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15 **New insight on vertebral anomalies in cultured Senegalese sole (*Solea senegalensis*,**  
16 **Kaup) at early stages of development**

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26 **Short running title:** Vertebral anomalies in cultured Senegalese sole

27 **Abstract**

28 Senegalese sole (*Solea senegalensis*, Kaup) is a promising flatfish species in aquaculture.  
29 However, skeletal anomalies are still a great concern in sole farming. Investigation of  
30 this issue is crucial to improving larval quality and optimizing production. The aim of this  
31 study was to thoroughly assess anomalies in the rachis of reared sole at early  
32 developmental stages. Sole (n = 507) were sampled at 31 or 32 days after hatching (dah).  
33 The specimens were stained with alcian blue and alizarin red, and evaluated for the  
34 detection of vertebral deformities. Most fish presented 9:34:3 vertebrae in abdominal,  
35 caudal, and caudal complex regions, respectively. Remarkably, all specimens showed at  
36 least one spinal anomaly. Alterations of neural/haemal elements, as well as deformities  
37 of hypurals, parhypural, and epural, were recurrent. Vertebral body anomalies (VBA)  
38 and/or vertebral column deviations (VCD) were identified in 52% of the individuals.  
39 Vertebral deformations and fusions were common, especially in caudal complex.  
40 "Minor" anomalies were predominant and some of the detected disorders might be a  
41 result of non/low-pathological processes. These results contribute a new insight into the  
42 main skeletal anomalies affecting cultured sole larvae. Further research is required to  
43 determine their impact on fish welfare and external appearances at commercial stages.

44 **Keywords:** Skeletal anomalies; Senegalese sole (*Solea senegalensis*); larval quality;  
45 vertebrae; skeletogenesis

46 **Introduction**

47 Senegalese sole (*Solea senegalensis*, Kaup) is a flatfish upon which Iberian aquaculture  
48 has placed great expectations (Howell, Conceição, Prickett, Cañavate & Mañanos 2009;  
49 Imsland, Foss, Conceição, Dinis, Delbare, Schram, Kamstra, Rema & White 2003;  
50 APROMAR 2015). Currently, fish farms are able to complete the production cycle and  
51 commercialize sole. However, its intensive culture is limited by the incidence of skeletal  
52 anomalies (Fernández, Pimentel, Ortiz-Delgado, Hontoria, Sarasquete, Estévez,  
53 Zambonino-Infante & Gisbert 2009; Gavaia, Dinis & Cancela 2002; Boglino, Darias, Ortiz-  
54 Delgado, Özcan, Estevez, Andree, Hontoria, Sarasquete & Gisbert 2012b).  
55 Approximately, 40% of juveniles present vertebral deformities in some Senegalese sole  
56 hatcheries, at the grading point for transfer to ongrowing farms (Riaza, personal  
57 communication). Similarly, in a number of scientific investigations, skeletal  
58 malformations have ranged from 44% (Gavaia *et al.* 2002) to around 80% (100% in some  
59 cases) in Senegalese sole larvae (Engrola, Conceição, Gavaia, Cancela & Dinis 2005;  
60 Gavaia, Domingues, Engrola, Drake, Sarasquete, Dinis & Cancela 2009; Fernández *et al.*  
61 2009).

62 In other fish species, the incidence of skeletal anomalies can be also elevated, as in  
63 gilthead sea bream (*Sparus aurata*, L.), with frequencies of 100% of fish affected in some  
64 cases (Boglione, Gagliardi, Scardi & Cataudella 2001; Fernández, Hontoria, Ortiz-  
65 Delgado, Kotzamanis, Estévez, Zambonino-Infante & Gisbert 2008). In European sea bass  
66 larvae (*Dicentrarchus labrax*, L.), from 10% to 30% of the specimens showed vertebral  
67 column deformities (Mazurais, Darias, Gouillou-Coustans, Le Gall, Huelvan,  
68 Desbruyeres, Quazuguel, Cahu & Zambonino-Infante 2008), whereas a range of 4-9%

69 vertebral anomalies was seen in Atlantic salmon juveniles (*Salmo salar*, L.) (Fjelldal,  
70 Hansen & Albrektsen 2012b). Moreover, studies on other flatfish species have reported  
71 skeletal abnormalities, with halibut (*Hippoglossus hippoglossus*, L.) at an incidence of  
72 41% to 89% of specimens showing at least one malformation (Lewis, Lall & Witten 2004);  
73 Japanese flounder (*Paralichthys olivaceus*, Temminck & Schlegel), with a range of 41-  
74 53% (Haga, Takeuchi, Murayama, Ohta & Fukunaga 2004), and turbot (*Scophthalmus*  
75 *maximus*, L.), where 51% of individual fish were deformed (Tong, Liu, Xu, Ma, Xiao, Xiao  
76 & Li 2012).

77 Skeletal deformities seriously hinder farming of many species since they affect growth,  
78 food efficiency level, susceptibility to diseases, and welfare of affected individuals  
79 (Boglione, Gavaia, Koumoundouros, Gisbert, Moren, Fontagné & Witten 2013a; Fjelldal,  
80 Hansen, Breck, Ørnsrud, Lock, Waagbø, Wargelius & Witten 2012a), leading to economic  
81 repercussions (Koumoundouros, Oran, Divanach, Stefanakis & Kentouri 1997b). Due to  
82 the significance of this problem, skeletal anomalies have been studied in numerous  
83 aquaculture species (for review, see Boglione *et al.* 2013a; Boglione, Gisbert, Gavaia,  
84 Witten, Moren, Fontagné & Koumoundouros 2013b). In some species, the development  
85 of skeletal malformations was associated with nutritional (Cahu, Zambonino Infante &  
86 Takeuchi 2003; Fernández *et al.* 2008; Fernández *et al.* 2009; Darias, Mazurais,  
87 Koumoundouros, Glynatsi, Christodouloupoulou, Huelvan, Desbruyeres, Le Gall,  
88 Quazuguel, Cahu & Zambonino-Infante 2010b), environmental, and genetic factors (Lall  
89 & Lewis-McCrea 2007; Mazurais, Glynatsi, Darias, Christodouloupoulou, Cahu,  
90 Zambonino-Infante & Koumoundouros 2009; Georgakopoulou, Katharios, Divanach &  
91 Koumoundouros 2010; Gjerde, Pante & Baeverfjord 2005), although parasites and  
92 bacteria could also cause these anomalies (Roberts & Rodger 2001; Madsen, Arnbjerg &

93 Dalsgaard 2001). In particular, intensive rearing protocols could be characterized by high  
94 incidences of skeletal abnormalities (Russo, Prestinicola, Scardi, Palamara, Cataudella &  
95 Boglione 2010). Therefore, research on production conditions at the first stages of  
96 development is important in order to improve larval quality and hone Senegalese sole  
97 farming practices. Former descriptions of skeletal deformities in this species were  
98 conducted mainly in research facilities (Fernández *et al.* 2009; Gavaia *et al.* 2009; Gavaia  
99 *et al.* 2002). Still, some gaps persist on the main typologies of vertebral anomalies  
100 affecting sole reared in fish farms, where multilevel factors may interact on fish in  
101 unknown manners, which experimental conditions are unable to mimic (Boglione *et al.*  
102 2013b). Moreover, from a commercial point of view, the establishment of bench marks  
103 to compare levels of deformity is highly required (Howell *et al.* 2009).

104 The aim of this study was to provide a thorough description and quantification of the  
105 most common anomalies in the vertebral column of reared Senegalese sole at early  
106 stages of development. This work also sought to provide simple definitions of some  
107 criteria to meet the demand for a uniform categorization of anomaly typologies  
108 (Boglione *et al.* 2013b) affecting Senegalese sole.

## 109 **Materials and methods**

110 Senegalese sole, reared in a fish farm in Northwest Spain, were randomly sampled (n =  
111 507). Fish were taken at 31 or 32 days after hatching (dah), humanely euthanized with  
112 an overdose of Tricaine methanesulfonate (MS-222, Sigma), and fixed in 10% buffered  
113 formalin. The specimens were stained for cartilage and bone, using a modified double  
114 staining technique, with alcian blue and alizarin red (Gavaia, Sarasquete & Cancela 2000;  
115 Darias, Lan Chow Wing, Cahu, Zambonino-Infante & Mazurais 2010a).

116 After staining, fish were measured, taking into account the standard length (StL; cm)  
117 from the rostral end of the skull to the end of hypurals; the standard height (StH; cm),  
118 which comprised the highest distance between the ventral and dorsal aspect of the fish,  
119 without taking into account fin rays. The StL/StH ratio was then calculated.

