

Biochemical and Molecular Study of the Influence of *Amaranthus hypochondriacus* Flour on Serum and Liver Lipids in Rats Treated with Ethanol

Viviana R. Lucero López · Gabriela S. Razzeto ·
Nora L. Escudero · M. Sofía Gimenez

Published online: 12 October 2013
© Springer Science+Business Media New York 2013

Abstract Hyperlipidemia and hepatic steatosis are frequent alterations due to alcohol abuse. Amaranth is a pseudocereal with hypolipidemic potential among other nutraceutical actions. Here we study the effect of *Amaranthus hypochondriacus* (*Ah*) seeds on serum and liver lipids, and the expression of genes associated to lipid metabolism and liver histology in male *Wistar* rats intoxicated with ethanol. The animals were divided into four groups; two groups were fed the American Institute of Nutrition 1993 for maintenance diet (AIN-93M), and the other two with AIN-93M containing *Ah* as protein source. One of each protein group received 20 % ethanol in the drinking water, thus obtaining: CC (control casein), EC (ethanol casein), *CAh* (control *Ah*) and *EAh* (ethanol *Ah*). When comparing *EAh* vs. EC, we found a positive effect of *Ah* on lipids, preventing the increment of serum cholesterol ($p < 0.001$), through the higher expression of the LDL receptor ($p < 0.001$); and it also decreased free ($p < 0.05$) and esterified cholesterol ($p < 0.01$) in liver, probably via the reduction of the 3-hydroxy-3-methylglutaryl coenzyme A reductase expression ($p < 0.001$). We also observed that amaranth contributed to the decrease of fat deposits in liver, probably through the decrease in acetyl-CoA carboxylase alpha ($p < 0.01$), glycerol-3-phosphate acyltransferase 1 ($p < 0.01$) and diacylglycerol O-acyltransferase 2 ($p < 0.05$) expression. The histological study showed a decrease in the fat deposits in the amaranth group when compared to casein;

this is consistent with the biochemical and molecular parameters studied in this work. In conclusion, amaranth could be recommended to avoid the alterations in the lipid metabolism induced by alcohol and other harmful agents.

Keywords *Amaranthus hypochondriacus* · Ethanol · Cholesterol · Triglycerides

Abbreviations

ACC α	Acetyl-CoA carboxylase alpha
<i>Ah</i>	<i>Amaranthus hypochondriacus</i>
AIN-93M	American Institute of Nutrition 1993 for maintenance diet
Apo B	Apolipoprotein B
BHT	Butylated hydroxytoluene
<i>CAh</i>	Control <i>Ah</i>
CC	Control casein
CPT-1	Carnitine palmitoil transferase 1
CYP7A1	Cholesterol 7-alpha-hydroxylase
DGAT-2	Diacylglycerol O-acyltransferase 2
<i>EAh</i>	Ethanol <i>Ah</i>
EC	Ethanol casein
GPAT-1	Glycerol-3-phosphate acyltransferase 1
HDL-C	High-density lipoprotein-cholesterol
HMG-CoAR	3-hydroxy-3-methylglutaryl coenzyme A reductase
LDL-C	Low-density lipoprotein-cholesterol
PPAR α	Peroxisome proliferator-activated receptor alpha
RT-PCR	Reverse transcription polymerase chain reaction
SREBP-2	Sterol regulatory element binding protein 2
TLC	Thin layer chromatography
Total-C	Total cholesterol
VLDL	Very-low-density lipoprotein

Electronic supplementary material The online version of this article (doi:10.1007/s11130-013-0388-3) contains supplementary material, which is available to authorized users.

V. R. L. López · G. S. Razzeto · N. L. Escudero · M. S. Gimenez
Department of Biochemistry and Biology Sciences, Faculty of Chemistry, Biochemistry and Pharmacy, National University of San Luis, IMIBIO-SL. CONICET, San Luis, Argentina

M. S. Gimenez (✉)
Chacabuco and Pedernera, San Luis 5700, Argentina
e-mail: marisofigime44@gmail.com

Introduction

Alcohol abuse affects all the socioeconomic levels in the world, generating serious problems in public health. Even though several studies have shown that a moderate intake of alcohol decreases the risk for cardiovascular diseases [1, 2], the chronic and excessive ingestion of ethanol induces changes in lipid metabolism, predisposing to the development of steatosis, hyperlipidemia and atherosclerosis [3]. On the other hand, it is important to remark that a healthy eating pattern such as the Mediterranean diet, consistent in whole grains, fruits and vegetables, is shown to be efficient in reducing cardiovascular risk and in improving the lipid profile [4].

