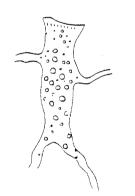
10 Secretion of Growth Factors and Other Regulatory Factors



Drawing (Fig. Id) modified from Sertoli's original article (1865).

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Growth Factors

Insulin-like Growth Factor

Transforming Growth Factor-a/ Epidermal Growth

Factor

Transforming Growth Factor β

Fibroblast Growth Factor

Nerve Growth Factor

Interleukin-1

Additional Testicular Growth Factors

Additional Regulatory Agents

Inhibin/Activin

Müllerian Inhibiting Substance

Sertoli/Leydig Factors

Summary

he Sertoli cell population forms a secretory epither lium that produce several categories of proteins. As previously discussed (see Ch. 7), these secretory products include transport and binding proteins; proteases and antiproteases; and extracellular matrix components (1, 2). These products have critical roles in maintaining the nutritional microenvironment and cytoarchitecture of the seminiferous tubule. Another category of secreted proteins are regulatory agents that are defined as substances that are secreted and subsequently bind to specific receptors to induce a signal transduction event to influence the function, growth or differentiation of a cell on a molecular level (2). These regulatory agents can act as autocrine factors in that they are produced by Sertoli cells and subsequently act on neighboring Sertoli cells. Alternatively, the regulatory agents can act as paracrine factors in that they are produced by Sertoli cells and act on adjacent cell types such as Leydig cells, peritubular myoid cells or developing germinal cells.

One of the major sub-categories of regulatory agents produced by Sertoli cells are growth factors. The properties of a number of the major growth factors identified in the testis are shown in Table 1. Observations regarding the production of the growth factors by Sertoli cells and subsequent actions are reviewed below. The major function for a growth factor is to regulate cell proliferation within a tissue. The ability of a growth factor to influence the differentiated function of a cell may be indirectly related to effects on cell proliferation. Previously growth and differentiation have been shown to be inversely related (3). Therefore, cells produce another sub-category of regulatory agents that are primarily involved in the control of cellular function and differentiation independent of cell growth. The production of a number of these hormone-like substances by Sertoli cells will also be discussed below. The production of these two types of regulatory agents.provides a mechanism for Sertoli cells to maintain and control cell proliferation and differentiated function at various stages of development. The production of these factors by Sertoli cells can influence Sertoli cell growth and differentiation, as well as the other cell types within the testis.

Growth Factors

Precise growth regulation is necessary for the development of the testis and maintenance of spermatogenesis (4). During fetal development all testicular cell types proliferate. The Leydig, peritubular, and Sertoli cells also actively grow in the prepubertal testis. Sertoli cells terminally differentiate and cease to divide at an early stage in pubertal development (5). Peritubular cells continue to proliferate in the adult with a defined turnover rate (6). Leydig cells appear in late fetal development and continue to grow and differentiate before puberty (7) and have a slowed but continuous rate of growth in the adult (6, 8). Germinal cells exhibit a delayed growth pattern, but some development begins shortly after birth when gonadocytes mitotically divide forming spermatogonia. At the onset of puberty germinal cell mitosis and meiosis begins initiating "waves" of spermatogenic cell proliferation. The control of cell proliferation of these various cell types throughout testis development requires the local production and action of various growth factors. Some of these growth factors will likely be Sertoli cell products.

Growth factor		Approx. size (kDa)	Examples of physiological action	Receptor(s)
Insulin-like Growth Factor-I	IGF-I	7.5	skeletal growth	IGF-I receptor
Insulin-like Growth Factor-II	IGF-II	7.5	fetal development	IGF-I and IGF-II
Epidermal Growth Factor	EGF	6	tissue growth	EGF receptor
Transforming Growth Factor Alpha	TGF-α	5	tissue growth	EGF receptor
Transforming Growth Factor Beta(s)	TGF-ß	25/dimer	growth inhibition/tissue repair	TGF- β , type 1, 2, and 3 receptors
Fibroblast Growth Factor	FGF	17	angiogenesis/tissue growth	FGF receptor
Nerve Growth Factor	NGF	13	neuronal development	NGF receptor
Interleukin-1	IL-1	17	immune response/ inflammation	IL-1 receptor

 Table 1

 Properties and Nomenclature of Several Common Growth Factors

Insulin-Like Growth Factors

The insulin-like growth factors (IGF) have structural similarity to insulin (9). IGF-I was previously termed somatomedin C and is considered an essential progression factor for cell growth and DNA synthesis. Production and secretion of IGF-I by the liver accounts for the high levels of IGF-I in serum and interstitial fluid (10). Another member of this family is IGF-II that may act as a growth factor during fetal development.

IGF-I mRNA was originally identified in whole testis (11). All testicular somatic cells have been shown to express and produce IGF-I (12-14) including Sertoli cells (15, 16). In addition, all testicular cell types appear to also respond to IGF. Both Sertoli and germinal cells contain receptors for IGF-I (17-19). IGF-I stimulates DNA synthesis (20) as well as increases transferrin and lactate production in immature Sertoli cells (19, 21). The presence of the blood-testis barrier prevents interstitial fluid-derived IGF-I from directly affecting sequestered germ cells. Thus, Sertoli cell production of IGF-I may allow for paracrine control of germinal cell proliferation. This is further suggested by the presence of IGF-I receptors and immunoreactivity in spermatocyctes and spermatids (22). Local production of IGF binding proteins may act as a mechanism to concentrate local levels of this factor (13). Although IGF-I may influence cell function, proposed cell-cell interactions involving IGF-I need to be questioned since all the somatic cell types are exposed to high levels of liver derived IGF-I present in interstitial fluid.

IGF-II has also been suggested to be involved in local interactions because both Sertoli and germinal cells contain IGF-II receptors (23). IGF-II, however, does not appear to be expressed locally (24). IGF-II appears to have the ability to stimulate Sertoli cell differentiation, perhaps by cross-reacting with IGF-I receptors (20). Neither IGF-I nor IGF-II have been demonstrated to act directly on germ cells and further study is necessary to understand the physiological importance of these factors.

Transforming Growth Factor- α /Epidermal Growth Factor

Transforming growth factor-alpha (TGF- α) is a structurally related member of the epidermal growth factor (EGF) family (25, 26). Due to similar protein structure, these factors act at the same receptor to stimulate cell growth (27). TGF- α is synthesized as a transmembrane precursor which may activate EGF receptors on neighboring cells or be proteolytically cleaved to release mature peptide. TGF- α is produced by non-transformed cells and appears to have an important role as a growth regulator in normal tissues.

