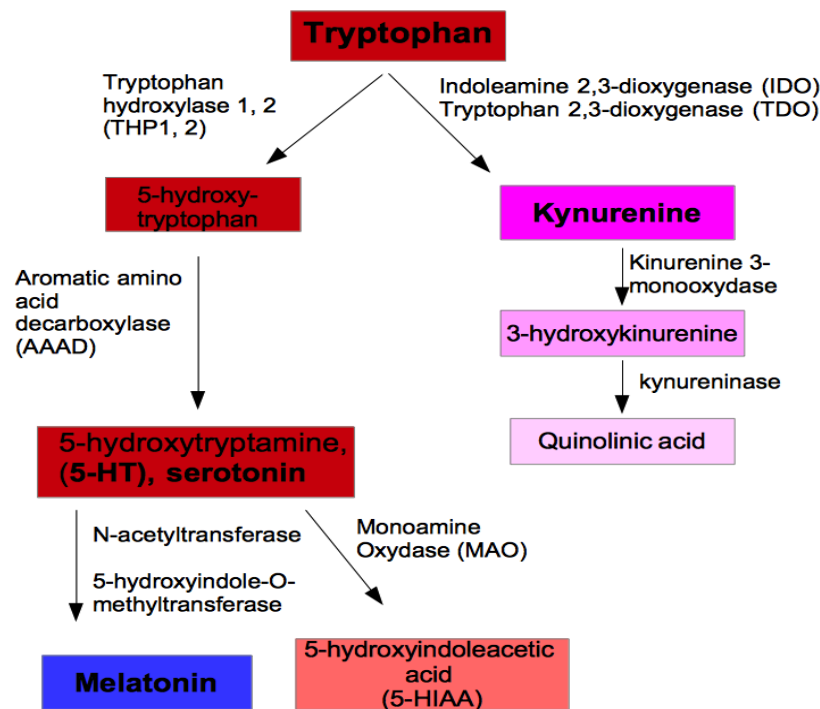


Figure 1. Metabolic pathways of tryptophan.

In the 1970s and 1980s, a further element to the evaluation of the reduced presence of 5-HT in the depressive condition was added by some authors with reference to the similarity between neurons and platelets in relation to finding a reduced amount of 5-HT in the neurons and the platelets in the depressive condition [10–17].

In more recent years, the results obtained by other authors through blood platelet fatty acids, in the interpretation of an artificial neural network, have enabled the possibility of carrying out a precise diagnostic evaluation of the two main psychopathologies, namely major depression (MD) and bipolar disorder (BD) [18–20].

Whereas 5-HT is mostly known for its role in mood, anxiety, psychosis, or memory in the central nervous system (CNS), more than 95% of total body 5-HT is present in the periphery. In addition, the majority of peripheral 5-HT is stored in platelets. On the other hand, platelets do not synthesize 5-HT, but they adsorb it from plasma through the 5-HT transporter and release it during their activation [21]. Actually, the function of 5-HT in platelets is not clear yet; studies suggest that it is important for the serotonylation of the proteins necessary in platelet aggregation [22].

A recent study revealed platelet contributions to inflammation in reactions involving antigen–antibody complexes [23]. Furthermore, some studies have found a significant correlation between the abnormal membrane fluidity of platelets and the severity of dementia [24–26].

Platelets and neurons are unquestionably different cells, however, they share common characteristics in molecular organization and in protein composition [27]. In particular, various proteins are typically expressed in both neurons and platelets, and circulating platelets have been proposed as an alternative model to investigate neuronal dysfunctions and as an accessible peripheral biomarker to monitor the onset and the progression of neurological diseases. In this respect, some studies have reported platelet alterations in autism spectrum disorders [28].

The hypothesis that is formulated refers to the possibility of using the platelet as a diagnostic element in psychopathology [29] and in accordance with Heron et al. [30]. The focus is on the role of membrane fluidity in the management of 5-HT uptake. The cell membrane is known to live in a

