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ORIGINAL ARTICLE

Big fish (and a smallish skate) eat small fish: diet variation and trophic level of *Sympterygia acuta*, a medium-sized skate high in the food web

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Keywords

Feeding activity; ontogenetic shifts; Rajidae; Southwest Atlantic.

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Abstract

The bignose fanskate, Sympterygia acuta, is a small-to-medium-sized species endemic to shallow coastal waters of the Southwest Atlantic. Sympterygia acuta displays a clear seasonal reproductive cycle, characterized by maximum egg-laying activity in spring and hatching in summer. We hypothesized that diet and feeding activity change with maturity stage and season and that, given its smallish size, the trophic level is low. Using a multiple-hypothesis modeling approach, the diet of S. acuta in relation to sex, body size, maturity stage, region (i.e. north and south) and season was analysed; and a potential relationship between feeding activity and the seasonal reproductive cycle was assessed. Sympterygia acuta fed on a broad spectrum of prey, but teleosts were more important (47.97% index of relative importance, %IRI), followed by decapods (39.84%IRI), cumaceans (8.31%IRI) and isopods (1.89%IRI). Maturity stage was a strong determinant of the ontogenetic diet shift of S. acuta, and relationships between number of prey consumed with season and region were found... Feeding activity was higher in the cold season than in the warm season, and was less important in the south region than in the north region. Unexpectedly, the specific trophic level was high (3.87). Sympterygia acuta shifts its diet with maturity stage, possibly by a combination of an improved ability to capture prey and a change in energy demand of mature individuals. Despite being a small-to-medium-sized skate, S. acuta showed a trophic level similar to that of large-bodied marine predators. It reduces its feeding activity seasonally because in the warm season this species may experience an increased predation risk from large sharks.

Introduction

Body size imposes important constraints on the manner in which predators interact with their prey (Werner & Gilliam 1984), and is one of the most important traits that shape the structure and dynamics of biological communities (Hairston & Hairston 1993; Arim *et al.* 2007). In aquatic communities, energy transfer between trophic levels is predicted to be strongly constrained by body size,

given that aquatic predators are generally gape limited (Hairston & Hairston 1993; Jennings *et al.* 2001; Arim *et al.* 2007). This morphological limitation results in a positive correlation between trophic level and body size in many fish species (Romanuk *et al.* 2011).

The feeding behavior of a predator can be affected by many factors, such as prey abundance, sex, body size and reproductive condition (Wetherbee & Cortés 2004). As a result, ontogenetic, regional and/or seasonal changes in

the dietary of a predator can occur. However, feeding behavior can also be influenced by a trade-off between foraging and predation risk and social behavior (Sih et al. 2000). In general, elasmobranch species (i.e. sharks and skates) that inhabit deep-water environments show yearround reproductive activity, whereas skates that inhabit shallow waters with high variability have seasonal reproductive cycles (Lucifora & García 2004). In the same way, species composition of benthic and demersal coastal communities is variable, as environmental conditions change seasonally (Jaureguizar et al. 2006). These changes in prev abundance and distribution affect the foraging ecology of their predators (Muto et al. 2001). Therefore, foraging behavior and reproductive strategy should be adapted to seasonal environmental fluctuations in species that dwell in shallow coastal waters.

The bignose fanskate, Sympterygia acuta Garman 1877 (Chondrichthyes, Rajidae), is endemic to the Southwest Atlantic and is distributed from Rio de Janeiro, Brazil (22°S) to the San Jorge Gulf, Argentina (45°43' S) (Cousseau et al. 2007; Bovcon et al. 2011). This species occurs in shallow coastal waters (up to 50 m depth) and has a small-to-medium body size (Cousseau et al. 2007). Males mature at 475 mm total length (TL), whereas females mature at 478 mm TL (Mabragaña et al. 2014). Off Uruguay and North Argentina, S. acuta displays a clear seasonal reproductive cycle, characterized by maximum egglaying activity in spring and hatching in summer (Mabragaña et al. 2014). Although S. acuta is common in this region, there has been no detailed study of the feeding ecology of this species. As S. acuta lives in shallow coastal waters with great environmental variability and shows a seasonal reproductive cycle, our hypothesis was that diet changes with maturity stage and season. Furthermore, given the rather small body size of S. acuta and the positive association between body size and trophic level observed in skates (Ebert & Bizzarro 2007), we also hypothesized that the trophic level of S. acuta is low.

The purposes of this study were to determine the feeding habits and the feeding activity of *S. acuta* off Uruguay and North Argentina. Our specific aims were to (i) quantify the diet; (ii) assess whether there are sexual, ontogenetic, regional and/or seasonal shifts in the diet composition; (iii) evaluate a potential relationship between feeding activity and the seasonal reproductive cycle; and (iv) estimate the trophic level of *S. acuta*.

Material and Methods

Study area

The study area was located in the coastal region off Uruguay and North Argentina between 34° and 41°S, from 5 to 50 m depth. This region consists of two large ecosystems: a northern (between 34° and 38°S) stratified coastal zone influenced by the very large discharge of continental waters of the Río de la Plata; and a southern (38°–41°S), more homogeneous coastal zone, called El Rincón, influenced by the smaller discharges of the Negro and Colorado rivers and by high-salinity waters of the San Matías Gulf (Guerrero & Piola 1997; Lucas *et al.* 2005).

Sampling

Individuals of *Sympterygia acuta* were collected during November and December 2005, February 2006 and June 2007 from scientific trawl surveys conducted by the Instituto Nacional de Investigación y Desarrollo Pesquero (INIDEP, Argentina; Fig. 1). Additional samples were obtained from commercial landings of the coastal fleet based in Mar del Plata harbor during October and November 2006 and February, June, July, August, October, November and December 2007 (Fig. 1).

All specimens used in this study were obtained already dead and frozen from both the scientific surveys and commercial fisheries. This ensured that our study did not impose any additional mortality upon the studied population

For each captured individual, sex, maturity stage and TL (mm) were recorded. Maturity stage (immature or mature) was determined according to the condition of

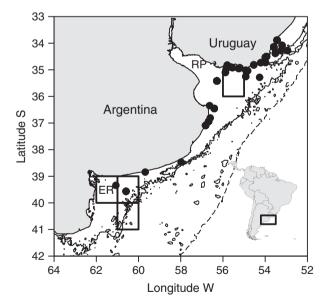


Fig. 1. Map of study area where individuals of *Sympterygia acuta* were collected off Uruguay and North Argentina. Black circles: trawl stations; black rectangles: cells of the fishing grid. The 50-m and 200-m isobaths are shown as solid and dashed lines, respectively. The rectangle in the inset shows the location of the study area in South America. ER = El Rincón; RP = Río de la Plata.

uteri, oviducal glands and ovarian follicles in females, and the development of sperm ducts and testes and the degree of clasper calcification in males (Stehmann 2002; Colonello *et al.* 2011). The stomachs were dissected from each individual and stored at -20 °C. In the laboratory, prey found in the stomachs were sorted, identified to the lowest possible taxonomic level using published catalogues (Boschi *et al.* 1992; Cousseau & Perrotta 2000), counted and weighed (± 0.01 g).

Data analyses

The importance of each prey in the diet was calculated using percentage by number (%N), percentage by mass (%M), percentage frequency of occurrence (%FO) and index of relative importance (IRI) (Pinkas *et al.* 1971). IRI was expressed as percentage (%IRI; Cortés 1997).

