

Sequence Analysis of nrDNA ITS Region in Paşa and Şah Apple (*Malus × domestica*) Genotypes

Emre SEVİNDİK1*, Zehra Tuğba MURATHAN²

¹ Aydın Adnan Menderes University, Faculty of Agriculture, Department of Agricultural Biotechnology, South Campus, Cakmar, Aydin, Türkiye ²Malatya Turgut Özal University, Battalgazi Vocational School, Malatya, Battalgazi, Türkiye * Corresponding author: E. SEVİNDİK e-mail: ph.d-emre@hotmail.com

RESEARCH ARTICLE

Abstract

In this study, phylogenetic analysis of Şah and Paşa apple genotypes was performed using nrDNA ITS sequences. After the plant leaves were brought to the laboratory, gDNAs were obtained by genomic DNA isolation method. PCR amplification was performed using primers ITS4 and ITS5A. ITS sequences of some apple and Rosaceae species were retrieved from NCBI, bioinformatics analyzes were made with Bioedit 7.2.3, Finch TV 1.4.0 and MEGA 6.0 programs and phylogenetic trees were constructed. In the study, ITS sequence length of the Paşa apple genotype was 656 bp, and the ITS sequence length of the Şah apple genotype was 649 bp. Only in the maximum likelihood phylogenetic tree constructed using sequences of apple genotypes, Paşa and Şah apple genotypes appeared in the same clade as *Malus domestica* cultivar="Casciana'' and *Malus domestica* cultivar="Rotella''. In the phylogenetic tree generated including other species belonging to the Rosaceae family; apple genotypes *Pyrus, Cotoneaster, Crataegus, Sorbus, Eriobotrya* and *Prunus* were detected in a same clade. Overall results clearly suggested that the ITS sequences were both suitable for differentiation between the selected genera and were compatible with previous phylogenetic studies.

Keywords: Apple, ITS, phylogenetic, sequences, Türkiye

INTRODUCTION

Rosace

Received: 29 August 2023 Accepted: 31 October 2023 Published: 15 November 2023

DOI: 10.15835/buasvmcn-agr:2023.0008

© 2023 Authors. The papers published in this journal are licensed under the Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License Rosaceae family members are widely grown worldwide and include plants of economic importance, famous fresh and dried fruits with important varieties (Pan et al., 2020). Rosaceae family contains economically important fruit trees such as almonds, peaches, plums, apples, pears, roses, strawberries, and raspberries (Jiang et al., 2019; Costa and Mafra, 2022). The genus Malus belongs to the Rosaceae family and includes about 35 species all over the world (Li et al., 2020). Plants of this genus are of great economic importance, notably the domesticated apple (Malus domestica Borkh.) and some horticultural ornamental plants (eg M. halliana Koehne, M. hupehensis (Pamp.) Rehder and M. micromalus Makino) (Kalkman, 2004; Liu et al., 2022). Apple (*Malus × domestica*) is the main fruit crop of temperate regions all over the world. Türkiye, one of the leading apple producers is among the apple spreading areas (Ercisli, 2004; Balta et al., 2022; Mignard et al., 2022). World apple production is approximately 86.5 million tons and Türkiye ranks 3rd in the world with its production exceeding 4.3 million tons (FAO, 2020). According to the data of 2021, in Türkiye, 4.493,264 tons of apple production was realized with a total apple production of 2.670,318 tons, approximately 60% of this production being in Isparta (1.130,134 tons), Nigde (552.617 tons), Karaman (535.350 tons), and Antalya (451.927 tons) (TÜİK, 2021). Apple, has beneficial properties for human health as one of the frequently consumed fruits, is known as a source of micronutrients such as dietary fiber, vitamin C, monosaccharides, organic acids, minerals, phenolic compounds and phytochemical compounds (Wu et al., 2007; Özdemir et al., 2009; Gokdogan and Baran, 2017; Mignard et al., 2022). Using molecular markers representing genome level differences has further advantages over the traditional phenotypic method that they are stable and can be detected in all tissues of the plant regardless of growth, development, differentiation or environment (Agarwal et al., 2008; Devi et al., 2022). nrDNA ITS (internal transcribed spacer) includes ITS1, 5.8S and ITS2 regions of plants. The rapid rate of evolution, high variability and bi-parental inheritances make it suitable for ITS region for genetic diversity analysis at species levels (Li et al., 2020). In addition, ITS genes are considered to be the most successful primers in plant DNA barcoding. In the past, ITS regions have been used in barcode studies of many plant species (Wang et al., 2013; Olivar et al., 2014; Castro et al., 2015; Liu et al., 2017; Hürkan, 2020; Ralte and Singh, 2021). In this study, using nrDNA ITS sequences, phylogenetic analysis of Şah and Paşa apple genotypes were conducted.

