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Plain language summary

Patients with long COVID suffer from

breathlessness during exercise, leading to

exercise intolerance. We know that SARS-CoV-2, the virus that causes COVID-19, can

infect carotid bodies which is a small sensory

regulating breathing and blood pressure. This

is called the carotid chemoreflex. However, it

carotid chemoreflex. Here, we examine whe-

patients with long COVID and if this is linked to

study shows that carotid chemoreflex is more sensitive in long COVID patients, who are

otherwise healthy. The carotid bodies could

breathlessness in patients with long COVID.

be a good therapeutic target for treating

is not clear if SARS-CoV-2 infection affects

ther the normal functioning of carotid che-

moreflex is disrupted in non-hospitalised

excessive breathing during exercise. Our

organ that sends signals to the brain for

Carotid body dysregulation contributes to Long COVID symptoms

Check for updates

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Abstract

Background The symptoms of long COVID, which include fatigue, breathlessness, dysregulated breathing, and exercise intolerance, have unknown mechanisms. These symptoms are also observed in heart failure and are partially driven by increased sensitivity of the carotid chemoreflex. As the carotid body has an abundance of ACE2 (the cell entry mechanism for SARS-CoV-2), we investigated whether carotid chemoreflex sensitivity was elevated in participants with long COVID.

Methods Non-hositalised participants with long-COVID (n = 14) and controls (n = 14) completed hypoxic ventilatory response (HVR; the measure of carotid chemoreflex sensitivity) and cardiopulmonary exercise tests. Parametric and normally distributed data were compared using Student's unpaired t-tests or ANOVA. Nonparametric equivalents were used where relevant. Peason's correlation coefficient was used to examine relationships between variables.

Results During cardiopulmonary exercise testing the V_E/VCO₂ slope (a measure of breathing efficiency) was higher in the long COVID group (37.8 ± 4.4) compared to controls (27.7 ± 4.8, P = 0.0003), indicating excessive hyperventilation. The HVR was increased in long COVID participants (-0.44 ± 0.23 l/min/ SpO₂%, R² = 0.77 ± 0.20) compared to controls (-0.17 ± 0.13 l/min/SpO₂%, R² = 0.54 ± 0.38, P = 0.0007). The HVR correlated with the V_E/VCO₂ slope (r = -0.53, P = 0.0036), suggesting that excessive hyperventilation may be related to carotid body hypersensitivity.

Conclusions The carotid chemoreflex is sensitised in long COVID and may explain dysregulated breathing and exercise intolerance in these participants. Tempering carotid body excitability may be a viable treatment option for long COVID patients.

Long COVID (post-COVID-19 syndrome), is a multi-organ, often debilitating condition associated with a range of symptoms. The UK's National Institute for Health Care and Excellence (NICE) defines long COVID as ongoing symptoms lasting for 12 or more weeks after initial SARS-CoV-2 infection, without alternative explanations¹. The estimated incidence of long COVID varies, and is reported to be up to 41% of non-hospitalised cases²⁻⁴ and up to 76% of hospitalised cases^{5,6}. The prevalence decreases in vaccinated populations⁷. In January 2023, 2 million people self-reported long COVID symptoms in the UK, with 77% experiencing adverse effects in their daily activities⁸. Persistent symptoms include chronic fatigue, 'brain fog,' cognitive impairment and memory loss, dyspnoea at rest and on exertion, exercise intolerance, orthostatic intolerance, inappropriate postural tachycardia, and episodic hyperadrenergic surges^{4,9,10}. A meta-analysis of 63 studies worldwide, with a total COVID-19 population of 257,348,

¹School of Physiology, Pharmacology and Neuroscience, University of Bristol, Bristol, UK. ²Department of Cardiology, Southmead Hospital, North Bristol NHS Trust, Bristol, UK. ³Bristol Heart Institute, University Hospitals Bristol and Weston NHS Foundation Trust, Bristol, UK. ⁴Department of Anaesthetics, Southmead Hospital, North Bristol NHS Trust, Bristol, UK. ⁵Department of Respiratory Medicine, University Hospitals Bristol and Weston NHS Foundation Trust, Bristol, UK. ⁶Manaaki Manawa, The Centre for Heart Research, University of Auckland, Auckland, New Zealand. ⁷These authors contributed equally: Ahmed El-Medany, Zoe H. Adams, Hazel C. Blythe. —email: Emma.hart@bristol.ac.uk reported that between 3–6 and 9–12 months post-infection, fatigue and dyspnoea were the most reported symptoms, with a prevalence of 32–47% and 21–25% respectively¹¹. Despite the prevalence of long COVID and severely disabling symptoms there are no treatment strategies available. Thus, it is crucial to identify the mechanisms involved in long COVID to inform urgently needed therapy. It is likely that mechanisms depend on the severity of the original infection and are different for hospitalised (e.g. long term sequalae from intensive care and intubation/ventilation) versus non-hospitalised patients who had mild to moderate initial symptoms.

Currently, the exact mechanisms driving long COVID in nonhospitalised patients remain unknown but are likely to be multiple¹². Studies have shown that exercise intolerance and disorganised breathing or breathing inefficiency during exercise are key features of long COVID¹³⁻¹⁶ even in patients who have normal lung function and no evidence of gas exchange abnormalities¹⁵. The carotid bodies are key oxygen, carbon dioxide and pH sensing organs that control ventilation, dyspnoea, and the circulation at rest and during exercise in health and disease^{17,18}. In fact, in chronic heart failure the carotid chemoreflex becomes chronically sensitised as a compensatory mechanism, and is associated with a worse prognosis¹⁹, exertional dyspnoea, dysfunctional or inefficient breathing, and poor exercise tolerance²⁰, similar to symptoms in patients with long COVID who do not have heart failure.

SARS-CoV-2 infects host cells via binding of its receptor-binding domain to the membrane bound angiotensin-converting enzyme 2 (ACE2)²¹. ACE2 is abundant in the carotid bodies²². A role of the carotid chemoreflex in the acute phase of the infection is supported by reports of silent hypoxia^{22,23}, SARS-CoV-2 invasion of glomus cells (main oxygen sensing cells) and microembolism within the small arteries suppling blood to the carotid body^{24,25}. The carotid bodies express their own renin-angiotensin system^{26,27}, where normal functioning is dependent on the balance of ACE1 and ACE2²⁶. Disruption of this local system causes increased carotid chemoreflex activity^{27,28}. Thus, disturbances in the carotid body following SARS-CoV-2 infection; by viral invasion, blood flow disruption and local immune responses could cause chemoreceptive dysfunction, by increasing local ACE1/ACE2 imbalance (in favour of higher ACE1 expression versus ACE2) and angiotensin II receptor stimulation^{24,29}.

