

# Comparative Phenotypic Resolution of Spontaneous, D<sub>2</sub>-Like and D<sub>1</sub>-Like Agonist-Induced Orofacial Movement Topographies in Congenic Mutants With Dopamine D<sub>2</sub> vs. D<sub>3</sub> Receptor “Knockout”

KATSUNORI TOMIYAMA,<sup>1,2</sup> FERGAL N. MCNAMARA,<sup>1</sup> JEREMIAH J. CLIFFORD,<sup>1</sup> ANTHONY KINSELLA,<sup>3</sup> JOHN DRAGO,<sup>4</sup> SARA FUCHS,<sup>5</sup> DAVID K. GRANDY,<sup>6</sup> MALCOLM J. LOW,<sup>7</sup> MARCELO RUBINSTEIN,<sup>8</sup> ORNA TIGHE,<sup>9</sup> DAVID T. CROKE,<sup>9</sup> NORIAKI KOSHIKAWA,<sup>2</sup> AND JOHN L. WADDINGTON<sup>1\*</sup>

<sup>1</sup>Department of Clinical Pharmacology and Institute of Biopharmaceutical Sciences, Royal College of Surgeons in Ireland, St Stephen's Green, Dublin 2, Ireland

<sup>2</sup>Department of Pharmacology and Dental Research Centre, Nihon University School of Dentistry, Tokyo 101-8310, Japan

<sup>3</sup>School of Mathematics, Dublin Institute of Technology, Dublin 8, Ireland

<sup>4</sup>Monash University Department of Medicine, Monash Medical Centre, Victoria 3168, Australia

<sup>5</sup>Department of Immunology, Weizmann Institute of Science, Rehovot 76100, Israel

<sup>6</sup>Department of Physiology and Pharmacology, Oregon Health Sciences University, Portland, Oregon 97201, USA

<sup>7</sup>Vollum Institute, Oregon Health Sciences University, Portland, Oregon 97201, USA

<sup>8</sup>Instituto de Investigaciones en Ingenieria Genetica y Biologia Molecular, Universidad de Buenos Aires, Buenos Aires 1428, Argentina

<sup>9</sup>Department of Biochemistry and Institute of Biopharmaceutical Sciences, Royal College of Surgeons in Ireland, St Stephen's Green, Dublin 2, Ireland

**KEY WORDS** orofacial movements; dopamine receptors; D<sub>2</sub> knockout; D<sub>3</sub> knockout; behavioural phenotype

**ABSTRACT** Using a novel system, the role of D<sub>2</sub>-like dopamine receptors in distinct topographies of orofacial movement was assessed in mutant mice with congenic D<sub>2</sub> vs. D<sub>3</sub> receptor knockout, and compared with findings in D<sub>1A</sub> mutants. Under spontaneous conditions, D<sub>2</sub> mutants evidenced increased vertical jaw movements and unaltered horizontal jaw movements, with reductions in tongue protrusions and incisor chattering; in D<sub>3</sub> mutants, only incisor chattering was reduced. Given previous evidence that D<sub>1A</sub> mutants show reduced horizontal but not vertical jaw movements, this indicates that apparent oppositional D<sub>1</sub>-like:D<sub>2</sub>-like interactions in the regulation of composited jaw movements may in fact reflect the independent actions of D<sub>2</sub> receptors to inhibit vertical jaw movements and of D<sub>1A</sub> receptors to facilitate horizontal jaw movements. Effects of the D<sub>2</sub>-like agonist RU 24213 to exert greater reduction in horizontal than in vertical jaw movements were not altered prominently in either D<sub>2</sub> or D<sub>3</sub> mutants. The D<sub>1</sub>-like agonists A 68930 and SK&F 83959 induced vertical jaw movements, tongue protrusions, and incisor chattering; induction of tongue protrusions by A 68930 was reduced in D<sub>2</sub> mutants. D<sub>2</sub> receptors exert topographically specific regulation of orofacial movements in a manner distinct from their D<sub>1A</sub> counterparts, while D<sub>3</sub> receptors exert only minor regulation of such movements. **Synapse 51:71–81, 2004.** © 2003 Wiley-Liss, Inc.

Contract grant sponsors: Nihon University (research grant to KT, NK), the Sato Fund (to NK), the Promotion and Mutual Aid Corporation for Private Schools of Japan (to NK), the Ministry of Education, Culture, Sports, Science & Technology, Japan (promotion of multidisciplinary research projects to KT, NK, JLW); Contract grant numbers: 14370609 (to NK), 14571802 (to KT); Contract grant sponsors: Science Foundation Ireland, Galen Fellowship from the Irish Brain Research Foundation, the Stanley Medical Research Institute, and the Research Committee of the Royal College of Surgeons in Ireland (to KT, FNM, JJC, DTC, JLW), in the Institute of Biopharmaceutical Sciences under the Higher Education Authority's Programme for Research in Third Level Institutions, Logan Research Fellowship from Monash University (to JD); Contract grant number: DA12062 (to DKG); Contract grant sponsor: the National Institute of Mental Health; Contract grant number: NO1 MH30003.

\*Correspondence to: Dr. John L. Waddington, Department of Clinical Pharmacology, Royal College of Surgeons in Ireland, St Stephen's Green, Dublin 2, Ireland. E-mail: jwadding@rcsi.ie

Received 21 May 2003; Accepted 2 September 2003

DOI 10.1002/syn.10284

## INTRODUCTION

In accordance with their origins in molecular biology rather than in classical functional considerations, attributing specific roles to individual members of the D<sub>1</sub>-like (D<sub>1A</sub>/D<sub>1</sub>, D<sub>1B</sub>/D<sub>5</sub>) and D<sub>2</sub>-like (D<sub>2L</sub>/S, D<sub>3</sub>, D<sub>4</sub>) families of dopamine (DA) receptors (Missale et al., 1998) in the regulation of behaviour remains problematic (Waddington et al., 1995, 2001). While they appear to exert important roles in the regulation of orofacial movements, with a particular focus on how these are regulated by the D<sub>1</sub>-like family, the D<sub>2</sub>-like family can also influence such processes, both independently and especially via D<sub>1</sub>-like:D<sub>2</sub>-like interactions (Rosengarten et al., 1983, 1986; Murray and Waddington, 1989; Collins et al., 1991; Waddington et al., 1994, 1998; Niznik et al., 2002).

In contrast to the availability of both agonists and antagonists which are highly selective for, and hence can discriminate readily *between*, these D<sub>1</sub>-like vs. D<sub>2</sub>-like receptor families, there are very few agents which can discriminate materially *within* each of these families; thus, the functions of D<sub>1</sub>-like and D<sub>2</sub>-like receptors are understood primarily at a "family" level only (Waddington et al., 1998, 2001). Over recent years, recombinant DNA techniques have been applied by several groups to construct mice with targeted gene deletion (knockout) of individual DA receptor subtypes (Sibley, 1999; Waddington et al., 2001). Yet their potential to clarify the roles of individual DA receptor subtypes in regulating orofacial movements remains to be realised; indeed, systematic assessment of such movements is only now being undertaken even in normal mice because of practical issues: mice are considerably smaller than rats and their orofacial movements more rapid, making for problems in assessment. These difficulties are exacerbated by considerable controversy, based primarily on data in rats, as to how orofacial movements should be defined phenomenologically and resolved empirically; generic terms such as "vacuous chewing" enjoy widespread usage despite uncertainty as to their relevance at a physiological level (Waddington, 1990; Waddington et al., 1998; Tomiyama et al., 2001, 2002).

For these reasons, we recently developed a novel system combined with a physiologically based approach to categorisation and quantification for the assessment of orofacial movement topography in mice (Tomiyama et al., 2001). This has been applied to describe the phenotype of orofacial movements and topographical responses to D<sub>2</sub>-like and D<sub>1</sub>-like agonists in mice with congenic D<sub>1A</sub> receptor knockout (Tomiyama et al., 2002). We now describe the application of this technique to characterise topographically, in a complementary, comparative manner, the phenotype of orofacial movements and topographical responses to the D<sub>2</sub>-like agonist RU 24213 and the D<sub>1</sub>-like agonists A 68930 and SK&F 83959 in mice with incipient con-

genic D<sub>2</sub> receptor knockout and congenic D<sub>3</sub> receptor knockout.

## MATERIALS AND METHODS

### Animals

The original F2 hybrid strain (129/Sv × C57BL/6J) containing the mutated D<sub>2</sub> receptor allele was generated as reported previously (Kelly et al., 1997). In outline, the targeted gene deletion was constructed in 129/Sv embryonic stem cells and male chimaeras mated with C57BL/6J females to produce heterozygous mutants (D<sub>2</sub><sup>+/-</sup>); homozygous mutants (D<sub>2</sub><sup>-/-</sup>) and wildtype (D<sub>2</sub><sup>+/+</sup>) littermates were identified among the progeny of heterozygous intermatings using polymerase chain reaction (PCR) analysis of isolated tail DNA. An incipient congenic D<sub>2</sub> line was established by backcrossing D<sub>2</sub><sup>+/-</sup> to wildtype C57BL/6 for five generations (Kelly et al., 1998). Incipient congenic D<sub>2</sub><sup>+/-</sup> mutants were transported to Dublin, where homozygous mutants (D<sub>2</sub><sup>-/-</sup>) and wildtype (D<sub>2</sub><sup>+/+</sup>) littermates were bred and genotyped by similar PCR of isolated tail DNA among the progeny of heterozygous intermatings (Clifford et al., 2001).

