

THE UNIVERSITY of EDINBURGH

Edinburgh Research Explorer

Expansion of the neurodevelopmental phenotype of individuals with EEF1A2 variants and genotype-phenotype study

Citation for published version:

Paulet, A, Bennett-Ness, C, Ageorges, F, Trost, D, Green, A, Goudie, D, Jewell, R, Kraatari-Tiri, M, Piard, J, Coubes, C, Lam, W, Lynch, SA, Samuel, G, Ramond, F, Fluss, J, Fagerberg, C, Brasch Andersen, C, Varvagiannis, K, Kleefstra, T, Gérard, B, Fradin, M, Vitobello, A, Tenconi, R, Denommé-Pichon, A-S, Vincent-Devulder, A, Haack, T, Marsh, JA, Laulund, LW, Grimmel, M, Riess, A, de Boer, E, Padilla-Lopez, S, Bakhtiari, S, Kruer, MC, Levy, J, Verloes, A, Abbott, CM & Ruaud, L 2024, 'Expansion of the neurodevelopmental phenotype of individuals with EEF1A2 variants and genotype-phenotype study', *European Journal of Human Comptices*, https://doi.org/10.1038/s41431-024-01560-8 European Journal of Human Genetics. https://doi.org/10.1038/s41431-024-01560-8

Digital Object Identifier (DOI):

10.1038/s41431-024-01560-8

Link:

Link to publication record in Edinburgh Research Explorer

Document Version: Peer reviewed version

Published In: European Journal of Human Genetics

Publisher Rights Statement:

Open Access. This article is licensed under a Creative Commons Attribution 4.0 International License (CC BY 4.0), which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http:// creat iveco mmons. org/ licen ses/ by/4. 0/.

General rights

Copyright for the publications made accessible via the Edinburgh Research Explorer is retained by the author(s) and / or other copyright owners and it is a condition of accessing these publications that users recognise and abide by the legal requirements associated with these rights.

Take down policy The University of Edinburgh has made every reasonable effort to ensure that Edinburgh Research Explorer content complies with UK legislation. If you believe that the public display of this file breaches copyright please contact openaccess@ed.ac.uk providing details, and we will remove access to the work immediately and investigate your claim.



Expansion of the neurodevelopmental phenotype of
 individuals with *EEF1A2* variants and genotype-phenotype
 study.

Alix Paulet¹, Cavan Benett-Ness², Faustine Ageorges¹, Detlef Trost³, 4 5 Andrew Green⁴, David Goudie⁵, Rosalyn Jewell⁶, Minna Kraatari-Tiri^{7,8}, Juliette PIARD⁹, Christine Coubes¹⁰, Wayne Lam¹¹, Sally Ann 6 Lynch¹², Groeschel Samuel¹³, Francis Ramond¹⁴, Joël Fluss¹⁵, Christina 7 8 Fagerberg¹⁶, Charlotte Brasch-Andersen¹⁷, Konstantinos Varvagiannis¹⁸, Tjitske Kleefstra^{19,20,21,22}, Bénédicte Gérard²³, Mélanie 9 10 Fradin²⁴, Antonio Vitobello²⁵, Romano Tenconi²⁶, Anne-Sophie Denommé-Pichon^{27,28}, Aline Vincent²⁹, Tobias Haack³⁰, Joseph 11 Marsh³¹, Lone Walentin Laulund³², Mona Grimmel³⁰, Angelika Riess³⁰, 12 Elke de Boer^{19,20,21}, Sergio Padilla Lopez³³, Somayeh Bakhtiari³³, 13 Michael Kruer³³, Jonathan Levy¹, Alain Verloes¹, Catherine Abbott², 14 15 Lyse Ruaud¹.

16

17 Correspondence to: Alix Paulet, Département de Génétique, Hôpital18 Robert-Debré, Paris, France.

19 Email: alix.paulet@hotmail.fr

20

21 1. Département de Génétique, Hôpital Robert-Debré, Paris, France.

22 2. Centre for Genomic and Experimental Medicine and Simons

23 Initiative for the Developing Brain, Institute of Genetics and Cancer,

24 Edinburgh, Scotland, UK.

25 3. Laboratoire Cerba, Saint-Ouen l'Aumône, France.

- 26 4. UCD School of Medicine and Medical Science Consultant in Clinical
- 27 Genetics, Dublin, Ireland.
- 28 5. Regional Genetics Service, NHS Tayside, Dundee, Scotland, UK.
- 29 6. Yorkshire Regional Genetics Service, Leeds Teaching Hospitals NHS
- 30 Trust, Leeds, England, UK.
- 31 7. Department of Clinical Genetics, Research unit of Clinical Medicine,
- 32 Medical Research Center Oulu, Finland.
- 33 8. Oulu University Hospital and University of Oulu, Finland.
- 34 9. Centre de Génétique Humaine, CHU Besançon, Besançon, France.
- 35 10. Service de Génétique Médicale, CHU de Montpellier, Montpellier,
- 36 France.
- 37 11. South-East of Scotland Clinical Genetics Service, General Hospital,
- 38 Edinburgh, Scotland, UK.
- 39 12. Clinical Genetics, Children's Health Ireland, Dublin, Ireland.
- 40 13. Department of Neuropediatrics, University Children's Hospital,
- 41 Tuebingen, Germany.
- 42 14. Service de Génétique, CHU Saint-Etienne Hôpital Nord, Saint-
- 43 Etienne, France.
- 44 15. University Hospitals of Geneva, Geneva, Switzerland.
- 45 16. Department of Clinical Genetics, Odense University Hospital,
- 46 Odense, Denmark.
- 47 17. Human Genetik, Syddansk Universitet, Odense, Denmark.
- 48 18. Service de Médecine Génétique, Hôpitaux universitaires de
- 49 Genève, Geneva, Switzerland.
- 50 19. Department of Clinical Genetics, Erasmus MC University Medical
- 51 Center, Rotterdam, The Netherlands.
 - 2

