

# A First Report of *Sclerotinia sclerotiorum* Causing Forsythia Twig Blight in Romania

Andreea-Mihaela Florea <sup>1</sup>, Andrei-Mihai Gafencu <sup>1</sup>, Florin-Daniel Lipşa <sup>2</sup> , Iulian Gabur <sup>1,\*</sup> and Eugen Ulea <sup>1,\*</sup>

<sup>1</sup> Department of Plant Science, Faculty of Agriculture, “Ion Ionescu de la Brad” Iasi University of Life Sciences, 3 Mihail Sadoveanu Alley, 700489 Iasi, Romania; amflorea@uaiasi.ro (A.-M.F.); agafencu@uaiasi.ro (A.-M.G.)

<sup>2</sup> Department of Food Technologies, Faculty of Agriculture, “Ion Ionescu de la Brad” Iasi University of Life Sciences, 3 Mihail Sadoveanu Alley, 700489 Iasi, Romania; flipsa@uaiasi.ro

\* Correspondence: gaburi@uaiasi.ro (I.G.); eulea@uaiasi.ro (E.U.)

**Abstract:** *Sclerotinia sclerotiorum* (Lib.) de Bary (1884) is a fungal plant pathogen with worldwide distribution and a varying host range from different botanical families. It can cause damage to a large variety of crops such as sunflower, soybean, dry bean, canola, some vegetables, and ornamental plants. This article reports the occurrence of twig blight on the forsythia plant from the NE region of Romania. The disease was observed on *Forsythia × intermedia* Zab. plants from the Arboretum Park of the Iasi University of Life Sciences (IULS), located in Iasi City, Romania. Infected tissue was investigated through morphological characteristics using Sanger sequencing. Genomic DNA was extracted from the isolate obtained from naturally infected plants, and the ribosomal internal transcribed spacer region was amplified using the ITS1, ITS2, and LSU D1 and D2. Based on the results of this study, molecular and morphological data suggest that Forsythia twig blight can be caused by *S. sclerotiorum*. Constant monitoring of *Sclerotinia sclerotiorum* across multiple hosts and time intervals will reduce potential spread and future economic losses in cultivated species.

**Keywords:** *Sclerotinia sclerotiorum*; new host; *Forsythia* spp.; Sanger sequencing; ornamental plants



**Citation:** Florea, A.-M.; Gafencu, A.-M.; Lipşa, F.-D.; Gabur, I.; Ulea, E. A First Report of *Sclerotinia sclerotiorum* Causing Forsythia Twig Blight in Romania. *Plants* **2023**, *12*, 3516. <https://doi.org/10.3390/plants12203516>

Academic Editor: Tika Adhikari

Received: 11 September 2023

Revised: 30 September 2023

Accepted: 3 October 2023

Published: 10 October 2023



**Copyright:** © 2023 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<https://creativecommons.org/licenses/by/4.0/>).

## 1. Introduction

*Sclerotinia sclerotiorum* (Lib.) de Bary (1884) is a fungal pathogen that causes substantial losses to a wide range of hosts worldwide. *Sclerotinia sclerotiorum* (SS) is capable of infecting many plant species from different botanical families and can cause damage to a large variety of crops, such as sunflower, soybean, dry bean, canola, some vegetables, and ornamental plants [1]. Often, yield losses due to *S. sclerotiorum* infection exceed 20–35%, but the literature describes more documented cases of losses over 50% and up to 80–100% in temperate climates [2]. *S. sclerotiorum* has no specificity for a particular host and is able to infect monocotyledonous and dicotyledonous plant species [2,3]. Recent studies have found that *S. sclerotiorum* can also grow endophytically in monocots, including rice, wheat, maize, barley, and oat [4–6]. Based on infected host plant species, bioclimatic data, and geographic locations, economic losses caused by *S. sclerotiorum* are substantial and can vary extremely [7,8].

In Romania, *Sclerotinia sclerotiorum* is known for causing significant damage to plants in field crops and forced crops in greenhouses, warehouses, and silos [9]. The number of *Sclerotinia sclerotiorum* plant hosts at the national level is certainly higher [10], but reports on the presence of the fungus *Sclerotinia sclerotiorum* on woody ornamental plants are extremely scarce. Although *Sclerotinia sclerotiorum* is considered the main pathogen for a large number of herbaceous and woody ornamental species, only some information on the confirmed infection process and pathogenesis is available for *Sclerotinia sclerotiorum* in woody ornamental plants. In the scientific literature, there are numerous examples of ornamental plant infections; however, most articles do not include a description of the symptoms or severity of the infection [11,12].

More than 60 different *Sclerotinia sclerotiorum* disease symptoms have been reported in the literature as related to a broad host range that comprises agronomically important plant species [1]. *S. sclerotiorum* can infect multiple plant organs and survive in vegetable rests and soil for a long period, as sclerotia [13]. Sclerotia can infect new hosts in asexual form, as mycelia, or self-fertilized sexual reproduction, as ascospores, providing *S. sclerotiorum* with the possibility of accessing a wide range of potential targets [14].

