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Abstract	*Cupriavidus* sp. UYMMa02A is a beta-rhizobia strain of the *Cupriavidus* genus, isolated from nodules of *Mimosa magentea* in Uruguay. This strain can form effective nodules with several *Mimosa* species, including its original host. Genome analyses indicate that *Cupriavidus* sp. UYMMa02A has a highly conserved 35 kb symbiotic island containing *nod, nif,* and *fix* operons, suggesting conserved mechanisms for the symbiotic interaction with plant hosts. However, while *Cupriavidus* sp. UYMMa02A produces functional nodules and promotes *Mimosa pudica* growth under nitrogen-limiting conditions, *nod* genes are not induced by luteolin or exposure to *Mimosa* spp. root exudate. To explore alternative mechanisms implicated in the *Cupriavidus*-*Mimosa* interaction, we assessed the proteomic profiles of *Cupriavidus* sp. UYMMa02A grown in the presence of pure flavonoids and co-culture with *Mimosa pudica* plants. This approach allowed us to identify 24 differentially expressed proteins potentially involved in bacterial-plant interaction. In light of the obtained results, a possible model for *nod*-alternative symbiotic interaction is proposed.

Keywords (separated by '-')	Beta-rhizobia - *Cupriavidus* - Luteolin - Apigenin - Proteomic - 2D-DIGE - *Mimosa pudica* - *Mimosa magentea* - Root exudates

Footnote Information	Cecilia Rodríguez-Esperón and Laura Sandes contributed equally to the present work.

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ORIGINAL ARTICLE

Nodulation in the absence of *nod* genes induction: alternative mechanisms involved in the symbiotic interaction between *Cupriavidus* sp. UYMMa02A and *Mimosa pudica*

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Abstract

Cupriavidus sp. UYMMa02A is a beta-rhizobia strain of the *Cupriavidus* genus, isolated from nodules of *Mimosa magentea* in Uruguay. This strain can form effective nodules with several *Mimosa* species, including its original host. Genome analyses indicate that *Cupriavidus* sp. UYMMa02A has a highly conserved 35 kb symbiotic island containing *nod, nif,* and *fix* operons, suggesting conserved mechanisms for the symbiotic interaction with plant hosts. However, while *Cupriavidus* sp. UYMMa02A produces functional nodules and promotes *Mimosa pudica* growth under nitrogen-limiting conditions, *nod* genes are not induced by luteolin or exposure to *Mimosa* spp. root exudate. To explore alternative mechanisms implicated in the *Cupriavidus-Mimosa* interaction, we assessed the proteomic profiles of *Cupriavidus* sp. UYMMa02A grown in the presence of pure flavonoids and co-culture with *Mimosa pudica* plants. This approach allowed us to identify 24 differentially expressed proteins potentially involved in bacterial-plant interaction. In light of the obtained results, a possible model for *nod*-alternative symbiotic interaction is proposed.

Keywords Beta-rhizobia · *Cupriavidus* · Luteolin · Apigenin · Proteomic · 2D-DIGE · *Mimosa pudica* · *Mimosa magentea* · Root exudates

Introduction

Rhizobia are soil bacteria that engage in symbiotic interactions with legume plants. During this interaction, new specialized organs called nodules are formed in the roots (and sometimes in stems) of host plants. Inside the nodules, bacteria convert atmospheric nitrogen into ammonia in a process known as Symbiotic Nitrogen Fixation (SNF) (Lindström and Mousavi 2020).

The ability to form nitrogen-fixing nodules in symbiosis with legumes is restricted to alpha and beta subgroups of proteobacteria (Andrews and Andrews 2017). The best-characterized rhizobia species; *Sinorhizobium meliloti*, *Rhizobium leguminosarum,* or *Bradyrhizobium japonicum,* belong to the *Rhizobiaceae* family of alpha-proteobacteria. In beta-proteobacteria, the *Burkholderiaceae* family is the best characterized and includes just three genera *Paraburkholderia, Trinickia,* and *Cupriavidus* (Chen et al. 2001, 2003; Dall'Agnol et al. 2017; Estrada-de los Santos et al. 2018). Both *Paraburkholderia* and *Cupriavidus* rhizobia strains were described at the beginning of the twenty-first

Cecilia Rodríguez-Esperón and Laura Sandes contributed equally to the present work.

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century. Since then, beta-rhizobia have been described as legume symbionts in America (Bontemps et al. 2010; Andam et al. 2007; dos Reis et al. 2010; Taulé et al. 2012), Africa (Garau et al. 2009; Howieson et al. 2013; Lemaire et al. 2015), Asia (Liu et al. 2012; Gehlot et al. 2013) and Oceania (Parker et al. 2007), primarily associated with legumes of Mimosoid clade, but also with some members of the *Papilionoideae* (Garau et al. 2009; Lemaire et al. 2015). Despite their worldwide distribution, the molecular mechanisms involved in the interaction between beta-rhizobia and host plants have been analyzed in a few model strains (Amadou et al. 2008; de Campos et al. 2017; Lardi et al. 2017; Klonowska et al. 2018; Bellés-Sancho et al. 2022; Rodríguez-Esperón et al. 2022). Genome analyses of beta-rhizobia that nodulate Mimosoid clade have shown the presence of a highly conserved and compact genomic region, known as the symbiotic island, that encodes for *nod, nif,* and *fix* genes (De Meyer et al. 2016; Zheng et al. 2017). The *nod* genes code for proteins involved in the synthesis (*nodBCHASUQ*) and exportation (*nodIJ)* of nodulation (Nod) factors, as well as their regulation (*nodD*). In turn, the *nif* genes encode proteins related to the nitrogenase complex (*nifH, nifD,* and *nifK),* regulation (*nifA*), and maturation processes (*nifEDXQ*). The *fix* genes encode membrane proteins required for electron transfer to generate the energy required for the SNF process (*fixABCX; fixNOPQ*).

In beta-rhizobia, *nod* genes are induced in the presence of pure flavonoids such as luteolin and apigenin (Marchetti et al. 2011; Rodríguez-Esperón et al. 2022) or root exudates of the host plant *Mimosa pudica* (Klonowska et al. 2018). In line with the conservation of this important recognition mechanism, other genes and molecules involved in bacteria-plant interaction have been identified in beta-rhizobia. These include the type-III secretion system (Saad et al. 2012) and synthesis of branched-chain amino acids in *Cupriavidus taiwanensis* LMG19424 (Chen et al. 2012); type VI secretion system (de Campos et al. 2017; Lardi et al. 2017) and synthesis of exopolysaccharide (EPS) cepacian in *Paraburkholderia phymatum* STM815 (Liu et al. 2020). Despite these few examples, there is still a lack of knowledge about the molecular mechanisms implicated in the symbiotic interaction between beta-rhizobia and legume hosts.

In Uruguay, beta-rhizobia have been identified as the main symbionts of *Parapiptadenia rigida* (Taulé et al. 2012) and *Mimosa* spp. (Platero et al. 2016; Pereira-Gómez et al. 2020). Phylogenetic analyses indicated that *Cupriavidus* strains symbiotically associated with these plants do not belong to the well-studied *C. taiwanensis* species but rather to the *C. necator* and other *Cupriavidus* species.

Cupriavidus sp. UYMMa02A was isolated from nodules of *Mimosa magentea, a* native legume found in the southeast region of Uruguay. Through a Multi-Locus Sequence Analysis (MLSA) approach, it was demonstrated that UYMMa02A does not belong to the previously described *C. taiwanensis* and *C. necator,* but it may represent a novel rhizobial species within the *Cupriavidus* genus (Platero et al. 2016). Genome sequencing of *Cupriavidus* sp. UYMMa02A (Iriarte et al. 2016) indicated the presence of a conserved and compact symbiotic island that encodes for the *nod, nif,* and *fix* genes observed in other beta-rhizobia (Amadou et al. 2008; Moulin et al. 2014; De Meyer et al. 2015a, b, 2016). In addition, this strain induces pink nodules on the roots of its original host and other *Mimosa* species, including *M. pudica* (Platero et al. 2016).