state of continuous mobility. Its fluidity is affected by the ratio of phospholipids to free cholesterol. In particular, long chain fatty acids, specifically arachidonic acid (AA) and docosahexaenoic acid (DHA), are integral components of neural membrane phospholipids and are extremely important for the maintenance of the optimum membrane fluidity of neurons [31]. There are several potential sources of AA in the brain, and an alteration in their components cannot only influence crucial intracellular and intercellular signaling but also alter many membrane physical properties, such as fluidity, phase transition temperature, bilayer thickness, and lateral domains, and therefore, the activity of membrane-dependent proteins [31,32]. Alterations in membrane proteins have been documented in depressed patients [33]. Walsh et al. reported an increase in the expression of the platelet adhesion receptor glycoprotein-Ib in depressed subjects [34]. In addition, Piletz et al. showed an increased P-selectin platelet content in depressed patients [35]. In particular, an excessive phospholipase-A 2 (PLA 2) activity could disrupt membrane fluidity, composition, and consequently, the activity of membrane-dependent proteins [33]. Membrane fluidity among depressed patients has been reported to be significantly involved, and brain levels of DHA have been shown to be reduced [36].

It has been hypothesized that membrane viscosity influences the reactivity of circulating platelets. The viscosity of the platelet membrane, the dislocation of the membrane components, and the distortion of the lipid–protein interactions could give rise to a signal mediated by an altered fluidity of the platelet membrane [37].

Therefore, the differential lipid fluidity of the membrane represents a crucial node of 5-HT signaling. Indeed, the differential accessibility of 5-HT receptors decreases with lipid fluidity [30]. Among the various subtypes of 5-HT receptors, the type-1A receptor serves as an important target in the development of therapeutic agents for neuropsychiatric disorders, including depression. In particular, it has been observed that the mobility of the 5-HT 1A receptor and the dynamics of the cell surface also depend on the receptor's interaction with G proteins [38].

A correlation between platelet fatty acid composition in humans (isolated from venous and arterial blood) and pigs (isolated from venous blood and the brain) has been performed [39]. Experiments have been conducted on the transfer of AA between platelets and the brain, in pigs compared to humans, where it has not been possible to investigate the human brain [36–38]; these studies have shown, through a mathematical extrapolation of the data, the increased concentration of AA [39] as the reason for the increased fluidity of the membrane and the reduction in the accessibility of 5-HT, as is also reported by Green et al. [40]. In particular, the concentration of AA in brain tissue from various areas of an animal model of depression has been shown to be higher than in the corresponding brain areas of the controls [40].

Under conditions of an alteration in the fluidity of the membrane due to an increase in AA, as happens in the depressive state, 5-HT would not be captured by the platelets and, moreover, could contribute to bone and cardiovascular damage [31,41,42].

In conclusion, there could be two mechanisms that induce the reduction in 5-HT in psychopathological subjects, both centrally and peripherally: on the one hand, the excessive hyper-expression of the kinurenine pathway, and on the other, the substantial modification of the mobility of the membrane in the direction of increased fluidity due to an excessive concentration of AA [31] (Figure 2).

The combination of the excessive expression of kinurenine and the increased fluidity of the membrane has never been considered in the meaning of a simultaneous effect in the determinism of depressive conditions, such as MD and BD, and in stress.

This observation appears to be relevant for the treatment of depressive psychopathology as well as for further cardiovascular and bone damage [41,42].

Therefore, how to intervene?

Evidence supports the relationship between the microbiota and depression, confirmed by research conducted on alterations of the microbiota in depressive pathology [8,43–45]. In particular, recent studies have shown how and to what extent the intestinal microbiota is involved in the regulation

of functional aspects of the brain and behavior, with particular regard to alterations in tryptophan metabolism and the serotonergic system. Research has been focused on the bidirectional communication between the brain and gut, looking at 5-HT as a molecular signaling factor in both the brain and the enteric nervous system, where 5-HT regulates functions such as secretion, vasodilation, peristalsis, and the perception of pain. The gut microbiota is a critical element of this axis and performs its function not only locally but on several levels. Alterations of 5-HT occur in disorders of the microbiota–gut–brain axis and in communication between the districts [8,46]. This axis explains the mood alterations in irritable bowel syndrome (IBS) and suggests that IBS pathogenesis is partly related to the dysfunctional control of 5-HT production by the gut microbiota. Furthermore, the development of IBS has been shown to be connected to tryptophan depletion. Mechanisms supporting the influence of the microbiota on the availability of tryptophan and therefore of 5-HT are possible through the enzymatic activities responsible for the degradation of tryptophan along the kinurenine pathway [47]. This pathway is involved in a reduction in the kinurenine/tryptophan ratio in germ-free mice (mice born and raised in the absence of microbiota). Furthermore, the alterations of IDO/tryptophan 2,3-dioxygenase (TDO) activity seem to have relevant clinical implications in various pathological conditions [48].