The *Forsythia* genus is a group of plants in the olive family (*Oleaceae*) with around 11 species that are primarily native to eastern Asia, with one species from Europe. In Romania, *Forsythia* spp. Vahl is a perennial plant, appreciated and well known only as an ornamental plant named “golden-bells”. The same plant variety is also used for a wide range of Chinese medicines and health diets [15].

The majority of reports of *S. sclerotiorum* in ornamental plants come from the commercial production of plants in greenhouses or nurseries [16]. Therefore, less is known about the epidemiology of *S. sclerotiorum* in ornamental plants grown in natural landscapes [17]. In flowering woody ornamentals, descriptions of disease indicate that ascospores initiate infection through flowers. In *Forsythia* spp., plant water-soaked lesions form on flower petals in cool and moist weather conditions that are favorable for carpogenic germination. Afterward, *Forsythia* spp. infections progress into branch tissue, resulting in girdling cankers and the wilting of individual branches [18]. Moreover, in 2023, Munoz et al. [19] described *S. sclerotiorum* as the cause of rose bent neck on commercial cut roses (*Rosa hybrida* L.), while Rahman et al. (2020) [20] observed flower and blossom blight due to white mold disease on garden roses (*Rosa chinensis*) in Bangladesh. *Sclerotinia sclerotiorum* (Lib.) de Bary infection of ornamental plants of *Forsythia* spp. Vahl was initially suggested in samples collected in spring 2021 from the Arboretum Park of the Iasi University of Life Sciences (IULS), located in Iasi City, Romania [21].

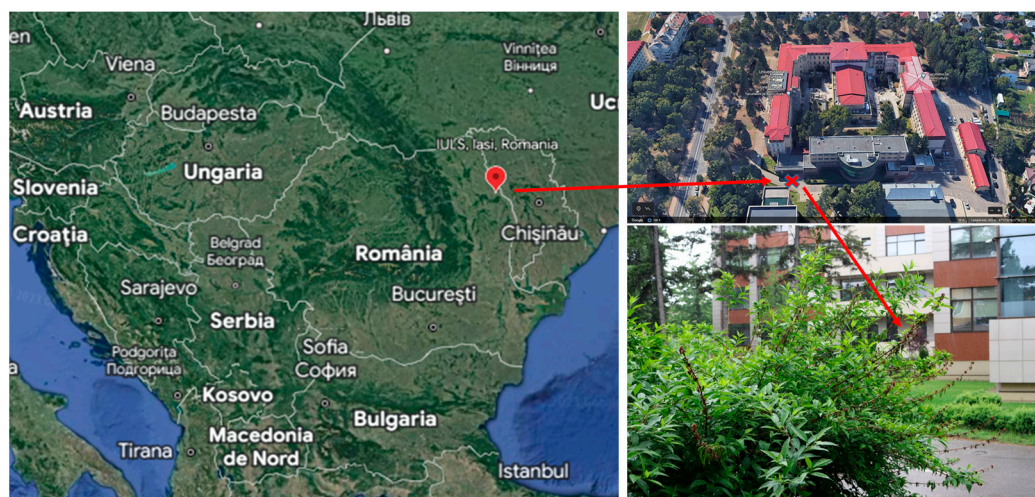
This article describes the first confirmed *Sclerotinia sclerotiorum* infection on ornamental plants of *Forsythia × intermedia* Zab. (*F. suspensa* × *F. viridissima*) tissue [22] through morphological characteristics and genetic investigation using PCR and Sanger sequencing. Genomic DNA was amplified using ribosomal internal transcribed spacer region-specific primers, and samples were analyzed using sequencing techniques.