120 All specimens were observed from both sides using a binocular model (Olympus®) SZX16  
121 to count the number of vertebrae and caudal fin rays, and to evaluate anomalies. The  
122 vertebral column was divided into three anatomical regions: abdominal, caudal, and  
123 caudal complex, according to Gavaia *et al.* (2002) (Figure 1). Each vertebra was assessed  
124 by considering anomalies in the neural arch and spine (dorsal aspect of the vertebra), in  
125 the parapophysis/haemal arch and spine (ventral aspect of the vertebra), and in the  
126 vertebral centrum. The terms for anatomical regions and anomalies used in this study  
127 were adapted from previous descriptions (Boglione *et al.* 2001; Gavaia *et al.* 2002; Lewis  
128 *et al.* 2004; Boglione *et al.* 2013a; Boglione *et al.* 2013b). Alterations in the arches/spines  
129 and parapophysis were categorized as bifurcation, number alteration, insertion  
130 alteration, fusion, incomplete arch, and deformation (alteration of the shape). Vertebral  
131 body anomalies (VBA) were considered as fusion or deformation, and three types of axis  
132 deviations (VCD) were assessed: kyphosis, lordosis, and scoliosis. Kyphosis and lordosis  
133 corresponded to the curvature of the axis towards, respectively, the dorsal and ventral  
134 aspects of the fish. Scoliosis was the bending of the rachis in the ocular/blind side of the  
135 body. Disorders in the caudal complex plates (parhypural, epural and hypurals) were  
136 also evaluated (bifurcation, number alteration, insertion alteration, fusion, and  
137 deformation).

138 During the development of this study, some considerations were made regarding types  
139 of anomalies:

140 1) Alterations in the number of parapophysis comprised the presence of one, or more  
141 than two parapophysis in each vertebral body, or the absence of these elements in a  
142 vertebra located between two vertebrae exhibiting parapophysis. 2) The presence of  
143 two complete and parallel neural/haemal spines (without forming an arch) was  
144 regarded as an alteration of the number of spines and an incomplete arch. 3) Partial  
145 fusions were considered as fusions. 4) Evident misalignments/subluxations of a vertebra  
146 were also considered as deviations. When a deviation towards more than one plane was  
147 present, the lesion was classified according to the deviation exhibiting the most obvious  
148 curvature. 5) When vertebral fusions or axis deviations affected more than one region  
149 in the same anomaly, they were registered in the anatomical area with the highest  
150 number of implicated vertebrae. 6) Hypural, epural, and parhypural fusions were  
151 regarded as anomalies, as were, also, the fusions between these elements and modified  
152 neural/haemal arches of the caudal complex.

## 153 **Results**

### 154 1. Measurements and number of vertebrae

155 Descriptive statistics on StL, StH, and the ratio StL/StH are displayed in Table 1 (mean,  
156 standard deviation (SD), mode, median, minimum and maximum values). The number  
157 of vertebral centra for each anatomical region and the number of caudal fin rays are  
158 shown in Table 2 (mean, SD, mode and percentage of individuals that showed this rate  
159 (%)), as well as minimum and maximum). The majority of the individuals presented 46  
160 vertebral centra distributed in 9:34:3 vertebrae in the abdominal region, caudal area,



161 and caudal complex, respectively. Just under 5% of specimens presented 4 vertebrae in  
162 the caudal complex, including the urostyle.

## 163 2. Skeletal anomalies

164 A summary of the results regarding skeletal anomalies is displayed in Table 3. All  
165 specimens presented at least one skeletal alteration, and around 52% had at least one  
166 anomaly in the vertebral centra or one rachis deviation. The number of observed  
167 skeletal anomalies reached 5,654 alterations, and the 16.1% of them corresponded to  
168 VBA and VCD.

169 The most frequent alterations of the vertebral column are illustrated in Figure 2 (A-E)  
170 and Figure 3 (A-E). Table 4 shows the number of deformed individuals (N.I.), the  
171 percentage they represent (%N.I.), the number of anomalies (N.A.), and the average  
172 number of anomalies per affected specimen (N.A./I.), for each anatomical region and  
173 malformation type.

174 In this investigation, the first abdominal vertebra presented two short neural spines; all  
175 individuals were lacking the neural arch (Figure 2 A). These data are not included in Table  
176 4. A high percentage of fish (83.2%) had alterations of the shape (deformations) of  
177 neural and/or haemal arches and spines in other vertebrae, with multiple deformations  
178 in the same specimen (Table 4) (Figure 2 A and B, and Figure 3 A). One typical  
179 modification in the morphology of these elements was the twisted tip of the haemal  
180 spines, especially in the front caudal vertebrae (Figure 2 B and Figure 3 A). Incomplete  
181 arches and alterations of the insertion of the spines were also common in caudal region  
182 (Figure 2 C and D and Figure 3 B), and were sometimes associated with mild  
183 abnormalities of the centra (Figure 2 D, and Figure 3 A and B). In the caudal complex,

184 neural and haemal arches and spines were also affected by deformations (Figure 3 C).  
185 Fusions between these structures were primarily related to fusions among vertebral  
186 bodies (Figure 3 D).

187 The main anomalies in the caudal complex consisted in minimal changes in the shape of  
188 hypurals, epural, and parhypural and/or fusions between these elements (Table 4). In  
189 particular, the epural, the hypural 5, and the parhypural were often slightly bent (Figure  
190 2 E and Figure 3 A, C and D). Fusions between hypurals were frequent, involving hypurals  
191 3 and 4 (Figure 2 E and Figure 3 C) in 84% of the affected individuals.

192 A number of alterations were observed throughout the vertebral column, especially  
193 affecting the parapophysis (Table 4). In some cases, these were associated with other  
194 alterations in the vertebra, whereas, in other cases they appeared in otherwise normal  
195 abdominal vertebrae as thin single, bony spicules (Figure 2 A). Other types of anomalies  
196 in the parapophysis were scarce. In addition, all specimens presented a variable number  
197 of abdominal vertebrae with two fused parapophysis, forming an arch (Figure 2 A). This  
198 datum is not reflected in Tables 3 and 4.

199 Regarding VBA and VCD, there was a high incidence of affected individuals (52%), as well  
200 as an elevated number of these anomalies. Approximately 70% of affected fish showed  
201 up to three alterations in these elements (with an average of 3.5 deformities per  
202 specimen), although ten fish presented more than ten anomalies (Table 3, Figure 3 A,  
203 and Figure 4). In general, the caudal region and the caudal complex showed a higher  
204 number of affected individuals and an increased number of VBA and/or VCD (Table 4).  
205 Figure 5 (A-B) and Figure 6 (A-C) show the number of sole affected by VBA and VCD along  
206 the vertebral axis, respectively. For every type of VBA, the critical location was preurals,

207 followed by the first two abdominal vertebrae (affected mainly by fusions) and the  
208 intermediary area between abdominal and caudal regions, in decreasing order of  
209 occurrence (Figure 5 A and B). Fusions and deformations were more frequent than VCD  
210 in every anatomical region (Table 4). In the abdominal region, 7.3% of the specimens  
211 showed partial fusions involving the first and the second centra. The two vertebral  
212 bodies were commonly bound in the dorsal or ventral portion of the endplates due to  
213 rotation or deformation of the centra. Fusions among vertebral bodies were  
214 distinguished in a certain frequency along the column (Figure 3 A-D and Figure 5 A) and  
215 involved a different number of vertebrae, with an average of 2.3. In some cases, fused  
216 vertebrae were almost the same length and shape as those around them, being almost  
217 totally reshaped into a single vertebra, although the line of fusion remained (Figure 3 B  
218 and C). In severe cases (more common in the caudal region), several vertebrae were  
219 deformed and fused (Figure 3 A). In the caudal complex, fusions comprising preurals 1  
220 and 2 were the most common (Figure 3 C and Figure 5 A). Curiously, in the same area,  
221 certain individuals showed four vertebrae, in which the intermediary (extra) vertebra  
222 (I.V.) was located between the two preurals and fused with preural 1 (Figure 3 D and  
223 Figure 5 A) in different degrees of severity.

224 Modifications of the shape (deformations) of vertebral bodies were frequently  
225 associated with fusions and/or axis deviations (Figure 2 E and Figure 3 A, D and E).  
226 Therefore, the distribution of deformations along the rachis was especially similar to the  
227 fusion's main location (Figure 5 B). Vertebral shortening (compressions) and lack of  
228 symmetry of the vertebral structure (Figure 2 D) were predominant. These modifications  
229 in the shape of the centra were pronounced in vertebrae with a "k" figure or a  
230 trapezoidal or triangular form (Figure 3 D). Some fish presented more severe

231 deformations, as incomplete vertebrae (vertebrae lacking a part of the centra) (Figure 2  
232 E) or vertebral bodies were aberrantly enlarged, some exhibiting also hyperostosis on  
233 the surface (Figure 3 E).

234 Axis deviations were a less frequent alteration of the vertebral column. The studied  
235 alterations of the rachis (kyphosis, lordosis, and scoliosis) displayed different bending  
236 angles and in certain cases, distinct types of curvatures were overlapped in the same  
237 vertebral segment (Figure 3 A and E). The caudal complex region was more affected by  
238 kyphosis (Figure 3 D and Figure 6 A). At the same time, lordosis was the most common  
239 deviation in the abdominal and caudal regions (Figure 3 E and Figure 6 B). In this  
240 investigation, there were few individuals (4%) showing a curved spine to the  
241 ocular/blind side of the body (scoliosis) (Figure 6 C). Moreover, axis deviations usually  
242 comprised diverse vertebrae (mean values are 3.0, 3.2, and 4.3 vertebrae for kyphosis,  
243 lordosis, and scoliosis, respectively) although four specimens presented a dorsal/ventral  
244 subluxated vertebra.