The present study aimed to analyze the initial steps of the *Cupriavidus-Mimosa* symbiotic interaction. Firstly, we determined the expression of *nod* genes in *Cupriavidus* sp. UYMMa02A strain when cultivated in the presence of pure flavonoids or *Mimosa* spp. root exudates. Then we analyzed the proteomic changes induced in the bacteria by the presence of pure flavonoids and the plant host. Finally, we integrated the obtained results into a model that reports the changes occurring during the initial steps of the *Cupriavidus* sp. UYMMa02A—*Mimosa pudica* symbiotic interaction.

Material and methods

Bacterial strains, plasmids, and growth conditions

The bacteria and plasmids used in this study are listed in Table 1. *Escherichia coli* strains were grown aerobically at 37 °C in Luria–Bertani (LB) medium. *Cupriavidus* strains were grown at 30 °C in LB or M9 minimal medium containing 14 mM sodium citrate as a carbon source (Sambrook et al. 1989). When indicated M9 cultures were supplemented with 5 µM luteolin or apigenin. Modified M9 media containing *Mimosa* spp. root exudates were prepared to replace the original water volume used in M9, by filter-sterilized root exudates. When required the following antibiotics were used for strain selection; Ampicillin 100 µg ml^{-1} (Ap), Nitrofurantoin 50 µg ml^{-1} (Nf), Chloramphenicol 25 µg ml^{-1} (Cf), and Tetracycline 8 µg ml^{-1} (Tc).

Sequence analysis

Homology searches and sequences retrieval were done via Internet server BLAST (NCBI, NIH, Bethesda, MD, USA: AQ4 http://www.ncbi.nlm.nih.gov) and RAST server (Aziz et al. 2008). Sequence alignments were done by Mega v7.0.26 (Kumar et al. 2016) and SnapGene® software (from Insightful Science; available at snapgene.com) was used for graphic presentation.

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Table 1 Strains and plasmids used in this work

Strain	Relevant characteristics	Reference
Escherichia coli DH5α	Cloning host; F^- λ^- *endA1 glnX44(AS) thiE1 recA1 relA1 spoT1 gyrA96*(Nal^R) *rfbC1 deoR nupG Φ80*(*lacZ*ΔM15) Δ(*argF-lac*)169 *hsdR*17	Hanahan (1983)
Cupriavidus sp. UYMMa02A	Wild-type strain, isolated from *Mimosa magentea* nodules in Uruguay	Platero et al. (2016)
Cupriavidus taiwanensis LMG19424^T	Type strain. Isolated from *Mimosa pigra* nodules in Taiwan	Chen et al. (2001)
Cupriavidus necator UYPR2.512	Wild-type strain, isolated from *Parapiptadenia rigida* nodules in Uruguay	Taulé et al. (2012)
Plasmid		
pRK600	Helper plasmid used for conjugation, Cf^R	Kessler et al. (1992)
pCZ388	pLAFR6 derivative containing a promoterless *lacZ* gene, Tc^R	Cunnac et al. (2004)
pCBM01	pCZ388 containing 401 bp of the *Cupriavidus taiwanensis* LMG19424 *nodB* promoter, Tc^R	Marchetti et al. (2010)

Reporter strains construction

To evaluate the expression of *nodB* gene promoter, plasmid pCBM01 containing the $pnodB_{19424}$-*lacZ* transcriptional fusion (Marchetti et al. 2010) or plasmid pCZ388 containing a promoterless *lacZ* (Cunnac et al. 2004) were introduced in *Cupriavidus* spp. strains by triparental mating as previously described (Rodríguez-Esperón et al. 2022).

Assessment of $pnodB_{19424}$-*lacZ gene* expression

Cultures of *Cupriavidus* spp. strains carrying the plasmids pCBM01 or pCZ388 were grown overnight at 30 °C using a rotary shaker at 200 rpm, in 5 mL of M9 supplemented with Tc. At the end of this time, a 1/100 (v/v) dilution was made in fresh medium supplemented with 5 µM luteolin (≥ 98% TLC, SIGMA, USA) or in a modified M9 prepared with roots exudates and incubated for 18 h at 30ºC. Uninduced control cultures were included for each assay. Beta-galactosidase assays were performed according to the standard Miller assay (Miller 1972).

Pre-treatment, surface sterilization, and germination of *Mimosa* spp. seeds

M. pudica seeds were obtained commercially from Outsidepride Seeds, LLC (Oregon, USA). *M. magentea* seeds were collected from native plant populations growing in Uruguay (Platero et al. 2016). The seeds were submerged in ethanol 95% for 2 min and dried in filter paper. After they were treated with 10 M sulfuric acid for 20 min, followed by seven washes with sterile distilled water. Finally, the seeds were treated with 4% sodium hypochlorite for 5 min followed by seven washes with sterile distilled water. Surface-sterilized seeds were germinated on 0.8% (wt/vol) agar-water plates, at 30 °C in dark conditions for 2 days.

Roots exudate collection method

Mimosa spp. pre-germinated seeds were sown in a 250 mL test glass jar (20 seeds per flask) containing 20 mL of sterile water and a stainless-steel grate for seedling support. Plants were incubated for five days under a photoperiod of 16 h light/8 h darkness at 26 °C. After that, the water solution containing the roots exudates was collected in 50 mL plastic conical tubes, centrifuged for 5 min at 6,000 g to remove cellular debris and supernatants were filtered-sterilized using 0.45 µm membrane filters (Millipore, USA). Filter-sterilized root exudates were used, instead of water, for preparing modified M9 media. All experiments including root exudates were performed with freshly (same day) prepared root exudates.

Gene expression based on quantitative reverse transcription-PCR (RT-qPCR)

The RT-qPCR experiments were performed with *Cupriavidus* sp. UYMMa02A growing in M9 with *M. pudica* root exudates or M9 media. For the bacterial RNA extraction, *Cupriavidus* sp. UYMMa02A was cultivated in 150 mL of modified M9 with *M. pudica* root exudates or M9 until reaching the mid-exponential phase (OD_{600} between 0.6–0.8). Cultures were then incubated for 20 min with 100 µg/mL chloramphenicol and then rapidly cooled by placing the tubes in a water–ice bath. Cells were recovered by centrifuging at 5000*g* for 5 min at 4 °C. Obtained pellets were suspended in 1.5 mL of lysis buffer (20 mM Tris–HCl Buffer pH 7.6; 50 mM $MgCl_2$; 150 mM NH_4Cl and transferred to 2 mL lysis tubes (MP Biomedicals Lysing Matrix Tubes # Matrix B). Cell lysis was carried out on Fast Prep equipment (MP Biomedicals) using a cycle of 6.0 m/s for 60 s. Finally, lysates were centrifuged at 10,000*g* for 20 min at 4 °C, and supernatants containing the cell extract were transferred to a new tube. One hundred microliters of the obtained extracts were used for total RNA extraction