Amaranth is a pseudocereal from the *Amaranthaceae* family that is grown in a wide range of agricultural and weather conditions. This small seed is an important source of vegetable protein, rich in methionine and lysine amino acids, being this last one a limiting in many grains. The grain has high levels of soluble fibers, magnesium, calcium, tocotrienols and unsaturated fatty acids, especially linoleic acid [5, 6]; there have also been reports about significant levels of squalene, an important precursor of all the steroids [7]. Besides its nutritional quality, this grain exerts several nutraceutical actions due to the presence of a series of phytochemical compounds with hypolipidemic action [8–11] and antihypertensive potential [12], as well as antioxidant capacity [13–15]. Amaranth is also a gluten free grain, so it can be incorporated in the diet of celiac patients and those allergic to cereals [16].

The aim of this work was to evaluate the *Amaranthus hypochondriacus* effects on cholesterol and triglycerides metabolism in the liver of rats that were sub-chronically treated with ethanol. To this end, we determined serum and liver lipids and the expression of genes related; in addition, a histological study was performed.

Materials and Methods

Plant Materials

Amaranthus hypochondriacus (*Ah*) var. *Antorcha* was provided by the Department of Agronomy at the National University of Río Cuarto, Argentina, from a 2009 harvest of an experimental cultivation. For the flour preparation, the dried whole grain was ground and sieved through a 200 μm nylon sieve.

Animals and Experimental Design

Twenty-four male *Wistar* rats weighing 200 g were used. They were divided into four groups of six rats each, housed in individual cages and maintained under a regular light–dark cycle, with free access to food and water. All groups were fed according to recommendations of the American Institute of

Nutrition 1993 (AIN-93M) for four weeks [17]. Two groups were fed with the AIN-93M diet and the other two with AIN-93 M containing *Ah* as protein source. One of each group received 20 % ethanol in the drinking water [18], thus obtaining: CC (control casein), EC (ethanol casein), *CAh* (control *Ah*) and *EAh* (ethanol *Ah*). An intake between 4 and 6 g/kg/day of alcohol per rat was estimated. All studies involving experimental animals were conducted in accordance to national and institutional guidelines for the protection of animal welfare. The study was approved by the Animal Care Committee of the National University of San Luis, Argentina.

Serum Parameters

Total cholesterol (total-C), HDL-cholesterol (HDL-C) and LDL-cholesterol (LDL-C) [19] were determined by enzymatic methods, using commercial kits (Wiener, Laboratorios S.A.I.C., Rosario Argentina). Proteins were determined by the Biuret method [20] with bovine serum albumin as standard.

Hepatic Lipids Determinations

Lipids were extracted from liver tissue [21]. Total lipids were determined by dry weight [22]. Lipids were resuspended in hexane and total cholesterol was determined using an aliquot [23, 24]. And other aliquots of the extracts were used for separating the different lipids on thin-layer chromatography (TLC). The lipids were detected by exposing the plates to iodine vapors; they were scraped off and were used for the determination of triglycerides [25] and free and esterified cholesterol [23]. Proteins were determined by the Biuret method [20].

Analysis of mRNA Expression by RT-PCR

The relative mRNA quantities of the genes coding for hydroxy-methylglutaryl coenzyme A reductase (HMG-CoAR), LDL receptor (LDLr), sterol regulatory element binding protein 2 (SREBP-2), cholesterol 7- α -hydroxylase (CYP7A1), apolipoprotein B (ApoB), glycerol-3-phosphate acyltransferase 1 (GPAT-1), diacylglycerol O-acyltransferase 2 (DGAT-2); acetyl-CoA carboxylase alpha (ACC α), peroxisome proliferator-activated receptor alpha (PPAR α) and carnitine palmitoyl transferase 1 (CPT-1), were determined by reverse transcription polymerase chain reaction (RT-PCR). The sequences of the different primers and sizes of the PCR products are shown in Table S1.

The PCR products were analyzed and photographed. The intensity of each band was measured using the NIH Image software Scion. The relative abundance of each band was then normalized according to the housekeeping gene β -actin, calculated as the ratio of the intensity values of each product to that of β -actin, and it was reported in arbitrary units (AU).

Morphological Study

A histological study of the liver was performed by light microscopy. Sections were stained with hematoxylin and eosin, and photographs were taken using a Leitz Dialux optical microscope supplied with a Leica camera (100×).

Statistical Analysis

Results are expressed as mean \pm standard deviations. Statistical differences were tested by using a one-way analysis of variance (ANOVA). A probability of 0.05 or less indicates significant difference [26].

