

Aljabri M (2022) Notulae Botanicae Horti Agrobotanici Cluj-Napoca Volume 50, Issue 3, Article number 12855 DOI:10.15835/nbha50312855 Review Article



Abiotic stress enhancement tools for improving crop tolerance

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Abstract

Abiotic stresses create an unfavourable environment for plant growth, increasing the possibilities of low yield and associated economic loss. Several steps have been taken to address this problem. During the last twenty years, techniques of genetic engineering/transgenic breeding have made significant advances in gene manipulation for inciting desirable traits in transgenic plants. Transgenic techniques allow us to identify potential genes, transcription factors (TFs), and miRNAs, engaged in certain processes in plants, allowing us to gain a comprehensive understanding of the processes at molecular and physiological levels which determine plant resilience and production. The reliability and specificity of this approach ensure that future plant enhancements will be a huge success. As a result, transgenic breeding has been determined to be a viable strategy for improving crop abiotic stress tolerance. The approach of CRISPR/Cas gene-editing technique to create stress-tolerant plant variants is gaining popularity right now. The researchers like this user-friendly technology because of its versatility. In the gene-editing process, the DNA sequence "CRISPR" and the endonuclease "Cas" collaborate under the supervision of specific guide RNA. In a variety of plant species, the CRISPR/Cas system is being utilized. In the majority of situations, Cas9 is employed. Various reports have surfaced which demonstrate the utilization of CRISPR/Cas9 technology to improve the abiotic stress tolerance of plants. Therefore, this review aimed to review the promising and effective applications of transgenic plant breeding for enhancing environmental stress tolerance and crop productivity, as well as its recent developments.

Keywords: abiotic stress; CRISPR; miRNAs; transcription factors; transgenic plants

Introduction

Extensive increases in several environmental stresses, exhibit a remarkable effect on crop productivity (Seleiman and Kheir, 2018; Seleiman *et al.*, 2020; 2021a; 2021b). Because of the current loss in the agricultural area, water scarcity, global warming, and climate change, it is approximated that crop productivity will decrease further in the future (Arora, 2019; Usman *et al.*, 2020). Abiotic stresses have a negative impact on crop development and yield (Seleiman, 2019). There are various kinds of abiotic stresses like drought, salinity, temperature stress, excessive water, heavy metal stress, and UV stress. Reduction in agricultural production has been noticed in the past few decades because of the decreased agricultural land area due to increased population and environmental stresses (Arora, 2019; Usman *et al.*, 2020). Thus, novel approaches in the agricultural sector like the application of new techniques and continuous agricultural innovation are critical.

Received: 16 Aug 2022. Received in revised form: 01 Sep 2022. Accepted: 09 Sep 2022. Published online: 12 Sep 2022. From Volume 49, Issue 1, 2021, Notulae Botanicae Horti Agrobotanici Cluj-Napoca journal uses article numbers in place of the traditional method of continuous pagination through the volume. The journal will continue to appear quarterly, as before, with four annual numbers.

To satisfy the growing need for food and counteract the negative impacts of abiotic stress on crop yield, it is essential to generate transgenic lines with increased tolerance to a broad range of abiotic stresses (Pandey et al., 2017; Noman et al., 2017). Conventional breeding approaches were utilized for genetic variability developing from intergeneric or interspecific hybridizations and diverse germplasms to stimulate cellular and tissue culture level mutations to make the plant more resilient to abiotic stresses. But there are various restrictions with these approaches (Dita et al., 2006). To introduce new plants much time is needed, and various undesired genes can be moved alongside desired genes, due to the intricate stress reactions and their performance, there is a modest yield improvement under stress conditions, and there is no assurance of getting a specific gene association among millions of crosses (Muehlbauer et al., 2006). The ability of gene identification and functional genomics in plants to govern a specific feature has altered dramatically as a result of recent biotechnology breakthroughs (Choudhary et al., 2012). Plant engineering and genetic manipulation strategies for abiotic stress resistance (Mushtaq et al., 2019) rely on gene expression, which is primarily concerned with regulatory and signalling networks that control genes. encoding stress-resistant proteins and enzymes for functional metabolite synthesis (Caño-Delgado et al., 2004; Noman et al., 2017). Therefore, the focus of this review is on the promising and effective applications of transgenic plant breeding for enhancing environmental stress tolerance and crop productivity, as well as its recent developments.

Abiotic stress control via transgenic approach

Abiotic stresses like salinity (Al-Ashkar et al., 2019; Al-Ashkar et al., 2020; Seleiman et al., 2020; Taha et al., 2020;), heat, and drought (Batool et al., 2020; Roy et al., 2021) are leading environmental cues which affect crop productivity of crops (Mukhtar et al., 2020; Elshayb et al., 2022). The use of traditional plant breeding methods is not much established in augmenting environmental stress so far. This could be because of the traits being regulated by several genes located at a quantitative trait locus (QTL). For impeding harmful impacts of numerous environmental conditions, it is required to verify the possible genes or QTLs (gene networks) linked to extensive tolerance to different abiotic stresses. These abiotic stresses negatively impact plant growth and yield by modifying vegetative and reproductive growth and development phases. plants undergo a range of physiological, biochemical, and molecular changes which are prompted by the abiotic stresses which impair the cellular machinery (Rai et al., 2011). These modifications comprise disorganization of cellular osmotic equilibrium which leads to deteriorated homeostasis, disposition of ions, and oxidative damage denaturing the intrinsic proteins of plants. The response of plants to various abiotic stresses occurs through various processes that stimulate the cell signalling, transcriptional controls, and generation of stresstolerant proteins, antioxidants, and osmolytes which sustain homeostasis and restore damaged integral proteins. Usually, stress-sensitive plants which are not able to produce such compounds under stress conditions become subjected to several stresses that obstruct their growth. Several genes have been recognized in several plants/organisms, which encode stress defending compounds and can be directed for genetic transformation into susceptible genotypes. These genes have been grouped into three classes as (a) osmolytes such as mannitol, glycine betaine, proline, and heat shock protein-coding genes, (b) genes accountable uptake of ion and water and transport like aquaporins and ion transporter, and (c) genes modulating transcriptional controls and signal transduction processes, like MAPK and DREBI.

Nanotechnology for abiotic stress tolerance

Abiotic stresses have an adverse impact on plant growth and productivity. Because of the increasing negative impacts of abiotic stresses, researchers have great interest to rationalize the impact of abiotic stresses and enhancing crop yield. According to FAO reports, by 2050 there will be an addition of 2.3 billion people

worldwide and to feed such an enormous population, there is a considerable challenge for plant researchers to enhance crop production to 70% of towards world agriculture. Hence there is an urgent need to recognize novel research areas to curb technological obstacles in tackling yield blockade, resource use efficiency, and development of environmentally accepted technology. Nanobiotechnology is becoming a promising area in mitigating the impacts linked with abiotic and biotic stress to acquire a sustainable future for agriculture globally. Nanoparticles are synthesized from metal or metal oxide by different modes i.e, physical, chemical, and biological. Dur to their smaller size, nanoparticles have gained some special properties which have opened new approaches in the agriculture sector. These particles are assessed for their possible function in plant development and protection from various stresses. Still, there is a long way to develop technology to attain sustainable agriculture (Saxena *et al.*, 2016) (Figure 1).



Figure 1. depicts the physiological, biochemical, and molecular basis of abiotic stress tolerance in plants under the treatment of different nanoparticles

The agricultural application of nanoparticles can increase production sustainably and productively there by adding to the agriculture reliability (Fincheira *et al.*, 2020). In agricultural fields Nanotechnology has been utilized to screen the quantity of water, nutrient availability, and plant stress caused by environmental and biotic factors (Singh and Singh, 2019). In *Cucumis sativus* L., increased growth and productivity were observed due to improved nutrient uptake by the supplementation of silicon dioxide (SiO₂) nanoparticles in presence of drought and salinity stress (Alsaeedi *et al.*, 2019). In presence of salt stress, the application of Ag nanoparticles amended the germination rate, shoot and root length, and fresh and dry mass of seedlings which led to alleviated development of *Lathyrus sativus* L. (grass pea) (Hojjat, 2019). In soybean (*Glycine max* L.) plants application of SiO₂ nanoparticles immobilized and inactivated the mercury thereby reducing the adverse impact of mercury (Li *et al.*, 2020). Supplementation of various levels of silica nanoparticles enhanced plant resilience to water deficit conditions in hawthorns (*Crataegus* sp.), hawthorn seedlings physiological and biochemical reactions vary in response to contrasting levels of silica nanoparticles during various stages of drought (Ashkavand *et al.*, 2015). It was demonstrated that silica nanoparticles had a positive impact on photosynthesis attributes. It was revealed that silica nanoparticles and fertilizer demonstrated a pronounced impact on the physiological and morphological characteristics of basil under salinity stress (Kalteh *et al.*, 2018).

Apart from the above effects, nanoparticles, due to their unusual attributes, have the potential to promptly transfer DNA particles inside cells of plants to modify their metabolic functioning, being able to

easily penetrate in plant cells (Giraldo *et al.*, 2014). The genotoxic impacts brought about by salinity in *Solanum lycopersicum* L. were overcome by the supplementation of zinc oxide (ZnO) by alteration of cytosine methylation (Haliloglu *et al.*, 2020). Currently, genome editing approaches are currently widely adopted perspectives for the crop enhancement program (Tiwari and Lata, 2019; Bhat *et al.*, 2020; Ansari *et al.*, 2020), as a result, a group of scientists has merged this method with nanotechnology and created a method for characterization of plant genome of plant species by employing carbon nanotubes and CRISPR-cas9 geneediting methods which leads to enhanced protein expression level (Demirer *et al.*, 2019), indicating that in future genome editing along with the use of nanoparticles may induce enhancement of several plant attributes including yield, quality, and stress resistance.

Application of Biotechnology in Abiotic Stress Tolerance

Various biotechnological techniques like molecular breeding and genetic engineering in plants propose certain alternatives for procuring enhanced and genome-edited plants in a brief period (Mishra *et al.*, 2017). Genetic engineering might repress reproductive hurdle in several plant species (Noman *et al.*, 2019). Transgenic breeding effectively enhanced yield via genetic alterations and improvements, with a small breeding phase (Gantait and Mondal, 2018). As advancement in the physiology of plants increased, understanding of complications in stress-resilient procedures and their association with various attributes, selection regulation employing genomic techniques will lead to recognition of quantitative trait loci (QTL) and genes associated with attributes (Young, 1996). Recognition of potential genes accountable for stress resilience in plants is necessary for creating transgenic plants with increased stress resistance (Muehlbauer *et al.*, 2006).