## 245 **Discussion**

246 In this study, skeletal anomalies were thoroughly assessed in reared Senegalese sole.  
247 The present work complements former descriptions of vertebral disorders (Fernández  
248 *et al.* 2009, Gavaia *et al.* 2002) and provides a new insight into the deformities profile in  
249 reared larvae of this species. A double staining technique for cartilage and bone was  
250 convenient for the accurate identification of skeletal features at early stages of  
251 development. Moreover, it can also be used for the detection of vertebral deformities  
252 and in ontogenetic studies (Gavaia *et al.* 2002; Darias *et al.* 2010a). The observation of  
253 both sides of the specimens was useful to confirm alterations that occur on only one

254 aspect of the fish, and it had special application in distinguishing the direction of axis  
255 deviations. In addition, since a standardized classification of skeletal anomalies is still  
256 lacking in the literature for some fish species (Boglione *et al.* 2013b), the terminology  
257 used in the present study was adapted from several reports to unify some criteria on  
258 anomaly categories affecting Senegalese sole. The comprehensive anomaly typologies  
259 evaluated in this work can serve as a basis for comparison for forthcoming research and  
260 industrial monitoring purposes.

261 In general, a very high frequency of anomalies was observed, given that all individuals  
262 presented at least one lesion. The most often repeated deformities concerned mainly  
263 the neural/haemal arches and spines and caudal complex elements. Curiously, the first  
264 abdominal vertebra presented two separated neural spines in all specimens. To our  
265 knowledge, this is the first description of such an abnormality, which is characterized by  
266 its specific setting in the neural aspect of the first abdominal vertebra and its very high  
267 incidence. A similar absence of fusion in the arch, affecting different locations, was  
268 reported in the present work, which is consistent with other studies in Senegalese sole  
269 (Boglino *et al.* 2012b). During arch development, two latero-dorsal intramembranous  
270 buds elongate and join together to form the arch (Gavaia *et al.* 2002). Sole affected by  
271 the former alteration probably present an impairment of this junction, although factors  
272 such as genetics should not be disregarded. Hence, it would be interesting to investigate  
273 the onset of this specific type of anomaly to deepen our understanding of its aetiology.  
274 Other types of skeletal disorders have contributed to the high incidence observed in this  
275 study, especially caudal haemal spine deformations, as well as anomalies in the caudal  
276 complex plates (deformations and fusions). Despite the scarcity of literature on the  
277 causes of such abnormalities, some nutritional factors have been reported, as they could

278 influence appearances, namely, high vitamin A content (Fernández *et al.* 2009) and low  
279 levels of phosphorous in the diet (Lewis-McCrea & Lall 2010; Fontagné, Silva, Bazin,  
280 Ramos, Aguirre, Surget, Abrantes, Kaushik & Power 2009). Fusions between hypurals  
281 were also frequent, which is in agreement with other studies on reared Senegalese sole  
282 (Gavaia *et al.* 2002; Gavaia *et al.* 2009), Japanese flounder (Hosoya & Kawamura 1998),  
283 and hatchery-reared and wild juveniles of gilthead sea bream (Boglione *et al.* 2001).  
284 Nevertheless, some authors have considered this particular alteration as a common and  
285 normal event in caudal complex development and ossification, rather than a skeletal  
286 deformity (Boglino, Darias, Estévez, Andree & Gisbert 2012a).

287 Another typical feature observed in this work was the presence of a variable number of  
288 vertebrae displaying fused parapophysis, forming an arch. These structures could be  
289 consistent with haemapophyses, occurring in several Pleuronectides genera (Sakamoto  
290 1984). It remains uncertain if fused parapophysis should be regarded as a disorder or a  
291 simple tendency of these skeletal elements.

292 The high incidence of anomalies observed in the present work is consistent with other  
293 studies that have reported broad ranges in the incidence of anomalies in reared sole,  
294 from 44% up to 100% in some cases (Gavaia *et al.* 2002; Gavaia *et al.* 2009; Fernández  
295 *et al.* 2009). In contrast with reared fish, wild Senegalese sole showed a lower incidence  
296 of skeletal anomalies (Gavaia *et al.* 2009), and these differences could indicate the  
297 presence of a selective mortality of the affected wild specimens and/or an effect of  
298 culture conditions on the development of skeletal malformations (Gavaia *et al.* 2009).  
299 In this regard, studies in Senegalese sole have reported an influence of environmental  
300 conditions like temperature, light, hypercapnia (Dionísio, Campos, Valente, Conceição,

301 Cancela & Gavaia 2012; Blanco-Vives, Villamizar, Ramos, Bayarri, Chereguini & Sánchez-  
302 Vázquez 2010; Pimentel, Faleiro, Dionísio, Repolho, Pousão-Ferreira, Machado & Rosa  
303 2014), and nutritional factors, such as the vitamin A dietary content (Fernández *et al.*  
304 2009). Two hypotheses were suggested by Fernández *et al.* (2009) in order to explain  
305 the high incidence of deformities in reared Senegalese sole: this species is more prone  
306 to the development of malformations than other species; and these anomalies are non-  
307 lethal, so deformed animals can survive and be detected. In this sense, as previously  
308 mentioned, the majority of the observed lesions concerned neural/haemal spines and  
309 caudal complex elements. This is in line with other studies in reared Senegalese sole  
310 (Gavaia *et al.* 2009) and could constitute a baseline level of deformities, intrinsic to the  
311 rearing process at research and production centres. Moreover, from an industrial  
312 approach, the slight defect of spines and caudal complex structures could eventually be  
313 considered as "minor" anomalies. However, the chondral bones as caudal complex  
314 plates present a higher sensitivity to high vitamin A and to the amount of ascorbic acid  
315 in the diet (Fernández & Gisbert 2010; Darias, Mazurais, Koumoundouros, Le Gall,  
316 Huelvan, Desbruyeres, Quazuguel, Cahu & Zambonino-Infante 2011). Therefore,  
317 alterations in their skeletogenesis may provide clues towards factors affecting other  
318 skeletal elements. Also, arches and spines have an important role in protecting vital  
319 organs, which should not be ignored (Boglione *et al.* 2013b; Stiasny 2000). In addition,  
320 neural and haemal process length may influence the fish shape (Harder 1975), and  
321 serious bone alterations in several of these elements could harm fish performance  
322 (Boglione *et al.* 2013b). On the other hand, resorption and remodelling are crucial  
323 processes for bone development, growth, and mechanical adaptation of the fish (Witten  
324 & Huysseune 2009). Likewise, as reported by Witten, Obach, Huysseune & Baeverfjord

325 (2006), some vertebral fusions seemed to start bone re/modelling, in which the  
326 outcome was a structure with almost the same shape and size of a single non-deformed  
327 vertebra. Hence, extrapolating this reshaping process to the mildly bent elements or  
328 fused caudal plates, it could be hypothesised that they could acquire a normal final  
329 shape as they develop. Since this study is limited to the larval period, further research is  
330 required to investigate and monitor the evolution of these anomalies and their  
331 repercussions on fish welfare and external appearances, up to commercial stages.

332 The results also highlighted a high incidence of sole showing VBA and/or VCD that was  
333 far greater than the 20% indicated as a good value at the end of the hatchery stage by  
334 Boglione *et al.* (2013a). VBA and VCD could be considered the most deleterious  
335 anomalies in intensive production, because some severe lesions in centra and the rachis  
336 may have repercussions on the external morphology of the fish and their welfare  
337 (Boglione *et al.* 2001; Fjellidal *et al.* 2012a; Cardeira, Mendes, Pousão-Ferreira, Cancela  
338 & Gavaia 2015), and lead to subsequent rejection by the consumer (Gavaia *et al.* 2002;  
339 Boglione *et al.* 2001; Ambrosio, Costa, Sanchez & Flos 2008). This supposes an added  
340 expense due to manual selection of malformed individuals (Koumoundouros *et al.*  
341 1997b; Gavaia *et al.* 2009). In Senegalese sole farms, sorting is usually performed by  
342 observing and palpating each fish (Rodríguez & Peleteiro 2014). Curiously, around 6% of  
343 sole categorized as “normal” by this method present severe skeletal deformities  
344 (Losada, de Azevedo, Barreiro, Barreiro, Ferreiro, Riaza, Quiroga & Vázquez 2014). This  
345 can lead to a decreased market value of a suboptimum product (Fernández *et al.* 2008;  
346 Koumoundouros, Gagliardi, Divanach, Boglione, Cataudella & Kentouri 1997a;  
347 Koumoundouros 2010), which, combined with other known issues, can cause serious  
348 economic losses (Fernández *et al.* 2008).



349 VBA and VCD appeared mainly in the caudal region and caudal complex area, which is in  
350 agreement with other studies in Senegalese sole (Gavaia *et al.* 2009; Gavaia *et al.* 2002).  
351 The same types of anomalies described here have also been reported in other fish  
352 species at different stages of development (Fernández *et al.* 2008; Dionísio *et al.* 2012;  
353 Fjelldal, Hansen & Berg 2007). Furthermore, Witten, Gil-Martens, Huysseune, Takle, &  
354 Hjelde (2009) have extensively characterized the main vertebral body deformities in  
355 Atlantic salmon by radiographic analysis. In the present study, the vertebrae most  
356 affected by VBA and/or VCD were the preurals, the first two abdominal vertebrae, and  
357 the centra in the intermediary area between the abdominal and caudal regions. Preural  
358 fusions were very common, as in other studies, in wild and reared Senegalese sole  
359 (Gavaia *et al.* 2009; Dionísio *et al.* 2012). In zebrafish (*Danio rerio*, Hamilton), the two  
360 vertebrae adjacent to the urostyle were often fused and could be more susceptible to  
361 this anomaly (Bensimon-Brito, Cancela, Huysseune, & Witten 2010). In the abdominal  
362 region, the most frequent alteration consisted in fusions between the first pair of  
363 vertebrae. It is known that fusions of one or two vertebral bodies to the occipital region  
364 of the skull could be considered as frequent non-pathological phenomena in  
365 osteichthyans (Witten *et al.* 2009; Boglione *et al.* 2013b). Still, different grades of fusion  
366 and deformity of the vertebral bodies were also distinguished along the rachis. In various  
367 situations, deformations were associated with other types of anomalies, such as fusions  
368 and VCD. Similarly, other studies reported some deformations concomitant with fusions  
369 or deviations affecting the same vertebral centra (Fjelldal *et al.* 2007; Cardeira *et al.*  
370 2015).