Results and Discussion

The aim of this study was to elucidate the possible effects of whole amaranth flour on the lipids content, using an experimental model of *Wistar* rats treated with ethanol (20 % ethanol solution) for four weeks. Serum and liver lipids were measured, as well as some genes involved in the lipid metabolism. In addition, a histological study of the liver was performed. We found that in presence of ethanol, *Ah* flour showed a beneficial effect on these parameters.

Table 1 shows that serum total-C and the lipoprotein fraction LDL + VLDL-C decreased in *EAh* vs. EC, as well as in *CAh* vs CC ($p < 0.001$); while there were no significant differences in HDL-C. In addition, EC showed the lowest value in the HDL/total cholesterol ratio when compared to the other groups. These results suggest susceptibility of EC to the harmful action of ethanol, and also the protective effect of amaranth in *EAh*.

Table 2 indicates that in EC vs. CC, the cholesterol and triglycerides liver content increased ($p < 0.05$) and ($p < 0.01$), respectively; free and esterified cholesterol also increased ($p < 0.05$) and ($p < 0.01$), respectively. In *EAh*, when compared to EC, there was a trend of decreasing cholesterol (13.7 %) and triglycerides (11.5 %); also a decrease of free ($p < 0.05$) and esterified ($p < 0.01$) cholesterol was observed. From these data we infer that in the groups exposed to ethanol, amaranth

would have a beneficial role in reducing the accumulation of lipids in the liver.

Since ethanol is metabolized in the liver and its effect on the lipid profile is complex, the more frequent alterations are reflected in the levels of lipoproteins and in the hepatic steatosis. The information available on alcohol impact on lipids is very controversial. In general, alcohol intake is frequently associated to an increase of HDL-C in blood and esterified cholesterol in liver [27]. Moreover, it is known that ethanol inhibits the release of VLDL particles, predisposing the development of fat liver [3]. All the modifications induced by ethanol can be modulated by molecular, genetic and environmental mechanisms; in addition, the patient factors are determinant as well as the multiple interactions of ethanol with the living organism.

Several components of grains have the ability to reverse some alterations in lipid metabolism. It has been suggested that the vegetal protein induces modifications in the cholesterol metabolism by the action of bioactive peptides, or by the differences in the amino acid pattern [28]. Furthermore, dietary fiber is known to cause a reduction in the intestinal absorption of cholesterol and/or bile acids, and it also decreases the triglyceride levels by inhibiting hepatic lipogenesis [29]. Some bioactive antioxidants, such as phenols, have cardio-protective actions by decreasing oxidized LDL particles [30], in addition to anti-inflammatory and anti-thrombotic effects [31].

Previous works coincide with our results regarding the cholesterol-lowering properties of amaranth. Czerwiński et al. [9] showed that amaranth flour positively influences the plasma lipid profile in rats fed with hypercholesterolemic diets, and they also proposed that this effect is related to the compounds with biological activity and the antioxidant activity in the whole seed. Escudero et al. [6] showed that the seed protein concentrate is efficient in reducing cholesterol and triglycerides levels in plasma of rats fed with hypercholesterolemic diets. Mendonça et al. [11] proposed that the bioactive peptides, present in the amaranth seed can affect the cholesterol absorption in the intestine or it can even have a direct effect on cholesterol synthesis and/or LDL uptake in liver, resulting in a cholesterol decrease in hamster's plasma. Shin et al. [32]

Table 1 Levels of lipids in serum

	CC	EC	CAh	EAh
Total-C (g/l)	67.67 \pm 2.58	67.60 \pm 2.30	54.8 \pm 3.11***	53.90 \pm 5.13 ^{oo}
HDL-C (g/l)	25.00 \pm 2.41	23.00 \pm 2.91	22.67 \pm 2.25	20.60 \pm 3.78
(LDL + VLDL)-C (g/l)	40.67 \pm 2.32	44.60 \pm 1.84	32.13 \pm 3.25***	33.30 \pm 2.90 ^{oo}
HDL-C/colesterol total	0.40	0.34	0.41	0.42

CC control casein; EC ethanol casein; CAh control Ah; Ah ethanol Ah; Total-C total cholesterol; HDL-C HDL-cholesterol; LDL + VLDL-C LDL + VLDL-cholesterol

Different symbols indicate statistical significance: *EAh* vs EC \rightarrow ^o : ^{oo} $p < 0.001$; *CAh* vs CC \rightarrow • : *** $p < 0.001$. Values are means \pm SD ($n = 6$)