The original F2 hybrid strain (129/Sv × C57BL/6) containing the mutated D<sub>3</sub> receptor allele was generated as reported previously (Accili et al., 1996). In outline, the targeted gene deletion was constructed in 129/Sv embryonic stem cells and male chimaeras mated with C57BL/6 female to produce heterozygous mutants (D<sub>3</sub><sup>+/-</sup>); homozygous mutants (D<sub>3</sub><sup>-/-</sup>) and wildtypes (D<sub>3</sub><sup>+/+</sup>) were identified among the progeny of heterozygous intermatings using PCR analysis of isolated tail DNA. To establish an essentially congenic line of D<sub>3</sub> knockouts, heterozygous mutants of this hybrid (129/Sv × C57BL/6) strain were backcrossed to wildtype C57BL/6 for seven generations. Heterozygous mutants of this seventh generation were then shipped to Dublin; here, this procedure was continued for an additional seven generations, giving a total of 14 backcrosses to wildtype C57BL/6 (McNamara et al., 2002). Analysis of isolated tail DNA was used similarly to identify congenic, homozygous mutants and wildtypes among the progeny of heterozygous intermatings.

Animals were housed in groups of 3–5 with food and water available *ad libitum* and were maintained at 21.0 ± 0.1°C on a 12/12 h (07.00 on; 19.00 off) light/dark schedule. Young adult mice from litters of the same generational age were used in behavioural assessments. These studies were approved by the Research Committee of the Royal College of Surgeons in Ireland and were conducted under license from the Department of Health in accordance with Irish legislation and European Communities Council Directive 86/609/EEC for the care and use of experimental animals.

### Restrictor system

As described previously (Tomiyama et al., 2001), the system consisted of a "restrictor," by which mice were lightly restrained around the neck by a clear perspex collar attached to a horizontal platform; this allowed visual observation to be focused onto the orofacial region with minimal disturbance to movements other than locomotion, rearing, and grooming. Circular collars were composed of two semicircular elements: one fixed to the platform and constituting a trough into which the neck was positioned; the other, inserted from above, completed light enclosure of the neck. Both the diameter of the collar and its height above the platform were adjustable according to body size, to allow a comfortable posture to be maintained. A piece of absorbent paper was spread over the platform of the restrictor. To facilitate observation of the orofacial region, small mirrors were fixed in inclined positions just under the snout of each mouse and lighting directed appropriately to illuminate the mouth. For each experimental session, five mice were placed individually into identical "restrictors," each separated by cardboard dividers to minimise visual and auditory disruption. The observer viewed each animal through slits in a cardboard screen in front of the array of "restrictors"; these slits were positioned optimally in relation to the mouth, mirrors, and illumination.

### Assessment of orofacial movement topography

On the basis of the natural repertoire of behaviours of the mouse at an ethological level, together with dental physiology, orofacial movement topography was categorised into the following four elements: vertical jaw movements, horizontal (lateral) jaw movements, tongue protrusions, and chattering (high-frequency rhythmical jaw movements with incisor tapping) (Tomiyama et al., 2001); general head movements and vibrissae movements were also recorded.

A rapid time-sampling behavioural checklist technique, used previously to resolve the topography of general exploratory and DA agonist-induced behaviour in knockout mice in an ethologically based, unrestricted paradigm (Clifford et al., 1998, 1999, 2000, 2001; Ross et al., 2000; McNamara et al., 2002, 2003), was applied similarly to resolve the topography of orofacial movement (Tomiyama et al., 2001, 2002): each of five mice was observed sequentially for 5-sec periods at 25-sec intervals; for each mouse, the presence or absence of each individual element (occurring alone or in any combination) was determined in each of the 5-sec periods. For assessment of spontaneous orofacial movement topography and its habituation profile, assessments commenced immediately after placement in restrictors and continued for 30-min periods over a total duration of 210 min; mice were used on a single occasion only. For assessment of orofacial movement topography in challenge studies, mice were habituated to

restrictors for a period of 3 h before administration of drug or vehicle, with assessments beginning thereafter over a total duration of 1 h; mice were used on two occasions only, separated by a drug-free interval of at least 1 week, with random allocation to one of the various treatments in each instance. All observations were made by a dentist (KT) experienced also in rodent psychopharmacology who was unaware of genotype and treatment given to each animal.

### Drugs

The selective D<sub>2</sub>-like agonist RU 24213 (*N-n*-propyl-*N*-phenyl-*p*-3-hydroxyphenylethylamine; Hoechst-Marion-Roussel, France) was dissolved in distilled water. The D<sub>1</sub>-like agonist A 68930 (([1*R*,3*S*]-1-aminomethyl-5,6-dihydroxy-3-phenylisochroman; Abbott Laboratories, North Chicago, IL, USA) was dissolved in dilute acetic acid and made up to volume with distilled water. The selective D<sub>1</sub>-like agent SK&F 83959 (3-methyl-6-chloro-7,8-dihydroxy-1-[3-methyl-phenyl]-2,3,4,5-tetrahydro-1*H*-3-benzazepine; Research Biochemicals International, Natick, MA / NIMH Chemical Synthesis Program, Bethesda, MD, USA) was dissolved in distilled water. Drugs or vehicle were injected subcutaneously into the flank in a volume of 2 ml/kg.

### Data analysis

For determination of habituation profiles of spontaneous orofacial movement topographies, total counts for each individual element were summed separately over the following periods: 0–30, 60–90, 120–150, 180–210 min. In drug challenge studies, these counts were summed over 0–60 min after the habituation period and subsequent drug administration. Data were expressed as mean  $\pm$  SEM and analysed using repeated-measures analysis of variance (ANOVA) after square-root transformation in the absence of appropriate non-parametric techniques for interaction terms. Individual group comparisons were then made using Student's *t*-test or Kruskal-Wallis nonparametric one-way ANOVA and Mann-Whitney *U*-test (McNamara et al., 2002, 2003; Ross et al., 2000; Tomiyama et al., 2001, 2002).

## RESULTS

### General parameters: spontaneous behaviour

On examining 40 (20 male, 20 female) incipient congenic D<sub>2</sub> mutants for spontaneous orofacial topography, mean body weight ( $18 \pm 1$  g; mean age  $114 \pm 4$  days) was significantly reduced ( $-22\%$ ;  $P < 0.001$ ) relative to 38 (18 male, 20 female) wildtype controls ( $23 \pm 1$  g; mean age  $116 \pm 6$  days). On qualitative inspection of posture, reactivity to handling, and general activity, no gross motor phenotype was apparent. These findings were as noted previously for D<sub>2</sub> mutants of this incipient congenic line (Kelly et al., 1997, 1998; Clifford et al., 2001).

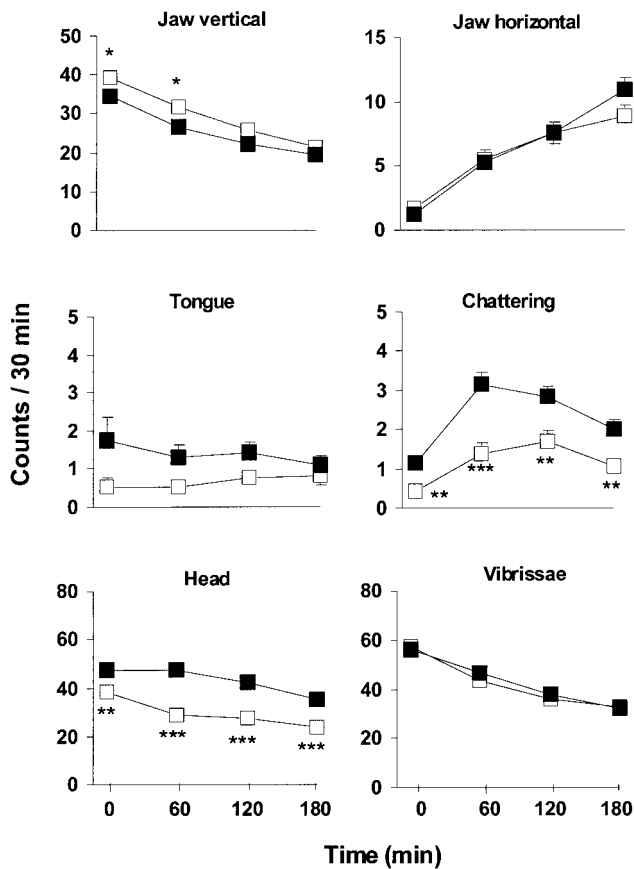


Fig. 1. Phenotype of spontaneous orofacial movement topographies in wildtypes ( $n = 38$ , filled squares) and incipient congenic  $D_2$  mutants ( $n = 40$ , open squares). Data are mean counts  $\pm$  SEM for vertical and horizontal jaw movements, tongue protrusions, incisor chattering, and general head and vibrissae movements over 30-min periods beginning at 0, 60, 120, and 180 min after commencing observations. \* $P < 0.05$ , \*\* $P < 0.01$ , \*\*\* $P < 0.001$  vs. wildtypes.