MATERIALS AND METHODS

Plant material and DNA extraction

In the study, Şah and Paşa apple genotypes were collected from certain regions in Ardahan/Türkiye. The green leaves of the collected samples were brought to the Plant Biotechnology laboratory and prepared for total genomic (gDNA) isolation. Total genomic DNA samples were extracted using the DNA plant kit (GeneMark, catalog no: DP022).

PCR and sequences analyses

PCR reactions were carried out using gDNAs with ITS4 (White et al., 1990) and ITS5A (Stanford et al., 2000) primers. Primer sequences, PCR components and PCR program are given in Table 1.

Primers	DNA Sequences (5'-3')	PCR components	PCR Amplification (35 Cycles except final extension step)
ITS5A (Forward)	5'-CCTTATCATTTAGAGGAAGGAG-3'	1 μL genomic DNA 1 μL primer (ITS4 and ITS5), 10 μL master mix (PCR	94 °C / 5 min 94 °C / 45 sec 50 °C / 45 sec
ITS4 (Reverse)	5'-TCCTCCGCTTATTGATATGC-3'	dNTP, 0.75 U Taq DNA polymerase) and 7 µL dH ₂ O	72 °C / 1 min 72 °C /10 min (final extension: 1 cycle)

Table 1. nrDNA ITS primer sequences, PCR components and protocol

PCR products were also run by electrophoresis on a 0.8% agarose gel. After the ITS regions were amplified, the PCR reactions were sent to Triogen biotechnology (Istanbul/Türkiye) with ITS4 and ITS5A primers for sequence analysis. Contigs were made and analyzed from forward and reverse sequences using Bioedit 7.2.3 (Hall, 1999) and Finch TV 1.4.0 programs. Subsequently, the contig sequence was blasted in NCBI. By using MEGA 6.0 (Tamura et al., 2013) software, both ITS sequences of apple genotypes and ITS sequences of some species belonging to Rosaceae family were taken from NCBI and phylogenetic trees were obtained with the maximum likelihood (Tamura Nei model) algorithm to evaluate the degree of support for given clades, a bootstrap analysis (1000 replicates) was also applied (Felsenstein, 1985). In addition, genetic distance matrices between other apple genotypes were calculated by MEGA 6.0 program.

RESULTS AND DISCUSSIONS

nrDNA ITS gene regions are frequently used in plant systematics to detect and document phylogenetic relationships (Yeşiltaş and Kolören, 2019). In the past, the ITS region has been used in phylogenetic studies of species belonging to the Rosaceae family (Potter et al., 2000; Bortiri et al., 2001; Yang and Pak, 2006; Faghir et al., 2014; Li et al., 2014; Liu et al., 2020). In this study, the ITS sequence length of the Paşa apple genotype was determined as 656 bp, and Şah apple genotype sequence length was determined 649 bp. Genotypes were uploaded to NCBI. Accession numbers: Paşa apple: OR704536, Şah apple: OR704543. To understand the phylogenetic relationship between Şah and Paşa genotypes and some apple genotypes, ITS sequences were retrieved from NCBI database for *Malus domestica* cultivar="Rotella (MH633851.1), *Malus domestica* cultivar="Casciana (MH633843.1), mestica* cultivar="Casciana (MH633848.1), *Malus domestica* cultivar="Bramley's Seedling (AF186479.1), *Malus domestica* cultivar Leathercoat (AF186477.1), *Malus domestica* cultivar Reinette (AF186478.1) *Malus × domestica* cultivar Ralls (EU150074.1), and *Malus domestica* cultivar Autumn Pearmain (AF186481.1), and a maximum likelihood phylogenetic tree was constructed. The phylogenetic tree consists of 2 clades (Figure 1). Clade 1 consisted of Casciana, Rotella cultivars and Şah and Paşa apple genotypes, and this group received a bootsrapt value of 99%. Clade 2 consists of the cultivars Bramleys, Ralls, Autumn Pearmain, Leathercoat, and Reinette.