## 2. Results

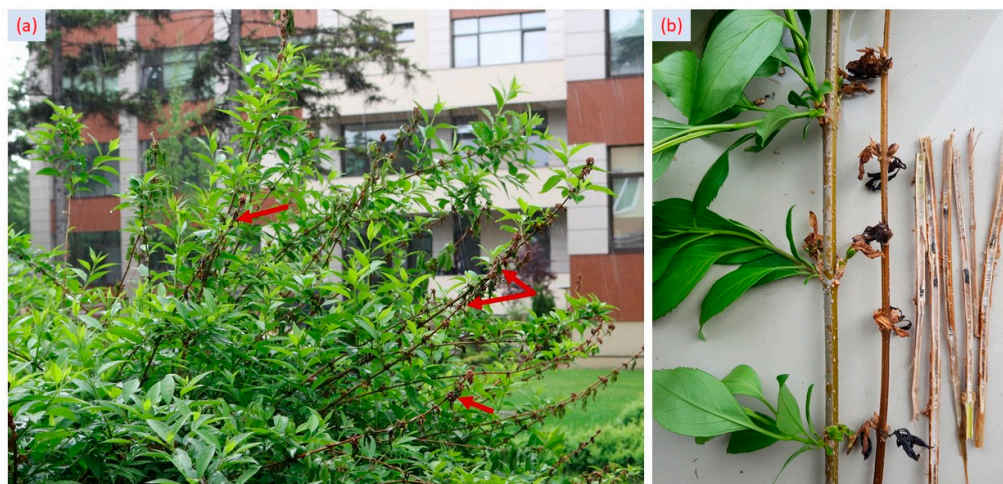
### 2.1. Fungal Infection and Symptoms

In the spring of 2022, environmental conditions were extremely favorable for *Sclerotinia sclerotiorum* (Lib.) de Bary initiation and evolution on *Forsythia × intermedia* Zab. ornamental plants from the Arboretum Park of the Iasi University of Life Sciences (IULS), located in Iasi, Romania (Figure 1).

*S. sclerotiorum* is a monocyclic pathogen, so there is only one primary mode of infection, depending on the type of infection incited [23]. Climatic conditions and plant vegetative growth stages have a major role in the infection spreading through ascospores. The floral elements, especially the stamens and the pistil, are highly sensitive to infection by ascospores. The ascospores completely colonized the mature and senescent flowers, and after a few days, the mycelium that colonized these flowers caused further infections through direct contact with the leaf and stem [24]. Water-soaked lesions progressed into branch tissue, resulting in the wilting of individual branches (Figure 2a). Necrotic tissues from the affected branches were examined, and we observed that they were covered with patches of fluffy white mycelia and sclerotia (Figure 2b).



**Figure 1.** *Sclerotinia sclerotium* on *Forsythia × intermedia* Zab. ornamental plant from the Arboretum Park of the Iasi University of Life Sciences (IULS), located in Iasi, Romania (GPS coordinates: [www.google.com/maps](http://www.google.com/maps), accessed on 6 April 2023, 47°11'30.7" N 27°33'25.2" E). Red arrows represent the exact location of the plant material investigated. Red x marks the physical location of the ornamental plant.

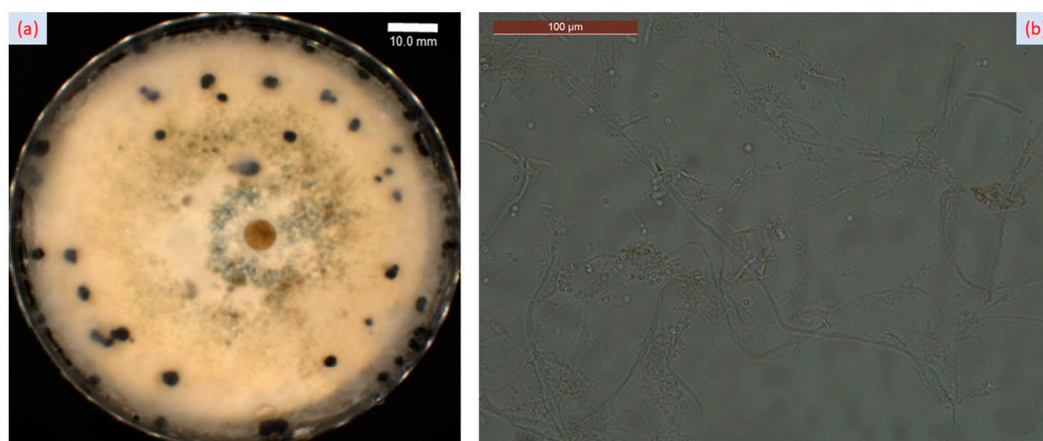


**Figure 2.** *Sclerotinia sclerotium* on *Forsythia*. (a) Wilted branches of *Forsythia* attacked by *Sclerotinia sclerotium*; (b) healthy tissues (left side) and necrotic tissues of *Forsythia* with patches of white mycelia and sclerotia (right side). Red arrows in (a) mark the sample extracted for further investigations and wilting of individual branches.

To investigate the presence of *Sclerotinia sclerotiorum*, the fungus was isolated from infected tissues of *Forsythia* and cultured on a PDA medium. The fungus started to produce white masses of mycelium that grew to the edge of the Petri dish (Figure 3a). The size of the mycelium masses became bigger, and their colors became darker as time progressed. Furthermore, we observed that many black sclerotia formed on the Petri dish (Figure 3a). Under microscopic analysis, *Sclerotinia sclerotiorum* showed hyaline hyphae, branched and multinucleate (Figure 3b).

Four days after the pathogenicity assay, inoculated branches initially displayed discoloration, followed by water-soaked lesions that progressively developed at a distance of 5–10 cm from the contact point. Then, in a short time, the inoculated branches completely withered.

