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REVIEW ARTICLE

Inaugural Review Prize 2023: The exercise hyperpnoea dilemma: A 21st-century perspective

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Abstract

During mild or moderate exercise, alveolar ventilation increases in direct proportion to metabolic rate, regulating arterial CO₂ pressure near resting levels. Mechanisms giving rise to the hyperpnoea of exercise are unsettled despite over a century of investigation. In the past three decades, neuroscience has advanced tremendously, raising optimism that the 'exercise hyperpnoea dilemma' can finally be solved. In this review, new perspectives are offered in the hope of stimulating original ideas based on modern neuroscience methods and current understanding. We first describe the ventilatory control system and the challenge exercise places upon blood-gas regulation. We highlight relevant system properties, including feedforward, feedback and adaptive (i.e., plasticity) control of breathing. We then elaborate a seldom explored hypothesis that the exercise ventilatory response continuously adapts (learns and relearns) throughout life and ponder if the memory 'engram' encoding the feedforward exercise ventilatory stimulus could reside within the cerebellum. Our hypotheses are based on accumulating evidence supporting the cerebellum's role in motor learning and the numerous direct and indirect projections from deep cerebellar nuclei to brain-stem respiratory neurons. We propose that cerebellar learning may be obligatory for the accurate and adjustable exercise hyperpnoea capable of tracking changes in life conditions/experiences, and that learning arises from specific cerebellar microcircuits that can be interrogated using powerful techniques such as optogenetics and chemogenetics. Although this review is speculative, we consider it essential to reframe our perspective if we are to solve the till-now intractable exercise hyperpnoea dilemma.

KEYWORDS

cerebellum, chemogenetics, exercise, hyperpnoea, learning, optogenetics, plasticity

1 | INTRODUCTION

Exercise is the greatest ventilatory stimulus routinely encountered in life. The hyperpnoea of exercise is a vital homeostatic response that must be robust and adaptable. Despite the physiological significance of exercise hyperpnoea, neural mechanisms underpinning the near-precise matching of alveolar ventilation to metabolic rate (i.e., $\Delta\dot{V}_A$

$\propto \Delta\dot{V}_{CO_2}$) with only minor alterations in arterial blood-gas tensions (Douglas & Haldane, 1909; Douglas et al., 1912; Haldane & Priestley, 1905) remain enigmatic. Consequently, exercise hyperpnoea has been termed the 'unanswered question' (Whipp, 2008), the 'ultra-secret' (Grodins, 1981) and the 'holy grail' (Dempsey, 2006) of respiratory and exercise physiology.

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By the end of the 19th century, German physiologists Julius Geppert and Nathan Zuntz articulated possible neural circuits and sources of stimuli involved in the control of breathing during exercise (Geppert & Zuntz, 1888; Zuntz & Geppert, 1886). In the 20th century, Dejours (1964), Grodins (1950) and Wasserman et al. (1979) provided updated theories at decade intervals on the nature of exercise hyperpnoea. Postulated factors were initially supported and then denied, generating more questions than answers. Since the late 1980s, major advances in our understanding of exercise hyperpnoea have been limited, despite continuing (but waning) interest (Bruce, 2022; Bruce et al., 2019; Casaburi, 2012; Dempsey et al., 2022; Forster et al., 2012; Haouzi, 2020; Parkes, 2017; Ward, 2019).

Over the past three decades, tremendous technological and conceptual advances in neuroscience have been made, including increased awareness of the capacity for neuroplasticity in respiratory control (Fuller & Mitchell, 2017; Mitchell & Babb, 2006; Mitchell & Baker, 2022; Vose et al., 2022). With increased knowledge, new ideas concerning exercise hyperpnoea have emerged, accompanied by powerful new techniques to test those ideas. The purposes of this review are to (a) define what is known about exercise hyperpnoea, its underlying mechanisms and the neural system through which it operates, and (b) pose a novel hypothesis that cerebellar learning is obligatory to develop a 'normal' exercise ventilatory response that continuously adapts (learns and relearns) to meet the demands on pulmonary ventilation and gas exchange imposed by the ever-changing physiological conditions experienced throughout life.

2 | PART I: THE VENTILATORY CONTROL SYSTEM

The ventilatory response to mild or moderate steady-state exercise consists of three distinct phases. Phase I describes an immediate (within one pedal stroke or step) increase in minute ventilation (\dot{V}_E ; see Table 1 for list of frequently used abbreviations) at exercise onset. Phase II describes an exponential increase in \dot{V}_E with a time constant of 60–90 s. Phase III describes a \dot{V}_E plateau (i.e., steady-state) beginning approximately 3 min after exercise onset. A successful exercise hyperpnoea is defined by regulation of arterial P_{CO_2} (P_{aCO_2}) near its set-point (~ 40 mmHg) due to proportional increases in \dot{V}_A and \dot{V}_{CO_2} (Dempsey & Welch, 2023). However, it must be appreciated that good P_{aCO_2} regulation (i.e., ± 2 mmHg of rest) is not evidence of a single mechanism that gives rise to precise $\dot{V}_A:\dot{V}_{\text{CO}_2}$ coupling. For example, even a 25% dissociation between the primary drive to \dot{V}_A and \dot{V}_{CO_2} yields only an ~ 2 mmHg change in P_{aCO_2} (Bennett & Fordyce, 1985) due to chemofeedback that 'layers' onto the primary (feedforward) drive to breathe once P_{aCO_2} regulation is lost (see Figure 1). Indeed, it is very difficult to distinguish between precise $\dot{V}_A:\dot{V}_{\text{CO}_2}$ feedforward coupling versus a moderate mismatch refined by chemofeedback. In most vertebrate species, mild hypocapnia is observed during mild or moderate steady-state exercise, whereas humans exhibit slight hypercapnia (Clark et al., 1980; Dempsey et al., 1984; Fordyce et al., 1982). Hence, despite widespread use of the term 'isocapnic hyperpnoea' to characterise

Highlights

- **What is the topic of this review?**

This review reframes our perspective on the control of breathing during mild or moderate exercise, emphasising concepts of learning, memory and plasticity.

- **What advances does it highlight?**

We propose that exercise hyperpnoea continuously adapts to ever-changing life conditions/experiences. We question if the memory 'engram' encoding exercise hyperpnoea resides within the cerebellum and outline powerful new methods to test this hypothesis. It is our hope that this review generates new ideas and renewed study of exercise hyperpnoea in the 21st century.

a human exercise ventilatory response, we suggest that the terms 'isocapnia' and 'hyperpnoea' should be used with caution since the precision of P_{aCO_2} regulation varies across species and with exercise intensity.

2.1 | Operational model of the ventilatory control system

An operational model of the ventilatory control system is illustrated in Figure 2; this simplified model is based upon control theory as described by Houk (1988) and mathematical simulations of the exercise stimulus described by Fred Grodins (Grodins et al., 1954;

TABLE 1 List of abbreviations.

Abbreviation	
G_{EX}	Exercise gain
G_{SYS}	System gain
LTM	Long-term modulation
P_{aCO_2}	Arterial CO_2 partial pressure
P_{aO_2}	Arterial O_2 partial pressure
pH_a	Arterial pH
S	Feedback gain
STM	Short-term modulation
\dot{V}_A	Alveolar ventilation
\dot{V}_{CO_2}	Rate of CO_2 production
\dot{V}_{O_2}	Rate of O_2 consumption
V_D	Dead space volume
\dot{V}_E	Minute ventilation
V_T	Tidal volume

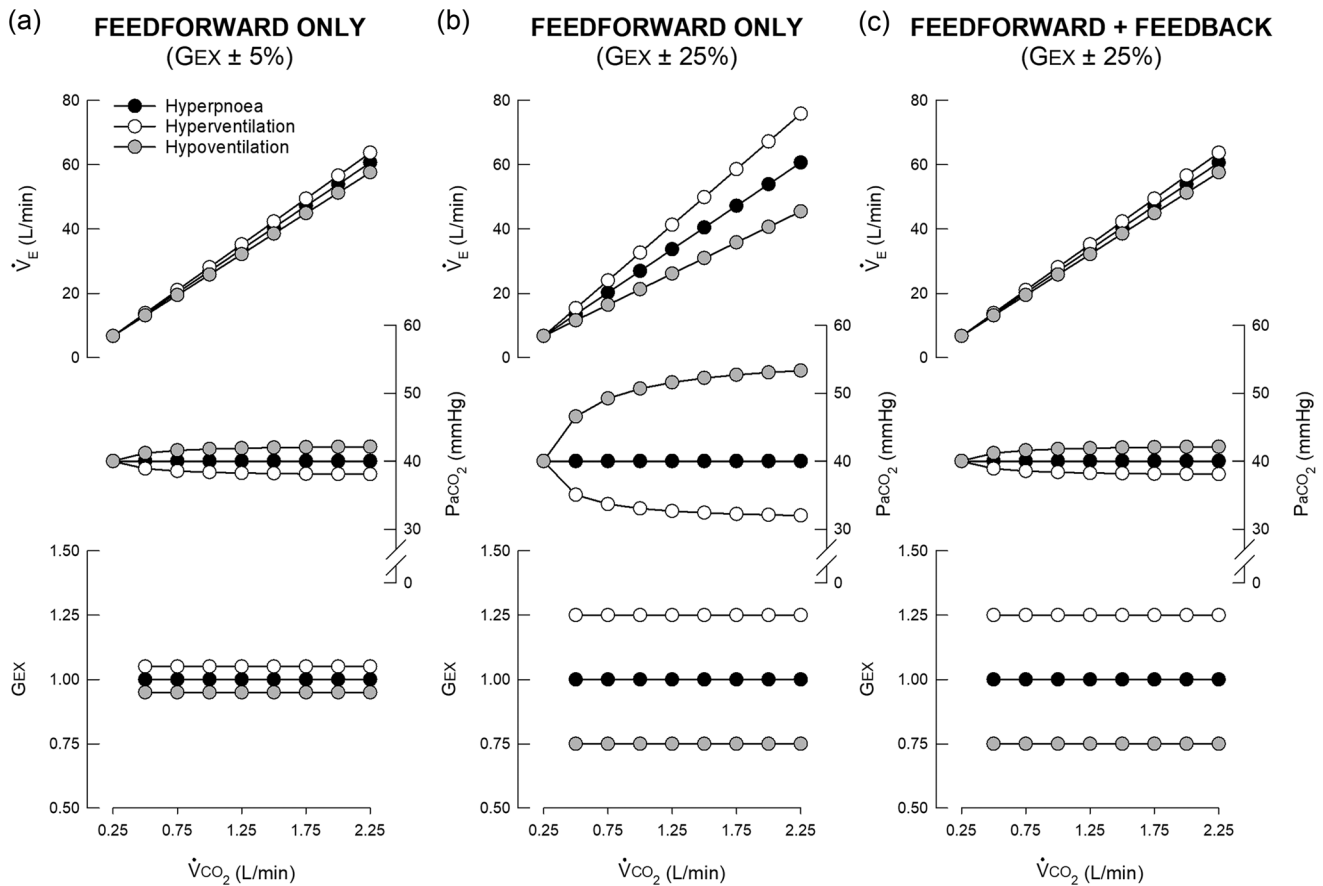


FIGURE 1 Feedforward and feedback interactions to exercise hyperpnoea. (a) Illustration of very good regulation of arterial CO₂ pressure (P_{aCO_2}) with small (5%) deviations in the feedforward exercise gain (G_{EX}) from an ‘ideal gain’ that would generate isocapnia (black symbols); there are corresponding 5% deviations in the system gain ($\dot{V}_E:\dot{V}_{CO_2}$). Slight increases in slope produce hypocapnia (white symbols), whereas slight decreases in slope produce modest hypercapnia (grey symbols). In this case, a 5% increase or decrease in G_{EX} above or below the isocapnic value would alter P_{aCO_2} by 2 mmHg from its resting level (40 mmHg) at a \dot{V}_{CO_2} of 2.25 L/min (10-fold above rest). (b) The hypothetical P_{aCO_2} regulation expected with larger (25%) deviations in G_{EX} from ‘ideal’, with corresponding deviations in the $\dot{V}_E:\dot{V}_{CO_2}$ relationship; in this simulation, an absence of chemofeedback is assumed. Such 25% variances in G_{EX} would decrease P_{aCO_2} by 8 mmHg (with excess G_{EX}) or increase P_{aCO_2} by 13 mmHg (with inadequate G_{EX}) from resting levels. (c) Restoration of very good (not perfect) P_{aCO_2} regulation with 25% deviations in G_{EX} but with CO₂ chemofeedback layered onto the feedforward drive, thereby minimising errors in P_{aCO_2} regulation. With the combination, only a small shift (5%) in the $\dot{V}_E:\dot{V}_{CO_2}$ slope from an isocapnic value occurs despite 25% excessive or inadequate feedforward drive. Thus, as in (a), P_{aCO_2} is once again regulated within ~2 mmHg of its resting value—a response difficult to discriminate from isocapnia without repeated P_{aCO_2} measurements. For all calculations in this simulation, the dead space to tidal volume ratio was kept at 0.2. \dot{V}_{CO_2} , rate of CO₂ production; \dot{V}_E , minute ventilation.

Grodins et al., 1967). Understanding mechanisms controlling physiological systems requires knowledge of the regulated variables. During mild or moderate exercise, the primary regulated variable is P_{aCO_2} . The relationships between the controlled (P_{aCO_2}), controlling (\dot{V}_A) and disturbing variables (\dot{V}_{CO_2}) are summarised in the alveolar ventilation equation:

$$P_{aCO_2} = \frac{R \times T \times \dot{V}_{CO_2}}{\dot{V}_A} \quad (1)$$

where R is the gas constant (2.785 [mL BTPS mmHg]/[mL STPD K]) and T is temperature in K. This assumes only conservation of mass, steady-state conditions and an inspired CO₂ fraction of zero.

Since the alveolar volume is equal to the tidal volume minus the physiological dead space (i.e., $\dot{V}_A = V_T - V_D$), \dot{V}_A can be replaced with \dot{V}_E :

$$P_{aCO_2} = \frac{R \times T \times \dot{V}_{CO_2}}{\dot{V}_E (1 - V_D/V_T)} \quad (2)$$

Figure 1 illustrates that isocapnia from rest to exercise can be achieved if \dot{V}_E and \dot{V}_{CO_2} increase in direct proportion, with a slope equal to $RT/[P_{aCO_2}(1 - V_D/V_T)]$ (Mitchell, 1990; Mitchell et al., 1984). Deviations in the $\dot{V}_E:\dot{V}_{CO_2}$ slope from this value, even if linear, lead to hypocapnia or hypercapnia. Equation 2 is extremely important in understanding exercise hyperpnoea since it expresses all possible system solutions during steady-state (phase III) exercise.

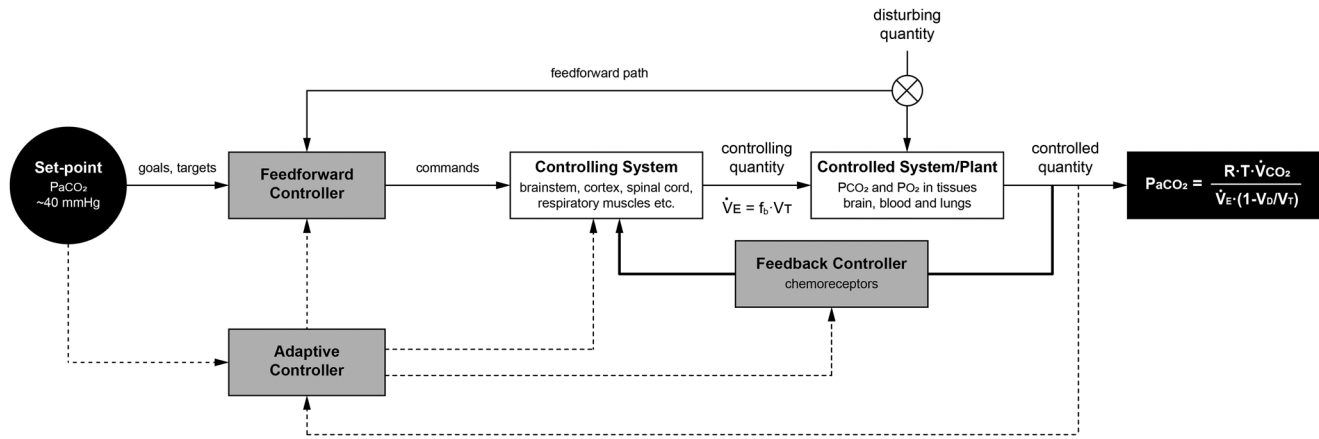


FIGURE 2 Operational model of the ventilatory control system. See text for description. Dashed lines indicate adaptive controller pathways. Thick black lines indicate feedback loop. f_b , breathing frequency; P_{aCO_2} , arterial CO_2 partial pressure; P_{aO_2} , arterial O_2 partial pressure; P_{CO_2} , CO_2 partial pressure; P_{O_2} , O_2 partial pressure; \dot{V}_{CO_2} , rate of CO_2 production; V_D , dead space volume; \dot{V}_E , minute ventilation; V_T , tidal volume.