Figure 1. The maximum likelihood (Tamura-Nei model) tree with bootstrap values (1000 replicates) generated using ITS sequences of some apple genotypes

According to the genetic distance method based on ITS sequences, the smallest distance was determined as 0.000, while the highest distance was determined as 0.037 (Table 2). Uzun et al. (2016), examined the genetic relationships of Turkish apple genotypes using ISSR markers. In their study, it was determined that the Şah apple and Paşa apple genotypes were in the same group. Sevindik et al. (2018), revealed the genetic diversity of apple genotypes with the ISSR marker technique. In the UPGMA dendogram, they determined that the Paşa and Şah apple genotypes were related to each other within the same clade. Sevindik et al. (2019) determined the molecular characterization of apple genotypes with cpDNA *trn*L-F sequences, and they found that the Paşa and Şah genotypes were in the same group.

Paşa apple		-	-	-	-	-	-	-	-	-	-	-
Şah apple		-	-	-	-	-	-	-	-	-	-	-
Malus domestica (USA)		0.026	-	-	-	-	-	-	-	-	-	-
Malus domestica (cultivar Rotella)		0.002	0.028	-	-	-	-	-	-	-	-	-
Malus domestica (cultivar Casciana)		0.003	0.026	0.005	-	-	-	-	-	-	-	-
Malus domestica (cultivar Rotella)		0.005	0.032	0.007	0.009	-	-	-	-	-	-	-
Malus domestica (cultivar Casciana)		0.005	0.032	0.007	0.009	0.010	-	-	-	-	-	-
Malus domestica (cultivarCasciana)		0.007	0.033	0.009	0.010	0.012	0.012	-	-	-	-	-
Malus domestica (cultivar Bramleys)		0.026	0.000	0.028	0.026	0.032	0.032	0.033	-	-	-	-
Malus domestica (cultivar Leathercoat)		0.030	0.017	0.032	0.033	0.035	0.035	0.037	0.017	-	-	-
Malus domestica (cultivar Reinette)		0.028	0.016	0.030	0.032	0.033	0.033	0.035	0.016	0.012	-	-
Malus x domestica (cultivar Ralls)		0.024	0.014	0.026	0.028	0.030	0.030	0.031	0.014	0.016	0.014	-
Malus domestica (cultivar Autumn Pearmain)		0.021	0.010	0.023	0.025	0.026	0.026	0.028	0.010	0.012	0.010	0.007

Table 2. Pairwise genetic distance (Tamura-Nei model) matrix obtained from nuclear ribosomal DNA internal transcribed spacer sequences (together with other *Malus domestica* cultivars)

In addition, in our study, ITS sequences of the taxa from Rosaceae family obtained from NCBI: Pyrus pyrifolia (KC895397.1), Pyrus ussuriensis (EU150070.1), Sorbus americana (FJ810037.1), Sorbus aucuparia (KY661717.1), Cotoneaster melanocarpus (JQ392405.1), Cotoneaster salicifolius (JQ392378.1), Prunus spinosa (EU669100.1), Prunus armeniaca (HF969271.1), Eriobotrya japonica (KJ170775.1), Eriobotrya cavaleriei (KJ170784.1), Potentilla anatolica (KT985669.1), Potentilla kotschyana (KT985720.1), Rosa arvensis

(KM353028.1), Rosa canina (KM353044.1), Rubus saxatilis (AF055747.1), Rubus idaeus (AF055755.1), Crataegus maximowiczii (EU683914.1), *Crataegus heldreichii* (EU500465.1), *Alchemilla faroensis* (EU072527.1), *Alchemilla vulgaris* (EU072573.1), *Fragaria vesca* (AF163510.1), *Fragaria × ananassa* (AF163494.1) were used to construct a maximum likelihood phylogenetic tree (Figure 2).



Figure 2. The maximum likelihood (Tamura-Nei model) tree generated using ITS sequences and other species sequences retrieved from NCBI

Apple genotypes, *Pyrus, Cotoneaster, Crataegus, Sorbus, Eriobotrya* and *Prunus* species detected in a same clade. The nrDNA ITS results showed discrimination at the genus level. Verbylaitė et al. (2006) determined *Malus* species together with *Pyrus, Kageneckia, Pyracantha, Cydonia, Sorbus,* and *Photinia* species using cpDNA *trnL-trnF* analyses. Sun et al. (2018), in their phylogenetic analysis, used 15 chloroplast gene regions, and *Malus, Docyniopsi, Docynia,* and *Eriolobus* were identified together. Sevindik and Murathan (2023) with cpDNA *rbcL* sequences, and Sevindik et al. (2023) with cpDNA *trnL* intron and *trnL*-F sequences detected *Malus* species together with *Cotoneaster, Sorbus, Prunus,* and *Pyrus.* Our results were consistent with previous cpDNA results. According to Judd et al. (1999), a combination of nuclear and chloroplast sequence data can provide complementary insights into the phylogenetic relationships of a genus.