- 52 20. Department of Human Genetics, Radboud University Medical
- 53 Center, Nijmegen, The Netherlands.
- 54 21. Donders Institute for Brain, Cognition and Behaviour, Radboud
- 55 University, Nijmegen, the Netherlands.
- 56 22. Center of Excellence for Neuropsychiatry, Vincent van Gogh
- 57 Institute for Psychiatry, Venray, The Netherlands.
- 58 23. Molecular Biology, Nouvel Hôpital Civil, Strasbourg, France.
- 59 24. Service de Génétique Médicale, Hôpital Sud, CHU de Rennes.
- 60 25. UMR-Inserm, Génétique des Anomalies du développement,
- 61 Université de Bourgogne Franche-Comté, Dijon.
- 62 26. Servizio di Genetica Medica, Dipartimento di Pediatra, Padova,
- 63 Italia.
- 64 27. Unité Fonctionnelle Innovation en Diagnostic génomique des
- 65 maladies rares, FHU-TRANSLAD, CHU Dijon Bourgogne, Dijon, France.
- 66 28. UMR1231 GAD, Inserm Université Bourgogne-Franche Comté,
- 67 Dijon, France.
- 68 29. Génétique Médicale, CHU de Caen, Caen France.
- 69 30. Institute of Medical Genetics and Applied Genomics, University of
- 70 Tübingen, Tübingen, Germany.
- 71 31. MRC Human Genetics Unit, Western General Hospital, University
- 72 of Edinburgh, Edinburgh, Scotland, UK.
- 73 32. H C Andersen Children's Hospital, Odense University Hospital,
- 74 Odense, Denmark.
- 75 33. Pediatric Movement Disorders Program, Division of Pediatric
 76 Neurology, Barrow Neurological Institute, Phoenix Children's
 77 Hospital, Phoenix, Arizona, USA.
 - 3

78 Keywords: *EEF1A2*, Intellectual disability, regression, epilepsy,
79 neurodevelopment.

80 Abstract

81 Translation elongation factor eEF1A2 constitutes the alpha subunit of 82 the elongation factor-1 complex, responsible for the enzymatic 83 binding of aminoacyl-tRNA to the ribosome. Since 2012, 21 84 pathogenic missense variants affecting EEF1A2 have been described in 42 individuals with a severe neurodevelopmental phenotype 85 86 including epileptic encephalopathy and moderate to profound 87 intellectual disability (ID), with neurological regression in some 88 patients.

89 Through international collaborative call, we collected 26 patients90 with *EEF1A2* variants and compared them to the literature.

91 Our cohort shows a significantly milder phenotype. 83% of the 92 patients are walking (vs. 29% in the literature), and 84% of the 93 patients have language skills (vs. 15%). Three of our patients do not 94 have ID. Epilepsy is present in 63% (vs. 93%). Neurological 95 examination shows a less severe phenotype with significantly less 96 hypotonia (58% vs. 96%), and pyramidal signs (24% vs. 68%). 97 Cognitive regression was noted in 4% (vs. 56% in the literature).

98 Among individuals over 10 years, 56% disclosed neurocognitive99 regression, with a mean age of onset at 2 years.

100 We describe 8 novel missense variants of EEF1A2. Modelling of the 101 different amino-acid sites shows that the variants associated with a 102 severe phenotype, and the majority of those associated with a 103 moderate phenotype, cluster within the switch II region of the protein 104 and thus may affect GTP exchange. In contrast, variants associated 105 with milder phenotypes may impact secondary functions such as 106 actin bundling. We report the largest cohort of individuals with 107 EEF1A2 variants thus far, allowing us to expand the phenotype 108 spectrum and reveal genotype-phenotype correlations.

109 Abbreviations:

- 110 ACMG: American College of Medical Genetics
- 111 ADHD: Attention Deficit and Hyperactivity Disorder
- 112 ASD: Autism Spectrum Disorder
- 113 CADD: Combined Annotation Dependent Depletion
- 114 CLB: Clobazam
- 115 FISQ : Full-Scale Intelligence Quotient
- 116 ID: Intellectual Disability
- 117 IQ: Intellectual Quotient
- 118 KETO DIET: Ketogenic Diet
- 119 LAM: Lamotrigin
- 120 LORAZEP: Lorazepam
- 121 MICROPAK: Micropakine
- 122 MRI: Magnetic Resonance Imaging
- 123 OFC: Occipito-Frontal Circumference
- 124 SD: Standard Deviation
- 125 TIQ: Total Intellectual Quotient
- 126 VA: Valproic Acid
- 127 WISC: Wechsler Intelligence Scale for Children

- 128 WM and GM: White Matter And Grey Matter
- 129 WPPSI-IV: Wechsler Preschool and Primary Scale of Intelligence

130 Introduction

131 The eEF1 family of eukaryotic elongation factor genes, which 132 comprises the two paralog eEF1A proteins eEF1A1 and eEF1A2 and 133 the 3 subunits of the eEF1B complex (eEF1B α , eEF1B β , and eEF1B γ) 134 encodes integral components of the translation elongation factor 135 complexes whose function is delivery of aminoacyl tRNA to ribosome 136 during the elongation step of protein synthesis¹. The two eEF1A 137 proteins, the second most abundant protein in the cell², share 92% 138 identity. eEF1A binds aa-tRNAs in a GTP-dependent manner, relying 139 on its cognate guanine exchange factor (GEF), eEF1B, to recycle the 140 inactive eEF1A-GDP complex into the active GTP-bound form. EEF1A1 141 gene is expressed almost ubiquitously. EEF1A2 is expressed mainly in muscle (including cardiac muscle) and in neurons^{3,4}. During 142 143 development, eEF1A1 is down-regulated in muscle and neurons and 144 is undetectable in mouse muscle by 3 weeks post-natal^{5,6}.

145 No pathogenic *EEF1A1* variants have been described, presumably 146 because they would not be compatible with life⁷. In contrast, in 1972, 147 a deletion of 15.8 kb that abolishes expression of eEF1A2 (MIM 148 01958) was discovered in mice that developed motor neuron degeneration, muscle atrophy, gait abnormalities and then died by 149 four weeks^{3,5}. A trio-based exome in 2012 revealed a *EEF1A2* variant 150 151 in a patient with early onset epilepsy, severely delayed psychomotor development, and autistic behavior⁸. Subsequent individual reports 152 and a series of 14 patients⁹ allowed to further delineate the 153 154 neurodevelopmental phenotype, combining moderate to severe

155	Intellectual Disability (ID), epilepsy, Autism Spectrum Disorder (ASD),					
156	and sleep disorders neurodegeneration and movement disorders ⁹⁻¹³ .					
157	Since 2012, 21 <i>EEF1A2</i> variants have been reported ⁸⁻²⁷ . All variants					
158	are missense. In one pedigree, a variant was inherited from a parent					
159	with less than 25% of mosaicism ⁹ . Other variants were <i>de novo</i> . To					
160	date, no genotype-phenotype correlation has been studied.					
161	Through an international collaboration, we collected data on 26					
162	unreported EEF1A2 patients, the largest cohort to date, and					

163 investigated possible genotype-phenotype correlations.