2.1.1 | Controlling system

Typically, P_{aCO_2} is regulated near a set-point of ~ 40 mmHg. With an increase in \dot{V}_{CO_2} , P_{aCO_2} can be defended only by changes in neural respiratory motor output that drive appropriate increases in \dot{V}_E . The neural control system (i.e., central controller) includes supra-pontine (e.g., primary motor cortex, periaqueductal grey, etc.) and ponto-medullary (e.g., ventral respiratory column) circuits that subserve voluntary (i.e., behavioural) and automatic breathing, respectively. These neural circuits activate more than 60 different respiratory muscles to produce changes in airflow with an inherent frequency and amplitude. The total volume of air displaced per minute in response to forcing functions generated by the central controlling system is the controlling quantity, or \dot{V}_E , given by the product of V_T and breathing frequency. Minute ventilation is determined by coordinated activation of spinal respiratory pump muscles (e.g., diaphragm, intercostal and abdominal muscles) and cranial upper airway resistance muscles (e.g., genioglossus) (Pilarski et al., 2019). The timing and pattern of respiratory muscle recruitment during exercise follows the 'principle of minimum effort' (Otis et al., 1950) to minimise respiratory muscle work.

The primary function of the ventilatory control system during exercise is to regulate P_{aCO_2} at or near its resting level. In this context, the term 'feedback' is defined with respect to errors in P_{aCO_2} regulation. Thus, chemofeedback mechanisms sense changes in P_{aCO_2} and/or arterial pH, driving or inhibiting \dot{V}_E to curtail P_{aCO_2} deviations from its set-point. For the purposes of this review, feedback does not refer to known sensory inputs from the lungs (e.g., pulmonary stretch receptors) or exercising limbs (e.g., metaboreceptors). If P_{aCO_2} deviates from rest due to excessive or inadequate feedforward drives to breathe, chemofeedback minimises the error in P_{aCO_2} regulation.

In contrast, 'feedforward' stimuli refers to neural drives to breathe that predict or anticipate the need for increased \dot{V}_E in response to

increased \dot{V}_{CO_2} , thereby regulating P_{aCO_2} without need for chemofeedback. In other words, feedforward commands translate goals, targets and information about potential disturbances to P_{aCO_2} that have not yet occurred (Houk, 1988). The distinction between feedforward versus feedback is an essential premise of this review and is defined by whether a neural mechanism is activated directly by changes in P_{aCO_2} . For example, P_{aCO_2} changes are not known to alter sensory afferent inputs from the lung or muscle, but do directly alter chemoreceptor inputs. Whilst feedforward and feedback control offer short-term protection that minimises errors in P_{aCO_2} regulation, adaptive control provides a longer-lasting solution to accommodate changing conditions, as discussed later in this review.

2.1.2 | Controlled system

The part of the system that is controlled is referred to as the 'plant'. The ventilatory control plant includes structures and processes involved in pulmonary gas exchange (e.g., alveoli, pulmonary capillaries) that directly influence P_{O_2} and P_{CO_2} in the lungs, tissues, blood and brain. The plant is disrupted by factors such as physiological dead space (expressed as V_D/V_T), pulmonary mechanics (compliance or resistance), inspired P_{O_2} and P_{CO_2} , and the respiratory exchange ratio ($\dot{V}_{CO_2}/\dot{V}_{O_2}$).

During mild or moderate exercise, P_{aCO_2} is tightly regulated and, as a result, arterial P_{O_2} (P_{aO_2}) and pH (pH_a) remain close to resting values. The relationship between alveolar P_{CO_2} and P_{O_2} is given by the alveolar gas equation (Fenn et al., 1946):

$$P_{AO_2} = P_{IO_2} - \frac{P_{ACO_2}}{RQ} \quad (3)$$

where P_{AO_2} is alveolar P_{O_2} , P_{IO_2} is inspired P_{O_2} , P_{ACO_2} is alveolar P_{CO_2} and RQ is respiratory quotient, $\dot{V}_{CO_2}/\dot{V}_{O_2}$.

TABLE 2 List of definitions.

Term	Definition
Adaptive control	P_{aCO_2} regulation due to short- and long-term adjustments in the moment-to-moment properties of the ventilatory control system
Central neurogenic	A feedforward stimulus for P_{aCO_2} regulation originating from cortical and subcortical structures
Exercise hyperpnoea	P_{aCO_2} regulation about its set-point due to proportional changes in \dot{V}_A and \dot{V}_{CO_2}
Feedback control	Closed-loop chemosensory regulation of P_{aCO_2}
Feedforward control	Open-loop P_{aCO_2} regulation by predictive or corrective commands
Long-term modulation	A persistent change in the exercise ventilatory response that outlasts the inducing stimulus
Peripheral neurogenic	A feedforward stimulus for P_{aCO_2} regulation originating from peripheral sensory receptors
Short-term modulation	An immediate change in the exercise ventilatory response that is reversed upon removal of the inducing stimulus

Abbreviations: \dot{V}_A , alveolar ventilation; \dot{V}_{CO_2} , rate of CO_2 production; P_{aCO_2} , arterial CO_2 partial pressure.

In Equation 3, alveolar and arterial P_{CO_2} are reasonable approximations of one another; however, a similar assumption cannot be made for alveolar and arterial P_{O_2} . The relationship between P_{aCO_2} and pH_a is formally expressed by the Henderson–Hasselbalch equation applied to the hydration/dehydration equilibrium of CO_2 :

$$pH_a = pK + \log \left[\frac{HCO_3^-}{0.03 \times P_{aCO_2}} \right] \quad (4)$$

where pK is the collective dissociation constant (6.1) and 0.03 is the solubility for CO_2 in plasma.

Thus, since alveolar (and arterial) P_{CO_2} is inversely related to alveolar P_{O_2} and pH_a , careful P_{aCO_2} regulation achieves satisfactory regulation of alveolar P_{O_2} and pH_a .

3 | PART II: LAYERS OF REGULATION

Based on the operational model of the ventilatory control system presented in Figure 2, three major control strategies (defined in Table 2), representing independent but interacting layers of regulation, govern the integrated exercise ventilatory response. Strategies include (a) feedback, (b) feedforward, and (c) adaptive control. Figure 3 illustrates some major identified sources of feedforward and feedback contributions to exercise hyperpnoea. Evidence for each control strategy in P_{aCO_2} regulation during exercise is described below; interested readers are directed elsewhere for more comprehensive discussion (Forster et al., 2012).

3.1 | Feedback control

Feedback control operates as closed-loop chemosensory regulation of P_{aCO_2} . During mild or moderate exercise in healthy humans, P_{aCO_2} is maintained close to the set-point, defined here as the mean resting value. Thus, classical arterial chemoreception via central and peripheral chemoreceptors has been dismissed as a major contributing factor to the control of breathing during exercise at sea level (Grodins,

1950; Shea et al., 1993; Wasserman, Whipp, Koyal et al., 1975). However, in all other mammalian species (as well as other terrestrial vertebrates) studied to date, breathing increases in excess of \dot{V}_{CO_2} , decreasing P_{aCO_2} slightly from rest to exercise in steady-state (Bainton, 1978; Dempsey et al., 1984; Smith et al., 1983). Thus, CO_2 chemoreception in species other than humans actually inhibits breathing in exercise (Mitchell, 1990).

Multiple studies confirm that carotid body chemoreceptors are not necessary for a successful exercise hyperpnoea, as shown by the essentially normal exercise ventilatory responses in carotid denervated ponies (Pan et al., 1983), goats (Mitchell et al., 1984) and dogs (Bainton, 1978). However, three caveats must be acknowledged. First, carotid denervation alone increases the resting P_{aCO_2} set-point and slightly reduces the $\dot{V}_E:\dot{V}_{CO_2}$ slope as required in accordance with Equation 2 (Bainton, 1978; Mitchell et al., 1984). Second, carotid denervated ponies exhibit exaggerated hyperventilation during phases I and II of the exercise ventilatory response (Forster et al., 1983) with a similar but opposite (exaggerated hypoventilation) response in asthmatic humans with carotid resection (Honda et al., 1979). Third, peripheral chemoreceptors contribute tonic respiratory drive at rest and during exercise, as indicated by the ~10% inhibition of breathing in response to a few breaths of hyperoxia (Dejours, 1962). Whilst peripheral chemoreceptors may ‘fine-tune’ \dot{V}_E by minimising blood-gas disturbances during steady-state transitions (Pan et al., 1983), the primary exercise stimulus is feedforward with respect to P_{aCO_2} regulation in humans and other vertebrate species.

Beyond mild to moderate exercise, peripheral chemoreceptors are a logical contributor to the hyperventilation of heavy exercise, sensing increases in core body temperature and circulating metabolites such as lactic acid, noradrenaline, adenosine, glucose and potassium (Paterson, 1992; Ward, 1994). Asthmatic patients with carotid resection (Wasserman, Whipp, Koyal et al., 1975) and children with central congenital hypoventilation syndrome (Shea et al., 1993) display normal steady-state hyperpnoea in moderate exercise, but blunted hyperventilation in heavy exercise, signifying peripheral chemoreceptor contributions. Conversely, other reports show that the hyperventilation of heavy exercise is independent of peripheral

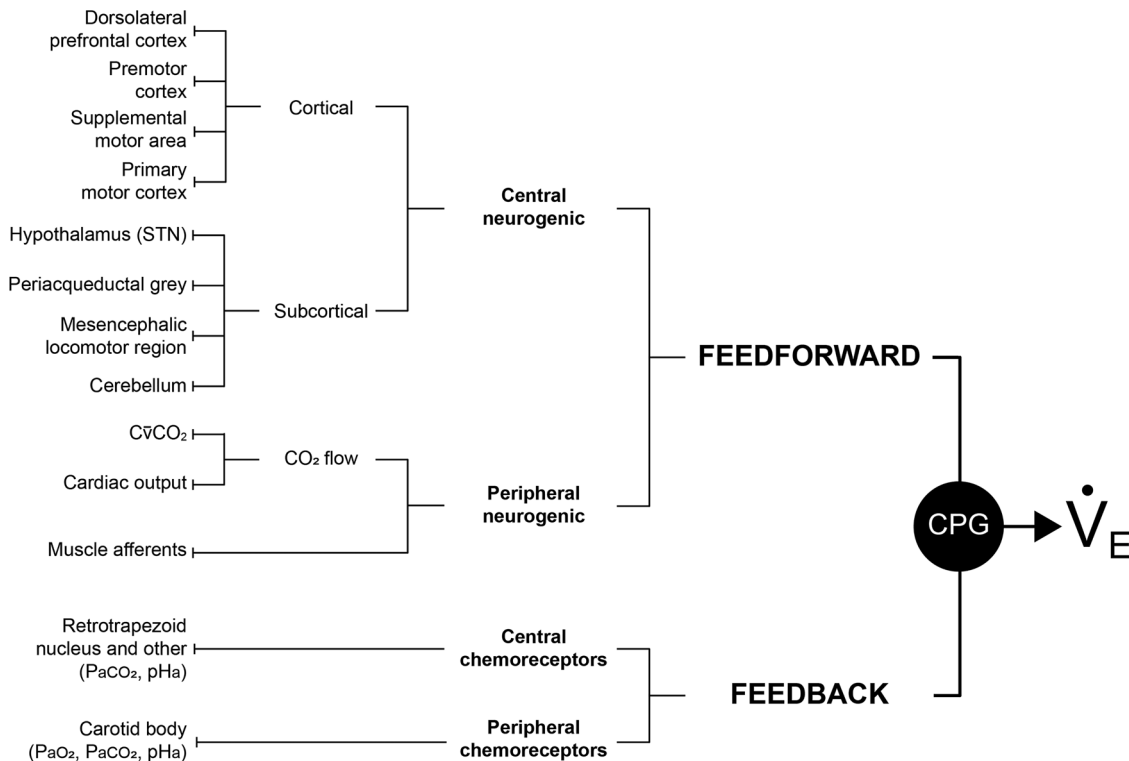


FIGURE 3 Sources of feedforward and feedback stimuli. The exercise ventilatory response results from combined (layered) feedforward and feedback drives to the brainstem respiratory central pattern generator (CPG). Many sources of feedforward and sensory stimuli have been identified, and are listed on the left. Muscle afferents include sensory afferent neurons from both respiratory and locomotor muscles. CPG, respiratory central pattern generators; $C_{\bar{v}CO_2}$, mixed-venous CO_2 content; P_{aCO_2} , arterial CO_2 partial pressure; P_{aO_2} , arterial O_2 partial pressure; pH_a , arterial pH; STN, subthalamic nucleus; \dot{V}_E , minute ventilation. Adapted from Cunningham et al. (1986).

chemoreceptors or metabolic acidosis (Pan et al., 1986). The view that changes in arterial potassium is sufficient to drive breathing during heavy exercise (Paterson, 1992; Paterson et al., 1989) is not borne out by studies in goats, since ventilatory responses to hyperkalaemia are insufficient to account for exercise hyperpnoea (Warner & Mitchell, 1990). Further, the ventilatory response to increased circulating noradrenaline is insufficient to account for exercise hyperpnoea or hyperventilation in moderate or heavy exercise (Pizarro et al., 1992; Warner & Mitchell, 1991).

3.2 | Feedforward control

Feedforward control is open-loop P_{aCO_2} regulation by predictive or corrective commands. The feedforward controller specifies goals or targets about the overall control process and receives sensory input from receptors that detect potential disturbances to P_{aCO_2} . Literature supports at least two types of feedforward mechanism in exercise hyperpnoea: (a) central neurogenic and (b) peripheral neurogenic.

3.2.1 | Central neurogenic mechanisms

Central command has been defined as a feedforward mechanism involving 'parallel activation of motor and cardiovascular centres'

(Goodwin et al., 1972). In this context, breathing should be regarded as a 'motor' (vs. autonomic) system. The phase I ventilatory response is remarkably fast and 'cannot be brought about by any chemical regulation as a result of the processes in the working muscles' (Krogh & Lindhard, 1913). Anticipation of exercise is sufficient to increase breathing in the transition from rest to work and was suggested to represent 'irradiation of impulses from the motor cortex' (Krogh & Lindhard, 1913). We prefer to use the term 'central neurogenic' (vs. central command), which we define as a feedforward stimulus with regard to P_{aCO_2} regulation that originates from cortical and/or subcortical structures of the central nervous system.

Studies using positron emission tomography to measure changes in regional cerebral blood flow as a reflection of regional neuronal activity during and/or after voluntary or imagined exercise demonstrate increased cortical and subcortical circuit activation, including the dorsolateral prefrontal cortex, premotor cortex, supplementary motor area, primary motor cortex and cerebellum (Fink et al., 1995; Thornton et al., 2001; Williamson et al., 2002; Williamson et al., 2006). Due to associations with learning and memory (formation/storage), some have postulated that activation of the aforementioned circuits may signify a learned or behavioural response at an unknown site (Fink et al., 1995; Guz, 1997; Thornton et al., 2001).

Evidence for subcortical central neurogenic contributions to breathing in exercise stem from 'fictive' respiration and locomotion

studies in neuromuscularly blocked decorticate cats. In these studies, stimulation of the hypothalamus and mesencephalic locomotor region elicited parallel increases in phrenic and limb motor output, as well as arterial blood pressure, accompanied by a decrease in P_{aCO_2} (Eldridge et al., 1981; Eldridge et al., 1985). Another potential site of interest is the midbrain periaqueductal grey, regarded as a cognitive integrator for central and peripheral neurogenic stimuli (Mantyh, 1983; Paterson, 2014), with functional connectivity to and from higher brain regions (e.g., dorsolateral prefrontal cortex) (An et al., 1998), basal ganglia (Horiuchi et al., 2009), brainstem cardiorespiratory central pattern generators (Bandler & Carrive, 1988) and group III/IV afferents (Kramer et al., 1996). An increase in periaqueductal grey activity (i.e., local field potentials) in humans undergoing deep brain stimulation for neuropathic pain occurs in anticipation of mild exercise, mirrored by changes in heart rate and blood pressure (Green & Paterson, 2008, 2020; Green et al., 2007).

Despite extensive literature supporting cortical and subcortical sites as primary mediators of the exercise ventilatory response, evidence from studies of $\dot{V}_E:\dot{V}_{CO_2}$ tracking during sinusoidal exercise indicates that the exercise ventilatory response can be uncoupled from limb movement (and presumably central neurogenic stimuli) (Casaburi et al., 1977; Haouzi et al., 2004). As the duration between sinusoidal oscillations in exercise workload decreases, \dot{V}_E and \dot{V}_{CO_2} changes also decrease but with an increased phase lag. Collectively, these observations led to the belief that the exercise ventilatory response is more closely associated with \dot{V}_{CO_2} changes vs. central neural (motor) command. In agreement, the exercise ventilatory response tracks \dot{V}_{CO_2} vs. limb movement in ponies (Forster et al., 1984). Whilst the relationship between \dot{V}_E and \dot{V}_{CO_2} (and the precision of P_{aCO_2} regulation) during treadmill exercise is similar when workload is increased via treadmill grade vs. speed, the pattern of breathing shifts in both dogs (Bainton, 1972) and goats (Smith et al., 1983). Thus, the pattern of locomotion (stride frequency vs. strength of contraction) impacts the pattern of breathing (frequency vs. volume) during exercise. Whether this shift arises from central neurogenic versus peripheral neurogenic stimuli from exercising limbs is unknown.