We propose that carotid body dysfunction occurs in long COVID, which contributes to dysregulation of ventilation and cardiovascular control, especially during exercise. Therefore, we conducted a casecontrol study to determine whether carotid chemoreflex sensitivity is elevated in non-hospitalised patients with long COVID and whether this could help to explain impairments in exercise tolerance and dysregulated breathing reported during exercise in non-hospitalised patients with ongoing symptoms. We hypothesised that long COVID patients would exhibit increased hypoxic ventilatory responses at rest and poorer ventilatory efficiency during exercise compared to controls. Ventilatory efficiency is defined as ventilation relative to the CO₂ production. The V_E/VCO_2 slope can be used to quantify the efficiency of ventilation in patient cohorts; where a steeper slope represents poorer efficiency^{30,31}. There is an exception, however; the V_E/VCO_2 slope does not reflect ventilatory efficiency in patients with moderate to advanced chronic obstructive lung disease³². In the current study, since the long COVID patients recruited did not have COPD, the V_E/ VCO₂ slope was used as the main measure of exercise ventilatory efficiency.

The findings of this study show that carotid chemoreflex sensitivity is amplified in non-hospitalised patients with long COVID (versus a control group) and that this correlates with hyperventilation and poor breathing efficiency during exercise. Elevated carotid chemoreceptor activity could explain several ongoing symptoms experienced by patients living with long COVID.

Methods

Design

This was a single-site case-control study.

Participants

Ethical approval for the study was granted by South Central Hampshire NHS Research Ethics Committee (21/SC/0260) and the Health Research Authority. Participants gave their written informed consent. All participants were asked to abstain from intense exercise and alcohol consumption 24 h before the study. All experimental protocols conformed to the Declaration of Helsinki. Inclusion criteria for all participants were: aged 18-80 years and a positive SARS-CoV-2 antibody test before vaccination, or a positive COVID-19 PCR antigen swab test. Long COVID participants had received a diagnosis of long COVID, where symptoms developed during or after an infection consistent with COVID-19 and continued for more than 12 weeks (and could not be explained by an alternative diagnosis) as per NHS (UK) National Institute for Health and Care Excellence guidelines³³. Participants without long COVID had symptoms lasting less than 4 weeks after their initial infection. See the online-only Supplement Note for exclusion criteria. However, due to scheduling issues (limited by the availability of staff support rather than recruitment), data sets from six healthy participants from a previous study (NHS REC numbers: 17/SW/0171 and 18/SW/0241), completed before the SARS-CoV-2 pandemic using the same methods, equipment, and location (temperature controlled clinical room) were added to the control group.

Experimental protocol

Participants attended the NIHR Bristol Clinical Research facility for 2 studies, completed at the same time of day, and the laboratory conditions were at a set temperature (22 °C). In visit one, informed consent, in-depth medical history, clinic blood pressure assessment, sit-to-stand test for orthostatic intolerance, lung function tests (spirometry), 12-lead ECG, pregnancy tests, and a symptom-limited incremental cardiopulmonary exercise test were completed. At this visit, participants were asked to list their ongoing symptoms (if any) and were specifically asked about symptoms associated with long COVID that have been previously published^{9,10}. The second visit involved resting ventilation and cardiovascular measurements followed by carotid chemoreflex assessment via the hypoxic ventilatory response.

Procedures

Clinic blood pressure. Participants rested in a chair for 10 min prior to clinic BP being assessed (Omron, 705IT, Omron Healthcare, Kyoto, Japan). Clinic BP was assessed in-line with European Hypertension Society Guidelines³⁴.

Orthostatic intolerance was assessed using a sit-to-stand test using the HYVET protocol³⁵ where BP was measured when sitting, immediately upon standing, and after 1, 2 and 3 minutes of standing.

Resting spirometry. Resting spirometry was used to assess lung function, to ensure no mechanical lung function abnormalities were present. Spirometry (Ergostick, CPET system, LoveMedical, UK) was completed in line with the joint American Thoracic and European Respiratory Society guidelines³⁶. The Global Lung Function Initiative (GLI) network reference values were used to calculate the percentage of predicted values and z-scores³⁷.

12-lead ECG: Resting 12-lead ECG was performed and checked by a Cardiologist at the Bristol Heart Institute for any ECG abnormalities, and to clear participants to exercise.

Cardiopulmonary exercise testing

After acclimatisation to the facemask and sitting on the cycle ergometer, participants completed a 5-min steady state resting period followed by 3 min of unloaded cycling. Participants then completed a continuous ramp incremental exercise test to volitional exhaustion where work rate increased by 15–30 W depending on their physical ability. Exercise tests were completed on an electronically braked cycle ergometer (Ergoselect 5; Ergoline, Germany). Cardiorespiratory data were recorded using a metabolic measurement system (Ergostik; LoveMedical, UK) with integrated 12-lead ECG

and finger pulse oximetry for heart rate and SpO₂% monitoring. Brachial arterial blood pressure was measured via an integrated automated auscultatory blood pressure cuff (LoveMedical, UK. Ratings of perceived exertion (6–20 Borg scale) and dyspnoea scores (modified Borg scale) were obtained at rest, and every minute during exercise, and at the end of exercise. Peak cardiopulmonary data were averaged over the last 30 seconds of exercise. The anaerobic threshold was measured via the V-slope method and Dual Criterion methods as recommended by the American Thoracic Society guidelines³⁸.

Measurement of exercise ventilatory efficiency

The main measurement of ventilatory efficiency was the VE/VCO2 slope, calculated as the minute ventilation, and VCO₂ values from initiation to peak exercise were used to measure V_E/VCO₂ slope via least squares linear regression³¹. Secondary measurements of ventilatory efficiency included, the V_E/VCO₂ nadir (calculated as the lowest V_E/VCO₂ ratio during exercise using 30 s of averaged breaths with 5-of-7 breath averaging³⁰), and the V_E/VCO₂ at anaerobic threshold.

