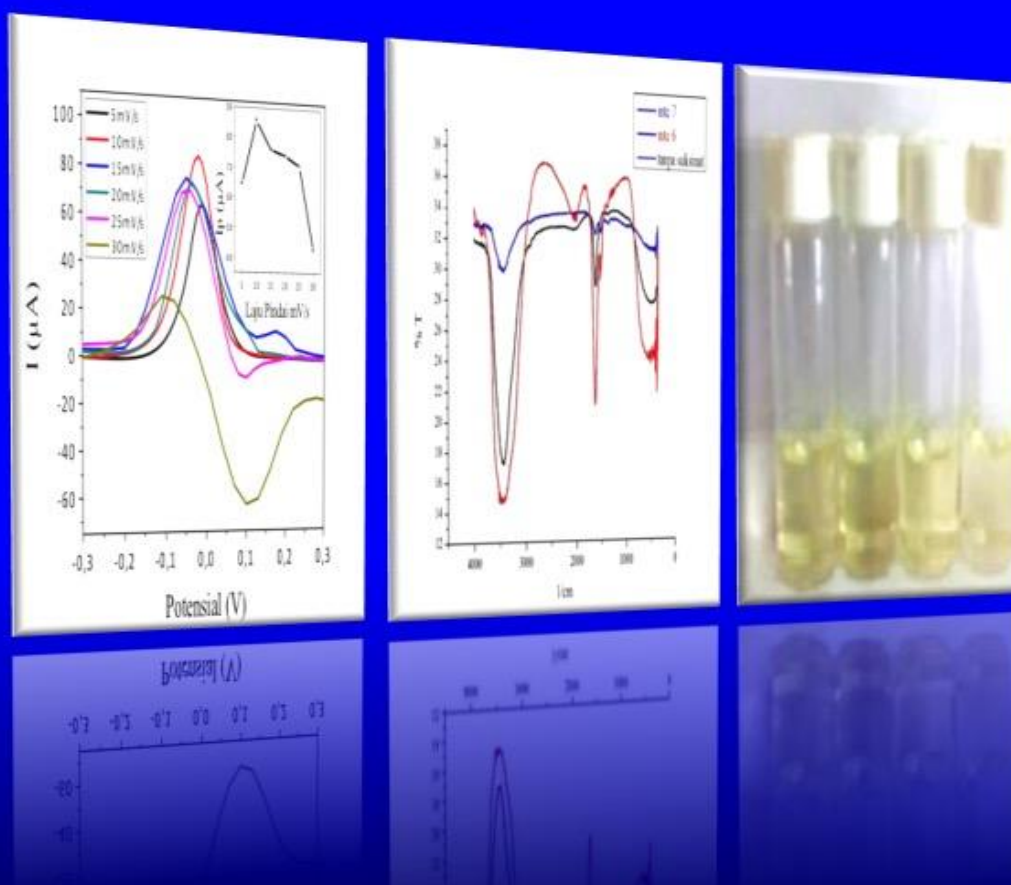
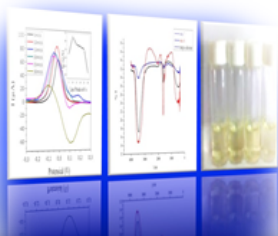


Jurnal KIMIA RISET

Volume 1 No. 2, Desember 2016; ISSN: 2528-0414; e-ISSN: 2528-0422



**Departemen Kimia
Fakultas Sains dan Teknologi
Universitas Airlangga
<http://e-journal.unair.ac.id/index.php/JKR>**



[Home](#) > [About the Journal](#) > [Editorial Team](#)

EDITORIAL TEAM

EDITOR-IN-CHIEF

[Dr. Purkan Purkan](#), Department of Chemistry, Faculty of Science and Technology, Airlangga University

VICE EDITOR-IN-CHIEF

[Dr. Abdulloh Abdulloh](#), Department of Chemistry, Faculty of Science and Technology, Airlangga University

EDITORIAL BOARD

[Prof. Ni Nyoman Tri Puspaningsih](#), Department of Chemistry, Faculty of Science and Technology, Airlangga University

[Prof. Chun-Hu Chen](#), Department of Chemistry, National Sun Yat-sen University, Kaohsiung, 80424, Taiwan

[Prof. Punnapayak Hunsa](#), Faculty of Science, Chulalongkorn University, Bangkok, 10330, Thailand

[Dr.rer.nat Ganden Supriyanto](#), Department of Chemistry, Faculty of Science and Technology, Airlangga University

[Dr. Hery Suwito](#), Department of Chemistry, Faculty of Science and Technology, Airlangga University

MANAGING EDITOR

[Dr. Imam Siswanto](#), Department of Chemistry, Faculty of Science and Technology, Airlangga University

[Rico Ramadhan, Ph.D](#), Department of Chemistry, Faculty of Science and Technology, Airlangga University

[Qurrota A'yuni, M.Si](#), Department of Chemistry, Faculty of Science and Technology, Airlangga University

[Fatiha Khairunnisa, M.Si](#), Department of Chemistry, Faculty of Science and Technology, Airlangga University

ADMINISTRATION SUPPORT

[Fendi Kristanto](#), Department of Chemistry Faculty of Science and Technology Universitas Airlangga

[Oktavia Wahyu Ningsih](#), Department of Chemistry Faculty of Science and Technology Universitas Airlangga

Instruction for Author

[Guide for authors](#)

[Online Submission](#)

[Document Template](#)

Journal Policy

[Focus and Scope](#)