164 **Material And Methods:**

165 Individual ascertainment:

Between December 2020 and December 2021, 26 individuals with variants in *EEF1A2* were identified through European Reference Network ERN-ITHACA (https://ern-ithaca.eu/) using its collaborative call system. Patients were evaluated by a geneticist. Written informed consents for DNA and data analyses were obtained from individuals or their legal guardians.

172 Cognitive assessment:

Out of the 26 patients included in our study, we assessed cognitive abilities for 13 individuals (Table 1): six underwent a Wechsler Intelligence Scales for Children according to age (Scales IV and V: 6 years to 16 years 11 months old, or Wechsler Preschool and Primary Scale of Intelligence III or IV: 4 years to 7 years 3 months old) and 7 patients were evaluated by clinicians (Table S1).

Among 6 patients which performed Wechsler tests, only 4 of the 6 patients had a computable Total Intelligence Quotient (TIQ). For the remaining two, they had too heterogenous profiles to allow TIQ calculation. We therefore utilized the available indices data (VCI, FRI, VSI, WMI, PSI) to estimate their intellectual levels.

For the 7 patients evaluated by clinicians, cognitive assessments wereprovided but we did not have quantitative data (Table S1).

Patients were divided into 4 categories such as "not ID" for TIQ \geq 70, "mild ID" for 50 \leq IQ<70, "moderate ID" for 35 \leq IQ<50, and "severe ID" for IQ<35, according to their IQ scores especially TIQ. In total, we classified 13 patients according to their level of intellectual efficiency (Table 1).

We excluded 13 individuals of cognitive analysis because they were
too young at the last follow-up appointment or they have been lost
to follow-up.

194 To categorize individuals from the literature when Weschler's FSIQ 195 was not available, we took into account the authors' assigned 196 category ID for each individual or we used the information on 197 individual developmental milestones (walking age, capacity for 198 language, verbal abilities...) to estimate the developmental delay as 199 "mild", "moderate" or "severe" phenotype (Table S2). The individuals 200 whose information was insufficient to be to be classified into these 201 categories were not scored.

202 Genetic investigations:

203 DNA was extracted from the peripheral blood leukocytes of the204 patients and parents (whenever possible) using standard procedures.

205 In 24/26 patients, genotyping was performed by exome sequencing 206 (ES) (single or trio) using routine methods. Confirmation and 207 segregation of variants in single ES were carried out by Sanger 208 sequencing. Variant prioritization was conducted according to the 209 transmission mode (de novo, autosomal recessive and X-linked), and 210 the frequency of the variants in the gnomAD database. Those variants 211 were classified according to American College of Medical Genetics 212 (ACMG) (ACMG and Combined Annotation Dependent Depletion 213 (CADD) score in Supplementary data, Table S3). There were no other 214 pathogenic variants in ClinVar and HGMD or loss-of-function variants 215 which could explain the patient phenotypes. Two variants were 216 identified on a NGS panel of 119 ID genes using standard NGS 217 procedures.

218 **Protein Modelling:**

The structures of eEF1A2*GDP (4COS), eEF1Ay*eEF1Bα (pdb: 1IJF),
aEF1A*GTP (pdb: 3AGJ), and EF-TU*EF-TS (pdb:1EFU) were used for
analysis using Pymol (The PyMOL Molecular Graphics System, Version
1.2r3pre, Schrödinger, LLC.) and CCP4MG²⁸. Structure-based variant
predictions made using Missense 3D²⁹ and dimer predictions using
FoldX³⁰. Briefly, eEF1A2 (pdb: 4COS) was repaired, then the protein

stability was estimated for each amino acid change in bothmonomeric and the dimeric eEF1A2.

227 Statistical analysis

- 228 We compared our patients to the previous reported ones. We used
- 229 Student Test with bilateral hypothesis, have tolerated a risk of error
- of less than 5% (*p-value* < 0.05).

231 Literature review:

PubMed was searched for peer-reviewed articles published in English
using the following keywords: « *EEF1A2* », « epileptic
encephalopathy gene », « *EEF1A* » «phenotype of *EEF1A2* » « *EEF1A2*with no intellectual disability » « epilepsy gene ».

236 **Results**

In our series, the mean age at last examination is 10.67 years vs. 9.6
years for literature patients (Table S4). The mean Occipito-frontal
circumference (OFC) is -0.67 SD (vs. -1.33 SD) (Table S4). Most
patients can walk on independently. The mean age for acquiring
walking is 30 months. For the 4 individuals in the literature this mean
is 39 months, *p*-value= 0.41 (Table S4).

Among 18 individuals aged more than 2 years, 83% are verbal (Figure 1) with a mean age of first words at 29 months (Table S4) (data not available for published cases). The average age at onset of epilepsy is 3.5 years (Table S4) (data not available for previously reported patients).

Neuropsychologist assessments were performed in six patients
(Table 1). For the 4 patients for whom a TIQ could be calculated, the
Full-Scale Intelligence Quotient (FSIQ) ranged from 40 to 77 (mean=
60.2; median= 58.5) (Table S1).

Patient 1 has a FSIQ in the low normal range (77) and the two others
(patients 6 and 19) had several indexes > 70 despite their
heterogeneous profile did not allow to compute FSIQ. Patient 19 has
a FSIQ of 69, but with low normal verbal comprehension index (76)
and fluid reasoning (71) (Table S1).

In our cohort of 13 for whom we have developmental information, 4patients have a phenotype described as severe (31%) compared to

those from the literature (76%) *p-value*=0.008 (-0.77;-0.13) (Table
1). Among published patients, 5/33 (15%) had profound ID (Table 2),
whereas there was no patient classified as having profound ID in our
cohort. There was no difference observed concerning mild and
moderate ID (Table 1).