Carotid chemoreflex assessment

Resting carotid chemoreceptor sensitivity tests were completed with participants in a semi-supine position with continuous monitoring of beat-to-beat blood pressure (Finapres), SpO₂ (ear-lobe pulse oximeter; Radical-7; Masimo Corp, USA), heart rate (lead II of 3-lead ECG, AD Instruments, Australia). Simultaneously, ventilation was measured via a facemask attached to a one-way non-rebreathing circuit (Hans Rudolph, Inc., USA). The inhalation part of the circuit delivered room air or 100% nitrogen gas (transiently) for carotid chemosensitivity testing. The exhalation arm of the circuit was connected to a gas

analyser (Ad Instruments) and flow head (MLT3000L; AD Instruments) fitted with a differential pressure transducer (FE141 Spirometer; AD Instruments) for the measurement of inspired and expired fractions of O_2 and CO_2 , tidal volume, breathing frequency and minute ventilation. All data were continuously monitored and recorded with a data acquisition system (Powerlab 16/30; AD Instruments) and stored for subsequent analyses using associated software (Lab-Chart 8.0 Pro; AD Instruments).

The transient hypoxic ventilatory response test was used to measure the sensitivity of the carotid chemoreflex^{20,39}. After a period of quiet rest breathing room air (10 min baseline), the researcher added extra-nitrogen to the room air being delivered to the face-mask. Nitrogen gas administration was controlled silently using a high-pressure electric valve. The nitrogen blended into the room air was delivered for 2-8 breaths, followed by a 3-minute recovery period or until ventilation and haemodynamic variables returned to baseline levels. This was repeated 6-8 times to obtain a range of oxygen saturations (SpO2: ~70-100%). The average of the two largest consecutive breaths in the 1 min proceeding the nitrogen exposure was used to calculate the ventilatory response to reductions in SpO2%¹⁹. The hypoxic ventilatory response was evaluated as the slope of the linear regression relating the minute ventilation to the nadir of SpO2 for each nitrogen exposure^{19,39} (Fig. 1b). The response of tidal volume and breathing frequency to reductions in SpO2 were also evaluated in the same way as that for minute ventilation.

The peak heart rate and blood pressure was determined following each hypoxic challenge using a 3-beat rolling average and plotted against the nadir oxygen saturation. The hypoxic heart rate (beats/min/%) and blood pressure (mmHg/%) were calculated as the slope of the simple linear regression obtained from baseline and the hypoxic challenges.



Fig. 1 | Measures of carotid chemoreflex sensitivity and ventilatory efficiency in humans. a Example carotid chemoreflex test in a control (n = 1, solid line) and long COVID (n = 1, dashed line) participants. Data shows the raw SpO₂ response to nitrogen exposure (bottom panel) and the resultant tidal volume, breathing frequency, and minute ventilation responses. **b** The minute ventilation (average of the largest 2 consecutive breaths) is plotted against the nadir SpO₂ for each nitrogen

exposure. The resultant slope of the linear regression is the hypoxic ventilatory response. Data are from a control (black circles) and a long-COVID (open red circles) participants, **c** the minute ventilation (V_E) plotted against the volume of expired carbon dioxide (VCO₂) for each breath during exercise in the same 2 participants (control; grey circles and long-COVID; open red circles). The slope of the regression is the V_E/VCO₂ slope and is used as a measure of breathing efficiency.

Statistics and reproducibility

Statistical analyses were completed in GraphPad Prism (V9.5.1). Participant demographics, resting spirometry, cardiopulmonary exercise and hypoxic ventilatory response data were analysed using an independent samples t-test or Mann Whitney U test if data were not normally distributed (Shapiro-Wilk test and visual inspection using Q-Q plots) or were non-parametric. Where data are compared across multiple time points between the groups, a mixed model ANOVA was used with a Bonferroni correction for pairwise comparisons. Data are reported as mean ± standard deviation (SD) or median (interquartile range) unless otherwise stated. A significance level of $\alpha < 0.05$ was used for all analyses.

Reporting summary

Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

Results Participants

Supplementary Fig. 1 shows recruitment, screening, cases excluded and the final sample size. Sixty-four individuals contacted the group about the study, and all were sent a participant information sheet. Of these, 32 replied and completed phone screening. Six were excluded due to screen failure (met exclusion criteria). Twenty-six participants were recruited (long COVID n = 16 and controls n = 10). Two participants (1 long COVID and 1 control) did not complete the study since they could not tolerate the mask for spirometry, 1 long COVID participant was excluded during the study visit due to identification of cardiac disease (valve disease or non-benign arrhythmia), and 1 participant was excluded between visits due to diagnosis of gout (control). The final sample size was 14 participants with long COVID and 8 controls (recovered from initial viral infection within 4 weeks

Table 1 | Demographic, spirometry, and SARS-CoV-2 infection data

Demographics	Controls (<i>n</i> = 14)	Long COVID (<i>n</i> = 14)	P-value
Sex (M/F)	3/11	2/12	_
Age (years)	35 ± 13	42 ± 12	0.1195
BMI (kg/m²)	24.5 ± 4.9	26.9 ± 5.3	0.2140
Height (cm)	166 ± 5	167 ± 7	0.5543
Body mass (kg)	67.2 ± 12.9	75.1 ± 16.3	0.1573
Received all recommended** COVID-19 vaccinations at time of study (%)	100%	100%	_
Time since initial infection** (days) [median (IQR)]	473 (265-543)	534 (400-709)	0.1006*
Clinic BP			
SBP (mmHg)	121±9	131 ± 17	0.0732
DBP (mmHg)	75 ± 7	83 ± 13	0.0594
HR (beats/min)	69 ± 12	75 ± 12	0.1895
Resting lung function			
n	11	14	
FEV ₁ (z-scores)	-0.32 ± 0.92	-0.90 ± 1.23	0.2109
FVC (z-scores)	-0.56 ± 1.01	-1.03 ± 0.86	0.2279
FEV ₁ /FVC (z-scores)	0.60 ± 0.95	0.21 ± 1.13	0.3727
FEF _{75%} (z-scores)	0.61 ± 1.01	0.43 ± 0.77	0.6191

*Mann Whitney U. BMI body mass index, IQR interquartile range, BP blood pressure, SBP systolic blood pressure, DBP diastolic blood pressure, HR heart rate, FEV1 forced expiratory volume in 1 s, FVC forced vital capacity, FEF75% forced expiratory flow after 75% of the FVC has been exhaled. Mean ± SD unless otherwise stated, **Note that the control group includes data from 6 participants who took part in studies in the same laboratory before the pandemic and did not have COVID-19. Thus, the data pertaining to vaccines and infections is for the 8 participants recruited who had COVID-19 infection but did not develop ongoing symptoms

without ongoing symptoms). Data sets from six healthy participants from a previous study (NHS REC numbers: 17/SW/0171 and 18/SW/0241), completed before the SARS-CoV-2 pandemic using the same methods, equipment, and location (clinical room) were added to the control group. These participants were identified based on age, sex, and body mass index (BMI) so that they matched the participants in the long COVID study.