Mean cumulative Shannon diversity index was plotted as a function of sample size by sex, maturity stage, season and region, randomized 100 times (Magurran 2004), to assess whether the number of stomachs analysed was sufficient to conduct statistical analyses. Sample size was considered sufficient to describe diet if the cumulative diversity reached an asymptote.

For statistical purposes, the prey were grouped separately in seven zoological categories (teleosts, shrimps, crabs, amphipods, cumaceans, isopods and polychaetes). To identify changes in the diet composition of Sympterygia acuta with sex, ontogeny, region and season, we adopted a multiple-hypothesis modeling approach (Franklin et al. 2001; Johnson & Omland 2004; Symonds & Moussalli 2011). Ontogenetic changes in the diet were assessed for body size (TL) and for maturity stage between immature and mature individuals. Regional dietary changes were assessed between north (between 34 and 38°S) and south regions (between 38 and 41°S). Seasonal differences in the diet were evaluated between warm (October to March) and cold seasons (April to September). For each prey category, we built generalized linear models (GLMs; Venables & Ripley 2002) where the response variable was the number of prey consumed and independent variables were sex, maturity stage, TL, region and season. Further, models with containing up to two independent variables in different cominations were fitted. We also fitted a model without any of the independent variables (i.e. a null model) to test the hypothesis that none of the variables considered had an effect on the consumption of a particular prey. The models with number of prey as response variable had large variances because of the large number of zero-values in the samples; therefore, a negative binomial error distribution and a log link were used (Crawley 2005).

For each model fitted, we calculated the Akaike information criterion (AIC) and Akaike weights (w). AIC measures the amount of information lost when fitting a model; hence, the model with the lowest AIC was selected as the best model (Crawley 2005). To obtain the likelihood of each model fitted given the data, w was calculated (Franklin *et al.* 2001). If the best model was not clearly superior to some of the others in the set, we used model averaging to estimate the parameters of the variables included in the best model (Johnson & Omland 2004; Symonds & Moussalli 2011).

To test the hypothesis that feeding activity changes with maturity stage and season, we built a GLM with diet breadth (i.e. number of prey species consumed, N_{species}), number of prey consumed (Nprey) and proportion of empty stomachs as response variables, and sex, maturity stage, TL, region and season as independent variables. As explained above, models containing multiple independent variables were fitted and the best model was selected by using a multiple-hypothesis modeling approach. To account for the different number of prey in each sample in the models having N_{species} as response variable, the log of the total number of prey consumed (log N_{prey}) by individual was included as an independent variable (Barbini & Lucifora 2012a). Models with N_{species} as the response variable had a Poisson error distribution and a log link because this variable consists of count data, which take only positive or zero values (Crawley 2005). Models with N_{prey} as the response variable had a negative binomial error distribution and a log link because the variance was much greater than the mean (Crawley 2005). Models with proportion of empty stomachs as the response variable had a binomial error distribution and a logit link due to their binary nature (i.e. containing food or empty; Crawley 2005). We interpreted the changes in the type and amount of prey consumed as a reflection of the feeding activity of S. acuta, i.e. a broad diet breadth and a higher number of prey consumed and a proportion of stomachs with food, indicate high feeding activity and vice versa. All statistical analyses were performed using the R statistical software, version 3.1.0. (R Development Core Team, 2014). The trophic level of S. acuta was calculated using the trophic index (TL_k) proposed by Cortés (1999) as:

$$TL_k = 1 + \left(\sum_{j=1}^n P_j \times TL_j\right) \tag{1}$$

where P_j is the proportion of each prey category based on %IRI, TL_j is the trophic level of each prey category j, and n is the total number of prey categories. Trophic level of each prey category was obtained from Ebert & Bizzarro (2007). The trophic dynamics of a species may vary

substantially along its life history (Garrison & Link 2000; Ebert & Bizzarro 2007), then the relationship between maturity stage and TLk was analysed. For that purpose, the observations were randomly sampled 100 times with replacement in order to obtain the frequency distribution of TL_k . A GLM was then used to model TL_k as a function of maturity stage of *S. acuta*. We fitted a null model to judge whether or not maturity stage had an effect on TL_k .

Results

A total of 417 individuals of *Sympterygia acuta* was examined, with 301 (72.18%) containing food in their stomachs. The sample was composed of 85 immature females (253–512 mm TL), 57 mature females (435–620 mm TL), 80 immature males (260–516 mm TL) and 79 mature males (448–570 mm TL). The cumulative diversity curves reached an asymptote in all subgroups, indicating that the sample size was large enough (Fig. 2).

Overall, 2556 prey with a total weight of 1458.3 g were found in the stomachs. In total, 68 different prey taxa were identified within stomachs: 21 decapods, 16 teleosts, 10 isopods, seven polychaetes, three amphipods, cumaceans and mollusks, two chondrichthyans and one mysidacean and lancelet. In terms of %IRI, the dominant prey were teleosts and decapods, followed by cumaceans and isopods. Amphipods, mysidaceans and polychaetes had minor importance (%IRI <1; Table 1). The most important prey by weight were teleosts. Decapods were the most important prey by number and frequency of occurrence. Among the teleosts, Dules auriga was the most important in number, weight and frequency of occurrence, followed by Engraulis anchoita. The most common decapod prey by number, weight and frequency of occurrence was Artemesia longinaris, the second by number and frequency of occurrence was Cyrtograpsus affinis, and Libinia spinosa was the second by weight.

Ontogenetic, regional and seasonal patterns in the diet composition of S. acuta were found (Table 2 and Fig. 3). The consumption of all prey categories was independent of sex. Immature individuals preyed on shrimps, crabs, cumaceans, isopods and polychaetes more heavily than mature individuals. By contrast, teleosts were consumed more by mature than by immature individuals. The consumption of amphipods decreased with increasing TL of S. acuta (Table 3). The consumption of teleosts, shrimps and crabs was affected by the region. Sympterygia acuta preyed more heavily on teleosts in the south region than in the north region, whereas shrimps and crabs were consumed more in the north region than in the south region. Seasonally, S. acuta preyed more heavily on cumaceans, isopods and polychaetes in the cold season than in the warm season.

For shrimp, amphipod and polychaete prey, there were no single models that prevailed over the others, since all models had a low w; therefore, an average of all models was computed (i.e. model averaging). For amphipods, the model averaging slope was -0.045 for TL (w = 0.812). The averaged coefficients for polychaetes were 0.821 for immature individuals (w = 0.515) and 1.307 for the cold season (w = 0.901).

The best models explaining $N_{species}$, N_{prey} and the proportion of empty stomachs included season and region (Table 3). These response variables were not affected by sex, maturity stage or TL. For both $N_{species}$ and N_{prey} , consumption was higher in the cold season than in the warm season, and less important in the south region than in the north region. Seasonally, the proportion of *S. acuta* with empty stomachs was lower in the cold season than in the warm season, and regionally the proportion of empty stomachs was higher in the south region than in the north region. The best model for $N_{species}$ had a low w; therefore, we computed model averaging: coefficients were 0.172 for the cold season (relative to the warm season, w = 0.794) and 0.079 for the south region (relative to the north region, w = 0.344).

The mean TL_k of S. acuta was 3.87 (SD = 0.022). The TL_k was significantly different between maturity stages (TL immature = 3.56; TL mature individuals = 4.15). The parameters of the model were: intercept = 4.15, maturity stage = 3.56 immature (relative to mature); AIC = -1120.1; w = 0.99.