EGF has been implicated to be involved in the maintenance of spermatogenesis (28). Removal of the salivary glands, a major site of EGF productions, from mice show 50% reduction of mature sperm and EGF replacement returns spermatogenesis to normal levels (29). Circulating concentrations of EGF, however, are considered too low

to mediate endocrine effects (30). EGF does not appear to be expressed in the rodent testis (31). EGF actions may be mediated by a locally produced EGF-like factor that blocks EGF from binding to its receptor (32). TGF- α is an EGF-like factor that is expressed by both peritubular cells and Sertoli cells, but not germinal cells (31). TGF- α has also been immunohistochemically detected in Leydig cells (33). Scatchard analysis indicates that high-affinity EGF receptors are present on peritubular cells, but not Sertoli cells (31). TGF- α stimulates DNA synthesis and cell division in peritubular cells, but not Sertoli cells (31). Both peritubular and Sertoli cell production of TGF- α may contribute to peritubular cell growth. At present it is unclear whether Sertoli or germinal cells contain functional receptors for EGF. Scatchard analysis and histochemistry do not indicate the presence of receptors on differentiated Sertoli cells (31, 34). However, another report presents immunological evidence that Sertoli cells may contain EGF receptors (35). Sertoli cells and germinal cells were recently found to contain very low, but detectable, levels of EGF-receptor mRNA (36). Recent literature suggests EGF may alter Sertoli cell function, including stimulation of lactate and inhibin production (37, 38). However, some actions of TGF- α /EGF on Sertoli cells may be mediated indirectly by peritubular cell production of other factors (31). The role of TGF- α in Sertoli cell-germ cell interactions is not clear. Speculation that developing spermatogonia respond to TGF- α (39) might provide an appropriate mechanism for Sertoli cells to influence spermatogonial growth. Two models utilizing TGF- α in transgenic mice, which overexpress TGF- α in the testis, show no abnormal features in testis morphology or spermatogenesis (36, 40, 41).

Transforming Growth Factor-B

Transforming growth factor-beta (TGF- β) is a multifunctional regulatory molecule which can stimulate or inhibit aspects of cellular growth and differentiation (42). TGF- β acts by inhibiting the actions of growth factors such as EGF/TGF- α . TGF- β can also promote cellular differentiation, extracellular matrix production and chemotaxis. Different sub-types of TGF- β (TGF- β 1, TGF- β 2, and TGF- β 3 in mammals) are produced as latent secreted precursors. Most cell types contain receptors for this ubiquitous factor.

TGF- β may act as a growth inhibitor in the testis to prevent spermatogonial growth before puberty and to terminate growth of the maturing Sertoli cell. Studies suggest that TGF- β is produced by Sertoli cells and may be modulated by gonadotrophins (39, 43, 44). Interestingly, the testis appears to be one of the few tissues where TGF- β_3 is expressed (45). Sertoli cells express all three forms of TGF- β (46). Sertoli cell TGF- β_1 expression is high in prepubertal animals and declines during puberty to low levels in the adult. TGF- β_2 expression is also high in prepubertal animals and at the onset of puberty in response to FSH is reduced to very low levels (46). TGF- β_3 interestingly is primarily expressed by the Sertoli cells for a short period during development at the onset of puberty until spermatogenesis is inhibited, days 10-15 of rat testis development (46). This pattern of TGF- β expression by Sertoli cells during development suggests a potential Sertoli-germinal cell interactions. Sertoli cell TGF- β 2 expression may be needed to prevent prepubertal germinal cell mitosis. Sertoli cell TGF β -3 expression may be needed to induce spermatogonial development and/or Sertoli cell differentiation at the onset of the spermatogenesis. Although germinal cells contain TGF- β receptors (47), the specific functions of the Sertoli cell TGF- β expression on germinal cells remains to be elucidated.

TGF- β does not appear to dramatically affect immature Sertoli cell growth or cellular differentiation (43). However, TGF- β may be important in regulating environmental interactions necessary for spermatogenesis. TGF- β increases Sertoli cell plasminogen activator production that may be involved in tissue remodeling for germ cell development (48). Due to the antagonistic growth regulation of TGF- α by TGF- β , the local production of TGF- β may act to limit TGF- α actions in the tubule.

Peritubular cells also express and produce TGF-B (43). TGF-B acts as a growth inhibitor for peritubular cells and blocks TGF-induced peritubular proliferation (46). A number of observations suggest that TGF-B is important in peritubular cell differentiation. TGF-B may regulate the production of extracellular matrix components by peritubular cells (43) and increase production of plasminogen activator inhibitor type 1 (PAI-1) by peritubular cells. TGF-ß induces peritubular cell contractility that is potentially required for sperm transport in the tubule (49) and for migration and colony formation of peritubular cells in culture (43). TGF-B-stimulated chemotaxis may be a mechanism to recruit non-differentiated fibroblasts to the exterior of the tubule in development. Therefore, TGF-B may influence morphogenesis and structural formation of the seminiferous tubule. The possibility that Sertoli cellderived TGF-B may act as a paracrine factor for peritubular cells remains to be determined.

TGF- β also may regulate Leydig cell growth and differentiation. Similar to TGF- α , TGF- β inhibits LHinduced steroidogenesis, possibly by decreasing LH receptor binding (50, 51). During development, the growth of maturing Leydig cells slows and may require a growth inhibitor like TGF- β . TGF- β decreases DNA synthesis in a transformed Leydig cell line; however, TGF- β has little affect on Leydig growth in primary culture (44, 52). The local production of TGF- β in the interstitium needs to be determined to elucidate the importance of Sertoli cell derived TGF- β for Sertoli-Leydig cell interactions.

Fibroblast Growth Factor

Fibroblast growth factor (FGF) can influence aspects of both cellular growth and differentiation (53). Aside from growth stimulation, recent studies indicate that FGF may play a critical role in angiogenesis and tissue repair. FGF has many cellular targets and widespread tissue distribution and is important in many organ systems, including the testis (54).

Basic FGF (bFGF) has been isolated from bovine and human testis (55-56). Sertoli cells appear to produce an FGF-like substance (57). Recently, Sertoli cells have been shown to express the FGF gene and secrete basic FGF in response to FSH (58). The angiogenic properties of FGF suggest that this factor may be involved in vascularization of this tissue during development. FGF is mitogenic for immature Sertoli cells (57, 59). The ability of FSH to stimulate Sertoli cell growth may be mediated indirectly by the ability of FSH to stimulate FGF production (58). Basic FGF may also be important in tissue remodeling for spermatogenesis in its ability to stimulate Sertoli cell plasminogen activator activity (59). FGF action on germ cells has not been demonstrated; however, immunolocalization of bFGF in germ cells has been demonstrated (60). Further molecular studies are necessary to localize cellular expression of FGF and its receptor to determine the potential function of Sertoli derived FGF.

Nerve Growth Factor

Nerve growth factor (NGF) is another mitogen which may mediate intercellular interactions involving growth (61). NGF is important for the development and maintenance of sympathetic neurons in the peripheral nervous system and cholinergic neurons in the central nervous system. NGF expression typically correlates with the amount of sympathetic innervation. Surprisingly, NGF is expressed at higher levels than expected in testosteronedependent organs, including the testis.

NGF mRNA is present in spermatocytes and early spermatids of the adult mouse (62, 63), while Sertoli cells express NGF receptor (64). Hypophysectomy increases NGF receptor mRNA in whole testis, while luteinizing hormone (LH), but not FSH replacement returns expression to basal levels. This observation suggests that testosterone down-regulates the receptor and may be an example of a gene that is negatively regulated by androgens. Interestingly, levels of NGF receptor may also correlate with stage VI-VIII of the seminiferous cycle, perhaps stimulating the Sertoli cell for later steps in germ cell maturation. The function of NGF in the testis is not known and requires further study.

Interleukin-1

The interleukins (ILs) are a family of cytokines produced by activated lymphocytes and macrophages. One of these factors, IL-1, may play an important role in mediating cellular activation during inflammation and infection (65). The β form of IL-1 is typically secreted by lymphocytes; however, IL-1 α is produced by non-immune tissues. The mitogenic properties of these factors suggest that IL-1 may mediate growth regulation.