[Publication Ethics](#)

[Article Processing Charge](#)

[Peer Reviewers](#)

[Editorial Team](#)

[Open Access Statement](#)

[Plagiarism](#)

[Copyright](#)

[Contact](#)

USER

Username

Password

Remember me

NOTIFICATIONS

[View](#)

[Subscribe](#)

JOURNAL CONTENT

Search

Search Scope

All

Browse

[By Issue](#)

[By Author](#)

[By Title](#)

[Other Journals](#)

INFORMATION

[For Readers](#)

[For Authors](#)

[For Librarians](#)

KEYWORDS

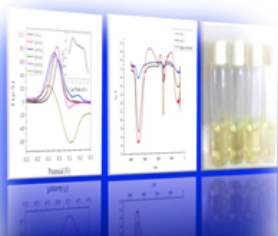
Alkaline protease, Chicken feces, Gallus gallus bankiva, Pseudomonas sp, Alkaloid turunan furokuinolin, skimmianin, Toddalia asiatica L., antikanker **Analytical Chemistry** Antifungal preparations, Candida albicans, Betel leaf (SDS), Gelinggang leaf extract (Cassia alata L.) Antioxidant, S. polycystum, T. decurrens, Dutungan island Biochemistry Biodiesel, In situ,



View JKR Stats

00063053





Home > Archives > Vol 1, No 2 (2016)

VOL 1, NO 2 (2016)

DESEMBER

TABLE OF CONTENTS

ARTICLES

Pengembangan Elektroda Pasta Karbon termodifikasi Molecularly Imprinted Polymer sebagai Sensor Potensiometri Asam Urat

doi 10.20473/jkr.v1i2.3085

Mirartul Khasanah, Handoko Darmokoemo, Nesti Widayanti

PDF
65-72

OPTIMASI KONSENTRASI SUBSTRAT XILAN AMPAS TAHU TERHADAP ENDO-B-1,4-D-XYLANASE UNTUK MEMPRODUKSI XILOOLIGOSAKARIDA

doi 10.20473/jkr.v1i2.3084

Anak Agung Istri Ratnadewi, Wuryanti Handayani, Siti Nur Avida

PDF
73-80

Skopoletin Senyawa Fenilpropanoid dari Kulit Umbi Ubi Jalar (*Ipomoea batatas* L.) varietas IR-melati

doi 10.20473/jkr.v1i2.3087

Citra Putri Pramitha, Nanik Siti Aminah, Alfinda Novi Kristanti

PDF
81-85

Cangkang Buah Karet Dengan Perekat Limbah Plastik Polipropilena Sebagai Alternatif Papan Partikel

doi 10.20473/jkr.v1i2.3091

Charles Banon, Teja Dwi Sutanto, Irfan Gustian, Ilman Koharudin, Widia Rahmi

PDF
86-93

STUDI PELEPASAN TERKONTROL TERHADAP NANOENKAPSULASI DIMETOKSI AMINO CALKON SEBAGAI DESAIN KANDIDAT SENYAWA ANTI KANKER YANG EFEKTIF

doi 10.20473/jkr.v1i2.3089

Mochamad Zakki Fahmi, Hery Suwito, Shofi Yasmin Nurain, Yogi Putra Hidayatullah

PDF
94-100

Korelasi hasil karakterisasi XRD, N2 adsorpsi-desorpsi dan TEM pada Karbon Mesopori dari Gelatin Tulang Sapi

doi 10.20473/jkr.v1i2.3088

Maria Ulfa

PDF
101-110

ISOLATION OF BIOACTIVE COMPOUNDS FROM DICRANACEAE MOSSES

doi 10.20473/jkr.v1i2.3086

Junairiah Junairiah, Tri Nurhariyati, Ni'matuzahroh Ni'matuzahroh, Lilis Sulistyorini

PDF
111-121

TEKNIK VOLTAMETRI PELUCUTAN ANODIK GELOMBANG PERSEGI UNTUK PENENTUAN KADAR LOGAM Cu DALAM KANGKUNG AIR

doi 10.20473/jkr.v1i2.3094

Irdhawati Irdhawati, Liana Sari, Ida Ayu Raka Astiti Asih

PDF
122-128

STUDI HUBUNGAN KUANTITATIF STRUKTUR AKTIVITAS SENYAWA TURUNAN MEISOINDIGO SEBAGAI INHIBITOR CDK4

PDF

Instruction for Author

Guide for authors

Online Submission

Document Template

Journal Policy

Focus and Scope

Publication Ethics

Article Processing Charge

Peer Reviewers

Editorial Team

Open Access Statement

Plagiarism

Copyright

Contact

USER

Username

Password

Remember me

Login

NOTIFICATIONS

View
Subscribe

JOURNAL CONTENT

Search

Search Scope

All

Search

Browse

By Issue

By Author

By Title

Other Journals

INFORMATION

For Readers

For Authors

For Librarians

KEYWORDS

Alkaline protease, Chicken feces, Gallus gallus bankiva, Pseudomonas sp Alkaloid turunan furokuinolin, skimmianin, Toddalia asiatica L., antikanker Analytical Chemistry Antifungal preparations, Candida albicans, Betel leaf (SDS), Gelinggang leaf extract (Cassia alata L.) Antioxidant, S. polycystum, T. decurrens, Dutungan island Biochemistry Biodiesel, In situ