CONCLUSIONS

As a result, in phylogenetic trees with ITS sequences, Paşa and Şah apple genotypes were detected together with *Malus domestica* cultivar="Casciana" and *Malus domestica* cultivar="Rotella", *Pyrus, Cotoneaster, Crataegus, Sorbus, Eriobotrya* and *Prunus* species in the same clade. ITS sequences were both suitable for differentiation between genera and were compatible with previous phylogenetic studies.

Author Contributions: E.S. and Z.T.M. Conceived and designed the analysis; Z.T.M. Collected the data; E.S. Performed the analysis; E.S. and Z.T.M. Wrote the paper.

Funding Source: The authors received no specific funding for this work.

Conflicts of Interest

The authors declare that they do not have any conflict of interest.

REFERENCES

- 1. Agarwal M, Shrivastava N, Padh H. Advances in molecular marker techniques and their applications in plant sciences. Plant Cell Reports. 2008; 27:617–631. <u>https://doi.org/10.1007/s00299-008-0507-z</u>
- 2. Balta MF, Karakaya O, Kurt H, Yılmaz M, Uzun S, Balta F. Phytochemical variation of native apple germplasm resources from the Eastern Black sea region, Turkey. Erwerbs-Obstbau. 2022; 64(4):685-695. https://doi.org/10.1007/s10341-022-00735-1
- 3. Bortiri E, Oh SH, Jiang JG, Baggett S, Granger A, Weeks C, Buckingham M, Potter D, Parfitt DE. Phylogeny and systematics of *Prunus* (Rosaceae) as determined by sequence analysis of ITS and the chloroplast trnL-trnF Spacer DNA. Systematic Botany. 2001; 26(4):797–807. <u>https://doi.org/10.1043/0363-6445-26.4.797</u>
- 4. Castro C, Hernandez A, Alvarado L, Flores D. DNA barcodes in fig cultivars (*Ficus carica* L.) using ITS regions of ribosomal DNA, the *psbA-trn*H spacer and the *mat*K coding sequence. American Journal of Plant Sciences. 2015; 6(01):95.
- 5. Costa J, Mafra I. Rosaceae food allergy: a review. Critical Reviews in Food Science and Nutrition. 2022; 1-38. https://doi.org/10.1080/10408398.2022.2045897
- Devi ML, Thorat SS, Devi KK, Sharma KC, Singh YD, Mishra A, Das S. Internal Transcribed Spacer (ITS) region of nuclear ribosomal DNA as a suitable DNA barcode for identification of *Zanthoxylum armatum* DC from Manipur. Molecular Biotechnology. 2022; 64(12):1454-1467. <u>https://doi.org/10.1007/s12033-022-00518-9</u>
- Ercisli S. A short review of the fruit germplasm resources of Turkey. Genetic Resources and Crop Evolution, 2004; 51(4):419–435. <u>https://doi.org/10.1023/B:GRES.0000023458.60138.79</u>
- Faghir MB, Attar F, Farazmand A, Osaloo SK. Phylogeny of the genus *Potentilla* (Rosaceae) in Iran based on nrDNA ITS and cpDNA *trn*L-F sequences with a focus on leaf and style characters' evolution. Turkish Journal of Botany. 2014; 38(3):417-429. <u>https://doi.org/10.3906/bot-1303-67</u>
- 9. Food and Agriculture Organization. Production quantities by country. FAO; 2020. https://www.fao.org/faostat/en/#data/QCL: accessed on 22.08.2022.
- Felsenstein J. Confidence limits on the phylogenies: an approach using the bootstrap. Evolution. 1985; 39:783– 791. <u>https://doi.org/10.1111/j.1558-5646.1985.tb00420.x</u>
- 11. Gokdogan O, Baran M (2017). Determination of energy use efficiency of some apple (*Malus × domestica*) production in Turkey: a case study of Eğirdir Region. Erwerbs-Obstbau. 2017; 59(1):13-18. https://doi.org/10.1007/s10341-016-0290-x
- 12. Hall TA. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. Nucleic Acids Symp Ser. 1999; 41:95–98.
- Hürkan K. Analysis of various DNA barcodes on the Turkish protected designation of origin apricot "Iğdır Kayısısı" (*Prunus armeniaca* cv. Şalak). Turkish Journal of Agriculture - Food Science and Technology. 2020; 8(9):1982–1987. <u>https://doi.org/10.24925/turjaf.v8i9.1982-1987.3594</u>
- Jiang F, Zhang J, Wang S, Yang L, Luo Y, Gao S, Zhang M, Wu S, Hu S, Sun H, Wang Y. The apricot (*Prunus armeniaca* L.) genome elucidates Rosaceae evolution and beta-carotenoid synthesis. Horticulture Research. 2019; 6:128. <u>https://doi.org/10.1038/s41438-019-0215-6</u>
- 15. Judd WS, Campbell CS, Kellogg EA, Stevens PF. Plant systematics: a phylogenetic approach. Sinauer Associates, Massachusetts, USA. 1999
- 16. Kalkman C. Rosaceae. In: Kubitzki, K. ed. The Families and Genera of Vascular Plants. Vol. VI. Flowering Plants. Dicotyledons. Celastrales, Oxalidales, Rosales, Cornales, Ericales. Berlin: Springer; 2004. p. 343–386.
- 17. Li F, Fan Q, Li Q, Chen S, Guo W, Cui D, Liao W. Molecular phylogeny of *Cotoneaster* (Rosaceae) inferred from nuclear ITS and multiple chloroplast sequences. Plant Systematics and Evolution. 2014; 300: 1533-1546. https://doi.org/10.1007/s00606-014-0980-5
- 18. Li Y, Liu Y, Wu P, Zhou S, Wang L, Zhou S. The complete chloroplast genome sequence of Malus toringoides
(Rosaceae). MitochondrialDNAPartB.2020.5(3):2787-2789.https://doi.org/10.1080/23802359.2020.1780977