371 Few sole showed axis deviations and some presented several types of curvatures in the  
372 same lesion, presenting a 3-D configuration. Kyphosis, lordosis, and scoliosis were

373 observed in a diversity of fish species, including flatfish (Nagano, Hozawa, Fujiki,  
374 Yamada, Miyaki, Sakakura & Hagiwara 2007; Cardeira, Bensimon-Brito, Pousão-Ferreira,  
375 Cancela & Gavaia 2012; Lewis *et al.* 2004; Boglione *et al.* 2001). Lordosis was mainly  
376 located in the intermediary vertebral segment between the abdominal and caudal  
377 regions, as pointed out by Sfakianakis, Georgakopoulou, Papadakis, Divanach, Kentouri  
378 & Koumoundouros (2006) in European sea bass. This deviation was related to  
379 temperature and hydrodynamics due to increased muscular activity in the caudal fin  
380 and/or the action of the muscles on the vertebral axis (Kihara, Ogata, Kawano, Kubota  
381 & Yamaguchi 2002; Kranenbarg, Waarsing, Muller, Weinans & van Leeuwen 2005;  
382 Sfakianakis *et al.* 2006). Further research would be required in order to determine the  
383 effects of nutritional and environmental causes of vertebral centre deformities and  
384 deviations in Senegalese sole larvae.

385 In conclusion, these data complement former studies, and provide new insights into the  
386 main skeletal anomalies that affect early stages of farmed Senegalese sole. This  
387 thorough evaluation constitutes a baseline and serves as a tool with which to compare  
388 future investigations. The results highlighted a high incidence of skeletal anomalies,  
389 affecting mainly the neural/haemal arches and spines and caudal complex elements. On  
390 the other hand, a considerable number of specimens presented VBA and VCD, especially  
391 deformations and fusions in caudal complex region. However, some of the detected  
392 disorders might be a result of non/low-pathological significance processes. Further  
393 research is required to determine the impact of such anomalies on the physiology and  
394 external appearance of the adult fish, as well as on innovative rearing protocols in order  
395 to optimize larval quality.

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403 **References**

- 404 Ambrosio P.P., Costa, C., Sanchez, P. & Flos, R. (2008) Stocking density and its influence  
405 on shape of Senegalese sole adults. *Aquaculture International* **16**, 333-343.
- 406 APROMAR (2015) *La acuicultura en España 2015*.
- 407 Bensimon-Brito A., Cancela, M.L., Huysseune, A. & Witten, P.E. (2010) The zebrafish  
408 (*Danio rerio*) caudal complex - a model to study vertebral body fusion. *Journal of*  
409 *Applied Ichthyology* **26**, 235-238.
- 410 Blanco-Vives B., Villamizar, N., Ramos, J., Bayarri, M.J., Chereguini, O. & Sánchez-  
411 Vázquez, F.J. (2010) Effect of daily thermo- and photo-cycles of different light  
412 spectrum on the development of Senegal sole (*Solea senegalensis*) larvae.  
413 *Aquaculture* **306**, 137-145.
- 414 Boglino A., Darias, M.J., Estévez, A., Andree, K.B. & Gisbert, E. (2012a) The effect of  
415 dietary arachidonic acid during the *Artemia* feeding period on larval growth and

416 skeletogenesis in Senegalese sole, *Solea senegalensis*. *Journal of Applied*  
417 *Ichthyology* **28**, 411-418.

418 Boglino A., Darias, M.J., Ortiz-Delgado, J.B., Özcan, F., Estevez, A., Andree, K.B.,  
419 Hontoria, F., Sarasquete, C. & Gisbert, E. (2012b) Commercial products for *Artemia*  
420 enrichment affect growth performance, digestive system maturation, ossification  
421 and incidence of skeletal deformities in Senegalese sole (*Solea senegalensis*)  
422 larvae. *Aquaculture* **324**, 290-302.

423 Boglione C., Gagliardi, F., Scardi, M. & Cataudella, S. (2001) Skeletal descriptors and  
424 quality assessment in larvae and post-larvae of wild-caught and hatchery-reared  
425 gilthead sea bream (*Sparus aurata* L. 1758). *Aquaculture* **192**, 1-22.

426 Boglione C., Gavaia, P., Koumoundouros, G., Gisbert, E., Moren, M., Fontagné, S. &  
427 Witten, P.E. (2013a) Skeletal anomalies in reared European fish larvae and  
428 juveniles. Part 1: normal and anomalous skeletogenic processes. *Reviews in*  
429 *Aquaculture* **5**, S99-S120.

430 Boglione C., Gisbert, E., Gavaia, P., Witten, P.E., Moren, M., Fontagné, S. &  
431 Koumoundouros, G. (2013b) Skeletal anomalies in reared European fish larvae and  
432 juveniles. Part 2: main typologies, occurrences and causative factors. *Reviews in*  
433 *Aquaculture* **5**, S121-S167.

434 Cahu C., Zambonino Infante, J. & Takeuchi, T. (2003) Nutritional components affecting  
435 skeletal development in fish larvae. *Aquaculture* **227**, 245-258.

436 Cardeira J., Bensimon-Brito, A., Pousão-Ferreira, P., Cancela, M.L. & Gavaia, P.J. (2012)  
437 Lordotic-kyphotic vertebrae develop ectopic cartilage-like tissue in Senegalese sole  
438 (*Solea senegalensis*). *Journal of Applied Ichthyology* **28**, 460-463.

439 Cardeira J., Mendes, A.C., Pousão-Ferreira, P., Cancela, M.L. & Gavaia, P.J. (2015)  
440 Micro-anatomical characterization of vertebral curvatures in Senegalese sole *Solea*  
441 *senegalensis*. *Journal of Fish Biology* **86**, 1796-1810.

442 Darias M.J., Lan Chow Wing, O., Cahu, C., Zambonino-Infante, J.L. & Mazurais, D.  
443 (2010a) Double staining protocol for developing European sea bass (*Dicentrarchus*  
444 *labrax*) larvae. *Journal of Applied Ichthyology* **26**, 280-285.

445 Darias M.J., Mazurais, D., Koumoundouros, G., Glynatsi, N., Christodouloupoulou, S.,  
446 Huelvan, C., Desbruyeres, E., Le Gall, M.M., Quazuguel, P., Cahu, C.L. &  
447 Zambonino-Infante, J.L. (2010b) Dietary vitamin D<sub>3</sub> affects digestive system  
448 ontogenesis and ossification in European sea bass (*Dicentrarchus labrax*, Linnaeus,  
449 1758). *Aquaculture* **298**, 300-307.

450 Darias M.J., Mazurais, D., Koumoundouros, G., Le Gall, M.M., Huelvan, C., Desbruyeres,  
451 E., Quazuguel, P., Cahu, C.L. & Zambonino-Infante, J.L. (2011) Imbalanced dietary  
452 ascorbic acid alters molecular pathways involved in skeletogenesis of developing  
453 European sea bass (*Dicentrarchus labrax*). *Comparative Biochemistry and*  
454 *Physiology Part A: Molecular & Integrative Physiology* **159**, 46-55.

455 Dionísio G., Campos, C., Valente, L.M.P., Conceição, L.E.C., Cancela, M.L. & Gavaia, P.J.  
456 (2012) Effect of egg incubation temperature on the occurrence of skeletal  
457 deformities in *Solea senegalensis*. *Journal of Applied Ichthyology* **28**, 471-476.

458 Engrola S., Conceição, L.E.C., Gavaia, P.J., Cancela, M.L. & Dinis, M.T. (2005) Effects of  
459 pre-weaning feeding frequency on growth, survival, and deformation of  
460 Senegalese sole, *Solea senegalensis* (Kaup, 1858). *Israeli Journal of Aquaculture -*  
461 *Bamidgeh* **57**, 10-18.

462 Fernández I. & Gisbert, E. (2010) Senegalese sole bone tissue originated from chondral  
463 ossification is more sensitive than dermal bone to high vitamin A content in  
464 enriched *Artemia*. *Journal of Applied Ichthyology* **26**, 344-349.

465 Fernández I., Hontoria, F., Ortiz-Delgado, J.B., Kotzamanis, Y., Estévez, A., Zambonino-  
466 Infante, J.L. & Gisbert, E. (2008) Larval performance and skeletal deformities in  
467 farmed gilthead sea bream (*Sparus aurata*) fed with graded levels of Vitamin A  
468 enriched rotifers (*Brachionus plicatilis*). *Aquaculture* **283**, 102-115.