doi 10.20473/jkr.v1i2.3090

Muhammad Arba, Riki Andriansyah, Messi Leonita

INSERSI GEN pncA KE DALAM PLASMID pGEM-T

doi 10.20473/jkr.v1i2.3092

Eli Hendrik Sanjaya

129-
134

PDF

135-
144

Microwave, Dedak Padi DENV-2, 2-isopropanol, Methanol, Chloroform, Ethanol 96%, RNA Isolation Dioscorea hispida Dennst, Dioscoreaceae, phenolic, 4-ethoxy-3-methoxyphenol, methyl-3,4-dihydroxybenzoic, antioxidant Drug loading, Ibuprofen, UIO-66, Zr-Metal Organic Framework Ganoderma lucidum, senyawa fitokimia, GCMS Koenzym Q10, Rice bran oil, Surfaktan, Nanoemulsi, Nanoemulsi Gel RNA, Precipitation Solvent, DMF, EDTA, Ultrapure H2O Selada merah (Lactuca sativa var. Crispa), sitotoksik, Artemia salina Leach. ZSM-5, metakaolin, terdealuminasi, tanpa templat, pengaturan suhu dan waktu asam amino, besi, nanomagnetit, dan mineral enzim, biodegradation, fungus enzyme fenil propanoid, skopoletin, dan Ipomoea batatas L. logam Fe, anodic stripping voltammetry, terong ungu, elektroda glassi karbon methanol, Phytochemical screening, Piper betle L. var Nigra. methyloleate, catalyst, Ni/H5NZ, hydrocracking

CURRENT ISSUE

ATOM	1.0
RSS	2.0
RSS	1.0



View JKR Stats

00063056



Pusat Pengembangan Jurnal dan Publikasi Ilmiah
Universitas Airlangga
copyright@2017 Template PPJPI

Journal Profile

Jurnal Kimia Riset

eISSN : 2528-0422 | pISSN : 2528-0414

Science

Universitas Airlangga



S3

Sinta Score



Indexed by GARUDA

3

H-Index

3

H5-Index

31

Citations

31

5 Year Citations

ISOLATION OF BIOACTIVE COMPOUNDS FROM DICRANACEAE MOSSES

Junairiah Junairiah^{1*}, Tri Nurhariyati¹, Ni'matuzahroh¹, Lilis Sulistyorini²

¹ Department of Biology, Faculty of Science and Technology, Airlangga University, Surabaya

² Faculty of Public Health, Airlangga University, Surabaya

email: alip.jun1@gmail.com

Received 3 July 2016

Accepted 30 November 2016

Abstract

Dicranoloma reflexum and *Dicranella coarctata* are mosses from Dicranaceae family. This study was purposed to identify bioactive compounds contained from both species. *Dicranoloma reflexum* and *Dicranella coarctata* collected from Cangar forest, Batu, East Java. Mosses was rinsed, dried and crushed into powder. Extraction was performed using maceration method with n-hexane, acetic acid, and methanol solvent. Compounds obtained then identified using Gas Chromatography Mass Spectra. Result showed that n-hexane, acetic ethyl, and methanol extract of *Dicranoloma reflexum* contained 61, 16, and 58 compounds respectively. Main component of each extract was 1-octadecene, phenol, and 9-octadecanoic acid. N-hexane, acetic ethyl, and methanol extract of *Dicranella coarctata* contained 5, 38, and 23 compounds respectively. Main component of each extract was thiosulphuric acid, E-15 heptedecenal, and n-hexadecanoic acid.

Keywords : *Dicranaceae*, *bioactive compounds*

Introduction

Bryophyte or mosses is source for various secondary metabolites (Asakawa, 2008). Mosses from Bryophyte is rich on secondary metabolites, such as terpenoid, flavonoid, and bibenzyles, also other compounds like fatty acid (Asakawa, 2007). With such secondary metabolites contents, many species of Bryophyta possess various biological activities, such as anti-bacteria, anti-fungal, anti-oxidant, and anti-inflammatory properties (Saxena dan Harinder, 2004).

For example Indian mosses *Bryum argenteum* had anti-pyretic, anti-rhynitic, and anti-bacterial properties. Himalayan *Fissidens laxitextus* has been known as an anti-diuretic. *Funaria hygrometrica* from India was also used as medicine for skin disease. *Entodon myurus* was found to be potential as anti-bacterial agent towards

Enterobacter aerogenes and *Klebsiella pneumoniae* (Alam et al., 2015).

Dicranuloma reflexum and *Dicranella coarctata* are mosses species from Bryopsida (Musci) class, Dicranales order, and Dicranaceae family. Both species has different habitat. *Dicranuloma reflexum* is arboreal moss, while *Dicranella coarctata* is terrestrial moss. Up until now, bioactive compounds from both species has yet to be studied. This study was aimed to isolate and identify bioactive compounds from *Dicranuloma reflexum* and *Dicranella coarctata*. This study was expected to give new lights on bioactive compounds contained on both moss species as basic information which then used for evaluating biological activities.

Material and method

Main ingredient used on this study was moss gametophyte of *Dicranoloma reflexum* and *Dicranella coarctata*. Both species was collected from Cangar forest, Batu, Malang, East Java.

Extraction

Moss gametophyte *Dicranoloma reflexum* and *Dicranella coarctata* was rinsed, dried, cut into small pieces, and weighted. Total weigh of *Dicranoloma reflexum* collected was 17.1 g, while *Dicranella coarctata* weighted 4.5 g. Moss was divided into three parts, each weighted 5.7 g and 1.5 g. Each part extracted using n-hexane, acetic ethyl, and methanol. Solvent volume used for extraction of *Dicranoloma reflexum* and *Dicranella coarctata* were 200 ml and 100 ml respectively. Extraction was performed using maceration method. Simplicia soaking was conducted for 4 days and repeated for 4 times. Each extract of *Dicranoloma reflexum* and *Dicranella coarctata* was dried until all solvent had evaporated perfectly.