The individuals of this series are more ambulant than previous
reported ones: 82% (18/22) vs. 29% (10/34), *p-value=*0.00004 (0.29;
0.76). They are more likely to have acquired language (83% vs. 14% *p-value=*10E-7 (0.47;0.91)) (Figure 1).

268 In our cohort, less patients had hypotonia and pyramidal syndrome 269 (Figure 1). There was no difference in the incidence of ataxia and ASD. 270 A majority of our patients have sleep disorders (11/24), with limited 271 effectiveness of melatonin treatment (Table S1). Half of our patients 272 (53%-9/17) had ADHD (Table S1). More than a half of our patients 273 (14/25) have gross motor issues with mainly unstable walk, and 77% 274 (17/22) have fine motor issues particularly concerning coordination 275 (Table S1).

We also compared our patients to the previously described ones regarding several items. The percentage of patients with epilepsy refractory to therapy, the percentage with epileptic encephalopathy were compared so as our series brain MRI features. The results are presented in Table 2.

We report 8 new *EEF1A2* variants (T24M, R96H, D97N, T104R, G356S,
D362N, P420L, V437F) with detailed phenotyping. Phenotypes

283	observed varies from mild to severe (Figure 2a). The patients with
284	variants T24M and R96C have no ID but are symptomatic: have
285	epilepsy (R96C) or speech delay with firsts sentences at age 6 (T24M).
286	For patients with P420L and V437F we could not document ID
287	because of the lack of psychometric evaluation (Table S1).

288 Molecular Modelling

We explored the impact of the variants through 3D-modelling of
eEF1A2, comparing the GDP-bound form of eEF1A2 (pdb:4COS) with
eEF1Ay*eEF1Bα (pdb:1IJF), aEF1A*GTP (pdb:3AGJ), and EF-TU*EF-TS
(pdb:1EFU). All variants have CADD scores above 20 (Table S3).

293 Mapping of the variants onto the structure of eEF1A2 (Figure 2b) 294 showed that several of the variants cluster around the switch II 295 region, a flexible motif key for GTP hydrolysis and GEF-mediated GDP 296 dissociation. R96C, R96H, and D97N are found in a loop in the Switch 297 II region, whilst T104R, R381W, and V437F are situated nearby. These 298 variations are anticipated to contribute towards switch II 299 destabilization and interfere with GTP recycling, whilst another 300 variant, T24M, is located at the GTP-binding pocket, and is predicted 301 to directly disrupt nucleotide binding (Supplementary modelling in 302 Supplemental data).

303 Further from the GDP-binding or switch regions, several variations 304 (E297K, G356S, D362N, and P420L) are located near the actin binding 305 region (Figure 2b) and are anticipated to disrupt actin-related 306 functions (Supplementary modelling). Three of these variants, E297K, 307 D362N and P420L are located near the tRNA-binding site, and may 308 affect eEF1A2 interactions with aminoacyl-tRNA. Given the overlap 309 between the binding sites for actin and aa-tRNAs and the eEF1B-310 binding region, some of these variants may also impact eEF1B 311 binding. Additionally, the presence of variants T104R and D362N on 312 the dimer interface, and the change in amino acid charge, suggest

- 313 they may influence eEF1A2 dimer formation (Supplementary
- 314 modelling).

315 **Discussion**

316 Widening *EEF1A2*-related spectrum

Our results show that patients with *EEF1A2* variants can have efficient cognitive abilities. Indeed, in our cohort, 3 individuals do not have an intellectual disability (IQ > 70), while in the literature, no patients are reported with preserved intellectual efficiency. In addition, within our cohort, the severity of ID is lower, with a majority of patients with mild to moderate ID.

In terms of development, the majority of our individuals have access
to language and walking, contrary to what has already been
described.

326 Taken together, the phenotypic spectrum associated with EEF1A2 variants is wider than previously suggested. All previously reported 327 328 patients had intellectual disabilities and the majority (93%) were 329 epileptic and nonverbal. We have shown that individuals with EEF1A2 330 variants may have milder phenotypes and that nearly half of them do 331 not have epilepsy at time of report. Epilepsy can be late-onset, as 332 illustrated by patient 7 (Ref. 20) with the D252H variant who did not 333 develop epilepsy until age 8 and the average age in the cohort was 334 10.7 years.

Young individuals are difficult to assess using the WPPSI-IV or WISC scales. There are 13 individuals who have received cognitive evaluation but only 4/13 have a valid TIQ because profiles are too heterogenous. For this study, we chose to take the heterogenous

profiles into account because even if partial Weschler 'scales remainan objective way to evaluate individuals' cognitive capacities.

341 To date, the only reported patient of autosomal dominant inherited 342 *EEF1A2* variant was from a mother with <25% of mosaicism⁹ but there 343 was no clinical information available. We newly report 2 patients 344 (patient 6 (whose father is patient 7) and patient 20) with inherited 345 variants asymptomatic from an parent (in terms of 346 neurodevelopmental disorder) which suggests an incomplete 347 penetrance of these variants (G356S and R96C). G356S is not 348 reported in the database and the position of the residue is highly 349 conserved. For G356S, the asymptomatic parent has a history of 350 irregular heartbeat, and he has at least 3 relatives who had seizures 351 (the relatives have not been tested for the variant yet). One individual 352 (patient 6) has the variant R96C inherited from his affected father 353 (patient 7), this variant had been shown to be associated with (Genetic Generalized epilepsies) GGE²⁷. No mosaicism detected in 354 355 unaffected parents.

These two examples of inherited *EEF1A2* variants strongly suggest incomplete penetrance. Although only 3 patients (including our 2) are reported so far, incomplete penetrance should be considered in *EEF1A2* variants. More individuals are needed to confirm this hypothesis and we recommend to test proband's parents even if there are apparently not affected. This incomplete penetrance is relevant to genetic counselling.

We can notice a fairly variability of expression in patients with the same variation. For example, in our series, patients 15, 16 and 17 have the same E124K variant already reported by Kaur et al¹⁵. Whereas patients 15,16 and Kaur et al patient acquired independent walk before 2 years with good language abilities and epilepsy, our patient 17 acquired walk at 4 years and had no epilepsy.