On examining 20 (10 male, 10 female) congenic  $D_3$  mutants for spontaneous orofacial topography, mean body weight ( $23 \pm 1$  g; mean age  $113 \pm 5$  days) did not differ from 20 (10 male, 10 female) wildtype controls ( $22 \pm 1$  g; mean age  $109 \pm 6$  days). On qualitative inspection of posture, reactivity to handling, and general activity, no gross motor phenotype was apparent. These findings were as noted previously for  $D_3$  mutants both on a mixed (Accili et al., 1996; Xu et al., 1997) and on a congenic (McNamara et al., 2002) genetic background.

### Spontaneous orofacial topography over habituation

#### $D_2$ mutants

In wildtypes, vertical jaw movements were initially prominent but declined subsequently over the habituation period (Fig. 1). Congenic  $D_2$  mutants evidenced a small overall increase in such movements, which habituated similarly (overall effect of genotype,  $F(1,74) = 4.71$ ,  $P < 0.05$ ; no genotype  $\times$  time interaction). Con-

versely, horizontal jaw movements occurred initially at a low level but increased markedly thereafter in wildtypes. This profile was unaltered in  $D_2$  mutants (no overall effect of genotype or time  $\times$  genotype interaction). Tongue protrusions and incisor chattering occurred at relatively low levels throughout habituation in wildtypes. These movements were reduced in  $D_2$  mutants (overall effects of genotype: tongue protrusions,  $F(1,74) = 12.10$ ,  $P < 0.001$ ; incisor chattering,  $F(1,74) = 30.28$ ,  $P < 0.001$ ; no time  $\times$  genotype interactions).

General head movements were initially prominent but then declined over habituation in wildtypes. In  $D_2$  mutants these movements were reduced and habituated more rapidly over the early period of assessment (overall effect of genotype,  $F(1,74) = 27.22$ ,  $P < 0.001$ ; time  $\times$  genotype interaction,  $F(3,222) = 4.95$ ,  $P < 0.01$ ). General movements of the vibrissae were initially prominent and declined over the habituation period. This profile was unaltered in  $D_2$  mutants (no overall effect of genotype or time  $\times$  genotype interaction).

In relation to gender, tongue protrusions and incisor chattering were more prominent in males of both genotypes (overall effects of gender: tongue protrusions,  $F(1,74) = 10.84$ ,  $P < 0.01$ ; incisor chattering,  $F(1,74) = 4.14$ ,  $P < 0.05$ ; no genotype  $\times$  gender interactions), while the rate of habituation of general vibrissae movements was more rapid in males of both genotypes (time  $\times$  gender interaction,  $F(3,222) = 3.78$ ,  $P < 0.05$ ; no time  $\times$  gender  $\times$  genotype interaction). No other effects of gender were encountered.

#### $D_3$ mutants

In wildtypes, vertical jaw movements, horizontal jaw movements, and tongue protrusions evidenced habituation profiles (Fig. 2) similar to those described above, with this profile not altered materially in congenic  $D_3$  mutants (no overall effects of genotype or time  $\times$  genotype interactions) other than some transient reduction in horizontal jaw movements. The habituation profile of incisor chattering in wildtypes was also similar to that described above, while in  $D_3$  mutants the level of this movement was decreased (overall effect of genotype,  $F(1,36) = 9.48$ ,  $P < 0.01$ ). General head and vibrissae movements were initially prominent and declined over habituation in wildtypes, as described above. This profile was not altered materially in  $D_3$  mutants (no overall effects of genotype or time  $\times$  genotype interactions), other than some transient increase in head movements.

In relation to gender, vertical jaw movements were less prominent in males than in females for both genotypes (overall effect of gender,  $F(1,36) = 9.14$ ,  $P < 0.01$ ; no genotype  $\times$  gender interaction), while horizontal jaw movements showed more rapid increase with time in males than in females for both genotypes (no overall



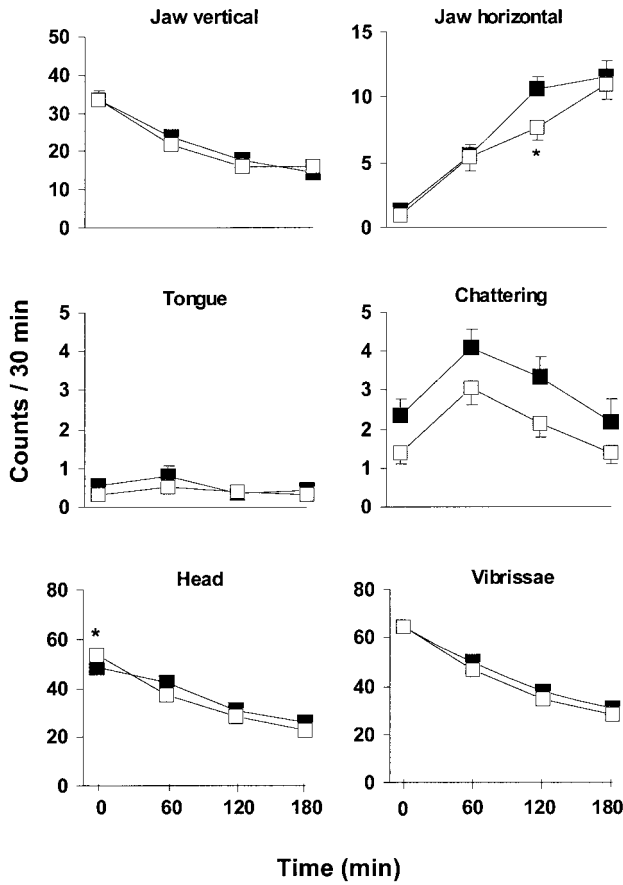


Fig. 2. Phenotype of spontaneous orofacial movement topographies in wildtypes (n = 20, filled squares) and congenic D<sub>3</sub> mutants (n = 20, open squares). Data are mean counts ± SEM for vertical and horizontal jaw movements, tongue protrusions, incisor chattering, and general head and vibrissae movements over 30-min periods beginning at 0, 60, 120, and 180 min after commencing observations. \**P* < 0.05 vs. wildtypes.

effect of gender or genotype × gender interaction; time × gender interaction,  $F(3,108) = 4.43$ ,  $P < 0.01$ ; no time × gender × genotype interaction). The time course of incisor chattering in males relative to females differed marginally between the genotypes (no time × genotype interaction; time × genotype × gender interaction,  $F(3,108) = 2.87$ ,  $P < 0.05$ ). Apart from the isolated instances noted above, there were no other overall effects of gender, genotype × gender, or time × genotype × gender interactions.

**General parameters: RU 24213 challenge**

On examining eight male incipient congenic D<sub>2</sub> mutants for orofacial topography in response to RU 24213, mean body weight (15 ± 1 g; mean age 86 ± 7 days) was reduced (−38%;  $P < 0.001$ ) relative to 12 male wildtype controls (24 ± 1 g; mean age 91 ± 7 days).

On examining 10 male congenic D<sub>3</sub> mutants for orofacial topography in response to RU 24213, mean body weight (23 ± 1 g; mean age 89 ± 2 days) did not differ from 10 male wildtype controls (23 ± 1 g; mean age

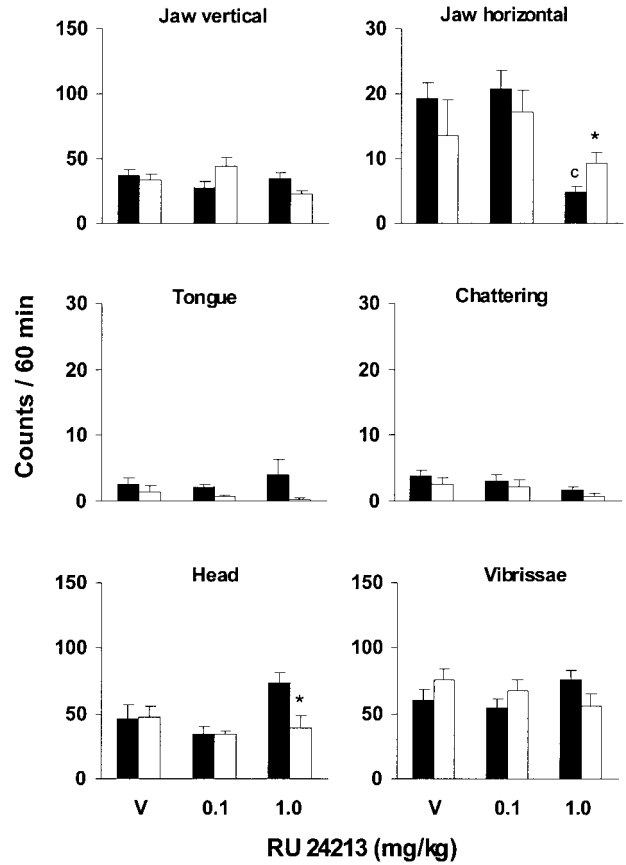


Fig. 3. Phenotype of orofacial movement topographies in wildtypes (n = 8, filled columns) and incipient congenic D<sub>2</sub> mutants (n = 5–6, open columns) following challenge with 0.1–1.0 mg/kg RU 24213 or vehicle (V). Data are mean counts ± SEM for vertical and horizontal jaw movements, tongue protrusions, incisor chattering, and general head and vibrissae movements over a 60-min period. <sup>c</sup>*P* < 0.001 vs. vehicle; \**P* < 0.05 vs. wildtypes receiving same dose.