Bioactive Compound Identification

Each extract obtained was analyzed using Gass Chromatography Mass Spectra.

Results and discussion

Dicranoloma reflexum and *Dicranella coarctata* were mosses lived on arboreal

and terrestrial habitat respectively (Fig 1 and 2).



Figure 1. Morphology of *Dicranoloma reflexum* moss.



Figure 2. Morphology of *Dicranella coarctata* moss.

Extract obtained from *Dicranoloma reflexum* and *Dicranella coarctata* were presented on Table 1.

Table 1. Extraction of *Dicranoloma reflexum* and *Dicranella coarctata* using various solvent.

No	Moss species	Simplicia weigh (g)	Solvent volume (ml)	N-hexane extract weigh (g)	Acetic ethyl extract weigh (g)	Methanol extract weigh (g)
1	<i>Dicranoloma reflexum</i>	5.7	200	0.1531	0.0851	0.7110
2	<i>Dicranella coarctata</i>	1.5	100	0.0063	0.0222	0.0493

GCMS Analysis of *Dicranoloma reflexum* and *Dicranella coarctata* Extract

Based on GC-MS analysis, chromatogram profile of n-hexane, acetic ethyl, and methanol of *Dicranoloma reflexum* and *Dicranella coarctata* were presented on Figure 2, 3, 4, 5, and 6.

Result of identification on bioactive compounds from n-hexane, acetic ethyl, and methanol extract were presented on table 2, 3, 4, 5, 6, dan 7. N-hexane, acetic ethyl, and methanol extract was found to contain 61, 16, and 58 compounds respectively (Table 2, 3, and 4). N-hexane, acetic ethyl, and methanol extract of *Dicranella coarctata* contained respectively 5, 38, and 23 compounds (Table 5, 6, and 7).

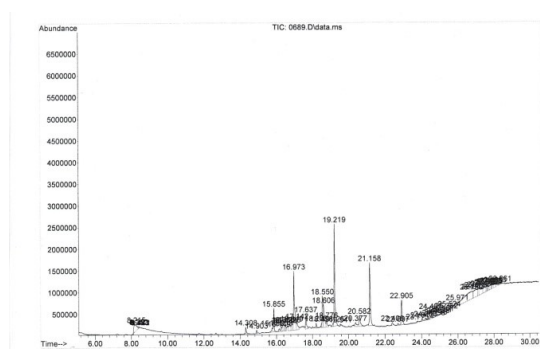


Figure 2. Chromatogram profile of n-hexane extract from *Dicranoloma reflexum*

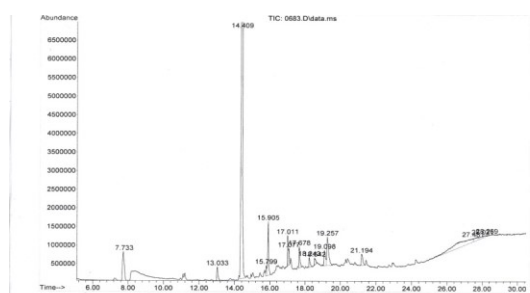


Figure 3. Chromatogram profile of acetic ethyl extract from *Dicranoloma reflexum*

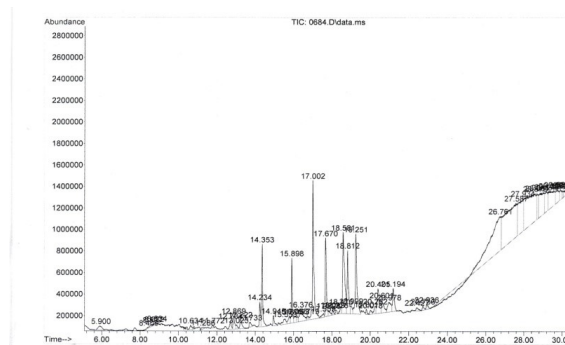


Figure 4. Chromatogram profile of methanol extract from *Dicranoloma reflexum*

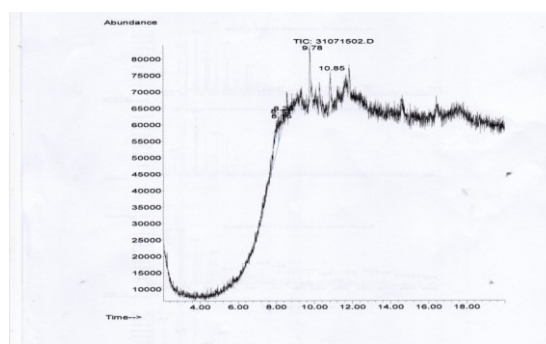


Figure 5. Chromatogram profile of n-hexane extract from *Dicranella coarctata*

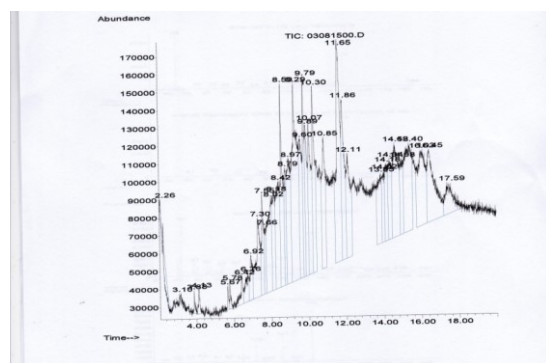


Figure 6. Chromatogram profile of acetic ethyl extract from *Dicranella coarctata*