Over the last two decades, plant engineering approaches for environmental stress resistance were extensively investigated. Scientists examined gene expression which was possibly associated with plant defence signalling pathways that coded proteins involved in abiotic stress tolerance. Currently, several achievements have been attained to enhance plant stress tolerance by genetic transformations (Noman *et al.*, 2019); moreover, the complicated genetic procedures for abiotic stress tolerance make the work complex (Mishra *et al.*, 2017). Thus, there should be a combination of plant biotechnology and conventional breeding and physiology.

Quantitative Trait Loci (QTL)

In abiotic stress resilience, Quantitative trait loci (QTL) mapping turned out as remarkable approach in plant breeding and is an excellent procedure for exploring genetically complex systems (Shen *et al.*, 2018). Under various abiotic stress conditions, improved crop yield was attained via the use of QTL, which regulated explicit agronomic attributes and physiological mechanisms for increased crop yield. Corresponding to their constancy and reaction to several environmental stresses QTL can be classified as "adaptive and constitutive" (Collins *et al.*, 2008). The later ones are common in environments, while adaptive QTL occurs environmental conditions, like QTL that are expressed in presence of high or low temperature, specifying the role of QTL in regulating temperature stresses (Shen *et al.*, 2018).

In rice (Oryza sativa L.) plants growing under salinity stress, several QTL were distinguished to regulate fundamental attributes in plants. In these plants meta-analysis of QTLs which regulate early-stage salt resilience was carried out by employing QTL data from numerous studies. 11 meta-QTLs for three attributes of little intervals were located on chromosomes 1 and 2 with the use of a consensus map. Upon analyzing 56 various genotypes and breeding lines, six salt-resistant genotypes (Bharathy, I Kung Ban 4-2 Mutant, Langmanbi, Fatehpur 3, CT-329, and IARI 5823) were described. A study of meta-QTL regions unveiled various prospect genes consociated with salinity resilient traits (Islam *et al.*, 2019). Genetic studies and QTL mapping in cucumber were conducted for thermo-tolerance utilizing recombinant inbred line (RILs; HR) population and a doubled haploid (DH; HP) population which were deduced from heat-sensitive (65G) and

tolerant (02245) parents. Upon inheritance investigation, it was proposed that multiple genes in cucumber (Cucumis sativus L.) seedlings influence both short-term extreme and long-term mild heat tolerance. Six QTLs for heat tolerance were observed which include qHT3.1, qHT3.2, qHT3.3, qHT4.1, qHT4.2, and qHT6.1. Amongst these, qHT3.2 was frequently observed thrice in HR and HP at various conditions, which interpreted phenotypic variations. The 481.2 kb region harboured 79 genes, nine of which might be taking part in heat stress reactions (Dong et al., 2020). In triticale, freezing resilience is a major attribute that leads to its winter hardiness. QTL and molecular markers related to cold resistance in winter triticale were identified by Wasek et al. (2021). They recognized three coherent QTL for various fluorescence parameters with complex interval mapping (CIM) and single-marker analysis (SMA). The first locus Qfr.hm-7A.1 justified the observed 9% disparity of both leakages of ions and plants recovery after freezing. The chief locus Qchl.hm-5A.1 was noticed for chlorophyll fluorescence traits that accounted for 19.6% of phenotypic dissimilarity. Co-location of QTL on chromosomes 7A.1, 4R, and 5R, clearly defined physiological and genetic link of the plant survival after freezing with the capability of exerting favourable photochemical activity of the photosystem II and fixing the integrity of cell membranes. Genes present in observed QTL were those which encode BTR1-like protein, transmembrane helix proteins like potassium channel, and phosphoric ester hydrolase concerned with osmotic stress response and proteins which regulate expression of the gene, chloroplast RNA processing, and pyrimidine salvage pathway (Wasek et al., 2021). QTL linked with attributes like seed germination and early plant growth were studied in wheat (Triticum aestivum L.) cultivars, 'Pasban 90' (salt-tolerant), and 'Frontana' (salt susceptible) using recombinant inbred lines. Composite interval mapping (CIM) was employed for analyzing QTL for various traits and in this analysis total 44 QTLs were distinguished. 26 Eleven main QTLs were present in 1B, 2B, 3B, 5B, 6B, and 7B chromosomes of the mapping population under salinity stress conditions. The results divulged that QTLs play an important part in salinity resistance at the early seedling growth stage (Batool et al., 2018). It has been deduced that conventional QTL mapping, QTL-sequencing, can quickly distinguish candidate genes, thereby altering the breeding methods and enhancing the breeding efficacy (Sharma et al., 2017). Through functional genomic strategies, QTL can be an excellent tool for translational genomics and the generation of quantitative characteristics in some plants (Adhikari et al., 2019; Wen et al., 2019). These studies established the essentiality of QTL identification and therefore proposed a novel and unique marker with huge potential to ameliorate abiotic stress resistance and crop production.

miRNAs in abiotic stress resistance

MicroRNAs are small about 22-24 nucleotide, non-coding regulatory RNAs which are required in sequence-specific composite regulatory mechanisms of gene silencing in plants. miRNAs are also involved in modulating abiotic stress resistance in plants. Defence responses in plants during biotic or abiotic stresses are arbitrated by alterations in both genes and miRNAs expression levels which are then implicated in regulating stress response and tolerance in plants. Several pieces of evidence have proposed the remarkable contribution of high-throughput sequencing technologies towards the recognition and functional description of various miRNAs in plants. Recently it has been revealed that primary miRNA transcripts (pri-miRNAs) code for regulatory peptides, which play a role in amending numerous agronomic attributes (Jatan and Lata, 2019).

With the utilization of a recent machine learning pattern, it was depicted that miRNA169, miRNA159, miRNA396, and miRNA393 have the most important function in plant reaction to water deficit conditions, salt stress, and temperature stress respectively (Vakilian, 2020). In maize, impact of miRNAs on its biochemical pathways was ascertained and several new miRNAs were discovered which increased the amelioration of drought stress. microRNAs modulate their target gene expression by their up-regulation or down-regulation which affects metabolic reactions of maize to drought conditions (Upadhyay *et al.*, 2019). It was found that miRNA393 was vigorously up-regulated by dehydration, freezing, high salt, and ABA administration in several plants (Zhang, 2015). In drought-stressed *Arabidopsis*, miR169 was down-regulated, and its target nuclear factor YA5 (NF-YA5) was remarkably stimulated when subjected to drought stress (Nadarajah and Kumar,

2019). Plants overexpressing miRNA169a- were susceptible to drought, in comparison to wild plants, whilst the miRNA169a-targeted NF-YA5-overexpression enhanced drought stress resistance in plants (Ni et al., 2013). Similarly, in Arabidopsis thaliana, GmNF-YA3 gene, a target of miR169, remarkably increased drought stress resilience in these plants (Zhang et al., 2011). In guava, 40 potential microRNAs which belonged to 19 families were differentiated. These differentiated miRNA precursors constituted stable stem-loop structures and demonstrated high sequence conservation among various plant species. Under salinity stress distinctive expression pattern of seven selected guava miRNAs (pgu-miR156f-5p, pgu-miR160c-5p, pgu-miR162-3p, pgumiR164b-5p, pgu-miR166t, pgu-miR167a-5p, and pgu-miR390b-5p) were recorded and among these pgumiR162-3p, pgu-miR164b-5p and pgu-miR166t were most affected ones. A total of 49 putative target transcripts of the differentiated guava miRNAs in this work were discovered to be involved in metabolic pathways, cellular development, and stress response signalling using the psRNA Target tool (Sharma et al., 2020). In cotton extreme sequencing was done to unravel significant roles of miRNAs in reaction to salinity and drought (Xie et al., 2015). Wang et al. (2016) classified the range of temperature stress-responsive miRNAs and their targets in cotton by using deep sequencing. In Cd stressed Brassica napus 44 known miRNAs (belonging to 27 families) and 103 new miRNAs were recognized. An intense investigation of miRNA expression description revealed 39 distinctively expressed miRNAs between control and Cd-stressed plants. The functions of matching target genes were found in transcription factor modulation, biotic stress reaction, ion homeostasis, and secondary metabolism, among other processes. These results indicated the role of miRNAs in the modulation of TFs and said processes in response to Cd stress in Brassica napus (Jian et al., 2018) (Figure 2).



Figure 2. Interaction between different miRNAs and TFs and their regulatory pathways in response to different stresses

CRISPR (Clustered Regularly Interspaced Short Palindromic Repeats)/Cas9 and Abiotic stress tolerance

Various environmental stresses restrain crop yield around the globe (Pandey *et al.*, 2017). Several traditional methods have been used to enhance crop production, but those methods are now becoming restrained because of the decrease in genetic resources of plants and those methods are very time consuming (Gantait and Mondal, 2018). There is an urgent demand for effective crop enhancement programmes with new genome editing approaches, like CRISPR-Cas9 (Tang *et al.*, 2017), that is quick, swift, and precise in getting genome-edited plants.

CRISPR-Cas9 system is a structured genome engineering tool, that is effectively implemented in several organisms like bacteria, animals, and plants (Sander *et al.*, 2014). Because of its effectiveness in specific gene editing via targeted traits, CRISPR-Cas9 system can be employed in crop increment, and this will open numerous approaches concerning this. CRISPR/Cas9-mediated genome editing for the generation of diverse abiotic stress resilient crops is depicted in Figure 3 as a step-by-step process.