using a PureLink RNA Mini Kit (Thermo Fisher Scientific #12183018A). Obtained RNA was treated with 5 units of DNase I (Thermo Fisher Scientific EN0521) for 5 min at 37 °C. One microgram of total RNA was converted to cDNA using the High-Capacity cDNA Reverse Transcription Kit (Applied Biosystem), following the manufacturer's recommendations. Quantitative PCR (qPCR) analyses were performed essentially as described (Rodríguez-Esperón et al. 2022). The UYMMa02A *efg* gene (ODV42482.1) which encodes for the elongation factor G and the *s14* gene (ODV43065.1) which encodes for the ribosome protein S14, were used as reference genes. The UYMMa02A *nodA, nodB,* and *nodC* genes were selected to analyze relative gene expression. Primer Blast software was used to design primer sets for *nodA, nodB*, *nodC, efg, and s14*. The following primers were used: efg-for (5′-GCGATCATTTGGGACGAAGC-3′); efg-rev (5′-CGGACTCGACCATCTTCTCG-3′); s14-for (5′-CTGTTCTACGTGTCAG-3′); s14-rev (5′-TGATGTTGATGCGGTGTTC-3′); nodA-for (5′-ACGTCCTCGCTGTGATTCTG-3′); nodA-rev (5′-AGGTCCGTTGCGTTCGATAG-3′); nodB-for (5′-TGGGGCAATTTCAGCTTCCA-3′; nodB-rev (5′-AGCGACTTCGTGTCCTTCAG-3′); nodC-for (5′-CAGAGCTTGCCTCACTTCCA-3′); nodC-rev (5′-TGCATCGTCCTCATAGTCGC-3′). qPCRs were performed on a C1000 Touch Thermal Cycler (Bio-Rad) using the iQTM SYBR Green Supermix (Bio-Rad). qPCRs conditions were as follows: 5 min at 95 °C, 40 cycles of 15 s at 95ºC, 30 s at 60 °C and 30 s at 72 °C. Primer specificity and dimer formation were checked by dissociation curves. A mixture of cDNA from induced and non-induced samples was used for calculating primer efficiency. The relative gene expression level was calculated using the $2^{-\Delta\Delta Cq}$ method, statistical analyses were performed in the InfoStat statistical program (https://www.infostat.com.ar/). Normal distribution of data was confirmed using the Shapiro–Wilk test and one-way ANOVA was used for sample comparisons. Differences were considered statistically significant if the p-value < 0.05.

Assessment of growth promotion capacity

Germinated seeds prepared as described above were transferred into glass tubes containing 15 mL of Jensen's N-free medium (36) solidified with 0.8% (wt/vol) agar. Seedlings were inoculated with 1 mL of a rhizobial suspension containing 1×10^7 cfu. One milliliter of sterile water was added to negative controls. Plants were grown under a photoperiod of 16 h light/8 h darkness at 26 °C. The presence and coloration of root nodules were evaluated periodically. Three months after inoculation, plants were harvested, and plant height was measured and used as a proxy for the plant growth promotion capacity of inoculated strains. The experiment was repeated three times with at least 10 plants per condition. Differences were considered statistically different if the p-value < 0.01 according to the t-test.

Cupriavidus sp. UYMMa02A-Mimosa pudica co-culture assays

A co-culture device was constructed allowing bacterial-plant signal exchange, but avoiding physical contact between both organisms (Fig. 9). Bacterial containers were assembled using 10 cm dialysis membrane tubing pieces (D9652, Sigma Aldrich). Membranes were washed with distillate water and one of their ends was sealed with a double knot, the other end was attached to a 10 cm silicone tubing and sealed with a plastic seal. For a complete and tight seal, a small glass tube was inserted inside the silicone tubing. Bacterial container sterilization was achieved by gamma-irradiation using a 21 kGy dose in the irradiation facilities of the Uruguayan Technological Laboratory (LATU). Test glass jars of 250 mL containing 50 mL of N-free Howieson (Howieson et al. 1993) liquid media diluted 1/10 (v/v) and polypropylene balls as seedling support, were autoclaved for 20 min at 121 °C. After cooling, the jars were opened in the laminar flow hoods, and bacterial containers were aseptically added, submerging the membrane in the media and the silicone tubing facing the jar lids. A total of 50 M. *pudica* germinated seeds were sown in each jar and incubated under a photoperiod of 16 h light/8 h darkness. After 5 days, the bacterial containers were filled through the silicone tubing, with 5 mL of 1×10^8 cfu mL^{-1} of *Cupriavidus* sp. UYMMa02A, and incubated for another five days under the same conditions of temperature and photoperiod. As control treatments, plant-free systems were used. In these cases, Howieson liquid media was supplemented with 2 mM sodium citrate as a carbon source and 2 mM ammonium chloride as a nitrogen source, to allow bacterial survival. Cell suspensions were carefully transferred from the membranes to clean and sterile plastic tubes and centrifuged for 5 min at 6000*g*. Bacterial pellets were used for total protein extraction. A minimum of three biological replicates were performed for each sample.

Total protein extraction and solubilization

For protein extraction, bacterial pellets were washed three times with 2 mL of phosphate-buffered saline (PBS) and resuspended in 10% of the original culture volume in PBS containing 1× complete-EDTA-Free protease inhibitor (Roche). Cell lysis was performed by sonication (7 cycles × 30 s) in continuous mode and at a relative power of 4 alternated with 30 s of rest in an ice-water bath, using an ultrasonic homogenizer (Cole-Parmer Instruments Co.). Cell lysates were separated by centrifugation at 12,000*g* for 15 min at 4 °C and the supernatant containing the total

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soluble proteins was kept at − 20 °C. At least three biological replicates were performed per sample.

Protein quantification

Protein concentration was estimated by the Bradford method (Bradford 1976) using bovine serum albumin (BSA) for calibration curves. The quality of protein fractions was verified by SDS-PAGE (Laemmli 1970).

Protein precipitation

Previous to two-dimensional (2D) electrophoresis, proteins were precipitated, washed, and concentrated, using the commercial kit "2-D Clean Up Kit" (GE Healthcare, Amersham Biosciences) according to the manufacturer's instructions. Finally, proteins were resuspended in a rehydration buffer (8 M urea, 2 M thiourea, 4% CHAPS, 40 mM DTT, 1.2% (v/v) IPG buffer) pH 8.5.

Protein labelling

Thirty-five µg of total proteins from each condition were labeled using the Refraction-2D™ Labelling Kit (NH DyeAGNOSTICS GmbH, Germany), following the manufacturer's recommendations. The samples were normalized and labeled with the Cy3 dye ($\lambda_{ex} = 532$ nm/$\lambda_{em} = 580$ nm) or Cy5 ($\lambda_{ex} = 633$ nm/$\lambda_{em} = 670$ nm), while Cy2 dye ($\lambda_{ex} = 488$ nm/$\lambda_{em} = 520$ nm) was used to label the internal standard which consists of a pooled sample comprising equal amounts (10 µg) of all samples to be compared.

2D-DIGE electrophoresis

The labeled samples were mixed in hydration buffer (8 M urea, 2 M thiourea, 2% (p/v) CHAPS, and 1.2% (v/v) IPG-buffer) and loaded on 24-cm Immobiline DryStrips previously hydrated in the same buffer (nonlinear pH range 3–10, GE Healthcare). Isoelectric focusing (IEF) was run using an IPGphor III apparatus (GE Healthcare). The voltage profile used was adapted to run overnight with the following voltage program: constant phase of 500 V/2 h, constant phase of 2000 V/2 h, a linear increase to 4000 V/2 h, a linear increase to 8000 V/2 h, and a constant final phase at 8000 V for 4 h. After that, the IPG strips were allowed to equilibrate for 10 min with mechanical agitation in 1 mL of equilibration buffer (6 M urea, 50 mM Tris–HCl, pH 8.6, 30% glycerol, 2% sodium dodecyl sulfate) containing 1% DDT for 10 min. Then, reactions were quenched by immersing the strips in SDS-PAGE running buffer (25 mM Tris, 192 mM glycine, 0.1% SDS). As a second dimension, proteins were separated by SDS-PAGE gels (12% acrylamide-bisacrylamide) using an Ettan DALT-Six apparatus Electrophoresis System electrophoresis cell maintained at 20 °C with the Multitemp III cooling unit (GE Healthcare). Each strip was placed on the corresponding acrylamide gel and bound to it, using a running buffer containing 0.2% agarose and 0.002% (w/v) bromophenol blue. The runs were carried out at 100 mA and 80 V for the first 5 min and then the voltage was increased to 120 V until the run front reached the end of the gel.