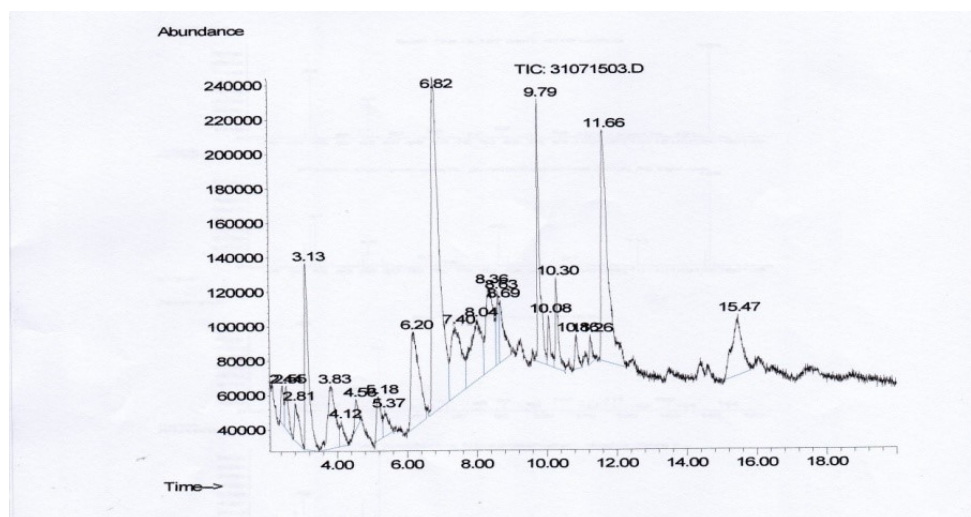


Figure 7. Chromatogram profile of methanol extract from *Dicranella coarctata*

Table 2. Chemical compounds from n-hexane extract of *Dicranoloma reflexum*

Peak	Retention time	Compound name	Areal range (%)
1	8.217	Naphthali Azulene	2.67
2	8.354	Naphthalene	0.21
3	8.383	Naphthalene	0.14
4	8.423	Naphthalene	0.04
5	8.440	Naphthalene	0.03
6	14.311	Pentanoic acid	0.81
7	14.900	5,7-Diamino-1,2,3,4-tetrahydro-1,1,4,4,6-pentamethylnaphthalene	0.36
8	15.764	Heptadecane	0.71
9	15.856	Cyclopentanepropanoic acid	2.51
10	16.147	2',6'-Acetoxyidide	0.48
11	16.268	Trimethylphenylsilane	0.62
12	16.353	Nonyphenol	1.21
13	16.411	4- Nonylphenol	0.89
14	16.479	Nonylphenol	0.47
15	16.497	Nonylphenol	0.68
16	16.714	Nonylphenol	2.05
17	16.851	Phenol	1.33
18	16.971	1-Octadecene	9.95
19	17.143	Eicosane	1.66
20	17.635	Hexahydrofarnesyl acetone	2.08
21	18.202	Nonadecane	0.49
22	18.499	1- Pentadecane	0.26
23	18.551	Pentadecanoic acid	4.17
24	18.608	Hexadecanoic acid	3.70
25	18.774	Hexadecanoic acid	1.18

Continued of Table 2

Peak	Retention time	Compound name	Areal range (%)
26	19.129	Dodecynilsuccinic anhydride 9 Nonadecene	0.38
27	19.220	1- Octadecene	12.53
28	19.541	I-Propyl 14 methyl-Pentadecanoate	0.15
29	20.376	Oleic acid	1.15
30	20.582	Octadecanoic acid	2.10
31	21.160	1- Octadecene	8.25
32	22.402	Eicisanoic acid	0.39
33	22.699	9-Tricosene (Z)-	0.31
34	22.905	9-Tricosene (Z)-	3.89
35	23.775	9-Tricosene (Z)-	0.22
36	24.015	9-Octadecanoic acid	0.30
37	24.204	2-Pyrazoline, 5-hydroxy-3-metyl-5-trifluoromethyl-2-(2-isopropyl-5-methylphenox	0.32
38	24.490	1-nonadecene	1.28
39	24.656	Elaidic acid	0.07
40	24.776	9-Tricosene (Z)-	0.02
41	24.908	Elaidic acid	0.13
42	25.051	9-Octadecenoic acid	0.14
43	25.160	9-Octadecenoic acid	0.07
44	25.360	Elaidic acid	0.35
45	25.526	9-Octadecenoic acid	0.45
46	25.972	Elaidic acid	2.27
47	26.750	9-Octadecenoic acid	10.93
48	26.842	9-Octadecenoic acid	0.68
49	26.996	9-Octadecenoic acid	3.85
50	27.151	2,3- Dihydroxypropyl elaidate	1.02
51	27.340	9-Octadecenoic acid	2.96
52	27.534	Elaidic acid	2.09
53	27.574	Oleic acid	0.54
54	27.620	n-Propyl 9- Octadecanoate	0.41
55	27.689	16-Octadecenoic acid	0.61
56	27.717	n-Propyl 9- Octadecanoate	0.50
57	27.803	9-Octadecenoic acid	0.83
58	27.843	9-Octadecenoic acid	0.26
59	27.889	9-tricosane, (Z)-	0.34
60	28.084	9-Octadecenoic acid	0.98
61	28.358	i-Propyl 9-Octadecenoate	0.55