Discussion

The typical pattern found in the diet composition of skate species is as follows: small-bodied skates are benthophagic, consuming worms and small crustaceans; midsized skates are predators of small and large crustaceans and, occasionally, cephalopods and fishes; and large-bodied skates consume mainly fishes (Orlov 1998). Our results indicate that Sympterygia acuta consumed mainly teleosts. As S. acuta is a small-to-medium-sized species, this pattern does not follow the general pattern of diet composition in skates. Food partitioning among sympatric skate species is mainly associated with body size (Ebert et al. 1991; Bizzarro et al. 2007; Treloar et al. 2007). Sympterygia acuta uses a trophic resource (i.e. teleosts) that is not available to other skates of small and medium sizes. This competitive advantage may be associated with morphologic traits (i.e. large snout size). A diet based on teleosts is a phenomenon widely observed in skates that have an elongated snout and occur in deep water, as for example in some species of the genera Dipturus (Kyne et al. 2008), Zearaja (Lucifora et al. 2000; Koen Alonso et al. 2001) and Raja (Robinson et al.

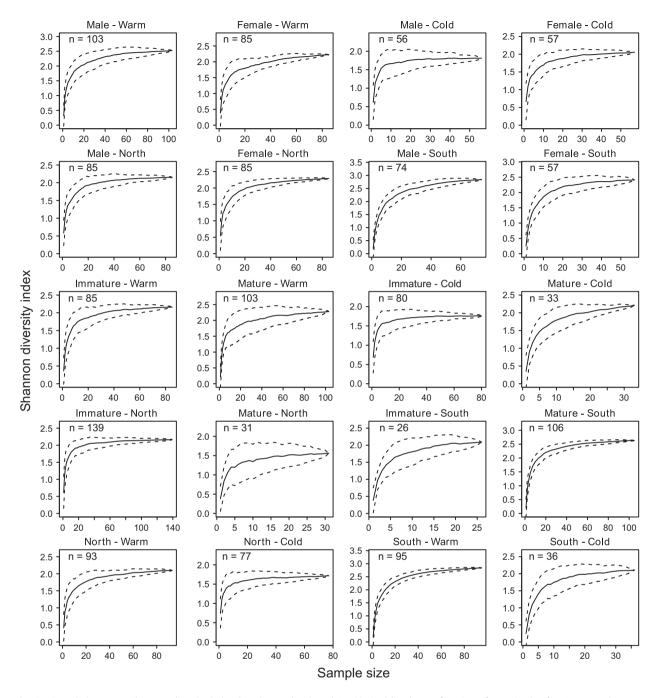


Fig. 2. Cumulative mean Shannon diversity index (continuous lines) and SD (dashed lines) as a function of sample size for *Sympterygia acuta* from off Uruguay and North Argentina.

2007). Among skates, *S. acuta* is the only species with a relatively small body size and a very elongated snout that occurs in shallow waters. The large snout of *S. acuta* may improve its sensory capacity to catch teleosts, in a similar way as the cephalofoil increases the sensory capabilities in prey searching of hammerhead sharks (Kajiura 2001). Further studies are needed to test this hypothesis.

Body size influences trophic hierarchies in aquatic communities (Hairston & Hairston 1993); therefore, small species such as *S. acuta* would be expected to feed at lower trophic levels. This prediction has been confirmed in elasmobranch fishes, such as some shark (Cortés 1999) and skate species (Ebert & Bizzarro 2007), and in many other fish species (Romanuk *et al.* 2011).

Table 1. Diet composition of *Sympterygia acuta* expressed as percentage by number (%N), mass (%M), frequency of occurrence (%FO), index of importance relative (IRI) and percentage of IRI (%IRI).

%N %M %FO IRI %IRI prey Teleostei 6.69 75.45 45.18 3711 47.97 Unidentified teleosts 3.32 17.37 25.25 522.7 Engraulidae Anchoa marinii 0.04 0.07 0.33 0.03 Engraulis anchoita 0.58 13.08 4.65 63.6 Phycidae Urophycis brasiliensis 0.04 0.02 0.33 0.02 Ophidiidae Raneya brasiliensis 0.08 1.28 0.66 0.90 Batrachoididae Porichthys porosissimus 0.31 4.79 2.32 11.87 Triglidae Prionotus nudigula 0.04 0.57 0.33 0.20 Serranidae 20.81 Dules auriga 0.82 5.31 115 Carangidae Trachurus lathami 0.12 0.66 1.84 2.64 Sciaenidae Cynoscion guatucupa 2.53 0.99 0.12 2.64 Micropogonias furnieri 0.04 0.64 0.33 0.22 Paralonchurus 0.12 0.82 0.99 0.93 brasiliensis Achiropsettidae Achiropsetta tricholepis 0.12 0.57 0.66 0.45 Paralichthyidae Etropus longimanus 0.39 3.98 2.66 11.61 Cynoglossidae Unidentified 0.99 0.12 0.61 0.73 Cynoglossidae Symphurus spp. 0.43 5.65 2.32 14.14 Chondrichthyes 0.23 1.10 1.33 1.77 0.02 Rajidae Unidentified skates 0.19 0.68 0.99 0.87 Atlantoraja cyclophora 0.04 0.42 0.33 0.15 Crustacea Decapoda 31.73 18.42 61.46 3082 39.84 Unidentified shrimps 5.05 1.46 12.96 84.27 Penaeidae Artemesia longinaris 14.75 5.67 24.25 495.2 Solenoceridae Pleoticus muelleri 0.51 2.18 2.66 7.16 Sergestidae 0.07 0.33 Sergia mirin 0.12 0.06 Peisos spp. 0.31 0.02 0.66 0.22 Anomura Unidentified anomurans 0.12 0.07 0.66 0.13 Diogenidae 0.31 Loxopagurus 0.36 1.66 1.11 loxochelis Paguridae Pagurus exilis 0.31 0.52 1.99 1.67 Albuneidae Blepharipoda 0.39 1.15 2.66 4.1 doelloi