IL-1 α -like activity was isolated from cultures of

mature Sertoli cells, while not found in cultures of other testicular cell types (66, 67). IL-1 activity in conditioned media increases at puberty, coinciding with the onset of spermatogenesis (68). IL-1 is mitogenic for a variety of cell types, thus the Sertoli cell might directly stimulate germ cell development through production of IL-1. One study indicates that intratesticular injection of IL-1 into hypophysectomized rats stimulates [³H]thymidine incorporation in spermatogonia (69). Presently, it is not known which cells contain IL-1 receptors; however, IL-1 can inhibit Leydig steroidogenesis (70). Another potential role for this cytokine is to mediate immune suppression.

Additional Testicular Growth Factors

Other mitogenic factors found in the testis include seminiferous growth factor (SGF) and Sertoli cellsecreted growth factor (SCSGF). SGF was the first mitogenic factor identified in the tubule (71). This 16 kDa mitogen was isolated from Sertoli cells based on its affinity for heparin and appears immunologically distinct from FGF (72, 73). SGF stimulates growth in transformed TM4 Sertoli, TM3 Leydig cells, and 6 day-old mouse Sertoli cells (74). SGF activity has been detected in many species and is predominant during prepubertal development (71). Another Sertoli cell secreted growth factor SCSGF has also been partially purified and appears mitogenic for a number of cell lines (75). SCSGF has some similarities to TGF- α , including its apparent molecular weight of 8 kDa and its ability to displace radiolabelled EGF from binding its receptor (75). Both SGF and SCSGF have not been fully characterized, and whether these factors are previously identified growth factors remains to be thoroughly investigated.

A number of additional growth factors have been shown to act on specific cell types or to be localized in interstitial fluid. The site of production and specific functions of these agents remains to be investigated. Factors such as activin have been shown to influence germ cell proliferation *in vivo* (76). Whether the actions of such agents are direct or indirectly mediated through alterations in the production of more classic growth factors previously discussed remains to be elucidated.

Additional Regulatory Agents

The regulation of cellular function and differentiation is required at various stages of testis development. Sertoli cell and germinal cell differentiation is induced at the onset of puberty and maintained at optimal levels in the adult. Peritubular myoid cells also differentiate at the early stages of pubertal development. Leydig cells undergo an initial stage of differentiation prenatally and then progressively develop throughout pubertal development. The ability of Sertoli derived regulatory agents to control aspects of this process is not understood; however, a number of unique regulatory agents have been shown to be produced by Sertoli cells.

Inhibin/Activin

The Sertoli cells have been known for several decades to produce a regulatory agent termed inhibin (77) that inhibits FSH production by the pituitary (78, 79). Two precursor subunit gene products of inhibin exist, α and β , which upon formation of a mature $\alpha\beta$ dimer forms inhibin to inhibit gonadotrophin production, while a ß dimer forms a molecule termed activin that can stimulate FSH production (80-82). Inhibin and activin are now known to have a wide variety of biological functions and are produced by a number of different tissues. Sertoli cells under the control of FSH or agents that alter cAMP levels produce inhibin (83-88). Although a major function for inhibin is to act on the pituitary to regulate FSH production, potential local actions of inhibin have been postulated. Inhibin and activin both can influence Leydig cell steroidogenesis (89-90). Leydig cells have been postulated to be involved in the regulation of Sertoli cell^{*} inhibin production (91-92). Therefore, inhibin may mediate a regulatory interaction between Sertoli cells and Leydig cells. Leydig cells, however, have also been shown to produce both inhibin and activin (93-96). The ability of both Sertoli cells and Leydig cells to produce inhibin and related peptides questions the relevance of inhibin mediated interactions between the cells. The observation that other cell types, such as the germinal cells (97), may provide additional sites of action for inhibin or activin suggest that understanding the role of inhibin/activin in the testis will require further investigation of sites of action and production. Therefore, the function for Sertoli cell produced inhibin remains to be fully elucidated, but will likely be both an endocrine agent for the pituitary and a paracrine factor within the testis.

Müllerian Inhibiting Substance

Müllerian inhibiting substance (MIS) is a 140 kDa factor that causes regression of the Müllerian ducts during development and has also been referred to as anti-Müllerian hormone (98, 99). MIS was first identified in fetal and neonatal testes (100) and was subsequently found to be produced by Sertoli cells of the neonatal testes (101-102). MIS has been cloned (103) and shown to be a member of the TGF-B superfamily due to sequence similarity. FSH appears to be an important modulator of MIS production by apparently inhibiting MIS production as the Sertoli cell differentiates in response to FSH (104). MIS is primarily produced in the fetal testis with minimal levels in the adult (105). Therefore, a major function for MIS production by neonatal Sertoli cells will be to assist in sexual development and fetal testis differentiation. Whether MIS has additional paracrine roles to modulate germinal cell development or effect cellular functions at later stages of development remains to be elucidated.

Sertoli/Leydig Factors

The ability of Sertoli cells to affect Leydig cell function was first proposed from observations that Leydig cell

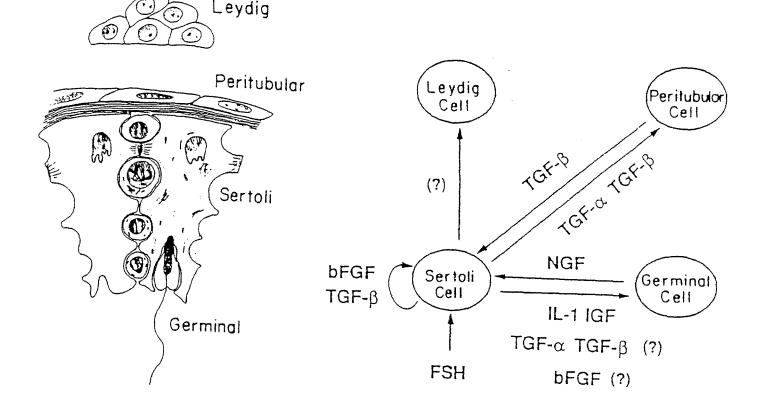
morphology was altered by seminiferous tubules with abnormal function and spermatogenesis (106-107). Damage of the seminiferous tubule with cytotoxic agents, cryptorchidism or pathological conditions causes an altered Leydig cell function and morphology (108-120). Leydig cell morphology also changes with the stage at the seminiferous tubule cycle (121-123). Therefore, the ability of Sertoli cells to produce regulatory agents that influence Leydig cell function has been examined. A number of different laboratories have used conditioned medium from cultures of Sertoli cells or seminiferous tubules. Investigators have found that Sertoli cell conditioned media contains factor(s) that can increase basal and hormone stimulated Leydig cell function (124-136), as well as decrease Leydig cell function (68, 137-143). The specific regulatory agents present in Sertoli cell conditioned media that influence Leydig cell function remain to be purified and characterized. It is likely that several of the growth factors produced by Sertoli cells previously discussed may contribute to the ability of Sertoli cell conditioned media to influence Leydig cell function.