Table 2 Levels of lipids in liver tissue

(μg/g tissue)	CC	EC	CAh	EAh
Total cholesterol	2755.98±269.36	3672.93±239.39*	2940.75±327.84	3170.51±368.46
Free cholesterol	1018.75±79.94	1328.80±77.54*	1132.02±57.59	1141.20±62.15°
Esterified cholesterol	1737.22±103.41	2344.13±82.09**	1808.73±112.19	2029.31±94.24°°
Triglycerides	655.23±68.66	876.10±75.34 **	728.27±70.32	774.93±63.71

CC control casein; EC ethanol casein; CAh control Ah; EAh ethanol Ah

Different symbols indicate statistical significance: EAh vs EC → ° : °p<0.05, °° p<0.01; EC vs CC → * : * p<0.05, ** p<0.01. Values are means ± SD (n=6)

published that the squalene found in amaranth can reduce cholesterol by increasing fecal elimination of steroids and interfering with cholesterol absorption in rats. Kulakoba et al. [33] showed that amaranth oil has hypocholesterolemic actions in rats. Zeashan et al. [34] revealed that ethanol extract of the whole amaranth plant, could yield a significant protection against galactosamine/lipopolysaccharide-induced hepatocellular injury, reverting the increased serum and hepatic lipids values. Despite the available information, there are no reports studying the protective effect of amaranth on alcohol intoxication.

For these reasons, we studied the 3-hydroxy-3-methylglutaryl coenzyme A reductase (HMG-CoAR) and the LDL receptor (LDLr), which play an important role in the biosynthesis and uptake of cholesterol in the liver, respectively. Both can be regulated by the sterol regulatory element binding protein 2 (SREBP-2). On the other hand, cholesterol 7-alpha-hydroxylase (CYP7A1) is the key enzyme for cholesterol elimination through bile acids. We also measured the expression of apolipoprotein B (Apo B) that is the main

protein constituent in the afferent way of cholesterol, being the most abundant protein in LDL, VLDL, IDL lipoproteins, and in chylomicrons.

Figure 1 shows the results of the expression of genes involved in cholesterol metabolism. The expression of HMG-CoAR was lower in the EAh group when compared to EC (p<0.001) and CAh (p<0.01), and that of LDLr was higher in EAh when compared to EC and CAh (p<0.001). HMG-CoAR mRNA levels increased in EC when compared to CC (p<0.001). There were no changes in the expression of SREBP-2, CYP7A1 or Apo B between the groups. From these results we infer that in the groups that received ethanol in a sub-chronic manner, amaranth had a positive effect due to the lower expression of HMG-CoAR, while the transcriptional activation of LDLr would be part of the regulatory mechanism in response to the cholesterol content, and it may also contribute to prevent the serum cholesterol increase due to ethanol.

Figure 2 shows the data of the expression of genes involved in fatty acids and triglycerides metabolism. The expression of the enzymes involved in the synthesis of triglycerides,

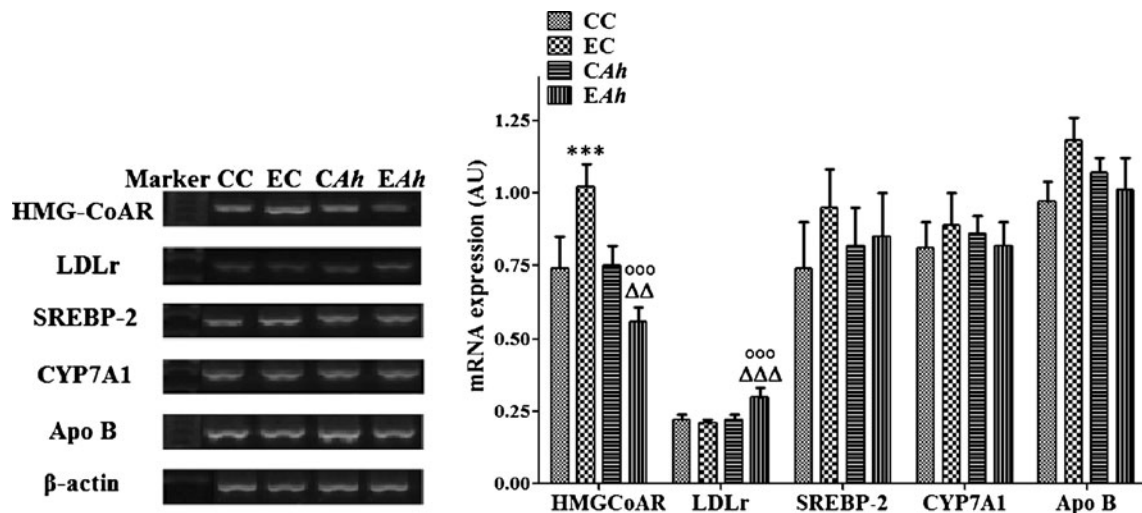


Fig. 1 RT-PCR analysis of the expression of cholesterol-related genes against β-actin as internal standard. Agarose gel images showing the PCR products and densitometric representation of the results. CC control casein; EC ethanol casein; CAh control Ah; EAh ethanol Ah; AU arbitrary units. HMG-CoAR 3-hydroxy-3-methylglutaryl coenzyme A