3.2.2 | Peripheral neurogenic mechanisms

Peripheral neurogenic mechanisms are a feedforward stimulus (with respect to P_{aCO_2} regulation) originating from peripheral sensory neurons. Two main peripheral neurogenic stimuli considered in the context of exercise hyperpnoea are (a) CO_2 flux to the lung, and (b) group III/IV afferents.

By infusing CO_2 into the femoral vein of exercising rats, Yamamoto and Edwards Jr (1960) reported a proportional increase in \dot{V}_A without appreciable change in P_{aCO_2} , claiming that ' CO_2 is sufficient stimulus for the regulation of its own concentration'. However, the authors were unable to prevent (and account for) large changes in cardiac output and blood volume during CO_2 loading, which may independently stimulate breathing (Ponte & Purves, 1978). In follow-up studies, some supported the findings of Yamamoto and Edwards Jr (Green & Schmidt,

1984; Wasserman et al., 1975), whereas others contradicted them (Bennett et al., 1984; Reischl et al., 1979). Carbon dioxide flow to the lungs (\dot{Q}_{CO_2}) is given by the product of cardiac output (\dot{Q}) and mixed-venous CO_2 content ($C_{\dot{v}CO_2}$):

$$\dot{Q}_{CO_2} = \dot{Q} \times C_{\dot{v}CO_2} \quad (5)$$

For decades, it was hypothesised that an unknown mixed-venous chemoreceptor capable of detecting changes in mixed-venous P_{CO_2} and/or CO_2 flow to the lungs could account for an isocapnic exercise hyperpnoea. However, although intrapulmonary CO_2 chemoreceptors exist in birds and reptiles (Milsom et al., 2004; Powell et al., 1988; Tallman & Grodins, 1982), analogous receptors have never been found in mammals despite considerable effort (Bartoli et al., 1973; Mitchell et al., 1980). Some authors claimed to provide evidence of venous P_{CO_2} sensors located in the pulmonary artery (Duke et al., 1963), pulmonary C-fibre endings (Trenchard et al., 1984), aorticopulmonary glomus tissue (Hughes, 1965) and CO_2 -modulated pulmonary stretch receptors (Green et al., 1986), while others refuted such claims (Coleridge et al., 1966; Cropp & Comroe, 1961; Dejours et al., 1955; Fordyce & Grodins, 1980; Sylvester et al., 1973). Abrupt release of venous tourniquets elicits hyperpnoea within 4 s, but this response has been attributed to activation of cardiopulmonary baroreceptors sensitive to changes in blood volume, stretch and pressure versus changes in venous P_{CO_2} or CO_2 flow (Mills, 1944).

In the 1980s, Phillipson and co-workers provided evidence supporting CO_2 flow per se as a unique stimulus to breathing. By removing CO_2 from the venous circulation at a rate equal to its production in resting sheep, prolonged apnoea was observed, despite maintenance of normal P_{aCO_2} , pH_a and P_{aO_2} (Phillipson et al., 1981b). Thus, the authors claimed breathing depends upon CO_2 flow to the lungs by an unknown mechanism. Using extracorporeal perfusion, Phillipson et al. (1981a) infused or removed CO_2 from the venous circulation in exercising sheep. Blood drained from the inferior vena cava and jugular veins was pumped through extracorporeal membrane lungs; \dot{V}_{CO_2} was raised using 100% CO_2 or lowered using O_2/N_2 mixtures, and the blood then returned to the superior vena cava. In 51 steady-state measurements, loading or scrubbing CO_2 with or without exercise led to proportional changes in \dot{V}_A without significant change in P_{aCO_2} . However, the statistical grounds on which these conclusions were made have been challenged (Bennett et al., 1984) on the basis that: (a) the use of pooled data makes it difficult to discern true isocapnia versus predicted changes in P_{aCO_2} when \dot{V}_{CO_2} changes are small (in this case, ~3-fold above rest); and (b) the authors considered lack of statistically significant difference as 'no change', whereas a more robust conclusion could be achieved by showing no significant difference from baseline whilst also demonstrating significant differences from predicted increases in P_{aCO_2} based upon simple mathematical models assuming only conventional CO_2 chemofeedback (Bennett & Fordyce, 1988; Bennett & Fordyce, 1993; Mitchell, 1990).

The possibility of intrathoracic pulmonary chemoreceptors capable of detecting changes in CO_2 flow to the lungs received its greatest

support in dual extracorporeal bypass experiments isolating the pulmonary from systemic circulation in dogs. Manipulating either side of the CO₂ flow equation (Equation 5) independently (up to 8-fold increases in pulmonary blood flow; up to 75 mmHg pulmonary arterial P_{CO₂}) increased \dot{V}_E in direct proportion to CO₂ flow whilst preserving P_{aCO₂} at baseline levels (Green & Sheldon, 1983; Sheldon & Green, 1982). These effects were abolished by bilateral cervical vagotomy.

The 'cardiodynamic hyperpnoea' theory was proposed from observations of an immediate increase in \dot{V}_E (and maintenance of end-tidal P_{CO₂}) in response to increased cardiac output caused by intravenous isoprenaline infusion or cardiac pacing in anaesthetised and unanaesthetised dogs (Wasserman et al., 1974). Mechanical distention of the heart and pulmonary arteries may reflexively stimulate breathing during exercise, but again, opposing evidence exists (Orr et al., 1988). Banner et al. (1988) observed blunted cardiovascular responses but normal ventilatory responses during voluntary and electrically induced leg exercise in pulmonary denervated heart-lung transplant recipients. The absence of cardiopulmonary afferents in this population casts doubt on the conclusions of Sheldon and Green, and the notion that the lungs are a site for mixed-venous CO₂ sensation. The 'vascular distention' hypothesis posits that changes in cardiac output could be detected by systemic (limb) smooth muscle mechanoreceptors, which activate thin-fibre group III/IV afferents (Haouzi, 2014; Haouzi et al., 1999). Thus, CO₂ flow to the lungs may be detected through means other than mixed-venous chemoreceptors, such as changes in local muscle blood flow (and cardiac output).

Since the late 19th century, it has been known that muscle activity stimulates breathing (Johansson, 1894). Evidence for the role of muscle sensory afferent neurons in exercise hyperpnoea comes from classical papers by Comroe and Schmidt (1943), Coote et al. (1971), McCloskey and Mitchell (1972), and Kao (1963). Inspired by the work of Alam and Smirk (1937), Comroe and Schmidt (1943) observed an increase in ventilation during post-exercise ischaemia, suggesting a metabolite-induced reflex originating in the exercising muscles. The cross-circulation experiments of Kao (1963) isolated the effects of muscle afferents from central neurogenic and blood-borne breathing stimuli in anaesthetised dogs. Electrical stimulation of the hind limbs of a neurologically intact (i.e., 'neural') dog with hind-limb venous blood diverted to a second, non-exercised (i.e., 'humoral') dog via interposed abdominal anastomoses elicited immediate hyperventilation in the neural dog, which was abolished by spinal cord transection. Coote et al. (1971) demonstrated that electrical stimulation of ventral roots S6-L1 increased ventilation, heart rate and blood pressure concurrent with tetanic hind-limb muscle contraction in anaesthetised cats. These responses were reduced by dorsal root transection and muscle paralysis. McCloskey and Mitchell (1972) then noted that the exercise pressor reflex persisted when large myelinated (group I/II) fibres were prevented from transmitting sensory information to the spinal cord via nodal blockade of dorsal roots. Conversely, the pressor reflex was arrested by local anaesthetic blockade of dorsal roots that preferentially suppresses sensory inputs from small myelinated and unmyelinated (group III/IV) fibres. In humans, the steady-state exercise ventilatory response is blunted by lumbar epidural fentanyl (μ -opioid

receptor agonist) injections that partially block sensory input from the exercising limbs (Amann et al., 2010; Amann et al., 2011). On balance, available evidence suggests that muscle afferents do contribute to exercise hyperpnoea.

However, once again, conflicting evidence exists. For example, Cross et al. (1982) confirmed that spinal cord transmission was not necessary for hyperpnoea during electrically induced hind-limb activation in anaesthetised dogs. Further counter-evidence comes from studies using percutaneous electrical stimulation of the quadriceps muscles to simulate aspects of exercise in intact (Asmussen et al., 1943) and paraplegic humans (Adams et al., 1984; Asmussen et al., 1943; Brice et al., 1988; Brown et al., 1990). In each instance, subjects displayed a 'normal' exercise hyperpnoea. However, muscle stimulation does not stimulate all, or even the most important, aspects of spontaneous exercise since it eliminates the possibility of a feedforward central neurogenic drive to breathe. What is more, the magnitude of increase in \dot{V}_{CO_2} during muscle stimulation is limited, making it difficult to discriminate between isocapnic- versus chemoreflex-driven hyperpnoea (Bennett & Fordyce, 1988).

In summary, although there is evidence for both central and peripheral neurogenic contributions to exercise hyperpnoea, none of the evidence to date is without controversy and none of the claimed mechanisms (even in combination with P_{aCO₂} feedback) are sufficient to explain exercise hyperpnoea. New hypotheses and/or layers of regulation must be considered.

3.3 | Adaptive control

The combination of (as yet unknown) feedforward and (CO₂) feedback neural mechanisms can explain exercise hyperpnoea in all vertebrate species studied to date, at least in normal conditions. However, physiological conditions inevitably change throughout life, requiring adaptation of the neural system controlling breathing to assure regulation of P_{aCO₂} with the same precision from rest to exercise (Mitchell, 1990; Mitchell et al., 1984). With growth and development, gain or loss of body mass, pregnancy, the onset of disease or injury, and the transition to old age, a fixed and immutable feedforward exercise stimulus cannot produce the precise blood-gas regulation observed, even after fine-tuning via chemofeedback. When conditions change, adaptive control makes short- and long-term adjustments in properties of feedforward and/or feedback mechanisms to restore homeostasis (Priban & Fincham, 1965). Two distinct but related mechanisms of adaptive control in the exercise ventilatory response are modulation and plasticity (Babb et al., 2010; Mitchell & Babb, 2006). It has been proposed that exercise hyperpnoea may even represent a learned response guided by accumulating life experience in development, consistent with Somjen's memory theory proposed to account for missing error signals (Somjen, 1992).

Several lines of evidence support lifetime adjustments to the exercise ventilatory response. First, lung mechanics and gas exchange characteristics degrade with age, increasing physiological dead space and V_D/V_T. Despite this challenge, elderly individuals increase the

slope of the relationship between \dot{V}_E and \dot{V}_{CO_2} as required (Equation 2) to regulate P_{aCO_2} from rest to exercise with similar precision to young adults (Johnson et al., 1994). Second, although adults with congestive heart failure also exhibit an increased V_D/V_T due to widening ventilation–perfusion inequality, they too augment their exercise ventilatory responses to preserve P_{aCO_2} regulation (Sullivan et al., 1988). Third, in carotid denervated ponies, hyperventilation during work transitions is less 1–2 years post-surgery versus 2–4 weeks post-surgery (Pan et al., 1998). Mechanisms of these adjustments have never been formally addressed, although similar regulation occurs in other conditions that increase resting ventilatory drive, such as serotonin depletion (Mitchell et al., 1984), peripheral chemoreceptor stimulation via domperidone (Schaefer & Mitchell, 1989) or increased respiratory dead space (Mitchell, 1990). The uniformity of these adjustments points to an adaptive neural control system capable of modulation and/or plasticity (Mitchell et al., 1990; Priban & Fincham, 1965)

Influenced by engineering principles of systems control theory (Francis & Wonham, 1975), the internal model paradigm described by Poon et al. (2007) posits that the respiratory controller continuously integrates sensorimotor (afferent and efferent) signals during exercise and adapts according to an ‘optimality principle’. Whilst the ‘self-tuning’ hypothesis is difficult to test experimentally, it does allude to modulation and plasticity in the neural system controlling breathing. Demonstrated examples of adaptive control in exercise hyperpnoea includes ‘short-term modulation’ (STM) and ‘long-term modulation’ (LTM) of the exercise ventilatory response (Bach et al., 1993; Martin & Mitchell, 1993; Mitchell, 1990; Mitchell et al., 1984; Mitchell et al., 1990; Wood et al., 2003; Wood et al., 2008).

3.3.1 | Defining neuromodulation and neuroplasticity in the exercise ventilatory response

Inclusive definitions of modulation and plasticity in respiratory control can be found elsewhere (Babb et al., 2010; Mitchell & Babb, 2006; Mitchell & Johnson, 2003). Briefly, modulation is a change in the control system that occurs when the initiating stimulus is present, but reverses rapidly once that stimulus is removed (minutes). Plasticity, on the other hand, is a change that outlasts the initiating stimulus, persisting hours, days or even a lifetime after the stimulus has ended. With these simple definitions in mind, STM is an immediate change in the exercise ventilatory response that reverses when the inducing stimulus is removed. In contrast, LTM is a persistent change in the exercise ventilatory response beyond the duration of the inducing stimulus. Thus, the term ‘LTM’ as originally defined is somewhat misleading since it is an example of neuroplasticity. Confusion in definitions arose because the field was not welcoming to the concept of plasticity at the time LTM was first described (Martin & Mitchell, 1993), and definitions of modulation and plasticity in respiratory control had not yet been formalised (Mitchell & Johnson, 2003). The effects of STM and LTM on $\dot{V}_E:\dot{V}_{CO_2}$ coupling during exercise are illustrated conceptually in Figure 4.

3.3.2 | Short-term modulation of the exercise ventilatory response

Short-term modulation can be demonstrated in goats and humans with acute presentation of external dead space (Mitchell, 1990; Wood et al., 2008). Increased dead space using a wide bore tube with hyperoxic background conditions represents a CO_2 load with unique properties in comparison with increasing inspired CO_2 concentrations (Mitchell & Osborne, 1980). In goats, increased dead space (200–600 ml) increases V_D/V_T , reduces \dot{V}_A and increases resting P_{aCO_2} (~1 to 4 mmHg). With mild or moderate treadmill exercise, P_{aCO_2} is regulated slightly below (~2 mmHg) this new baseline level through increased system gain (Mitchell, 1990). Increased system gain was partitioned into the feedforward exercise (G_{EX} , Equation 6) versus feedback gain ($S = \Delta\dot{V}_E/\Delta P_{aCO_2}$) in accordance with a simple mathematical model (Mitchell, 1990):

$$G_{EX} = \left(\frac{\Delta\dot{V}_E}{\Delta\dot{V}_{CO_2}} \right) - S \times \left(\frac{\Delta P_{aCO_2}}{\Delta\dot{V}_{CO_2}} \right) \quad (6)$$

In this model, increased system gain ($G_{SYS} = \Delta\dot{V}_E/\Delta\dot{V}_{CO_2}$) with increased dead space could be accounted for by increased G_{EX} with unchanging S . Since G_{SYS} increased within a single exercise trial and was reversed in subsequent trials exercise without dead space, G_{EX} was increased by STM.

In healthy young men, a similar increase in G_{SYS} occurs during mild exercise with added dead space, regulating end-tidal P_{CO_2} with the same precision from rest to exercise at its new elevated resting level (Wood et al., 2008, 2009). In subsequent studies, STM was reported in elderly men (Wood et al., 2010), younger and elderly women (Wood et al., 2011), and people with obesity and obstructive sleep apnoea (Bernhardt et al., 2017). Thus, external dead space (and increased V_D/V_T) elicits STM, possibly accounting for the increased exercise ventilatory responses characteristic of old age and heart failure (see above).