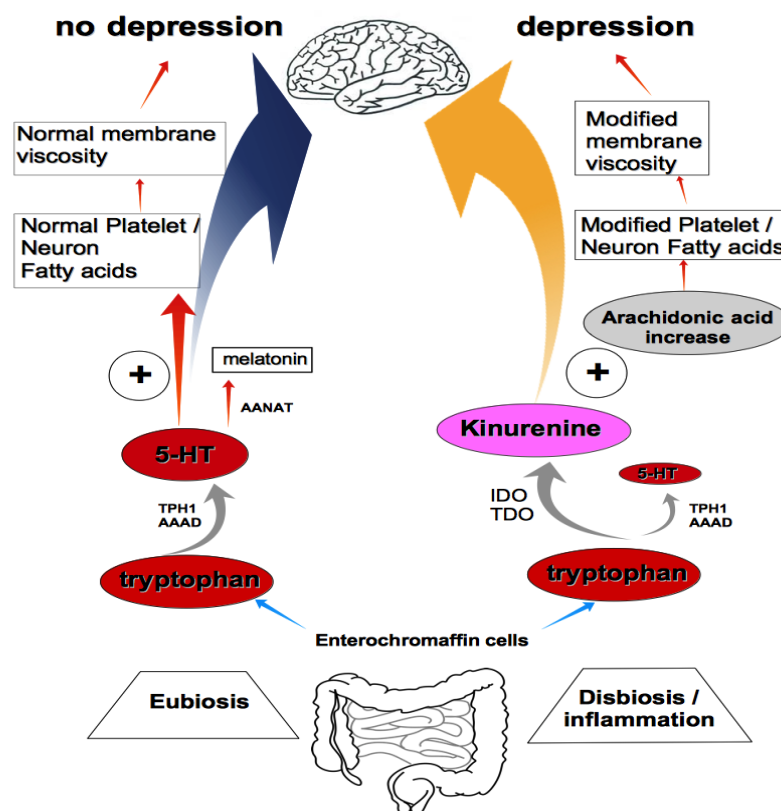


Figure 2. Tryptophan pathway from gut to brain. On the left, a schematic description of the metabolic pathway of tryptophan from enterochromaffin cells to platelets is shown under normal conditions. To the right, the same hypothetical pathway is modified in conditions of intestinal inflammation or dysbiosis (loss of stability of gut microbiota) and under an alteration in the fluidity of the membrane due to the excessive presence of arachidonic acid. In this case, 5-HT would not be captured by platelets and would remain free, thus contributing to possible cardiovascular and bone damage. In addition, the enzymes IDO and TDO metabolize tryptophan to kinurenine and, furthermore, stop tryptophan from entering the brain. AAAD, aromatic L-amino acid decarboxylase; AANAT, N-acetyl transferase; IDO, indoleamine 2,3-dioxygenase; TDO, tryptophan 2,3-dioxygenase; 5-HT, 5-hydroxytryptamine, serotonin; TPH1, tryptophan hydroxylase 1.

Interestingly, the conversion of tryptophan to 5-HT occurs predominantly in the intestine, in the enterochromaffin cells of the intestinal mucosa. As previously mentioned, the vast majority of 5-HT is not found in the CNS but in the gastrointestinal tract [49]. The levels of 5-HT vary depending on plasma tryptophan levels and on the current availability of tryptophan from nutrition [50].

Several species of enteric microorganisms, such as *Lactobacillus*, *Bifidobacterium*, *Escherichia*, *Bacillus*, and *Saccharomyces*, are able to produce neurotransmitters, including 5-HT. Unlike 5-HT, tryptophan produced by gut microbiota is permeable with respect to the blood–brain barrier and produces positive effects on mood through an increase in 5-HT levels in the brain [51]. In addition, gut microbiota can use tryptophan directly. Preclinical studies have reported direct and indirect mechanisms by which the gut microbiota is able to regulate tryptophan availability for kynurenine pathway metabolism, with downstream effects on CNS function [52].

Finally, impaired rapid eye movement (REM) sleep in depression is partially caused by the hyperactivation of REM-on cholinergic neurons. This is supported by the discovery that cholinergic stimulation leads to a more severe REM sleep inhibition in depressed patients compared to healthy controls [53]. Sleep disturbances are common in depressed subjects and also in subjects with dysbiosis and intestinal inflammation [54]. This is probably also attributable to an altered level of melatonin consequent to the altered level of its precursor, 5-HT. A recent study reports that total microbiome diversity was positively correlated with increased sleep efficiency and total sleep time and was negatively correlated with waking after sleep onset [55].