reductasa; LDLr LDL receptor; SREBP-2 sterol regulatory element binding protein 2; CYP7A1 cholesterol 7-alpha-hydroxylase; Apo B apolipoprotein B. Different symbols indicate statistical significance: EAh vs EC → ° : °°° p<0.001; EAh vs CAh → Δ : ΔΔ p<0.01, ΔΔΔ p<0.001; EC vs CC → * : *** p<0.001. Values are means ± SD (n=6)

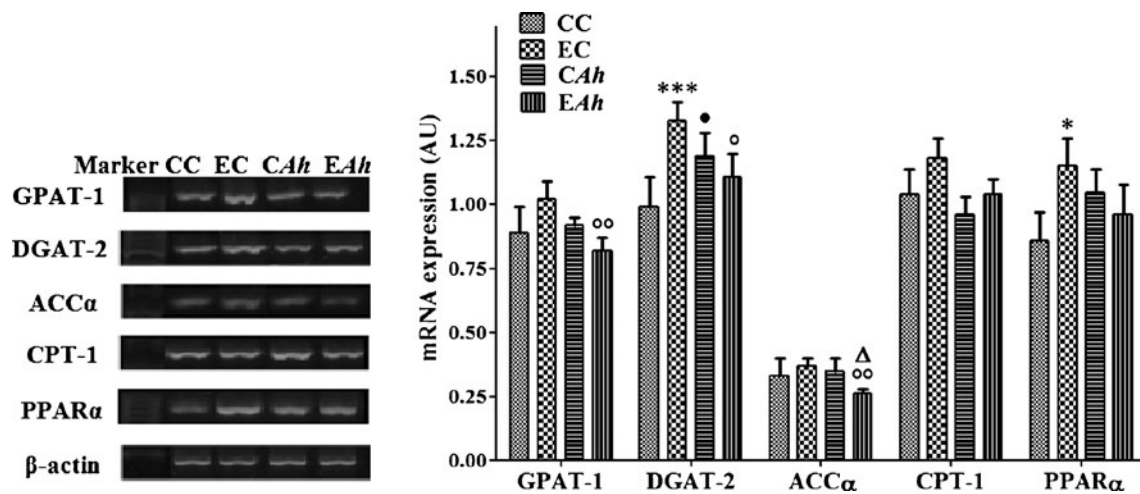


Fig. 2 RT-PCR analysis of the expression of cholesterol-related genes against β -actin as internal standard. Agarose gel images showing the PCR products and densitometric representation of the results. *CC* Control Casein; *EC* Ethanol Casein; *CAh* Control *Ah*; *EAh* Ethanol *Ah*; *AU* arbitrary units. *GPAT-1* glycerol-3-phosphate acyltransferase 1; *DGAT-2* diacylglycerol O-acyltransferase 2; *ACC α* acetyl-CoA carboxylase

alpha; *CPT-1* carnitine palmitoyl transferase 1; *PPAR α* peroxisome proliferator-activated receptor alpha. Different symbols indicate statistical significance: *EAh* vs *EC* \rightarrow° : $p < 0.05$, $^{\circ\circ}$ $p < 0.01$; *CAh* vs *CC* \rightarrow^{\bullet} : $p < 0.05$; *EAh* vs *CAh* \rightarrow^{Δ} : $p < 0.05$; *EC* vs *CC* \rightarrow^* : $p < 0.05$, *** $p < 0.001$. Values are means \pm SD ($n = 6$)