After thoracic dorsal rhizotomy from T2–T12, goats are no longer able to compensate for increased respiratory dead space during exercise with STM, at least initially (Mitchell et al., 1990). With a respiratory mask in place, treadmill-trained goats exhibit overt ventilatory failure and hypercapnia, although the magnitude of failure diminished with each successive exercise trial. Strikingly, goats were once again able to generate a normal exercise hyperpnoea after multiple exercise trials. Thus, although these data may suggest some inability to elicit STM, another mechanism intervenes and restores function. Gradual restoration of exercise hyperpnoea with repeated exercise trials inspired studies that led to the discovery of LTM (Martin & Mitchell, 1993). In *post mortem* studies on the goats with thoracic dorsal rhizotomy, it was verified that regeneration of thoracic sensory afferent neurons could not be observed. Nevertheless, elevations in cervical and thoracic monoamine concentrations (e.g., serotonin, noradrenaline and dopamine) (Mitchell et al., 2000) led to further exploration of mechanisms giving rise to functional recovery. In rats, cervical dorsal rhizotomy increases serotonin (Kinkead et al., 1998)

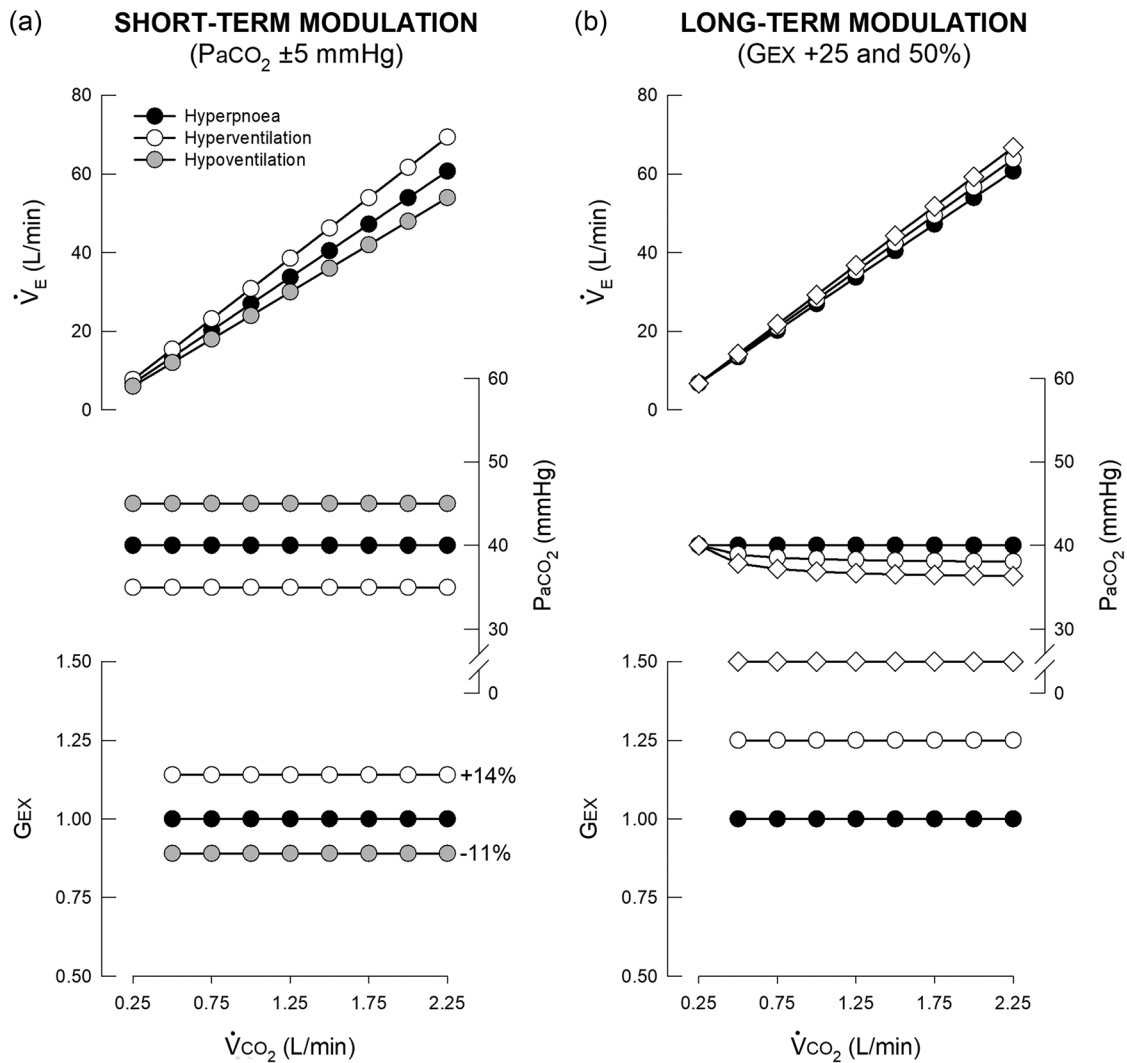


FIGURE 4 Short- and long-term modulation of the exercise ventilatory response. Black symbols represent an isocapnic hyperpnoea, white symbols represent hyperventilation, and grey symbols represent hypoventilation at rest. (a) Required (active) changes in the feedforward exercise gain (G_{EX}) and the resulting changes in system gain ($\dot{V}_E:\dot{V}_{CO_2}$ slope) to regulate arterial CO₂ pressure (P_{aCO_2}) with the same precision from rest to exercise at different P_{aCO_2} set-points. To regulate P_{aCO_2} at 45 mmHg, G_{EX} must increase by 14%. To regulate P_{aCO_2} at 35 mmHg, G_{EX} must decrease by 11% from the value required to regulate P_{aCO_2} at 40 mmHg. Changes in the P_{aCO_2} set-point and immediate (appropriate) changes in G_{EX} to enable similar P_{aCO_2} regulation demonstrate short-term modulation. (b) A 25% and 50% increase in G_{EX} with corresponding changes in the $\dot{V}_E:\dot{V}_{CO_2}$ slope of +5% and +10%, respectively. The former is identical to Figure 1c, whereby CO₂ chemofeedback layers onto the primary feedforward exercise stimulus to produce only a ~2 mmHg decrease in P_{aCO_2} from rest at a \dot{V}_{CO_2} of 2.25 L/min. The latter represents an additional layer of neuroplasticity (i.e., long-term modulation), denoted by diamonds, causing a ~4 mmHg decrease in P_{aCO_2} from rest. \dot{V}_{CO_2} , rate of CO₂ production; \dot{V}_E , minute ventilation.

and neurotrophic factors (Johnson et al., 2000), and enhances phrenic motor plasticity following acute intermittent hypoxia (Kinkead et al., 1998). These results are consistent with the idea that monoamines (e.g., serotonin) play key roles in respiratory motor plasticity and contribute to functional recovery of exercise hyperpnoea after thoracic sensory denervation.

In concordance, systemic administration of a broad-spectrum serotonin receptor antagonist (methysergide) abolishes STM with increased dead space in goats (Bach et al., 1993). Since methysergide and a more selective serotonin 5-HT_{2A} receptor antagonist, ketanserin, delivered to the thoracic cerebrospinal fluid via an indwelling catheter

abolish STM (Mitchell et al., 2008), spinal serotonin receptor activation is necessary for STM.

3.3.3 | Long-term modulation of the exercise ventilatory response

Long-term modulation occurs in goats and humans in response to repeated trials of combined exercise with CO₂ loading (dead space or inspired CO₂). After repeated trials of hypercapnic exercise, LTM manifests as profound hyperventilation during initial exercise trials

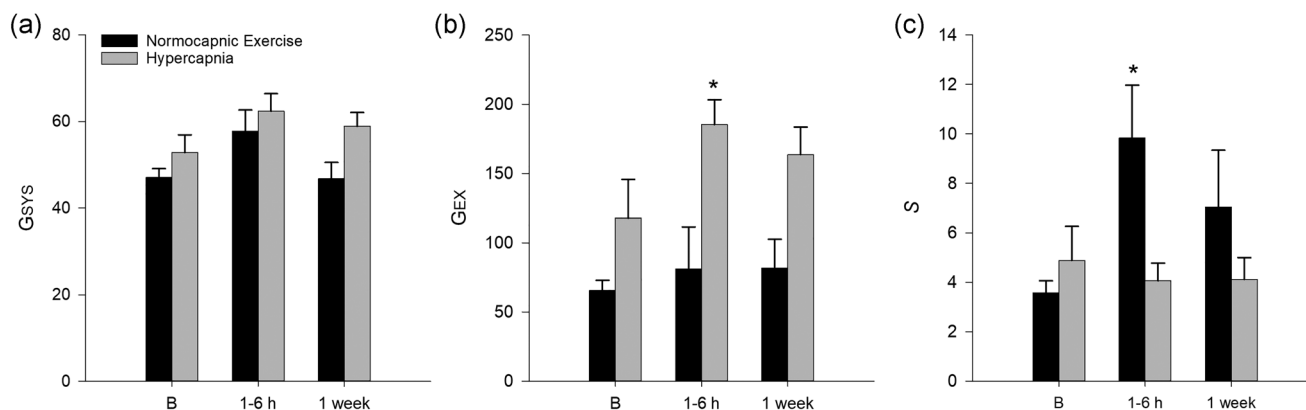


FIGURE 5 Long-term modulation of the exercise ventilatory response in goats. Changes in system gain ($G_{SYS} = \Delta\dot{V}_E/\Delta\dot{V}_{CO_2}$; a), feedforward gain ($G_{EX} = [(\Delta\dot{V}_E/\Delta\dot{V}_{CO_2}) - S(\Delta P_{aCO_2}/\Delta\dot{V}_{CO_2})]$; b), and feedback gain ($S = \Delta\dot{V}_E/\Delta P_{aCO_2}$; c) are shown during moderate exercise (4 km/h, 5% incline) in goats after 14–16 trials of normocapnic exercise training (black bars) and 14–16 trials of hypercapnic training (grey bars). Time points are baseline (B), 1–6 h post-training and 1 week post-training. These data demonstrate that G_{EX} is increased to some extent after hypercapnic training at rest, but not after repeated exercise without hypercapnia (b). Reproduced from data published only in abstract form (Martin & Mitchell, 1992; Martin et al., 1993).

without CO_2 loading (Martin & Mitchell, 1993; Wood et al., 2003). However, with subsequent (unloaded) trials, the exercise ventilatory response returns to normal, demonstrating a lasting but reversible form of learning or memory in the exercise ventilatory response.

Although LTM is induced by repeated exercise paired with CO_2 loading, it is not observed following repeated bouts of normocapnic (Martin & Mitchell, 1992) or hypoxic exercise (Turner et al., 1995). On the other hand, repeated CO_2 loading at rest (without exercise) partially elicits LTM (Martin et al., 1993), as shown in Figure 5. The capacity for LTM after repeated hypercapnia at rest, and the failure of repeated hypoxic exercise to elicit LTM, are perplexing since they suggest that the explicit pairing of hypercapnia with exercise, and carotid body chemoreceptor feedback, are not necessary for full LTM expression.

Attempts to replicate LTM in humans are somewhat inconsistent. Whilst some studies report LTM in phase I (Helbling et al., 1997; Turner & Stewart, 2004; Turner & Summers, 2002) and/or phases II or III (Reed & Coates, 2001; Wood et al., 2003), others do not (Al-Harbi et al., 2001; Cathcart et al., 2005; Cathcart et al., 2010; Moosavi et al., 2002; Summers & Turner, 2003). Many factors could explain the discrepancy in findings, particularly the number and duration of training trials, severity of hypercapnia, mode of inducing hypercapnia (e.g., dead space vs. inspired CO_2), participant training status, and history of physical activity. Notably, no human investigation to date directly measured P_{aCO_2} or respiratory mechanics, and most fail to report the slope of the $\dot{V}_E:\dot{V}_{CO_2}$ relationship (i.e., G_{SYS}) or G_{EX} . Thus, conditions necessary for LTM in humans are unclear. Other open questions are the need for explicit association of hypercapnia and exercise (Turner & Summers, 2002), and the task specificity of exercise training (Summers & Turner, 2003). In the most robust protocol to date (Wood et al., 2003), demonstration of LTM was compelling since it presented the highest number of hypercapnic exercise trials ($n = 70$) among published studies. An identical protocol was used to demonstrate learning in the exercise cardiovascular response. Repeated associations of

moderate exercise with neck suction to activate arterial baroreceptors attenuated increases in systolic blood pressure during subsequent exercise without neck suction for up to 2 days post-training (Herigstad et al., 2007).

As with STM, serotonin is believed to play a key role in LTM. In goats with serotonin depletion due to inhibition of the rate limiting enzyme in serotonin biosynthesis, tryptophan hydroxylase, LTM is no longer observed (Johnson & Mitchell, 2001). Hypercapnia (particularly when combined with exercise) activates brainstem raphe serotonergic neurons, releasing serotonin in the spinal cord. In a single exercise trial, serotonin release triggers STM. With repeated spinal serotonin release, LTM is expected. By way of analogy in anaesthetised rats, repeated cervical spinal serotonin receptor activation elicits long-term facilitation of phrenic motor neuron activity (MacFarlane & Mitchell, 2009; MacFarlane et al., 2011). Thus, STM may transition to LTM after repeated trials (and serotonin receptor activation).

In our working model, spinal serotonin release and 5-HT₂ receptor binding on respiratory motor neurons activates intracellular kinases that increase motor neuron excitability by closing potassium channels (i.e., STM) and/or phosphorylation of glutamate receptors that strengthen synaptic inputs onto respiratory motor neurons (i.e., LTM). Increased excitability potentiates motor neuron output for the same descending respiratory drive, augmenting \dot{V}_E at rest and during exercise. Reports that serotonin depletion by tryptophan hydroxylase inhibition with *p*-chlorophenylalanine abolishes both STM (Bach et al., 1993) and LTM (Johnson & Mitchell, 2001) suggest a common mechanism. How repeated serotonin receptor activation engages this mechanism is unknown, but may be akin to the transition from phrenic long-term facilitation (i.e., plasticity) to enhanced phrenic long-term facilitation (i.e., metaplasticity) when rats are exposed to acute intermittent hypoxia on consecutive days (Fields & Mitchell, 2015).

In summary, the exercise ventilatory response exhibits considerable modulation and plasticity—features infrequently considered in earlier

discussions of exercise hyperpnoea. Below, we address the exercise hyperpnoea dilemma, reasoning that the primary feedforward exercise stimulus is under constant adaptive control, and that mechanisms of learning, memory and plasticity may explain the near-perfect P_{aCO_2} regulation during moderate exercise.

4 | PART III: THE EXERCISE HYPERPNOEA DILEMMA

Major challenges must be met before we can solve the exercise hyperpnoea dilemma, including: (a) identifying factors that drive exercise hyperpnoea and accurately 'calibrate' \dot{V}_A across a wide \dot{V}_{CO_2} range; (b) generating unequivocal evidence that these factors are both necessary and sufficient for a normal exercise hyperpnoea in intact animal models or humans; and (c) differentiating between the primary exercise stimulus versus compensatory mechanisms that substitute when the primary stimulus is impaired (referred to as 'redundancy').

Notwithstanding the challenges listed above and the long history of attempts to solve the exercise hyperpnoea dilemma, there is reason for optimism since exceptional strides have been made in the field of neuroscience over the past few decades and concepts of modulation and plasticity are now established in the neural control of breathing (Feldman et al., 2003; Mitchell & Baker, 2022; Vose et al., 2022). Merging new knowledge with technical neuroscience innovations creates exciting new opportunities to probe specific neural substrates, cells and circuits that drive (or at least contribute to) exercise hyperpnoea.

4.1 | A new perspective on exercise hyperpnoea

A principal aim of this review is to reframe our approach to understanding exercise hyperpnoea. We propose that adaptive control (i.e., plasticity/learning) adjusts exercise hyperpnoea so that it remains appropriate in the face of ever-changing physiological conditions from birth to the end of life. The goal of an adaptive exercise hyperpnoea is to regulate P_{aCO_2} with the same precision from rest to exercise despite unavoidable changes in intrinsic respiratory system characteristics, such as lung compliance or pulmonary gas exchange that accompanies development and ageing. Other disturbances that challenge the accuracy of exercise hyperpnoea include permanent or reversible changes in body weight, diet, pregnancy and changes in ambient conditions (e.g., high altitude).

As a corollary, we posit that modulation and plasticity compensate for the onset of lung (e.g., asthma) or neural injury/diseases (e.g., spinal cord injury, ALS) that impair breathing function. Hence, increased understanding of how respiratory modulation and/or plasticity influences the exercise ventilatory response across the lifespan is of paramount significance from a translational perspective, particularly as it pertains to the development of novel therapeutics to preserve/restore breathing in clinical disorders that cause exercise intolerance due to respiratory compromise. Although

speculative, the hypotheses presented herein are put forth in the hope they will stimulate original investigations into the nature and mechanisms of exercise hyperpnoea—the largest (yet still unexplained) ventilatory response experienced by all individuals during normal life.

4.2 | Learned respiratory behaviours

Exercise hyperpnoea, like other homeostatic regulatory behaviours, may gradually accumulate throughout development in response to repeated error signals (Somjen, 1992). In the newborn, CO_2 chemoreception is already intact, although it can take a week or more to come 'online' in some species (Davis et al., 2006; Stunden et al., 2001; Wickstom et al., 1999). Thus, repeated CO_2 perturbations associated with physical activity may trigger neuroplasticity, adjusting system responses to meet changing needs. Once established, neuroplasticity can once again adjust the exercise ventilatory response if new error signals indicate the need for further system refinement.

If physical activity is associated with repeated error signals near birth, it may evoke neuroplasticity (i.e., learning) and guide the development of (or refine) exercise hyperpnoea. However, bilateral carotid denervation in neonatal goats (1–3 days of age) does not impact the adult (3–18 months of age) exercise ventilatory response (Lowry et al., 1999), casting some doubt on the developmental hypothesis. Nevertheless, the reliance of LTM on CO_2 chemoreception (Mitchell et al., 2008; Turner & Mitchell, 1997; Turner et al., 1995) versus other forms of respiratory motor plasticity that rely on O_2 chemoreception (Feldman et al., 2003; Mitchell & Baker, 2022) suggests that this question has not been adequately tested.