Table 1. Continued

| Prey No. No. | Table 1. Continued | | | | | |
|--|--------------------------|------|-------|-------|-------|------|
| Unidentified brachyurans | prey | %N | %M | %FO | IRI | %IRI |
| Unidentified brachyurans | Brachyura | | | | | |
| Majidae | - | 1.88 | 1.95 | 11.29 | 43.24 | |
| Leurocyclus tuberculosus 0.04 0.05 0.33 0.03 Libinia spinosa 0.16 1.93 1.33 2.77 Pelia rotunda 0.09 0.04 0.33 0.04 Atelecyclidae Peltarion spinosulum 0.39 0.49 0.66 0.59 Portunidae Ovalipes trimaculatus 0.59 0.57 1.33 1.54 Xanthidae Pillimnus reticulatus 0.04 0.02 0.33 0.02 Belliidae Corystoides abbreviatus 0.62 0.21 1.33 1.12 Grapsidae Cyrtograpsus affinis 4.26 1.09 7.97 42.72 Pinnixa patagoniensis 0.39 0.10 2.66 1.31 1.71 Pinnixa patagoniensis 0.39 0.10 2.66 1.31 1.71 Pinnixa patagoniensis 0.39 0.10 2.66 1.31 1.71 Pinnixa patagoniensis 0.39 0.55 7.97 76.13 0.98 Mpinphipoda 8.99 0 | | | | | | |
| tuberculosus Libinia spinosa 0.16 1.93 1.33 2.77 Pella rotunda 0.09 0.04 0.33 0.04 Atelecyclidae Peltarion spinosulum 0.39 0.49 0.66 0.59 Portunidae 0.04 0.02 0.33 1.54 Xanthidae Pilimus reticulatus 0.04 0.02 0.33 0.02 Belliidae Corystoides abbreviatus 0.62 0.21 1.33 1.12 Grapsidae Cyrtograpsus affinis 4.26 1.09 7.97 42.72 Pinnixa patagoniensis 0.39 0.10 2.66 1.31 Pinnixa patagoniensis 0.39 0.10 2.66 1.31 Pinnixa brevipollex 1.25 0.33 6.64 10.50 Dissodactylus 0.04 <0.01 | Collodes rostratus | 0.12 | 0.10 | 0.33 | 0.07 | |
| Libinia spinosa 0.16 1.93 1.33 2.77 Pelia rotunda 0.09 0.04 0.33 0.04 Atelecyclidae Peltarion spinosulum 0.39 0.49 0.66 0.59 Portunidae 0.04 0.02 0.33 1.54 Aunthidae Pilimnus reticulatus 0.04 0.02 0.33 0.02 Belliidae Corystoides abbreviatus 0.62 0.21 1.33 1.12 Grapsidae Cyrtograpsus affinis 4.26 1.09 7.97 42.72 Pinnotheridae Pinnixa patagoniensis 0.39 0.10 2.66 1.31 1.12 Pinnixa patagoniensis 0.39 0.10 2.66 1.31 1.50 | Leurocyclus | 0.04 | 0.05 | 0.33 | 0.03 | |
| Pelia rotunda 0.09 0.04 0.33 0.04 | tuberculosus | | | | | |
| Atelecyclidae Reltarion spinosulum 0.39 0.49 0.66 0.59 Portunidae 0040/lipes trimaculatus 0.59 0.57 1.33 1.54 Vanithidae Pilimnus reticulatus 0.04 0.02 0.33 0.02 Belliidae Corystoides abbreviatus 0.62 0.21 1.33 1.12 Grapsidae Cyrtograpsus affinis 4.26 1.09 7.97 42.72 Pinnotheridae Pinnixa patagoniensis 0.39 0.10 2.66 1.31 Pinnixa brevipollex 1.25 0.33 6.64 10.50 Dissodactylus 0.04 <0.01 0.33 0.01 Crinitichelis 8.99 0.55 7.97 76.13 0.98 Amphipoda 8.99 0.55 7.97 76.13 0.98 Unidentified amphipods 8.53 0.54 6.31 57.23 Ampeliscidae 0.43 0.01 1.33 0.58 Caprellidae 0.27 0.01 0.33 | Libinia spinosa | 0.16 | 1.93 | 1.33 | 2.77 | |
| Peltarion spinosulum | Pelia rotunda | 0.09 | 0.04 | 0.33 | 0.04 | |
| Portunidae | Atelecyclidae | | | | | |
| Ovalipes trimaculatus 0.59 0.57 1.33 1.54 Xanthidae Pilimnus reticulatus 0.04 0.02 0.33 0.02 Belliidae Corystoides abbreviatus 0.62 0.21 1.33 1.12 Grapsidae Cyrtograpsus affinis 4.26 1.09 7.97 42.72 Pinnixa patagoniensis 0.39 0.10 2.66 1.31 Pinnixa brevipollex 1.25 0.33 6.64 10.50 Dissodactylus crinitichelis 0.04 <0.01 0.33 0.01 Amphipoda 8.99 0.55 7.97 76.13 0.98 Unidentified amphipods 8.53 0.54 6.31 57.23 Ampeliscidae 0.43 0.01 1.33 0.58 Caprellidae 0.04 <0.01 0.33 0.01 Cumacea 34.54 1.98 14.20 14.20 Diastyliidae 32.04 1.83 15.61 528.9 Isopoda 0.43 0.09 | Peltarion spinosulum | 0.39 | 0.49 | 0.66 | 0.59 | |
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| Pilimnus reticulatus 0.04 0.02 0.33 0.02 Belliidae Corystoides abbreviatus 0.62 0.21 1.33 1.12 Grapsidae Cyrtograpsus affinis 4.26 1.09 7.97 42.72 Pinnotheridae Pinnixa patagoniensis 0.39 0.10 2.66 1.31 Pinnixa brevipollex 1.25 0.33 6.64 10.50 Dissodactylus crinitichelis 0.04 <0.01 0.33 0.01 Amphipoda 8.99 0.55 7.97 76.13 0.98 Unidentified amphipods 8.53 0.54 6.31 57.23 Ampeliscidae 0.43 0.01 1.33 0.58 Caprellidae 0.04 <0.01 0.33 0.01 Cumacea 34.54 1.98 17.61 643.2 8.31 Unidentified cumaceans 0.27 0.01 0.33 0.09 Bodotiriidae 2.23 0.14 5.98 14.20 Disstylidae 5.98< | Ovalipes trimaculatus | 0.59 | 0.57 | 1.33 | 1.54 | |
| Belliidae Corystoides abbreviatus 0.62 0.21 1.33 1.12 Grapsidae Cyrtograpsus affinis 4.26 1.09 7.97 42.72 Pinnotheridae Pinnixa patagoniensis 0.39 0.10 2.66 1.31 Pinnixa brevipollex 1.25 0.33 6.64 10.50 Dissodactylus 0.04 <0.01 0.33 0.01 Ciritichelis Amphipoda 8.99 0.55 7.97 76.13 0.98 Unidentified amphipods 8.53 0.54 6.31 57.23 57.23 Ampeliscidae 0.43 0.01 1.33 0.08 Caprellidae 0.44 0.01 0.33 0.01 0.08 Caprellidae 0.04 <0.01 0.33 0.09 Cumacea 34.54 1.98 17.61 643.2 8.31 Unidentified cumaceans 0.27 0.01 0.33 0.09 146.8 1.89 Isopoda 5.98 0.81 21.59 14 | Xanthidae | | | | | |
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| Grapsidae Cyrtograpsus affinis 4.26 1.09 7.97 42.72 Pinnotheridae Pinnixa patagoniensis 0.39 0.10 2.66 1.31 Pinnixa patagoniensis 0.03 0.10 2.66 1.31 Pinnixa brevipollex 1.25 0.33 6.64 10.50 Dissodactylus crinitichelis 0.04 <0.01 | Belliidae | | | | | |
| Cyrtograpsus affinis 4.26 1.09 7.97 42.72 Pinnotheridae Pinnixa patagoniensis 0.39 0.10 2.66 1.31 Pinnixa brevipollex 1.25 0.33 6.64 10.50 Dissodactylus 0.04 <0.01 | Corystoides abbreviatus | 0.62 | 0.21 | 1.33 | 1.12 | |
| Pinnotheridae Pinnixa patagoniensis 0.39 0.10 2.66 1.31 Pinnixa brevipollex 1.25 0.33 6.64 10.50 Dissodactylus 0.04 <0.01 0.33 0.01 crinitichelis 0.04 <0.01 0.33 0.01 Amphipoda 8.99 0.55 7.97 76.13 0.98 Unidentified amphipods 8.53 0.54 6.31 57.23 Ampeliscidae 0.43 0.01 1.33 0.58 Caprellidae 0.04 <0.01 0.33 0.01 Cumacea 34.54 1.98 17.61 643.2 8.31 Unidentified cumaceans 0.27 0.01 0.33 0.09 Bodotriidae 2.23 0.14 5.98 14.20 Diastylidae 32.04 1.83 15.61 528.9 Isopoda 5.98 0.81 21.59 146.8 1.89 Unidentified isopods 0.43 0.09 3.65 1.89 | Grapsidae | | | | | |
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| Sopoda 5.98 0.81 21.59 146.8 1.89 Unidentified isopods 0.43 0.09 3.65 1.89 Sphaeromatidae Sphaeroma serratum 0.16 <0.01 0.99 0.16 Cirolanidae Excirolana armata 0.12 <0.01 0.99 0.12 Cirolana spp. 0.04 0.01 0.33 0.02 Serolidae Serolis spp. 0.39 0.04 3.32 1.45 Serolis gaudichaudii 0.08 0.02 0.66 0.06 Serolis marplatensis 2.46 0.42 9.30 26.84 Serolis polaris 0.09 0.06 0.66 0.09 Idoteidae Idoteidae Idotea baltica 1.68 0.12 3.65 6.59 Macrochiriditea spp. 0.55 0.02 2.66 1.52 Cephalochordata 0.43 0.03 1.99 0.93 0.01 Branchiostomidae Branchiostomidae Branchiostomidae 8 Franchiostoma platae 0.43 0.03 1.99 0.93 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.36 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 0.03 D.05 Constitution 0.04 0.05 0.33 0.03 0.05 Constitution 0.05 0.05 0.03 0.05 | | | | | | |
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| Cirolanidae Excirolana armata 0.12 <0.01 0.99 0.12 Cirolana spp. 0.04 0.01 0.33 0.02 Serolidae Serolis spp. 0.39 0.04 3.32 1.45 Serolis gaudichaudii 0.08 0.02 0.66 0.06 Serolis marplatensis 2.46 0.42 9.30 26.84 Serolis polaris 0.09 0.06 0.66 0.09 Idoteidae 1.68 0.12 3.65 6.59 Macrochiriditea spp. 0.55 0.02 2.66 1.52 Cephalochordata 0.43 0.03 1.99 0.93 0.01 Branchiostomidae Branchiostoma platae 0.43 0.03 1.99 0.93 Mysidacea 9.47 0.36 3.99 39.19 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.35 3.98 4.36 Maldanidae 0.08< | · | 0.16 | <0.01 | 0.00 | 0.16 | |
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| Serolis gaudichaudii 0.08 0.02 0.66 0.06 Serolis marplatensis 2.46 0.42 9.30 26.84 Serolis polaris 0.09 0.06 0.66 0.09 Idoteidae 1.68 0.12 3.65 6.59 Macrochiriditea spp. 0.55 0.02 2.66 1.52 Cephalochordata 0.43 0.03 1.99 0.93 0.01 Branchiostomidae Branchiostomidae 8 0.03 1.99 0.93 0.01 Mysidacea 9.47 0.36 3.99 39.19 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.35 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | | 0.39 | 0.04 | 3 32 | 1 45 | |
| Serolis marplatensis 2.46 0.42 9.30 26.84 Serolis polaris 0.09 0.06 0.66 0.09 Idoteidae 0.09 0.06 0.66 0.09 Idoteidae 0.00 0.00 0.00 0.00 Macrochiriditea spp. 0.55 0.02 2.66 1.52 Cephalochordata 0.43 0.03 1.99 0.93 0.01 Branchiostomidae 0.43 0.03 1.99 0.93 0.01 Mysidacea 9.47 0.36 3.99 39.19 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.36 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | | | | | | |
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| Idoteidae Idotea baltica 1.68 0.12 3.65 6.59 Macrochiriditea spp. 0.55 0.02 2.66 1.52 Cephalochordata 0.43 0.03 1.99 0.93 0.01 Branchiostomidae 0.43 0.03 1.99 0.93 0.51 Mysidacea 9.47 0.36 3.99 39.19 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.36 5.98 6.61 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | ' | | | | | |
| Macrochiriditea spp. 0.55 0.02 2.66 1.52 Cephalochordata 0.43 0.03 1.99 0.93 0.01 Branchiostomidae 0.43 0.03 1.99 0.93 0.51 Mysidacea 9.47 0.36 3.99 39.19 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.36 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | ' | | | | | |
| Macrochiriditea spp. 0.55 0.02 2.66 1.52 Cephalochordata 0.43 0.03 1.99 0.93 0.01 Branchiostomidae 0.43 0.03 1.99 0.93 0.51 Mysidacea 9.47 0.36 3.99 39.19 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.36 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | Idotea baltica | 1.68 | 0.12 | 3.65 | 6.59 | |
| Cephalochordata 0.43 0.03 1.99 0.93 0.01 Branchiostomidae 0.43 0.03 1.99 0.93 0.91 Mysidacea 9.47 0.36 3.99 39.19 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.36 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | | | | | | |
| Branchiostomidae Branchiostoma platae 0.43 0.03 1.99 0.93 Mysidacea 9.47 0.36 3.99 39.19 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.36 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | • • • | | | | | 0.01 |
| Mysidacea 9.47 0.36 3.99 39.19 0.51 Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.36 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | • | | | | | |
| Polychaeta 1.79 1.04 11.96 33.92 0.44 Unidentified polychaetes 0.74 0.36 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | Branchiostoma platae | 0.43 | 0.03 | 1.99 | 0.93 | |
| Unidentified polychaetes 0.74 0.36 5.98 6.61 Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | Mysidacea | 9.47 | 0.36 | 3.99 | 39.19 | 0.51 |
| Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | • | | | | | |
| Flabelligeridae 0.74 0.35 3.98 4.36 Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | Unidentified polychaetes | 0.74 | 0.36 | 5.98 | 6.61 | |
| Maldanidae 0.08 0.25 0.66 0.22 Nephthyidae 0.04 0.05 0.33 0.03 | | 0.74 | 0.35 | 3.98 | 4.36 | |
| . , | | 0.08 | 0.25 | 0.66 | 0.22 | |
| Onuphidae 0.16 0.02 1.33 0.23 | Nephthyidae | 0.04 | 0.05 | 0.33 | 0.03 | |
| | Onuphidae | 0.16 | 0.02 | 1.33 | 0.23 | |