469 Fernández I., Pimentel, M.S., Ortiz-Delgado, J.B., Hontoria, F., Sarasquete, C., Estévez,  
470 A., Zambonino-Infante, J.L. & Gisbert, E. (2009) Effect of dietary vitamin A on  
471 Senegalese sole (*Solea senegalensis*) skeletogenesis and larval quality. *Aquaculture*  
472 **295**, 250-265.

473 Fjelldal P.G., Hansen, T., Breck, O., Ørnsrud, R., Lock, E.J., Waagbø, R., Wargelius, A. &  
474 Witten, P.E. (2012a) Vertebral deformities in farmed Atlantic salmon (*Salmo salar*  
475 L.) - etiology and pathology. *Journal of Applied Ichthyology* **28**, 433-440.

476 Fjelldal P.G., Hansen, T.J. & Berg, A.E. (2007) A radiological study on the development  
477 of vertebral deformities in cultured Atlantic salmon (*Salmo salar* L.). *Aquaculture*  
478 **273**, 721-728.

479 Fjellidal P.G., Hansen, T. & Albrektsen, S. (2012b) Inadequate phosphorus nutrition in  
480 juvenile Atlantic salmon has a negative effect on long-term bone health.  
481 *Aquaculture* **334–337**, 117-123.

482 Fontagné S., Silva, N., Bazin, D., Ramos, A., Aguirre, P., Surget, A., Abrantes, A., Kaushik,  
483 S.J. & Power, D.M. (2009) Effects of dietary phosphorus and calcium level on  
484 growth and skeletal development in rainbow trout (*Oncorhynchus mykiss*) fry.  
485 *Aquaculture* **297**, 141-150.

486 Gavaia P.J., Dinis, M.T. & Cancela, M.L. (2002) Osteological development and  
487 abnormalities of the vertebral column and caudal skeleton in larval and juvenile  
488 stages of hatchery-reared Senegal sole (*Solea senegalensis*). *Aquaculture* **211**, 305-  
489 323.

490 Gavaia P.J., Sarasquete, C. & Cancela, M.L. (2000) Detection of mineralized structures  
491 in early stages of development of marine Teleostei using a modified alcian blue-  
492 alizarin red double staining technique for bone and cartilage. *Biotechnic &*  
493 *Histochemistry : official publication of the Biological Stain Commission* **75**, 79-84.

494 Gavaia P.J., Domingues, S., Engrola, S., Drake, P., Sarasquete, C., Dinis, M.T. & Cancela,  
495 M.L. (2009) Comparing skeletal development of wild and hatchery-reared  
496 Senegalese sole (*Solea senegalensis*, Kaup 1858): evaluation in larval and postlarval  
497 stages. *Aquaculture Research* **40**, 1585-1593.

498 Georgakopoulou E., Katharios, P., Divanach, P. & Koumoundouros, G. (2010) Effect of  
499 temperature on the development of skeletal deformities in Gilthead seabream  
500 (*Sparus aurata* Linnaeus, 1758). *Aquaculture* **308**, 13-19.

- 501 Gjerde B., Pante, M.J.R. & Baeverfjord, G. (2005) Genetic variation for a vertebral  
502 deformity in Atlantic salmon (*Salmo salar*). *Aquaculture* **244**, 77-87.
- 503 Haga Y., Takeuchi, T., Murayama, Y., Ohta, K. & Fukunaga, T. (2004) Vitamin D<sub>3</sub>  
504 compounds induce hypermelanosis on the blind side and vertebral deformity in  
505 juvenile Japanese flounder *Paralichthys olivaceus*. *Fisheries Science* **70**, 59-67.
- 506 Harder W. (1975) *Anatomy of Fishes*. Schweizerbart, Stuttgart.
- 507 Hosoya K. & Kawamura, K. (1998) Skeletal formation and abnormalities in the caudal  
508 complex of the Japanese flounder, *Paralichthys olivaceus* (Temminck & Schlegel).  
509 *Bulletin of the National Research Institute of Fisheries Science* **12**, 97-110.
- 510 Howell B., Conceição, L., Prickett, R., Cañavate, P. & Mañanos, E. (2009) Sole farming:  
511 nearly there but not quite? *Aquaculture Europe* **34**, 24-27.
- 512 Imsland A.K., Foss, A., Conceição, L.E.C., Dinis, M.T., Delbare, D., Schram, E., Kamstra,  
513 A., Rema, P. & White, P. (2003) A review of the culture potential of *Solea solea* and  
514 *S. senegalensis*. *Reviews in Fish Biology and Fisheries* **13**, 379-407.
- 515 Kihara M., Ogata, S., Kawano, N., Kubota, I. & Yamaguchi, R. (2002) Lordosis induction  
516 in juvenile red sea bream, *Pagrus major*, by high swimming activity. *Aquaculture*  
517 **212**, 149-158.
- 518 Koumoundouros G. (2010) Morpho-anatomical abnormalities in Mediterranean marine  
519 aquaculture. In: *Recent Advances in Aquaculture Research* (ed. by G.  
520 Koumoundouros), pp. 125-148. Transworld Research Network, Kerala, India.



521 Koumoundouros G., Gagliardi, F., Divanach, P., Boglione, C., Cataudella, S. & Kentouri,  
522 M. (1997a) Normal and abnormal osteological development of caudal fin in *Sparus*  
523 *aurata* L. fry. *Aquaculture* **149**, 215-226.

524 Koumoundouros G., Oran, G., Divanach, P., Stefanakis, S. & Kentouri, M. (1997b) The  
525 opercular complex deformity in intensive gilthead sea bream (*Sparus aurata* L.)  
526 larviculture. Moment of apparition and description. *Aquaculture* **156**, 165-177.

527 Kranenbarg S., Waarsing, J.H., Muller, M., Weinans, H. & van Leeuwen, J.L. (2005)  
528 Lordotic vertebrae in sea bass (*Dicentrarchus labrax* L.) are adapted to increased  
529 loads. *Journal of Biomechanics* **38**, 1239-1246.

530 Lall S.P. & Lewis-McCrea, L.M. (2007) Role of nutrients in skeletal metabolism and  
531 pathology in fish — An overview. *Aquaculture* **267**, 3-19.

532 Lewis L.M., Lall, S.P. & Witten, P.E. (2004) Morphological descriptions of the early  
533 stages of spine and vertebral development in hatchery-reared larval and juvenile  
534 Atlantic halibut (*Hippoglossus hippoglossus*). *Aquaculture* **241**, 47-59.

535 Lewis-McCrea L.M. & Lall, S.P. (2010) Effects of phosphorus and vitamin C deficiency,  
536 vitamin A toxicity, and lipid peroxidation on skeletal abnormalities in Atlantic  
537 halibut (*Hippoglossus hippoglossus*). *Journal of Applied Ichthyology* **26**, 334-343.

538 Losada A.P., de Azevedo, A.M., Barreiro, A., Barreiro, J.D., Ferreiro, I., Riaza, A.,  
539 Quiroga, M.I. & Vázquez, S. (2014) Skeletal malformations in Senegalese sole  
540 (*Solea senegalensis* Kaup, 1858): gross morphology and radiographic correlation.  
541 *Journal of Applied Ichthyology* **30**, 804-808.

542 Madsen L., Arnbjerg, J. & Dalsgaard, I. (2001) Radiological examination of the spinal  
543 column in farmed rainbow trout *Oncorhynchus mykiss* (Walbaum): experiments  
544 with *Flavobacterium psychrophilum* and oxytetracycline. *Aquaculture Research* **32**,  
545 235-241.

546 Mazurais D., Darias, M.J., Gouillou-Coustans, M.F., Le Gall, M.M., Huelvan, C.,  
547 Desbruyeres, E., Quazuguel, P., Cahu, C. & Zambonino-Infante, J.L. (2008) Dietary  
548 vitamin mix levels influence the ossification process in European sea bass  
549 (*Dicentrarchus labrax*) larvae. *American Journal of Physiology-Regulatory,*  
550 *Integrative and Comparative Physiology* **294**, R520-R527.

551 Mazurais D., Glynatsi, N., Darias, M.J., Christodoulopoulou, S., Cahu, C.L., Zambonino-  
552 Infante, J. & Koumoundouros, G. (2009) Optimal levels of dietary vitamin A for  
553 reduced deformity incidence during development of European sea bass larvae  
554 (*Dicentrarchus labrax*) depend on malformation type. *Aquaculture* **294**, 262-270.

555 Nagano N., Hozawa, A., Fujiki, W., Yamada, T., Miyaki, K., Sakakura, Y. & Hagiwara, A.  
556 (2007) Skeletal development and deformities in cultured larval and juvenile seven-  
557 band grouper, *Epinephelus septemfasciatus* (Thunberg). *Aquaculture Research* **38**,  
558 121-130.

559 Pimentel M.S., Faleiro, F., Dionísio, G., Repolho, T., Pousão-Ferreira, P., Machado, J. &  
560 Rosa, R. (2014) Defective skeletogenesis and oversized otoliths in fish early stages  
561 in a changing ocean. *Journal of Experimental Biology* **217**, 2062-2070.