A prominent model to study learned respiratory behaviours is the gill withdrawal reflex of the marine invertebrate *Aplysia californica*, whereby many forms of learning have been identified, such as sensitization, habituation and classical conditioning (Walters et al., 1979, 1982). With classical conditioning, learning results from the explicit association of a conditioned stimulus with the unconditioned stimulus (Pavlov, 1927). In *Aplysia*, the conditioned stimulus (siphon touch) elicits only weak siphon and gill withdrawal when presented alone. However, when paired with an aversive (unconditioned) stimulus such as a tail shock, the response to siphon touch is sensitised. If repeated, paired associations are presented (~3 to 15 trials), long-lasting memory is formed (Carew et al., 1981; Carew et al., 1983). Similar to respiratory motor plasticity in vertebrates (e.g., phrenic long-term facilitation and LTM), serotonin mediates the classically conditioned gill withdrawal reflex in *Aplysia* (Glanzman et al., 1989).

In unanaesthetised cats, Orem and Netick (1986) paired an auditory tone (conditioned stimulus) with a noxious respiratory stimulus (ammonia inhalation; unconditioned stimulus), which terminates inspiration and prolongs expiration. When the conditioned and unconditioned stimuli were repeatedly paired, the conditioned tone triggers apnoea, demonstrating a form of associative learning and memory in the neural circuits controlling breathing. In humans, Gallego and Perruchet (1991) showed that eight pairings of an auditory tone

(conditioned stimulus) with hypoxia (unconditioned stimulus) for 12 breath cycles elicits a conditioned hyperventilation. Based upon this evidence for classical, associative conditioning of respiratory motor behaviours, it is worthwhile to contemplate what is known about associative conditioning mechanisms in non-respiratory motor behaviours, such as the conditioned eye-blink response in rabbits. In this model of associative motor conditioning, the cerebellum has been implicated as a key site of memory engram storage (Krupa et al., 1993; McCormick et al., 1981).

4.3 | Could a cerebellar memory engram encode the hyperpnoea of exercise?

In their comprehensive review of respiratory control during exercise, Forster and colleagues stated, 'To our knowledge it has not been postulated nor is there rationale or evidence to suggest that cerebellar nuclei are the primary source of the stimulus for the exercise hyperpnoea' (Forster et al., 2012). On the other hand, there is also no evidence to rule out a salient cerebellar role since research on this topic has been minimal. Here, we outline evidence that the cerebellum is more than a site of sensorimotor integration and coordination, as it has well-documented roles in multiple forms of motor learning. Exercise hyperpnoea is a complex motor behaviour that requires contributions from many well-coordinated respiratory muscles. As a highly orchestrated motor behaviour modified by neuroplasticity, there is sufficient evidence to consider that the memory engram for exercise hyperpnoea at least partially resides within the cerebellum. For the purposes of this review, an engram is defined as physical and/or chemical changes elicited by learning that underlie newly formed memory associations (Josselyn & Tonegawa, 2020).

4.3.1 | Role of the cerebellum in motor learning

The cerebellum contains approximately 69 billion neurons—over 80% of all neurons in the brain (Azevedo et al., 2009). The cerebellum receives inputs carrying information about outgoing motor commands from the motor cortex and brainstem, as well as sensory information via the brainstem and spinal cord (Brodal, 1981). These anatomical connections exhibit functional topography (Stoodley & Schmahmann, 2018). Thus, the cerebellum is an ideal network for sensorimotor learning (Brindley, 1964; Hansel et al., 2001) and functions as an 'array of adjustable motor pattern generators' that stores and retrieves learned motor behaviours (Houk, 1987).

Evidence that the cerebellum is critical for normal motor function and the formation of motor memories is shown by individuals with cerebellar degeneration that have persistent deficits in visuomotor learning (Martin et al., 1996), speech adaptation (Parrell et al., 2017) and locomotor adaptation (Morton & Bastian, 2006). These deficits arise from impaired sensorimotor memory formation (Hadjiiosif et al., 2023). Although important, such observations do not shed light on mechanisms of cerebellum-dependent motor learning.

Guided by detailed study of cerebellar neural networks by Eccles et al. (1967), Marr (1969) and Albus (1971) recognised that cerebellar synaptic connections are modified by experience. The crux of the Marr–Albus model of associative cerebellar memory is that parallel fibre-to-Purkinje cell synapses undergo long-lasting changes due to concurrent climbing fibre activation, which serves as the 'teacher' for new motor behaviours through error signal detection. An excellent example of cerebellar associative memory is the conditioned eye-blink response in rabbits (Houk et al., 1996; Krupa et al., 1993; McCormick et al., 1981).

Conditioned eye-blink responses occur when a conditioned stimulus (e.g., audio tone) is repeatedly paired with an aversive, unconditioned stimulus (e.g., air-puff into the eye). After explicitly paired presentations with a specific time interval between the conditioned and unconditioned stimuli, the conditioned stimulus subsequently evokes the conditioned response (i.e., eye-blink). Cells that comprise the engram for this simple motor memory are localised in the cerebellar interpositus nucleus—a site that is both necessary and sufficient for classically conditioned eye-blink responses (Christian & Thompson, 2003).

Recent experiments established that cerebellar Purkinje cells encode signals associated with the predicted sensory consequences of movement versus actual movement (Pasalar et al., 2006; Popa et al., 2012). Sensory prediction errors occur if there is a mismatch between the predicted sensory consequence of movement computed by the feedforward controller and actual sensory feedback. Brooks et al. (2015) recorded neural responses of the rostral fastigial nucleus in two adult rhesus monkeys during voluntary head movements with unexpected perturbations (resistive torque). The sensitivities (i.e., response gain) of rostral fastigial nucleus neurons dynamically tracked the initial introduction of error signals and its subsequent decline during motor learning, mirroring responses of the vestibular nuclei. These results elegantly demonstrate that deep cerebellar nuclei learn to expect the unexpected, ensuring that movements remain accurately calibrated over time.

4.3.2 | Role of the cerebellum in respiratory control

The cerebellum has elaborate direct and indirect projections to regions relevant to respiratory motor control (Krohn et al., 2023), including the nucleus of the solitary tract (Andrezik et al., 1984), rostral ventral respiratory group (Gaytan & Pasaro, 1998), trigeminal motor nucleus (Judd et al., 2021), facial nucleus (Moolenaar & Rucker, 1976), hypoglossal motor nucleus (Guo et al., 2020), Kölliker–Fuse nucleus (Fujita et al., 2020), locus coeruleus (Teune et al., 2000), dorsal raphe nucleus (Cavdar et al., 2018), periaqueductal grey (Anand et al., 1959; Frontera et al., 2020; Koutsikou et al., 2014) and hypothalamus (Dietrichs et al., 1994). Although the cerebellum's role in respiratory control is under-studied, it is known that deep cerebellar nuclei modulate breathing (Xu & Frazier, 1997, 2002), and are involved in certain forms of respiratory motor plasticity. For example, phrenic long-term facilitation induced by acute intermittent hypoxia is obliterated by

removing the cerebellar vermis (i.e., the part of the spinocerebellum containing the rostral fastigial nucleus) (Hayashi et al., 1993). As phrenic long-term facilitation is cerebellar and serotonin-dependent (Feldman et al., 2003; Mitchell et al., 2001), it may share common features with serotonin-dependent LTM (Mitchell & Babb, 2006; Mitchell et al., 2008).

Electrical stimulation of the rostral fastigial nucleus in anaesthetised cats increases respiratory frequency and/or amplitude (Bassal & Bianchi, 1982). Similarly, tachypnoea is observed during electrical stimulation within close proximity of the rostral fastigial nucleus in humans (Hirano et al., 1993). The rostral fastigial nucleus receives and integrates information from respiratory afferent neurons, including chemoreceptors (Lutherer et al., 1989). Additional evidence suggests that the rostral fastigial nucleus is CO₂ sensitive, as demonstrated by increased respiratory amplitude after localised acetazolamide micro-injections in rats (Xu et al., 2001).

There are data to suggest that the rostral fastigial nucleus may also play a role in the neural control of breathing during exercise. In goats, bilateral neurotoxic lesions of the rostral and caudal fastigial nucleus attenuates the normal exercise-induced hypocapnia by 1.3–2.8 mmHg during mild steady-state treadmill exercise (Martino et al., 2006). In these experiments, only ~55% of fastigial neurons were destroyed, and post-lesion studies were delayed by more than 1 week when the goats had recovered walking ability. Two important considerations when interpreting these findings are: (a) elimination of exercise-induced hypocapnia (~2 mmHg) represents a nearly 25% reduction in the exercise feedforward stimulus once the inhibitory role of chemofeedback is accounted for (Mitchell, 1990); and (b) since ~50% of fastigial neurons were lesioned, spared neurons may have compensated, similar to respiratory motor neuron compensation in ALS (Seven & Mitchell, 2019). Thus, cerebellar contributions to exercise hyperpnoea may have been underestimated. In humans undergoing functional imaging, the cerebellum (and possibly the fastigial nucleus) increases activity during active inspiration (Colebatch et al., 1991), active expiration (Ramsay et al., 1993), imagined exercise (Thornton et al., 2001), voluntary hyperpnoea (McKay et al., 2003) and asphyxia (McKay et al., 2010).

Collectively, these studies give credence to the possibility of a major cerebellar role in exercise hyperpnoea, particularly any learned component. The cerebellum is important for somatic motor learning (as envisioned by Marr and Albus) and discrete areas of the cerebellum (e.g., fastigial nucleus) are clearly engaged in the control of breathing. However, the specific role of the cerebellum in acquisition or storage of a learned exercise hyperpnoea has never been formally tested to our knowledge.

4.4 | Working hypothesis

Here, we pose the hypothesis that cerebellar learning is obligatory for normal development and maintenance of exercise hyperpnoea in humans and other mammals. Our hypothesis is largely based on demonstrations that exercise hyperpnoea exhibits plasticity in adult

mammals (Martin & Mitchell, 1993; Mitchell et al., 1990), that the cerebellum contributes to respiratory control, including forms of respiratory 'memory' (Hayashi et al., 1993), and that deep cerebellar nuclei are crucial in other forms of motor learning (Clark et al., 1992; Krupa & Thompson, 1995, 1997; Krupa et al., 1993). For example, the cerebellar interpositus nucleus is central to both the acquisition and retention of conditioned eye-blink responses (Krupa et al., 1993). Once acquired, the conditioned response is eliminated and then restored by reversibly inhibiting nucleus interpositus neurons. In contrast, inactivation of relevant brainstem motor nuclei also prevents the unconditioned reflex, without impairing acquisition of the conditioned response (Krupa et al., 1996). Thus, strong evidence supports the stance that the engram (vs. unconditioned reflex) is stored in the cerebellar interpositus nucleus. Similar approaches using more contemporary techniques to activate/inactivate target neurons without impacting off-target neurons can be used to investigate cerebellar contributions to exercise hyperpnoea.

Plasticity in the exercise ventilatory response has been shown in previous studies using repeated associations of external dead space or elevated inspired CO₂ to elicit hypercapnia (unconditioned stimulus) with moderate exercise (conditioned stimulus) (Martin & Mitchell, 1993; Wood et al., 2003). This model of associative learning in exercise hyperpnoea may be useful to test suspected cerebellar engram locations. Powerful chemogenetic and optogenetic techniques are now available to manipulate specific neural pathways of interest and some have already begun to apply such methods to the control of breathing during exercise (Herent et al., 2023; Korsak et al., 2018). Thus, it is possible to test if a given site is sufficient (e.g., excitatory designer receptors activated by designer drugs, DREADDs) and necessary (e.g., inhibitory DREADDs) to account for exercise hyperpnoea. For instance, inactivation of the inferior olivary nucleus, climbing fibres, Purkinje cells or the rostral fastigial nucleus during versus after associative training may prevent (or minimise) the feedforward exercise stimulus, leaving residual chemofeedback-driven ventilation characterised by elevated P_{aCO_2} from rest to exercise. Alternatively (or in addition), inhibition of these neurons may prevent the ability to acquire LTM. If inactivation of cerebellar circuits prevents LTM, activation of those same circuits should (a) increase \dot{V}_E substantially at rest, driving severe hypocapnia, and/or (b) elicit LTM if paired with exercise without CO₂ loading. Long-term modulation would be expressed as a reversibly enhanced feedforward exercise gain and exacerbated exercise hypocapnia.

Figure 6 is a simplified diagram of cerebellar neural networks depicting our working model. We propose that feedforward inputs activate mossy fibres whereas changes in P_{aCO_2} activate climbing fibres. During hypercapnic exercise, mossy and climbing fibres are activated in close temporal association. Deep cerebellar nuclei subsequently modulate breathing during exercise. Inhibitory chemogenetics targeting Purkinje cells and/or neurons in the rostral fastigial nucleus as the site of plasticity would theoretically prevent learning (i.e., acquisition of LTM), or even block the exercise hyperpnoea if it represents a learned response from birth with the engram stored at this site. Optogenetics may also be useful to target inferior

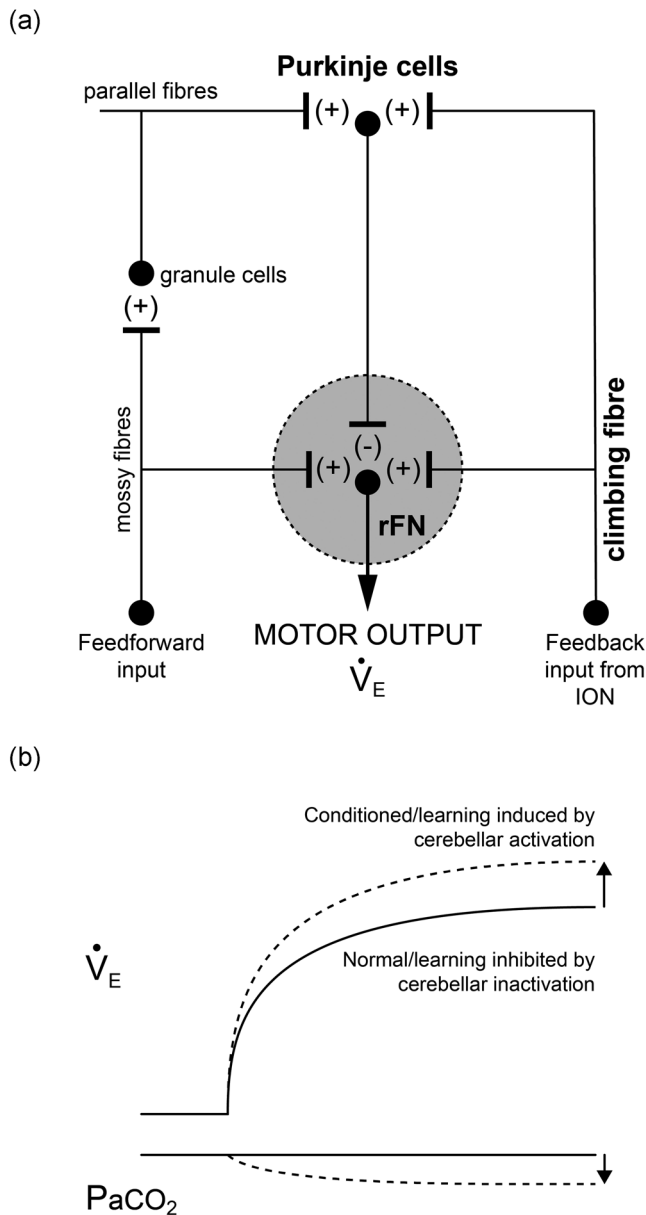


FIGURE 6 Hypothesis. (a) A diagram of a cerebellar neural network including a possible site of plasticity at the climbing fibre-to-Purkinje cell synapse. (b) A normal ventilatory response to mild or moderate steady-state exercise with good regulation of arterial P_{aCO_2} (P_{aCO_2} ; unbroken lines). Repeated trials of hypercapnic training elicit hyperventilation during subsequent exercise trials without CO_2 loading due to associative learning (i.e., long-term modulation of the exercise ventilatory response; dashed lines). We hypothesise that learning can be inhibited by chemogenetic inactivation of Purkinje cells or the rostral fastigial nucleus (rFN) during conditioning or induced by optogenetic activation of the inferior olivary nucleus (ION) or the climbing fibre during exercise without need for CO_2 loading. See text for further details. \dot{V}_E , minute ventilation.

olive/climbing fibre activation (simulating error signals) to test their requirement for plasticity.

4.5 | Limitations and other considerations

Our hypotheses are highly speculative, but are presented in an effort to stimulate new research on a topic that has seemingly 'run out of good ideas'. We do not yet know: (a) if exercise hyperpnoea is an example of life-long learning versus a response modified by neuroplasticity; (b) if the cerebellum is implicated in LTM of the exercise ventilatory response; (c) where the engram for exercise hyperpnoea is stored and what cells types are involved; and (d) if the engram is an ensemble of widely distributed cells in multiple regions of the central nervous system, as with other forms of learning and memory, such as fear conditioning (Josselyn & Tonegawa, 2020).