Table 1. Continued

| %N | %M | %FO | IRI | %IRI |
|------|------------------------------|---|---|----------------|
| | | | | |
| 0.04 | < 0.01 | 0.33 | 0.01 | |
| 0.12 | 0.25 | 0.99 | 0.36 | < 0.01 |
| | | | | |
| 0.04 | 0.05 | 0.33 | 0.03 | |
| | | | | |
| 0.04 | 0.09 | 0.33 | 0.05 | |
| | | | | |
| | | | | |
| 0.04 | 0.10 | 0.33 | 0.05 | |
| | 0.04 0.12 0.04 0.04 | 0.04 <0.01 0.12 0.25 0.04 0.05 0.04 0.09 | 0.04 <0.01 0.33 0.12 0.25 0.99 0.04 0.05 0.33 0.04 0.09 0.33 | 0.04 <0.01 |

However, *S. acuta* has a higher trophic level than expected from its body size, especially for mature individuals, with similar values to large-bodied marine predators, such as some mammals (Pauly *et al.* 1998; Tucker & Rogers 2014) and large skates and sharks (Cortés 1999; Ebert & Bizzarro 2007). This unexpectedly high trophic level for a small elasmobranch suggests that *S. acuta* may play a unique functional role in Southwest Atlantic food webs.