369 We also report 2 patients (11 and 5) of de novo EEF1A2 variant 370 mosaicism which did not cause milder phenotypes. Patient 11 371 presents epileptic encephalopathy and has TIQ of 52 at 3y. Patient 5 372 can speak in sentences. Nevertheless, they have similarities: they 373 both have white and grey matter abnormalities on brain MRI and 374 focal epilepsy, controlled with antiepileptic treatment. Concerning 375 patient 5, the variation D91N has been reported three times in the 376 Literature in patients with severe to profound ID and early onset 377 epilepsy^{9,15,18}, whereas our patient is obviously less severe affected 378 than the three others. This mosaicism (estimated to 23%) could have 379 caused bias in comparison because this milder phenotype is possibly 380 only a result of mosaicism. So, mosaicism state should be taken into 381 account while considering an EFF1A2 variant, although the 382 impossibility to estimate percentage of mosaicism in brain makes the 383 phenotype difficult to be predicted.

Ten patients^{9-10,12,22-23} have been reported to have regression in childhood. In our cohort, only patient 22 (R381W) has shown regressive traits which began in her third decade (but she is also the only individual to have reached this age). She has started to show less

interest in activities she used to like such as writing, swimming and she lost some abilities. This individual suggest that regression may appear later than we thought while reviewing the literature patients (where most of patients showed regression in infancy⁹). It's clear that patients must be followed up and supported to prevent complications during time.

394 Molecular insights

Plotting the patients' variants and those previously described on the 395 396 surface of the eEF1A2 protein (Figure 2b) suggests that variants, 397 resulting in severe ID are generally clustered around the switch I and 398 switch II regions, or nucleotide/GEF binding sites for GTP and GTP 399 exchange factor eEF1B. The growing cluster of variants around the 400 switch II region or GDP-binding site (Figure 2b) suggests that 401 disruption of GDP-binding and GEF-induced GDP dissociation may be 402 a key mechanism for the NDD-causing EEF1A2 missense variants, 403 adding to the evidence from Carvill et al⁹. Four of the 19 variants 404 classified as severe directly coincide with defined eEF1B binding sites (Figure 2b). 405

406 Many of the milder variants map further away but could affect tRNA 407 or actin binding (Figure 2b). There is evidence that E295K, the 408 equivalent E297K mutation in yeast, affects translational fidelity. 409 Variants affecting translational fidelity might be less likely to affect 410 neurodevelopment, but would be anticipated to lead to 411 neurodegeneration, which is observed in a subset of individuals with 412 EEF1A2 variants. There is also the possibility that these variants are 413 not pathogenic and the cause of the individual phenotype was 414 misunderstood.

415 **Conclusion**

416 We have described a cohort of individuals with a less severe 417 phenotype, though they share characteristics such as developmental 418 delay (especially speech delay), mild to severe intellectual disability, 419 ASD, ADHD, early onset epilepsy, hypotonia, ataxia and sleep disorder 420 which are concordant with the features related to variants in this 421 gene in the literature. We suggest the existence of incomplete 422 penetrance of certain variants which was not described so far with 423 EEF1A2. Our series illustrate how the evolution of diagnostic 424 strategies may lead to redefine the phenotypic spectrum of known 425 genes that have been initially reported with a homogeneous and 426 usually "severe" phenotype. There was probably an ascertainment 427 bias in older patients that were more likely to be reported because 428 they were severe, and were perhaps selected on this basis. The 429 widespread adoption of the whole genome and whole exome 430 sequencing, which results in an agnostic pan-genomic evaluation lead 431 to the diagnosis of patients that would not be sent to targeted panel 432 diagnosis. The first genotype-phenotype correlations are emerging, 433 but new patients will be necessary to confirm these correlations.

To conclude, we expanded the phenotype spectrum and describednew *EEF1A2* variants.

- 436 All anonymized data and related documentation from this study are
- 437 available on reasonable request.
- 438 Declaration to public database: All variants are reported and
- 439 annotated in ClinVar website (accession number SCV004171535-
- 440 SCV004171551): https://www.ncbi.nlm.nih.gov/clinvar/
- 441 **Bibliographie**
- Kahns S, Lund A, Kristensen P, et al. The elongation factor 1 A-2
 isoform from rabbit: cloning of the cDNA and characterization of
 the protein. *Nucleic Acids Res.* 1998;26(8):1884-1890.
 doi:10.1093/nar/26.8.1884
- Abbott CM, Newbery HJ, Squires CE, Brownstein D, Griffiths LA,
 Soares DC. eEF1A2 and neuronal degeneration. *Biochem Soc Trans*. 2009;37(Pt 6):1293-1297. doi:10.1042/BST0371293
- 3. Newbery HJ, Loh DH, O'Donoghue JE, et al. Translation
 elongation factor eEF1A2 is essential for post-weaning survival in
 mice. *J Biol Chem*. 2007;282(39):28951-28959.
 doi:10.1074/jbc.M703962200
- Knudsen SM, Frydenberg J, Clark BF, Leffers H. Tissue-dependent
 variation in the expression of elongation factor-1 alpha isoforms:
 isolation and characterisation of a cDNA encoding a novel variant
 of human elongation-factor 1 alpha. *Eur J Biochem*.
 1993;215(3):549-554. doi:10.1111/j.1432-1033.1993.tb18064.x
- 458 5. Chambers DM, Peters J, Abbott CM. The lethal mutation of the459 mouse wasted (wst) is a deletion that abolishes expression of a
 - 25

tissue-specific isoform of translation elongation factor 1alpha,
encoded by the Eef1a2 gene. Proc Natl Acad Sci U S A. 1998 Apr
14;95(8):4463-8. doi: 10.1073/pnas.95.8.4463. PMID: 9539760;
PMCID: PMC22512.