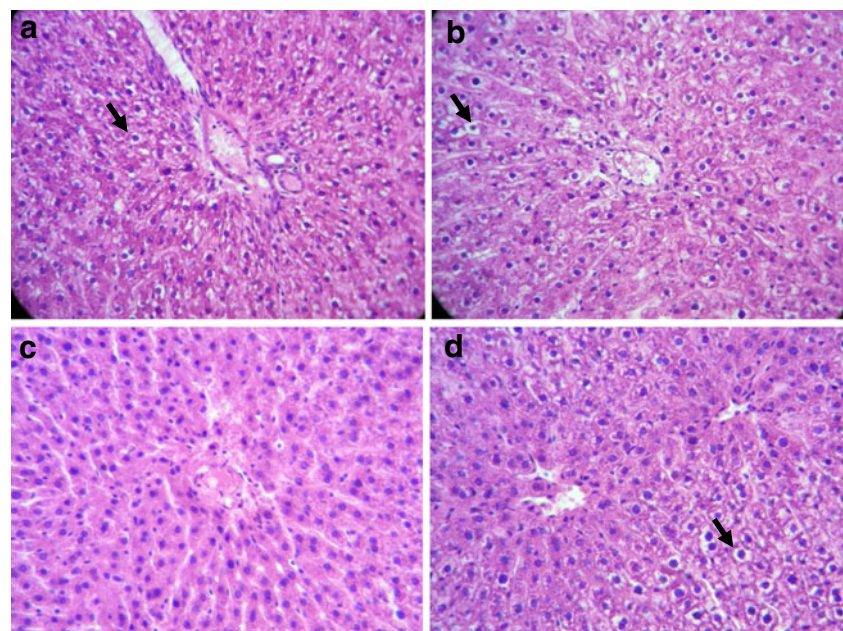
glycerol-3-phosphate acyltransferase 1 (*GPAT-1*) and diacylglycerol O-acyltransferase 2 (*DGAT2*) decreased in *EAh* vs. *EC* ($p < 0.01$) and ($p < 0.05$), respectively. Considering these two genes, it is clear that in rats that were exposed to ethanol, amaranth exerts a control effect on the triglycerides synthesis; this does not happen when the protein source is casein, since an increment in *DGAT-2* expression is observed when comparing *EC* vs. *CC* ($p < 0.001$). In addition, as previously expressed, in rats treated with ethanol amaranth tends to reduce the triglycerides liver deposit; this effect would be supported by the lower expression of their synthesis enzymes. For a better

understanding of these results, we determined the expression of some genes involved in the fatty acids metabolism.

We studied acetyl-CoA carboxylase alpha (*ACC α*), which is the regulatory enzyme in the fatty acids biosynthesis, and carnitine palmitoyl transferase 1 (*CPT-1*), which catalyzes the entrance of fatty acids into the mitochondria and it is an important enzyme in the β -oxidation, which can be regulated by peroxisome proliferator-activated receptor alpha (*PPAR α*).

Figure 2 also shows that the expression of *ACC α* decreased in *EAh* when compared to *EC* ($p < 0.01$) and *CAh* ($p < 0.05$). Animals fed with amaranth did not show

Fig. 3 Hematoxylin and eosine stained liver sections (100 \times). **a** *CC* (control casein): hepatocytes are displayed with fatty infiltrates (arrow). **b** *EC* (ethanol casein): fatty deposits are widely distributed occupying most of the cytoplasm. **c** *CAh* (control *Ah*): the hepatocytes show a normal histoarchitecture. **d** *EAh* (ethanol *Ah*): the fatty deposits are seen by sector



differences in the levels of CPT-1 or PPAR α mRNA; instead, a significant increase in PPAR α was observed in EC when compared to CC ($p < 0.05$).

It is known that ethanol can suppress the expression of PPAR α , attenuating fatty acid β -oxidation and leading to an accumulation of triglycerides in the liver [35]. Amaranth could reduce the lipogenesis inductor effect of ethanol through the decrease in the expression of ACC α and the triglycerides synthesis enzymes. In addition, there would be an increased expression of PPAR α in EC, when compared to CC, in response to the increased level of triglycerides in liver. This was not observed in the groups fed with amaranth, since there are no changes in the liver triglycerides content.

In general, it has been observed that increases or decreases in the expression of genes involved in the synthesis or degradation of fatty acids, cholesterol and triglycerides, where not affected by the transcription factors expression. This is due to the multiple ways of regulation, at the post-transcriptional and post-translational levels. It is important to note that in this model, the administration of ethanol is sub-chronical before the development of a more severe liver damage, so hyperlipidemia prevails and thus, we observe the first changes in liver gene expression.

Based on the results of lipids content in serum and liver, we thought it was necessary to determine if there is a simultaneous fat infiltration, since this is an important factor for the development of alcoholic hepatic disease. Therefore, we performed a hematoxylin and eosin stain.