2. Conclusions

Membrane mobility changes the uptake of serotonin, which decreases in the case of increased fluidity and increases in the case of increased viscosity. The gut microbiota affects depression-like behavior through influencing brain tryptophan accessibility and the serotonergic system [56].

The overlapping of the two phenomena, a common finding in depressive psychopathology, helps to reduce the availability of serotonin and, therefore, its regulatory function on mood.

Furthermore, for the reasons previously analyzed and explained, inflammation, osteoporosis risk, and ischemic cardiovascular risk can also exist in psychopathology.

In this context, the microbiota–gut–brain axis, in the complexity of its interactions, appears crucial in governing our thoughts, actions, and weaknesses. For these reasons, we can hypothesize that “depression” should be understood as a biological and cultural synthesis of human existence.

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Abbreviations

AA	arachidonic acid
BD	bipolar disorder
CNS	central nervous system
DHA	docosahexaenoic acid
5-HT	5-hydroxytryptamine, serotonin
IBS	irritable bowel syndrome
IDO	indoleamine 2,3-dioxygenase
MD	major depression
REM	rapid eye movement
TDO	tryptophan 2,3-dioxygenase

References

1. Lapin, I.P.; Oxenkrug, G.F. Intensification of the central serotonergic processes as a possible determinant of the thymoleptic effect. *Lancet* **1969**, *1*, 32–39. [[CrossRef](#)]
2. Frazer, A.; Pandey, G.N.; Mendels, J. Metabolism of Tryptophan in Depressive Disease. *Arch. Gen. Psychiatry* **1973**, *29*, 528–535. [[CrossRef](#)] [[PubMed](#)]
3. Cowen, P.J.; Parry-Billings, M.; Newsholme, E.A. Decreased plasma tryptophan levels in major depression. *J. Affect. Disord.* **1989**, *16*, 27–31. [[CrossRef](#)]
4. Gabbay, V.; Ely, B.A.; Babb, J.; Liebes, L. The possible role of the kynurenine pathway in anhedonia in adolescents. *J. Neural. Transm.* **2012**, *119*, 253–260. [[CrossRef](#)]
5. Zhang, X.; Gainetdinov, R.R.; Beaulieu, J.M.; Sotnikova, T.D.; Burch, L.H.; Williams, R.B.; Schwartz, D.A.; Krishnan, R.R.; Caron, M.G. Loss-of-Function mutation in tryptophan hydroxylase-2 identified in unipolar major depression. *Neuron* **2005**, *45*, 11–16. [[CrossRef](#)]
6. Lemonde, S.; Turecki, G.; Bakish, D.; Du, L.; Hrdina, P.D.; Bown, C.D.; Sequeira, A.; Kushwaha, N.; Morris, S.J.; Basak, A.; et al. Impaired repression at a 5-hydroxytryptamine 1A receptor gene polymorphism associated with major depression and suicide. *J. Neurosci.* **2003**, *23*, 8788–8799. [[CrossRef](#)]
7. Hendriksen, E.; van Brgeijk, D.; Oosting, R.S.; Redegeld, F.A. Mast cells in neuroinflammation and brain disorders. *Neurosci. Biobehav. Rev.* **2017**, *79*, 119–133. [[CrossRef](#)]
8. Traina, G. Mast cells in the brain- Old cells, new target. *J. Integr. Neurosci.* **2017**, *16*, S69–S83. [[CrossRef](#)]