101 ± 5 days); the age of D<sub>3</sub> mutants was slightly less than wildtype counterparts (−12%,  $P < 0.05$ ).

**RU 24213-induced orofacial topography D<sub>2</sub> mutants**

In wildtypes, RU 24213 did not exert any consistent effect on vertical jaw movements (Fig. 3). In congenic D<sub>2</sub> mutants this profile was not altered materially, although there were some subtle differences between the genotypes in relation to dose (no overall effects of dose or genotype; dose × genotype interaction,  $F(1,36) = 7.80$ ,  $P < 0.01$ ). Horizontal jaw movements were reduced by increasing doses of RU 24213. This effect did not differ prominently between the genotypes (overall effect of dose,  $F(1,36) = 6.61$ ,  $P < 0.02$ ; no overall effect of genotype or dose × genotype interaction), although it appeared marginally attenuated in D<sub>2</sub> mutants at the higher dose of RU 24213. Tongue protrusions were not influenced by RU 24213. These movements were reduced among D<sub>2</sub> mutants relative to wildtypes in a manner unrelated to treatment (no

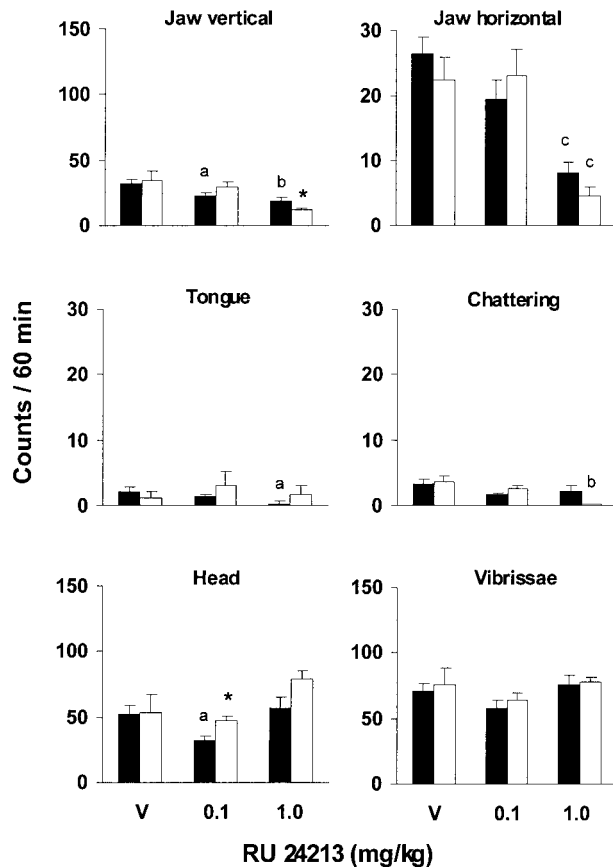


Fig. 4. Phenotype of orofacial movement topographies in wildtypes ( $n = 6-8$ , filled columns) and congenic  $D_3$  mutants ( $n = 6-7$ , open columns) following challenge with 0.1–1.0 mg/kg RU 24213 or vehicle (V). Data are mean counts  $\pm$  SEM for vertical and horizontal jaw movements, tongue protrusions, incisor chattering, and general head and vibrissae movements over a 60-min period. <sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$ , <sup>c</sup> $P < 0.001$  vs. vehicle; \* $P < 0.05$  vs. wildtypes receiving same dose.

overall effect of dose or dose  $\times$  genotype interaction; overall effect of genotype,  $F(1,36) = 5.26$ ,  $P < 0.05$ ). Incisor chattering was not influenced materially by RU 24213 (no overall effects of dose or of genotype, or dose  $\times$  genotype interaction). RU 24213 exerted only subtle, biphasic effects on head movements, which did not differ materially between the genotypes (overall effect of dose,  $F(1,36) = 4.17$ ,  $P < 0.05$ ; no overall effect of genotype or dose  $\times$  genotype interaction), although the effect of the higher dose of RU 24213 appeared marginally attenuated in  $D_2$  mutants. There were no alterations in vibrissae movements (no overall effects of dose or of genotype, or dose  $\times$  genotype interaction).

### $D_3$ mutants

In wildtypes, RU 24213 reduced both vertical and particularly horizontal jaw movements, with a low baseline level of tongue protrusions being reduced further (Fig. 4). In congenic  $D_3$  mutants this profile was not altered materially (overall effects of dose: vertical jaw movements,  $F(2,34) = 15.29$ ,  $P < 0.001$ ; horizontal

jaw movements,  $F(2,34) = 28.76$ ,  $P < 0.001$ ; no effects of genotype or dose  $\times$  genotype interactions), other than a marginally greater effect on vertical jaw movements in those receiving the highest dose of RU 24213. Incisor chattering was reduced by RU 24213, somewhat more prominently in  $D_2$  mutants than in wildtypes (overall effect of dose,  $F(2,34) = 8.57$ ,  $P < 0.001$ ; no overall effect of genotype; dose  $\times$  genotype interaction,  $F(2,34) = 3.76$ ,  $P < 0.05$ ). RU 24213 exerted a biphasic effect on head movements, which did not differ materially between the genotypes (overall effect of dose,  $F(2,34) = 6.05$ ,  $P < 0.01$ ; no overall effect of genotype or dose  $\times$  genotype interaction), although the action of the lower dose of RU 24213 to reduce such movements appeared marginally attenuated in  $D_3$  mutants. There were no alterations in vibrissae movements (no overall effects of dose or of genotype, or dose  $\times$  genotype interaction).

### General parameters: A 68930 challenge

On examining 10 male incipient congenic  $D_2$  mutants for orofacial topography in response to A 68930, mean body weight ( $20 \pm 1$  g; mean age  $168 \pm 12$  days) was reduced ( $-29\%$ ;  $P < 0.001$ ) relative to 10 male wildtype controls ( $28 \pm 1$  g; mean age  $169 \pm 10$  days). Limited availability precluded A 68930 challenge studies in  $D_3$  mutants.

### A 68930-induced orofacial topography

In wildtypes, A 68930 readily induced vertical jaw movements (Fig. 5). This response was unaltered in incipient congenic  $D_2$  mutants (overall effect of dose,  $F(2,32) = 26.32$ ,  $P < 0.001$ ; no overall effect of genotype or dose  $\times$  genotype interaction). There was no induction of horizontal jaw movements in either genotype (no overall effects of dose or of genotype, or dose  $\times$  genotype interaction), with a marginal reduction in baseline levels in vehicle-treated mutants. A 68930 readily induced tongue protrusions in wildtypes. This effect was substantially diminished in  $D_2$  mutants at the higher dose of A 68930 (overall effect of dose,  $F(2,34) = 21.21$ ,  $P < 0.001$ ; overall effect of genotype,  $F(1,34) = 5.64$ ,  $P < 0.05$ ; dose  $\times$  genotype interaction,  $F(2,34) = 6.26$ ,  $P < 0.01$ ). While A 68930 readily induced incisor chattering in both genotypes, levels were slightly reduced in  $D_2$  mutants relative to wildtypes across all treatment groups (overall effect of dose,  $F(2,34) = 36.22$ ,  $P < 0.001$ ; overall effect of genotype,  $F(1,34) = 4.74$ ,  $P < 0.05$ ; no dose  $\times$  genotype interaction). There was no consistent induction of general head movements by A 68930 in either genotype, although at the higher dose these were reduced in  $D_2$  mutants (no overall effect of dose; overall effect of genotype,  $F(1,34) = 6.69$ ,  $P < 0.05$ ; dose  $\times$  genotype interaction,  $F(2,34) = 5.48$ ,  $P < 0.01$ ). There were no effects on movements of the vibrissae.