Image analysis

After 2D-DIGE electrophoresis, gels were fixed with ethanol:acetic acid:H_2O (5:1:4) solution for 30 min. They were then scanned using a Typhoon 9500 FLA scanner (GE Healthcare), using the parameters recommended by the manufacturer, and analyzed using ImageQuant TL v8.1 software (GE Healthcare). The SameSpots software (Totallab, Newcastle, UK) was used to match and analyze protein spots, allowing the detection, normalization with the internal standard, and quantification of the spots. Differential-in-gel analysis was used to calculate protein abundance alterations between samples on the same gel. The resulting spot maps for each biological replicate were then analyzed through biological variation analysis to provide statistical data on the differential protein expression. Spots that exhibited differences in the level of fluorescence with a p-value ≤ 0.05 and a rate of change ≥ 1.25 were considered as differentially regulated and selected for identification.

Spot picking, protein digestion, and MALDI-TOF/TOF protein identification

To identify the differentially expressed proteins, gels were stained using Coomassie Brilliant Blue G-250 (Bio-Rad, Hercules, CA). Spots presenting significant differences were excised from gels, unstained with a solution of 0.1 M NH_4HCO_3 in acetonitrile (ACN) 50% (v/v) and in-gel digested overnight using modified sequencing grade trypsin (Promega, Madison, USA). Peptide extraction was performed as previously described (Gil et al. 2019) and samples were desalted using Zip-Tip C18 reverse phase microcolumns (Millipore, Merck, USA) eluted directly on the plate using matrix solution (α-cyano-4-hydroxycinnamic acid in 60% ACN, 0.1% TFA). The mixture was spotted onto an Opti-TOF plate of 384 positions (Ab Sciex). Spectra acquisition was performed on a MALDI-TOF/TOF MS (4800 Analyzer Abi Sciex) operated in positive reflector mode. The collected MS and MS/MS spectra of selected ions were externally calibrated using a standard peptide mix (Applied Biosystems). Protein identification was carried out using Mascot (Matrix Science, London, UK, (http://www.matrixscience.com) in Sequence query mode, using the genome of *Cupriavidus* sp. UYMMa02A (GCA_001725945.1) or the NCBI NR databases. The

search parameters were: Unrestricted taxonomy; allowable trypsin cleavage jumps = 1; partial modifications: oxidation of methionine and alkylation of cysteine by carbamidomethylation; peptide mass tolerance = 0.05 Da and MS/MS tolerance = 0.45 Da. For protein identification at least one MS/MS spectra per protein was required (with a Mascot peptide ion score $p < 0.05$) and a p-value $p < 0.05$ in the Mascot protein score. Proteins were classified into COGs functional categories and assigned to KEGG pathways using eggNOG-mapper (Huerta-Cepas et al. 2016) and also classified according to their subcellular location with the CELLO v.2.5 (Cheng et al. 2014). The genomic context of the proteins and the location of their gene sequence at the chromosomal level were analyzed by blast searches against the *Cupriavidus* sp. UYMMa02A genome annotated in the RAST server (Aziz et al. 2008).

Results

The genome of Cupriavidus sp. *UYMMa02A encodes a highly conserved symbiotic island*

The draft genome of *Cupriavidus* sp. UYMMa02A was published in 2016 (Iriarte et al. 2016). Genome comparison among beta-rhizobial strains revealed a high level of synteny and nucleotide identity in operons containing the *nod, nif,* and *fix* genes (Fig. 1). However, there were differences in non-symbiotic and transposon-related genes within the symbiotic islands leading to length heterogeneity among them. The symbiotic island of *Cupriavidus* sp. UYMMa02A was the shortest with a predicted length of 33,908 bp.

A closer inspection of the *nod* operon in *Cupriavidus* sp. UYMMa02A indicated that eight *nod* genes, involved in Nod factors biosynthesis and exportation, namely *nodBCIJAH-SUQ,* are arranged in a single operon, while the *nodD* gene encoding a LysR-type transcriptional regulator, was located in the opposite orientation of *nodB* gene (Fig. 2). Moreover, a conserved NodD DNA binding motif known as nod-box (Schlaman et al. 1992) was identified in the intergenic region between *nodD* and *nodB* genes (Fig. 2). Minor differences in length and sequence were observed among the compared sequences, with the symbiotic islands of *Cupriavidus* sp. UYMMa02A and *Cupriavidus necator* UYPR2.512, showing the highest sequence homology.

The UYMMa02A nod operon is not induced by pure flavonoids

The flavonoids luteolin and apigenin are known to induce the expression of *nod* genes in rhizobial *Cupriavidus* sp. strains (Amadou et al. 2008; Marchetti et al. 2010; Rodríguez-Esperón et al. 2022). The observed syntenies lead us to hypothesize that the *nod* genes of *Cupriavidus* sp. UYMMa02A would be regulated by the same mechanism. To analyze this, *Cupriavidus* sp. UYMMa02A was transformed with the pCBM01 plasmid containing a $pnodB_{19424}$-*lacZ* transcriptional fusion and beta-galactosidase (B-gal) activity was compared with *C. necator* UYPR2.512 and *C. taiwanensis* LMG19424, both harboring the same plasmid. As

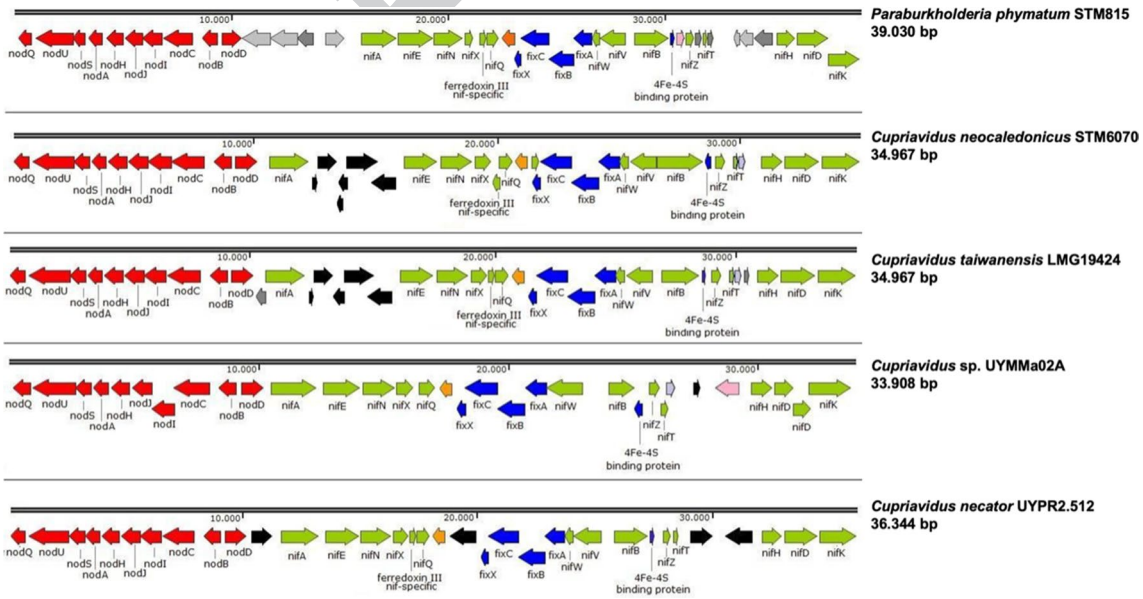

Fig. 1 Comparison of symbiotic islands of different beta-rhizobia. The size of each symbiotic island is indicated below strain names. Genes belonging to the *nod/nif/fix* operons are coloured in red/green/blue respectively, and transposons-related genes are coloured in black. Other colours represent genes that do not belong to the aforementioned operons