Participant demographics are shown in Table 1. Age, BMI, height, and body mass were similar between the control and long COVID groups (P > 0.05). Clinic systolic blood pressure (SBP), diastolic BP (DBP) and heart rate (HR), were not different between groups (P > 0.05).

All participants who had COVID-19 were first infected from April 2020 to December 2021, when the dominant variants of SARS-CoV-2 were the original virus, Alpha, Delta, and Omicron. The average number of days since first infection to the study visit, in the long COVID and control groups, were similar (Table 1, P = 0.1006). As per the exclusion criteria, none of the participants were hospitalised (admitted) for COVID-19 and had a previous positive PCR test at the time of the initial infection or were seropositive for SARS-CoV-2 antibodies (see inclusion/exclusion criteria in methods). The initial infection caused mild to moderate symptoms in all participants. Table 2 outlines the most common ongoing symptoms previously reported^{9,40,41} and the percentage of the long COVID participants in this study reporting these symptoms. Long COVID participants reported at least 3 symptoms, with the most common symptoms being dyspnoea at rest and on exertion, extreme fatigue, "brain fog" and chest pain.

Ten out of the 14 long COVID participants had been seen in a cardiology outpatient long COVID clinic at a UK based University Hospital NHS Trust, where structural cardiac disease (e.g., myocarditis, heart failure, ischaemia) had been excluded. All the participants reporting chest pain were reviewed in a cardiology clinic, which was diagnosed as non-cardiac in origin. None of the long COVID participants (or controls) had been diagnosed with pre-existing cardiovascular or pulmonary diseases. Four of the long COVID participants had been prescribed ivabradine in the cardiology outpatient long COVID clinic. These participants stopped taking ivabradine 48 h before their study visits (plasma half-life: 2 h and effective half-life: 11 h). Severe asthma was an exclusion criterion, however, 2 controls and 6 long COVID participants had been previously diagnosed with mild asthma (before COVID). All participants had a normal resting 12-lead ECG, which was completed as part of the study screening process. Twelve of the 14 long

Table 2 Common ongoing symptoms reported in post COVID-19 syndrome and the % of participants in the present study, self-reporting these symptoms

Symptoms	Long COVID (n = 14)
At least 3 symptoms	100%
Dyspnoea on exertion	92%
Dyspnoea at rest	86%
Brain fog	86%
Fatigue	79%
Chest pain	50%
Palpitations or tachycardia	36%
Dizziness	36%
Post-exertional malaise	36%
Muscle or joint pain	21%
Oedema in extremeties	14%
Nausea	14%
Chronic dry cough	14%
Impaired taste or smell	7%
Alopecia	7%
Chronic fever	0%
Diarrhoea	0%

COVID participants self-reported having a chest x-ray, all of which were normal. Finally, Supplementary Table 1 provides information on drugs prescribed in both groups.

Resting spirometry

Spirometry data are summarised in Table 1 and are presented as z-scores with a normal z-score being between $\pm 1.64^{42}$. There were no differences between the two groups for FEV₁ (forced expiratory volume in 1 s), FVC (forced vital capacity), FEV₁/FVC and FEF_{75%} (forced expiratory flow after 75% of the FVC has been exhaled; P > 0.05). No participants had an FEF_{75%} smaller than -1.64 z-score, therefore excluding small airway disease. One control subject and 3 long COVID participants had a reduced FEV₁ and FVC with a normal FEV₁/FVC ratio, suggestive of a restrictive ventilatory defect. One long COVID participant had a reduced FEV₁ and FVC, with an FEV₁/FVC ratio consistent with an obstructive ventilatory defect (z-score = -3.25) indicating severe airflow obstruction (z-score = -3.85)⁴².

Sit to standing blood pressure test

To check for orthostatic intolerance in both groups, a sit-to-standing test (like that used in the HYVET study³⁵) was completed (3 mins of standing). Sit-to-stand was completed in all long COVID participants and 8 of the controls. BP was measured at seated rest, immediately after standing, 1 min, 2 min, and 3 min of standing. According to HYVET study guidelines, a fall in SBP > 15 and DBP > 7 mmHg was used as a diagnosis of orthostatic intolerance. During standing, the mean SBP and DBP did not fall by more than 15 and 7 mmHg in either group at any time point³⁵ (Supplementary Fig. 2; Supplementary Table 2 for mean change data). Two control participants had a fall in SBP > 15 or DBP > 7 mmHg, and 1 participant from the long COVID group met this criterion (Supplementary Fig. 2). Mean data at each timepoint were analysed using a mixed model ANOVA. Interestingly, we found that the increase in HR was larger in the control group versus the long COVID group (main Time*Group effect; P = 0.0258, see Supplementary Data for ANOVA details and Fig. 2).

Fig. 2 | The hypoxic ventilatory response (HVR) based on minute ventilation, tidal volume (V_T) and breathing frequency (f_B). A more negative response indicates a greater HVR. Closed circles are control and open circles are long-COVID participants. The red dots denote control participants that did not have COVID-19 (n = 6). Panels **a**-**c** show the hypoxic ventilatory response based on minute ventilation, tidal volume and breathing frequency, respectively. There was a higher HVR based on minute ventilation and tidal volume in the participants with long COVID (n = 14) versus control (n = 14). Panels **d**-**f** show the heart rate, systolic and diastolic blood pressure hypoxic responses. Data are the slope of the linear regression where SpO2% was plotted against the heart rate, and BP during each hypoxic exposure. There was no difference in the HR or BP response to hypoxia between groups. Data are mean +/-SD.

Resting cardiopulmonary data

The resting HR, BP, minute ventilation, tidal volume and breathing frequency were similar between groups (Table 3, P > 0.05). Resting partial pressure of the end tidal CO₂ (P_{ET}CO₂) was lower in the long COVID group (range: 22 to 34 mmHg versus controls (range: 30–36 mmHg, Table 3, P = 0.0378). The minute ventilation was similar between groups despite a lower P_{ET}CO₂ in the long COVID group. This was coupled with a higher ventilation for the volume of CO₂ expired (V_E/VCO₂ ratio, P = 0.0246); suggesting that there is some hyperventilation occurring at rest in the long COVID group. There were no differences in the expiratory and inspiratory times, or the ratio of inspiration to expiratory times between groups (P > 0.05; Table 3 for exact *P*-values). Interestingly, the tidal volume/ inspiratory time ratio (V_T/Ti; an index of inspiratory flow⁴³) was 16% higher in the long COVID group versus the controls (P = 0.0483) indicative of increased inspiratory drive.