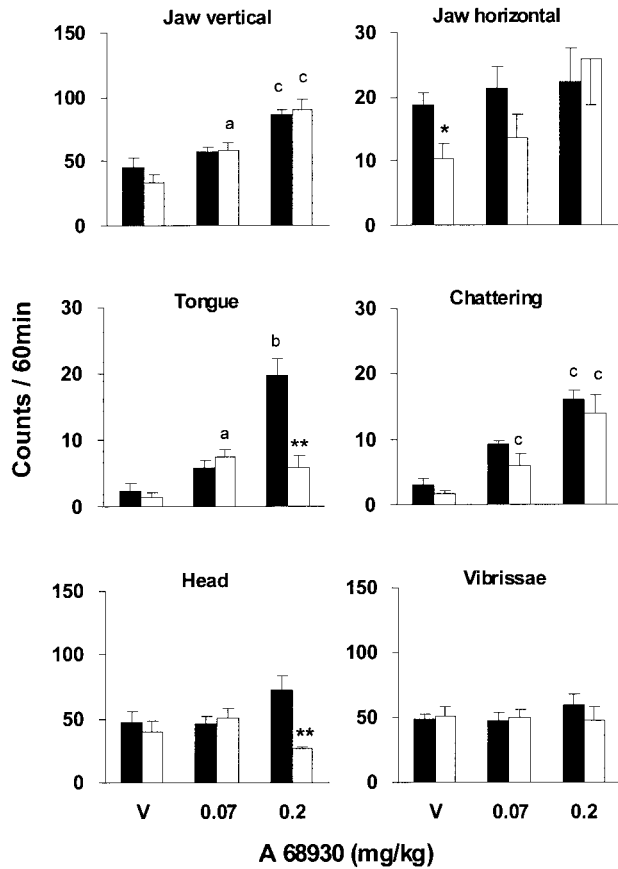


Fig. 5. Phenotype of orofacial movement topographies in wildtypes (n = 6–7, filled columns) and incipient congenic D<sub>2</sub> mutants (n = 6–7, open columns) following challenge with 0.07–0.2 mg/kg A 68930 or vehicle (V). Data are mean counts ± SEM for vertical and horizontal jaw movements, tongue protrusions, incisor chattering, and general head and vibrissae movements over a 60-min period. <sup>a</sup>P < 0.05, <sup>b</sup>P < 0.01, <sup>c</sup>P < 0.001 vs. vehicle; \*P < 0.05, \*\*P < 0.01 vs. wildtype receiving the same dose.

**General parameters: SK&F 83959 challenge**

On examining five male incipient congenic D<sub>2</sub> mutants for orofacial topography in response to SK&F 83959, mean body weight (18 ± 3 g; mean age 221 ± 48 days) was lower (–18%) relative to five male wildtype controls (22 ± 1 g; mean age 340 ± 24 days).

On examining 10 male congenic D<sub>3</sub> mutants for orofacial topography in response to SK&F 83959, mean body weight (27 ± 1 g; mean age 146 ± 9 days) did not differ from 10 male wildtype controls (26 ± 1 g; mean age 144 ± 8 days).

**SK&F 83959-induced orofacial topography D<sub>2</sub> mutants**

In wildtypes, SK&F 83959 induced vertical jaw movements, tongue protrusions, and incisor chattering and reduced horizontal jaw movements (Fig. 6). These responses were unaltered in congenic D<sub>2</sub> mutants (overall effects of dose: vertical jaw movements, F(1,16) = 27.94, P < 0.001; tongue protrusions,

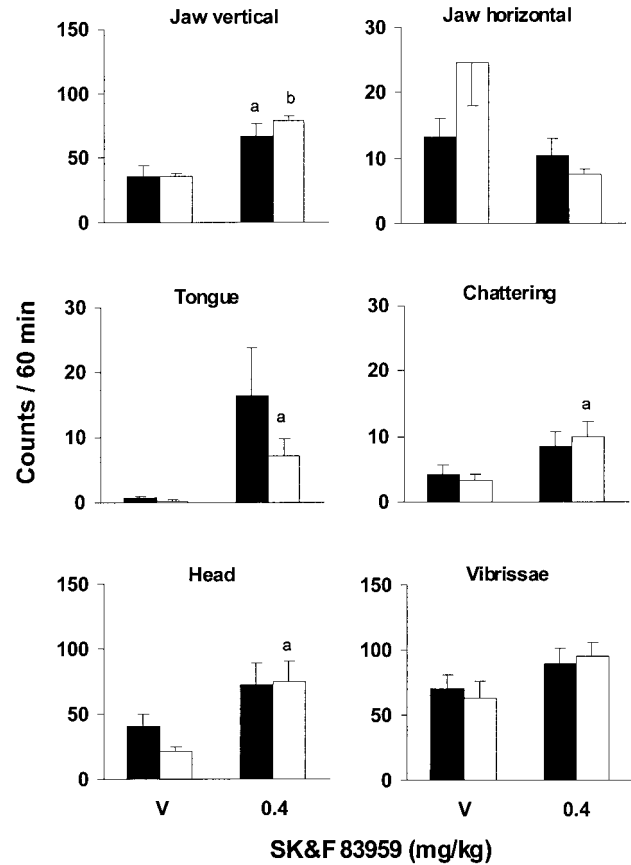


Fig. 6. Phenotype of orofacial movement topographies in wildtypes (n = 5, filled columns) and incipient congenic D<sub>2</sub> mutants (n = 5, open columns) following challenge with 0.4 mg/kg SK&F 83959 or vehicle (V). Data are mean counts ± SEM for vertical and horizontal jaw movements, tongue protrusions, incisor chattering, and general head and vibrissae movements over a 60-min period. <sup>a</sup>P < 0.05, <sup>b</sup>P < 0.01 vs. vehicle.

F(1,16) = 16.27, P < 0.001; incisor chattering, F(1,16) = 9.86, P < 0.01; horizontal jaw movements, F(1,16) = 5.61, P < 0.05; no overall effects of genotype or dose × genotype interactions). Similarly, SK&F 83959 induced general head and vibrissae movements in a manner that did not differ between the genotypes (overall effects of dose: head movements, F(1,16) = 15.38, P < 0.01; vibrissae movements, F(1,16) = 5.15, P < 0.05; no overall effects of genotype or dose × genotype interactions).

**D<sub>3</sub> mutants**

In wildtypes, SK&F 83959 induced vertical jaw movements, tongue protrusions, and incisor chattering and reduced horizontal jaw movements (Fig. 7). These responses were unaltered in congenic D<sub>3</sub> mutants (overall effects of dose: vertical jaw movements, F(2,34) = 70.68, P < 0.001; tongue protrusions, F(2,34) = 5.43, P < 0.01; incisor chattering, F(2,34) = 7.06, P < 0.01; horizontal jaw movements, F(2,34) = 8.47, P < 0.001; no overall effects of genotype or dose ×

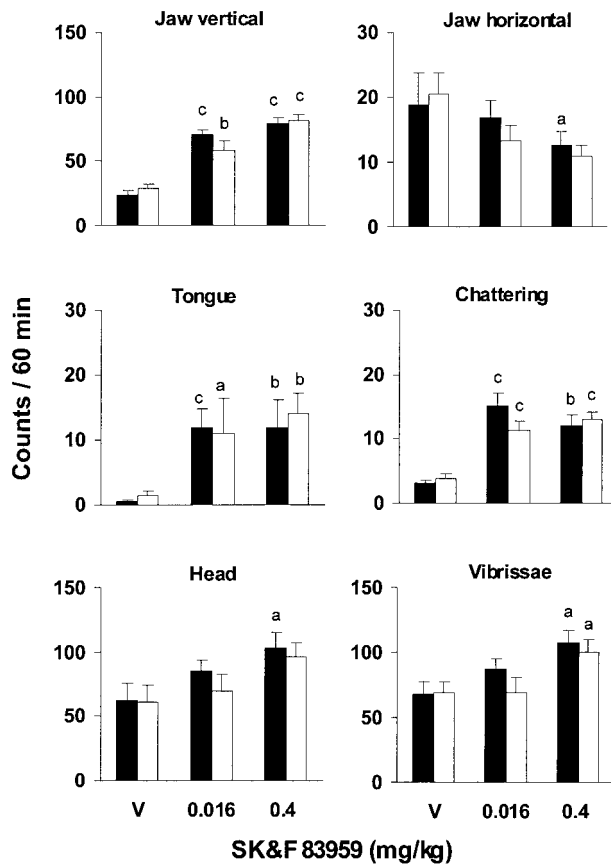


Fig. 7. Phenotype of orofacial movement topographies in wildtypes ( $n = 6-7$ , filled columns) and congenic  $D_3$  mutants ( $n = 6-7$ , open columns) following challenge with 0.016–0.4 mg/kg SK&F 83959 or vehicle (V). Data are mean counts  $\pm$  SEM for vertical and horizontal jaw movements, tongue protrusions, incisor chattering, and general head and vibrissae movements over a 60-min period. <sup>a</sup> $P < 0.05$ , <sup>b</sup> $P < 0.01$ , <sup>c</sup> $P < 0.001$  vs. vehicle.

genotype interactions). Similarly, SK&F 83959 induced general head and vibrissae movements in a manner that did not differ between the genotypes (overall effects of dose: head movements,  $F(2,34) = 21.65$ ,  $P < 0.001$ ; vibrissae movements,  $F(2,34) = 42.43$ ,  $P < 0.001$ ; no overall effects of genotype or dose  $\times$  genotype interactions).