- Khalyfa A, Carlson BM, Dedkov EI, Wang E. Changes in protein
 levels of elongation factors, eEF1A-1 and eEF1A-2/S1, in longterm denervated rat muscle. Restor Neurol Neurosci. 2003;21(12):47-53. PMID: 12808202.
- 468 7. McLachlan F, Sires AM, Abbott CM. The role of translation
 469 elongation factor eEF1 subunits in neurodevelopmental
 470 disorders. *Hum Mutat*. 2019;40(2):131-141.
 471 doi:10.1002/humu.23677
- De Ligt J, Willemsen MH, van Bon BWM, et al. Diagnostic exome
 sequencing in persons with severe intellectual disability. *N Engl J Med.* 2012;367(20):1921-1929. doi:10.1056/NEJMoa1206524
- Carvill GL, Helbig KL, Myers CT, et al. Damaging de novo missense
 variants in EEF1A2 lead to a developmental and degenerative
 epileptic-dyskinetic encephalopathy. *Hum Mutat*.
 2020;41(7):1263-1279. doi:10.1002/humu.24015
- 479 10. Veeramah KR, Johnstone L, Karafet TM, et al. Exome sequencing
 480 reveals new causal mutations in children with epileptic
 481 encephalopathies. *Epilepsia*. 2013;54(7):1270-1281.
 482 doi:10.1111/epi.12201

- 483 11. Long K, Wang H, Song Z, Yin X, Wang Y. EEF1A2 mutations in
 484 epileptic encephalopathy/intellectual disability: Understanding
 485 the potential mechanism of phenotypic variation. *Epilepsy Behav*486 *EB*. 2020;105:106955. doi:10.1016/j.yebeh.2020.106955
- 12. Inui T, Kobayashi S, Ashikari Y, et al. Two cases of early-onset
 myoclonic seizures with continuous parietal delta activity caused
 by EEF1A2 mutations. *Brain Dev.* 2016;38(5):520-524.
 doi:10.1016/j.braindev.2015.11.003
- 491 13. Cao S, Smith LL, Padilla-Lopez SR, et al. Homozygous EEF1A2
- 492 mutation causes dilated cardiomyopathy, failure to thrive, global
 493 developmental delay, epilepsy and early death. *Hum Mol Genet*.
- 494 2017;26(18):3545-3552. doi:10.1093/hmg/ddx239
- 495 14. Kaneko M, Rosser T, Raca G. Dilated cardiomyopathy in a patient
 496 with autosomal dominant EEF1A2-related neurodevelopmental
 497 disorder. *Eur J Med Genet*. 2021;64(1):104121.
 498 doi:10.1016/j.ejmg.2020.104121
- 499 15. Kaur S, Van Bergen NJ, Gold WA, et al. Whole exome sequencing
 500 reveals a de novo missense variant in EEF1A2 in a Rett syndrome501 like patient. *Clin Case Rep.* 2019;7(12):2476-2482.
 502 doi:10.1002/ccr3.2511
- 503 16. Zacher P, Mayer T, Brandhoff F, et al. The genetic landscape of
 504 intellectual disability and epilepsy in adults and the elderly: a
 505 systematic genetic work-up of 150 individuals. *Genet Med Off J*

506 Am Coll Med Genet. 2021;23(8):1492-1497. doi:10.1038/s41436-

507 021-01153-6

- 508 17. Lopes F, Barbosa M, Ameur A, et al. Identification of novel genetic
- 509 causes of Rett syndrome-like phenotypes. J Med Genet.

510 2016;53(3):190-199. doi:10.1136/jmedgenet-2015-103568

- 18. Lam WWK, Millichap JJ, Soares DC, et al. Novel de novo EEF1A2
 missense mutations causing epilepsy and intellectual disability. *Mol Genet Genomic Med*. 2016;4(4):465-474.
 doi:10.1002/mgg3.219
- 515 19. de Kovel CGF, Brilstra EH, van Kempen MJA, et al. Targeted
 516 sequencing of 351 candidate genes for epileptic encephalopathy
 517 in a large cohort of patients. *Mol Genet Genomic Med*.
 518 2016;4(5):568-580. doi:10.1002/mgg3.235
- 20. Nakajima J, Okamoto N, Tohyama J, et al. De novo EEF1A2
 mutations in patients with characteristic facial features,
 intellectual disability, autistic behaviors and epilepsy. *Clin Genet*.
 2015;87(4):356-361. doi:10.1111/cge.12394
- 523 21. Ostrander BEP, Butterfield RJ, Pedersen BS, et al. Whole-genome
- 524 analysis for effective clinical diagnosis and gene discovery in early
- 525 infantile epileptic encephalopathy. *NPJ Genomic Med*. 2018;3:22.
- 526 doi:10.1038/s41525-018-0061-8
- 527 22. De Rinaldis M, Giorda R, Trabacca A. Mild epileptic phenotype
 528 associates with de novo eef1a2 mutation: Case report and
 - 28

529 review. Brain Dev. 2020;42(1):77-82.

530 doi:10.1016/j.braindev.2019.08.001

23. Lance EI, Kronenbuerger M, Cohen JS, Furmanski O, Singer HS,
Fatemi A. Successful treatment of choreo-athetotic movements
in a patient with an EEF1A2 gene variant. SAGE Open Med Case *Rep.* 2018;6:2050313X18807622.

- 535 doi:10.1177/2050313X18807622
- 536 24. O'Roak BJ, Stessman HA, Boyle EA, et al. Recurrent de novo
 537 mutations implicate novel genes underlying simplex autism risk.
 538 Nat Commun. 2014;5:5595. doi:10.1038/ncomms6595

539 25. Helbig KL, Farwell Hagman KD, Shinde DN, et al. Diagnostic
540 exome sequencing provides a molecular diagnosis for a
541 significant proportion of patients with epilepsy. *Genet Med.*542 2016;18(9):898-905. doi:10.1038/gim.2015.186

26. Vogt LM, Lorenzo M, B Prendergast D, Jobling R, Gill PJ. EEF1A2
pathogenic variant presenting in an infant with failure to thrive
and frequent apneas requiring respiratory support. Am J Med
Genet A. 2022 Oct;188(10):3106-3109. doi:
10.1002/ajmg.a.62932. Epub 2022 Aug 8. PMID: 35938194.