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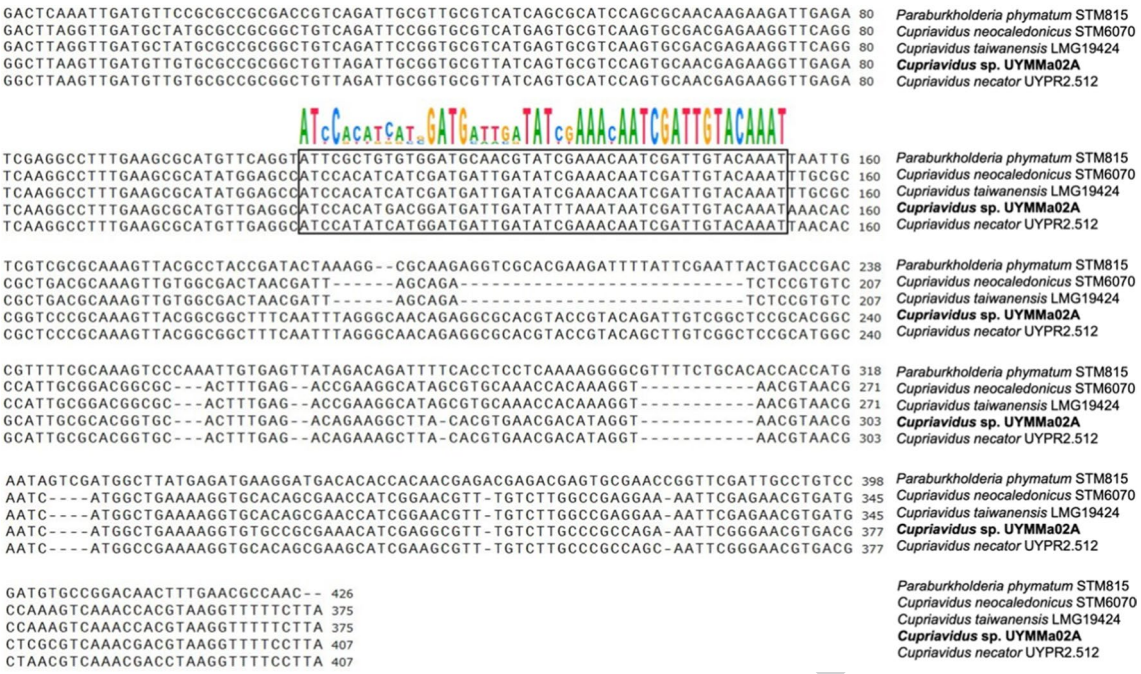

Fig. 2 Sequence alignment of the *nodB-nodD* sequence in different Beta-rhizobia strains. Aligned sequences include the intergenic region between *nodB* and *nodD* and the first 100 bp of both genes. The black rectangle indicates nod-box localization. Conserved motifs are highlighted in colour letters

expected, a strong induction of B-gal activity was observed for *C. necator* UYPR2.512 and *C. taiwanensis* LMG19424 in the presence of luteolin. However, *Cupriavidus* sp. UYMMa02A remained unresponsive to the presence of pure flavonoids (Fig. 3).

Cupriavidus sp. UYMMa02A nod genes are not induced in the presence of *Mimosa* spp. root exudates

Considering that *nod* genes expression is regulated by different compounds in different rhizobial strains (Hungria et al. 1991; Schmidt et al. 1994; Jiménez-Guerrero et al. 2017), we decided to investigate whether *Mimosa pudica* and *Mimosa magentea* root exudates could serve as potential inducers for *nod* genes expression in *Cupriavidus* sp. UYMMa02A. Root exudates are complex solutions containing different

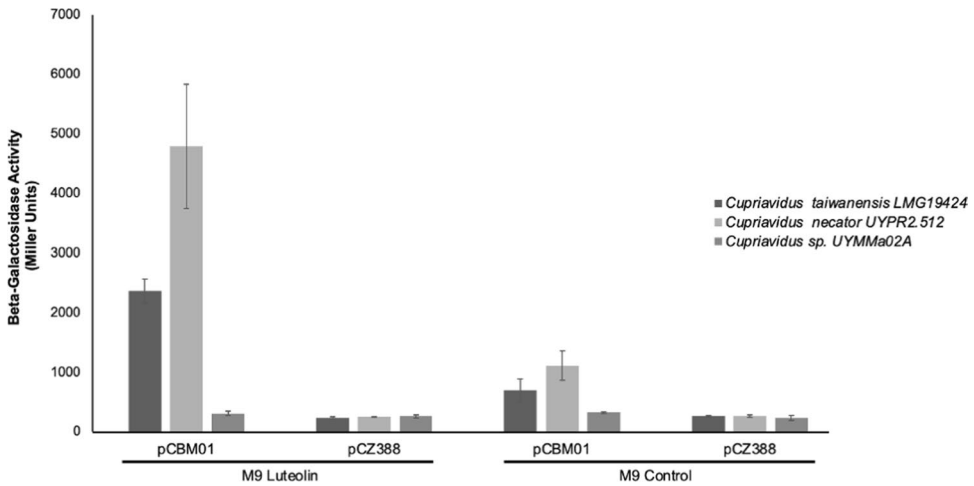

Fig. 3 Expression of the $pnodB_{19424}$-*lacZ* fusion in *Cupriavidus taiwanensis* LMG 19424, *Cupriavidus necator* UYPR2.512, and *Cupriavidus* sp. UYMMa02A in response to luteolin

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amino acids, organic acids, sugars, and phenolic compounds, which have been shown to induce *nod* gene expression in *C. taiwanensis* LMG19424 (Klonowska et al. 2018).

A clear induction of *nod* gene expression was observed, as indicated by an increase in B-gal activity when *C. taiwanensis* LMG19424 and C. necator UYPR2.512 were grown in the presence of *Mimosa* spp. root exudates (Figs. 4 and 5). These results indicate that *Mimosa pudica* and *Mimosa magentea* root exudates contain inducers for *nod* gene expression. However, when *Cupriavidus* sp. UYMMa02A was exposed to these root exudates, we did not observe any change in B-gal activity, suggesting that *nod* genes expression was not induced in this strain (Figs. 4 and 5). Similar results were observed when pure flavonoids were used.

Altogether these findings suggest that the *pnodB*$_{19424}$*-lacZ* is not responsive in *Cupriavidus* sp. UYMMa02A. However, we cannot exclude the possibility that endogenous *nod* genes in *Cupriavidus* sp. UYMMa02A *nod* genes could be induced but were not detected by the assay used. To directly assess this, we analyzed the mRNA levels of *nodA, nodB,* and *nodC* in *Cupriavidus* sp. UYMMa02A in the presence or absence of *M. pudica* root exudates (Fig. 6). No changes in the relative *nod* gene expression were observed when *Cupriavidus*

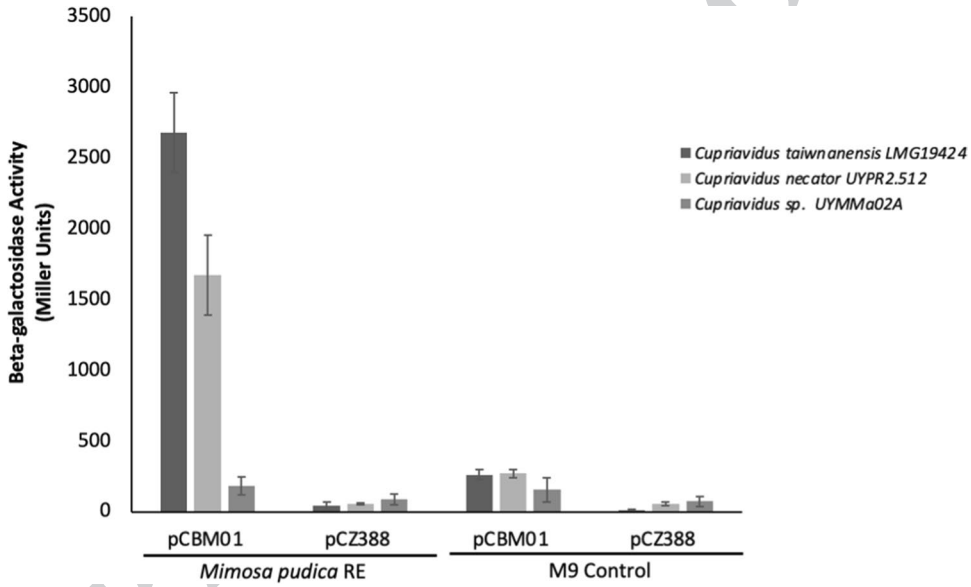

Fig. 4 Expression of the *pnodB*$_{19424}$*-lacZ* fusion in *Cupriavidus taiwanensis* LMG19424, *Cupriavidus necator* UYPR2.512, and *Cupriavidus* sp. UYMMa02A in response to *Mimosa pudica* root exudates (RE)