Cardiopulmonary exercise testing

Cardiopulmonary exercise testing (CPET) was completed on a cycle ergometer to peak oxygen consumption (VO₂ peak) to assess exercise tolerance and identify any breathing or cardiovascular abnormalities that might not be evident at rest. We also wanted to understand whether any dysfunctional breathing (defined here as inappropriate hyperventilation or poor breathing efficiency characterised by elevated minute ventilation/volume of CO₂ expired (V_E/VCO₂) slopes¹⁶) during exercise could be linked to carotid chemoreflex hyperactivity in the long COVID group versus control.

To assess whether maximal effort was achieved and as an objective assessment of the quality of the test, a respiratory exchange ratio>1.15, maximum predicted HR > 85%, rating of perceived exertion (RPE; 6–20 Borg Scale) of 17–20 and a plateau in VO₂ were used. Overall, 86% of the control group and 86% of the long COVID group achieved these criteria. Thus, differences between groups are likely not due to differences in effort or quality of the CPET. In fact, the control group reported a lower RPE at peak exercise (17; 17–18 (median, IQR)) versus the long COVID group (19; 18–19, P = 0.0081; Mann Whitney U). Table 4 shows the mean ± SD (range)



Table 3 | Baseline cardiopulmonary variables in controls versus long COVID

Baseline cardio- pulmonary data	Controls (n = 14)	Long COVID (n = 14)	P-value
SBP (mmHg)	112 ± 9	118 ± 16	0.1603
DBP (mmHg)	75 ± 8	80 ± 16	0.1985
HR (beats/min)	76 ± 9	83 ± 11	0.0886
V _E (L/min)	9.2 ± 2.1	10.1 ± 2.8	0.3434
V ₇ (mL)	609 ± 115	659 ± 173	0.3546
f _B (breaths/min)	15 ± 3	16 ± 3	0.8938
Ti (secs)	1.65 ± 0.37	1.60 ± 0.41	0.7934
Te (secs)	2.86 ± 0.61	2.77 ± 0.81	0.7509
Ti/Te	0.59 ± 0.08	0.59 ± 0.12	0.9062
V ₇ /Ti	0.37 ± 0.07	0.43 ± 0.09	0.0483
P _{ET} CO ₂ (mmHg)	33.5 ± 2.6	30.9 ± 3.7	0.0378
VO ₂ (L/min)	0.25 ± 0.05	0.24 ± 0.05	0.5781
VCO ₂ (L/min)	0.22 ± 0.04	0.21 ± 0.05	0.8006
V _E /VCO ₂ ratio	42.2 ± 4.4	48.0 ± 8.3	0.0246

SBP systolic blood pressure, DBP diastolic blood pressure, HR heart rate, VE minute ventilation, V_T tidal volume, f_B breathing frequency, T_I inspiratory time, Te expiratory time, V_T/T_i inspiratory flow index, $P_{ET}CO_2$ end-tidal partial pressure of carbon dioxide, VO_2 volume of inspired oxygen, VCO_2 volume of expired carbon dioxide. Mean \pm SD. Data compared via unpaired Student's t-test. Bold data are where P < 0.05.

for all CPET variables. None of the participants desaturated during the test (defined as SpO_2 %<95%, where peak SpO_2 % was similar between groups), and there were no exercise induced cardiac ischaemic changes observed on the 12-lead ECG.

The VO₂ peak measured in L/min, mL/kg/min and as a % of peak predicted were lower in the long COVID group versus controls (Table 4; P < 0.05). Absolute VO₂ at anaerobic threshold (AT) was also lower (P = 0.0025), but the % of peak VO₂ at which the AT occurred was similar between groups (P = 0.6490), indicating normal metabolic function. Along these lines there were no other indications of defects in muscle metabolic pathways indicating no muscle metabolic limitations to VO2 peak in either group (i.e. normal peak respiratory exchange ratio (RER) and a normal VO₂/work rate relationship⁴⁴ (Table 4). Maximum HR was lower in the long COVID group versus controls, but there was no difference between groups when HR was measured as the % of predicted maximum (P = 0.1326). The slope of the HR plotted against the %VO₂ peak was lower in the long COVID group (Table 4; P = 0.0114) versus controls indicating a blunted HR response to exercise and potentially some degree of chronotropic incompetence. Heart rate at AT was also lower in the long COVID group versus controls (P = 0.0453). Finally, the peak oxygen pulse (VO₂/HR) was lower in the long COVID group versus controls, indicating a cardiovascular limitation to exercise. Despite this, the HR recovery at 1 min was similar between groups (P = 0.4653); indicating that parasympathetic engagement post-exercise in the long COVID group was similar to control.

The VCO₂ at peak was lower in the long COVID group (Table 4; P = 0.0477), whereas minute ventilation, tidal volume and breathing frequency at peak were similar between groups (Table 4, P > 0.05). Supplementary Fig. 3 shows the minute ventilation versus the VCO₂ and VO₂ at rest, AT, and peak exercise in both groups. The mean breathing reserve (max minute ventilation/predicted maximal voluntary ventilation (calculated as FEV1 * 40) was similar between groups (P = 0.8708). Thus it is likely that the lower peak VO₂ in the long COVID group is not a result of pulmonary mechanical limitations⁴⁵.

The V_E/VCO₂ *ratio* at peak (ANOVA; P = 0.0051) and at AT (P = 0.0477) was higher in the long COVID group versus control (see Supplementary Fig. 3 for ANOVA details and V_E/VCO₂ ratio plotted against the VO₂ at rest, AT, and peak exercise). The V_E/VCO₂ slope (Table 4, P = 0.0008) and the V_E/VCO₂ nadir (P = 0.0020) during exercise

Table 4 | Cardiopulmonary exercise testing data in the control and long COVID groups