Table 3. Chemical compounds from acetic ethyl extract of *Dicranoloma reflexum*

Peak	Retention time	Compound name	Areal range (%)
1	7.730	Cyclohexene,3,5,5-timethyl-(CAS)	3.97
2	13.035	Phenol	13.035
3	14.408	Pentan-1,3-Dioldiisobutyrate,2,2,4-Trimethyl	67.78
4	15.798	Heptadecane	0.64
5	15.907	Allyldimethyl(prop-1-ynyl)silane	3.77
6	17.011	1-Octadecene	2.02
7	17.074	1-Octadecene	1.00
8	17.675	2-Pentadecanone,6,10,14-trimethyl-	1.68
9	18.242	Nonadecane	0.92
10	18.545	4-(91Z)-N-hydroxythanimidoyl)2-methylpyridazin-3(2H)-one	0.70
11	19.100	1,2-Benzenedicarboxylic acid, dibutyl ester	1.77
12	19.260	1-Octadecene	3.81
13	21.194	9-Tricosane	1.29
14	27.483	11-Octadecenoic acid	7.81
15	28.020	n-Propyl 9-Octadecenoate	1.41
16	28.267	Erucic acid	0.29

Table 4. Chemical compounds from methanol extract of *Dicranoloma reflexum*

Peak	Retention time	Compound name	Areal range (%)
1	5.899	Cycloheptane	0.50
2	8.411	4-Fluorothiophenol	0.06
3	8.594	Azulene	0.14
4	8.680	Naphthalene	0.07
5	8.835	Naphthalene	0.12
6	10.632	1,3-Cyclopentanedione,4-hydroxy-2methyl	0.32
7	11.284	Hexenoic acid	0.15
8	11.770	3-Chloropropionic acid	0.14
9	12.703	2-Furanmethanol	0.45
10	12.869	2,5-Difluorobenzenoic acid	0.71
11	13.023	Heptadecanoic acid	0.21
12	13.224	Phenol	0.66
13	13.733	2(4H)-Benzofuranone,5,6,7,71-tetrahydro-4,4,7a-trimethyl	0.30
14	14.236	2-Tetradecene	0.94
15	14.351	Propanoic acid	2.60
16	14.946	8-(N-(Etfyl) amino)-5,6-dimethoxyquinoline	0.52
17	15.552	Cyclotetradecane	0.58
18	15.781	Cyclopentadecane	0.45
19	15.896	Pentanoic acid	2.01
20	16.096	n-Tetradecane	0.46
21	16.193	1-Tetradecane	0.22
22	16.376	2,6-Diisopropyl naphthalene	1.44
23	16.720	1-Octadecene	0.17

Continued of Table 4

Peak	Retention time	Compound name	Areal range (%)
24	17.000	1-Octadecene	6.32
25	17.561	Cyclohexanone	0.26
26	17.670	2-pentadecanone	2.99
27	17.830	4-((1Z)-(N-Hydroxyethanimydoyl)-2-methylpyridazine-3-(2H)-one	0.31
28	18.047	9-Octadecenoic acid	0.21
29	18.230	2-Hexyldecanol	0.23
30	18.476	11-Octadecenoic acid	0.32
31	18.579	Hexadecenoic acid	5.27
32	18.814	Methyl-3-(3,5-Diterbutyl-4-Hydroxyphenyl)Propionate	2.75
33	18.997	1-Octadecene	0.51
34	18.249	1-Octadecene	4.26
35	18.798	9-Octadecanal	0.21
36	20.16	9-Octadecenoic acid	0.26
37	20.262	Tert-Hexadenanethiol	0.40
38	20.405	9-Octadecenoic acid	2.24
39	20.599	10-Octadecenoic acid	1.07
40	20.977	9-Octadecenoic acid	1.10
41	21.194	1-Heptacosanol	1.29
42	22.430	Thiosulfuric acid	0.24
43	22.739	9-Octadecenoic acid	0.23
44	22.934	9-Octadecenoic acid	0.33
45	26.762	9-Octadecenoic acid	11.06
46	27.586	9-Octadecenoic acid	15.03
47	27.935	9-Octadecenoic acid	5.71
48	28.576	Elaidic acid	10.92
49	28.679	Elaidic acid	1.15
50	29.022	9-Tricosane, (Z)-	3.83
51	29.176	Elaidic acid	1.95
52	29.400	9-Octadecenoic acid	3.19
53	29.680	Elaidic acid	1.35
54	29.926	Elaidic acid	0.64
55	30.029	Elaidic acid	0.24
56	30.098	9-Octadecenoic acid	0.29
57	30.218	9-Tricosane, (Z)-	0.27
58	30.338	9-Octadecenoic acid	0.49

Table 5. Chemical compounds from n-hexane extract of *Dicranella coarctata*

Peak	Retention time	Compound name	Areal range (%)
1	8.12	Thiosulfuric acid	42.52
2	8.15	Tetrahydroxycyclopentadienone	2.72
3	8.26	9-Octadecenoic acid	12.83
4	9.79	Cyclopentane	18.43
5	10.85	9-Octadecenoic acid	23.50