Ontogenetic diet shifts in many skates are usually a function of body size (Bizzarro et al. 2007; Robinson et al. 2007) and the effect of reproductive condition (i.e. maturity stage) on feeding habits is less known. In S. acuta, maturity stage has a strong effect in producing ontogenetic dietary shifts: invertebrates (i.e. shrimps, crabs, isopods, and cumaceans) are preyed upon by immature individuals, while teleosts are consumed mainly by mature individuals. This pattern may occur by the combination of an improved ability to capture prey and a change in energy demand of large individuals. Mature individuals may be more efficient predators than immature individuals. An increase in the metabolic requirements that mature individuals need for reproductive success may also contribute to generate the observed pattern. There is no single explanation for this pattern, but the effect of maturity stage on dietary shifts may be part of a more complex behavioral response to maturation (Koen Alonso *et al.* 2001). This pattern is reflected in the higher trophic level of mature individuals compared with immature individuals.

Spatial and seasonal differences in the diet composition of S. acuta were found. In the study area, the same spatial patterns in diet composition were found for both the Rio skate, Rioraja agassizii (Barbini & Lucifora 2011), and the spotback skate, Atlantoraja castelnaui (Barbini & Lucifora 2012b): crustaceans were consumed more in the north region and teleosts were more preved upon in the south region. Possible regional differences in prev availability resulting from differences in environmental conditions and oceanographic characteristics between the north and south regions (Acha et al. 2004; Giberto et al. 2004) may explain this dietary pattern. Seasonal variations in the diet composition of skates have also been found previously in the study area (e.g. Psammobatis extenta, Braccini & Perez 2005; R. agassizii, Barbini & Lucifora 2011; A. castelnaui, Barbini & Lucifora 2012b). Seasonal fluctuations in the abundance and distribution of prey may explain this pattern (Muto et al. 2001).

The feeding activity of *S. acuta* varied clearly between seasons and regions. The effect of season in producing a shift in feeding activity cannot be explained by the seasonal reproductive cycle. The level of feeding activity of *S. acuta* was independent of maturity stage. Spatially, the higher feeding activity in the north region than in the south region might be explained by the same reason as the spatial differences in diet composition described above.

The responses of prey to predators can have important effects on prey refuge use, activity, vigilance, and feeding and mating behavior (Sih *et al.* 2000). Besides, the level of perceived predation risk by prey can vary over time owing to seasonal changes in predator occurrence (Lima & Dill 1990; Sih *et al.* 2000). From a trophic viewpoint, some times are more dangerous for feeding than others, influencing foraging activity of prey because predation

Table 2. Best models explaining the consumption (in number of individuals) of the prey categories of Sympterygia acuta.

| prey | intercept | coefficients | AIC | W |
|-------------|-----------|-------------------------------|--------|-------|
| teleosts | -0.444 | -1.181 immature + 0.534 south | 513.85 | 0.671 |
| shrimps | -0.286 | 1.403 immature - 0.457 south | 913.99 | 0.428 |
| crabs | -0.646 | 1.155 immature – 1.981 south | 616.92 | 0.836 |
| amphipods | 3.545 | -0.012 TL | 229.66 | 0.458 |
| cumaceans | -22.314 | 22.031 immature + 2.609 cold | 633.26 | 0.882 |
| isopods | -4.541 | 3.789 immature + 1.039 cold | 425.18 | 0.845 |
| polychaetes | -3.788 | 1.606 immature + 1.254 cold | 245.49 | 0.477 |

The coefficients of Maturity stage and Region are relative to Mature and North, respectively. AIC = Akaike information criterion; w = Akaike's weight; TL = total length.

Table 3. Best models explaining the number of prey species consumed (N_{species}), number of prey consumed (N_{prey}) and proportion of empty stomachs of *Sympterygia acuta*.

| response variable | intercept | coefficients | AIC | W |
|------------------------------------|-----------|---|--------|-------|
| N_{species} | 0.338 | 0.235 cold – 0.235 south + 0.244 log(N _{prey}) | 855.05 | 0.297 |
| N_{prey} | 2.251 | 0.636 cold – 1.538 south | 1788.8 | 0.875 |
| proportion of empty stomachs | -1.333 | -1.087 cold + 1.090 south | 453.74 | 0.989 |

AIC = Akaike information criterion; w = Akaike's weight.

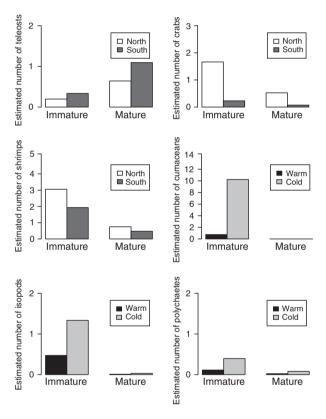


Fig. 3. Changes in consumption of prey categories (by number) with maturity stage, season and region of *Sympterygia acuta* from off Uruguay and North Argentina estimated by generalized linear models. The models had a log link and a negative binomial error distribution.

risk varies seasonally (Lima & Dill 1990). Off North Argentina, *S. acuta* is subject to the threat of predation by large sharks, such as the broadnose sevengill shark, *Notorynchus cepedianus* (Lucifora *et al.* 2005), the school shark, *Galeorhinus galeus* (Lucifora *et al.* 2006), and the sand tiger shark, *Carcharias taurus* (Lucifora *et al.* 2009). In the Southwest Atlantic, these sharks migrate seasonally

in coastal waters among Brazil, Uruguay and Argentina, being present during spring and summer in shallow waters off Northern Argentina (Lucifora 2003). Sympterygia acuta may reduce its feeding activity seasonally as a response to the presence of these large predators in the warm season. Possibly, this pattern may be associated with a change in foraging behavior (i.e. less frequent feeding activity) as a predator avoidance response.

Uruguayan and Northern Argentinean coastal waters are among the most heavily impacted by fisheries in the Southwest Atlantic (Tyedmers et al. 2005). Sympterygia acuta is captured by the multi-species fleet that exploits the coastal demersal fish assemblage (Massa et al. 2004). In addition, this species is caught commonly by recreational fishermen along the coast of Buenos Aires Province, Argentina (Cedrola et al. 2011). The coastal habitat of S. acuta is exposed to a high intensity of bottom trawling (Brazeiro et al. 2003) that alters the physical structure of the bottom, affecting indirectly the benthic community (Kaiser 1998; Watling & Norse 1998). Sympterygia acuta completes its life cycle from egg to adult in very shallow waters (Mabragaña et al. 2014) and feeds mainly on benthic prey. Therefore, this species is threatened by both heavy fishing and habitat destruction. Consequently, its biomass has declined (Massa & Hozbor 2004, 2011) and the International Union for Conservation of Nature has assessed this species as Vulnerable (Massa & Hozbor 2004). All of this increases the concern and the need for appropriate measures for the conservation of this species.

Conclusions

Our results indicate that *Sympterygia acuta* feeds on a broad spectrum of prey, but teleosts are the most important followed by decapods. In this skate, maturity stage is a strong determinant of the ontogenetic diet shift that is associated with region and season. The level of feeding activity is also associated with season and region. *Sympterygia acuta* is peculiar among the skates studied so far in having a high trophic level despite its rather small body size, which indicates that it may have a unique functional role in Southwest Atlantic food webs.