- 562 Roberts R.J. & Rodger H.D. (2001) The pathophysiology and systematic pathology of  
563 teleosts. In: *Fish Pathology* (ed. by R. J. Roberts), pp. 55-132. W. B. Saunders,  
564 London.
- 565 Rodríguez J.L. & Peleteiro J.B. (2014) *Cultivo del lenguado senegalés (Solea*  
566 *senegalensis)*. Fundación Observatorio Español de Acuicultura, Madrid.
- 567 Russo T., Prestinicola, L., Scardi, M., Palamara, E., Cataudella, S. & Boglione, C. (2010)  
568 Progress in modeling quality in aquaculture: an application of the Self-Organizing  
569 Map to the study of skeletal anomalies and meristic counts in gilthead seabream  
570 (*Sparus aurata*, L. 1758). *Journal of Applied Ichthyology* **26**, 360-365.
- 571 Sakamoto K. (1984) Interrelationships of the family Pleuronectidae (Pisces:  
572 Pleuronectiformers). *Memoirs of the Faculty of Fisheries, Hokkaido University* **31**,  
573 95-215.
- 574 Sfakianakis D.G., Georgakopoulou, E., Papadakis, I.E., Divanach, P., Kentouri, M. &  
575 Koumoundouros, G. (2006) Environmental determinants of haemal lordosis in  
576 European sea bass, *Dicentrarchus labrax* (Linnaeus, 1758). *Aquaculture* **254**, 54-64.
- 577 Stiassny M.L.J. (2000) Chapter 6 - Skeletal System. In: *The Laboratory Fish* (ed. by G. K.  
578 Ostrander), pp. 109-118. Academic Press, London.
- 579 Tong X., Liu, Q., Xu, S., Ma, D., Xiao, Z., Xiao, Y. & Li, J. (2012) Skeletal development and  
580 abnormalities of the vertebral column and of the fins in hatchery-reared turbot  
581 *Scophthalmus maximus*. *Journal of Fish Biology* **80**, 486-502.

- 582 Witten P.E., Gil-Martens, L., Huysseune, A., Takle, H. & Hjelde, K. (2009) Towards a  
583 classification and an understanding of developmental relationships of vertebral  
584 body malformations in Atlantic salmon (*Salmo salar* L.). *Aquaculture* **295**, 6-14.
- 585 Witten P.E. & Huysseune, A. (2009) A comparative view on mechanisms and functions  
586 of skeletal remodelling in teleost fish, with special emphasis on osteoclasts and  
587 their function. *Biological Reviews* **84**, 315-346.
- 588 Witten P.E., Obach, A., Huysseune, A. & Baeverfjord, G. (2006) Vertebrae fusion in  
589 Atlantic salmon (*Salmo salar*): development, aggravation and pathways of  
590 containment. *Aquaculture* **258**, 164-172.

591 **Tables:**

<b>Measurements</b>	<b>Mean</b>	<b>SD</b>	<b>Mode</b>	<b>Median</b>	<b>Min</b>	<b>Max</b>
<b>StL (cm)</b>	1.44 ± 0.14		1.5	1.4	0.7	1.8
<b>StH (cm)</b>	0.46 ± 0.05		0.5	0.5	0.3	0.5
<b>Ratio StL/StH</b>	3.29 ± 0.42		3.0	3.2	2.0	5.3

592 Table 1. Specimen measurements (StL, StH, and Ratio StL/StH): mean ± standard

593 deviation (SD), mode, median, minimum (Min), and maximum (Max).

<b>Meristic counts</b>	<b>Mean</b>	<b>SD</b>	<b>Mode</b>	<b>(%)</b>	<b>Median</b>	<b>Min</b>	<b>Max</b>
<b>Abdominal vertebrae</b>	8.93	± 0.28	9	92.11	9	8	11
<b>Caudal vertebrae</b>	33.57	± 0.68	34	51.48	34	30	35
<b>Caudal complex vertebrae</b>	3.05	± 0.23	3	94.67	3	2	4
<b>Caudal fin rays</b>	20.25	± 0.54	20	79.09	20	20	24

594 Table 2. Number of vertebrae in abdominal, caudal, and caudal complex regions, as  
595 well as caudal fin ray counts: mean ± standard deviation (SD), mode, percentage of  
596 individuals exhibiting the most frequent value (%), median, minimum (Min), and  
597 maximum (Max).

<b>Number of individuals with at least one anomaly (%)</b>	100.0
<b>Total number of anomalies</b>	5,654
<b>Number of anomalies/number of affected individuals</b>	11.2
<b>Number of individuals with VBA and/or VCD (%)</b>	51.7
<b>Total number of VBA and VCD</b>	913
<b>Number of VBA and VCD/number of individuals with VBA and/or VCD</b>	3.5
<b>Number of VBA and VCD/total number of anomalies (%)</b>	16.1

598 Table 3. General results on the incidence of skeletal anomalies in the studied fish:  
599 number of individuals displaying at least one anomaly (%); number of individuals with  
600 vertebral body anomalies (VBA) and/or rachis deviations (VCD) (%); total number of  
601 skeletal abnormalities; number of VBA and VCD and the percentage of VBA and VCD in  
602 the total number of anomalies; anomalies and VBA and/or VCD loads.

		Anomaly typology	N.I.	%N.I.	N.A.	N.A./I.
Abdominal region	Parapophysis	Bifurcation	1	0.20	1	1.00
		Number alteration	65	12.82	71	1.09
		Insertion alteration	3	0.59	3	1.00
		Fusion	8	1.58	9	1.13
		Deformation	4	0.79	4	1.00
	Neural arches and spines	Bifurcation	3	0.59	3	1.00
		Number alteration	12	2.37	24	2.00
		Insertion alteration	8	1.58	13	1.63
		Fusion	8	1.58	9	1.13
		Incomplete arch	16	3.16	36	2.25
	VBA	Fusion	55	10.85	62	1.13
		Deformation	28	5.52	65	2.32
	VCD	Kyphosis	4	0.79	4	1.00
		Lordosis	10	1.97	10	1.00
		Scoliosis	3	0.59	3	1.00
Caudal region	Neural arches and spines	Bifurcation	2	0.39	2	1.00
		Number alteration	31	6.11	46	1.48
		Insertion alteration	30	5.92	58	1.93
		Fusion	14	2.76	15	1.07
		Incomplete arch	61	12.03	101	1.66
		Deformation	79	15.58	224	2.84
	Haemal arches and spines	Bifurcation	3	0.59	3	1.00
		Number alteration	21	4.14	30	1.43
		Insertion alteration	53	10.45	71	1.34
		Fusion	22	4.34	30	1.36
		Incomplete arch	75	14.79	111	1.48
	VBA	Fusion	81	15.98	110	1.36
		Deformation	83	16.37	252	3.04
	VCD	Kyphosis	28	5.52	30	1.07
		Lordosis	36	7.10	43	1.19
Scoliosis		15	2.96	16	1.07	
Caudal complex	Neural arches and spines	Bifurcation	1	0.20	1	1.00
		Number alteration	32	6.31	32	1.00
		Insertion alteration	4	0.79	4	1.00
		Fusion	31	6.11	31	1.00
		Incomplete arch	13	2.56	13	1.00
		Deformation	56	11.05	59	1.05
	Haemal arches and spines	Bifurcation	2	0.39	2	1.00
		Number alteration	13	2.56	13	1.00
		Insertion alteration	6	1.18	8	1.33
		Fusion	45	8.88	45	1.00
		Incomplete arch	24	4.74	26	1.08
	Hypurals epural parhypural	Bifurcation	0	0.00	0	0.00
		Number alteration	0	0.00	0	0.00
		Insertion alteration	0	0.00	0	0.00
		Fusion	231	45.56	282	1.22
Deformation		471	92.90	777	1.65	
VBA	Fusion	95	18.74	95	1.00	
	Deformation	127	25.05	206	1.62	
VCD	Kyphosis	13	2.56	13	1.00	
	Lordosis	1	0.20	1	1.00	
	Scoliosis	3	0.59	3	1.00	

603 Table 4. Frequencies of skeletal alterations by location, affected skeletal structure, and  
604 type: number of fish showing at least one skeletal anomaly (N.I.), percentage of affected



605 individuals (%N.I.), number of observed alterations (N.A.), and average number of  
606 malformations per specimen (N.A./I.). VBA: Vertebral body anomalies. VCD: Vertebral  
607 column deviations.

608 **Figure legends:**

609 Figure 1. Sole (*Solea senegalensis*) larva stained with alcian blue and alizarin red,  
610 showing anatomical regions considered: abdominal, caudal, and caudal complex. Note  
611 the slightly twisted front caudal haemal spines and bending of caudal plates. P:  
612 parapophysis; N: neural arch and spine; H: haemal arch and spine; V: vertebral centrum;  
613 Pu1: preural 1; Pu2: preural 2; U: urostyle; H1-H5: hypurals 1-5; E: epural; Ph:  
614 parhypural; R: caudal fin rays. Bar = 5 mm. Insert bar = 500  $\mu$ m.

615 Figure 2. Main skeletal alterations in 31/32 dah Senegalese sole (*Solea senegalensis*). A.  
616 Absence of the neural arch of the first abdominal vertebra (white arrowhead); slight  
617 deformities in neural arches (black arrowheads) and alteration in the number of  
618 parapophyses (black arrow); note that some parapophyses were fused, forming an arch  
619 (asterisk). B. Twisted haemal elements in the caudal region (black arrowheads). C.  
620 Slightly incomplete neural and haemal arches (white arrowheads). D. Alteration of the  
621 haemal spine insertion, note the lack of symmetry of vertebral bodies (white asterisks).  
622 E. Slight bending of all caudal plates; hypurals 3 and 4 are fused (white star); fusion  
623 between the last caudal vertebra and preural 2, with severe deformations of the  
624 vertebral bodies (white asterisks) and fusion among neural arches (white arrowhead),  
625 preural 2 showed an incomplete vertebral centrum (lacked a part of the vertebra);  
626 twisted haemal spines (black arrowhead) associated to this lesion. Bars = 500  $\mu$ m.