## DISCUSSION

These studies seek to clarify the functional roles of individual DA receptor subtypes, specifically here the involvement of members of the  $D_2$ -like family in regulating orofacial movements (Rosengarten et al., 1983, 1986; Murray and Waddington, 1989; Collins et al., 1991; Waddington et al., 1998; Niznik et al., 2002). Described here are such phenotypic aspects of mice with incipient congenic  $D_2$  and congenic  $D_3$  receptor knockout.

### Methodological issues

A number of methodological refinements have been incorporated into these studies: 1) Because of general

concerns over potential effects of knockout on a mixed (hybrid) genetic background, which might influence apparent phenotype independent of the entity deleted (Gerlai, 1996; Crawley et al., 1997; Kelly et al., 1998; Phillips et al., 1999; Waddington et al., 2001; McNamara et al., 2003), incipient congenic  $D_2$  and congenic  $D_3$  mutant lines were established by repeated backcrossing into a single strain. This work, with counterpart studies of orofacial movement topography in a congenic  $D_{1A}$  mutant line (Tomiya et al., 2002), constitutes a first systematic examination of such phenotype in a congenic  $D_2$  or  $D_3$  mutant line. 2) To allow the detailed study of orofacial movements in the mouse, a novel system was utilised (Tomiya et al., 2001). 3) A physiologically based approach to the resolution and quantification of orofacial movements was adopted, with specification of individual topographies rather than recourse to widely adopted, generic terms such as “vacuous chewing” which composite multiple components that may be regulated differentially (Waddington, 1990; Waddington et al., 1998). This approach allowed resolution in wildtypes of four topographies: vertical jaw movements, horizontal jaw movements, tongue protrusions, and incisor chattering, as well as general movements of the head and vibrissae. Comparisons are made with our recent report on these same parameters, assessed using identical methods by the same investigators, in congenic  $D_{1A}$  mutants (Tomiya et al., 2002).

### Spontaneous orofacial topography

Under spontaneous conditions, vertical jaw movements were increased in incipient congenic  $D_2$  mutants in a manner that did not change materially with time, but were unaltered in congenic  $D_{1A}$  mutants. Conversely, horizontal jaw movements were unaltered in  $D_2$  mutants but essentially abolished in  $D_{1A}$  mutants. Thus, the present approach indicates that apparent oppositional  $D_1$ -like: $D_2$ -like interactions in the regulation of composited jaw movements (Waddington et al., 1994) may in fact reflect the independent actions of  $D_2$  receptors to inhibit vertical jaw movements and of  $D_{1A}$  receptors to facilitate horizontal jaw movements. These differential regulatory effects may be masked by a composite index of jaw movements and are revealed only on resolving individual topographies of behaviour. There appears to be little material involvement of  $D_3$  receptors in these processes.

Low baseline levels of tongue protrusions and incisor chattering were reduced in  $D_2$  mutants and in  $D_{1A}$  mutants in a time-dependent fashion, but were less altered in  $D_3$  mutants. These findings indicate distinct, facilitatory rather than inhibitory roles in their regulation. Among general head and vibrissae movements, only the former were reduced and only in  $D_2$  mutants. Conversely,  $D_{1A}$  mutants evidenced an altered time-course of habituation for these movements, which



would have gone unrecognised using composite indices over shorter periods of assessment.

Thus, the present findings extend to the oral domain our recent findings that D<sub>2</sub>, and less so D<sub>3</sub>, receptors exert essentially motoric effects on exploratory behaviour. While initial studies in D<sub>2</sub> mutants on a mixed genetic background indicated impaired motor function and some Parkinsonian features (Baik et al., 1995; Jung et al., 1999), motor deficits in D<sub>2</sub> mutants were subsequently reported to be much less prominent both on a mixed (Clifford et al., 2000) and on an incipient congenic (Kelly et al., 1998; Clifford et al., 2001) background. Although the present abnormalities in orofacial movement were found in the same incipient congenic D<sub>2</sub> mutants which showed less prominent deficits in motor function (Kelly et al., 1998; Clifford et al., 2001), the relationship of deficits in motor function to abnormalities in orofacial movements is not clear; while striatal D<sub>2</sub> receptors are likely to be involved, the contribution of extrastriatal limbic and possibly cortical mechanisms cannot be discounted. Also, we cannot exclude the possibility of different effects on orofacial movements in D<sub>2</sub> mutants on a mixed background.

Conversely, D<sub>1A</sub> receptors also influence psychomotor processes that regulate change in behaviour over time as an animal interacts with and habituates to its environment (Clifford et al., 1998, 2001; McNamara et al., 2002, 2003), as noted here in relation to orofacial movement. As for D<sub>2</sub> receptors, while striatal D<sub>1A</sub> receptors are likely to be involved, extrastriatal limbic and possibly cortical mechanisms may also contribute to these processes.

As with all "knockouts," it cannot be excluded that aspects of phenotype are influenced also by compensatory mechanisms consequent to the developmental absence of the entity deleted (Clifford et al., 1998, 2000, 2001; Kelly et al., 1998; Sibley, 1999; Waddington et al., 2001; Tomiyama et al., 2002; McNamara et al., 2003).

### **D<sub>2</sub>-like agonist-induced orofacial topography**

The D<sub>2</sub>-like agonist RU 24213 (Euvrard et al., 1980; Clifford et al., 2000; McNamara et al., 2002) reduced vertical and particularly horizontal jaw movements; this effect on horizontal jaw movements was partially reduced in incipient congenic D<sub>2</sub> but not in congenic D<sub>3</sub> mutants. This could suggest, in addition to D<sub>2</sub> mechanisms, some additional involvement of D<sub>4</sub>-mediated or non-DAergic effects. These findings are complementary to the D<sub>1A</sub> receptor exerting a primary, facilitative role in the regulation of horizontal jaw movements under spontaneous conditions and only a minor role under RU 24213 challenge. Low baseline levels of tongue protrusions and incisor chattering, with head and vibrissae movements, were not influenced in any substantive manner by RU 24213, without prominent differences between the genotypes. This indicates some

specificity of RU 24213 to influence jaw movements in the horizontal plane. Only in relation to head movements did a low but not a high dose of RU 24213 exert an inhibitory effect, and this effect was attenuated in congenic D<sub>3</sub> mutants. This complements attenuation in congenic D<sub>3</sub> mutants of the effect of a low dose of RU 24213 to reduce only certain topographies of exploratory behaviour and only following a similar period of habituation (McNamara et al., 2002).

### **D<sub>1</sub>-like agonist-induced orofacial topography**

A 68930 is a D<sub>1</sub>-like agonist of high selectivity and full efficacy to stimulate adenylyl cyclase (DeNinno et al., 1991; Daly and Waddington, 1993; Clifford et al., 2001). That it induced vertical, but not horizontal, jaw movements which were unaltered in incipient congenic D<sub>2</sub> mutants would suggest mediation via D<sub>1</sub>-like receptors independent of D<sub>2</sub> receptor modulation. Conversely, A 68930-induced tongue protrusions were reduced in D<sub>2</sub> mutants, suggesting a facilitatory role for D<sub>2</sub> receptors in this D<sub>1</sub>-like agonist-induced response. Furthermore, both spontaneous and A 68930-induced incisor chattering were reduced in D<sub>2</sub> mutants, suggesting a facilitatory role for D<sub>2</sub> receptors in this orofacial topography.

SK&F 83959 shows high selectivity for D<sub>1</sub>-like over D<sub>2</sub>-like receptors at which it acts to inhibit rather than stimulate DA-sensitive adenylyl cyclase (Arnt et al., 1992; Andringa et al., 1999; Gnanalingham et al., 1995) yet it shows all the behavioural characteristics of cyclase-stimulating D<sub>1</sub>-like agonists, including induction in rats of "vacuous chewing"/perioral movements (Arnt et al., 1992; Deveney and Waddington, 1995; Adachi et al., 1999; Hasegawa et al., 2001) and in mice of vertical but not horizontal jaw movements together with tongue protrusions and incisor chattering (Tomiyama et al., 2001), in a manner similar to A 68930. Thus, it may act at a putative D<sub>1</sub>-like site linked to a transduction system other than / additional to adenylyl cyclase, with phosphoinositide turnover being the most widely entertained candidate (Mahan et al., 1990; Undie and Friedman, 1990; Undie et al., 1994; Waddington et al., 1995, 1998; Panchalingham and Undie, 2001; Niznik et al., 2002). The effects of D<sub>2</sub> receptor ablation on topographical responsivity to SK&F 83959, confined to some attenuation of tongue protrusions, were similar to effects on responsivity to A 68930; conversely, such responsivity was essentially abolished in D<sub>1A</sub> mutants (Tomiyama et al., 2002).