- Li W, Li B, Zhang P, Hu D, Wang A. Potential biological mechanisms underlying the endangered status of *Glehnia littoralis* revealed by nrDNA ITS and RAPD analyses. Biotechnology & Biotechnological Equipment. 2020; 34(1):1243-1251. <u>https://doi.org/10.1080/13102818.2020.1830713</u>
- 20. Liu ZF, Ci XQ, Li L, Li HW, Conran JG, Li J. DNA barcoding evaluation and implications for phylogenetic relationships in Lauraceae from China. PloS one. 2017; 12(4):e0175788. https://doi.org/10.1371/journal.pone.0175788
- 21. Liu BB, Liu GN, Hong DY, Wen J. *Eriobotrya* belongs to *Rhaphiolepis* (Maleae, Rosaceae): evidence from chloroplast genome and nuclear ribosomal DNA data. Frontiers in Plant Science. 2020; 10:1731. https://doi.org/10.3389/fpls.2019.01731
- 22. Liu BB, Ren C, Kwak M, Hodel RG, Xu C, He J, et al. Phylogenomic conflict analyses in the apple genus *Malus* sl reveal widespread hybridization and allopolyploidy driving diversification, with insights into the complex biogeographic history in the Northern Hemisphere. Journal of Integrative Plant Biology. 2022; 64(5):1020-1043. https://doi.org/10.1111/jipb.13246
- 23. Mignard P, Beguería S, Giménez R, Font i Forcada C, Reig G, Moreno MÁ. Effect of genetics and climate on apple sugars and organic acids profiles. Agronomy. 2022; 12(4):827. <u>https://doi.org/10.3390/agronomy12040827</u>
- 24. Olivar JEC, Brillantes RY, Rubite RR, Alejandro GJD. Evaluation of three candidate DNA barcoding loci in selected *Ficus* L.(Moraceae). International Journal of Scientific and Technology Research. 2014; 3(9):43-48.
- 25. Özdemir Y, Akçay ME, Özkan M. Fonksiyonel bir meyve olarak elma. Tarım Bilimleri Araştırma Dergisi. 2009; 1:51-55.
- 26. Pan M, Zhu H, Bonthond G, Tian C, Fan X. High diversity of *Cytospora* associated with canker and dieback of Rosaceae in China, with 10 new species described. Frontiers in Plant Science. 2020; 11:690. https://doi.org/10.3389/fpls.2020.00690
- 27. Potter D, Luby JJ, Harrison RE. Phylogenetic relationships among species of *Fragaria* (Rosaceae) inferred from non-coding nuclear and chloroplast DNA sequences. Systematic Botany. 2000; 25(2):337-348. https://doi.org/10.2307/2666646
- 28. Ralte L, Singh YT. Use of *rbc*L and ITS2 for DNA barcoding and identification of Solanaceae plants in hilly state of Mizoram, India. Research on Crops. 2021; 22(3):616-623. <u>https://doi.org/10.31830/2348-7542.2021.110</u>
- 29. Sevindik E, Uysal H, Murathan ZT. Genetic diversity based on ISSR markers of apple genotypes in Ardahan/Turkey. Notulae Scientia Biologicae. 2018; 10(4):554-558. <u>https://doi.org/10.15835/nsb10410347</u>
- 30. Sevindik E, Murathan ZT, Filiz S, Yalçin K. Molecular characterization based on chloroplast (*trn*L-F) DNA sequence of the apple genotypes in Ardahan/Turkey. Bangladesh Journal of Botany. 2019; 48(4):1099-1106. https://doi.org/10.3329/bjb.v48i4.49058
- Sevindik E, Efe F, Murathan ZT. Molecular genetic diversity and phylogenetic investigation of *Pyrus communis* L.(Rosaceae) genotypes using cpDNA sequences with RAPD and ISSR analyses. Erwerbs-Obstbau. 2023. 65(2):231-240. <u>https://doi.org/10.1007/s10341-022-00798-0</u>
- 32. Sevindik E, Murathan ZT. DNA Barcoding and Sequence Analysis of the cpDNA *rbc*L Region in Some *Pyrus communis* L. (Rosaceae) Genotypes from Ardahan/Türkiye. Erwerbs-Obstbau. 2023; 65:1315–1320. https://doi.org/10.1007/s10341-023-00860-5
- 33. Sun J, Shi S, Li J, Yu J, Wang L, Yang X, Ling G, Zhou S. Phylogeny of Maleae (Rosaceae) based on multiple chloroplast regions: implications to genera circumscription. BioMed Research International. 2018; 7627191. doi: 10.1155/2018/7627191
- 34. Stanford AM, Harden R, Parks CR. Phylogeny and biogeography of *Juglans* (Juglandaceae) based on *mat*K and ITS sequence data. American Journal of Botany. 2000; 87(6):872–882. <u>https://doi.org/10.2307/2656895</u>
- 35. Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. MEGA6: molecular evolutionary genetics analysis version 6.0. Molecular Biology and Evolution. 2013; 30(12):2725–2729. <u>https://doi.org/10.1093/molbev/mst197</u>
- 36. TÜİK. Türkiye İstatistik Kurumu Bitkisel Üretim İstatistikleri Veri Tabanı; 2021. <u>https://biruni.tuik.gov.tr/medas/?kn=92&locale=tr</u>; accessed on 22.08.2022.
- Uzun A, Ozongun S, Gulsen O, Yılmaz KU, Kaymak S, Ercisli S. Determination of genetic relatedness among Turkish apple germplasm based on ISSR markers. Journal of Applied Botany and Food Quality. 2016; 89:82-88. <u>https://doi.org/10.5073/JABFQ.2016.089.010</u>
- 38. Verbylaitė R, Ford-Lloyd B, Newbury J. The phylogeny of woody Maloideae (Rosaceae) using chloroplast trnLtrnF sequence data. Biologija. 2006; 52(1).