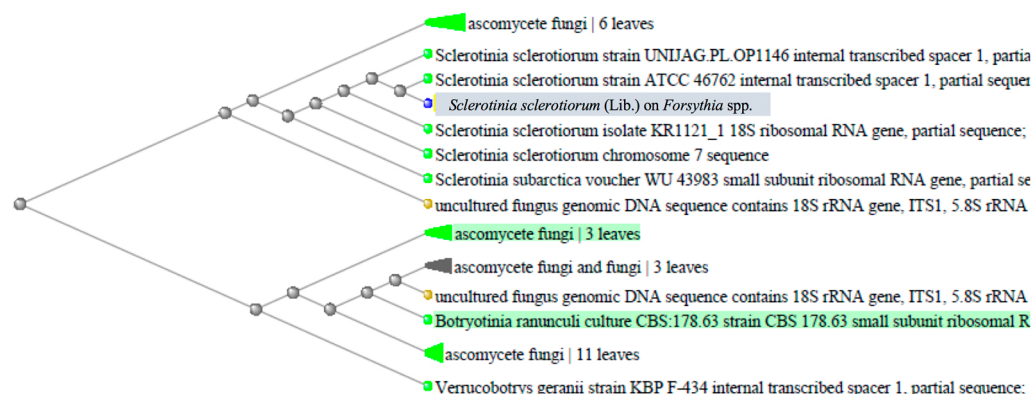




**Figure 3.** *Sclerotinia sclerotiorum*. (a) White masses of mycelium and sclerotia developed on PDA medium plate; (b) hyphae of *Sclerotinia sclerotiorum* at 400× magnification.

## 2.2. Sequence Alignment and Phylogenetic Analysis/Molecular Characterization

Genomic DNA isolated from plant material was subjected to PCR amplification of the 5.8S rRNA region, followed by Sanger sequencing. The PCR amplicon sizes obtained from the selected primers, ITS1/ITS2 and LSU D1/D2, were 580 bp. Information regarding the similarity of the investigated sequence was obtained by conducting BLAST searches (*blastn*) [25] against the GenBank database [26]. Genetic distance was investigated using simple neighbor-joining clustering based on uncorrected p-distances. BLAST results indicated that the nucleotide sequence on the selected sample had 99.91% similarity with the *Sclerotinia sclerotiorum* chromosome 7 sequence (Sequence ID: CP017820.1) and the *Sclerotinia sclerotiorum* isolate KR1121\_1 (Sequence ID: KC311494.1). The phylogenetic analysis revealed that isolate SS\_R was phylogenetically identical to other *S. sclerotiorum* isolates available in the GenBank database (Figure 4 and Supplementary File S1).



**Figure 4.** Genetic distance tree using the NCBI-BLAST tool, with “neighbor joining” method, max seq difference = 0.5. The sample analyzed in this paper is marked with grey. Sample sequence has a length of 580 bp and was amplified using PCR specific primers ITS1/ITS2 and LSU D1/D2.

A set of ten publicly available *S. sclerotiorum* sequences from different locations (accession numbers: UNIJSG.PL.OP1146, ATCC46762, KR1121\_1, WU43983, KC311494.1), together with uncultured fungal genomic DNA sequences, were grouped together and formed a cluster. High similarities were observed among the SS\_R isolate and other ascomycete fungi sequences obtained from leaf samples [19]. Phylogenetic analysis of the Forsythia twig blight sample with other fungal isolates from NCBI GeneBank indicated a high similarity of isolate SS\_F to other *S. sclerotiorum* isolates. Based on the results of this study, molecular and morphological data suggest that Forsythia twig blight is caused by *Sclerotinia sclerotiorum*.

### 3. Discussion

The world's cropping system has faced increasing challenges in recent years as climate change has advanced, creating optimal developmental conditions for fungal outbreaks, for example, white mold disease caused by *Sclerotinia sclerotiorum* [27]. The severe economic consequences of white mold infections can be addressed in major crops through integrated agro-management strategies. However, these strategies rely on a vast amount of fungicides that, in some cases, could trigger pathogen resistance to chemical compounds [28]. Moreover, increased quantities of chemical fungicides could also affect environmental and microbiota development at the plant level, suggesting that new disease management programs are necessary to avoid potential outbreaks or undesirable environmental, cultural, or biological side effects.

It is well known that *Sclerotinia sclerotiorum* is a typical necrophytic pathogen that produces severe economic losses, and recent studies showed that this fungus induces the production of reactive oxygen species (ROS), which leads to necrosis in the host, allowing the pathogen to absorb nutrients from the dead tissues. An important cofactor for enzymes scavenging ROS is represented by copper, which is an essential nutrient for microbial pathogens [29,30]. In fact, the metals most recognized for their crucial role in fungal homeostasis are elements like copper (Cu), iron (Fe), zinc (Zn), and manganese (Mn), which are extremely necessary for various biochemical processes, usually as enzyme cofactors. Other non-essential elements with an unknown biological role that can adversely affect microorganisms are cadmium (Cd), lead (Pb), and mercury (Hg) [31,32].