5 | SUMMARY

Our understanding of the neural control of breathing continues to expand. Recent conceptual and technological breakthroughs may allow researchers, for the first time, to precisely target specific neural pathways associated with learning, memory and plasticity in automatic (respiratory) or autonomic (cardiovascular) responses to exercise. Explaining the mechanistic basis of exercise hyperpnoea remains a great challenge facing respiratory and exercise physiologists. Julius Comroe once stated '... respiratory physiologists are not necessarily good neurobiologists or good control system engineers, and maybe they must be to solve the problem' (Comroe, 1965). Nearly 60 years later, we echo this sentiment, and hope to reopen discussions that generate new ideas and renewed study of exercise hyperpnoea in the 21st century.

Our perspective is that the hyperpnoea of exercise is (continuously) adjusted throughout life, and may drive development of a normal exercise ventilatory response, or adapt a congenital, rudimentary feed-forward exercise stimulus so that it generates appropriate exercise hyperpnoea despite changing conditions. Relevant conditions that require adaptation include healthy ageing, weight gain/loss, injury, onset of lung disease, sojourn to high altitude and many others. Each of these experiences requires active adjustment (i.e., modulation and/or plasticity) of the neural system driving breathing, or the superb regulation of P_{aCO_2} typical of mild or moderate exercise will degrade, limiting human performance or even life. These ideas are not adequately tested and considerable research efforts are needed to determine if they have merit.

We speculate that the memory engram encoding the hyperpnoea of exercise resides within the cerebellum. Although there are certainly other possibilities, there is ample evidence from other automatic motor systems to strongly consider this possibility. Of great importance, these and alternate hypotheses can be tested using modern neuroscience methods (and concepts) that were unavailable to previous

generations of researchers interested in the exercise hyperpnoea dilemma (Swanson, 1978).

AUTHOR CONTRIBUTIONS

Both authors have read and approved the final version of this manuscript and agree to be accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. Both persons designated as authors qualify for authorship, and all those who qualify for authorship are listed.

CONFLICT OF INTEREST

The authors declare no conflicts of interest.

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REFERENCES

- Adams, L., Frankel, H. L., Garlick, J., Guz, A., Murphy, K., & Semple, S. J. G. (1984). The role of spinal cord transmission in the ventilatory response to exercise in man. *The Journal of Physiology*, 355, 85–97.
- Al-Harbi, W., Coates, J. C., & Reed, J. W. (2001). Associative conditioning of the exercise ventilatory response. *Respiration Physiology*, 2, P25.
- Alam, M., & Smirk, F. H. (1937). Observations in man upon a blood pressure raising reflex arising from the voluntary muscles. *The Journal of Physiology*, 89(4), 372–383.
- Albus, J. S. (1971). A theory of cerebellar function. *Mathematical Biosciences*, 10(1-2), 25–61.
- Amann, M., Bain, G. M., Proctor, L. T., Sebranek, J. J., Pegelow, D. F., & Dempsey, J. A. (2010). Group III and IV muscle afferents contribute to ventilatory and cardiovascular responses to rhythmic exercise in humans. *Journal of Applied Physiology*, 109(4), 966–976.
- Amann, M., Blain, G. M., Proctor, L. T., Sebranek, J. J., Pegelow, D. F., & Dempsey, J. A. (2011). Implications of group III and IV muscle afferents for high-intensity endurance exercise performance in humans. *The Journal of Physiology*, 589(Pt 21), 5299–5309.
- An, X., Bandler, R., Ongur, D., & Price, J. L. (1998). Prefrontal cortical projections to longitudinal columns in the midbrain periaqueductal grey in macaque monkeys. *Journal of Comparative Neurology*, 401(4), 455–479.
- Anand, B. K., Malhotra, C. L., Singh, B., & Dua, S. (1959). Cerebellar projections to limbic system. *Journal of Neuroscience*, 22(4), 451–457.
- Andreziak, J. A., Dormer, K. J., Foreman, R. D., & Person, R. J. (1984). Fastigial nucleus projections to the brain stem in beagles: Pathways for autonomic regulation. *Neuroscience*, 11, 497–507.
- Asmussen, E., Nielsen, M., & Wieth-Pedersen, G. (1943). Cortical or reflex control of respiration during muscular work? *Acta Physiologica*, 6, 168–175.
- Azevedo, F. A. C., Carvalho, L. R. B., Grinberg, L. T., Farfel, J. M., Ferretti, R. E. L., Leite, R. E. P., Filho, W. J., Lent, R., &erculano-Houzel, S. (2009). Equal numbers of neuronal and nonneuronal cells make the human brain an isometrically scaled-up primate brain. *Journal of Comparative Neurology*, 513(5), 532–541.
- Babb, T. G., Wood, H. E., & Mitchell, G. S. (2010). Short- and long-term modulation of the exercise ventilatory response. *Medicine and science in sports and exercise*, 52, 1681–1687.
- Bach, K. B., Lutcavage, M. E., & Mitchell, G. S. (1993). Serotonin is necessary for short-term modulation of the exercise ventilatory response. *Respiratory Physiology*, 91(1), 57–70.
- Bainton, C. R. (1972). Effect of speed vs. grade and shivering on ventilation in dogs during active exercise. *Journal of Applied Physiology*, 33, 778–787.
- Bainton, C. R. (1978). Canine ventilation after acid-base infusions, exercise, and carotid body denervation. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 44(1), 28–35.
- Bandler, R., & Carrive, P. (1988). Integrated defence reaction elicited by excitatory amino acid microinjection in the midbrain periaqueductal grey region of the unrestrained cat. *Brain Research*, 439, 95–106.
- Banner, N., Guz, A., Heaton, R., Innes, J. A., Murphy, K., & Yacoub, M. (1988). Ventilatory and circulatory responses at the onset of exercise in man following heart or heart-lung transplantation. *The Journal of Physiology*, 399, 437–449.
- Bartoli, A., Cross, B., Guz, A., Jain, S. K., Noble, M. I., & Trenchard, D. (1973). Proceedings: A vagal reflex modifying ventilation activated by CO₂ in the lungs. *The Journal of Physiology*, 234(2), 93P–94P.
- Bassal, M., & Bianchi, A. L. (1982). Inspiratory onset or termination induced by electrical stimulation. *Respiration Physiology*, 50(1), 23–40.
- Bennett, F. M., & Fordyce, W. E. (1985). Characteristics of the ventilatory exercise stimulus. *Respiration Physiology*, 59(1), 55–63.
- Bennett, F. M., & Fordyce, W. E. (1988). Gain of the ventilatory exercise stimulus: Definition and meaning. *Journal of Applied Physiology*, 66, 2011–2017.
- Bennett, F. M., & Fordyce, W. E. (1993). Regulation of PaCO₂ during rest and exercise: A modeling study. *Annals of Biomedical Engineering*, 21, 545–555.
- Bennett, F. M., Tallman, R. D. J., & Grodins, F. S. (1984). Role of VCO₂ in control of breathing of awake exercising dogs. *Journal of Applied Physiology*, 56(5), 1335–1339.
- Bernhardt, V., Mitchell, G. S., Lee, W. Y., & Babb, T. G. (2017). Short-term modulation of the ventilatory response to exercise is preserved in obstructive sleep apnea. *Respiratory Physiology and Neurobiology*, 236, 42–50.
- Brice, A. G., Forster, H. V., Pan, L. G., Funahashi, A., Hoffman, M. D., Murphy, C. L., & Lowry, T. F. (1988). Is the hyperpnoea of muscular contractions critically dependent on spinal afferents? *Journal of Applied Physiology*, 64, 226–233.
- Brindley, G. S. (1964). The use made by the cerebellum of the information that it receives from sense organs. *IRBO Bull.*, 3, 80.
- Brodal, A. (1981). *Neurological Anatomy*. Oxford University Press.
- Brooks, J. X., Carriot, J., & Cullen, K. E. (2015). Learning to expect the unexpected: Rapid updating in primate cerebellum during voluntary self-motion. *Nature Neuroscience*, 18(9), 1310–1317.
- Brown, D. R., Forster, H. V., Pan, L. G., Brice, A. G., Murphy, C. L., Lowry, T. F., Gutting, S. M., Funahashi, A., Hoffman, M. D., & Powers, S. K. (1990). Ventilatory response to spinal cord-lesioned subjects to electrically induced exercise. *Journal of Applied Physiology*, 68(6), 2312–2321.
- Bruce, R. M. (2022). Investigating the control of exercise hyperpnoea: A synergy of contributions. *Experimental Physiology*, 107(2), 103–105.
- Bruce, R. M., Jolley, C., & White, M. J. (2019). Control of exercise hyperpnoea: Contributions from thin-fibre skeletal muscle afferents. *Experimental Physiology*, 104(11), 1605–1621.
- Carew, T. J., Hawkins, R. D., & Kandel, E. R. (1983). Differential classical conditioning of a defensive withdrawal reflex in *Aplysia californica*. *Science*, 219(4583), 397–400.
- Carew, T. J., Walters, E. T., & Kandel, E. R. (1981). Classical conditioning in a simple withdrawal reflex in *Aplysia californica*. *Journal of Neuroscience*, 1(12), 1426–1437.
- Casaburi, R. (2012). The mechanism of the exercise hyperpnoea: The ultrasecret revisited. *American Journal of Respiratory and Critical Care Medicine*, 186, 578–579.
- Casaburi, R., Whipp, B. J., Wasserman, K., Beaver, W. L., & Koyal, S. N. (1977). Ventilatory and gas exchange dynamics in response to sinusoidal work. *Journal of Applied Physiology*, 42, 300–301.
- Cathcart, A. J., Herrold, N., Turner, A. P., Wilson, J., & Ward, S. A. (2005). Absence of long-term modulation of ventilation by dead-space loading during moderate exercise in humans. *European Journal of Applied Physiology*, 93(4), 411–420.

- Cathcart, A. J., Whipp, B. J., Turner, A. P., Wilson, J., & Ward, S. A. (2010). Influence of cycling history on the ventilatory response to cycle-ergometry in humans: A role for respiratory memory? *Advances in Experimental Medicine and Biology*, 669, 311–314.
- Cavdar, S., Ozgur, M., Kuvvet, Y., Bay, H., & Aydogmus, E. (2018). Cortical, subcortical and brain stem connections of the cerebellum via the superior and middle cerebellar peduncle in the rat. *Journal of Integrative Neuroscience*, 17(3-4), 609–618.
- Christian, K. M., & Thompson, R. F. (2003). Neural substrates of eyeblink conditioning: Acquisition and retention. *Learning & Memory*, 10, 427–455.
- Clark, J. M., Sinclair, R. D., & Lenox, J. B. (1980). Chemical and nonchemical components of ventilation during hypercapnic exercise in man. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 48(6), 1065–1076.
- Clark, R. E., Zhang, A. A., & Lavond, D. G. (1992). Reversible lesions of the cerebellar interpositus nucleus during acquisition and retention of a classically conditioned behavior. *Behavioral Neuroscience*, 106(6), 879–888.
- Colebatch, J. G., Adams, L., Murphy, K., Martin, A. J., Lammertsma, A. A., Tochon-Danguy, H. J., Clark, J. C., Friston, K. J., & Guz, A. (1991). Regional cerebral blood flow during volitional breathing in man. *The Journal of Physiology*, 443, 91–103.
- Coleridge, H. M., Coleridge, J. C., & Howe, A. (1966). Aorticopulmonary glomus tissue in the cat. *Nature*, 211(5054), 1187.
- Comroe, J. H. J. (1965). *Physiology of Respiration: An Introductory Text*. Year Book Medical Publishers.
- Comroe, J. H. J., & Schmidt, C. F. (1943). Reflexes from the limbs as a factor in hyperpnea of muscular exercise. *American Journal of Physiology*, 138, 536–547.
- Coote, J. H., Hilton, S. M., & Perez-Gonzalez, J. F. (1971). The reflex nature of the pressor response to muscular exercise. *The Journal of Physiology*, 215(3), 789–804.
- Cropp, G. J. A., & Comroe, J. H. J. (1961). Role of mixed venous blood PCO₂ in respiratory control. *Journal of Applied Physiology*, 16, 1029–1033.
- Cross, B. A., Davey, C. A., Guz, A., Katona, P. G., Maclean, M., Murphy, K., Semple, S. J. G., & Stidwill, R. (1982). The role of spinal cord transmission in the ventilatory response to electrically induced exercise in the anesthetized dog. *The Journal of Physiology*, 329, 37–55.
- Cunningham, D. J. C., Robbins, P. A., & Wolff, C. B. (1986). Integration of respiratory responses to changes in alveolar partial pressures of CO₂ and O₂ and in arterial pH. In A. P. Fishman, N. S. Cherniack, & J. G. Widdicombe (Eds.), *Handbook of Physiology* pp. 475–528. American Physiological Society.
- Davis, S. E., Solhied, G., Castillo, M., Dwinell, M. R., Brozoski, D., & Forster, H. V. (2006). Postnatal developmental changes in CO₂ sensitivity in rats. *Journal of Applied Physiology*, 101(4), 1097–1103.
- Dejours, P. (1962). Chemoreflexes in breathing. *Physiological Reviews*, 42, 335–358.
- Dejours, P. (1964). Control of respiration during muscular exercise. In *Handbook of Physiology*, pp. 631–648. American Physiological Society.
- Dejours, P., Mithoefer, J. C., & Teillac, A. (1955). Essai de mise en évidence de chemorecepteurs veineux de ventilation. *The Journal of Physiology*, 47(1), 160–163.
- Dempsey, J. A. (2006). Challenges for future research in exercise physiology as applied to the respiratory system. *Exercise and Sport Sciences Reviews*, 34(3), 92–98.
- Dempsey, J. A., Mitchell, G. S., & Smith, C. A. (1984). Exercise and chemoreception. *American Review of Respiratory Disease*, 129, S31–S34.
- Dempsey, J. A., Neder, J. A., Phillips, D. B., & O'Donnell, D. E. (2022). The physiology and pathophysiology of exercise hyperpnea. In R. Chen, & P. G. Guyenet (Eds.), *Handbook of Clinical Neurology* pp. 201–232.
- Dempsey, J. A., & Welch, J. F. (2023). Control of breathing. *Seminars in Respiratory and Critical Care Medicine*, 44, 627–649.
- Dietrichs, E., Haines, D. E., Roste, G. K., & Roste, L. S. (1994). Hypothalamocerebellar and cerebellohypothalamic projections—circuits for regulating nonsomatic cerebellar activity? *Histology and Histopathology*, 9(3), 603–614.
- Douglas, C. G., & Haldane, J. S. (1909). The regulation of normal breathing. *The Journal of Physiology*, 38(5), 420–440.
- Douglas, C. G., Haldane, J. S., Henderson, Y., & Schneider, E. C. (1912). The physiological effects of low atmospheric pressures, as observed on Pike's peak, Colorado. *Proceedings of the Royal Society of London. Series B: Biological Sciences*, 85, 65–67.
- Duke, H. N., Green, J. H., Heffron, P. F., & Stubbens, W. J. (1963). Pulmonary chemoreceptors. *Experimental Physiology*, 48, 164–175.
- Eccles, J., Ito, M., & Szentagothai, J. (1967). *The cerebellum as a neuronal machine*. Springer Verlag.
- Eldridge, F. L., Millhorn, D. E., Kiley, J. P., & Waldrop, T. G. (1985). Stimulation by central command of locomotion, respiration and circulation during exercise. *Respiration Physiology*, 59(3), 313–337.
- Eldridge, F. L., Millhorn, D. E., & Waldrop, T. G. (1981). Exercise hyperpnea and locomotion: Parallel activation from the hypothalamus. *Science*, 211(4484), 844–846.
- Feldman, J. L., Mitchell, G. S., & Nattie, E. E. (2003). Breathing: Rhythmicity, plasticity, chemosensitivity. *Annual Review of Neuroscience*, 26, 239–266.
- Fenn, W. O., Rahn, H., & Otis, A. B. (1946). A theoretical study of the composition of the alveolar air at altitude. *American Journal of Physiology*, 156, 637–653.
- Fields, D. P., & Mitchell, G. S. (2015). Spinal metaplasticity in respiratory motor control. *Frontiers in Neural Circuits*, 9, 2.
- Fink, G. R., Adams, L., Watson, J. D., Innes, J. A., Wuyam, B., Kobayashi, I., Corfield, D. R., Murphy, K., Jones, T., & Frackowiak, R. S. J. (1995). Hyperpnea during and immediately after exercise in man: Evidence of motor cortical involvement. *The Journal of Physiology*, 489, 663–675.
- Fordyce, W. E., Bennett, F. M., Edelman, S. K., & Grodins, F. S. (1982). Evidence in man for a fast neural mechanism during the early phase of exercise hyperpnea. *Respiration Physiology*, 48(1), 27–43.
- Fordyce, W. E., & Grodins, F. S. (1980). Ventilatory responses to intravenous and airway CO₂ administration in anesthetized dogs. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 48(2), 337–346.
- Forster, H. V., Haouzi, P., & Dempsey, J. A. (2012). Control of breathing during exercise. *Comprehensive Physiology*, 2, 743–777.
- Forster, H. V., Pan, L. G., Bisgard, G. E., Flynn, C., Dorsey, S. M., & Britton, M. S. (1984). Independence of exercise hypocapnia and limb movement frequency in ponies. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 57(6), 1885–1893.
- Forster, H. V., Pan, L. G., Bisgard, G. E., Kaminski, R. P., Dorsey, M., & Busch, M. A. (1983). Hyperpnea of exercise at various PIO₂ in normal and carotid body-denervated ponies. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 54, 1387–1393.
- Francis, B., & Wonham, W. (1975). The internal model principles for linear multivariable regulators. *Applied Mathematics & Optimization*, 2, 170–174.
- Frontera, J. L., Baba Aissa, H., Sala, R. W., Maihes-Hamon, C., Georgescu, I. A., Lena, C., & Popa, D. (2020). Bidirectional control of fear memories by cerebellar neurons projecting to the ventrolateral periaqueductal grey. *Nature Communications*, 11(1), 5207.
- Fujita, H., Kodama, T., & du Lac, S. (2020). Modular output circuits of the fastigial nucleus for diverse motor and nonmotor functions of the cerebellar vermis. *eLife*, 9, e58613.
- Fuller, D. D., & Mitchell, G. S. (2017). Respiratory neuroplasticity—overview, significance and future directions. *Experimental Neurology*, 287, 144–152.
- Gallego, J., & Perruchet, P. (1991). Classical conditioning of ventilatory responses in humans. *Journal of Applied Physiology*, 70(2), 676–682.
- Gaytan, S. P., & Pasaro, R. (1998). Connections of the rostral ventral respiratory neuronal cell group: an anterograde and retrograde tracing study in the rat. *Brain Research Bulletin*, 47(6), 625–642.