Figure 3. Step-by-step presentation of CRISPR/Cas9 mediated genome editing for the development of different abiotic stress resistance varieties of crops

CRISPR/Cas9 has also been utilized in plants like wheat, maize (*Zea mays* L.), tomato (*Solanum lycopersicum* L.), soybean (*Glycine max* L.), and sorghum (*Sorghum bicolor* L.) besides its utilization in model plants (Osakabe and Osakabe, 2017). CRISPR/Cas9-interceded editing technique in tomato to develop its long shelf life was employed through targeted mutagenesis and gene replacement (Yu *et al.*, 2017), through complex genome-editing to increase lycopene content (Li *et al.*, 2018) and through concomitant editing of miR482b and miR482c for increased tolerance to *phytophthora infestans* (Hong *et al.*, 2020). Likewise, in soybean, plant height and internode length were modulated with the use of CRISPR/Cas9 editing approach and it was also utilized to regulate the flowering time of the plants (Cheng *et al.*, 2019; Li *et al.*, 2020). Apart from this, the utilization of this technique to enhance abiotic stress-resilience in plants has evoked the curiosity of several researchers. The multiplex of genes is engaged in environmental stress reactions modulating metabolism, signalling, and regulatory pathways of plants. CRISPR/Cas9 system can target single or multiple genes to enhance plant potentiality to stress resistance. Point mutations, insertions or deletions or

transcriptional modulations might be produced for the target-based gene-editing via CRISPR/Cas9 system (Jain, 2015). Important things which need to be taken into consideration in this system for enhancing environmental stress tolerance in plants are the designation of the desired promoter for cas9 expression, guide RNA outline, formation of novel alleles for abiotic stress-reactive genes, and the development appropriate gene delivery system (Osakabe and Osakabe, 2017). In Arabidopsis a genetic character, ABRE1 was utilized for enhancing water deficit tolerance since abscisic acid-responsive element-binding protein 1/ABRE binding factor 2(ABRE1/ABF2) are involved in modulating the drought stress reactions (Roca Paixão et al., 2019). In this study, CRISPR/Cas9 genome-editing was employed for stimulation of endogenous ABRE1 promoter and in this study inactive Cas9 (dCas9) was combined with HAT or histone acetyltransferase. Utilization of CRISPR/dCas9^{HAT} in *Arabidopsis thaliana* exhibited an ameliorated tolerance against drought stress by positively regulating ABRE1 (Roca Paixão et al., 2019). With CRISPR/Cas9 technology, Alfatih et al. (2020) developed rice PARAQUAT TOLERANCE 3 knockout mutants (OsPQT3). These mutants exhibited higher resilience to salt and oxidative stresses (Alfatih et al., 2020). To enhance drought stress resistance in Arabidopsis CRISPR/Cas9 was employed to establish novel alleles of OST2 (playing an effective role in stomatal movement), by the co-expression of Cas9 and GFP (green fluorescent protein) for enhancing tolerance against drought (Osakabe et al., 2016). These authors employed truncated guide RNA (tru-guide RNA) for sitespecific alterations not causing any off-target impacts. The modified stress responses in Arabidopsis through a high expression of OST2 in germline cells propose an increased inheritable stress-resilience in the plant (Osakabe et al., 2016). CRISPR/cas9 stimulated ARGOS8 variations in maize resulting in improved grain yield of the plants under drought stress (Shi et al., 2016). All these studies showed the capability of CRISPR/cas9 system to produce crop plants that are resistant to drought stress by developing new allelic variations. CRISPR/cas9 approach has also been employed to decrease the metal toxicity of plants. With CRISPR/Cas9 system, a group of researchers successfully developed rice plants with less cesium by disabling K+ transporter OsHAK1 (Nieves-Cordones et al., 2017). Similarly, a rice plant with less cadmium was developed by knocking out the metal transporter gene OsNramp5 (Tang et al., 2017). Additionally, rice plants resistant to herbicide were developed through CRISPR/Cas9 interceded homologous recombination of acetolactate synthase (ALS). Research is advancing for utilizing CRISPR/Cas9 genome-editing technique to surpass problems occurring via various abiotic stress conditions. In Table 1, some selected studies for a better understanding of the role of biotechnology in abiotic stress tolerance in plants are presented. While the elevation of different abiotic stresses with different techniques on plant is shown in Table 2.

Transcription Factors linked with abiotic stress

Crop plants are negatively impacted by a diverse environmental stress that have a pronounced effect on crop production. Plants have developed advanced approaches, and genes encoding transcription factors (TFs), which are prime modulators of stress-receptive genes which are eminent prospects for crop enhancement. Examples associated with recent works comprise TF gene regulation and overexpression perspectives in plants to increase stress resilience. But there is still a lot to learn regarding various plant TFs. Of more than 80 TF families, only a few have been extensively studied for their role in abiotic as well as biotic stress reactions and among these, some of the important TFs are NAC, MYB, WRKY, bZIP, and ERF/DREB (Baillo *et al.*, 2019).

When plant cells perceive abiotic stress via receptors or sensors, ROS levels increase, resulting in cell oxidative damage and, finally, cell death. ROSs have recently been recognized as important second messengers in complicated signalling networks of abiotic stress reactions in plants. As a result, controlling ROS signalling and homeostasis is an essential technique for improving stress resistance in plants under adverse environmental conditions (You and Chan, 2015). TFs play a key function downstream of ROS signalling pathways as the main modulatory proteins taking part in abiotic stress responses. During abiotic stress reactions in plants,

members of the MYB, bHLH, WRKY, bZIP, and NAC families have been shown to play essential roles in modulating ROS signal transduction.

| Biotechnological approaches | Туре | Species | Function | References |
|--------------------------------|---------------|----------------------------------|--|-----------------------------------|
| •• | | Oryza sativa L. | Drought stress tolerance | Shim <i>et al.</i> , 2018 |
| | | Triticumaestivum L. | Dehydration tolerance | Chen <i>et al.</i> , 2018 |
| | NAC | Pyrusbetulifolia L. | Chilling and drought stress resistance | Jin <i>et al.</i> , 2017 |
| | | Tamarix hispida L. | Tolerance to salinity and osmotic stress | He <i>et al.</i> , 2019 |
| | | Solanum lycopersicum L. | Heat stress tolerance | Meng et al., 2015 |
| | | Oryza sativa L. | Heat and salinity stress tolerance | Deeba <i>et al</i> ., 2017 |
| | | Zea mays L. | High temperature and drought stress tolerance | Casaretto <i>et al.</i> , 2016 |
| | MVB | Arabidopsis thaliana (L.) Heynh. | Heat and drought stress tolerance | Zhao <i>et al</i> ., 2017 |
| | IVI I D | Nicotiana tabacum L. | Cold stress tolerance | Xing et al., 2019 |
| | | Arabidopsis thaliana (L.) Heynh. | Drought tolerance | Yu <i>et al</i> ., 2016 |
| Transcription | | Oryza sativa L. | Drought tolerance | Fávero <i>et al.</i> , 2018 |
| Factors (TFs) | | Oryza sativa L. | Cold and salt stress tolerance | Yang <i>et al.</i> , 2012 |
| | | Triticumaestivum L. | Drought and oxidative stress tolerance | Wei <i>et al.</i> , 2017 |
| | WRKY | Arabidopsis thaliana (L.) Heynh. | Heat stress tolerance | Li <i>et al.</i> , 2011 |
| | | Triticum aestivum L. | Drought stress tolerance | He et al., 2016 |
| | | Triticum aestivum L. | Heat stress tolerance | Wang <i>et al.</i> , 2017 |
| | | Arabidopsis thaliana (L.) Heynh. | Resistance to waterlogging | Raineri <i>et al.</i> , 2015 |
| | | Oryza sativa L. | Salt tolerance | Kobayashi <i>et al.</i> , 2017 |
| | | Hordeum vulgare L. | Salt stress tolerance | Han <i>et al.</i> , 2018 |
| OTI | | Triticum aestivum L. | Salt stress tolerance | Batool <i>et al.</i> , 2018 |
| QIL | | Oryza sativa L. | Salt stress tolerance | Islam <i>et al.</i> , 2019 |
| | | Cucumis sativus L. | High-temperature stress tolerance | Dong <i>et al.</i> , 2020 |
| | | Triticum aestivum L. | Freezing tolerance | Wąsek <i>et al.</i> , 2021 |
| | miR396b | Arabidopsis thaliana (L.) Heynh. | Cold tolerance | Zhang <i>et al.</i> , 2016 |
| MicroRNAs | miR319 | Oryza sativa L. | Cold tolerance | Liu <i>et al.</i> , 2017 |
| | miR393a | Agrostis stolonifera L. | Salt tolerance, drought tolerance, heat tolerance | Zhao <i>et al</i> ., 2018 |
| | miR172a | Glycine max (L.) Merr. | Salt stress tolerance | Pan <i>et al</i> ., 2016 |
| | miRNA169 a | Arabidopsis thaliana (L.) Heynh. | Drought stress tolerance | Ni <i>et al.</i> , 2013 |
| | miR169 | Solanum lycopersicum L. | Drought stress tolerance | Rao <i>et al.</i> , 2020 |
| | Osa-miR319 | Oryza sativa L. | Cold stress tolerance | Sun <i>et al.</i> , 2020 |

Table 1. Some selected studies for a better understanding of the role of biotechnology in abiotic stress tolerance in plants

| | | 1 | |
|----------------------|---|---|----------------------------------|
| Abiotic stress tool | Abiotic stress | Plant species | Reference |
| | Salt stress | Hordeum spontaneum L. | Kuang <i>et al</i> ., 2019 |
| | Cold and drought stress | Camellia sinensis L. | Wang <i>et al.</i> , 2016 |
| | Salt stress | Gossypium hirsutum L. | Gao <i>et al.</i> , 2016 |
| | Drought and salinity stress | Gossypium hirsutum L. | Liang <i>et al.</i> , 2016 |
| | Salt stress | Paulownia fortune L. | Fan <i>et al.</i> , 2016 |
| | Salt stress | Glycine max L. | Pan <i>et al.</i> , 2016 |
| | Salt stress | Populus cathayana L. | Qiu <i>et al.</i> , 2021 |
| Biotechnological | Salt stress | Oryza sativa L. | Ai <i>et al.</i> , 2021 |
| approach | Drought stress | Arabidopsis thaliana L. | Roca Paixão <i>et al.</i> , 2019 |
| | Salt stress | Oryza sativa L. | Farhat <i>et al</i> ., 2019 |
| | Drought stress | Phaseolus vulgaris L. | Dramadri <i>et al.</i> , 2019 |
| | Salt stress | Solanum lycopersicum L. | Tran <i>et al.</i> , 2021 |
| | Drought stress | Triticum aestivum L. | Shi <i>et al.</i> , 2018 |
| | Drought stress | Perennial ryegrass L. | Hang <i>et al.</i> , 2021 |
| | Salt stress | Arabidopsis thaliana L. | Denver <i>et al.</i> , 2019 |
| | Drought stress | Arachis hypogaea L. | Kiranmai <i>et al.</i> , 2018 |
| | Drought stress | Triticum aestivum L. | Ikram <i>et al.</i> , 2020 |
| | Salt stress | Fragaria × ananassa | Avestan <i>et al.</i> , 2019 |
| Nanahiatashnalagiasl | Drought stress | Zea mays L. | Karvar <i>et al</i> ., 2022 |
| approach | Drought stress | Oryza sativa L. 25S Arabidopsis thaliana L. Oryza sativa L. 25S Phaseolus vulgaris L. 25S Phaseolus vulgaris L. 25S Triticum aestivum L. 25S Perennial ryegrass L. 25S Arabidopsis thaliana L. 25S Perennial ryegrass L. 25S Arachis hypogaea L. 25S Triticum aestivum L. 25S Triticum aestivum L. 25S Triticum aestivum L. 25S Triticum aestivum L. 25S Solanum melongena L. 25S Solanum lycopersicum L. 25S Solanum lycopersicum L. 25S Glycine max L. 25S Ipomoea batatas [L.] Lam Transgenic chrysanthemum L. Arabidopsis thaliana L. 4 Stress Gossypium Hirsutum L. 4 stress Arachis hypogaea L. | Semida <i>et al.</i> , 2021 |
| approach | ical Drought stress Tr Salt stress Fr Drought stress Sola Drought stress Sola Salt stress Sola Salt stress Trace Drought and heat stress Tr Drought stress | Trachyspermum ammi L. | Abdoli <i>et al.</i> , 2020 |
| | Salt stress | Camellia sinensis L. Gossypium hirsutum L. Gossypium hirsutum L. Paulownia fortune L. Glycine max L. Populus cathayana L. Oryza sativa L. Arabidopsis thaliana L. Oryza sativa L. Phaseolus vulgaris L. Solanum lycopersicum L. Triticum aestivum L. Perennial ryegrass L. Arabidopsis thaliana L. Arabidopsis thaliana L. Arachis hypogaea L. Triticum aestivum L. Fragaria × ananassa Zea mays L. Solanum melongena L. Trachyspermum ammi L. Solanum lycopersicum L. Triticum aestivum L. Glycine max L. Ipomoea batatas [L.] Lam Transgenic chrysanthemum L. Gossypium Hirsutum L. Arabidopsis thaliana L. Arachis hypogaea L. Oryza sativa L. Agrostis stolonifera L. | Almutairi <i>et al.</i> , 2016 |
| | Drought and heat stress | Triticum aestivum L. | El-Esawi <i>et al.</i> , 2019 |
| | Drought stress | Glycine max L. | Ning et al., 2017 |
| | Drought stress | Ipomoea batatas [L.] Lam | Zhang <i>et al.</i> , 2019 |
| | Salt stress | Transgenic chrysanthemum L. | Liang <i>et al.</i> , 2017 |
| Conoria annroach | Drought and Salt Stress | Gossypium Hirsutum L. | Salisu <i>et al.</i> , 2021 |
| Genetic approach | Salt stress | Arabidopsis thaliana L. | Zhao <i>et al</i> ., 2019 |
| | Salt and drought stress | Arachis hypogaea L. | Banavath <i>et al.</i> , 2018 |
| | Drought stress | Oryza sativa L. | Takeuchi <i>et al.</i> , 2016 |
| | Drought and salt stress | Agrostis stolonifera L. | Takeuchi <i>et al.</i> , 2016 |