CPET data	Controls (n = 14)	Long COVID (<i>n</i> = 14)	P-value
HR at peak (beats/min)	170 ± 17	158 ± 18	0.0619
(% predicted)	97 ± 6	92 ± 9	0.1326
VO ₂ at peak (L/min)	1.78 ± 0.57	1.40 ± 0.34	0.0378
(mL/kg/min)	26.7 ± 7.4	18.6 ± 4.7	0.0015
(% predicted)	91 ± 18	76 ± 18	0.0340
HR/%VO ₂ slope	1.00 ± 0.15	0.77 ± 0.26	0.0114
RER at peak	1.28 ± 0.11	1.27 ± 0.09	0.7483
RPE at peak (median, IQR)	17 (16-18)	19 (18-19)	0.0081*
Dyspnoea at peak (median, IQR)	5 (4-8)	8 (5-9)	0.0515*
SpO ₂ (%) at peak	99 ± 1	99 ± 1	0.8966
VCO ₂ at peak (L/min)	2.29 ± 0.83	1.76 ± 0.52	0.0477
$P_{ET}CO_2$ at peak	39.6 ± 4.7	33.6 ± 3.8	0.0007
O ₂ pulse peak (mL/beat)	10.6±2.8	8.9 ± 2.0	0.0650
Peak SBP (mmHg)	177 ± 21	171 ± 26	0.5115
Peak DBP (mmHg)	88 ± 12	90 ± 15	0.6286
V _E at peak (L/min)	71.5 ± 29.2	66.4 ± 22.4	0.6012
Breathing reserve	43 ± 18	42 ± 24	0.8708
f _B at peak (breaths/min)	36 ± 10	37 ± 9	0.8705
V _T at peak (L)	1.98 ± 0.47	1.83 ± 0.40	0.3440
V_E/VCO_2 ratio at peak	31.4 ± 4.9	38.7 ± 5.4	0.0005
V_E/VO_2 ratio at peak	39.8 ± 7.6	48.4 ± 8.6	0.0095
V _E /VCO ₂ slope	27.7 ± 4.8	37.8 ± 4.4	0.0003
Nadir V _E /VCO ₂	25.0 ± 2.4	30.1 ± 5.0	0.0020
$\mathrm{VO}_{2}\mathrm{at}\mathrm{AT}(\mathrm{mL/kg/min})$	18.7 ± 5.6	12.6 ± 4.3	0.0025
(% of VO ₂ peak)	68 ± 10	66 ± 13	0.6490
P _{ET} CO ₂ at AT (mmHg)	44 ± 4	38 ± 6	0.0041
V _E /VCO ₂ at AT	25.5 ± 2.4	30.4 ± 5.4	0.0042
HR at AT (beats/min	132 ± 13	121 ± 15	0.0453
(% of peak HR)	76±5	77 ± 8	0.6181
Δ from rest (beats/min)	61 ± 18	38±16	0.0009
VO ₂ /WR slope	8.2 ± 1.2	8.2 ± 1.5	0.9283
Heart rate recovery 1 min (beats/min)	19±7	17±5	0.4635

*Mann Whitney U. *HR* heart rate, *VO*₂ volume of inspired oxygen, *RER* respiratory exchange ratio, *RPE* rating of perceived exertion, *SpO*₂ oxygen saturation, *VCO*₂ volume of expired carbon dioxide, *SBP* systolic blood pressure, *DBP* diastolic blood pressure, *VE* minute ventilation, *Bf* breathing frequency, *TV* tidal volume, *VE/VCO*₂ ventilatory efficiency, *AT* anaerobic threshold, *P_{ET}CO*₂ partial pressure of end-tidal carbon dioxide, *WR* work rate. Bold data are where *P* < 0.05.

were also higher in the long COVID group versus controls indicating lower breathing efficiency (a higher minute ventilation to remove a given volume of CO₂) indicating hyperventilation at any point during exercise (Fig. 1 for example slopes (raw data) in a control and long COVID participant). Thirteen percent of the control group had a V_E/VCO₂ slope higher than the normal range (20–30⁴⁶) versus 88% in the long COVID group. Since V_E/VCO₂ has been shown to be higher in people with hypertension; we completed an ANCOVA with clinic SBP as a covariate to check that the slightly elevated SBP in the long COVID group was driving this relationship. The SBP did not adjust the relationship between V_E/VCO₂ and long COVID (SBP effect; *P* = 0.811) where V_E/VCO₂ remained elevated in the long

COVID group versus controls (group effect, P = 0.002 (SPSS v28)). Finally, the $P_{ET}CO_2$ was lower at the AT and at peak exercise in the long COVID group versus the control group (time*group effect; P = 0.0119, mixed model ANOVA), but the magnitude of rise of the $P_{ET}CO_2$ from rest to AT and from rest to peak exercise was the same in both groups (P = 0.0904, Supplementary Fig. 4, mixed model ANOVA).

Hypoxic ventilatory response

Fig. 1a shows examples carotid chemoreflex tests in a control and a long COVID participants and the resultant hypoxic ventilatory responses (calculated from the minute ventilation) in these participants (Fig. 1b). The hypoxic ventilatory response (minute ventilation response to reductions in SpO₂%) was elevated in the long COVID $(-0.44 \pm 0.23 \text{ l/min/ SpO}_2\%)$ $R^2 = 0.77 \pm 0.20$) group compared to controls (-0.17 ± 0.13 l/min/SpO2%, $R^2 = 0.54 \pm 0.38$, Fig. 2a, P = 0.0007); thus, for a given decrease in SpO₂, the participants with long COVID had a greater increase in minute ventilation. This was driven by a greater increase in tidal volume (Fig. 2b) in the participants with long COVID rather than a greater increase in breathing frequency compared to the controls (Fig. 2c). Taken together this indicates that the participants with long COVD have an elevated carotid chemoreflex sensitivity to hypoxia. Since elevated SBP is thought to be linked to heightened HVR - an ANCOVA was completed with clinic SBP as a covariate. The level of clinic SBP did not adjust the relationship between HVR and long COVID (SBP effect; P = 0.503), where having long COVID remained associated with an elevated HVR (group effect adjusted for SBP; P < 0.001 (completed in SPSS v28).

There was no difference in the HR response to hypoxia (Fig. 2d) between groups (control; -0.66 ± 0.35 beats/min/ SpO₂% versus long COVID; -0.58 ± 0.60 beats/min/SpO2%, P = 0.5978). Additionally, there was no difference in the SBP or DBP response to hypoxia between groups (Figs. 2e, 2f). The BP response to hypoxia was variable between individuals with some participants showing a depressor response to hypoxia and others showing a pressor (most common, where the slope of the regression is negative in Figs. 2e, 2f) or biphasic response to hypoxia.

The hypoxic ventilatory response was correlated with the V_E/VCO₂ slope during exercise (r = -0.53, P = 0.0036) and the V_E/VCO₂ nadir (r = -0.45, P = 0.0159; supplementary Fig. 7); indicating that the high carotid chemoreflex sensitivity in the long COVID participants may partially explain the higher V_E/VCO₂ slope (poorer breathing efficiency) during exercise.