Table 6. Chemical compounds from acetic ethyl extract of *Dicranella coarctata*

Peak	Retention time	Compound name	Areal range (%)
1	2.26	Cyclotetrasiloxane	0.74
2	3.10	2-hexanamine, 4 methyl	0.18
3	3.88	2-hexanamine, 4 methyl	0.18
4	4.13	Naphthalene	0.32
5	5.67	Benzeneethanamine	0.21
6	5.78	Benzeneethanamine	0.23
7	6.42	Benzeneethanamine	0.57
8	6.76	2,4(1H,3G=H)-Pyrimidinedione	0.80
9	6.92	Metramaminol bitartrate	0.92
10	7.30	o-hydroxybiphenyl	2.74
11	7.55	(cis)-2-nonadecene	1.60
12	7.65	Tetrahydroxycyclopentadienone	0.91
13	8.02	2Hydroxymino-N-(P-methoxyphen)	3.25
14	8.18	2Hydroxymino-N-(P-methoxyphen)	2.33
15	8.42	Hexadecanoic acid	3.58
16	8.59	Cyclophenyl 2-Methylenebutanyl	3.94
17	8.80	2Hydroxymino-N-(P-methoxyphen)	1.63
18	8.97	Hydroxymethapyrilene	4.03
19	9.29	E-15-Heptadecenal	8.61
20	9.61	1-Hentetracontanol	3.28
21	9.79	1,2-Cyclohexadiol	2.61
22	9.90	14-BETA-H-Pregna	2.89
23	10.07	Trans pinene	3.62
24	10.30	Citronellyl valerat	3.56
25	10.85	Cyclohexane	3.42
26	11.65	Hexadecanoic acid	7.21
27	11.86	1-Pentadecene	3.85
28	12.11	(trans)-2-nonadecene	3.47
29	13.85	17-Pentatriacontene	2.37
30	14.02	4-Hexenoic acid	1.45
31	14.19	4-Hexenoic acid	1.80
32	14.34	17-Pentatriacontene	2.10
33	14.62	17-Pentatriacontene	4.46
34	14.97	17-Pentatriacontene	1.86
35	15.40	9-Octadecenal	5.15
36	16.03	1-Octadecene	5.05
37	16.45	1-Eiocosene	4.58
38	17.59	Dimer of Coleon F	1.94

Table 7. Chemical compounds from methanol extract of *Dicranella coarctata*

Peak	Retention time	Compound name	Areal range (%)
1	2.44	1,2,3,4-Tetrahydroxybutane	0.88
2	2.56	Pentanal	1.24
3	2.81	N,N'-Dimethylpiperazine	1.51
4	3.13	Cyclopentanone	6.34
5	3.84	Propylamine	4.28
6	4.13	Methyl 2-(phenylsulfonyl)-5-deu	0.90
7	4.55	2-Furancarboxaldehyde,5-(hydro	0.78
8	5.18	N-(2-methoxycarbonylethylidene)	1.61
9	5.37	(E)-4-chloro-2,3-dimethyl	1.04
10	6.20	Isothiazole	6.93
11	6.82	6,6 Dideutero-nonen-1-ol	22.92
12	7.40	Nonanoic acid	6.47
13	8.05	Benzene,1-methyl-2-(2-propenyl	5.85
14	8.36	1H-indene,2,3-dihydro-	5.83
15	8.63	Undecanal	1.57
16	8.69	Hexadecanoic acid	2.42
17	9.80	Neophytadiene	5.73
18	10.08	1-formyl-2,2,6-trimethyl-3,(3-m	1.16
19	10.30	1,8-Nonadiene,2,8-dimethyl-	2.23
20	10.86	14-Beta-H-Pregna	0.89
21	11.26	Methyl-3-(3,5-Diterbutyl-4-Hyd	0.61
22	11.66	n-Hexadecanoic acid	13.62
23	15.48	Cyclopropaneoctanal	5.19

Based on GC-MS analysis, n-hexane extract of *Dicranoloma reflexum* was found to consist 61 peaks, indicating that it contained 61 compounds. Main compound found was 1-octadecene with areal total 12.53% (Table 2). Acetic ethyl extract of *Dicranoloma reflexum* contained 16 compounds, the main compound was phenol with areal range 13.035% (Table 3). Methanol extract of *Dicranoloma reflexum* contained 58 compounds. Main compound found was 9-octadecanoic acid with areal range of 15.03% (Table 4).

1-octadecene was a hydrocarbon compound usually found from *Moringa oleifera*. This compound possessed anti-oxidant, anti-bacterial, and anti-fungal properties towards *Bacillus cereus*, *Escherichia coli*, *Pseudomonas*

aeruginosa, *Penicillium aurantio*, *Penicillium griceum*, *Penicillium expansum*, *Penicillium digitatum*, and *Aspergillus niger* (Maruffo et al., 2013). N-hexane extract of *Prunus dome* was also found to contain 1-octadecene⁹¹. This compound had anti-bacterial activity towards *Salmonella* and anti-fungal towards *Microsporum canis*. In addition, this compound also had potential as anti-oxidant. Other compound found was phenol with chemical formula of C₆H₅OH and possessed hydroxyl group bound to phenyl ring on its structure. Cowan (1999) reported that phenol compound without hydroxyl group had higher anti-bacterial activity caused of higher affinity towards microbes plasma membrane. Quoumarin and quercetin were phenolic compounds with anti-bacterial activity

towards *Escherichia coli*, *Enterobacter aerogenes*, *Salmonella typhimurium*, and *Salmonella infantis* (Nitiema et al., 2012).