To our knowledge, this study represents the first report of *Sclerotinia sclerotiorum* infection in *Forsythia × intermedia* Zab. (*F. suspensa* × *F. viridissima*). Plant samples were collected from the Arboretum Park of the Iasi University of Life Sciences (IULS), and fungal infection was identified and validated through molecular analysis. The primary focus was given to the sclerotial mycoparasite morphological and molecular investigation and mode of action against *Forsythia × intermedia* Zab., an ornamental species frequently encountered in NE Romanian areas. Plant developmental stages, temperature, and humidity have an important role in the onset of *Sclerotinia sclerotiorum* infections [19]. In recent years, climatic changes like cool, wet weather in spring and variable winter conditions in the NE region of Romania have led to a more frequent appearance and increased incidence of *Sclerotinia sclerotiorum* on numerous ornamental plants.

Our understanding of the process of infection and pathogenesis for *Sclerotinia*-induced disease is lacking key components, while details regarding physiology, biochemistry, and molecular aspects are limited in the literature [33]. In this study, morphological analysis showed that the fungus *Sclerotinia sclerotiorum* infects the shoots of *Forsythia* only through the withering flowers, especially the stamens and the pistil, and extends up and down the woody branches from those points, causing wilting on the individual branches. Fungal-caused lesions progressed into branch tissue and led to a wilting effect. Taking into account the infection mechanism, effective management practices are necessary to limit the presence of this pathogen in other potential hosts and reduce the possibility of outbreaks.

Molecular identification of the isolated fungus, *Sclerotinia* sp., was further confirmed through PCR using ITS1, ITS2, and LSU D1 and D2 primers. PCR products produced amplicons of various sizes of about 586 bp, corresponding to the ITS and LSU regions. The amplicon was sequenced using Sanger. Munoz et al. observed similar results and reported the molecular characterization of *Sclerotinia sclerotiorum* using ITS 1 and ITS 4 on commercial cut roses (*Rosa hybrida* L.) [19].

Currently, twig blight symptoms are rare, and during this evaluation, we observed a relatively small area affected. However, constant monitoring of *Sclerotinia sclerotiorum* across multiple hosts and times will reduce the potential spread and future economic losses in cultivated species.

## 4. Materials and Methods

### 4.1. Diseased Sample Collection and Fungus Isolation

*Sclerotinia sclerotiorum* was observed on 18 May 2022 in *Forsythia* spp. plants from the Arboretum Park of the Iasi University of Life Sciences (IULS), located in Iasi, Romania (GPS coordinates: [www.google.com/maps](https://www.google.com/maps), accessed on 6 April 2023, 47°11'30.7" N 27°33'25.2" E). Infected tissues of the *Forsythia* host plant were collected and investigated in the research laboratory of the phytopathology discipline within the "Ion Ionescu de la Brad" Iasi University of Life Sciences (IULS). After a putative micromycete identification based on visual symptoms, fungal morphology, microscopic preparations, and a specialized guidebook, we followed a standard procedure [34] for fungal isolation to confirm the presence of *Sclerotinia sclerotiorum* in *forsythia* plants. Infected tissues of *Forsythia* × *intermedia* Zab. [22] were cut into small pieces and then rinsed 3~4 times with diluted water after being treated with 70% (*v/v*) ethanol for 2~3 s. The treated tissues were transferred to potato dextrose agar (PDA) medium and cultured at 25 °C. Then, whitish, cottony mycelium was transferred into PDA plates for 10 days to obtain pure cultures. White masses of mycelium developed to the edge of the Petri dish, and finally, many black sclerotia formed on the PDA medium of the selected isolate coded SS\_F.

### 4.2. Pathogen Identification—Morphologic

Kock's postulates were fulfilled using the isolate SS\_F to inoculate six branches with a length of 25–30 cm of *Forsythia* × *intermedia* Zab. by placing a 1 cm agar plug with actively growing hyphal tips from a colony of *Sclerotinia sclerotiorum*. Six other branches were inoculated with an agar plug with no hyphae on the branch. All branches were maintained in a disinfected area with conditions for relative humidity close to 90% and a temperature of 28 °C with 10 h of light daily, after which water-soaked lesions similar to those in Arboretum Park samples were observed on the six inoculated branches.