One factor postulated to be produced by Sertoli cells and which can modulate Leydig cell function is an LHRH-like substance (144-149). Leydig cells from some species contain receptors for LHRH (150-153) and LHRH has long term inhibitory effects on Leydig cell steroidogenesis (154, 155). The production of an LHRH-like substance by Sertoli cells, however, has been questioned (156) and appears somewhat species specific. Further analysis of species specificity, sites of production, sites of action and biochemical characterization is required.

Although the production of factors by Sertoli cells to influence Leydig cell function may be needed during development of the testis, the function of such agents in the adult needs further consideration. The concentration of androgen present in the adult testis is significantly higher than the concentrations needed to maintain Sertoli cell or germinal cell function (157-160). Reduction of androgen levels by 80-90% may not have major effects on testis function. Therefore the physiological need to regulate Leydig cell steroidogenesis needs to be carefully considered; however, alternate Leydig cell functions may require a more active regulation by Sertoli cells.

Summary

The literature reviewed indicates that Sertoli cells produce a number of regulatory agents that can have both paracrine and autocrine roles in the regulation of testis cell growth and differentiation. The majority of the regulatory agents shown to be produced by Sertoli cells are growth factors. The integrated actions of various factors such as TGF- α and TGF- β could provide an efficient mechanism to regulate cell proliferation during gonadal development. The potential role that Sertoli cell derived growth factors may have in the regulation of various cell types is shown in Figure 1. Observations obtained imply that growth factors will likely be critical regulatory agents



Secretory Product	Proposed Site Action	Potential Function	
Growth Factors			
IGF-1	Sertoli/germinal/peritubular/Leydig	metabolism/growth	
TGF-α	peritubular/?germinal/?Sertoli	growth stimulation	
TGF-β	peritubular/?germinal/?Sertoli	growth inhibition/cellular differentiation	
IL-1	?germinal	growth regulation	
FGF	Sertoli/?germinal	growth stimulation	
Other Regulatory Agents			
Inhibin	Leydig/pituitary	alter steroidogenesis/regulate FSH	
MIS	fetal gonad	promote gonadal development	
LHRH-like factor	Leydig	decrease steroidogenesis	

 Table 2

 Regulatory Agents Produced by Sertoli Cells

(?) denotes speculated site of action.

involved in gonadal cell-cell interactions. The endocrine regulation of testis growth may be influenced by indirect effects on growth factor production. An example of this is the ability of FSH to increase FGF production and suppress TGF-82 production. Besides growth factors, Sertoli cells also produce a number of regulatory agents that influence cellular function and differentiation. A partial list of the regulatory agents produced by Sertoli cells is shown in Table 2. Although numerous factors have been shown to be produced by Sertoli cells, their physiological roles in regulation of testis growth and differentiation remains to be elucidated. Further analysis of the regulatory agents produced by Sertoli cells will provide insight into the importance Sertoli cells have in the control and maintenance of testis function on a molecular level.

References

- Griswold MD. Protein secretions of Sertoli cells. Int. Rev. Cytol. 1988; 110:133-156.
- Skinner MK. Cell-cell interactions in the testis. Endocrine Rev 1991; 12:45-77.
- Stein GS, Lian JB, Owen TA. Relationship of cell growth to the regulation of tissue-specific gene expression during osteoblast differentiation. FASEB Journal 1990; 4:3111-3123.
- Clermont Y, Perey B. Quantitative study of the cell population of the seminiferous tubules in immature rats. Am J Anat 1957; 100:241-267.
- 5. Orth J. Proliferation of Sertoli cells in fetal and postnatal rats: a quantitative auto-radiographic study. Anat Rec 1982; 203:485-492.
- Teerds KJ, DeRooij DG, Rommerts FFG, Van Der Tweel I, Wensing CJG. Turnover time of Leydig cells and other interstitial cells in the testis of the adult rat. Arch Androl 1989; 23:105-111.
- 7. Lording DW, de Kretser DM. Comparative ultra structural and histochemical studies of the interstitial studies of the rat

testis during fetal and postnatal development. J Reprod Fert 1972; 29:261-275.

- Hardy MP, Zirkin BR, Ewing LL. Kinetic studies on the development of the adult population of Leydig cells in testes of the pubertal rat. Endocrinology 1989; 124:762-770.
- Froesch ER, Schmid C, Schwander J, Zapf J. Actions of insulin-like growth factors. Ann Rev of Physiol 1985; 47:443-467.
- 10. Daughaday WH, Rotwein P. Insulin-like growth factors I and II: peptide, messenger RNA and gene structure, serum, and tissue concentrations. Endocrine Rev 1989; 10:68-91.
- Casella SJ, Smith EP, VanWyk JJ, Joseph DR, Hynes MA, Hoyt EC, Lund PK. Isolation of a rat testis cDNA encoding an insulin-like growth factor I precursor. DNA 1987; 6:325-330.
- Closset J, Gothot A, Sente B, Scippo ML, Igout A, Vandebroeck M, Dombrowicz D, Hennen G. Pituitary hormones dependent expression of insulin-like growth factors I and II in the immature hypophysectomized rat testis. Molr Endocrinol 1989; 3:1125-1131.
- Cailleau J, Vermeire S, Verhoeven G. Independent control of the production of insulin-like growth factor I and its binding protein by cultured testicular cells. Mol Cel Endocrinol 1990; 69:79-89.
- Naville D, Chatelain PC, Avallet O, Saez JM. Control of production of insulin-like growth factor I by pig Leydig and Sertoli cells cultured alone or together: cell-cell interactions. Mol Cel Endocrinol 1990; 70:217-224.
- Chatelain PG, Naville D, Saez JM. Somatomedin-c/insulinlike growth factor I-like material secreted by porcine Sertoli cells in vitro: characterization and regulation. Biochem Biophy Res Comm 1987; 146:1009-1017.
- Smith EP, Svoboda ME, VanWyk JJ, Kierszenbaum AL, Tres LT. Partial characterization of a somatomedin-like peptide from the medium of cultured rat Sertoli cells. Endocrinology 1987; 120:186-193.
- Handelsman DJ, Spaliviero JA, Scott CD, Baxter RC. Identification of insulin-like growth factor-I and its receptor in the rat testis. Acta Endocrinol 1985; 109:543-549.
- Hansson HA, Billig H, Isgaard J. Insulin-like growth factor I in the developing and mature rat testis: immunohistochemical aspects. Biol Reprod 1989; 40:1321-1328.