- Geppert, J., & Zuntz, N. (1888). Ueber die Regulation der Athmung. *Pflügers Archiv*, 42, 189–245.
- Glanzman, D. L., Mackey, S. L., Hawkins, R. D., Dyke, A. M., Lloyd, P. E., & Kandel, E. R. (1989). Depletion of serotonin in the nervous system of Aplysia reduces the behavioural enhancement of gill withdrawal as well as the heterosynaptic facilitation produced by tail shock. *Journal of Neuroscience*, 9(12), 4200–4213.
- Goodwin, G. M., McCloskey, D. I., & Mitchell, J. H. (1972). Cardiovascular and respiratory responses to changes in central command during isometric exercise at constant muscle tension. *The Journal of Physiology*, 226(1), 173–190.
- Green, A. L., & Paterson, D. J. (2008). Identification of neurocircuitry controlling cardiovascular function in humans using functional neurosurgery: Implications for exercise control. *Experimental Physiology*, 93(9), 1022–1028.
- Green, A. L., & Paterson, D. J. (2020). Using deep brain stimulation to unravel the mysteries of cardiorespiratory control. *Comprehensive Physiology*, 10, 1085–1104.
- Green, A. L., Wang, S., Purvis, S., Owen, S. L. F., Bain, P. G., Stein, J. F., Guz, A., Aziz, T. Z., & Paterson, D. J. (2007). Identifying cardiorespiratory neurocircuitry involved in central command during exercise in humans. *The Journal of Physiology*, 578, 605–612.
- Green, J. F., Schertel, E. R., Coleridge, H. M., & Coleridge, J. C. (1986). Effect of pulmonary arterial PCO₂ on slowly adapting pulmonary stretch receptors. *Journal of Applied Physiology*, 60, 2048–2055.
- Green, J. F., & Schmidt, N. D. (1984). Mechanism of hyperpnea induced by changes in pulmonary blood flow. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 56(5), 1418–1422.
- Green, J. F., & Sheldon, M. I. (1983). Ventilatory changes associated with changes in pulmonary blood flow in dogs. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 54(4), 997–1002.
- Grodins, F. S. (1950). Analysis of factors concerned in regulation of breathing in exercise. *Physiological Reviews*, 30(2), 220–239.
- Grodins, F. S. (1981). Exercise hyperpnea: The ultra secret. *Advances in Physiology Science*, 10, 243–251.
- Grodins, F. S., Buell, J., & Bart, A. J. (1967). Mathematical analysis and digital simulation of the respiratory control system. *Journal of Applied Physiology*, 22(2), 260–276.
- Grodins, F. S., Gray, J. S., Schroeder, K. R., Norins, A. L., & Jones, R. W. (1954). Respiratory responses to CO₂ inhalation: A theoretical study of a nonlinear biological regulator. *Journal of Applied Physiology*, 7(3), 283–308.
- Guo, H., Yuan, X. S., Zhou, J. C., Chen, H., Li, S. Q., Qu, W. M., & Huang, Z. L. (2020). Whole-brain monosynaptic inputs to hypoglossal motor neurons in mice. *Neuroscience Bulletin*, 36, 585–597.
- Guz, A. (1997). Brain, breathing and breathlessness. *Respiration Physiology*, 109, 197–204.
- Hadjiosif, A. M., Gibo, T. L., & Smith, M. A. (2023). Cerebellar damage impairs long-term but not short-term sensorimotor memory. *BioRxiv*, <https://doi.org/10.1101/2023.08.11.553008>
- Haldane, J. S., & Priestley, J. G. (1905). The regulation of the lung-ventilation. *The Journal of Physiology*, 32(3-4), 225–266.
- Hansel, C., Linden, D. J., & D'Angelo, E. (2001). Beyond parallel fiber LTD: The diversity of synaptic and non-synaptic plasticity in the cerebellum. *Nature Neuroscience*, 4(5), 467–475.
- Haouzi, P. (2014). Tracking pulmonary gas exchange by breathing control during exercise: Role of muscle blood flow. *The Journal of Physiology*, 592, 453–461.
- Haouzi, P. (2020). The ventilatory component of the muscle metaboreflex: Catch me if you can! *Experimental Physiology*, 105(12), 2246–2249.
- Haouzi, P., Cheunel, B., & Chalon, B. (2004). The control of ventilation is dissociated from locomotion during walking in sheep. *The Journal of Physiology*, 559, 315–325.
- Haouzi, P., Hill, J. M., Lewis, B. K., & Kaufman, M. P. (1999). Responses of group III and IV muscle afferents to distension of the peripheral vascular bed. *Journal of Applied Physiology*, 87(2), 545–553.
- Hayashi, F., Coles, S. K., Bach, K. B., Mitchell, G. S., & McCrimmon, D. R. (1993). Time-dependent phrenic nerve responses to carotid afferent activation: Intact vs. decerebrate rats. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 34, R811–R819.
- Helbling, D., Boutellier, U., & Spengler, C. M. (1997). Modulation of the ventilatory increase at the onset of exercise humans. *Respiration Physiology*, 109, 219–229.
- Herent, C., Diem, S., Usseglio, G., Fortin, G., & Bouvier, J. (2023). Upregulation of breathing rate during running exercise by central locomotor circuits in mice. *Nature Communications*, 14(1), 2939.
- Herigstad, M., Balanos, G. M., & Robbins, P. A. (2007). Can human cardiovascular regulation during exercise be learnt from feedback from arterial baroreceptors? *Experimental Physiology*, 92(4), 695–704.
- Hirano, T. K., Yoshida, H. K., Furuse, M., Taniguchi, K., & Inao, S. (1993). Fastigial pressor response observed during an operation on a patient with cerebellar bleeding. An anatomical review and clinical significance. *Neurosurgery*, 32(4), 675.
- Honda, Y., Myojo, S., Hasegawa, S., Hasegawa, T., & Severinghaus, J. W. (1979). Decreased exercise hyperpnea in patients with bilateral carotid chemoreceptor resection. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 46, 908–912.
- Horiuchi, J., McDowall, L. M., & Dampney, R. A. L. (2009). Vasomotor and respiratory responses evoked from the dorsolateral periaqueductal grey are mediated by the dorsomedial hypothalamus. *The Journal of Physiology*, 587(Pt 2), 5149–5162.
- Houk, J. C. (1987). Model of the cerebellum as an array of adjustable pattern generators. In M. Glickstein, C. Yeo, & J. Stein (Eds.), *Cerebellum and Neuronal Plasticity*. Springer.
- Houk, J. C. (1988). Control strategies in physiological systems. *Federation of American Societies for Experimental Biology Journal*, 2(2), 97–107.
- Houk, J. C., Buckingham, J. T., & Barto, A. G. (1996). Models of the cerebellum and motor learning. *Behavioral and Brain Sciences*, 19, 368–383.
- Hughes, T. (1965). Portal blood supply to glomus tissue and its significance. *Nature*, 205, 149–151.
- Johansson, J. E. (1894). Ueber die Einwirkung der Muskelthätigkeit auf die Athmung und die Herzthätigkeit. *Skand Archives of Physiology*, 5, 20–66.
- Johnson, B. D., Badr, M. S., & Dempsey, J. A. (1994). Impact of the aging pulmonary system on the ventilatory response to exercise. *Clinics in Chest Medicine*, 15(2), 229–246.
- Johnson, R. A., & Mitchell, G. S. (2001). p-Chlorophenylalanine eliminates long-term modulation of the exercise ventilatory response in goats. *Respiration Physiology*, 128, 161–169.
- Johnson, R. A., Okragly, A. J., Haak-Frendscho, M., & Mitchell, G. S. (2000). Cervical dorsal rhizotomy increases brain-derived neurotrophic factor and neurotrophin-3 expression in the ventral spinal cord. *Journal of Neuroscience*, 20(10), RC77.
- Josselyn, S. A., & Tonegawa, S. (2020). Memory engrams: Recalling the past and imagining the future. *Science*, 367(6473), eaaw4325.
- Judd, E. N., Lewis, S. M., & Person, A. L. (2021). Diverse inhibitory projections from the cerebellar interposed nucleus. *eLife*, 10, e66231.
- Kao, F. F. (1963). An experimental study of the pathway involved in exercise hyperpnea employing cross-circulation techniques. In D. Cunningham & B. Lloyd (Eds.), *Regulation of Human Respiration*. Blackwell.
- Kinthead, R., Zhan, W. Z., Prakash, Y. S., Bach, K. B., Sieck, G. C., & Mitchell, G. S. (1998). Cervical dorsal rhizotomy enhances serotonergic innervation of phrenic motoneurons and serotonin-dependent long-term facilitation of respiratory motor output in rats. *Journal of Neuroscience*, 18(20), 8436–8443.
- Korsak, A., Sheikhabahaei, S., Machhada, A., Gourine, A. V., & Huckstepp, R. T. R. (2018). The role of parafacial neurons in the control of breathing during exercise. *Scientific Reports*, 10, 400.

- Koutsikou, S., Crook, J. J., Earl, E. V., Leith, J. L., Watson, T. C., Lumb, B. M., & Apps, R. (2014). Neural substrates underlying fear-evoked freezing: The periaqueductal grey-cerebellar link. *The Journal of Physiology*, 592(10), 2197–2213.
- Kramer, J., Jarboe, M., & Waldrop, T. G. (1996). Periaqueductal grey neuronal responses to hindlimb muscle contraction in the cat. *Society of Neuroscience Abstracts*, 22, 89.
- Krogh, A., & Lindhard, J. (1913). The regulation of respiration and circulation during the initial stages of muscular work. *Scandinavian Archives of Physiology*, 30, 112–136.
- Krohn, F., Novello, M., van der Giessen, R. S., De Zeeuw, C. I., Pel, J. J., & Bosman, L. W. J. (2023). The integrated brain network that controls respiration. *eLife*, 12, e83654.
- Krupa, D. J., Thompson, J. K., & Thompson, R. F. (1993). Localization of a memory trace in the mammalian brain. *Science*, 260(5110), 989–991.
- Krupa, D. J., & Thompson, R. F. (1995). Inactivation of the superior cerebellar peduncle blocks expression but not acquisition of the rabbit's classically conditioned eye-blink response. *Proceedings of the National Academy of Sciences, USA*, 92(11), 5097–5101.
- Krupa, D. J., & Thompson, R. F. (1997). Reversible inactivation of the cerebellar interpositus nucleus completely prevents acquisition of the classically conditioned eye-blink response. *Learning & Memory*, 3, 545–556.
- Krupa, D. J., Weng, J., & Thompson, R. F. (1996). Inactivation of brainstem motor nuclei blocks expression but not acquisition of the rabbit's classically conditioned eyeblink response. *Behavioral Neuroscience*, 110(2), 219–227.
- Lowry, T. F., Forster, H. V., Pan, L. G., Korducki, M. A., Probst, J., Franciosi, R. A., & Forster, M. (1999). Effect of carotid body denervation on breathing in neonatal goats. *Journal of Applied Physiology*, 87, 1026–1034.
- Lutherer, L. O., Williams, J. L., & Everse, S. J. (1989). Neurons of the rostral fastigial nucleus are responsive to cardiovascular and respiratory challenges. *Journal of the Autonomic Nervous System*, 27(2), 101–112.
- MacFarlane, P. M., & Mitchell, G. S. (2009). Episodic spinal serotonin receptor activation elicits long-lasting phrenic motor facilitation by an NADPH oxidase-dependent mechanism. *The Journal of Physiology*, 587(Pt 22), 5469–5481.
- MacFarlane, P. M., Vinit, S., & Mitchell, G. S. (2011). Serotonin 2A and 2B receptor-induced phrenic motor facilitation: Differential requirement for spinal NADPH oxidase activity. *Neuroscience*, 178, 45–55.
- Mantyh, P. W. (1983). Connections of midbrain periaqueductal grey in the monkey. II. Descending efferent projections. *Journal of Neurophysiology*, 49(3), 582–594.
- Marr, D. (1969). A theory of the cerebellar cortex. *The Journal of Physiology*, 202, 437–470.
- Martin, P. A., Bloomer, S. E. M., & Mitchell, G. S. (1993). Repeated hypercapnia at rest augments future exercise ventilatory responses. *Federation of American Societies for Experimental Biology Journal*, 7, A858.
- Martin, P. A., & Mitchell, G. S. (1992). Effects of repeated exercise hyperpnea on future ventilatory responses to exercise and hypercapnia at rest in goats. *Federation of American Societies for Experimental Biology Journal*, 6, A1757.
- Martin, P. A., & Mitchell, G. S. (1993). Long-term modulation of the exercise ventilatory response in goats. *The Journal of Physiology*, 470, 601–617.
- Martin, T., Keating, J., Goodkin, H., Bastian, A. J., & Thach, W. (1996). Throwing while looking through prisms: I. Focal olivocerebellar lesions impair adaptation. *Brain*, 119(Pt 4), 1183–1198.
- Martino, P. F., Davis, S., Opansky, C., Krause, K., Bonis, J. M., Czerniak, S. G., Pan, L. G., Qian, B., & Forster, H. V. (2006). Lesions in the cerebellar fastigial nucleus have a small effect on the hyperpnea needed to meet the gas exchange requirements of submaximal exercise. *Journal of Applied Physiology*, 101, 1199–1206.
- McCloskey, D. I., & Mitchell, J. H. (1972). Reflex cardiovascular and respiratory responses originating in exercising muscle. *The Journal of Physiology*, 224(1), 173–186.
- McCormick, D. A., Lavond, D. G., Clark, G. A., Kettner, R. E., Rising, C. E., & Thompson, R. F. (1981). The engram found? Role of the cerebellum in classical conditioning of nictitating membrane and eyelid responses. *Bulletin of the Psychonomic Society*, 18, 103–105.
- McKay, L. C., Critchley, H. D., Murphy, K., Frackowiak, R. S. J., & Corfield, D. R. (2010). Sub-cortical and brainstem sites associated with chemostimulated increases in ventilation in humans. *Neuroimage*, 49(3), 2526–2535.
- McKay, L. C., Evans, K. C., Frackowiak, R. S. J., & Corfield, D. R. (2003). Neural correlates of voluntary breathing in humans. *Journal of Applied Physiology*, 95(3), 1170–1178.
- Mills, J. N. (1944). Hyperpnoea in man produced by sudden release of occluded blood. *The Journal of Physiology*, 103(2), 244–252.
- Milsom, W. K., Abe, A. S., Andrade, D. V., & Tattersall, G. J. (2004). Evolutionary trends in airway CO₂/H⁺ chemoreception. *Respiratory Physiology and Neurobiology*, 144, 191–202.
- Mitchell, G. S. (1990). Ventilatory control during exercise with increased respiratory dead space in goats. *Journal of Applied Physiology*, 69, 718–727.
- Mitchell, G. S., & Babb, T. G. (2006). Layers of exercise hyperpnea: Modulation and plasticity. *Respiratory Physiology and Neurobiology*, 151(2-3), 251–266.
- Mitchell, G. S., Bach, K. B., Martin, P. A., Foley, K. T., Olson, E. B., Jr, Brownfield, M. S., Miletic, V., Behan, M., McGuiirk, S., & Sloan, H. E. (2000). Increased spinal monoamine concentrations after chronic thoracic dorsal rhizotomy in goats. *Journal of Applied Physiology*, 89(4), 1266–1274.
- Mitchell, G. S., & Baker, T. L. (2022). Respiratory neuroplasticity: Mechanisms and translational implications of phrenic motor plasticity. *Handbook of Clinical Neurology*, 188, 409–432.
- Mitchell, G. S., Baker, T. L., Nanda, S. A., Fuller, D. D., Zabka, A. G., Hodgeman, B. A., Bavis, R. W., Mack, K. J., & Olson, E. B. Jr. (2001). Invited review: Intermittent hypoxia and respiratory plasticity. *Journal of Applied Physiology*, 90(6), 2466–2475.
- Mitchell, G. S., Cross, B. A., Hiramoto, T., & Scheid, P. (1980). Effects of intrapulmonary CO₂ and airway pressure on phrenic activity and pulmonary stretch receptor discharge in dogs. *Respiration Physiology*, 41, 29–48.
- Mitchell, G. S., Douse, M. A., & Foley, K. T. (1990). Receptor interactions in modulation ventilatory activity. *American Journal of Physiology*, 259, R911–R920.
- Mitchell, G. S., & Johnson, S. M. (2003). Neuroplasticity in respiratory motor control. *Journal of Applied Physiology*, 94(1), 358–374.
- Mitchell, G. S., & Osborne, J. L. (1980). A comparison between carbon dioxide inhalation and increased dead space ventilation in chickens. *Respiration Physiology*, 40, 227–239.
- Mitchell, G. S., Smith, C. A., & Dempsey, J. A. (1984). Changes in the VI-VCO₂ relationship during exercise in goats: Role of the carotid bodies. *Journal of Applied Physiology*, 57(6), 1894–1900.
- Mitchell, G. S., Turner, D. L., Henderson, D. R., & Foley, K. T. (2008). Spinal serotonin receptor activation modulates the exercise ventilatory response with increased dead space in goats. *Respiratory Physiology and Neurobiology*, 161, 230–238.
- Moolenaar, G. M., & Rucker, H. K. (1976). Autoradiographic study of brain stem projections from fastigial pressor areas. *Brain Research*, 114(3), 492–496.
- Moosavi, S. H., Guz, A., & Adams, L. (2002). Repeated exercise paired with “imperceptible” dead space loading does not alter VE of subsequent exercise in humans. *Journal of Applied Physiology*, 92, 1159–1168.
- Morton, S. M., & Bastian, A. J. (2006). Cerebellar contributions to locomotor adaptations during split belt treadmill walking. *Journal of Neuroscience*, 26, 9107–9116.
- Orem, J., & Netick, A. (1986). Behavioral control of breathing in the cat. *Brain Research*, 366(1-2), 238–253.