### 4.3. Molecular Characterization of *Sclerotinia Sclerotiorum* Isolates

Genomic DNA was isolated using dried specimen samples of naturally infected plant material with the fungi. DNA was purified using the PCR BIO Rapid Extract Lysis Kit (<https://pcrbio.com/row/products/dna-extraction/pcrbio-rapid-extract-lysis-kit/>, accessed on 6 April 2023, PCR Biosystems Ltd., London, UK), following the manufacturer's protocol. PCR amplification was conducted using the Platinum™ PCR SuperMix High Fidelity (<https://www.thermofisher.com/order/catalog/product/12532016>, accessed on 6 April 2023, Thermo Fisher Scientific, Waltham, MA, USA) and purification with ExoSap (<https://www.thermofisher.com/order/catalog/product/78205.10.ML>, accessed on 6 April 2023, Thermo Fisher Scientific, Waltham, MA, USA). The PCR reaction amplified four gene loci using the nuclear ribosomal internal transcribed spacer (ITS) and the large subunit ribosomal rRNA (LSU) as potential markers for species identification. The following primers were used: ITS1 (sequence: 5'- TCCGATGGTGAAC-CTGCGG-3'), ITS2 (sequence: 5'-ATGCGATACTTGGTGTGAAT-3'), and LSU D1 and D2 primers specific for the internal transcribed spacer region of the ribosomal DNA [35]. The ITS amplicon was amplified through an initial denaturation step at 5 min at 95 °C, 35 cycles of 30 s at 94 °C, 30 s at 55 °C, 40 s at 55 °C, and a final extension stage of 5 min at 72 °C. PCR products were verified using 1% agarose gel electrophoresis, comparing them with 2 Kb DNA markers. BMR Genomics (Padova, Italy) conducted genomic DNA extraction, PCR, and Sanger sequencing.

### 4.4. Sequencing and Phylogenetic Analysis

Sequencing was conducted using the BrilliantDye™ Terminator (v1.1) Cycle Sequencing Kit (1000 rxn) (<https://www.nimagen.com/shop/products/brd1-1000/brilliantdye-terminator-v11-cycle-sequencing-kit-1000-rxn>, accessed on 6 April 2023, NimaGen B.V., Nijmegen, The Netherlands) on a 3730xl DNA Analyzer (Thermo Fisher Scientific, Waltham, MA, USA). Results were submitted to rDNA homology sequence alignment using BLAST (<https://blast.ncbi.nlm.nih.gov/Blast.cgi>, accessed on 6 April 2023) and compared with

the NCBI GenBank (GenBank; <http://www.ncbi.nlm.nih.gov/BLAST/incidence.html>, accessed on 6 April 2023) to verify the genetic similarity of the samples.

The phylogenetic tree was created using the NCBI-BLAST tool with the method “neighbor joining”, with a maximum seq difference of 0.5.

## 5. Conclusions

Awareness of favorable environmental conditions and knowledge of the life cycles of the host species can help to prevent *Sclerotinia sclerotiorum* infections. Familiarity with environmental conditions, disease dispersion methods, and continued scouting and sampling techniques should be included in an integrated management program in order to prevent *S. sclerotiorum* fungus incidence. Further investigations are needed for a better understanding of the germination and dormancy requirements of sclerotia caused by *S. sclerotiorum*. The inclusion of morphological, molecular, and climate data is crucial to the improvement of disease forecasting models.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/plants12203516/s1>, File S1: Sanger sequences alignment with the NCBI database.

**Author Contributions:** Conceptualization, A.-M.F. and E.U.; methodology, A.-M.F. and I.G.; software, A.-M.G.; validation, A.-M.F., F.-D.L., E.U. and I.G.; formal analysis, A.-M.F. and A.-M.G.; investigation, A.-M.F.; resources, E.U.; data curation, F.-D.L.; writing—original draft preparation, A.-M.G.; writing—review and editing, A.-M.F., E.U. and I.G.; visualization, A.-M.F., E.U. and I.G.; supervision, A.-M.F., E.U. and I.G.; project administration, A.-M.F. and E.U.; funding acquisition, A.-M.F. All authors have read and agreed to the published version of the manuscript.

**Funding:** This research received no external funding.

**Data Availability Statement:** Data can be found in Supplementary Materials.

**Acknowledgments:** The work of Iulian GABUR was supported by the project “PROINVENT”, Contract no. 62487/03.06.2022—POCU/993/6/13 Code 153299, financed by The Human Capital Operational Programme 2014–2020 (POCU), Romania.