Discussion

The pathophysiological mechanisms driving ongoing symptoms in patients with long COVID, after an initial mild infection, are unclear. Here we show for the first time that carotid chemoreflex sensitivity is amplified in nonhospitalised patients with long COVID (versus a control group) and that this was correlated with hyperventilation and poor breathing efficiency during exercise. Elevated carotid chemoreceptor activity could explain several of the ongoing symptoms experienced by patients living with long COVID.

In this population of participants with long COVID, we show that despite similar lung function (resting spirometry and ventilatory reserve during exercise) to the control group, these participants hyperventilate at rest and during exercise. At rest, this is evidenced by a similar minute ventilation compared to control despite a lower resting $P_{ET}CO_2$. Resting levels of arterial CO₂ provides the key stimulus for respiratory drive, mainly via the central chemoreceptors⁴⁷. Reductions in $P_{ET}CO_2$ indicate decreases in PaCO₂^{48,49} which is a stimulus to lower ventilation; however, this has not occurred in the long COVID participants since their level of ventilation was the same as the controls suggesting a resetting of chemoreceptor set-point for breathing. Additionally, the V_E/VCO_2 ratio at rest is higher in the long COVID participants, showing that they are breathing more to remove the same volume of CO₂ as controls. Altered V_E/VCO_2 ratio (and $P_{ET}CO_2$) could result from changes to gas exchange between the alveoli and pulmonary circulation. However, we found that during exercise the magnitude

of the rise in $P_{ET}CO_2$ was the same in the long COVID participants versus the controls. During exercise (with a normal gas exchange) $P_{ET}CO_2$ rises and peaks around anaerobic threshold. When gas exchange is impaired by abnormal ventilation or perfusion of the lung, the $P_{ET}CO_2$ will show smaller rises, no rise, or even decrease as seen in COPD, heart failure and pulmonary hypertension⁵⁰. As such, the similar rise in $P_{ET}CO_2$ with exercise in long COVID patients and control participants suggests no differences in gas exchange between these groups. However, we acknowledge that this needs to be confirmed with gas diffusion studies. A recent study by Beaudry et al. ⁵¹ showed that gas diffusion was normal in long COVID participants versus controls.

We also found evidence of elevated resting V_T/Ti, an index of inspiratory flow, in the long COVID group versus the control group. The V_T /Ti is used as surrogate of primary neural drive for inspiration⁴³ and thus could indicate a higher 'inspiratory drive' at rest in the long COVID participants which may contribute to their feelings of dyspnoea. High inspiratory flow can also indicate changes in airway mechanics⁵², however, this is unlikely in the long COVID group, since resting lung function (spirometry) was similar between groups. During exercise, hyperventilation or poor breathing efficiency was also evidenced by elevated V_F/VCO₂ slopes and an elevated V_E/VCO₂ ratio at anaerobic threshold and peak exercise. Since the carotid chemoreflex plays an important part in the control of breathing both at rest and exercise (including inspiratory drive⁵³); it is possible that it is contributing to hyperventilation in long COVID. However, we did not directly measure PaCO₂, thus the assumption that the lower P_{ET}CO₂ and the higher V_E for a given level of VCO₂ during exercise indicates hyperventilation (and not increased dead space ventilation for example) should be taken with caution⁵⁴.

The mammalian carotid chemoreflex is a protective reflex that contributes to ventilatory and cardiovascular control⁵⁵. In addition to signalling the need for increased ventilation during hypoxia (and other stimuli¹⁸), the carotid body chemoreceptors contribute to resting ventilatory drive, as demonstrated by their tonic activity in humans and animal models^{53,56}. It is well established that certain disease states exhibit exaggerated carotid chemoreflex sensitivity¹⁹ and tonicity¹⁷. Given that elevated carotid chemoreflex sensitivity measured by the hypoxic ventilatory response predicts symptom burden, exercise intolerance, dyspnoea, and hyperventilation in heart failure^{20,57}, we aimed to assess carotid chemoreflex sensitivity in long COVID. The hypoxic ventilatory response was 145% higher in the long COVID group versus age-, BMI- and sex-matched controls indicating amplified carotid chemoreceptor sensitivity in the long COVID group. The higher hypoxic ventilatory response was observed despite a lower resting P_{ET}CO₂ in the long COVID group even though hypocapnia normally depresses carotid chemoreflex sensitivity to hypoxia⁵⁸. The hypoxic ventilatory response was in fact similar to that measured in a group of patients with heart failure reduced ejection fraction in our lab (-0.44 \pm 0.23 L/min/ SpO2% versus -0.48 ± 0.30 L/min/SpO2%; mean \pm SD) using the same equipment before March 2020 (see Supplementary Fig. 5). The elevated hypoxic ventilatory response in the long COVID participants was driven by amplified responses in tidal volume rather than breathing frequency, which supports previous reports in humans where the increase in minute ventilation during mild hypoxia is driven mainly by increases in tidal volume⁵⁹.

The hypoxic ventilatory response was correlated with the V_E/VCO₂ slope, so that a person with high chemoreflex sensitivity had a higher V_E/VCO₂ slope (or vice versa). This suggests that elevated carotid chemoreflex sensitivity may partially explain reduced breathing efficiency and hyperventilation during exercise in participants with long COVID^{13,14,16}. In fact, breathing efficiency was poor in some of the participants in this study; 64% of the long COVID group had a V_E/VCO₂ slope >34 (versus only 7% (one person) in the control group), which is a powerful prognostic indicator of future outcomes for people with heart failure⁶⁰. It is possible that in long COVID the carotid chemoreflex has a similar function as that described in heart failure, partially explaining the dysregulated breathing and feelings of breathlessness in patients with ongoing symptoms after their initial mild infection. Elevated V_E/VCO₂ slopes are also caused by increased dead space

ventilation due to 1) inadequate aeration of the alveoli and 2) poor perfusion of the aerated lung spaces, affecting gas exchange. However, in this study there was no evidence of mechanical lung or small airway issues (due to a similar prevalence of normal breathing reserve in the two groups and similar resting spirometry between the groups). No obvious evidence of gas exchange issues were observed either, because the $P_{\rm ET}CO_2$ during exercise (to AT) increased by a similar magnitude in the participants with long COVID versus the controls.