Table 2. Elevation of different abiotic stresses with various techniques used

NAC TFs

NAC is a category of TFs which plays a part in growth modulation of plants and is related to abiotic stress, morphogenesis, and metabolism. In a study conducted on Celery (*Apium graveolens* L.), a total of 111 putative NAC TFs were ascertained based on the celery transcriptome and genome database. These 111 NAC TFs were classified into 18 subfamilies based on their NAC domain. Real-time quantitative PCR (RT-qPCR) depicted that few *AgNAC* genes were distinctively expressed in presence of extreme environmental conditions (heat, cold, drought, and salt). It was observed that *AgNAC63* (ortholog of *ANAC072/RD26*) was stimulated by heat, cold, and salt conditions. From this study, it was concluded that AgNAC TFs might be involved in celery stress resistance (Duan *et al.*, 2020). In drought-stressed peanut plants, 132 AhNACs were described by the genomic study of the plants and were grouped into eight subgroups (I–VIII) based on their association with *Arabidopsis* NAC proteins. Transcriptomics exhibited several *AhNAC* genes that countered the water deficit and abscisic acid (ABA) stresses. Additionally, 20 of them were chosen in response to PEG and ABA application and were assessed by quantitative real-time polymerase chain reaction. Results depicted these genes

remarkably retaliated to water deficit and ABA in roots and/or leaves. The work was useful in directing the useful depiction and enhanced drought-resilient germplasms in peanuts (Li *et al.*, 2021). Heterologous overexpression of *Arabidopsis ANAC042/AtJUB1* in tomato (*S. lycopersicum*) and *Arabidopsis* enhanced abiotic stress resilience in these plants (Alshareef *et al.*, 2019). In wheat, it was revealed that *TaNAC47* gene is stimulated by low temperature, salinity, water deficiency, and ABA, depending on distinctive levels of expression. Overexpression of *TaNAC47* in transgenic Arabidopsis resulted in ABA vulnerability and increased resistance to above-mentioned stresses. *TaNAC47* overexpression also caused various metabolic alterations, because of the stimulation of downstream genes such as *AtRD29A*, *AtRD29B*, and *AtP5CS1*; the alterations might have facilitated the transgenic plants to curb the stress. ONAC5/6/9 and ONAC10, four rice NAC genes, were found to be overexpressed in a study, that led to enhanced drought resistance and decreased grain loss in presence of drought (Chung *et al.*, 2018). These researchers deduced that ONACs function as the cellular constituent that modulate several target genes which might change root structure for drought resistance (Chung *et al.*, 2018).

The MYB TFs

The MYB family is an extensive and diversified class of transcription factors in eukaryotes. The proteins belonging to this family are largely linked to protein-protein association, DNA attaching and regulatory activity control of proteins (Roy *et al.*, 2016). A number of MYB proteins have been described in modulating different cellular processes like stress responses and cell morphogenesis in various crop species (Ambawat *et al.*, 2013). Several MYB TFs are noticed to have remarkable function in modulating the temperature stress of temperature several plants. In tomatoes, overexpression of *LeAN2* enhanced plant resilience to high-temperature conditions via the higher activity of antioxidants (non-enzymatic) and decreased ROS accumulation (Meng *et al.*, 2015). It was also revealed that overexpression of *OsMYB1* gene enhances the resilience of rice plants to heat and salt stresses (Deeba *et al.*, 2017). Whilst *OsMYB55* in maize was effectual in improving high temperature and dehydration tolerance through increased growth of plants (Casaretto *et al.*, 2016). Six MYB genes were identified in wheat which were associated with heat stress and among them *TaMYB80* was effectual in imparting tolerance against high temperature and water deficit conditions in transgenic *Arabidopsis* (Zhao *et al.*, 2017). Likewise, *PbrMYB5* from *Pyrusbetulafolia* conferred resistance against cold stress by regulating the ascoribic acid synthesis in tobacco (Xing *et al.*, 2019).

WRKY TFs

WRKY is one of the most distinguished groups of a plant being TFs and in plants, it influences the various developmental, physiological, and metabolic processes (Raineri *et al.*, 2015). Recently it has been observed that WRKY proteins take part in several environmental stresses in numerous plants (Chen *et al.*, 2017; Qin *et al.*, 2015), Like in Arabidopsis, WRKY46, WRKY54, and WRKY70 were observed to function with BES1 TF to elevate brassinosteroid (BR)-modulated plant development, however, it contrastively modulated water deficiency tolerance (Raineri *et al.*, 2015). In Arabidopsis, salt stress stimulated *WRKY71* has been found to induce flowering in plants, hence, facilitating the early completion of the life cycle of the plants to avoid salt stress (Yu *et al.*, 2018). In addition to this, varied expression of maize *ZmWRKY17* in salt-stressed Arabidopsis led to decreased ABA susceptibility which was exhibited by thriving green cotyledons and long roots—concerning exogenously applied ABA however enhanced plant susceptibility to salt stress (Cai *et al.*, 2017). In rice, *OsWRKY30* and *OsWRKY47* have been revealed to bestow drought resistance. Concurrently, *OsWRKY71* acts as a promising modulator of cold stress resistance by modulating downstream target gene expression, such as *OsTGFR* and *WSI76* (Kim *et al.*, 2016). *TaWRKY146* from wheat showed a remarkable

expression in wheat seedlings after osmotic stress and manifested dehydration resilience to transgenic Arabidopsis by promoting closure of stomatal (Ma *et al.*, 2017).

Conclusions

Abiotic stresses pose a significant threat to crop productivity around the world, and their impact is expected to worsen in the future. Plant reactions to abiotic stressors are mediated by extremely complicated signalling networks, necessitating a multi-pronged effort to identify the genes involved and untangle the reaction for practical use. Plant abiotic stress has been an important topic for researchers in the last two decades, prompting the identification of candidate genes and transcriptional factors as well as the use of diverse biotechnological techniques. To combat abiotic challenges such as temperatures, drought stress, and metal stress, numerous transcription factors (NAC, ARF, MYB, SOC, MAPK, CBFs, and so on) have been found. MiRNAs have been discovered to play a role plant development, and stress responses in recent studies. Identification of miRNAs taking part in plant life processes and stress responses has been sped up because of better sequencing and bioinformatics technologies. In recent years, several in silico methods and databases have been of great assistance in identifying miRNAs implicated in plant development, and stress reactions, as well as predicting their target gene networks. For speedy deployment and adoption of the technique in the field, genome-edited crops using CRISPR/Cas9 must be deemed BE (bioengineered) crops. We believe that use of CRISPR/Cas9 approach in a variety of plant species could usher in a second green revolution, ensuring that need for need and nutritional security are satisfied for the world's growing population. As previously said, the world population is quickly expanding, and this has a negative impact on the environment. We can boost crop yield to fulfil the demand for food by employing genome editing technologies. We should exploit this chance to boost crop output and save the lives of millions of people who are starving due to lack of food globally, particularly in developing countries.

Authors' Contributions

The author read and approved the final manuscript.

Ethical approval (for researches involving animals or humans)

Not applicable.

Acknowledgements

This research received no specific grant from any funding agency in the public, commercial, or not-forprofit sectors.

Conflict of Interests

The authors declare that there are no conflicts of interest related to this article.