- 39. Wang M, Zhao HX, Wang L, Wang T, Yang RW, Wang XL, Zhou YH, Ding CB, Zhang L. Potential use of DNA barcoding for the identification of *Salvia* based on cpDNA and nrDNA sequences. Gene. 2013; 528(2):206-215. https://doi.org/10.1016/j.gene.2013.07.009
- 40. White TJ, Bruns T, Lee S, Taylor J. Amplifications and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand J, Sninsky T (eds) PCR Protocols: A Guide to Methods and Applications. Academic Press, San Diego, CA, USA; 1990. p. 315–322.
- 41. Wu J, Gao H, Zhao L, Liao X, Chen F, Wang Z, Hu X. Chemical compositional characterization of some apple cultivars. Food Chemistry. 2007; 3:88–93. <u>https://doi.org/10.1016/j.foodchem.2006.07.030</u>
- 42. Yang JY, Pak JH. Phylogeny of Korean *Rubus* (Rosaceae) based on ITS (nrDNA) and *trn*L/F intergenic region (cpDNA). Journal of Plant Biology. 2006; 49:44-54. <u>https://doi.org/10.1007/BF03030787</u>
- 43. Yeşiltaş BN, Kolören O. Molecular characterization of *Sicyos* species in Ordu and Giresun provinces. Turkish Journal of Weed Science. 2019; 22(1):37–44.