9. Traina, G. Mast cells in gut and brain and their potential role as an emerging therapeutic target for neural diseases. *Front. Cell. Neurosci.* **2019**, *13*, 345. [[CrossRef](#)]
10. Takahashi, S. Reduction of Blood Platelet Serotonin Levels in Manic and Depressed Patients. *Folia Psych. Neurol. Jap.* **1976**, *30*, 476–486. [[CrossRef](#)]
11. Stahl, S.M. The Human Platelet. A Diagnostic and Research Tool for the Study of Biogenic Amines in Psychiatric and Neurologic Disorders. *Arch. Gen. Psychiatry* **1977**, *34*, 509–516. [[CrossRef](#)] [[PubMed](#)]
12. Pletscher, A.; Laubscher, A. Blood Platelets as Models for Neurons: Uses and Limitations. *J. Neural Transm. Suppl.* **1980**, *16*, 7–16.
13. Da Prada, A.M.; Cesura, J.M.; Launay, J.G.; Richards, J.G. Platelets as a Model for Neurones? *Cell. Mol. Life Sci.* **1988**, *44*, 115–126. [[CrossRef](#)] [[PubMed](#)]
14. Kim, H.L.; Plaisant, O.; Leboyer, M.; Gay, C.; Kamal, L.; Devynck, M.A.; Meyer, P. Reduction of Platelet Serotonin in Major Depression (Endogenous Depression). *Comptes Rendus Seances Acad. Sci. III* **1982**, *295*, 619–622.
15. Musselman, D.L.; Tomer, A.; Manatunga, A.K.; Knight, B.T.; Porter, M.R.; Kasey, S.; Marzec, U.; Harker, L.A.; Nemeroff, C.B. Exaggerated Platelet Reactivity in Major Depression. *Am. J. Psychiatry* **1996**, *153*, 1313–1317.
16. Camacho, A.; Dimsdale, J.E. Platelets and Psychiatry: Lessons Learned from Old and New Studies. *Psychos. Med.* **2000**, *62*, 326–336. [[CrossRef](#)]
17. Plein, H.; Berk, M. The Platelet as a Peripheral Marker in Psychiatric Illness. *Hum. Psychopharm.* **2001**, *16*, 229–236. [[CrossRef](#)]
18. Cocchi, M.; Tonello, L.S.; Tsaluchidu, S.; Puri, B.K. The use of Artificial Neural Networks to Study Fatty Acids in Neuropsychiatric Disorders. *BMC Psychiatry* **2008**, *8*, S3. [[CrossRef](#)]
19. Cocchi, M.; Tonello, L. Bio Molecular Considerations in Major Depression and Ischemic Cardiovascular Disease. *Cent. Nerv. Syst. Agents Med. Chem.* **2010**, *10*, 97–107. [[CrossRef](#)]
20. Benedetti, S.; Bucciarelli, S.; Canestrari, F.; Catalani, S.; Mandolini, S.; Marconi, V.; Mastrogiacomo, A.; Silvestri, R.; Tagliamonte, M.; Venanzini, R.; et al. Platelet's Fatty Acids and Differential Diagnosis of Major Depression and Bipolar Disorder through the use of an Unsupervised Competitive- Learning Network Algorithm (SOM). *Open J. Depress.* **2014**, *3*, 52–73. [[CrossRef](#)]
21. Mammadova-Bach, E.; Mauler, M.; Braun, A.; Duerschmied, D. *Immuno-Thrombotic Effects of Platelet Serotonin*; IntechOpen: London, UK, 2017.
22. Walther, D.J.; Peter, J.U.; Winter, S.; Hölte, M.; Paulmann, N.; Grohmann, M.; Vowinckel, J.; Alamo-Bethencourt, V.; Wilhelm, C.S.; Ahnert-Hilger, G.; et al. Serotonylation of small GTPases is a signal transduction pathway that triggers platelet alpha-granule release. *Cell* **2003**, *115*, 851–862. [[CrossRef](#)]