**Conflicts of Interest:** The authors declare no conflict of interest.

## References

- Purdy, L.H. *Sclerotinia sclerotiorum*: History, Diseases and Symptomatology, Host Range, Geographic Distribution, and Impact. *Phytopathology* **1979**, *69*, 875. [[CrossRef](#)]
- Hossain, M.M.; Sultana, F.; Li, W.; Tran, L.P.; Mostofa, M.G. *Sclerotinia sclerotiorum* (Lib.) de Bary: Insights into the Pathogenomic Features of a Global Pathogen. *Cells* **2023**, *12*, 1063. [[CrossRef](#)]
- Jahan, R.; Siddique, S.S.; Jannat, R.; Hossain, M.M. Cosmos white rot: First characterization, physiology, host range, disease resistance, and chemical control. *J. Basic Microbiol.* **2022**, *62*, 911–929. [[CrossRef](#)] [[PubMed](#)]
- Islam, M.R.; Akanda, A.M.; Hossain, M.M.; Hossain, M.M. First characterization of a newly emerging phytopathogen, *Sclerotinia sclerotiorum* causing white mold in pea. *J. Basic Microbiol.* **2021**, *61*, 923–939. [[CrossRef](#)] [[PubMed](#)]
- Islam, M.R.; Prova, A.; Akanda, A.M.; Hossain, M.M. First report of white mould caused by *Sclerotinia sclerotiorum* on pea in Bangladesh. *J. Plant. Pathol.* **2020**, *102*, 941. [[CrossRef](#)]
- Tian, B.; Xie, J.; Fu, Y.; Cheng, J.; Li, B.O.; Chen, T.; Zhao, Y.; Gao, Z.; Yang, P.; Barbetti, M.J.; et al. A cosmopolitan fungal pathogen of dicots adopts an endophytic lifestyle on cereal crops and protects them from major fungal diseases. *ISME J.* **2020**, *14*, 3120–3135. [[CrossRef](#)]
- Cohen, S.D. Estimating the Climate Niche of *Sclerotinia sclerotiorum* Using Maximum Entropy Modeling. *J. Fungi* **2023**, *9*, 892. [[CrossRef](#)]
- Borah, T.R.; Dutta, S.; Barman, A.R.; Helim, R.; Sen, K. Variability and host range of *Sclerotinia sclerotiorum* in Eastern and North Eastern India. *J. Plant Pathol.* **2021**, *103*, 809–822. [[CrossRef](#)]
- Sesan, T.E.; Crisan, A. Putregaiul alb al plantelor de cultura-*Sclerotinia sclerotiorum*-Prevenire si combatere, Edit. *Ceres Bucur.* **1998**, 15–27.
- Bontea, V. *Parasitic and Saprophytic Fungi of Romania*; Academia Republicii Socialiste România: Bucuresti, Romania, 1985.
- Boland, G.J.; Hall, R. Index of plant hosts of *Sclerotinia sclerotiorum*. *Can. J. Plant Pathol.* **1994**, *16*, 93–108. [[CrossRef](#)]
- Farr, D.F.; Rossman, A.Y. *Fungal Databases*; U.S. National Fungus Collections; ARS, USDA: Washington, DC, USA, 2017. Available online: <https://nt.arsgrin.gov/fungaldatabases/> (accessed on 15 May 2023).