- Oonk RB, Jansen R, Grootegoed JA. Differential effects of FSH, insulin and insulin-like growth factor-I on hexose uptake and lactate production by rat Sertoli cells. J Cell Physiol 1989; 139:210-218.
- Borland K, Mita M, Oppenheimer CL, Blinderman LA, Massague J, Hall PF, Czech MP. The actions of insulin-like growth factors I and II on cultured Sertoli cells. Endocrinology 1984; 114:240-246.
- Skinner MK. Griswold MD. Multiplication stimulating activity (MSA) can substitute for insulin to stimulate the secretion of testicular transferrin by cultured Sertoli cells. Cell Biol Internat Rep 1983; 7:441-446.
- Tres LL, Smith EP, VanWyk JJ, Kierszebaum AL. Immunoreactive sites and accumulation of somatomedin-c in rat Sertoli-spermatogenic cell cocultures. Exptl Cell Res 1986; 162:33-50.
- O'Brien DA. Gabel CA, Rockett DK, Eddy EM. Receptormediated endocytosis and differential synthesis of mannose 6-phosphate receptors in isolated spermatogenic and Sertoli cells. Endocrinology 1989; 125:2973-2984.
- Murphy LJ, Bell GI, Friesen HG. Tissue distribution of insulin-like growth factor I and II messenger ribonucleic acid in the adult rat. Endocrinology 1987; 120:1279-1282.
- 25. Derynck R. Transforming growth factor alpha. Cell 1988; 54:593-595.
- 26. Carpenter G, Cohen S. Epidermal growth factor. J Biol Chem 1990; 265:7709-7712.
- Carpenter G. Receptors for epidermal growth factor and other polypeptide mitogens. Ann Rev of Biochem 1987; 56:881-914.
- Stastny M. Cohen S. The stimulation of ornithine decarboxylase activity in testes of the neonatal mouse. Biochem Biophys Acta 1972; 261:177-180.
- Tsutsumi O, Kurachi H, Oka T. A physiological role of epidermal growth factor in male reproductive function. Science 1986; 233:975-977.
- Carpenter G, Zendegui J. A biological assay for epidermal growth factor and related polypeptides. Anal Biochem 1986; 153:279-282.
- Skinner MK. Takacs K, Coffey RJ. Transforming growth factor-alpha gene expression action in the seminiferous tubule: peritubular cell-Sertoli cell interactions. Endocrinology 1989; 124:845-854.
- 32. Holmes SD, Spotts G, Smith RG. Rat Sertoli cells secrete a growth factor that blocks epidermal growth factor binding to its receptor. J Biol Chem 1986; 261:4076-4080.
- Teerds KJ, Rommerts FFG, Dorrington JH. Immunohistochemical detection of transforming growth factor-alpha in Leydig cells during development of the rat testis. Mol Cell Endocrinol 1990; 69:R1-R6.
- Stubbs SC, Hargreave TB, Habib FK. Localization and characterization of epidermal growth factor receptors on human testicular tissue by biochemical and immunohisto-chemical techniques. J Endocrinol 1990; 125:485-492.
- Suárez-Quian CA, Dai M, Onada M, Kriss RM, Dym M. Epidermal growth factor receptor localization in the rat and monkey testis. Biol Reprod 1989; 41:921-932.
- Mullaney BP, Skinner MK. Transforming growth factor-alpha and epidermal growth factor receptor expression and action during pubertal development of the seminiferous tubule (1992)(submitted).
- Mallea LE, Machado AJ, Navoroli F, Rommerts FFG. Epidermal growth factor stimulates lactate production and inhibits aromatization in cultured Sertoli cells from immature rats. Int J Androl 1986; 9:201-208.
- Welsh TH, Hsueh AJW. Mechanisms of the inhibitory action of epidermal growth factor on testicular androgen biosynthesis. Endocrinology 1982; 11:1498-1506.
- 39. Mullaney BP, Skinner MK. Hormonal regulation of Sertoli

cell expression of transforming growth factors alpha and beta: potential role in Sertoli cell-germinal cell interactions. (1992a)(submitted).

- Jhappan C, Stahle C, Harkins RN, Fausto N. Smith GH, Merlino GT. TGF-alpha overexpression in transgenic mice induces liver neoplasia and abnormal development of the mammary gland and pancreas. Cell 1990; 61:1137-1146.
- Matsui Y, Halter SA, Holt JT, Gogan BLM, Coffey RJ. Development of mammary hyperplasia and neoplasia in MMTV-TGF-alpha transgenic mice. Cell 1990; 61:1147-1155.
- 42. Roberts AB & Sporn MB. Transforming growth factor beta. Adv Can Res 1988; 51:107-145.
- Skinner MK, Moses HL. Transforming growth factor beta gene expression and action in the seminiferous tubule: peritubular-Sertoli cell interactions. Mol Endocrinol 1989; 3:625-634.
- Benahmed M, Esposito G, Sordoillet C, dePeretti E, Chauvin MA, Ghiglieri C, Revol A, Morera AM. Transforming growth factor beta and its related peptides in the testis: an intragonadal polypeptide control system. Persp Androl 1989; 53:191-201.
- 45. Miller DA, Lee A, Matsui Y, Chen EY, Moses HL, Derynck R. Complementary DNA cloning of the murine transforming growth factor-β₃ (TGF₃) precursor and the comparative expression of TGF₃ and TGFβ₁, messenger RNA in murine embryos and tissues. Molr Endocrinol 1989; 3:1926-1934.
- Mullaney BP, Skinner MK. Transforming growth factor-beta expression and action during the pubertal development of the seminiferous tubule (1992b)(submitted).
- Naz RK, Kumar R. Transforming growth factor B1 enhances expression of 50 kDa protein related to 2'-5' oligoadenylate synthetase in human sperms. J. Cell Physiol. 1991; 146:156-163.
- Nargolwalla C. McCabe D. Fritz IB. Modulation of levels of messenger RNA for tissue-type plasminogen activator in rat Sertoli cells, and levels of messenger RNA for plasminogen activator inhibitor in testis peritubular cells. Mol Cel Endocrinol 1990; 70:73-80.
- Ailenberg M, Tung PS, Fritz IB. Transforming growth factor beta elicits shape changes and increases contractility of testicular peritubular cells. Biol Reprod 1990; 42:499-509.
- Avallet D, Vigier M, Perrard-Sapori MH, Saez JM. Transforming growth factor beta inhibits Leydig cell functions. Bioch and Bioph Res Comm 1987; 146:575-581.
- Lin T, Blaisdell J, Haskell JF. Transforming growth factor beta inhibits Leydig cell steroidogenesis in primary culture. Biochemical and Bioph Res Comm 1987; 137:387-393.
- 52. Gonzalez-Manchon C, Vale W. Activin-A, inhibin and transforming growth factor beta modulate growth of two gonadal cell lines. Endocrinology 1989: 125:1662-1672.
- Gospodarowicz D, Ferrara N, Schweigerer L. Neufeld G. Structural characterization and biological functions of fibroblast growth factor. Endocrine Rev 1987; 8:95-1114.
- Gospodarowicz D, Ferrara N. Fibroblast growth factor and the control of pituitary and gonad development and function. J Steroid Biochem 1989; 32:183-191.
- Ueno N, Naird A, Esch F, Ling N, Guillemin R. Isolation and partial characterization of basic fibroblast growth factor from bovine testis. Mol Cell Endocrinol 1987; 49:189-194.
- Story MT, Sasse J, Kakuska D, Jacobs SC, Lawson RK. A growth factor in bovine and human testes structurally related to basic fibroblast growth factor. J Urol 1988: 140:422-427.
- Smith EP, Hall SH, Monaco L, French FS, Wilson MW, Conti M. A rat Sertoli cell factor similar to basic fibroblast growth factor increases c-fos messenger ribonucleic acid in cultured Sertoli cells. Mol Endocrinol 1989; 3:954-961.
- Mullaney BP, Skinner MK. Basic fibroblast growth factor expression during pubertal development of the seminiferous

tubule (1992c)(submitted).