On the basis of evidence in D<sub>3</sub>-null mice on a mixed genetic background (Xu et al., 1997; Jung and Schmauss, 1999; Karasinska et al., 2000), and in other paradigms (Levavi-Sivan et al., 1998; Ridray et al., 1998; Schwartz et al., 1998) it has been suggested that D<sub>3</sub> receptors may participate in and modulate aspects of well-described D<sub>1</sub>-like:D<sub>2</sub>-like interactions that are fundamental regulators of DAergic function (Wadding-

ton et al., 1994, 2001). Congenic  $D_3$  mutants evidenced no material alteration in the topography of orofacial movements in response to these  $D_1$ -like agonists, indicating no substantive role for the  $D_3$  receptor either in their genesis or in their modulation.

### Regulation of orofacial movement topography

These findings help to resolve the individual roles of  $D_2$  and  $D_3$  receptors in orofacial topographies mediated through  $D_2$ -like receptors, yet some anomalies are apparent. For example, A 68930-induced orofacial topographies were either uninfluenced or diminished in  $D_2$  mutants; however, we have recently reported A 68930-induced "vacuous chewing" to be enhanced in these same incipient congenic  $D_2$  mutants under more naturalistic conditions, in accordance with their regulation by oppositional  $D_1$ -like: $D_2$ -like interactions (Waddington et al., 2001; Clifford et al., 2001). It should be emphasised that "vacuous chewing" constitutes a generic term which composites into a single measure a diversity of orofacial movements as assessed under naturalistic circumstances, while the present study involves resolution of specific topographies of orofacial movement assessed under restraint. In particular, it remains difficult to resolve the paradox that naturalistic assessment, while constituting a more physiologically relevant situation, precludes detailed topographical resolution, while restraint allows such topographical resolution but under less physiological circumstances. Such factors may contribute to these differing profiles of effect of  $D_2$  receptor ablation between the two conditions. Thus, an important methodological issue is highlighted (Waddington et al., 2001): profiles of phenotypic effect of gene deletion obtained under one set of conditions may not necessarily generalise to other conditions.

Furthermore, some differences in the effect of  $D_2$  receptor ablation were evident according to whether topographies of orofacial movement were occurring spontaneously or induced by  $D_2$ -like vs.  $D_1$ -like selective agonists. This would appear to attest the following realities: under spontaneous conditions, all DA receptor subtypes are subjected to endogenous, tonic stimulation by DA; conversely, under challenge with agonists, specific families of DA receptor subtypes are subjected to exogenous, phasic stimulation, on a background of possibly modified endogenous, tonic stimulation by DA. Such physiological differences may contribute to differing profiles of effect of  $D_2$  receptor ablation between the two conditions.

In summary,  $D_2$  receptors exert differential regulation of individual orofacial movement topographies in a manner that is distinct from, and sometimes opposite to, that of their  $D_{1A}$  counterparts. Conversely,  $D_3$  receptors exert only minor regulation of such movements.

### ACKNOWLEDGMENTS

We thank Abbott for A 68930 and Hoechst-Marion-Roussel for RU 24213. SK&F 83959 was provided by Research Biochemicals International as part of the Chemical Synthesis Program (NIMH).

### REFERENCES

- Acili D, Fishbourne CS, Drago J, Steiner H, Lachowicz JE, Park BH, Gauda EB, Lee EJ, Cool MH, Sibley DR, Gerfen CR, Westphal H, Fuchs S. 1996. A targeted mutation of the  $D_3$  dopamine receptor gene is associated with hyperactivity in mice. *Proc Natl Acad Sci USA* 93:1945–1949.
- Adachi K, Ikeda H, Hasegawa M, Nakamura S, Waddington JL, Koshikawa N. 1999. SK&F 83959 and non-cyclase-coupled dopamine  $D_1$ -like receptors in jaw movement via dopamine  $D_1$ -like/ $D_2$ -like receptor synergism. *Eur J Pharmacol* 367:143–149.
- Andringa G, Drukarch B, Leysen JE, Cools AR, Stoof JC. 1999. The alleged dopamine  $D_1$  receptor agonist SKF 83959 is a dopamine  $D_1$  receptor antagonist in primate cells and interacts with other receptors. *Eur J Pharmacol* 364:33–41.
- Arnt J, Hyttel J, Sanchez C. 1992. Partial and full dopamine  $D_1$  receptor agonists in mice and rats: relation between behavioural effects and stimulation of adenylate cyclase activity in vitro. *Eur J Pharmacol* 213:259–267.
- Baik JH, Picetti R, Saiardi A, Thiriet G, Dierich A, Depaulis A, Le Meur M, Borrelli E. 1995. Parkinsonian-like locomotor impairment in mice lacking dopamine  $D_2$  receptors. *Nature* 377:424–428.
- Clifford JJ, Tighe O, Croke DT, Sibley DR, Drago J, Waddington JL. 1998. Topographical evaluation of the phenotype of spontaneous behaviour in mice with targeted gene deletion of the  $D_{1A}$  dopamine receptor: paradoxical elevation of grooming syntax. *Neuropharmacology* 37:1595–1602.
- Clifford JJ, Tighe O, Croke DT, Kinsella A, Sibley DR, Drago J, Waddington JL. 1999. Conservation of behavioural topography to dopamine  $D_1$ -like receptor agonists in mutant mice lacking the  $D_{1A}$  receptor implicates a  $D_1$ -like receptor not coupled to adenylyl cyclase. *Neuroscience* 93:1483–1489.
- Clifford JJ, Usiello A, Vallone D, Kinsella A, Borrelli E, Waddington JL. 2000. Topographical evaluation of behavioural phenotype in a line of mice with targeted gene deletion of the  $D_2$  dopamine receptor. *Neuropharmacology* 39:382–390.
- Clifford JJ, Kinsella A, Tighe O, Rubinstein M, Grandy DK, Low M, Croke DT, Waddington JL. 2001. Comparative, topographically-based evaluation of behavioural phenotype and specification of  $D_1$ -like: $D_2$ -like interactions in a line of incipient congenic mice with  $D_2$  dopamine receptor "knockout." *Neuropsychopharmacology* 25:527–536.
- Collins P, Broekkamp CLE, Jenner P, Marsden CD. 1991. Drugs acting at D-1 and D-2 receptors induce identical purposeless chewing in rats which can be differentiated by cholinergic manipulation. *Psychopharmacology* 103:503–512.
- Crawley JN, Belknap JK, Collins A, Crabbe JC, Frankel W, Henderson N, Hitzemann RJ, Maxson SC, Miner LL, Silva AJ, Wehner JM, Wynshaw-Boris A, Paylor R. 1997. Behavioural phenotypes of inbred mouse strains: implications and recommendations for molecular studies. *Psychopharmacology* 132:107–124.
- Daly SA, Waddington JL. 1993. Behavioural evidence for "D-1-like" dopamine receptor subtypes in rat brain using the new isochroman agonist A 68930 and isoquinoline antagonist BW 737C. *Psychopharmacology* 113:45–50.
- DeNinno MP, Schoenleber R, MacKenzie R, Britton DR, Asin KE, Briggs C, Trugman JM, Ackerman M, Artman L, Bednarz L, Bhatt R, Curzon P, Gomez E, Kang CH, Stittsworth J, Keababian JW. 1991. A68930: a potent agonist selective for the dopamine  $D_1$  receptor. *Eur J Pharmacol* 199:209–219.
- Euvrard C, Ferland L, Di Paolo T, Beaulieu M, Labrie F, Oberlander C, Raynaud JP, Boissier JR. 1980. Activity of two new potent dopaminergic agonists at the striatal and anterior pituitary levels. *Neuropharmacology* 19:379–386.
- Gerlai R. 1996. Gene-targeting studies of mammalian behavior: is it the mutation or the background genotype? *Trends Neurosci* 19:177–181.
- Gnanalingham KK, Hunter AJ, Jenner P, Marsden CD. 1995. Stimulation of adenylate cyclase activity by benzazepine D-1 dopamine agonists with varying efficacies in 6-hydroxydopamine lesioned rat — relationship to circling behaviour. *Biochem Pharmacol* 49:1185–1193.