Figure 3a and b show pictures from the CC and EC groups, respectively. Both groups exhibit fat deposits with structural modifications in the liver parenchyma. This effect is more pronounced in EC, where the fat infiltrates are widely distributed, occupying most of the cytoplasm, and have a spongy aspect. Figure 3c shows a picture of CAh, where we can visualize a normal liver parenchyma, with aligned hepatocytes and conserved white spaces corresponding to the sinusoids. Figure 3d shows a picture of EAh, where we can observe some lipid droplets localized by sectors. When comparing CAh vs. CC (Fig. 3c and a), and in lesser extent in EAh vs. EC (Fig. 3d and b), we can clearly see that amaranth decreased the fat deposit when compared to casein. These data is consistent with the biochemical and molecular results shown in this study.

Conclusion

Our findings suggest that in the presence of ethanol, the integral flour of *Amaranthus hypochondriacus* has a positive effect on lipids, which may be preventing the increase in serum cholesterol through a higher expression of the LDL receptor, as well as a decrease of liver cholesterol, due to changes in the expression of HMG-CoAR. On the other hand,

amaranth seems to attenuate the tendency of ethanol to stimulate the fat deposit in liver, by decreasing the expression of ACC α and enzymes involved in triglycerides synthesis. These properties could be mainly attributed to the bioactive components in the amaranth flour, such as the proteins, fatty acid profile, soluble dietary fiber, squalene and flavonoids content.

For all these reasons, besides being recognized as a nutritive source and an antioxidant resource, amaranth should also be recommended to prevent the alterations in the lipid metabolism induced by alcohol and other harmful agents. Figure S2 shows a schematic diagram illustrating possible effects of *Amaranthus hypochondriacus* flour on the lipid metabolism, in the liver of rats sub-chronically treated with ethanol.

Acknowledgments We are grateful to Dr. Nidia Gómez for her assistance in the morphological study.

Conflict of Interest The authors declare that they have no conflict of interest.

References

- Sillanaukee P, Koivula T, Jokela H, Pitkajarvi T, Seppa K (2000) Alcohol consumption and its relation to lipid based cardiovascular risk factors among middle-aged women: the role of HDL(3) cholesterol. *Atherosclerosis* 152:503–510
- Di Castelnuovo A, Costanzo S, Donati MB, Iacoviello L, de Gaetano G (2010) Prevention of cardiovascular risk by moderate alcohol consumption: epidemiologic evidence and plausible mechanisms. *Intern Emerg Med* 5(4):291–297
- Baraona E, Lieber CS (1998) Alcohol and lipids. *Recent Dev Alcohol* 14:97–134
- Schlienger JL, Pradignac A (2009) Nutrition approaches to prevent chronic disease. *Rev Prat* 59(1):61–65
- Marcone MF, Kakuda Y, Yada RY (2003) Amaranth as a rich dietary source of beta-sitosterol and other phyosterols. *Plant Foods Hum Nutr* 58(3):207–211
- Escudero NL, de Arellano ML, Luco JM, Giménez MS, Mucciarelli SI (2004) Comparison of the chemical composition and nutritional value of *Amaranthus cruentus* flour and its protein concentrate. *Plant Foods Hum Nutr* 59(1):15–21
- Rodas B, Bressani R (2009) The oil, fatty acid and squalene content of varieties of raw and processed amaranth grain. *Arch Latinoam Nutr* 59(1):82–87
- Plate AYA, Arêas JAG (2002) Cholesterol-lowering effect of extruded amaranth (*Amaranthus caudatus* L.) in hypercholesterolemic rabbits. *Food Chem* 76(1):1–6
- Czerwiński J, Bartnikowska E, Leontowicz H, Lange E, Leontowicz M, Katrich E, Trakhtenberg S, Gorinstein S (2004) Oat (*Avena sativa* L.) and amaranth (*Amaranthus hypochondriacus*) meals positively affect plasma lipid profile in rats fed cholesterol-containing diets. *J Nutr Biochem* 15(10):622–629
- Escudero NL, Zirulnik F, Gomez NN, Mucciarelli SI, Giménez MS (2006) Influence of a protein concentrate from *Amaranthus cruentus* seeds on lipid metabolism. *Exp Biol Med (Maywood)* 231(1):50–59
- Mendonça S, Saldiva PH, Cruz RJ, Arêas JAG (2009) Amaranth protein presents cholesterol-lowering effect. *Food Chem* 116(3):738–742