- Jaillard C, Chatelain PG, Saez JM. *In vitro* regulation of pig Sertoli cell growth and function: effects of fibroblast growth factors and somatomedin-c. Biol Reprod 1987; 37:665-674.
- Mayerhofer A, Russell LD, Grothe C, Rudolf M, Gratzl M. Presence of localization of a 30-kDc basic fibroblast growth factor-like protein in rodent testis. Endocrinology 1991; 129:921-924.
- Yanker BA, Shooter EM. The biology and mechanism of action of nerve growth factor. Ann Rev of Biochem 1982; 51:845-868.
- Olson L. Ayer-LeLievre C, Ebendal T, Sieger A. Nerve growth factor-like immuno-reactivities in rodent salivary glands and testis. Cell Tiss Res 1987; 248:275-286.
- Ayer-LeLievre C, Olson L, Ebendal T, Hallbrook F, Persson H. Nerve growth factor mRNA and protein in the testis and epididymis of mouse and rat. Proc Nat Acad Sci USA 1988; 85:2628-2632.
- Persson H, Ayer-LeLievre C, Soder O, Villar MJ, Metsis M, Olson L. Ritzen M & Hokfelt T. Expression of beta-nerve growth factor receptor mRNA in Sertoli cells down-regulated by testosterone. Science 1990; 247:704-707.
- Durum SK, Schmidt JA, Oppenheim J. Interleukin-1: an immunological perspective. Ann Rev Immunol 1985; 3:263-287.
- Gustafsson K, Soder O, Pollanen P, Ritzen EM. Isolation and partial characterization of an interleukin-1-like factor from rat testis interstitial fluid. J Reprod Immunol 1985; 14:139-150.
- Khan SA, Schmidt K, Hallin P, DiPauli R, DeGeyter C, Nieschlag E. Human testis cytosol and ovarian follicular fluid contain high amounts of interleukin-1-like factor(s). Mol Cell Endocrinol 1988; 58:221-230.
- Syed V, Soder O, Arver S, Lindh M, Khan S, Ritzen EM. Ontogeny and cellular origin of an interleukin-1-like factor in the reproductive tract of the male rat. Int J Androl 1988; 11:437-447.
- Pollanen P, Soder O & Parvinen M. Interleukin-1-alpha stimulation of spermatogonial proliferation *in vivo*. Reprod Fertil Develop 1989; 1:85-87.
- Calkins JH, Sigel MM, Nankin HR, Lin T. Interleukin-1 inhibits Leydig cell steroidogenesis in primary culture. Endocrinology 1988; 123:1605-1610.
- Feig LA, Bellve AR, Erickson NH, Klagsbrun M. Sertoli cells contain a mitogenic polypeptide. Proc Nat Acad Sci USA 1980; 77:4774-4778.
- Feig LA, Kagsbrun M, Bellve AR. Mitogenic polypeptide of the mammalian seminiferous epithelium: biochemical characterization and partial purification. J Cell Biol 1983; 97:1435-1443.
- 73. Bellvé AR, Zheng W. Growth factors as autocrine and paracrine modulators of male gonadal functions. J Reprod Fertil 1989: 85:771-793.
- Bellvé AR, Feig LA. Cell proliferation in the mammalian testis: biology of the seminiferous growth factor. Rec Prog Horm Res 1984; 40:531-561.
- 75. Buch JP, Lamb DL, Lipschultz LI, Smith RG. Partial characterization of a unique growth factor secreted by human Sertoli cells. Fertil Steril 1988; 49:658-665.
- Mather JP, Attie KM, Woodruff TK, Rice GC, Phillips DM. Activin stimulates spermatogonial proliferation in germ-Sertoli cell cocultures from immature rat testis. Endocrinology 1990; 127:3206-3214.
- 77. McCullagh DR. Dual endocrine activity of the testis. Science 1932; 76:19-20.
- 78. Rivier C, Vale W, Rivier J. Studies of the inhibin family of hormonesic review. Hormone Res 1987; 28:104-118.
- Risbridger GP, Robertson DM, deKretser DM. Current perspectives of inhibin biology. Acta Endocrinol 1990;

122:673-82.

- de Jong FH, Robertson DM. Inhibin: 1985 update on action and purification. Molecular and Cellular Endocrinology 1988; 68:555-560.
- Franchimont P, Verstraelen-Proyard J, Hazee-Hagelstein MT, Renard CH, Demoulin A, Bourguignon JP, Hustin J. Inhibin: from concepts to reality. Vitamins and Hormones (New York) 1979; 37:243-250.
- Ying SY. Inhibins, activins and follistatins: gonadal proteins modulating the secretion of follicle-stimulating hormone. Endo Rev 1988; 9:267-293.
- Steinberger A, Steinberger E. Secretion of an FSH inhibiting factor by cultured Sertoli cells. Endocrinology 1976; 99:918-921.
- Bicsak TA, Vale W, Vaughan J, Tucker EM, Cappel S, Hsueh AJ. Hormonal regulation of inhibin production by cultured Sertoli cells. Mol Cell Endocrinol 1987; 49:211-921.
- Morris PL, Vale WW, Cappel S, Bardin CW. Inhibin production by primary Sertoli cell-enriched cultures: regulation by follicle-stimulating hormone, androgens, and epidermal growth factor. Endocrinology 1988; 122:717-725.
- 86. Gonzales GF, Risbridger GP, de Kretser DM. In vitro synthesis and release of inhibin in response to FSH stimulation by isolated segments of seminiferous tubules from normal to adult male rats. Mol Cell Endocrinol 1988; 59:179-185.
- Conti M, Culler MD, Negro-Vilar A. Adenosine receptordependent modulation of inhibin secretion in cultured immature rat Sertoli cells. Mol Cell Endocrinol 1988; 59:255-259.
- Gonzales GF, Risbridger GP, de Kretser DM. The effect of insulin on inhibin production in isolated seminiferous tubule segments from adult rats cultured in vitro. Mol Cell Endocrinol 1989; 61:209-216.
- Hsueh AJ, Dahl KD, Vaughan J, Tucker E, Rivier J, Bardin CW, Vale W. Heterodimers and homodimers of inhibin subunits have different paracrine action in the modulation of luteinizing hormone-stimulated androgen biosynthesis. Proc Natl Acad Sci USA 1987; 84:5082-5086.
- Lin T, Calkins JK, Morris PL, Vale W, Bardin CW. Regulation of Leydig cell function in primary culture by inhibin and activin. Endocrinology 1989; 125:2134-2140.
- Sharpe RM, Kerr JB, Maddocks S. Evidence for a role of the Leydig cells in control of the intratesticular secretion of inhibin. Mol Cell Endocrinol 1988; 60:243-247.
- Drummond AE, Risbridger GP, de Kretser DM. The involvement of Leydig cells in the regulation of inhibin secretion by the testis. Endocrinol 1989; 125:510-515.
- Risberger GP, Clements J, Robertson DM, Drummond AE, Muir J, Berger HG, de Kretser DM. Immuno- and bioactive inhibin and inhibin alpha-subunit expression in rat Leydig cell cultures. Mol Cell Endocrinol 1989; 66:119-122.
- Roberts V, Meunier H. Sawchenko PE, Vale W. Differential production and regulation of inhibin subunits in rat testicular cell types. Endocrinology 1989; 125:2350-2359.
- Shaha C, Morris PL. Chen CL, Vale W, Bardin CW. Immunostainable inhibin subunits are in multiple types of testicular cells. Endocrinology 1989; 125:1941-1950.
- Lee W, Mason AJ, Schwall R, Szonyi E, Mather JP. Secretion of activin by interstitial cells in the testis. Science 1989: 243:396-398.
- van Dissel-Emiliani FM, Grootenhuis AJ, de Jong FH, de Rooij DG. Inhibin reduces spermatogonial numbers in testes of adult mice and Chinese hamsters. Endocrinology 1989: 125:1899-1903.
- Liu MA, Oliff A. TGBB and MIS family of growth regulators. Can Invest 1991; 9:325-336.
- Donahoe PK, Takahashi M, Cleno S, Manganaro TF. Müllerian inhibiting substancein the ovary. Prog Clin Biol Res 1988; 267:153-175.
- 100. Jost A. Problems of fetal endocrinology: the gonadal and

hypophyseal hormones. Recent Prog Hormone Res 1953; 8:379-418.