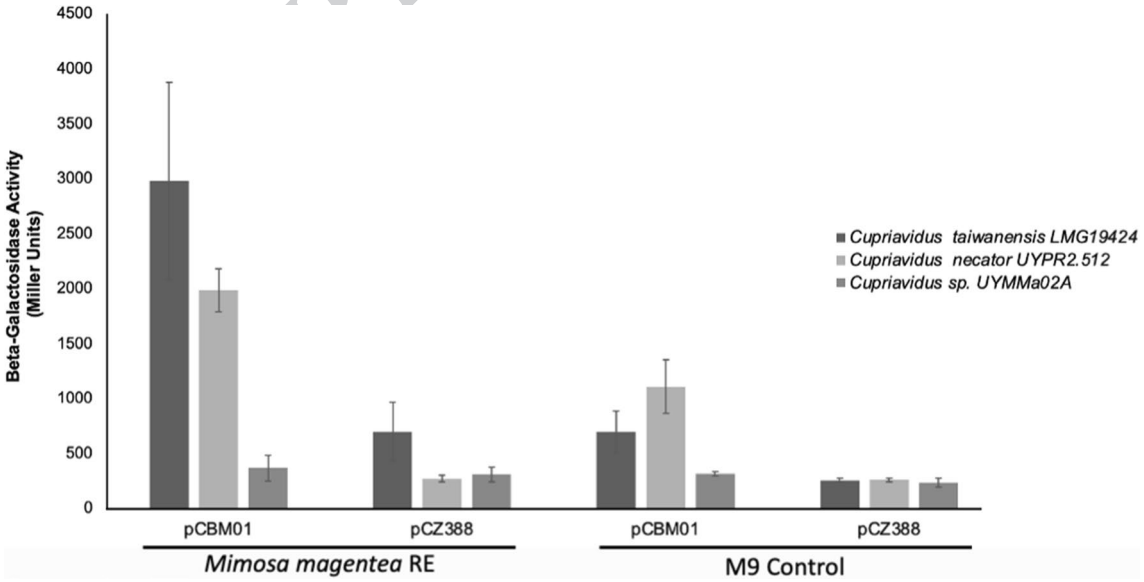

Fig. 5 Expression of the *pnodB*$_{19424}$*-lacZ* fusion in *Cupriavidus taiwanensis* LMG19424, *Cupriavidus necator* UYPR2.512, and *Cupriavidus* sp. UYMMa02A in response to *Mimosa magentea* root exudates (RE)

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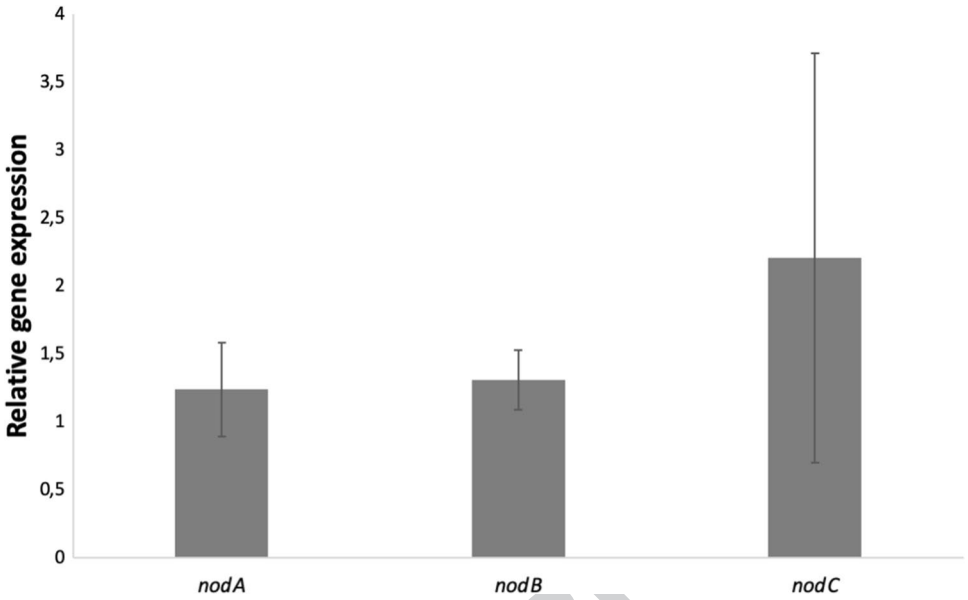

Fig. 6 UYMMa02A *nodA, nodB, and nodC* expression in response to *M. pudica* root exudates. The bars represent the relative expression (Fold Change) of the *nodA, nodB,* and *nodC* genes to the housekeeping genes *elongation factor G (efg)* and *ribosomal protein S14* (S14) when growing in M9 media supplemented with *M. pudica* root exudates versus in M9 media without exudates

sp. UYMMa02A was grown in the presence of *M. pudica* root exudates compared to growth in M9 minimal media. These results confirm the absence of *nod* genes induction in *Cupriavidus* sp. UYMMa02A.

Cupriavidus sp. UYMMa02A induces functional nodules and promotes *M. pudica* plant growth in nitrogen-limiting conditions

We have previously shown that *Cupriavidus* sp. UYMMa02A can form nodules in the roots of several *Mimosa* sp. including *M. pudica* plants (Platero et al. 2016). However, there was no direct evidence of this symbiotic interaction's plant growth promotion ability. When *M. pudica* plants growing in nitrogen-free media were inoculated with *Cupriavidus* sp. UYMMa02A, we observed the formation of reddish nodules in the roots and a significant increase in plant height (Fig. 7), strongly supporting the proficiency of this symbiotic pair.

Proteomic changes induced by luteolin and apigenin in *Cupriavidus* sp. UYMMa02A

The lack of induction of *nod* genes suggests that alternative mechanisms could be involved in the establishment of the *Cupriavidus* sp. UYMMa02A-*M. pudica* symbiotic interaction. To determine the significance of this finding, we decided to employ a proteomic approach known as Differential In Gel Expression (DIGE) (Meleady 2018; Mozejko-Ciesielska and Mostek 2019). In the first approach, bacteria were grown in the presence of luteolin or apigenin, and total protein profiles were compared with bacteria growing in the absence of the flavonoids. Two-dimensional electrophoresis allowed us to separate 373 different spots, representing around 4.5% of the CDS encoded by *Cupriavidus* sp. UYMMa02A genome (Iriarte et al. 2016). DIGE analysis showed 17 spots with altered expression levels in the presence of luteolin while 16 spots were differentially expressed in the presence of apigenin. When we performed a combined analysis of differentially expressed spots in the presence of the flavonoids versus the control condition, a total of 22 differentially expressed spots were detected, 8 overrepresented in the luteolin condition, 8 in the apigenin condition, and 6 in the control condition. After Coomassie blue staining and gel excision, a total of 9 proteins were identified by MALDI-TOF analyses (Table 2). According to COG analyses, 3 of the identified proteins belong to Category E (Amino acid transport and metabolism) and 3 to category C (Energy conversion and production), one protein to Category I (Lipids transport and metabolism), and one belong to category P (inorganic ion transport). Except for the tricarboxylate transporter protein TctC with a periplasmic localization, all the identified proteins were predicted to be cytoplasmic. A model summarizing the major findings using this approach is shown in Fig. 8.