23. Cloutier, N.; Allaey, I.; Marcoux, G.; Machlus, K.R.; Mailhot, B.; Zufferey, A.; Levesque, T.; Becker, Y.; Tessandier, N.; Melki, I.; et al. Platelets release pathogenic serotonin and return to circulation after immune complex-mediated sequestration. *Proc. Natl. Acad. Sci. USA* **2018**, *115*, E1550–E1559. [[CrossRef](#)]
24. Zubenko, G.S.; Kopp, U.; Seto, T.; Firestone, L.L. Platelet Membrane Fluidity Individuals at Risk for Alzheimer's Disease: A Comparison of Results From Fluorescence Spectroscopy and Electron Spin Resonance Spectroscopy. *Psychopharmacology* **1999**, *145*, 175–180. [[CrossRef](#)]
25. Zubenko, G.S.; Cohen, B.M.; Reynolds, C.F.; Boller, F.; Malinakova, I.; Keefe, N. Platelet membrane fluidity in Alzheimer's disease and major depression. *Am. J. Psychiatry* **1987**, *144*, 860–868. [[PubMed](#)]
26. van Rensburg, S.J.; Carstens, M.E.; Potocnik, F.C.V.; Aucamp, A.K.; Taljaard, J.J.F.; Koch, K.R. Membrane fluidity of platelets and erythrocytes in patients with Alzheimer's disease and the effect of small amounts of aluminium on platelet and erythrocyte membranes. *Neurochem. Res.* **1992**, *17*, 825–829. [[CrossRef](#)] [[PubMed](#)]
27. Leiter, O.; Walker, T.L. Platelets in Neurodegenerative Conditions—Friend or Foe? *Front. Immunol.* **2020**, *11*, 747. [[CrossRef](#)]
28. Padmakumar, M.; Van Raes, E.; Van Geet, C.; Freson, K. Blood platelet research in autism spectrum disorders: In search of biomarkers. *Res. Pract. Thromb. Haemost.* **2019**, *3*, 566–577. [[CrossRef](#)]
29. Tonello, L.; Cocchi, M. The Cell Membrane: Is it a bridge from psychiatry to quantum consciousness? *NeuroQuantology* **2010**, *1*, 54–60. [[CrossRef](#)]
30. Heron, D.S.; Shinitzky, M.; Hershkowitz, M.; Samuel, D. Lipid Fluidity Markedly Modulates the Binding of Serotonin to Mouse Brain Membranes. *Proc. Natl. Acad. Sci. USA* **1980**, *77*, 7463–7467. [[CrossRef](#)]
31. Traina, G.; Cocchi, M. Mast Cells, Astrocytes, Arachidonic Acid: Do They Play a Role in Depression? *Appl. Sci.* **2020**, *10*, 3455. [[CrossRef](#)]
32. Horrocks, L.A.; Farooqui, A.A. Docosahexaenoic acid in the diet: Its importance in maintenance and restoration of neural membrane function. *Prostag. Leukotr. Essent. Fat. Acids* **2004**, *70*, 361–372. [[CrossRef](#)] [[PubMed](#)]
33. Hibbeln, J.R.; Palmer, J.W.; Davis, J.M. Are disturbances in lipid-protein interactions by phospholipase-A2 a predisposing factor in affective illness? *Biol. Psychiatry* **1989**, *25*, 945–961. [[CrossRef](#)]
34. Walsh, M.T.; Dinan, T.G.; Condren, R.M.; Ryan, M.; Kenny, D. Depression is associated with an increase in the expression of the platelet adhesion receptor glycoprotein Ib. *Life Sci.* **2002**, *70*, 3155–3165. [[CrossRef](#)]
35. Piletz, J.E.; Zhu, H.; Madakasira, S.; Pazzaglia, P.; DeVane, C.L.; Goldman, N.; Halaris, A. Elevated P-selectin on platelets in depression: Response to bupropion. *J. Psychiatr. Res.* **2000**, *34*, 397–404. [[CrossRef](#)]
36. Müller, C.P.; Reichel, M.; Muhle, C.; Rhein, C. Brain membrane lipids in major depression and anxiety disorders. *Bioch. Biophys. Acta* **2015**, *1851*, 1052–1065. [[CrossRef](#)] [[PubMed](#)]
37. Watala, C.; Golański, J.; Boncler, M.A.; Pietrucha, T.; Gwoździński, K. Membrane Lipid Fluidity of Blood Platelets: A Common Denominator That Underlies the Opposing Actions of Various Agents that Affect Platelet Activation in Whole Blood. *Platelets* **1998**, *9*, 315–327.
38. Kalipatnapu, S.; Pucadyil, T.J.; Chattopadhyay, A. Membrane Organization and Dynamics of the Serotonin1A Receptor Monitored Using Fluorescence Microscopic Approaches. In *Serotonin Receptors in Neurobiology*; Chattopadhyay, A., Ed.; CRC Press/Taylor & Francis: Boca Raton, FL, USA, 2007; Chapter 3.
39. Cocchi, M.; Tonello, L.; Amato, P.; De Lucia, A. Platelet and Brain Fatty acid transfer: Hypothesis on Arachidonic Acid and its relationship to Major Depression. *J. Biol. Res.* **2009**, *82*, 47–53. [[CrossRef](#)]
40. Green, P.; Gispan-Herman, I.; Yadid, G. Increased Arachidonic Acid Concentration in the Brain of Flinders Sensitive Line Rats, an Animal Model of Depression. *J. Lipid. Res.* **2005**, *46*, 1093–1096. [[CrossRef](#)]
41. Cocchi, M.; Tonello, L.; Lercker, G. Fatty acids, membrane viscosity, serotonin and ischemic heart disease. *Lipids Health Dis.* **2010**, *9*, 97. [[CrossRef](#)]
42. Cocchi, M.; Tonello, L.; Gabrielli, F.; Pregnolato, M. Depression, osteoporosis, serotonin and cell membrane viscosity between biology and philosophical anthropology. *Ann. Gen. Psychiatry* **2011**, *10*, 9. [[CrossRef](#)]
43. Bear, T.L.K.; Dalziel, J.E.; Coad, J.; Roy, N.C.; Butts, C.A.; Gopal, P.K. The Role of the Gut Microbiota in Dietary Interventions for Depression and Anxiety. *Adv. Nutr.* **2020**, *11*, 890–907. [[CrossRef](#)] [[PubMed](#)]
44. Dinan, T.G.; Cryan, J.F. Melancholic microbes: A link between gut microbioma and depression? *Neurogastroenterol. Motil.* **2013**, *25*, 713–719. [[CrossRef](#)] [[PubMed](#)]
45. Dinan, T.G.; Cryan, J.F. The impact of gut microbiota on brain and behaviour: Implications for psychiatry. *Curr. Opin. Clin. Nutr. Metab. Care* **2015**, *18*, 552–558. [[CrossRef](#)]