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References

- Acha E.A., Mianzan H.W., Guerrero R.A., Favero M., Bava J. (2004) Marine fronts at the continental shelves of austral South America. Physical and ecological processes. *Journal of Marine Systems*, **44**, 83–105.
- Arim M., Bozinovic F., Marquet P.A. (2007) On the relationship between trophic position, body mass and temperature: reformulating the energy limitation hypothesis. *Oikos*, **116**, 1524–1530.
- Barbini S.A., Lucifora L.O. (2011) Feeding habits of the Rio Skate, *Rioraja agassizi* (Chondrichthyes, Rajidae), from off Uruguay and north Argentina. *Journal of the Marine Biological Association of the United Kingdom*, **91**, 1175–1184.
- Barbini S.A., Lucifora L.O. (2012a) Ontogenetic diet shifts and food partitioning between two small sympatric skates (Elasmobranchii, Rajidae) in the Southwestern Atlantic. *Marine and Freshwater Research*, **63**, 905–913.
- Barbini S.A., Lucifora L.O. (2012b) Feeding habits of a large endangered skate from the Southwest Atlantic: the spotback skate *Atlantoraja castelnaui*. *Marine and Freshwater Research*, **63**, 180–188.
- Bizzarro J.J., Robinson H.J., Rinewalt C.S., Ebert D.A. (2007) Comparative feeding ecology of four sympatric skate species off central California, USA. *Environmental Biology of Fishes*, **80**, 197–220.
- Boschi E.E., Fischbach C.E., Iorio M.I. (1992) Catálogo ilustrado de los crustáceos estomatópodos y decápodos marinos de Argentina. *Frente Marítimo*, **10**, 7–94.
- Bovcon N.D., Cochia P.D., Góngora M.E., Gosztonyi A.E. (2011) New records of warm-temperate water fishes in central Patagonian coastal waters (Southwestern South Atlantic Ocean). *Journal of Applied Ichthyology*, **27**, 832–839.
- Braccini J.M., Perez J.E. (2005) Feeding habits of the sand skate *Psammobatis extenta* (Garman, 1913): sources of variation in dietary composition. *Marine and Freshwater Research*, **56**, 395–403.
- Brazeiro A., Acha E.M., Mianzan H.W., Gómez M., Fernández V. (2003) Aquatic Priority Areas for the Conservation and Management of the Ecological Integrity of the Río de la Plata and its Maritime Front. FrePlata, Proyecto PNUD-GEF RLA/99/631, Montevideo-Buenos Aires: 81 pp.
- Cedrola P., Bovcon N., Bruno C., Bustamante C., Caille G., Calvo S., Chiaramonte G., García Liotta R., González R., Laura D., Massola V., Mendia L., Moronta M., Perier R., Remes Lenicov M., Tombesi M.L. (2011) La pesca deportiva de condrictios en el Mar Argentino. In: Wöhler O.C., Cedrola P., Cousseau M.B. (Eds), Contribuciones sobre biología, pesca y comercialización de tiburones en la Argentina. Aportes para la elaboración del Plan de Acción Nacional. Consejo Federal Pesquero, Buenos Aires: 193–205.
- Colonello J.H., Christiansen H.E., Macchi G.J. (2011) Escala de madurez sexual para peces cartilaginosos de la Plataforma Continental Argentina. In: Wöhler O.C., Cedrola

- P., Cousseau M.B. (Eds), Contribuciones sobre biología, pesca y comercialización de tiburones en la Argentina. Aportes para la elaboración del Plan de Acción Nacional. Consejo Federal Pesquero, Buenos Aires: 115–127.
- Cortés E. (1997) A critical review of methods of studying fish feeding based on analysis of stomach contents: application to elasmobranch fishes. *Canadian Journal of Fisheries and Aquatic Sciences*, **54**, 726–738.
- Cortés E. (1999) Standardized diet compositions and trophic levels of sharks. *ICES Journal of Marine Science*, **56**, 707–717.
- Cousseau M.B., Perrotta R.G. (2000) Peces marinos de Argentina: biología, distribución y pesca. Instituto Nacional de Investigación y Desarrollo Pesquero, Mar del Plata: 167.
- Cousseau M.B., Figueroa D.E., Díaz de Astarloa J.M., Mabragaña E., Lucifora L.O. (2007) Rayas, chuchos y otros batoideos del Atlántico Sudoccidental (34°S-55°S). Instituto Nacional de Investigación y Desarrollo Pesquero, Mar del Plata: 102.
- Crawley M.J. (2005) Statistics. An introduction using R. Wiley, London: 327.
- Ebert D.A., Bizzarro J.J. (2007) Standardized diet compositions and trophic levels of skates (Chondrichthyes, Rajiformes, Rajoidei). *Environmental Biology of Fishes*, **80**, 221–237.
- Ebert D.A., Cowley P.O., Compagno L.J. (1991) A preliminary investigation of the feeding ecology of skates (Batoidea: Rajidae) off west coast of southern Africa. *South African Journal of Marine Science*, **10**, 71–81.
- Franklin A.B., Sheik T.M., Anderson D.R., Burnham K.P. (2001) Statistical model selection: an alternative to null hypothesis testing. In: Shenk T.M., Franklin A.M. (Eds), *Modeling in Natural Resources Management: Development, Interpretation, and Application*. Island Press, Washington: 75, 90
- Garrison L.P., Link J.S. (2000) Fishing effects on spatial distribution and trophic guild structure of the fish community in the Georges Bank region. *ICES Journal of Marine Science*, **57**, 723–730.
- Giberto D.A., Bremec C.S., Acha E.M., Mianzan H. (2004) Large-scale spatial patterns of benthic assemblages in the SW Atlantic: the Río de la Plata estuary and adjacent shelf waters. *Estuarine, Coastal and Shelf Science*, **61**, 1–13.
- Guerrero R.A., Piola R.A. (1997) Masas de agua de la plataforma continental. In: Boschi E.E. (Ed.), *El Mar Argentino y sus Recursos Pesqueros*, Vol 1. Instituto Nacional de Investigación y Desarrollo Pesquero, Mar del Plata: 107–118.
- Hairston N.G., Hairston N.G. (1993) Cause-effect relationships in energy flow, trophic structure, and interspecific interactions. *The American Naturalist*, **143**, 379–411.
- Jaureguizar A.J., Menni R., Lasta C., Guerrero R. (2006) Fish assemblages of the northern Argentine coastal system: spatial patterns and their temporal variations. *Fisheries Oceanography*, 14, 326–344.