- 101. Hayashi M, Shimu H, Hayashi K, Trelstad R, Donahoe PK. Immunocytochemical localization of Müllerian inhibiting substance in the rough endoplasmic reticulum and golgi apparatus in Sertoli cells of the neonatal calf testes using a monoclonal antibody. J Histochem Cytochem 1984; 32:649-654.
- 102. Blanchard M. Josso N. Source of anti-Müllerian hormone synthesized by the fetal testis: Mullerian inhibiting activity of fetal bovine Sertoli cells in tissue culture. Pediatric Res 1974; 8:968-971.
- 103. Cate R. Mattaliano R. Hession C, Tizard R, Farber N, Cheung A, Ninfa E, Frey A, Gash D, Chow E, Fisher R, Bertonis J, Torres G, Wallner B, Ramachandran K, Ragin R, Manganaro T, MacLaughlin D, Donahoe PK. Isolation of the bovine and human genes for Müllerian inhibiting substance and expression of the human gene in animal cells. Cell 1986; 45:685-698.
- 104. Kuroda T. Lee M. Hagg C. Powell D, Manganero T, Donahoe PK. Müllerian inhibiting substance ontogeny and its modulation by follicle stimulating hormone in the rat testis. Endocrinology 1990; 127:1825-1832.
- 105. Hutson JM, Ikawa H, Donahoe P. The ontogeny of Müllerian inhibiting substance in the gonads of the chicken. J Pediatric Surg 1981; 16:822-827
- 106. Rich KA, de Kretser DM. Effect of differing degrees of destruction of the rat seminiferous epithelium on levels of serum follicle stimulating hormone and androgen binding protein. Endocrinology 1977; 101:959-968.
- 107. Aoki A, Fawcett DW. Is there a local feedback from the seminiferous tubules affecting activity of the Leydig cells? Biol Reprod 1978; 19:144-158.
- Rich KA, Kerr JB, de Kretser DM. Evidence for Leydig cell dysfunction in rats with seminiferous tubule damage. Mol Cell Endocrinol 1979; 13:123-135.
- 109. Surina MN. Dynamics of development of the interstitial tissue and seminiferous tubules in the testis of human fetuses. Problemy Endokrinologii (Moskva) 1980; 26:41-45.
- 110. Kula K, Romer TE, Włodarczyk WP. Somatic and germinal cells' interrelationship in the course of seminiferous tubule maturation in man. Arch Androl 1980; 4:9-16.
- Steinberger E, Root A, Ficher M, Smith KD. The role of androgens in the initiation of spermatogenesis in man. J Clin Endocrinol Metab 1973; 37:746-750.
- Neaves WB, Johnson L, Petty CS. Seminiferous tubules and daily sperm production in older adult men with varied numbers of Leydig cells. Biol Reprod 1987; 36:301-308.
- 113. Kerr JB, Rich KA, de Kretser DM. Alterations of the fine structure and androgen secretion of the interstitial cells in the experimentally cryptorchid rat testis. Biol Reprod 1979; 20:409-412.
- 114. deKretser DM, Sharpe RM, Swanston IA. Alternations in steroidogenesis and human chorionic gonadotrophin binding in the cryptorchid rat testis. Endocrinology 1979; 105:135-138.
- 115. Risberger GP, Kerr JB, Peake R, Rich KA, de Kretser DM. Temporal changes in rat Leydig cell function after the induction of bilateral cryptorchidism. J Reprod Fert 1981a; 63:415-420.
- 116. Risberger GP, Kerr JB, de Kretser. Evaluation of Leydig cell function and gonadotrophin binding in unilateral and bilateral cryptochidism: evidence for local control of Leydig cell function by the seminiferous tubule. Biol Reprod 1981b; 24:534-540.
- 117. Risberger GP, Kerτ JB, Peake RA, deKretser DM. An assessment of Leydig cell function after bilateral or unilateral efferent duct ligation: further evidence for local control of Leydig cell function. Endocrinology 1981c; 109:1234-1241.

- 118. Bergh A, Damber JE. Local regulation of Leydig cells by the seminiferous tubules. Effect of short-term cryptorchidism. Int J Androl 1984; 7:409-418.
- 119. Jégou B, Laws AO, de Kretser DM. The effect of cryptorchidism and subsequent orchidopexy on testicular function in adult rats. J Reprod Fert 1983; 69:137-142.
- 120. Jégou B, Peake RA, Irby DC, de Kretser DM. Effects of the induction of experimental cryptorchidism and subsequent orchidopexy on testicular function in immature rats. Biol Reprod 1984; 30:179-187.
- 121. Bergh A. Local differences in Leydig cell morphology in the adult rat testis: evidence for a local control of Leydig cells by adjacent seminiferous tubules. Int J Androl 1982; 5:325-330.
- 122. Bergh A. Paracrine regulation of Leydig cells by the seminiferous tubules. Int J Androl 1983; 6:57-65.
- 123. Bergh A. Development of stage-specific paracrine regulation of Leydig cells by the seminiferous tubules. Int J Androl 1985; 8:80-85.
- 124. Benahmed M, Grenot C, Tabone E, Sanchez P, Morera AM. FSH regulates cultured Leydig cell function via Sertoli cell proteins: and *in vitro* study. Bioch Biophys Res Comm 1985; 132:729-734.
- 125. Janecki A, Jakubowiak A, Lukaszyk A. Stimulatory effect of Sertoli cell secretory products on testosterone secretion by purified Leydig cells in primary culture. Mol Cell Endocrinol 1985; 42:235-243.
- 126. Verhoeven G, Cailleau J. A factor in spent media from Sertoli cell-enriched cultures that stimulates steroidogenesis in Leydig cells. Molecular and Cellular Endocrinology 1985; 40:57-68.
- 127. Papadopoulos V, Carreau S, Drosdowsky MA. Effects of seminiferous tubule secreted factor(s) on Leydig cell cyclic AMP production in mature rat. FEBS Letters 1986; 202:74-78.
- 128. Benahmed M, Tabone E, Grenot C, Sanchez P, Chauvin MA, Morera AM. Paracrine control of Leydig cell activity by FSH dependent proteins from Sertoli cells: an *in vitro* study. J Steroid Biochem 1986a; 24:311-315.
- 129. Verhoeven G, Cailleau J. Specificity and partial purification of a factor in spent media from Sertoli cell-enriched cultures that stimulates steroidogenesis in Leydig cells. J Steroid Biochem 1986; 25:393-402.
- 130. Papadopoulos V, Kamtchouing P, Drosdowsky MA, Hochereau de Reviers MT. Carreau S. Adult rat Sertoli cells secrete a factor or factors which modulate Leydig cell function. J of Endocrinology 1987a; 114:459-467.
- Verhoeven G, Cailleau J. A Leydig cell stimulatory factor produced by human testicular tubules. Mol Cell Endocrinol 1987; 49:137-147.
- 132. Perrard-Sapori MH, Chatelain PC, Rogemond N, Saez JM. Modulation of Leydig cell functions by culture with Sertoli cells or with Sertoli cell-conditioned medium: effect of insulin, somatomedin-C and FSH. Mol Cell Endocrinol 1987; 50:193-201.
- Liu YX, Dahl KD. A factor(s) produced by Sertoli cells stimulates androgen biosynthesis by Leydig cells in neonatal rats. Scientia Sinica (Peking)[B] 1988; 31:818-827.
- 134. Carreau S, Papadopoulos V, Drosdowsky MA. Paracrine regulation of Leydig cell aromatase activity in the rat: development with age. Pathologie Biologie (Paris) 1988a; 36:1002-1006.
- Carreau S, Papadopoulos V, Drosdowsky MA. Stimulation of adult rat Leydig cell aromatase activity by a Sertoli cell factor. Endocrinology 1988b; 122:1103-1109.
- 136. Saez JM, Sanchez P, Berthelon MC, Avallet O. Regulation of pig Leydig cell aromatase activity by gonadotropins and Sertoli cells. Biol Reprod 1989; 41:813-820.
- 137. Syed V, Khan SA, Ritzen EM. Stage-specific inhibition of