627 Figure 3. Distinct abnormalities observed in the vertebral column. A. Multiple severe  
628 VBA as fusions (black arrows) and deformations of the affected vertebrae, mostly in  
629 abdominal and caudal regions. Misaligned vertebrae (black asterisk), kyphosis (K), and  
630 lordosis (L) were also present. In the lordotic segment, vertebrae were also slightly

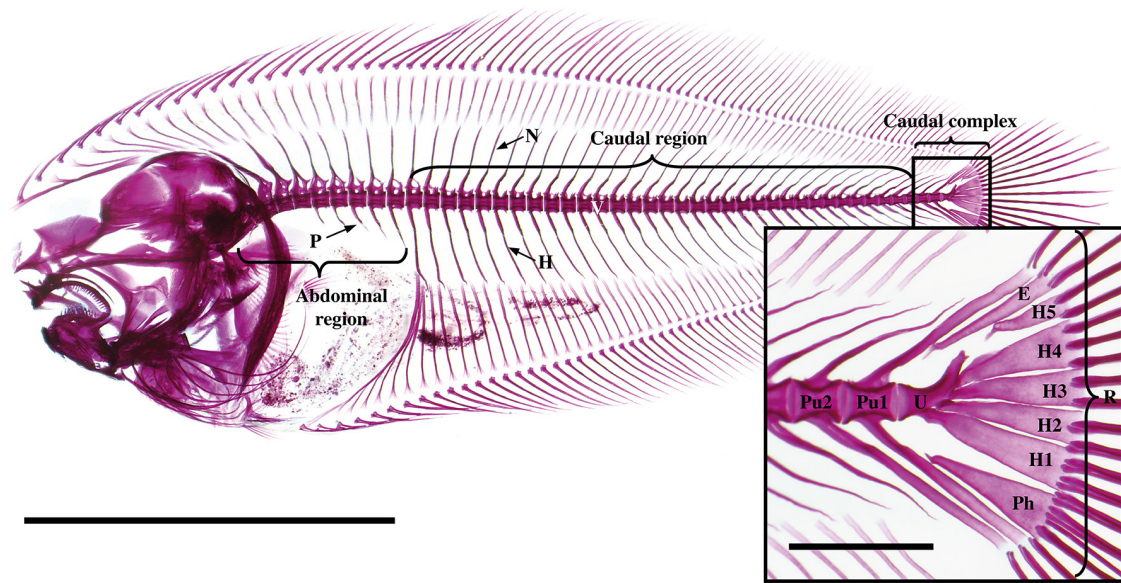
631 deviated to the blind side of the fish. Note the alterations in the insertion of neural and  
632 haemal arches in the affected segments, and the slight bending of some neural and  
633 haemal spines and caudal complex plates. B. Caudal vertebral fusion and incomplete  
634 neural arch (white arrowhead). C. Fusions among preurals, note the remaining line of  
635 fusion (arrow) and mild deformations of neural and haemal spines (black arrowheads).  
636 Fusion of hypurals 3-4 (white star), and slight bending of the caudal complex plates. D.  
637 Presence of an extra element between the preurals (arrow), fused with preural 1, with  
638 kyphosis (K) and deformations in the caudal complex vertebrae and fusion among the  
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640 of haemal spines (black arrowhead) and in caudal complex plates were observed. E.  
641 Gross deformations of abdominal vertebral centres (white asterisks) and lordosis (L),  
642 with slight deviation towards the blind aspect of the individual. Note the enlargement  
643 and the darker and irregular articular surface in the affected abdominal vertebrae. The  
644 first abdominal vertebra lacked a neural arch (white arrowhead). Deformations of neural  
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647 Figure 4. Frequency of vertebral anomalies (VBA and VCD) per specimen. Most affected  
648 fish presented one or three alterations (90 and 61 *Solea senegalensis*, respectively),  
649 although ten of them showed more than 10 VBA or VCD. VBA: Vertebral body anomalies.  
650 VCD: Vertebral column deviations.

651 Figure 5. Distribution of the vertebral body anomalies (VBA) throughout the spinal  
652 column. A. Fusions. The first two abdominal vertebrae and preurals were the most  
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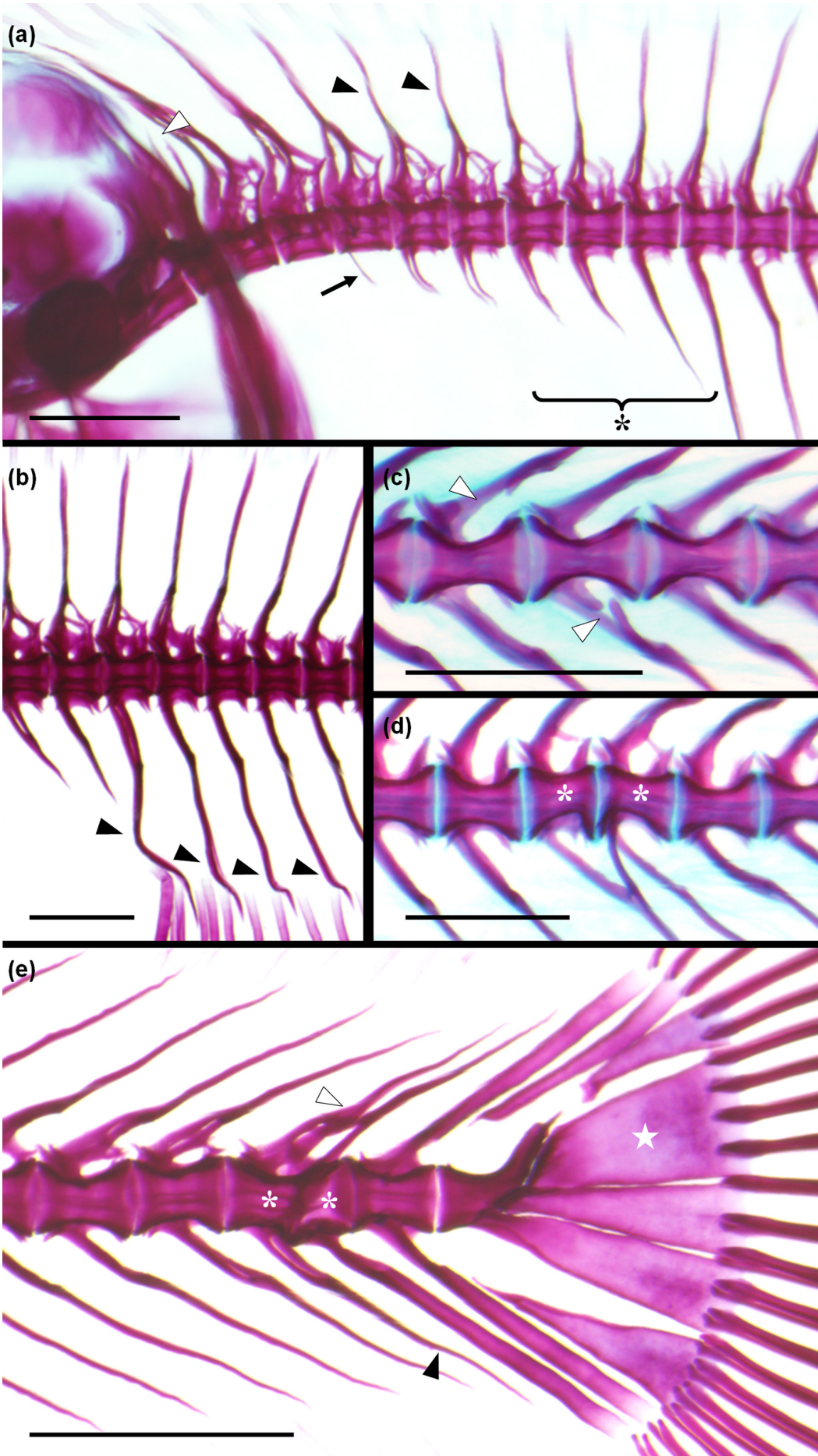
656 Figure 6. Distribution of the vertebral column deviations (VCD) throughout the spinal  
657 column. A. Kyphosis. These deviations were more frequent in the caudal complex region.  
658 B. Lordosis. The intermediary vertebral segment between abdominal and caudal regions  
659 was the most affected by lordotic curvatures. C. Scoliosis. Scoliotic lesions were  
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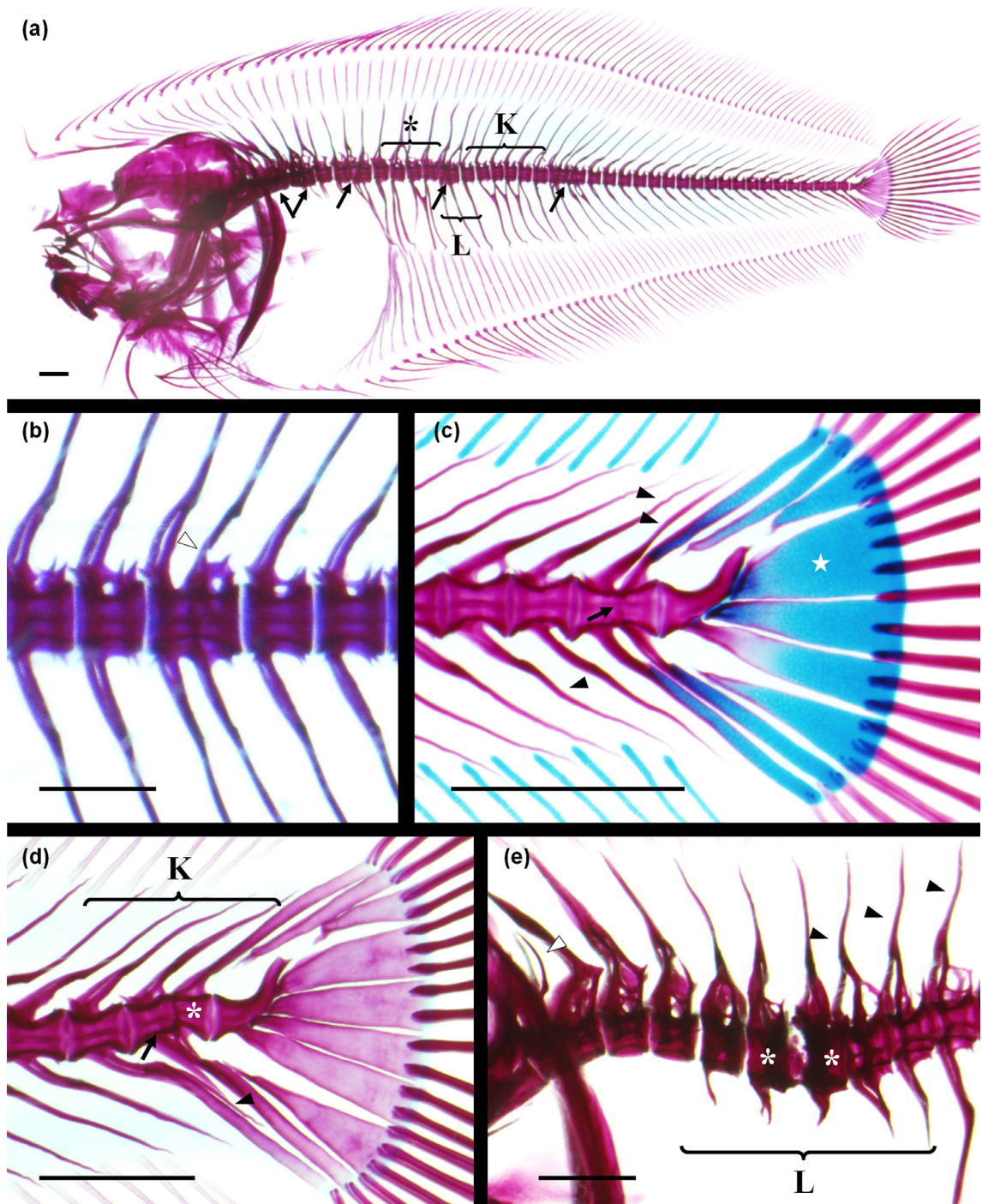
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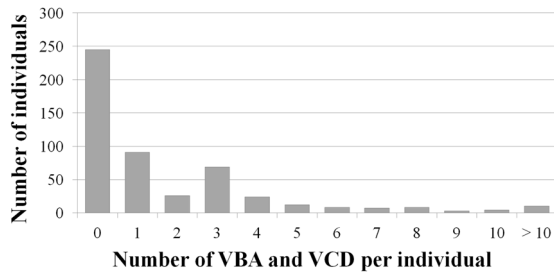