12. Fritz M, Vecchi B, Rinaldi G, Añón MC (2011) Amaranth seed protein hydrolysates have *in vivo* and *in vitro* antihypertensive activity. *Food Chem* 126(3):878–884
13. López VR, Razzeto GS, Giménez MS, Escudero NL (2011) Antioxidant properties of *Amaranthus hypochondriacus* seeds and their effect on the liver of alcohol-treated rats. *Plant Foods Hum Nutr* 66(2):157–162
14. Nsimba R, Kikuzaki H, Konishi Y (2008) Antioxidant activity of various extracts and fractions of *Chenopodium quinoa* and *Amaranthus* spp. seeds. *Food Chem* 106(2):760–766
15. Tironi VA, Añón MC (2010) Amaranth proteins as a source of antioxidant peptides: effect of proteolysis. *Food Res Int* 43(1):315–322
16. de la Barca AM, Rojas-Martínez ME, Islas-Rubio AR, Cabrera-Chávez F (2010) Gluten-free breads and cookies of raw and popped amaranth flours with attractive technological and nutritional qualities. *Plant Foods Hum Nutr* 65(3):241–246
17. Reeves PG, Nielsen FH, Feahy GC Jr (1993) AIN-93 purified diets for laboratory rodents: final report of the American Institute of Nutrition *ad hoc* writing committee on the reformulation of the AIN-76A rodent diet. *J Nutr* 123(11):1939–1951
18. Vengeliene V, Vollmayr B, Henn FA, Spanagel R (2005) Voluntary alcohol intake in two rat lines selectively bred for learned helpless and non-helpless behavior. *Psychopharmacology (Berl)* 178(2–3):125–132
19. Eblen-Zajjur A, Eblen-Zajjur M (2001) Estimation of low density lipoprotein cholesterol concentration: regression analysis versus Friedewald's formula. *Rev Med Chil* 129:1263–1270
20. Layne E (1957) Spectrophotometric and turbidimetric methods for proteins. *Methods Enzymol* 3:447–454
21. Hara A, Radin NS (1978) Lipid extraction of tissues with a low-toxicity solvent. *Anal Biochem* 90:420–426
22. Folch J, Lees M, Sloane-Stanley GH (1957) A simple method for the isolation and purification of total lipides from animal tissues. *J Biol Chem* 226:497–509
23. Zak B, Moss N, Boyle AJ, Zlatkis A (1954) Reactions of certain unsaturated steroids with acid iron reagent. *Anal Chem* 26:776–777
24. Abel LL, Levy BB, Brodie BB, Kendall FE (1952) A simplified method for the estimation of total cholesterol in serum and demonstration of its specificity. *J Biol Chem* 195:357–366
25. Sardesai VM, Manning JA (1968) Determination of triglycerides in plasma and tissues. *Clin Chem* 14(2):156–161
26. Snedecor GW, Cochran WG (1980) *Statistical methods*, 7th edn. Iowa State University Press
27. Feinman L, Lieber CS (1999) Ethanol and lipid metabolism. *Am J Clin Nutr* 70(5):791–792
28. Yoshikawa M, Takahashi M, Yang S (2003) Delta opioid peptides derived from plant proteins. *Curr Pharm Des* 9(16):1325–1330
29. Venkatesan N, Devaraj SN, Devaraj H (2003) Increased binding of LDL and VLDL to Apo B, E receptors of hepatic plasma membrane of rats treated with Fibernat. *Eur J Nutr* 42(5):262–271
30. Kaliora AC, Dedoussis GV, Schmidt H (2006) Dietary antioxidants in preventing atherogenesis. *Atherosclerosis* 187(1):1–17
31. Michalska M, Gluba A, Mikhailidis DP, Nowak P, Bielecka-Dabrowa A, Rysz J, Banach M (2010) The role of polyphenols in cardiovascular disease. *Med Sci Monit* 16(5):RA110–RA119
32. Shin DH, Heo HJ, Lee YJ, Kim HK (2004) Amaranth squalene reduces serum and liver lipid levels in rats fed a cholesterol diet. *Br J Biomed Sci* 61:11–14
33. Kulakova SN, Pozdniakov AL, Korf II, Karagodina ZV, Medvedev FA, Viktorova EV, Gonor KV, Kamysheva IM, Gadzhieva ZM (2006) Amaranth oil: peculiarities of chemical composition and influence on lipid metabolism by rats. *Vopr Pitan* 75:36–42
34. Zeashan H, Amresh G, Singh S, Rao CV (2010) Protective effect of *Amaranthus spinosus* against D-galactosamine/lipopolysaccharide-induced hepatic failure. *Pharm Biol* 48(10):1157–1163
35. Ringseis R, Muschick A, Eder K (2007) Dietary oxidized fat prevents ethanol-induced triacylglycerol accumulation and increases expression of PPAR alpha target genes in rat liver. *J Nutr* 137(1):77–83