13. O'Sullivan, C.A.; Belt, K.; Thatcher, L.F. Tackling Control of a Cosmopolitan Phytopathogen: Sclerotinia. *Front. Plant Sci.* **2021**, *12*, 707509. [CrossRef] [PubMed]
14. Ekins, M.G.; Hayden, H.L.; Aitken, E.A.B.; Goulter, K.C. Population structure of *Sclerotinia sclerotiorum* on sunflower in Australia. *Australas. Plant Pathol.* **2011**, *40*, 99–108. [CrossRef]
15. Kim, H.J.; Ono, E.; Morimoto, K.; Yamagaki, T.; Okazawa, A.; Kobayashi, A.; Satake, H. Metabolic engineering of lignan biosynthesis in *Forsythia* cell culture. *Plant Cell Physiol.* **2009**, *50*, 2200–2209. [CrossRef] [PubMed]
16. Daughtrey, M.L.; Wick, R.L.; Peterson, J.L. *Compendium of Flowering Potted Plant Diseases*; APS Press: Saint Paul, MN, USA, 1995.
17. Grabowski, M.A. Exploring the host range of *Sclerotinia sclerotiorum* in herbaceous ornamental plants. Ph.D. Thesis, University of Minnesota, Minneapolis, MN, USA, 2017. Available online: <https://conservancy.umn.edu/handle/11299/201043> (accessed on 10 May 2023).
18. Jones, R.K.; Benson, D.M. *Diseases of Woody Ornamentals and Trees in Nurseries*; APS Press: Saint Paul, MN, USA, 2001; pp. 100–125.
19. Muñoz, M.; Behnke, L.E.; Faust, J.E.; Schnabel, G. First Report of Rose BentNeck Caused by *Sclerotinia sclerotiorum* on Commercial Cut Roses (*Rosa hybrida* L.). *Horticulturae* **2023**, *9*, 646. [CrossRef]
20. Rahman, M.M.E.; Suzuki, K.; Islam, M.M.; Dey, T.K.; Harada, N.; Hossain, D.M. Molecular characterization, mycelial compatibility grouping, and aggressiveness of a newly emerging phytopathogen, *Sclerotinia sclerotiorum*, causing white mold disease in new host crops in Bangladesh. *J. Plant Pathol.* **2020**, *102*, 775–785. [CrossRef]
21. Florea, A.M.; Gafencu, A.M.; Lipşa, F.D.; Ulea, E. New host for *Sclerotinia sclerotiorum* in the NE region of Romania, Scientific Papers. Series, A. *Agronomy* **2022**, *65*, 69–72.
22. Zanoschi, V.; Sârbu, I.; Toniuc, A. *Flora Lemnoasă Spontană și Cultivată din România*; Universitatea Alexandru Ioan Cuza: Iași, Romania, 2004; Volume 4, p. 290.
23. Heffer Link, V.; Johnson, K.B. White Mold. *Plant Health Instr.* **2007**. Available online: <https://www.apsnet.org/edcenter/disandpath/fungalasco/pdlessons/Pages/WhiteMoldPort.aspx> (accessed on 10 January 2023). [CrossRef]
24. Abawi, G.S.; Grogan, R.G. Epidemiology of diseases caused by *Sclerotinia* Species. *Am. Phytopathol. Soc.* **1979**, *69*, 899–904. [CrossRef]
25. Altschul, S.F.; Gish, W.; Miller, W.; Myers, E.W.; Lipman, D.J. Basic local alignment search tool. *J. Mol. Biol.* **1990**, *215*, 403–410. [CrossRef]
26. National Center for Biotechnology Information (NCBI). Bethesda (MD): National Library of Medicine (US), National Center for Biotechnology Information. 1988. Available online: <https://www.ncbi.nlm.nih.gov/> (accessed on 6 June 2023).
27. Dutta, S.; Borah, T.; Roy Barman, A.; Hansda, S.; Ghosh, P. Overview of *Sclerotinia sclerotiorum* in India with special reference to its emerging severity in Eastern Region. *Indian Phytopathol.* **2016**, *69*, 236–239.
28. Firoz, M.J.; Xiao, X.; Zhu, F.-X.; Fu, Y.-P.; Jiang, D.-H.; Schnabel, G.; Luo, C.-X. Exploring mechanisms of resistance to dimethachlone in *Sclerotinia sclerotiorum*. *Pest Manag. Sci.* **2016**, *72*, 770–779. [CrossRef] [PubMed]
29. Ding, Y.; Mei, J.; Chai, Y.; Yang, W.; Mao, Y.; Yan, B.; Yu, Y.; Disi, J.O.; Rana, K.; Li, J.; et al. *Sclerotinia sclerotiorum* utilizes host-derived copper for ROS detoxification and infection. *PLoS Pathog.* **2020**, *16*, e1008919. [CrossRef] [PubMed]
30. Ding, Y.; Mei, J.; Chai, Y.; Yu, Y.; Shao, C.; Wu, Q.; Disi, J.O.; Li, Y.; Wan, H.; Qiam, W. Simultaneous transcriptome analysis of host and pathogen highlights the interaction between Brassica oleracea and *Sclerotinia sclerotiorum*. *Phytopathology* **2019**, *109*, 542–550. [CrossRef]
31. Robinson, J.R.; Isikhuemhen, O.S.; Anike, F.N. Fungal–Metal Interactions: A Review of Toxicity and Homeostasis. *J. Fungi* **2021**, *7*, 225. [CrossRef] [PubMed]
32. Gajewska, J.; Floryszak-Wieczorek, J.; Sobieszczuk-Nowicka, E.; Mattoo, A.; Arasimowicz-Jelonek, M. Fungal and oomycete pathogens and heavy metals: An inglorious couple in the environment. *IMA Fungus* **2022**, *13*, 6. [CrossRef] [PubMed]
33. Saharan, G.S.; Naresh, M. Chapter: Infection and Pathogenesis. In *Sclerotinia Diseases of Crop Plants: Biology, Ecology and Disease Management*; Springer: Berlin/Heidelberg, Germany, 2008; pp. 215–217. [CrossRef]
34. Wang, A.R.; Lin, W.W.; Chen, X.T.; Lu, G.D.; Zhou, J.; Wang, Z.H. Isolation and identification of *Sclerotinia* stem rot causal pathogen in *Arabidopsis thaliana*. *J. Zhejiang Univ. Sci. B* **2008**, *9*, 818–822. [CrossRef]
35. White, T.J.; Bruns, T.D.; Lee, S.B.; Taylor, J.W.; Innis, M.A.; Gelfand, D.H.; Sninsky, J. Amplification and Direct Sequencing of Fungal Ribosomal RNA Genes for Phylogenetics. In *PCR Protocols: A Guide to Methods and Applications*; Innis, M.A., Gelfand, D.H., Sninsky, J.J., White, T.J., Eds.; Academic Press, Inc.: New York, NY, USA, 1990; pp. 315–322.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of MDPI and/or the editor(s). MDPI and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.