References

- Abdoli S, Ghassemi-Golezani K, Alizadeh-Salteh S (2020). Responses of ajowan (*Trachyspermum ammi* L.) to exogenous salicylic acid and iron oxide nanoparticles under salt stress. Environmental Science and Pollution Research 27(29):36939-36953. https://doi.org/10.1007/s11356-020-09453-1
- Adhikari L, Makaju SO, Missaoui AM (2019). QTL mapping of flowering time and biomass yield in tetraploid alfalfa (*Medicago sativa* L.). BMC Plant biology 19:359. *https://doi.org/10.1186/s12870-019-1946-0*
- Ai B, Chen Y, Zhao M, Ding G, Xie J, Zhang F (2021). Overexpression of miR1861h increases tolerance to salt stress in rice (*Oryza sativa* L.). Genetic Resources and Crop Evolution 68(1):87-92. https://doi.org/10.1007/s10722-020-01045-9
- Al-Ashkar I, Alderfasi A, Ben Romdhane W, Seleiman MF, El-Said RA, Al-Doss A (2020). Morphological and Genetic Diversity within Salt Tolerance Detection in Eighteen Wheat Genotypes. Plants 9(3):287. https://doi.org/10.3390/plants9030287
- Al-Ashkar I, Alderfasi A, El-Hendawy S, Al-Suhaibani N, El-Kafafi S, Seleiman MF (2019). Detecting Salt Tolerance in Doubled Haploid Wheat Lines. Agronomy 9(4): 211. https://doi.org/10.3390/agronomy9040211
- Alfatih A, Wu J, Jan SU, Zhang Z, Xia J, Xiang C (2020). Loss of rice PARAQUAT TOLERANCE 3 confers enhanced resistance to abiotic stresses and increases grain yield in field. Plant, Cell & Environment 43:2743-2754. https://doi.org/10.1111/pce.13856
- Almutairi, ZM (2016). Influence of silver nano-particles on the salt resistance of tomato (Solanum lycopersicum) during germination. International Journal of Agriculture and Biology 18(2):449-457. https://doi.org/10.17957/IJAB/15.0114
- Alsaeedi A, El-Ramady H, Alshaal T, El-Garawany M, Elhawat N, Al-Otaibi A (2019). Silica nanoparticles boost growth and productivity of cucumber under water deficit and salinity stresses by balancing nutrients uptake. Plant Physiology and Biochemistry 139: 1–10. *https://doi.org/10.1016/j.plaphy.2019.03.008*
- Alshareef NO, Wang JY, Ali S, Al-Babili S, Tester M, Schmöckel SM (2019). Overexpression of the NAC transcription factor JUNGBRUNNEN1 (JUB1) increases salinity tolerance in tomato. Plant Physiology and Biochemistry 140:113-121. https://doi.org/10.1016/j.plaphy.2019.04.038
- Ambawat S, Sharma P, Yadav NR, Yadav RC (2013). MYB transcription factor genes as regulators for plant responses: An overview. Physiology and Molecular Biology of Plants 19:307-321. https://doi.org/10.1007/s12298-013-0179-1
- Ansari WA, Chandanshive SU, Bhatt V, Nadaf AB, Vats S, Katara JL, Sonah H, Deshmukh R (2020). Genome editing in cereals: approaches, applications and challenges. International Journal of Molecular Sciences 21(11):4040. https://doi.org/10.3390/ijms21114040
- Arora NK (2019). Impact of climate change on agriculture production and its sustainable solutions. Environmental Sustainability 2:95-96. https://doi.org/10.1007/s42398-019-00078-w
- Ashkavand P, Tabari M, Zarafshar M, Tomásková I, Struve D (2015). Effect of SiO2 nanoparticles on drought resistance in hawthorn seedlings. Leśne Prace Badawcze 76(4).
- Avestan S, Ghasemnezhad M, Esfahani M, Byrt CS (2019). Application of nano-silicon dioxide improves salt stress tolerance in strawberry plants. Agronomy 9:246. https://doi.org/10.3390/agronomy9050246
- Baillo EH, Kimotho RN, Zhang Z, Xu P (2019). Transcription factors associated with abiotic and biotic stress toleranc and their potential for crops improvement. Genes 10: 771. https://doi.org/10.3390/genes10100771
- Banavath JN, Chakradhar T, Pandit V, Konduru S, Guduru KK, Akila CS, Podha S, Puli CO (2018). Stress inducible overexpression of AtHDG11 leads to improved drought and salt stress tolerance in peanut (*Arachis hypogaea* L.). Frontiers in Chemistry 2:6-34. https://doi.org/10.3389/fchem.2018.00034
- Batool N, Ilyas N, Shahzad A, Hauser BA, Arshad M (2018). Quantitative trait loci (QTLs) mapping for salt stress tolerance in wheat at germination stage. Pakistan Journal of Agricultural Sciences 55(1):47-55. https://doi.org/10.21162/PAKJAS/18.5426
- Batool T, Ali S, Seleiman MF, Naveed NH, Ali A, Ahmend K, Abid M, Rizwan M, Shahid MR, Alotaibi M, et al. (2020). Plant growth promoting rhizobacteria alleviates drought stress in potato in response to suppressive oxidative stress and antioxidant enzymes activities. Scientific Report 10:16975. https://doi.org/10.1038/s41598-020-73489-z

- Bhat MA, Bhat MA, Kumar V, Wani IA, Bashir H, Shah AA, Rahman S, Jan AT (2020). The era of editing plant genomes using CRISPR/Cas: a critical appraisal. Journal of Biotechnology 324:34-60. https://doi.org/10.1016/j.jbiotec.2020.09.013
- Cai R, Dai W, Zhang C, Wang Y, Wu M, Zhao Y, Ma Q, Xiang Y, Cheng B (2017). The maize WRKY transcription factor ZmWRKY17 negatively regulates salt stress tolerance in transgenic *Arabidopsis* plants. Planta 246:1215-1231. https://doi.org/10.1007/s00425-017-2766-9
- Caño-Delgado A, Yin Y, Yu C, Vafeados D, Mora-García S, Cheng JC, Nam KH, Li J, Chory J (2004). BRL1 and BRL3 are novel brassinosteroid receptors that function in vascular differentiation in *Arabidopsis*. Development 131:5341-5351. https://doi.org/10.1242/dev.01403
- Casaretto JA, El-kereamy A, Zeng B, Stiegelmeyer SM, Chen X, Bi Y-M, Rothstein SJ (2016). Expression of OsMYB55 in maize activates stress-responsive genes and enhances heat and drought tolerance. BMC Genomics 17:312. https://doi.org/10.1186/s12864-016-2659-5
- Chen D, Chai S, McIntyre CL, Xue G-P (2018). Overexpression of a predominantly root-expressed NAC transcription factor in wheat roots enhances root length, biomass and drought tolerance. Plant Cell Reports 37:225-237. https://doi.org/10.1007/s00299-017-2224-y
- Chen F, Hu Y, Vannozzi A, Wu K, Cai H, Qin Y, Mullis A, Lin Z, Zhang L (2017). The WRKY transcription factor family in model plants and crops. Critical Reviews in Plant Sciences 36:311-335. https://doi.org/10.1080/07352689.2018.1441103
- Cheng Q, Dong L, Su T, Li T, Gan Z, Nan H, Lu S, Fang C, Kong L, Li H, Hou Z (2019). CRISPR/Cas9-mediated targeted mutagenesis of GmLHY genes alters plant height and internode length in soybean. BMC Plant Biology 19(1):1-11. https://doi.org/10.1186/s12870-019-2145-8
- Choudhary SP, Yu JQ, Yamaguchi-Shinozaki K, Shinozaki K, Tran LS (2012). Benefits of brassinosteroid crosstalk. Trends in Plant Sciences 17:594-605. *https://doi.org/10.1016/j.tplants.2012.05.012*
- Chung PJ, Jung H, Choi YD, Kim JK (2018). Genome-wide analyses of direct target genes of four riceNAC-domain transcription factors involved in drought tolerance. BMC Genomics 19:40. *https://doi.org/10.1186/s12864-017-4367-1*
- Collins NC, Tardieu F, Tuberosa R (2008). Quantitative trait loci and crop performance under abiotic stress: Where do we stand? Plant Physiology 147:469-486. *https://doi.org/10.1104/pp.108.118117*
- Deeba F, Sultana T, Javaid B, Mahmood T, Naqvi S (2017). Molecular characterization of a MYB protein from Oryzasativa for its role in abiotic stress tolerance. Brazilian Archives of Biology and Technology 60:1-12. https://doi.org/10.1590/1678-4324-2017160352
- Demirer GS, Zhang H, Goh NS, Gonz 'alez-Grandío E, Landry MP (2019). Carbon nanotube–mediated DNA delivery without transgene integration in intact plants. Nature Protocols 14:2954-2971. *https://doi.org/10.1038/s41596-019-0208-9*
- Denver JB, Ullah H (2019). miR393s regulate salt stress response pathway in Arabidopsis thaliana through scaffold protein RACK1A mediated ABA signaling pathways. Plant Signaling & Behavior 14(6):1600394. https://doi.org/10.1080/15592324.2019.1600394
- Dita MA, Rispail N, Prats E, Rubiales D, Singh KB (2006). Biotechnology approaches to overcome biotic and abiotic stress constraints in legumes. Euphytica 147:1-24. *https://doi.org/10.1007/s10681-006-6156-9*
- Dong S, Zhang S, Wei S, Liu Y, Li C, Bo K, Miao H, Gu X, Zhang S (2020). Identification of Quantitative Trait Loci controlling high-temperature tolerance in cucumber (*Cucumis sativus* L.) seedlings. Plants 9(9):155. https://doi.org/10.3390/plants9091155
- Dramadri IO, Nkalubo ST, Kelly JD (2019). Identification of QTL associated with drought tolerance in Andean common bean. Crop Science 59(3):1007-1020. *https://doi.org/10.2135/cropsci2018.10.0604*
- Duan AQ, Yang XL, Feng K, Liu JX, Xu ZS, Xiong AS (2020). Genome –ide analysis of NAC transcription factors and their response to abiotic stress in celery (*Apium graveolens* L.). Computational Biology and Chemistry 84:07186. https://doi.org/10.1016/j.compbiolchem.2019.107186
- El-Esawi MA, Al-Ghamdi AA, Ali HM, Ahmad M (2019). Overexpression of AtWRKY30 transcription factor enhances heat and drought stress tolerance in wheat (*Triticum aestivum* L.). Genes 10(2):163. https://doi.org/10.3390/genes10020163