548 27. Epi25 Collaborative. Electronic address:
549 jm4279@cumc.columbia.edu; Epi25 Collaborative. Sub-genic
550 intolerance, ClinVar, and the epilepsies: A whole-exome
551 sequencing study of 29,165 individuals. Am J Hum Genet. 2021
552 Jun 3;108(6):965-982. doi: 10.1016/j.ajhg.2021.04.009. Epub

- 553 2021 Apr 30. Erratum in: Am J Hum Genet. 2021 Oct
 554 7;108(10):2024. PMID: 33932343; PMCID: PMC8206159.
- 28. Presenting your structures: the CCP4mg molecular-graphics
 software S. McNicholas, E. Potterton, K. S. Wilson and M. E. M.
 Noble Acta Cryst. (2011). D67, 386-394
- Ittisoponpisan, S., Islam, S.A., Khanna, T., Alhuzimi, E., David, A.
 & Sternberg, M.J.E. (2019)_Can Predicted Protein 3D Structures
 Provide Reliable Insights into whether Missense Variants Are
 Disease Associated? J. Mol. Biol. 431, 2197-
- 562 2212._https://doi.org/10.1016/j.jmb.2019.04.009
- 30. Van Durme J, Delgado J, Stricher F, Serrano L, Schymkowitz J and
 Rousseau F. A graphical interface for the FoldX forcefield.
 Bioinformatics. 2011 Jun 15;27(12):1711-2.

566 Figure Legends:

Table 1. Cognitive abilities of patients in our series compared to
those previous reported.

This table shows comparison in terms of cognitive abilities between our patients and the previously reported ones. The 4 categories classification strategy has been explained in the Material and Method section. The number of patients are in parentheses. * means the comparison is statistically significant, p-value <0.05. CI : Confidence Interval.

575 Table 2. Cohort and literature comparison regarding epilepsy and

576 brain MRI characteristics

577 This table shows comparison between our cohort and the previous 578 reported cases regarding epilepsy and brain MRI features. *P-value* 579 and CI 95% are mentioned. CI : Confidence Interval. * when 580 significant.

581 Figure 1. Our series main features compared with the patients 582 from the literature.

The figure 1 shows the proportion of our patients (in black) in comparison to those from the literature (featured in grey) according to the main criteria studied. Walking abilities is represented by the motor category. Concerning the speech abilities, "verbal" means the patient can speak and be understandable, "words" means he can express himself in words but not in sentences, "sentences" stands for the ability of the patient to make proper sentences. On the right

part of the diagram, the neurological features are shown withe the
presence of hypotonia, pyramidal syndrome, epilepsy and the
presence of regression. * Stands for statistically significance p<0.05. *P-values* are framed at the top.

594 **Figure 2. Distribution of pathogenic** *EEF1A2* **variants.**

595 Figure 2a: distribution of variants from our cohort (up) and the 596 previous reported ones (down). Novel variants are underlined. 597 Variants are classified by their associated phenotype in terms of ID, 598 classification is as described as above with associated symbols (circle 599 for not deficient, triangle for mild ID, square for moderate ID and star 600 for severe). Blank when the variant is not clearly classified (for 601 example when associated with 2 ID categories). Figure 2b: variants 602 mapped onto the crystal structure of GDP-bound eEF1A2 (PDB:4C0S). 603 New variants are shown in black, with labels in bold, previously 604 described mutations are in white. T24M and V437F are buried. The 605 binding site of eEF1B is highlighted in white, and the GTP binding site 606 in dark grey.

607 Table S1. Cohort's patients characteristics

The table S1 shows the characteristics of our 26 patients ordered by the position of their variation along the *EEF1A2* gene. Each column represents a criterion of interest. "M" means Male, "F" means Female. The ages at last examination are presented in years "y". The ages at first steps achievement without falling and first comprehensive words achievement are presented in months "m". Concerning the languages abilities at last examination, classification 615 into 3 categories (absent/words/sentences) are presented, with 616 details of language level. About the neuropsychological assessment 617 column, Total Intellectual Quotients (TIQ) are presented when they 618 can be calculated. In case the TIQ could not be calculated the different 619 QI indicators are presented with their respective scores: VCI, Verbal 620 Comprehension Index, VSI, Visual Spatial Index, FRI, Fluid Reasoning 621 index, WMI, Working Memory Index, PSI, Processing Speed Index. 622 The age of the tests were carried out are in parentheses ("NA" when 623 non available), followed by the type of test performed: WISC-V or 624 WPPSI-IV. For the therapy and efficacity column, the drugs used are in parentheses. Regarding the weight, height and Occipito-frontal 625 626 circumference (OFC) columns; the results are presented in standard 627 deviation (SD). The gross motor feature indicates walking disorder. The fine motor features entail coordination disorder. The proportion 628 629 of patients are in parentheses.

Y: Yes // N: No // NA: Non available // NC: Non concerned //ADHD:
Attention Deficit Hyperactivity Disorder // VA: Valproic Acid // LAM:
Lamotrigin // CLB: Clobazam // Lorazep: Lorazepam // Micropak:
micropakine // Keto diet: Ketogenic Diet // TIQ: Total Intellectual
Quotient.

635 Table S2. Literature patients characteristics

The table S2 shows the characteristics of the 42 patients reported in literature. Each column represents a criterion of interest. The ages at last examination are presented in years. "y" stands for "years", "m" for "months". "M" means Male, "F" means Female. "NA": Non

available. "Y" stands for yes, "N" stands for no. Regarding the
Occipito-frontal circumference (OFC) at examination column, the
results are presented in SD. The proportion of patients for each
category are in parentheses.

644 Table S3. ACMG classification and CADD Score

645 WES :Whole Exome sequencing; ACMG: American College of Medical 646 Genetics; CADD: Combined Annotation Dependent Depletion. This 647 table presents the classification according to ACMG guidelines and 648 the CADD score for each variant ordered by their position along the 649 EEF1A2 gene. The aim of ACMG classification is to divide the variants 650 into 5 categories, classes "1" and "2" are "benign classes". The class "3" is for "unknown significance variants", the class "4" represents 651 "probably pathogenic variants" and the 5th is for "pathogenic 652 653 variants". The CADD score evaluates the pathogenicity : if it is > 20, it 654 supports the pathogenicity of the variant. The "mosaic state" column 655 indicates if there is mosaicism for the variant and the associated 656 percentage found in the patient's blood sample in case of mosaicism.

Table S4. Our series and previously reported patients description

This table shows the number of patients and the mean age at last examination (in years), the mean age at first steps achievement without falling (in months), the mean age at onset of epilepsy and the meanoccipito-frontal circumference (OFC) in standard deviation (SD). The comparison, when possible, was not significant, *p-value* <0.05.