- Orr, J. A., Fedde, M. R., Shams, H., Roskenbleck, H., & Sceid, P. (1988). Absence of CO₂-sensitive venous chemoreceptors in the cat. *Respiration Physiology*, 73(2), 211–224.
- Otis, A. B., Fenn, W. O., & Rahn, H. (1950). Mechanics of breathing in man. *Journal of Applied Physiology*, 2(11), 592–607.
- Pan, L. G., Forster, H. V., Bisgard, G. E., Kaminski, R. P., Dorsey, S. M., & Busch, M. A. (1983). Hyperventilation in ponies at the onset of and during steady-state exercise. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 54, 1394–1402.
- Pan, L. G., Forster, H. V., Bisgard, G. E., Murphy, C. L., & Lowry, T. F. (1986). Independence of exercise hyperpnea and acidosis during high-intensity exercise in ponies. *Journal of Applied Physiology*, 60(3), 1016–1024.
- Pan, L. G., Forster, H. V., Matino, P., Strecker, P. J., Beales, J., Serra, A., Lowry, T. F., Forster, M. M., & Forster, A. L. (1998). Important role of carotid afferents in control of breathing. *Journal of Applied Physiology*, 85(4), 1299–1306.
- Parkes, M. J. (2017). Reappraisal of systemic venous chemoreceptors: Might they explain the matching of breathing to metabolic rate in humans? *Experimental Physiology*, 102, 1567–1583.
- Parrell, B., Agnew, Z., Nagarajan, S., Houde, J., & Ivry, R. B. (2017). Impaired feedforward control and enhanced feedback control of speech in patients with cerebellar degeneration. *Journal of Neuroscience*, 37(38), 9249–9258.
- Pasalar, S., Roitman, A. V., Durfee, W. K., & Ebner, T. J. (2006). Force field effects on cerebellar Prkinje cell discharge with implications for internal models. *Nature Neuroscience*, 9, 1404–1411.
- Paterson, D. J. (1992). Potassium and ventilation in exercise. *Journal of Applied Physiology*, 72(3), 811–820.
- Paterson, D. J. (2014). Defining the neurocircuitry of exercise hyperpnoea. *The Journal of Physiology*, 592, 433–444.
- Paterson, D. J., Robbins, P. A., & Conway, J. (1989). Changes in arterial plasma potassium and ventilation during exercise in man. *Respiration Physiology*, 78(3), 323–330.
- Pavlov, P. I. (1927). Conditioned reflexes: An investigation of the physiological activity of the cerebral cortex. *Annals of Neurosciences*, 17, 136–141.
- Phillipson, E. A., Bowes, G., Townsend, E. R., Duffin, J., & Cooper, J. D. (1981a). Role of metabolic CO₂ production in ventilatory response to steady-state exercise. *Journal of Clinical Investigation*, 68, 768–774.
- Phillipson, E. A., Duffin, J., & Cooper, J. D. (1981b). Critical dependence of respiratory rhythmicity on metabolic CO₂ load. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 50(1), 45–54.
- Pilarski, J. Q., Leiter, J. C., & Fregosi, R. F. (2019). Muscles of breathing: Development, function, and patterns of activation. *Comprehensive Physiology*, 9(3), 1025–1080.
- Pizarro, J., Warner, M. M., Ryan, M., Mitchell, G. S., & Bisgard, G. E. (1992). Intracarotid norepinephrine infusions inhibit ventilation in goats. *Respiration Physiology*, 90(3), 299–310.
- Ponte, J., & Purves, M. J. (1978). Carbon dioxide and venous return and their interaction as stimuli to ventilation in the cat. *The Journal of Physiology*, 274, 455–475.
- Poon, C., Tin, C., & Yu, Y. (2007). Homeostasis of exercise hyperpnea and optimal sensorimotor integration: The internal model paradigm. *Respiratory Physiology & Neurobiology*, 159(1), 1–13.
- Popa, L. S., Hewitt, A. L., & Ebner, T. J. (2012). Predictive and feedback performance errors are signaled in the simple spike discharge of individual Purkinje cells. *Journal of Neuroscience*, 32, 15345–15358.
- Powell, F. L., Milsom, W. K., & Mitchell, G. S. (1988). Effects of intrapulmonary CO₂ and airway pressure on pulmonary vagal afferent activity in the alligator. *Respiration Physiology*, 74(3), 285–298.
- Priban, I. P., & Fincham, W. F. (1965). Self-adaptive control and respiratory system. *Nature*, 208, 339–343.
- Ramsay, S. C., Adams, L., Murphy, K., Corfield, D. R., Grootoonek, S., Bailey, D. L., Frackowiak, R. S. J., & Guz, A. (1993). Regional cerebral blood flow during volitional expiration in man: A comparison with volitional inspiration. *The Journal of Physiology*, 461, 85–101.
- Reed, J. W., & Coates, J. C. (2001). Induction of long-term modulation of the exercise ventilatory response in man. In H. Poon CS & Kazemi (Eds.), *Frontiers in Modeling and Control of Breathing Advances in Experimental Medicine and Biology*. Springer.
- Reischl, P., Gonzalez, F., Jr., ECJ, G., WE, F., & FS, G. (1979). Arterial PCO₂ response to intravenous CO₂ in awake dogs unencumbered by external breathing apparatus. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 47(5), 1099–1104.
- Schaefer, S. L., & Mitchell, G. S. (1989). Ventilatory control during exercise with peripheral chemoreceptor stimulation: Hypoxia vs. domperidone. *Journal of Applied Physiology*, 67(6), 2438–2446.
- Seven, Y. B., & Mitchell, G. S. (2019). Mechanisms of compensatory plasticity for respiratory motor neuron death. *Respiratory Physiology and Neurobiology*, 265, 32–39.
- Shea, S. A., Andres, L. P., Shannon, D. C., & Banzett, R. B. (1993). Ventilatory responses to exercise in humans lacking ventilatory chemosensitivity. *The Journal of Physiology*, 468, 623–640.
- Sheldon, M. I., & Green, J. F. (1982). Evidence for pulmonary CO₂ chemosensitivity: Effects on ventilation. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 52(5), 1192–1197.
- Smith, C. A., Mitchell, G. S., Jameson, L. C., Musch, T. I., & Dempsey, J. A. (1983). Ventilatory response of goats to treadmill exercise: Grade effects. *Respiration Physiology*, 54(3), 331–341.
- Somjen, G. G. (1992). The missing error signal-regulation beyond negative chemofeedback. *News in Physiological Sciences*, 7, 184–185.
- Stoodley, C. J., & Schmahmann, J. D. (2018). Functional topography of the human cerebellum. *Handbook of Clinical Neurology*, 154, 59–70.
- Stunden, C. E., Filosa, J. A., Garcia, A. J., Dean, J. B., & Putnam, R. W. (2001). Development of in vivo ventilatory and single chemosensitive neuron responses to hypercapnia in rats. *Respiration Physiology*, 127(2-3), 135–155.
- Sullivan, M. J., Higginbotham, M. B., & Cobb, F. R. (1988). Increased exercise ventilation in patients with chronic heart failure: Intact ventilatory control despite hemodynamic and pulmonary abnormalities. *Circulation*, 77(3), 552–559.
- Sumners, D. P., & Turner, D. L. (2003). Long-term modulation of the leg exercise ventilatory response is not elicited by hypercapnic arm exercise. *Respiratory Physiology & Neurobiology*, 138, 107–114.
- Swanson, G. D. (1978). The exercise hyperpnea dilemma. *Chest*, 73, (2 Suppl), 277–279.
- Sylvester, J. T., Whipp, B. J., & Wasserman, K. (1973). Ventilatory control during brief infusions of CO₂-laden blood in the awake dog. *Journal of Applied Physiology*, 35, 178–186.
- Tallman, R. D. J., & Grodins, F. S. (1982). Intrapulmonary CO₂ receptors and ventilatory response to lung CO₂ loading. *Journal of Applied Physiology: Respiratory, Environmental and Exercise Physiology*, 52(5), 1272–1277.
- Teune, T. M., van der Burg, J., van der Moer, J., Voogd, J., & Ruigrok, T. J. (2000). Topography of cerebellar nucleus projections to the brain stem in the rat. *Progress in Brain Research*, 124, 141–172.
- Thornton, J. M., Guz, A., Murphy, K., Griffith, A. R., Pedersen, D. L., Kardos, A., Leff, A., Adams, L., Casadei, B., & Paterson, D. J. (2001). Identification of higher brain centres that may encode the cardiorespiratory response to exercise in humans. *The Journal of Physiology*, 533(Pt 3), 823–836.
- Trenchard, D., Russell, N. J. W., & Raybould, H. E. (1984). Non-myelinated vagal lung receptors and their reflex effects on respiration in rabbits. *Respiration Physiology*, 55(1), 63–79.
- Turner, D. L., Martin, P. A., & Mitchell, G. S. (1995). Hypoxic exercise does not elicit long-term modulation of the normoxic exercise ventilatory response in goats. *Advances in Experimental Medicine and Biology*, 393, 245–248.
- Turner, D. L., & Mitchell, G. S. (1997). Long-term facilitation of ventilation following repeated hypoxic episodes in awake goats. *The Journal of Physiology*, 499, 543–550.

- Turner, D. L., & Stewart, J. D. (2004). Associative conditioning with cycling and inspiratory resistance enhances the early exercise ventilatory response in humans. *Journal of Applied Physiology*, 93(3), 333–339.
- Turner, D. L., & Summers, D. P. (2002). Associative conditioning of the exercise ventilatory response in humans. *Respiratory Physiology and Neurobiology*, 132, 159–168.
- Vose, A. K., Welch, J. F., Nair, J., Dale, E. A., Fox, E. J., Muir, G. D., Trumbower, R. D., & Mitchell, G. S. (2022). Therapeutic acute intermittent hypoxia: A translational roadmap for spinal cord injury and neuromuscular disease. *Experimental Neurology*, 347, 113891.
- Walters, E. T., Carew, T. J., & Kandel, E. R. (1979). Classical conditioning in *Aplysia californica*. *Proceedings of the National Academy of Sciences, USA*, 76(12), 6675–6679.
- Walters, E. T., Carew, T. J., & Kandel, E. R. (1982). Habituation, sensitization and associative learning in *Aplysia*. In A. L. Beckman (Ed.), *The Neural Basis of Behavior* pp. 117–138. Springer.
- Ward, S. A. (1994). Peripheral and central chemoreceptor control of ventilation during exercise in humans. *Canadian Journal of Applied Physiology*, 19, 305–333.
- Ward, S. A. (2019). Exercise physiology: Exercise hyperpnoea. *Current Opinion in Physiology*, 10, 166–172.
- Warner, M. M., & Mitchell, G. S. (1990). Ventilatory response to hyperkalemia and exercise normoxic and hypoxic goats. *Respiration Physiology*, 82(2), 239–249.
- Warner, M. M., & Mitchell, G. S. (1991). Role of catecholamines and beta-receptors in ventilatory response during hypoxic exercise. *Respiration Physiology*, 85, 41–53.
- Wasserman, K., Whipp, B. J., Casaburi, R., Golden, M., & Beaver, W. L. (1979). Ventilatory control during exercise in man. *Bulletin Europeen De Physiopathologie Respiratoire*, 15(1), 27–51.
- Wasserman, K., Whipp, B. J., Casaburi, R., Huntsman, D. J., Castagna, J., & Lugliani, R. (1975). Regulation of arterial PCO₂ during intravenous CO₂ loading. *Journal of Applied Physiology*, 38, 651–656.
- Wasserman, K., Whipp, B. J., & Castagna, J. (1974). Cardiodynamic hyperpnea: Hyperpnea secondary to cardiac output increase. *Journal of Applied Physiology*, 36(4), 457–464.
- Wasserman, K., Whipp, B. J., Koyal, S. N., & Cleary, M. G. (1975). Effect of carotid body resection on ventilatory and acid-base control during exercise. *Journal of Applied Physiology*, 39, 354–358.
- Whipp, B. J. (2008). Control of exercise hyperpnoea: The unanswered question. In *Adv Exp Med Biol*, pp. 16–21. Springer.
- Wickstom, H. R., Holgert, H., Hokfelet, T., & Lagercrantz, H. (1999). Birth-related expression of c-fos, c-jun and substance P mRNAs in the rat brainstem and pia mater: Possible relationship to changes in central chemosensitivity. *Brain Research Developmental Brain Research*, 112(2), 255–266.
- Williamson, J. W., Fadel, P. J., & Mitchell, J. H. (2006). New insights into central cardiovascular control during exercise in humans: A central command update. *Experimental Physiology*, 91(1), 51–58.
- Williamson, J. W., McColl, R., Mathews, D., Mitchell, J. H., Raven, P. B., & Morgan, P. W. (2002). Brain activation by central command during actual and imagined handgrip under hypnosis. *Journal of Applied Physiology*, 92, 1317–1324.
- Wood, H. E., Mitchell, G. S., & Robbins, P. A. (2003). A learned component of the ventilatory response to exercise in man. *The Journal of Physiology*, 553(Pt 3), 967–974.
- Wood, H. E., Mitchell, G. S., & Babb, T. G. (2008). Short-term modulation of the exercise ventilatory response in young men. *Journal of Applied Physiology*, 104, 244–252.
- Wood, H. E., Mitchell, G. S., & Babb, T. G. (2009). Breathing mechanics during exercise with added dead space reflect mechanisms of ventilatory control. *Respiratory Physiology and Neurobiology*, 168, 210–217.
- Wood, H. E., Mitchell, G. S., & Babb, T. G. (2010). Short-term modulation of the exercise ventilatory response in older men. *Respiratory Physiology and Neurobiology*, 173(1), 37–46.
- Wood, H. E., Mitchell, G. S., & Babb, T. G. (2011). Short-term modulation of the exercise ventilatory response in younger and older women. *Respiratory Physiology and Neurobiology*, 179(2-3), 235–247.
- Xu, F., & Frazier, D. T. (1997). Respiratory-related neurons of the fastigial nucleus in response to chemical and mechanical challenges. *Journal of Applied Physiology*, 82(4), 1177–1184.
- Xu, F., & Frazier, D. T. (2002). Role of the cerebellar deep nuclei in respiratory modulation. *The Cerebellum*, 1(1), 35–40.
- Xu, F., Zhang, Z., & Frazier, D. T. (2001). Microinjection of acetazolamide into the fastigial nucleus augments respiratory output in the rat. *Journal of Applied Physiology*, 91(5), 2342–2350.
- Yamamoto, W. S., & Edwards, M. W., Jr. (1960). Homeostasis of carbon dioxide during intravenous infusion of carbon dioxide. *Journal of Applied Physiology*, 15, 807–818.
- Zuntz, N., & Geppert, J. (1886). Über die Natur der normalen Atemreize und den Ort ihrer Wirkung. *Archiv für die gesamte Physiologie des Menschen und der Tiere*, 38, 337–338.

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