- Hasegawa M, Adachi K, Nakamura S, Sato S, Waddington JL, Koshikawa N. 2001. Ventral striatal vs. accumbal (shell) mechanisms and non-cyclase-coupled dopamine D<sub>1</sub>-like receptors in jaw movements. *Eur J Pharmacol* 423:171–178.
- Jung MY, Schmauss C. 1999. Decreased c-fos responses to dopamine D(1) receptor agonist stimulation in mice deficient for D(3) receptors. *J Biol Chem* 274:29406–29412.
- Jung MY, Skryabin BV, Aria M, Abbondanzo S, Fu D, Brosius J, Robakis NK, Polites HG, Pintar JE, Schmauss C. 1999. Potentiation of the D<sub>2</sub> mutant motor phenotype in mice lacking dopamine D<sub>2</sub> and D<sub>3</sub> receptors. *Neuroscience* 91:911–924.
- Karasinska JM, George SR, El-Ghundi M, Fletcher PJ, O'Dowd BF. 2000. Modification of dopamine D(1) receptor knockout phenotype in mice lacking both dopamine D(1) and D(3) receptors. *Eur J Pharmacol* 399:171–181.
- Kelly MA, Rubinstein M, Asa SL, Zhang G, Saez C, Bunzow JR, Allen RG, Hnasko R, Ben-Jonathan N, Grandy DK, Low MJ. 1997. Pituitary lactotroph hyperplasia and chronic hyperprolactinemia in dopamine D<sub>2</sub> receptor-deficient mice. *Neuron* 19:103–113.
- Kelly MA, Rubinstein M, Phillips TJ, Lessov CN, Burkhardt-Kasch S, Zhang G, Bunzow JR, Fang Y, Gerhardt GA, Grandy DK, Low MJ. 1998. Locomotor activity in D2 dopamine receptor-deficient mice is determined by gene dosage, genetic background, and developmental adaptations. *J Neurosci* 18:3470–3479.
- Levavi-Sivan B, Park BH, Fuchs S, Fishburn CS. 1998. Human D3 dopamine receptor in the medulloblastoma TE671 cell line: cross-talk between D1 and D3 receptors. *FEBS Lett* 439:138–142.
- Mahan LC, Bush RM, Monsma Jr FJ, Sibley DR. 1990. Expression of striatal D<sub>1</sub> dopamine receptors coupled to inositol phosphate production and Ca<sup>2+</sup> mobilization in *Xenopus oocytes*. *Proc Natl Acad Sci USA* 87:2196–2200.
- McNamara FN, Clifford JJ, Tighe O, Kinsella A, Drago J, Fuchs S, Croke DT, Waddington JL. 2002. Phenotypic, ethologically based resolution of spontaneous and D<sub>2</sub>-like vs. D<sub>1</sub>-like agonist-induced behavioural topography in mice with congenic D<sub>3</sub> dopamine receptor “knockout.” *Synapse* 46:19–31.
- McNamara FN, Clifford JJ, Tighe O, Kinsella A, Drago J, Croke DT, Waddington JL. 2003. Congenic D<sub>1A</sub> dopamine receptor mutants: ethologically-based resolution of behavioural topography indicates genetic background as a determinant of knockout phenotype. *Neuropsychopharmacology* 28:86–99.
- Missale C, Nash SR, Robinson SW, Jaber M, Caron MG. 1998. Dopamine receptors: from structure to function. *Physiol Rev* 78:189–225.
- Murray AM, Waddington JL. 1989. The induction of grooming and vacuous chewing by a series of selective D-1 dopamine receptor agonists: two directions of D-1:D-2 interaction. *Eur J Pharmacol* 160:377–384.
- Niznik HB, Sugamori KS, Clifford JJ, Waddington JL. 2002. D<sub>1</sub>-like dopamine receptors: molecular biology and pharmacology. In: Di Chiara G, editor. *Handbook of experimental pharmacology: dopamine in the CNS*. Heidelberg: Springer. p 122–158.
- Panchalingham S, Undie AS. 2001. SKF 83959 exhibits biochemical agonism by stimulating [<sup>35</sup>S]GTPγS binding and phosphoinositide hydrolysis in rat and monkey brain. *Neuropharmacology* 40:826–837.
- Phillips TJ, Hen R, Crabbe JC. 1999. Complications associated with genetic background effects in research using knockout mice. *Psychopharmacology* 147:5–7.
- Ridray S, Griffon N, Mignon V, Souil E, Carboni S, Diaz J, Schwartz JC, Sokoloff P. 1998. Coexpression of dopamine D1 and D3 receptors in islands of Calleja and shell of nucleus accumbens of the rat: opposite and synergistic functional interactions. *Eur J Neurosci* 10:1676–1686.
- Rosengarten H, Schweitzer JW, Friedhoff AJ. 1983. Induction of oral dyskinesia in naive rats by D-1 stimulation. *Life Sci* 33:2471–2482.
- Rosengarten H, Schweitzer JW, Friedhoff AJ. 1986. Selective dopamine D-2 receptor reduction enhances a D-1 mediated oral dyskinesia in rats. *Life Sci* 39:29–35.
- Ross SA, Wong JY, Clifford JJ, Kinsella A, Massalas JS, Horne MK, Scheffer IE, Kola I, Waddington JL, Berkovic SF, Drago J. 2000. Phenotypic characterization of an alpha 4 neuronal nicotinic acetylcholine receptor subunit knock-out mouse. *J Neurosci* 20:6431–6441.
- Schwartz JC, Diaz J, Bordet R, Griffon N, Perachon S, Pilon C, Ridray S, Sokoloff P. 1998. Functional implications of multiple dopamine receptor subtypes: the D1/D3 receptor coexistence. *Brain Res Rev* 26:236–242.
- Sibley DR. 1999. New insight into dopaminergic receptor function using antisense and genetically altered animals. *Annu Rev Pharmacol Toxicol* 39:313–341.
- Tomiyama K, McNamara FN, Clifford JJ, Kinsella A, Koshikawa N, Waddington JL. 2001. Topographical assessment and pharmacological characterization of orofacial movements in mice: dopamine D<sub>1</sub>-like vs. D<sub>2</sub>-like receptor regulation. *Eur J Pharmacol* 418:47–54.
- Tomiyama K, McNamara FN, Clifford JJ, Kinsella A, Drago J, Tighe O, Croke DT, Koshikawa N, Waddington JL. 2002. Phenotypic resolution of spontaneous and D<sub>1</sub>-like agonist-induced orofacial movement topographies in congenic dopamine D<sub>1A</sub> receptor “knock-out” mice. *Neuropharmacology* 42:644–652.
- Undie AS, Friedman E. 1990. Stimulation of a dopamine D<sub>1</sub> receptor enhances inositol phosphate formation in rat brain. *J Pharmacol Exp Ther* 253:987–992.
- Undie AS, Weinstock J, Sarau HM, Friedman E. 1994. Evidence for a distinct D<sub>1</sub>-like dopamine receptor that couples to activation of phosphoinositide metabolism in brain. *J Neurochem* 62:2045–2048.
- Waddington JL. 1990. Spontaneous orofacial movements induced in rodents by very long-term neuroleptic drug administration: phenomenology and putative relationship to tardive dyskinesia. *Psychopharmacology* 101:431–447.
- Waddington JL, Daly SA, McCauley PG, O'Boyle KM. 1994. Levels of functional interaction between D<sub>1</sub>-like and D<sub>2</sub>-like dopamine receptor systems. In: Niznik HB, editor. *Dopamine receptors and transporters: pharmacology, structure and function*. New York: Marcel-Dekker. p 511–537.
- Waddington JL, Daly SA, Downes RP, Deveney AM, McCauley PG, O'Boyle KM. 1995. Behavioural pharmacology of “D<sub>1</sub>-like” dopamine receptors: further subtyping, new pharmacological probes and interactions with ‘D<sub>2</sub>-like’ receptors. *Prog Neuro Psychopharmacol Biol Psychiatry* 19:811–831.
- Waddington JL, Deveney AM, Clifford JJ, Tighe O, Croke DT, Sibley DR, Drago J. 1998. D<sub>1</sub>-like dopamine receptors: regulation of psychomotor behaviour, D<sub>1</sub>-like: D<sub>2</sub>-like interactions and effects of D<sub>1A</sub> targeted gene deletion. In: Jenner P, Demirdama R, editors. *Dopamine receptor subtypes: from basic science to clinic*. Amsterdam: IOS Press. p 45–63.
- Waddington JL, Clifford JJ, McNamara FN, Tomiyama K, Koshikawa N, Croke DT. 2001. The psychopharmacology-molecular biology interface: exploring the behavioural roles of dopamine receptor subtypes using targeted gene deletion (“knockout”). *Prog Neuro Psychopharmacol Biol Psychiatry* 25:925–964.
- Xu M, Koeltzow TE, Santiago GT, Moratalla R, Cooper DC, Hu XT, White NM, Graybiel AM, White FJ, Tonegawa S. 1997. Dopamine D<sub>3</sub> receptor mutant mice exhibit increased behavioral sensitivity to concurrent stimulation of D<sub>1</sub> and D<sub>2</sub> receptors. *Neuron* 19:837–848.