Finally, the resting $P_{ET}CO_2$ was <30 mmHg in 4 participants with long COVID. Low $P_{ET}CO_2$ causes similar symptoms as those often experienced by patients with long COVID including feelings of brain fog (due to poor cerebral perfusion) and paraesthesia⁶¹. The level of $P_{ET}CO_2$ however did not reach apnoeic threshold levels⁶² in any patients because there were no apnoeas at rest and no evidence of periodic breathing patterns⁶³. Low resting $P_{ET}CO_2$ and hyperventilation is a hallmark of hyperventilatory syndrome which has been cited as a cause of ongoing in symptoms in some cohorts of patients with long COVID⁶⁴. It is possible that problems with breathing at rest and during exercise, leading to lower $P_{ET}CO_2$ levels and manifesting as hyperventilation syndrome could be partially driven by the carotid chemoreflex in patients with long COVID.

Possible mechanisms of increased carotid chemoreceptor sensitivity after SARS-CoV-2 infection include local changes within the parenchyma of the carotid body and/or dysfunction occurring in medullary regions that process afferent sensory information and control efferent ventilatory responses.

In the acute phase of the infection, local viral and immune cell invasion of the carotid body²⁴ could disrupt normal functioning, potentially via immune cells destroying infected glomus cells explaining silent hypoxia in some patients²². In the long-term these cells are likely replaced but there is the possibility of long-term local inflammation and disruption of ACE1 and ACE2 balance leading to elevated carotid chemoreceptor drive. Additionally, since the carotid bodies are highly sensitive to disturbances in perfusion, any blood flow disruption caused by microthrombi and endothelial dysfunction²⁴ could elevate carotid body activity⁶⁵. It is also possible that dysfunction of the petrosal ganglion, which carries afferent sensory input into the brainstem, causes hyper-reactivity in the carotid chemoreflex. Of note, the petrosal ganglion participates in mediating taste signals in the brain. However, only 7% of our long COVID participants reported ongoing impaired sense of taste and smell¹¹. Additionally, inflammation or neuronal damage in the medullary regions mediating afferent signals from the carotid body could contribute to increased chemoreceptor drive. Along these lines there is evidence from animal models that the original SARS-CoV infiltrates medullary brain regions⁶⁶, but there is no strong evidence of this occurring with existing variants of SARS-CoV-2.

Finally, some evidence suggests the SARS-CoV-2 may affect mitochondrial function⁶⁷. The glomus cells in the carotid body have unique mitochondria, which are important for O₂ sensing⁶⁸. Any mitochondrial dysfunction in glomus cells could lead to augmented chemoreflex activity⁶⁸. Since mitochondrial dysfunction could occur in any organ, the exercise intolerance and hyperventilation observed in long COVID could also be driven, for example, by the metaboreceptors in skeletal muscle due to poor mitochondrial function and this needs to be examined.

There are some limitations that need to be considered. Firstly, the control group are a combination of prospectively recruited participants who recovered from a SARS-CoV-2 infection in <4 weeks, and participants who were retrospectively identified from our previous studies (all measures were taken using the same equipment, procedures, and site). Thus 6 of the controls did not have a previous SARS-CoV-2 infection. We cannot rule out that SARS-CoV-2 infection has long lasting effects of the cardiopulmonary system even if individuals recover with no ongoing symptoms. However, our exploratory analyses (see Supplementary Fig. 6) indicates that even without the controls who did not have COVID-19, the carotid chemoreflex sensitivity and $V_{\rm E}/\rm VCO_2$ slope (and $V_{\rm E}/\rm VCO_2$ nadir) remained elevated in the long COVID patients versus the controls who did have COVID-19 (Supplementary Fig. 6). Secondly, we measured the carotid chemoreflex

sensitivity using poikilocapnic hypoxia, thus $P_{\rm ET}CO_2$ decreased after each hypoxic exposure (due to ventilatory adjustments). However, it is unlikely that this affected the ventilatory response to hypoxia because 1) the $P_{\rm ET}CO_2$ decreased only after the breaths that were used as measurements for ventilation and thus could not affect the data and 2) we waited for the $P_{\rm ET}CO_2$ to return to baseline after each hypoxic exposure. Thirdly, this was a single site study with a small sample size and thus needs to be evaluated in a larger population of patients with long COVID. Forth, we did not take blood gases measurements, thus we cannot be sure that the low $P_{\rm ET}CO_2$ observed in our long COVID patients is a true reflection of the low $PaCO_2$ and thus hyperventilation in these patients. Finally, although we see evidence of elevated carotid chemoreceptor sensitivity in patients with long COVID, we need to evaluate whether dampening down the hyperreflexia helps to improve symptoms.

Conclusions

Here, we show for the first time to the best of our knowledge that patients with long COVID have elevated carotid chemoreceptor sensitivity and that this is correlated with poor breathing efficiency or hyperventilation during exercise. Interventions that temper carotid body excitability could be explored as a treatment option for long COVID. Previously our group had shown that P2X3 receptors in the carotid body can be targeted to reduce carotid chemoreflex hyperreflexia in an animal model of hypertension¹⁷ and heart failure⁶⁹, and could be a viable target in humans with long COVID. Gefapixant, an oral P2X3 receptor antagonist, has recently demonstrated efficacy and an acceptable safety profile in chronic cough in phase 3 clinical trials⁷⁰. P2X3 receptors could therefore be a viable target in humans with long COVID.

Data availability

Source data underlying Figs. 1 and 2 are provided as Supplementary Data 1 and Supplementary Data 2 Excel files. The authors intend to share their underpinning research data to maximise reuse and evidence their findings. At the time these data were generated, participants were not asked for their permission to share data 'openly'. The data will be deposited at the University of Bristol Research Data Repository (data.bris.ac.uk/data) where once published, they will be assigned the following https://doi.org/10.5523/bris.2w4fl5nhvpl782hhvpbwsv3zw. A metadata record will be published openly by the repository and this record will clearly state how data can be accessed by bona fide researchers. Access requests will be directed to the Research Data team at Bristol, who will assess the motives of potential data re-users before granting access to the data. No authentic request for access will be refused and re-users will not be charged for any part of this process.

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Author contributions

ECH, AKN, JFRP, HB, AEM and ZA conceived and designed the study. AEM, ZA, HB and ECH recruited participants, collected data, performed the bulk of the analysis and drafted the initial manuscript. JFRP, AKN, KH, AK and APA provided ongoing review of and feedback on analyses over the course of multiple meetings, and reviewed and revised the manuscript.

Competing interests

The authors declare no competing interests.

Additional information

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