- Elshayb OM, Nada AM, Sadek AH, Ismail SH, Shami A, Alharbi BM, ... Seleiman MF (2022). The integrative effects of biochar and ZnO nanoparticles for enhancing rice productivity and water use efficiency under irrigation deficit conditions. Plants 11(11):1416. https://doi.org/10.3390/plants11111416
- Fan G, Li X, Deng M, Zhao Z, Yang L (2016). Comparative analysis and identification of miRNAs and their target genes responsive to salt stress in diploid and tetraploid *Paulownia fortunei* seedlings. PLoS One 11(2):e0149617. https://doi.org/10.1371/journal.pone.0149617
- Farhat S, Jain N, Singh N, Sreevathsa R, Dash PK, Rai R, Yadav S, Kumar, Sarkar AK, Jain A, Singh NK (2019). CRISPR-Cas9 directed genome engineering for enhancing salt stress tolerance in rice. Cell and Developmental Biology 96:91-99. https://doi.org/10.1016/j.semcdb.2019.05.003
- Fávero PJR, Mara DAL, Dos SBM (2018). Overexpression of ScMYBAS1 alternative splicing transcripts differentially impacts biomass accumulation and drought tolerance in rice transgenic plants. PLoS One 13:e0207534. https://doi.org/10.1371/journal.pone.0207534
- Fincheira P, Tortella G, Duran N, Seabra AB, Rubilar O (2020). Current applications of nanotechnology to develop plant growth inducer agents as an innovation strategy. Critical Reviews in Biotechnology 40:15-30. https://doi.org/10.1080/07388551.2019.1681931
- Gantait S, Mondal S (2018). Transgenic approaches for genetic improvement in groundnut (*Arachis hypogaea* L.) against major biotic and abiotic stress factors. Journal of Genetic Engineering and Biotechnology 16: 537–544. https://doi.org/10.1016/j.jgeb.2018.08.005
- Gao S, Yang L, Zeng HQ, Zhou ZS, Yang ZM, Li H, Sun D, Xie F, Zhang B (2016). A cotton miRNA is involved in regulation of plant response to salt stress. Scientific Reports 6(1):19736. *https://doi.org/10.1038/srep19736*
- Giraldo JP, Landry MP, Faltermeier SM, McNicholas TP, Iverson NM, Boghossian AA, Reuel NF, Hilmer AJ, Sen F, Brew JA, Strano MS (2014). Plant nanobionics approach to augment photosynthesis and biochemical sensing. Nature Materials 13:400-408. https://doi.org/10.1038/nmat3890.
- Haliloglu K, Hosseinpour A, Cinisli Kan T, Ozturk H, Ozkan G, Pour-Aboughadareh A, Poczai P (2020). Investigation of the protective roles of zinc oxide nanoparticles and plant growth promoting bacteria on DNA damage and methylation in tomato (*Solanum lycopersicum* L.) under salinity stress. Horticulture, Environment, and Biotechnology 12:245.
- Han Y, Yin S, Huang L, Wu X, Zeng J, Liu X, Qiu L, Munns R, Chen ZH, Zhang G (2018). A Sodium Transporter HvHKT1;1 Confers salt tolerance in barley via regulating tissue and cell ion homeostasis. Plant & Cell Physiology 59:1976-1989. https://doi.org/10.1093/pcp/pcy116
- Hang N, Shi T, Liu Y, Ye W, Taier G, Sun Y, Wang K, Zhang W (2021). Overexpression of Os-microRNA408 enhances drought tolerance in perennial ryegrass. Physiologia Plantarum 172(1):733-747. https://doi.org/10.1111/ppl.13276
- He G-H, Xu J-Y, Wang Y-X, Liu J-M, Li P-S, Chen M, Ma Y-Z, Xu Z-S (2016). Drought-responsive WRKY transcription factor genes TaWRKY1 and TaWRKY33 from wheat confer drought and/or heat resistance in Arabidopsis. BMC Plant Biology 16:116. *https://doi.org/10.1186/s12870-016-0806-4*
- He Z, Li Z, Lu H, Huo L, Wang Z, Wang Y, Ji X (2019). The NAC protein from Tamarixhispida, ThNAC7, confers salt and osmotic stress tolerance by increasing reactive oxygen species scavenging capability. Plants 8:221. https://doi.org/10.3390/plants8070221
- Hojjat S.S (2019). Effect of interaction between Ag nanoparticles and salinity on germination stages of *Lathyrus sativus* L. Open Access Journal of Environmental and Soil Sciences 2(2):193-198. https://doi.org/10.32474/OAJESS.2019.02.000132
- Hong Y, Meng J, He X, Zhang Y, Liu Y, Zhang C, Qi H, Luan Y (2020). Editing miR482b and miR482c simultaneously by CRISPR/Cas9 enhanced tomato resistance to *Phytophthora infestans*. Phytopathology 111(6):1008-1016. https://doi.org/10.1094/PHYTO-08-20-0360-R
- Ikram M, Raja NI, Javed B, Hussain M, Hussain M, Ehsan M, Rafique N, Malik K, Sultana T, Akram A (2020). Foliar applications of bio-fabricated selenium nanoparticles to improve the growth of wheat plants under drought stress. Green Processing and Synthesis 9(1):706-714. https://doi.org/10.1515/gps-2020-0067
- Islam M, Ontoy, Subudhi PK (2019). Meta-Analysis of quantitative trait loci associated with seedling-stage salt tolerance in rice (*Oryza sativa* L.). Plants 8(2):33. *https://doi.org/10.3390/plants8020033*

- Jain M (2015). Function genomics of abiotic stress tolerance in plants: a CRISPR approach. Frontiers in Plant Science 6:2011-2014. https://doi.org/10.3389/fpls.2015.00375
- Jatan R, Lata CH (2019). Role of microRNAs in abiotic and biotic stress resistance in plants. Proceedings of the Indian National Science Academy 85:553-567. *https://doi.org/10.1007/s12010-014-0914-2*
- Jian H, Yang BO, Zhang A, Ma J, Ding Y, Chen Z, Li J, Xu X, Liu L (2018). Genome wide identification of microRNAs in response to cadmium stress in oilseed rape (*Brassica napus* L.) using high-throughput sequencing. International Journal of Molecular Sciences 19:1431. https://doi.org/10.3390/ijms19051431
- Jin C, Li K-Q, Xu X-Y, Zhang H-P, Chen H-X, Chen Y-H, Hao J, Wang Y, Huang X-S, Zhang S-L (2017). A novel NAC transcription factor, PbeNAC1, of *Pyrus betulifolia* confers cold and drought tolerance via interacting with PbeDREBs and activating the expression of stress-responsive genes. Frontiers in Plant Science 8:1049. https://doi.org/10.3389/fpls.2017.01049
- Kalteh M, Alipour ZT, Ashraf S, Marashi Aliabadi M, Falah Nosratabadi A (2018). Effect of silica nanoparticles on basil (*Ocimum basilicum*) under salinity stress. Journal of Chemical Health Risks 4(3):49-55.
- Karvar M, Azari A, Rahimi A (2022). Titanium dioxide nanoparticles (TiO2-NPs) enhance drought tolerance and grain yield of sweet corn (*Zea mays* L.) under deficit irrigation regimes. Acta Physiologiae Plantarum 44:14. https://doi.org/10.1007/s11738-021-03349-4
- Kim C-Y, Vo KTX, Nguyen CD, Jeong D-H, Lee S-K, Kumar M, Kim S- R, Park S-H, Kim J-K, Jeon J-S (2016). Functional analysis of a cold-responsive rice WRKY gene, OsWRKY71. Plant Biotechnology Reports 10:13-23. https://doi.org/10.1007/s11816-015-0383-2
- Kiranmai K, Lokanadha Rao G, Pandurangaiah M, Nareshkumar A, Amaranatha Reddy V, Lokesh U, ... Sudhakar C (2018). A novel WRKY transcription factor, MuWRKY3 (*Macrotyloma uniflorum* Lam. Verdc.) enhances drought stress tolerance in transgenic groundnut (*Arachis hypogaea* L.) plants. Frontiers in Plant Science 9:346. https://doi.org/10.3389/fpls.2018.00346
- Kobayashi NI, Yamaji N, Yamamoto H, Okubo K, Ueno H, Costa A, Tanoi K, Matsumura H, Fujii-Kashino M, Horiuchi T, et al. (2017). OsHKT1;5 mediates Na⁺ exclusion in the vasculature to protect leaf blades and reproductive tissues from salt toxicity in rice. Plant Journal 91:657-670. *https://doi.org/10.1111/tpj.13595*
- Kuang L, Shen Q, Wu L, Yu J, Fu L, Wu D, Zhang G (2019). Identification of microRNAs responding to salt stress in barley by high-throughput sequencing and degradome analysis. Environmental and Experimental Botany 160:59-70. https://doi.org/10.1016/j.envexpbot.2019.01.006
- Li C, Yan C, Sun Q, Yuan C, Mou Y, Shan S, Zhao X (2021). The BHLH transcription factor AhbHLH112 improves the drought tolerance of peanut. BMC Plant Biology 21: 1-12. https://doi.org/10.1186/s12870-021-03318-6
- Li S, Fu Q, Chen L, Huang W, Yu D (2011). *Arabidopsis thaliana* WRKY25, WRKY26, and WRKY33 coordinate induction of plant thermotolerance. Planta 233:1237-1252. *https://doi.org/10.1007/s00425-011-1375-2*
- Li X, Wang Y, Chen S, Tian H, Fu D, Zhu B, Luo Y, Zhu H (2018). Lycopene is enriched in tomato fruit by CRISPR/Cas9-mediated multiplex genome editing. Frontiers in Plant Science 9:559. https://doi.org/10.3389/fpls.2018.00559
- Li Y, Zhu N, Liang X, Bai X, Zheng L, Zhao J, Li Y, Zhang Z, Gao Y (2020). Silica nanoparticles alleviate mercury toxicity via immobilization and inactivation of Hg(ii) in soybean (*Glycine max*). Environmental Science: Nano 7:1807-1817. https://doi.org/10.1039/D0EN00091D
- Li Z, Cheng Q, Gan Z, Hou Z, Zhang Y, Li Y, Li H, Nan H, Yang C, Chen L, Lu S (2020). Multiplex CRISPR/Cas9mediated knockout of soybean LNK2 advances flowering time. Crop Journal 9(4):767-776. https://doi.org/10.1016/j.cj.2020.09.005.
- Liang C, Meng Z, Meng Z, Malik W, Yan R, Lwin K.M, Lin F, Wang Y, Sun G, Zhou T, Zhu T (2016). GhABF2, a bZIP transcription factor, confers drought and salinity tolerance in cotton (*Gossypium hirsutum* L.). Scientific Reports 6(1):1-4. https://doi.org/10.1038/srep35040
- Liang QY, Wu YH, Wang K, Bai ZY, Liu QL, Pan YZ, Zhang L, Jiang BB (2017). Chrysanthemum WRKY gene DgWRKY5 enhances tolerance to salt stress in transgenic chrysanthemum. Scientific Reports 7(1):1-10. https://doi.org/10.1038/s41598-017-05170-x
- Liu Q, Yan S, Yang T, Zhang S, Chen Y-Q, Liu B (2017). Small RNAs in regulating temperature stress response in plants. Journal of Integrative Plant Biology 59:774-791. *https://doi.org/10.1111/jipb.12571*