- Jennings S., Pinnegar J.K., Polunin N.V.C., Boon T.W. (2001) Weak cross-species relationships between body size and trophic level belie powerful size-based trophic structuring in fish communities. *Journal of Animal Ecology*, **70**, 934–944.
- Johnson J.B., Omland K.S. (2004) Model selection in ecology and evolution. *Trends in Ecology and Evolution*, 19, 101–108.
- Kaiser M.J. (1998) Significance of bottom-fishing disturbance. *Conservation Biology*, **12**, 1230–1235.
- Kajiura S.M. (2001) Head morphology and electrosensory pore distribution of carcharhinid and sphyrnid sharks. *Environmental Biology of Fishes*, 61, 125–133.
- Koen Alonso M., Crespo E.A., Gracía N.A., Pedraza S.N., Mariotti P.A., Berón Vega B., Mora N.J. (2001) Food habits of *Dipturus chilensis* (Pisces: Rajidae) off Patagonia, Argentina. *ICES Journal of Marine Science*, 58, 288–297.
- Kyne P.M., Courtney A.J., Bennett M.B. (2008) Aspects of reproduction and diet of the Australian endemic skate *Dipturus polyommata* (Ogilby) (Elasmobranchii: Rajidae), by-catch of a commercial prawn trawl fishery. *Journal of Fish Biology*, **72**, 61–77.
- Lima S.L., Dill L.M. (1990) Behavioral decisions made under the risk of predation: a review and prospectus. *Canadian Journal of Zoology*, **68**, 619–640.
- Lucas A.J., Guerreo R.A., Mianzan H.W., Acha E.M., Lasta C.A. (2005) Coastal oceanographic regimes of the Northern Argentine Continental Shelf (34°–43°S). *Estuarine, Coastal and Shelf Science*, **65**, 405–420.
- Lucifora L.O. (2003) Ecología y conservación de los grandes tiburones costeros de Bahía Anegada, Provincia de Buenos Aires, Argentina. PhD thesis, Universidad Nacional de Mar del Plata, Mar del Plata, Argentina.
- Lucifora L.O., García V.B. (2004) Gastropod predation on egg cases of skates (Chondrichthyes, Rajidae) in the Southwestern Atlantic: quantification and life history implications. *Marine Biology*, **145**, 917–922.
- Lucifora L.O., Valero J.L., Bremec C.S., Lasta M.L. (2000)
 Feeding habits and prey selection by the skate *Dipturus chilensis* (Elasmobranchii: Rajidae) from the south-western Atlantic. *Journal of the Marine Biological Association of the United Kingdom*, **80**, 953–954.
- Lucifora L.O., Menni R.C., Escalante A.H. (2005)
 Reproduction, abundance and feeding habits of the broadnose sevengill shark *Notorynchus cepedianus* in north Patagonia, Argentina. *Marine Ecology Progress Series*, **289**, 237–244.
- Lucifora L.O., García V.B., Menni R.C., Escalante A.H. (2006) Food habits, selectivity, and foraging modes of the school shark, *Galeorhinus galeus*. *Marine Ecology Progress Series*, **315**, 259–270.
- Lucifora L.O., García V.B., Escalante A.H. (2009) How can the feeding habits of the sand tiger shark influence the success of conservation programs? *Animal Conservation*, **12**, 291–301.
- Mabragaña E., Lucifora L.O., Corbo M. L., Díaz de Astarloa J.M. (2014) Seasonal reproductive biology of the bignose

- fanskate *Sympterygia acuta* (Chondrichthyes, Rajidae). *Estuaries and Coasts*, **38**, 1466–1476.
- Magurran A.E. (2004) Measuring Biological Diversity. Blackwell, Malden: 256.
- Massa A., Hozbor N.M. (2004) Sympterygia acuta. In: IUCN 2013. IUCN Red List of Threatened Species. Version 2013.1. www.iucnredlist.org. Downloaded on 28 October 2013.
- Massa A.M., Hozbor N.M. (2011) Evolución de las estimaciones de abundancia de los peces carilaginosos demersales de mayor valor comercial del Atlántico Sudoccidental, capturados entre 34° y 41° S, a profundidades menores de 50 m. In: Wöhler O.C., Cedrola P., Cousseau M.B. (Eds), Contribuciones sobre biología, pesca y comercialización de tiburones en la Argentina. Aportes para la elaboración del Plan de Acción Nacional. Consejo Federal Pesquero, Buenos Aires: 193–205.
- Massa A., Lucifora L.O., Hozbor N.M. (2004) Condrictios de las regiones costeras bonaerenses y uruguayas. In: Boschi E.E. (Ed.), El Mar Argentino y sus recursos pesqueros, Tomo 4. Instituto Nacional de Investigación y Desarrollo Pesquero, Mar del Plata: 85–99.
- Muto E.Y., Soares L.S.H., Goiten R. (2001) Food resource utilization of the skates *Rioraja agassizi* and *Psammobatis extenta* on the continental shelf off Ubatuba, south-eastern Brazil. *Revista Brasileira de Biologia*, **61**, 217–238
- Orlov A.M. (1998) The diets and feeding habits of some deepwater benthic skates (*Rajidae*) in the Pacific waters off the Northern Kuril Islands and Southeastern Kamchatka. *Alaska Fishery Research Bulletin*, **5**, 1–17.
- Pauly D., Trites A.W., Capuli E., Christensen V. (1998) Diet composition and trophic levels of marine mammals. *ICES Journal of Marine Science*, 55, 467–488.
- Pinkas L.M., Oliphant S., Iverson I.L.K. (1971) Food habits of alba-core, bluefin tuna and bonito in Californian waters. California Fish and Game, 152, 1–105.
- R Core Team (2014) R: a Language and Environment for Statistical Computing. R Foundation for Statistical Computing, Vienna.
- Robinson H.J., Cailliet G.M., Ebert D.A. (2007) Food habits of the longnose skate, *Raja rhina* (Jordan and Gilbert, 1880), in central California waters. *Environmental Biology of Fishes*, **80**, 165–179.
- Romanuk T.N., Hayward A., Hutchings J.A. (2011) Trophic level scales positively with body size in fishes. *Global Ecology and Biogeography*, **10**, 231–240.
- Sih A., Ziemba R., Harding K.C. (2000) New insights on how temporal variation in predation risk shapes prey behavior. *Trends in Ecology and Evolution*, **15**, 3–4.
- Stehmann M.F.W. (2002) Proposal of a maturity stages scale for oviparous and viviparous cartilaginous fishes (Pisces, Chondrichthyes). Archive of Fishery and Marine Research, 50, 23–48.
- Symonds M.E., Moussalli A. (2011) A brief guide to model selection, multimodel inference and model averaging in

- behavioural ecology using Akaike's information criterion. *Behavioral Ecology and Sociobiology*, **65**, 13–21.
- Treloar M.A., Laurenson L.J.B., Stevens J.D. (2007) Dietary comparisons of six skate species (Rajidae) in south-eastern Australian waters. *Environmental Biology of Fishes*, **80**, 181–196.
- Tucker M.A., Rogers T.L. (2014) Examining predator—prey body size, trophic level and body mass across marine and terrestrial mammals. *Proceedings of The Royal Society B*, **281**, 20142103.
- Tyedmers P.H., Watson R., Pauly D. (2005) Fueling global fishing fleets. *Ambio*, **34**, 635–638.

- Venables W.N., Ripley B.D. (2002) Modern Applied Statistics with S-Plus, 4th edn. Springer, New York: 495.
- Watling L., Norse E.A. (1998) Disturbance of the seabed by mobile fishing gear: a comparison to forest clear cutting. *Conservation Biology*, **12**, 1180–1197.
- Werner E.E., Gilliam G.F. (1984) The ontogenetic niche and species interactions in size-structured populations. *Annual Review of Ecology and Systematics*, **15**, 393–425.
- Wetherbee B.M., Cortés E. (2004) Food consumption and feeding habits. In: Carrier J.C., Musick J.A., Heithaus M.R. (Eds), *Biology of Sharks and their Relatives*. CRC Press, Boca Raton: 223–244.