Differential protein expression during *Cupriavidus* sp. UYMMa02A-*Mimosa pudica* co-cultures

The next step in elucidating the mechanisms involved in the plant-bacteria interaction was to determine the proteomic responses of *Cupriavidus* sp. UYMMa02A to the presence of plant host *Mimosa pudica.* We implemented a co-culture device intended to allow plant-bacteria signal interchange, preventing physical contact between symbionts. In this device, plants were grown hydroponically

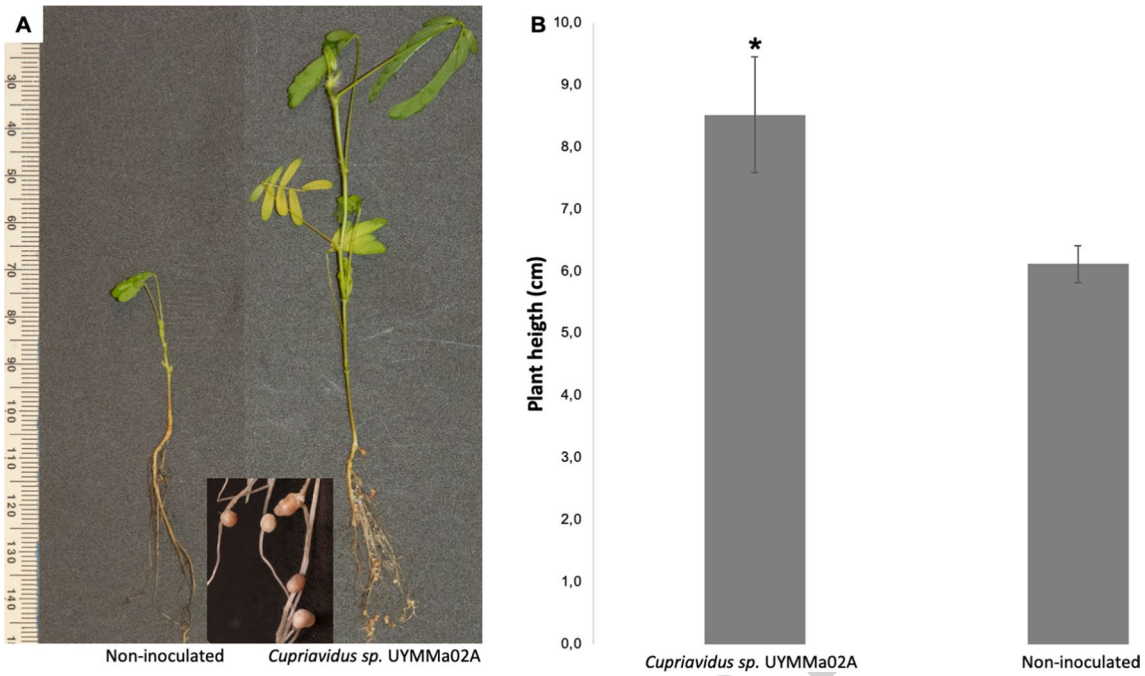

Fig. 7 *Mimosa pudica* growth promotion ability of *Cupriavidus* sp. UYMMa02A. A. Examples of *M. pudica* development phenotype when inoculated with *Cupriavidus* sp. UYMMA02A or not inoculated. Insert, detail of the nodules formed by *Cupriavidus* sp. UYMMa02A in *M. pudica* roots. B. *M. pudica* plant height was recorded 30 days post-inoculation with *Cupriavidus* sp. UYMMa02A or with water as non-inoculated control. Asterisk indicates significant differences (p-value < 0.01) between inoculated plants and non-inoculated plants

with bacteria placed in a closed dialysis membrane submerged in the same hydroponic solution (Fig. 9). Protein profile expression under co-culture conditions was compared with bacterial cells cultivated in the same conditions, but in that case, *Mimosa* plants were not included. The hydroponic solution was supplemented with minimal amounts of carbon and nitrogen sources to allow bacterial survival. Image analyses of the 2D gels captured a total of 674 spots, representing around 8% of the bacterial encode capacity. Thirty-seven differentially expressed spots were determined (26 and 11 spots were overexpressed and repressed respectively). After staining and excision from the gels, 17 different proteins were identified by MALDI-TOF, belonging to 11 different COG categories. Notably, proteins categorized in the categories O (posttranslational modifications and chaperones), G (carbohydrate transport and metabolism), and M (membrane and cell wall) contains both induced and repressed proteins, while proteins in category T (transduction signals mechanisms) were found overexpressed (Table 3). Subcellular localization analysis identified 9 cytoplasmic proteins, 3 periplasmic proteins, and 1 outer membrane protein, while 5 proteins were predicted to be found both at the bacterial cytoplasm and extracellularly. A model integrating the major findings using this approach is presented in Fig. 10.

Discussion

The presence of highly conserved symbiotic islands in the genome of rhizobial *Cupriavidus* spp. suggest the existence of conserved symbiotic mechanisms among these strains. Notwithstanding, gene expression analyses have indicated that pure flavonoids or *Mimosa* spp. root exudates are ineffective in inducing *Cupriavidus* sp. UYMMa02A *nod* genes transcription. This is intriguing since it has been shown that the presence of luteolin or apigenin induces the synthesis and exportation of Nod factors in *C. taiwanensis* LMG19424 (Amadou et al. 2008). In addition, these flavonoids were shown to induce the expression of the $pnodB_{19424}$-*lacZ* transcriptional fusion present in pCBM01, in both *C. taiwanensis* LMG19424 (Marchetti et al. 2010) and *C. necator* UYPR2.512 (Rodríguez-Esperón et al. 2022). It is well known that the flavonoid effect on *nod* genes regulation is dependent on the NodD regulatory protein (Schlaman et al. 1992). Different NodD proteins respond to different flavonoids, being this mechanism one of the bases of the rhizobium-legume specificity (Masson-Boivin et al. 2009). Small differences could be observed at *Cupriavidus* sp. UYMMa02A NodD protein sequence and in the *nodB-nodD* intergenic sequence in

Table 2 *Cupriavidus* sp. UYMMa02A proteins differentially expressed in the presence of apigenin and luteolin

Spot	Fold change	*p* value	Mascot score	Sequence coverage (%)	IP	MW (KDa)	Identified protein		COG category	Localization
							Protein name	ID		
Apigenin induced proteins										
103	1.5	0.019	163	18	5.5	60.8	Bifunctional aspartate transaminase/aspartate 4-decarboxylase	WP_224080690.1	E	Cytoplasmic
272	1.4	0.027	185	38	5.8	27.9	Enoyl-CoA hydratase	ODV41316.1	I	Cytoplasmic
257	1.5	0.004	157	16	9.6	33.4	Tripartite tricarboxylate transporter substrate binding protein	WP_011299555.1	C	Periplasmic
224	1.5	0.052	157	33	5.9	39.4	Branched chain amino acid aminotransferase	ODV41908.1	E	Cytoplasmic
Luteolin induced proteins										
103	1.5	0.050	163	18	6	60.8	Bifunctional aspartate transaminase/aspartate 4-decarboxylase	WP_224080690.1	E	Cytoplasmic
317	1.4	0.021	94	44	6	18.2	DNA starvation/stationary phase protection protein	WP_211952485.1	P	Cytoplasmic
335	3.4	0.001	239	33	6	55	Aldehyde dehydrogenase	ODV42183.1	C	Cytoplasmic
Apigenin repressed proteins										
155	1.4	0.002	143	36	9	32.1	E2 component of the 2-oxoglutarate dehydrogenase complex	WP_211945176.1	C	Cytoplasmic

comparison with *C. taiwanensis* LMG19424 and *C. necator* UYPR2.512. Thus, it is possible that *Cupriavidus* sp. UYMMa02A NodD protein does not become activated by the presence of flavonoids or this protein could not recognize the *C. taiwanensis* LMG19424 *nodB* promoter region present in the $pnodB_{19424}$-*lacZ* used here. To overcome these limitations, we used *M. magentea* and *M. pudica* root exudates for *nod* genes expression analyses, however no changes in the expression levels of *nod* genes in *Cupriavidus* sp. UYMMa02A were observed. These results indicated that in *Cupriavidus* sp. UYMMa02A *nod* genes may not be involved in the first steps of the symbiotic interaction and suggest the existence of alternative mechanisms to Nod factors for bacterial-plant interaction. It is now known that some rhizobial strains are naturally able to engage in symbiotic interactions with their plant host in a Nod-factor-independent way. Researchers have discovered strains of *Bradyrhizobium* sp. lacking the canonical *nodABC* genes that can effectively nodulate *Aeschynomene* spp. (Giraud et al. 2007; Miché et al. 2010) and *Arachis hypogaea* (Guha et al. 2022).