- Ma J, Gao X, Liu Q, Shao Y, Zhang D, Jiang L, Li C (2017). Overexpression of TaWRKY146 increases drought tolerance through inducing stomatal closure in *Arabidopsis thaliana*. Frontiers in Plant Science 8:2036. https://doi.org/10.3389/fpls.2017.02036
- Meng X, Wang J-R, Wang G-D, Liang X-Q, Li X-D, Meng Q-W (2015). An R2R3-MYB gene, LeAN2, positively regulated the thermo-tolerance in transgenic tomato. Journal of Plant Physiology 175:1-8. https://doi.org/10.1016/j.jplph.2014.09.018
- Mishra GP, Singh B, Seth T, Singh AK, Halder J, Krishnan N, Tiwari SK, Singh PM (2017). Biotechnological advancements and begomovirus management in Okra (*Abelmoschus esculentus* L.), status and perspectives. Frontiers in Plant Science 8:360. https://doi.org/10.3389/fpls.2017.00360
- Muehlbauer FJ, Cho S, Sarker A, McPhee KE, Coyne CJ, Rajesh PN, Ford R (2006). Application of biotechnology in breeding lentil for resistance to biotic and abiotic stress. Euphytica 147:149-165. *https://doi.org/10.1007/s10681-006-7108-0*
- Mukhtar T, Rehman Su, Smith D, Sultan T, Seleiman MF, Alsadon AA, Amna, Ali S, Chaudhary HJ, Solieman THI, Ibrahim AA, Saad MAO (2020) Mitigation of Heat Stress in *Solanum lycopersicum* L. by ACC-deaminase and Exopolysaccharide Producing Bacillus cereus: Effects on Biochemical Profiling. Sustainability 12(6):2159. https://doi.org/10.3390/su12062159
- Mushtaq M, Sakina A, Wani SH, Shikari AB, Tripathi P, Zaid A, Galla A, Abdelrahman M, Sharma M, Singh AK (2019). Harnessing genome editing techniques to engineer disease resistance in plants. Frontiers in Plant Science 10:550. https://doi.org/10.3389/fpls.2019.00550
- Nadarajah K, Kumar IS (2019). Drought response in rice: The miRNA story. International Journal of Molecular Sciences 20:3766. https://doi.org/10.3390/ijms20153766
- Ni Z, Hu Z, Jiang Q, Zhang H (2013). GmNFYA3, a target gene of miR169, is a positive regulator of plant tolerance to drought stress. Plant Molecular Biology 82:113-129. *https://doi.org/10.1007/s11103-013-0040-5*
- Nieves-Cordones M, Mohamed S, Tanoi K, Kobayashi NI, Takagi K, Vernet A, Guiderdoni E, Périn C, Sentenac H, Véry AA (2017). Production of low-Cs⁺ rice plants by inactivation of the K+ transporter OsHAK1 with the CRISPR-Cas system. The Plant Journal 92:43-56. *https://doi.org/10.1111/tpj.13632*
- Ning W, Zhai H, Yu J, Liang S, Yang X, Xing X, Huo J, Pang T, Yang Y, Bai X (2017). Overexpression of Glycine soja WRKY20 enhances drought tolerance and improves plant yields under drought stress in transgenic soybean. Molecular Breeding 37(2):19. https://doi.org/10.1007/s11032-016-0614-4
- Noman A, Aqeel M, Deng J, Khalid N, Sanaullah T, Shuilin H (2017). Biotechnological advancements for improving floral attributes in ornamental plants. Frontiers in Plant Science 8:530. *https://doi.org/10.3389/fpls.2017.00530*
- Osakabe Y, Osakabe K (2017). Genome editing to improve abiotic stress responses in plants. Progress in Molecular Biology and Translational Science 149:99-109. https://doi.org/10.1016/bs.pmbts.2017.03.007
- Osakabe Y, Watanabe T, Sugano SS, Ueta R, Ishihara R (2016). Optimization of CRISPR/Cas9 genome editing to modify abiotic stress responses in plants. Scientific Reports 6:26685. *https://doi.org/10.1038/srep26685*
- Paixão JFR, Gillet F, Ribeiro TP, Bournaud C, Lourenço-tess IT, Noriega DD, De Melo BP, De Almeida-engler J, Grosside-sa MF (2019). Improved drought stress tolerance in Arabidopsis by CRISPR/dCas9 fusion with a histone AcetylTransferase. Scientific Reports 9:8080. https://doi.org/10.1038/s41598-019-44571-y
- Pan WJ, Tao JJ, Cheng T, Bian XH, Wei W, Zhang WK, Ma B, Chen SY, Zhang JS (2016). Soybean miR172a improves salt tolerance and can function as a long-distance signal. Molecular Plant 9(9):1337-1340. https://doi.org/10.1016/j.molp.2016.05.010
- Pandey S, Fartyal D, Agarwal A, Shukla T, James D, Kaul T, Negi Y.K, Arora S, Reddy MK (2017). Abiotic stress tolerance in plants: Myriad roles of ascorbate peroxidase. Frontiers in Plant Science 8:581. https://doi.org/10.3389/fpls.2017.00581
- Qin Y, Tian Y, Liu X (2015). A wheat salinity-induced WRKY transcription factor TaWRKY93 confers multi pleabiotic stress tolerance in Arabidopsis thaliana. Biochemical and Biophysical Research Communications 464:428-433. https://doi.org/10.1016/j.bbrc.2015.06.128
- Qiu T, Du K, Jing Y, Zeng Q, Liu Z, Li Y, Ren Y, Yang J, Kang X (2021). Integrated transcriptome and miRNA sequencing approaches provide insights into salt tolerance in allotriploid *Populus cathayana*. Planta 254:25. https://doi.org/10.1007/s00425-021-03600-9

- Rai MK, Kalia RK, Singh R, Gangola MP, Dhawan AK (2011). Developing stress tolerant plants through in vitro selection—an overview of the recent progress. Environmental and Experimental Botany 71:89-98. https://doi.org/10.1016/j.envexpbot.2010.10.021
- Raineri J, Ribichich KF, Chan RL (2015). The sunflower transcription factor HaWRKY76 confers drought and flood of Arabidopsis thaliana plants without yield penalty. Plant Cell Reports 34:2065-2080. https://doi.org/10.1007/s00299-015-1852-3
- Rao S, Balyan S, Jha S, Mathur S (2020). Novel insights into expansion and functional diversification of MIR169family in tomato. Planta 251:55. *https://doi.org/10.1007/s00425-020-03346-w*
- Roy S (2016). Function of MYB domain transcription factors in abiotic stress and epigenetic control of stress response in plant genome. Plant Signaling & Behavior 11:e1117723. *https://doi.org/10.1080/15592324.2015.1117723*
- Roy R, Núez-Delgado A, Sultana S, Wang J, Mmunirf A, Battaglia M, Sarker T, Seleiman MF, Barmon M, Zhang RQ (2021). Additions of optimum water, spent mushroom compost and wood biochar to improve the growth performance of althaea rosea in drought-prone coal-mined spoils. Journal of Environmental Management 295:113076. https://doi.org/10.1016/j.jenvman.2021.113076
- Salisu SB, Mehari TG, Ahmad A, Tajo SM, Ibrahim S, Iqbal MS, Elasad M, Zhang J, Wei H, Yu S (2021). Genome wide identification and characterization of mitogen activated protein kinase (MAPK) genes reveals their potential in enhancing drought and salt stress tolerance in *Gossypium hirsutum*. Research Square PPR432727. https://doi.org/10.21203/rs.3.rs-1078536/v1
- Sander JD, Joung JK (2014). CRISPR-Cas systems for editing, regulating and targeting genomes. Nature Biotechnology 32(4):347-355. *https://doi.org/10.1038/nbt.2842*
- Saxena R, Tomar RS, Kumar M (2016). Exploring nano biotechnology to mitigate abiotic stress in crop plants. Journal of Pharmaceutical Sciences and Research 8(9):974.
- Seleiman MF (2019). Use of plant nutrients in improving abiotic stress tolerance in wheat. In: Hasanuzzaman M, Nahar K, Hossain M (Eds). Wheat Production in Changing Environments. Springer, Singapore: pp 481-495. https://doi.org/10.1007/978-981-13-6883-7_19
- Seleiman MF, Almutairi KF, Alotaibi M, Shami A, Alhammad BA, Battaglia ML (2021a). Nano-Fertilization as an Emerging Fertilization Technique: Why Can Modern Agriculture Benefit from Its Use? Plants 10(1):2. https://doi.org/10.3390/plants10010002
- Seleiman MF, Al-Suhaibani N, Ali N, Akmal M, Alotaibi M, Refay Y, Dindaroglu T, Abdul-Wajid HH, Battaglia ML (2021b). Drought Stress Impacts on Plants and Different Approaches to Alleviate Its Adverse Effects. Plants 10(2):259. https://doi.org/10.3390/plants10020259
- Seleiman MF, Kheir AM (2018). Saline soil properties, quality and productivity of wheat grown with bagasse ash and thiourea in different climatic zones. Chemosphere 193:538-546. https://doi.org/10.1016/j.chemosphere.2017.11.053
- Seleiman MF, Santanen A, Mäkelä P (2020). Recycling sludge on cropland as fertilizer-Advantages and risks. Resources, Conservation & Recycling 155:104647. *https://doi.org/10.1016/j.resconrec.2019.104647*
- Seleiman MF, Semida WM, Rady MM, Mohamed GF, Hemida KA, Alhammad BA, Hassan MM, Shami A (2020). Sequential Application of Antioxidants Rectifies Ion Imbalance and Strengthens Antioxidant Systems in Salt-Stressed Cucumber. Plants 9(12):1783. https://doi.org/10.3390/plants9121783
- Semida WM, Abdelkhalik A, Mohamed GF, Abd El-Mageed TA, Abd El-Mageed SA, Rady MM, Ali EF (2021). Foliar application of zinc oxide nanoparticles promotes drought stress tolerance in eggplant (Solanum melongena L.). Plants 10:421. https://doi.org/10.3390/plants10020421
- Sharma A, Ruiz-Manriquez LM, Serrano-Cano FI, Reyes-Perez PR, Tovar Alfaro CK, Baron Andrade YE, Hernandez Aros AK, Srivastava A, Paul S (2020). Identification of microRNAs and their expression in leaf tissues of guava (*Psidium guajava* L.) under salinity stress. Agronomy 10:1920. https://doi.org/10.3390/agronomy10121920
- Sharma DK, Torp AM, Rosenqvist E, Ottosen C-O, Andersen SB (2017). QTLs and potential candidate genes for heat stress tolerance identified from the mapping populations specifically segregating for Fv/Fm in wheat. Frontiers in Plant Science 8:1668. *https://doi.org/10.3389/fpls.2017.01668*
- Shen L, Wang C, Fu Y, Wang J, Liu Q, Zhang X, Yan C, Qian Q, Wang K (2018). QTL editing confers opposing yield performance in different rice varieties. Journal of Integrative Plant Biology 60:89-93. https://doi.org/10.1111/jipb.12501