interstitial cell testosterone secretion by rat seminiferous tubules *in vitro*. Mol Cell Endocrinol 1985; 40:257-264.

- Benahmed M. Morera AM, Chauvin MA. Evidence for a Sertoli cell. FSH-suppressible inhibiting factor(s) of testicular steroidogenic activity. Biochem Biophys Res Comm 1986b; 139:169-178.
- 139. Syed V, Khan SA, Lindh M, Ritzen EM. Ontogeny and cellular origin of a rat seminiferous tubule factor(s) that inhibits LH-dependent testosterone production by interstitial cells in vitro. Internat J Androl 1987; 10:711-720.
- 140. Papadopoulos V, Kamtchouing P, Drosdowsky MA, Carreau S. Spent media from immature seminiferous tubules and Sertoli cells inhibit adult rat Leydig cell aromatase activity. Horm Meta Res 1987b; 19:62-64.
- 141. Takase M, Tsutsui K, Kawashima S. Inhibitory effect of Sertoli cell-cultured media on LH binding to mouse Leydig cells in culture. Endocrinologica Japonica (Tokyo) 1988; 35:285-293.
- 142. Syed V, Lindh M, Khan SA, Ritzén EM. Hormonal regulation of a rat seminiferous tubules in intact and cryptorchid rats. Int J Androl 1989; 6:464-472.
- 143. Vihko KK, Huhtaniemi I. A rat seminiferous epithelial factor that inhibits Leydig cell cAMP and testosterone prodution: mechanism, stage-specific secretion, and partial cryptorchid rats. Mol Cell Endocrinol 1989; 65:119-127.
- 144. Sharpe RM, Fraser HM. Leydig cell receptors for luteinizing hormone releasing hormone and its agonists and their modulation of administration or deprivation of the releasing hormone. Biochem Biophys Res Comm 1980; 95:256-262.
- 145. Sharpe RM, Fraser HM, Cooper I, Rommerts FF. Sertoli-Leydig cell communication via an LHRH-like factor. Nature 1981; 290:785-787.
- 146. Sharpe RM, Fraser HM, Cooper I, Rommerts FF. The secretion, measurement, and function of a testicular LHRH-like factor. Ann New York Acad Sci 1982; 383:272-294.
- 147. Sharpe RM. Intratesticular factors controlling testicular function. Biol Reprod 1984; 30:29-49.
- 148. Sharpe RM. Cooper I. Comparison of the effects on purified Leydig cells of four hormones (oxytocin, vasopressin, opiates and LHRH) with suggested paracrine roles in the testis. J Endocrinol 1987; 113:89-96.
- 149. Saint Pol PS, Hermand E, Tramu G. Paracrine factors in adult rat testis gonadotrophin control of opioids and LHRH-like peptide. Andrologia 1988; 20:173-181.
- 150. Bourne GA, Regiani S, Payne AH, Marshall JC. Testicular GnRH receptors-characterization and localization on inter-

stitial tissue. J Clin Endocrinol Metab 1980; 51:407-409.

- 151. Clayton RN, Katikineni M, Chan V, Dufau ML. Catt KJ. Direct inhibition of testicular function by gonadotrophinreleasing hormone: mediation by specific gonadotrophinreleasing hormone receptors in interstitial cells. Proc Nat Acad Sei USA 1980; 77:4459-4463.
- 152. Lefebvre FA, Reeves JJ, Seguin C, Massicotte J, Labrie F. Specific binding of a potent LHRH agonist in rat testis. Mol Cell Endocrinol 1980; 20:127-131.
- 153. Hedger MP, Risbridger GP, deKretser DM.
 Autoradiographical localization of luteinizing hormone releasing hormone (LHRH) receptors on rat testicular intertubular cells fractionated on Percoll density gradients. Aus J Biol Sci 1985; 38:435-439.
- 154. Sharpe RM, Doogan DG, Cooper I. Direct effects of a luteinizing hormone-releasing hormone agonist on intratesticular levels of testosterone and interstitial fluid formation in intact male rats. Endocrinology 1983a; 113:1306-1313.
- 155. Sharpe RM, Doogan DG, Cooper I. Factors determining whether the direct effects of an LHRH agonist on Leydig cell function *in vivo* are stimulatory or inhibitory. Mol Cell Endocrinol 1983b; 32:57-65.
- 156. Hedger MP, Robertson DM, Tepe SJ, Browne CA, deKretser DM. Degradation of luteinizing hormone-releasing hormone (LHRH) and an LHRH agonist by the rat testis. Mol Cell Endocrinol 1986; 46:59-70.
- 157. Awoniyi CA, Santuilli R, Sprando RL, Ewing LL, Zirkin BR. Restoration of advanced spermatogenic cells in the experimentally regress rat testis: quantitative relationship to testosterone concentration within the testis. Endocrinology 1989a; 124:1217-1223.
- 158. Awoniyi CA, Santulli R, Chandrashekar V, Schanbacher BD, Zirkin BR. Quantitative restoration of advanced spermatogenic cells in adult male rats made azoospermic by active immunization against luteinizing hormone or gonadotrophin-releasing hormone. Endocrinology 1989b; 125:1303-1309.
- Sun Y-T, Irby DC, Robertson DM, deKretser D. The effects of exogenously administered testosterone on spermatogenesis in intact and hypophysectomized rats. Endocrinology 1989; 125:1000-1010.
- 160. Santulli R, Sprando RL, Awoniyi CA, Ewing LL, Zirkin BR. To what extent can spermatogenesis be maintained in the hypophysectomized adult rat testis with exogenously administered testosterone? Endocrinology 1990; 126:95-101.

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