Another compound identified was 9-octadecanoic acid with chemical formula $C_{19}H_{36}O_2$ and possessed anti-oxidative and anti-carcinogenic biological activities (Syeda et al., 2011 and Hema et al., 2011). N-hexadecanoic acid had chemical formula of $C_{16}H_{32}$ and biological properties as anti-fungi, anti-oxidant, hypocholesterolemic, nematocide, and anti-androgenic flavor, haemolytic-5-alpha reductase inhibitor, anti-microbe, and anti-malaria (Hema et al., 2011 and Pietro et al., 2010). The ethanol extract of leaves of *Indigofera suffruticosa* containing n-hexadecanoic acid (9.83%) and 9-octadecanoic acid (10.21%) (Vijisara and Arumugam, 2014). The same compound is also found in the ethanol extract of beans, respectively 7.02% and 0.89%. Markkas and Govindharajala (2015) showed that the methanol extract of *Mollugo cerviana* containing hexadecanoic acid and 9-octadecanoic

acid. Harkati et al. (2012) reported that *Scorzonera undulata* containing hexadecanoic acid (42.2%) and 9-octadecanoic acid (7.7%). Aja et al. (2014) reported that the methanol extract of *Moringa oleifera* contains 9-octadecanoic acid is more higher (20.8%) than hexadecanoic acid (1.31%)

Another one compound found was thiosulphuric acid with chemical formula of $C_{20}H_7NO_3S_2$ and 157 molecular weigh. This compound was also identified from methanol extract of *Clerodendron phlomidis* ((Lakshmi dan Viji Stella Bai, 2015). Kumar et al (2011) reported that methanol and acetone extract of *Spirulina platensis* also contained E-15 Heptadecanal. This compound possessed anti-bacterial properties against *Staphylococcus aureus* and *Salmonella typhimurium*.

Acknowledgement

Author would like to thank 2015 DIPA DITLITABMAS of Airlangga University for funding this study.

References

- Aja PM, Nwachukwu N, Ibiam US, Igwenyi IO, Offor LE, and Orji UO. 2014. Chemical Constituents of *Moringa oleifera* Leaves and Seeds from Abakaliki, Nigeria. American Journal of Phytomedicine and Clinical Therapeutics. 2(3):310-321.
- Alam A, Srama V, Rawat KK, and Verma PK. 2015. Bryophytes-the Ignored Medicinal plants. Sikkim Manipal University. Vol2. No.1:299-315.
- Asakawa Y. 2007. Biologically Active Compounds from Bryophytes. Faculty of Pharmaceutical Sciences. Tokushima Bunri University. Yamashiro-cho. Tokushima. Japan.
- Asakawa Y. 2008. Liverworts-Potential Source of Medicinal Compounds. Current Pharmaceutical Design 14(29): 3067-3085.
- Cowan MM. 1999. Plant Products as Antimicrobial Agents. Clinical Microbiology Reviews 12(4): 564-570.
- Harkati B, Akkae S, Franca MGD. 2012. Composition of The Volatile Components Extracted from The Roots of *Scorzonera undulata* ssp *deliciosa* (Guis) Maire: from Algeria. Green and Sustainable Chemistry (2): 59-61.
- Hema R, kusmaravel s, and Alagusundaram. 2011. GC/MS Determination of Bioactive components of *Murraya koenigii*. Journal of American Science. (7)1.

- Kumar V, Bhatnagar AK, and Srivastava JN. 2011. Antibacterial Activity of Crude Extracts of *Spirulina platensis* and its structural Elucidation of Bioactive compound. Journal of Medicinal Plants research Vol 5(32). Pp. 7043-7048
- Lakshmi V and Viji Stella Bai G. 2015. Determination of Biologically Active Compounds in *Clerodendrum phlomidis* (L.) leaf Extract Using GC-MS. International journal of multidisciplinary research and Development: 2(1):294-300
- Mahmood A, Ahmed R, and kosar s. 2009. Phytochemical Screening and Biological Activities of the Oil Components of *Prunus domestica* Linn. Journal of saudy Chemical Society Vol 13. Issue 3: 273-277.
- Marimuthu K, Nagaraj N, Ravi O. 2014. GC-MS Analysis of Phytochemicals, Fatty Acids and Antimicrobial Potency of Dry Christmas Lima Beans. Int. J. Pharm. Sci. Rev.Res 27(2):63-66.
- Markkas N and Govindharajalu M. 2015. Determination of Phytocomponents in The Methanolic Extract of *Mollugo cerviana* by GC-MS Analysis. International Journal of Research in Biological sciences. 5(4): 26-29.
- Maruffo T, Nazzaro F, Mancini E, Fratianni F, coppola F, De martino L, Agostinho AB, and de feo V. 2013. Chemical Composition and Biological Activity of The essential oil from Leaves of *Moringa oleifera* Lam. Cultivated in Mozambique. Molecules 18: 10989-11000.
- Pietro Z, Maurizio S, Maurizio B. Antonella M, sergio R, carmen f, and Felice S. 2010. Essential Oil composition of stems and Fruits of *Carraluma Europea* N.E.Br. (Apocynaceae). Molecules.15:627-638.
- Saxena DK and Harinder. 2004. Uses of Bryophytes. Resonance. pp. 56-65.
- Syeda FA, Habib-Ur-Rehman, Choudahry MI, and Ata-ur-rahman. 2011. Gass chromatography Mass Spectra (GC-MS) Analysis of Petroleum eter Extract (Oil) and Bioassys of Crude Extracts of *Iris germamica*. International Journal of Genetics and Molrcular Biology 3(7):95-100.
- Vijisaral Edand Arumugam S. 2014. Analysis of Bioactive Constituents of *Indigofera suffruticosa* Leaves. Journal of Chemical and Pharmaceutical research 6(8): 294-300.