- Shi GQ, Fu JY, Rong LJ, Zhang PY, Guo CJ, Kai XI (2018). TaMIR1119, a miRNA family member of wheat (*Triticum aestivum*), is essential in the regulation of plant drought tolerance. Journal of Integrative Agriculture 17(11): 2369-2378. https://doi.org/10.1016/S2095-3119(17)61879-3
- Shi J, Gao H, Wang H, Lafitte HR, Archibald R.L, Yang M, Hakimi SM, Mo H, Habben JE (2017). ARGOS8 variants generated by CRISPR-Cas9 improve maize grain yield under field drought stress conditions. Plant Biotechnology Journal 15:207-216. https://doi.org/10.1111/pbi.12603
- Shim JS, Oh N, Chung PJ, Kim YS, Choi YD, Kim JK (2018). Overexpression of OsNAC14 improves drought tolerance in rice. Frontiers in Plant Science 9:310. *https://doi.org/10.3389/fpls.2018.00310*
- Singh VK, Singh AK (2019). Role of microbially synthesized nanoparticles in sustainable agriculture and environmental management. Role of Plant Growth Promoting Microorganisms in Sustainable Agriculture and Nanotechnology. Woodhead Publishing 2019, pp 55-73. https://doi.org/10.1016/B978-0-12-817004-5.00004-X
- Sun M, Shen Y, Yang J, Cai X, Li H, Zhu Y, Jia B, Sun X (2020). miR535 negatively regulates cold tolerance in rice. Molecular Breeding 40:14. https://doi.org/10.1007/s11032-019-1094-0
- Taha R, Seleiman MF, Alotaibi M, Alhammad BA, Rady MM, Mahdi AHA (2020). Exogenous potassium treatments elevate salt tolerance and performances of *Glycine max* L. by boosting antioxidant defense system under actual saline field conditions. Agronomy 10(11):1741. *https://doi.org/10.3390/agronomy10111741*
- Takeuchi K, Hasegawa H, Gyohda A, Komatsu S, Okamoto T, Okada K, Terakawa T, Koshiba T (2016). Overexpression of RSOsPR10, a root-specific rice PR10 gene, confers tolerance against drought stress in rice and drought and salt stresses in bentgrass. Plant Cell, Tissue and Organ Culture 127(1):35-46. *https://doi.org/10.1007/s11240-016-1027-0*
- Tang L, Mao B, Li Y, Lv Q, Zhang L, Chen C, ... Zhao B (2017). Knockout of OsNramp5 using the CRISPR/Cas9 system produces low cd-accumulating indica rice without compromising yield. Scientific Reports 7:14438. https://doi.org/10.1038/s41598-017-14832-9
- Tang X, Lowder LG, Zhang T, Malzahn AA, Zheng X, Voytas DF, Zhong Z, Chen Y, Ren Q, Li Q (2017). A CRISPR– Cpf1 system for efficient genome editing and transcriptional repression in plants. Nature Plants 3:17018. https://doi.org/10.1038/nplants.2017.18
- Tiwari S, Lata C (2019). Genome engineering in rice: applications, advancements and future perspectives. In: Singh S, Upadhyay S, Pandey A, Kumar S (Eds). Molecular Approaches in Plant Biology and Environmental Challenges. Energy, Environment, and Sustainability. Springer, Singapore, pp 323-337. https://doi.org/10.1007/978-981-15-0690-1_15
- Tran MT, Doan DT, Kim J, Song YJ, Sung YW, Das S, Kim EJ, Son GH, Kim SH, Van Vu T, Kim JY (2021). CRISPR/Cas9-based precise excision of SlHyPRP1 domain (s) to obtain salt stress-tolerant tomato. Plant Cell Reports 40(6):999-1011. https://doi.org/10.1007/s00299-020-02622-z
- Upadhyay U, Singh P, Verma OP (2019). Role of microRNAs in regulating drought stress tolerance in maize. Journal of Pharmacognosy and Phytochemistry 8:328-331.
- Usman M, Farooq M, Wakeel A, Nawaz A, Cheema SA, Rehman H, Ashraf I, Sanaullah M (2020). Nanotechnology in agriculture: current status, challenges and future opportunities. Science of the Total Environment 721:137778. https://doi.org/10.1016/j.scitotenv.2020.137778
- Vakilian KA (2020). Machine learning improves our knowledge about miRNA functions towards plant abiotic stresses. Scientific Reports 10:3041. *https://doi.org/10.1038/s41598-020-59981-6*
- Wang J, Tao F, An F, Zou Y, Tian W, Chen X, Xu X, Hu X (2017). Wheat transcription factor TaWRKY70 ispositively involved in high-temperature seedling plant resistance to *Puccinia striiformis* f. sp. tritici. Molecular Plant Pathology 18:649-661. https://doi.org/10.1111/mpp.12425
- Wang Q, Liu N, Yang X, Tu L, Zhang X (2016). Small RNA-mediated responses to low- and high-temperature stresses in cotton. Scientific Reports 6:35558. https://doi.org/10.1038/srep35558
- Wang Y, Shu Z, Wang W, Jiang X, Li D, Pan J, Li X (2016). CsWRKY2, a novel WRKY gene from Camellia sinensis, is involved in cold and drought stress responses. Biologia Plantarum 60(3):443-451. https://doi.org/10.1007/s10535-016-0618-2
- Wąsek I, Dyda M, Gołębiowska G, Tyrka M, Rapacz M, Szechyńska-Hebda M, Wędzony M (2021). Quantitative trait loci and candidate genes associated with freezing tolerance of winter triticale (× Triticosecale Wittmack). Journal of Applied Genetics 63:15-33. https://doi.org/10.1007/s13353-021-00660-1

- Wei Q, Zhang F, Sun F, Luo Q, Wang R, Hu R, Chen M, Chang J, Yang G, He G (2017). A wheat MYB transcriptional repressor TaMyb1D regulates phenylpropanoid metabolism and enhances tolerance to drought and oxidative stresses in transgenic tobacco plants. Plant Science 265:112-123. https://doi.orgg/10.1016/j.plantsci.2017.09.020
- Wen J, Jiang F, Weng Y, Sun M, Shi X, Zhou Y, Yu L, Wu Z (2019). Identification of heat-tolerance QTL and hightemperature stress-responsive genes through conventional QTL mapping, QTL-seq and RNA-seq in tomato. BMC Plant Biology 19:398. https://doi.org/10.1186/s12870-019-2008-3
- Xie F, Wang Q, Sun R, Zhang B (2015). Deep sequencing reveals important roles of microRNAs in response to drought and salinity stress in cotton. Journal of Experimental Botany 66:789-804. *https://doi.org/10.1093/jxb/eru437*
- Xing C, Liu Y, Zhao L, Zhang S, Huang X (2019). A novel MYB transcription factor regulates ascorbic acid synthesis and affects cold tolerance. Plant, Cell & Environment 42:832-845. *https://doi.org/10.1111/pce.13387*
- Yang A, Dai X, Zhang W-H (2012). A R2R3-type MYB gene, OsMYB2 is involved in salt, cold, and dehydration tolerance in rice. Journal of Experimental Botany 63:2541-2556. *https://doi.org/10.1093/jxb/err431*
- You J, Chan Z (2015). ROS regulation during abiotic stress responses in crop plants. Frontiers in Plant Science 6:1092. https://doi.org/10.3389/fpls.2015.01092
- Young ND (1996). QTL mapping and quantitative disease resistance in plants. Annual Review of Phytopathology 34: 479-501. https://doi.org/10.1146/annurev.phyto.34.1.479
- Yu QH, Wang B, Li N, Tang Y, Yang S, Yang T, Xu J, Guo C, Yan P, Wang Q, Asmutola P (2017). CRISPR/Cas9induced targeted mutagenesis and gene replacement to generate long-shelf-life tomato lines. Scientific Reports 7:11874. https://doi.org/10.1038/s41598-017-12262-1
- Yu Y, Wang L, Chen J, Liu Z, Park CM, Xiang F (2018). WRKY71 acts antagonistically against salt-delayed flowering in Arabidopsis thaliana. Plant & Cell Physiology 59: 414-422. https://doi.org/10.1093/pcp/pcx201
- Yu YT, Wu Z, Lu K, Bi C, Liang S, Wang XF, Zhang DP (2016). Overexpression of the MYB37 transcription factor enhances abscisic acid sensitivity, and improves both drought tolerance and seed productivity in Arabidopsis thaliana. Plant Molecular Biology 90:267-279. https://doi.org/10.1007/s11103-015-0411-1
- Zhang B (2015). MicroRNA: A new target for improving plant tolerance to abiotic stress. Journal of Experimental Botany 66:1749-1761. https://doi.org/10.1093/jxb/erv013
- Zhang X, Wang W, Wang M, Zhang HY, Liu JH (2016). The miR396b of Poncirustrifoliata functions in cold tolerance by regulating ACC oxidase gene expression and modulating ethylene–polyamine homeostasis. Plant & Cell Physiology 57:1865-1878. https://doi.org/10.1093/pcp/pcw108
- Zhang X, Zou Z, Gong P, Zhang J, Ziaf K, Li H, Xiao F, Ye Z (2011). Over-expression of microRNA169confers enhanced drought tolerance to tomato. Biotechnology Letters 33:403-409. *https://doi.org/10.1007/s10529-010-0436-0*
- Zhang Y, Deng G, Fan W, Yuan L, Wang H, Zhang P (2019). NHX1 and eIF4A1-stacked transgenic sweet potato shows enhanced tolerance to drought stress. Plant Cell Reports 38(11):1427-1438. https://doi.org/10.1007/s00299-019-02454-6
- Zhao J, Yuan S, Zhou M, Yuan N, Li Z, Hu Q, Bethea FG, Liu H, Li S, Luo H (2018). Transgenic creeping bentgrass overexpressing OsamiR393aexhibits altered plant development and improved multiple stress tolerance. Plant Biotechnology Journal 17:233-251. https://doi.org/10.1111/pbi.12960
- Zhao Y, Tian X, Wang F, Zhang L, Xin M, Hu Z, Yao Y, Ni Z, Sun Q, Peng H (2017). Characterization of wheat MYB genes responsive to high temperatures. BMC Plant Biology 17:208. *https://doi.org/10.1186/s12870-017-1158-4*
- Zhao Y, Yang Z, Ding Y, Liu L, Han X, Zhan J, Wei X, Diao Y, Qin W, Wang P, Liu P (2019). Over-expression of an R2R3 MYB Gene, GhMYB73, increases tolerance to salt stress in transgenic Arabidopsis. Plant Science 286:28-36. https://doi.org/10.1016/j.plantsci.2019.05.021



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