46. Conte, C.; Sichetti, M.; Traina, G. Gut–Brain Axis: Focus on Neurodegeneration and Mast Cells. *Appl. Sci.* **2020**, *10*, 1828. [[CrossRef](#)]
47. Gao, J.; Xu, K.; Liu, H.; Liu, G.; Bai, M.; Peng, C.; Li, T.; Yin, Y. Impact of the Gut Microbiota on Intestinal Immunity Mediated by Tryptophan Metabolism. *Front. Cell. Infect. Microbiol.* **2018**, *8*, 13. [[CrossRef](#)] [[PubMed](#)]
48. Cervenka, I.; Agudelo, L.Z.; Ruas, J.L. Kynurenines: Tryptophan's Metabolites in Exercise, Inflammation, and Mental Health. *Science* **2017**, *357*, 9794. [[CrossRef](#)]
49. Banskota, S.; Ghia, J.E.; Khan, W.I. Serotonin in the gut: Blessing or a curse. *Biochimie* **2019**, *161*, 56–64. [[CrossRef](#)]
50. Jenkins, T.A.; Nguyen, J.C.D.; Polglaze, K.E.; Bertrand, P.P. Influence of tryptophan and serotonin on mood and cognition with a possible role of the gut-brain axis. *Nutrients* **2016**, *8*, 56. [[CrossRef](#)]
51. Kaur, H.; Bose, C.; Mande, S.S. Tryptophan Metabolism by Gut Microbiome and Gut-Brain-Axis: An in silico Analysis. *Front. Neurosci.* **2019**, *13*, 1365. [[CrossRef](#)]
52. Kennedy, P.J.; Cryan, J.F.; Dinan, T.G.; Clarke, G. Kynurenine Pathway Metabolism and the Microbiota–Gut–Brain Axis. *Neuropharmacology* **2017**, *112*, 399–412. [[CrossRef](#)]
53. Berger, M.; Riemann, D.; Höchli, D.; Spiegel, R. The rapid-movement cholinergic sleep induction test of the eye with RS-86. Indicator of state or trait of depression? *Arch. Gen. Psychiatry* **1989**, *46*, 421–428. [[CrossRef](#)] [[PubMed](#)]
54. Li, Y.; Hao, Y.; Fan, F.; Zhang, B. The Role of Microbiome in Insomnia, Circadian Disturbance and Depression. *Front. Psychiatry* **2018**, *9*, 669. [[CrossRef](#)] [[PubMed](#)]
55. Smith, R.P.; Easson, C.; Lyle, S.M.; Kapoor, R.; Donnelly, C.P.; Davidson, E.J.; Parikh, E.; Lopez, J.V.; Tartar, J.L. Gut microbiome diversity is associated with sleep physiology in humans. *PLoS ONE* **2019**, *14*, e0222394. [[CrossRef](#)] [[PubMed](#)]
56. Lukic, I.; Getselter, D.; Koren, O.; Elliott, E. Role of Tryptophan in Microbiota-Induced Depressive-Like Behavior: Evidence from Tryptophan Depletion Study. *Front. Behav. Neurosci.* **2019**, *13*, 123. [[CrossRef](#)]



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