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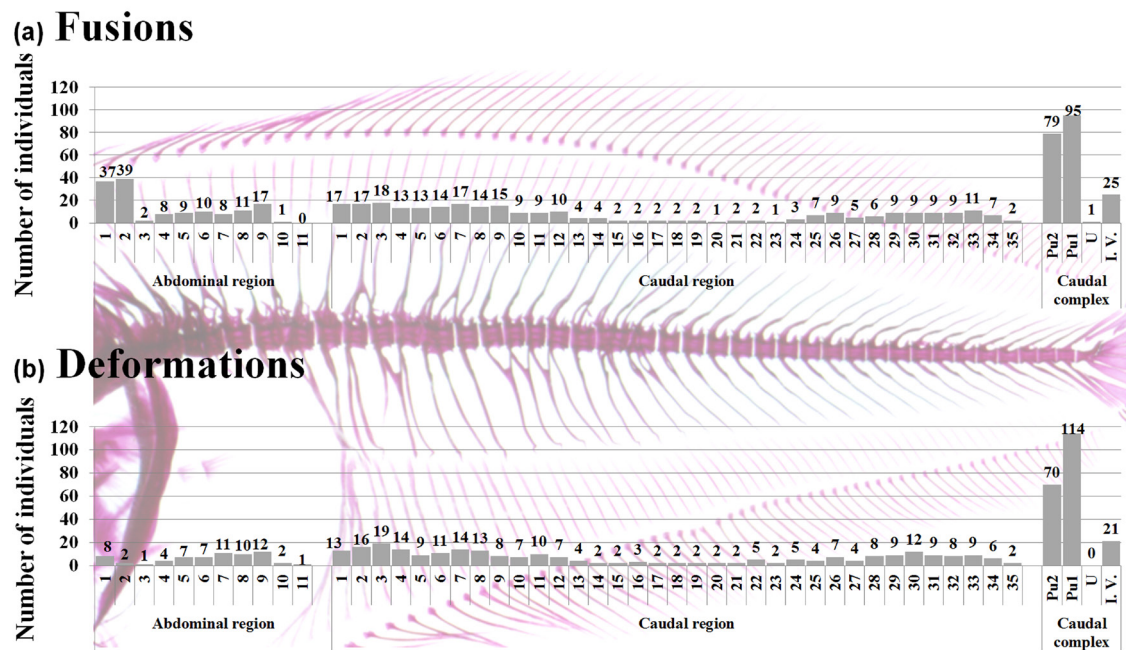
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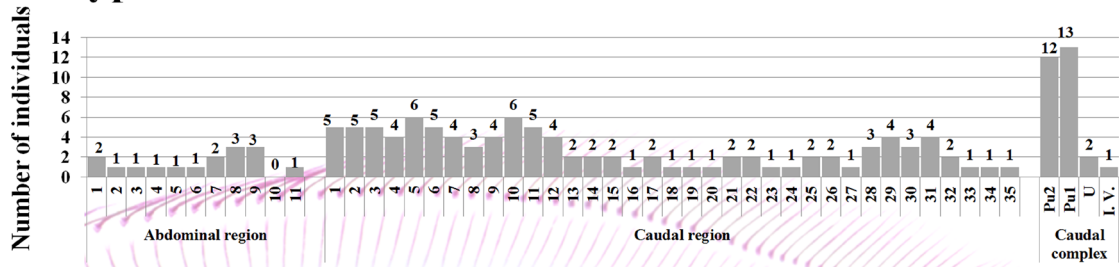
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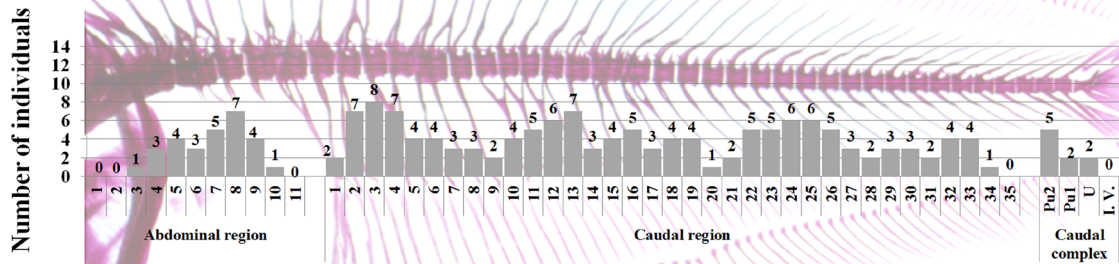
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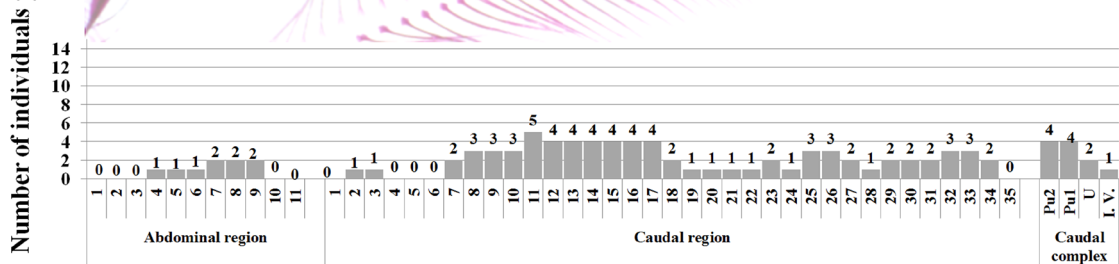
**(a) Kyphosis**



**(b) Lordosis**



**(c) Scoliosis**



720

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