663 Supplementary modelling figures:

664 Figure S1. Variations in the switch II loop

665 Crystal structure of eEF1A2:GDP (4COS), with wild-type residues in
666 blue, and variant residues in red. Variants R96C, R96H, D97N, and
667 R381W highlighted in the switch II region.

668 Figure S2. R96H and R96C disrupt hydrogen bond network in

669 **eEF1Bα-bound form.**

670 Crystal structure of eEF1A *eEF1Bα (1IJF), with wild-type residues in
671 blue and mutant residues in red. R96 forms hydrogen bonds with
672 E132 and E135 of helix C in eEF1B-bound form. Both variants, R96C
673 and R96H, orient away from helix C, likely preventing hydrogen bond
674 formation and potentially destabilising disrupting eEF1Ba-mediated
675 release of GDP.

676 Figure S3. Variants impact eEF1A2 dimerisation.

Crystal structure of eEF1A2 homodimer (4COS), with chain A in blue
and chain A in gold. WT residues shown in blue, mutant residues in
red. T104 interacts with Thr71 from chain B on the dimer interface.
Modelling the T104R variant suggests that the bulky Arg residue will
clash with chain B and disrupt dimer formation. D362 forms a salt
bridge with K62 across the dimer interface, and the loss of charge in
the D362N variant suggests that this interaction will be lost.

684 **Figure S4. Variant at the nucleotide-binding site.**

- 685 Crystal structure of eEF1A2:GDP (4COS), showing the GDP-binding
- 686 site. The T24M variant is depicted, adjacent to the GDP-binding

- 687 residues. The Thr residue is buried within a helix, so the Met
- 688 substitution disrupts side-chain hydrogen bonds and sterically
- clashes. The variant likely destabilises the helix and displaces Lys20,
- 690 disrupting GDP-binding.

692 Acknowledgements:

- We want to acknowledge AnDDI-Rares and the families for their participation. In addition, the collaboration in this study were facilitated by ERN ITHACA, one of the 24 European Reference Networks (ERNs) approved by the ERN Board of Member States, cofounded by European Commission. [EU Framework Partnership Agreement ID: 3HP-HP-FPA ERN-01-2016/739516]
- 699 The aims of this study contribute to the Solve-RD project, which has
- 700 received funding from the European Union's Horizon 2020 research
- and innovation program under grant agreement number 779257.

702 Author Contributions:

- 703 Conceptualization: AP, LR, CA, AV.
- 704 Formal Analysis: AP.
- 705 Investigation: AP, FA, LR, MCK, SB, JL, DT, SB, MCK, AV, RT, CBA,
- 706 MG, BG, SP.
- 707 Supervision: LR, AV, CA.
- 708 Visualization: AP, CA, CBN.
- 709 Writing Original Draft: AP, LR, CA, CBN.

- 710 Writing Review & Editing: AP, LR, CA, CBN, AV, FM, CC, RF, JP, SJ,
- 711 CF, LW, JF, AV, ASDP, TH, JM, SAL, WL, RJ, MK, DG, AG, EDB, JL, DT,
- 712 SB, MCK, AV, RT, CBA, MG, BG, SP.

713 Conflicts of interest:

714 The authors declare no conflicts of interest.

715 Ethical Approval:

- Ethical approval was not required because it was a retrospective observational study and patients have already signed consent for genetics analyses for diagnosis. Every patient has been anonymized by the clinician before collecting data (a number has been assigned

to each patient).

721 Funding:

- 722 The authors received no financial support for the research,
- authorship, and/or publication of this article.

CognitiveCohort (13Literature (33Evaluationpatients)patients)		P-value	CI 95%	
NoID	23% (3)	0%	0.082	[-0.03 ;
NOID				0.49]
Mild	31% (4)	3% (1)	0.063	[-0.02 ;
wind				0.57]
Moderate	15% (2)	6% (2)	0.418	[-0.14 ;
Woderate				0.33]
Sovoro*	21% (1)	76% (25)	0.008	[-0.77 ;
Severe	51% (4)	70% (23)	0.008	-0.13]
Brofound*	0	15% (5)	0.023	[-0.28 ;
FIOIOUIIU				-0.02]

		Cohort	Literature	p-value	CI 95%
Enilongy	Epilepsy refractory to therapy	25% 3/12	50% 15/30	0.13	[-0.58 ; 0.08]
гриерзу	Epileptic encephalopathy	7% 1/15	24% 9/38	0.085	[-0.36 ; 0.02]
	Normal	50% 11/22	47% 15/32	0.8258	[-0.25 ; 0.31]
	Thin corpus callosum	14% 3/22	17% 5/30	0.767	[-0.23 ; 0.17]
	Delayed myelinization	9% 2/22	13% 4/30	0.636	[-0.22 ; 0.13]
Brain MRI	White and grey matter abnormalities	27% 6/22	9% 3/32	0.114	[-0.04 ; 0.40]
	Cerebellar and cortical atrophy*	9% 2/22	34% 11/32	0.0207*	[-0.46 ; -0.04]



Figure 1. Our series main features compared with the patients from the literature. The figure 1 shows the proportion of our patients (in black) in comparison to those from the literature (featured in grey) according to the main criteria studied. Walking abilities is represented by the motor category. Concerning the speech abilities, "verbal" means the patient can speak and be understandable, "words" means he can express himself in words but not in sentences, "sentences" stands for the ability of the patient to make proper sentences. On the right part of the diagram, the neurological features are shown withe the presence of hypotonia, pyramidal syndrome, epilepsy and the presence of regression. * Stands for statistically significance p<0.05. *P-values* are framed at the top.



2b

2a



Figure 2. Distribution of pathogenic *EEF1A2* **variants**. Figure 2a: distribution of variants from our cohort (up) and the previous reported ones (down). Novel variants are underlined. Variants are classified by their associated phenotype in terms of ID, classification is as described as above with associated

symbols (circle for not deficient, triangle for mild ID, square for moderate ID and star for severe). Blank when the variant is not clearly classified (for example when associated with 2 ID categories). Figure 2b: variants mapped onto the crystal structure of GDP-bound eEF1A2 (PDB:4C0S). New variants are shown in black, with labels in bold, previously described mutations are in white. T24M and V437F are buried. The binding site of eEF1B is highlighted in white, and the GTP binding site in dark grey.