The lack of induction of *nod* genes in *Cupriavidus* sp. UYMMa02A prompted us to search for alternative mechanisms involved in bacterial-plant interaction. To identify proteins potentially implicated in the initial steps of this process, we analyzed the patterns of protein expression of *Cupriavidus* sp. UYMMa02A when exposed to pure flavonoids and during plant co-culture conditions. The response to different *nod* gene-inducing flavonoids and root exudates derived from the plant host has been studied in various rhizobial species, mostly belonging to the alpha-proteobacteria class, using transcriptomic and proteomic approaches (Jiménez-Guerrero et al. 2017; diCenzo et al. 2019). These studies have demonstrated, that in addition to *nod* genes, flavonoids, and root exudates can influence the expression of several bacterial genes encoded by different bacterial replicons. When comparing the responses to flavonoids, and root exudates, there is only partial overlap, particularly in *nod*-related genes, indicating that specific mechanisms are regulated in response to distinct stimuli (Capela et al. 2005). In addition, these responses are often specific to the genus and strains of rhizobia (Fagorzi et al. 2021).

Only two works that used omics approaches have been published for the analysis of rhizobial *Cupriavidus* strains response to flavonoids (Rodríguez-Esperón et al. 2022) or host root exudates (Klonowska et al. 2018). Both works showed the differential expression of hundreds of genes, located at different bacterial replicons, in response to these stimuli. In agreement with this, upon luteolin and apigenin or co-culture conditions, *Cupriavidus* sp. UYMMa02A experiences significant changes at the proteomic level. Approximately 6% (22 out of 357 spots) of the *Cupriavidus* sp. UYMMa02A proteome changed in response to the presence

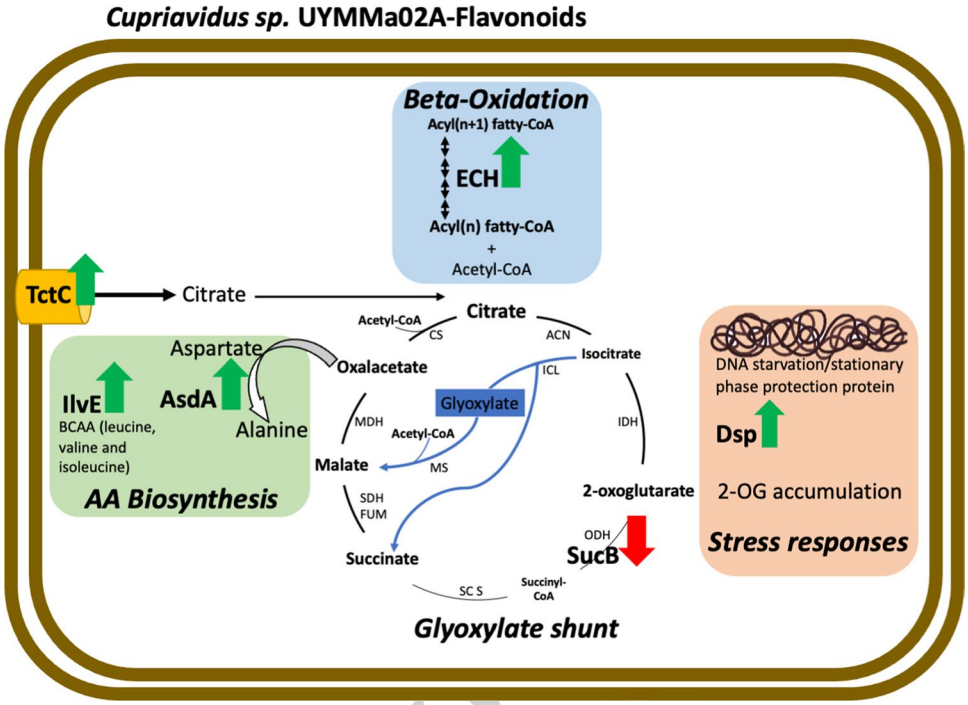

Fig. 8 UYMMa02A proteins and pathways observed in response to flavonoids. Proteins differentially expressed in the presence of apigenin and/or luteolin are highlighted in bold. Green and red arrows indicate if proteins were found either upregulated (up direction) or downregulated (down direction), with respect to the control condition. The main cellular processes affected are indicated in bold and cursive text. See the text for a full description

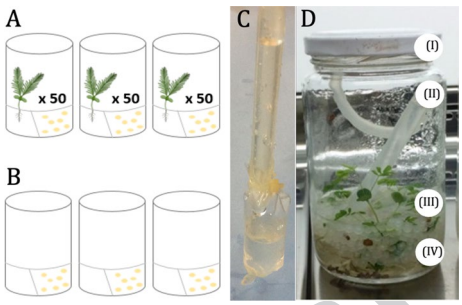

Fig. 9 Plant-microorganism co-culture system. Bacteria were grown in the presence (**A**) or absence (**B**) of *M. pudica* plants. To prevent direct physical interaction between symbionts, the bacteria were placed inside a membrane tubing (**C**). **D** Picture of the real co-culture assembled system (I) jar lid; (II) silicone tube connected to the membrane for bacterial inoculation; (III) *M. pudica* plants; (IV) Mineral plant culture medium containing polypropylene balls for seedling support. As control treatments, plant-free systems were used (**B**)

of flavonoids, suggesting that these molecules are indeed signal molecules for this *Cupriavidus* sp. strain. In agreement with the observed lack of induction of the *pnodB*$_{19424}$-*lacZ* transcriptional fusion, we did not detect the differential expression of proteins belonging to the symbiotic Island. Indeed, most of the affected proteins identified were related to the energy conversion and production (C) and amino acid metabolism (E) COG categories, indicating that flavonoids induce metabolic changes in *Cupriavidus* sp. UYMMa02A. The overexpression of the tricarboxylate transport protein (TctC) could reflect an increase in citrate transport, the only carbon source in the used media. Under these conditions, citrate would be incorporated into the *Cupriavidus* metabolism directly through the citrate cycle (TCA or Krebs cycle), serving as an energy source and a carbon skeleton source. However, the concomitant diminution of the E2 component of the 2-oxoglutarate dehydrogenase complex would slow down the conversion of 2-oxo-glutarate (2-OG) to Succinyl-CoA. This change would result in the accumulation of 2-OG, a key biosynthetic intermediate that connects carbon and nitrogen metabolism. The 2-OG molecule is also involved in modulating enzyme activity and detoxifying reactive oxygen species (ROS) (Huergo and Dixon 2015). Considering this, our result may indicate that some of the metabolic changes observed after exposure to luteolin and apigenin in *Cupriavidus* sp. UYMMa02A could be related to an oxidative stress response. In that sense, Mailloux and collaborators have shown that in response to oxidative stress, *Pseudomonas fluorescence* increased the activity of an NADP-dependent isocitrate dehydrogenase while decreasing the activity of the 2-oxoglutarate dehydrogenase complex, leading to the accumulation of 2-OG (Mailloux et al. 2007). Supporting this hypothesis, we also observed a rise in the DNA starvation/stationary phase protection protein (DpsA), which belongs to the ferritin superfamily and is involved in DNA binding and protection during starvation and/or oxidative stress conditions (Martinez and Kolter 1997; Gambino and Cappitelli 2016; Orban and Finkel 2022). Since luteolin and apigenin have been reported to scavenge ROS (Wu et al. 2015; Salehi et al. 2019; Caporali et al. 2022), it is unlikely that the flavonoids themselves are causing oxidative stress. Instead, the flavonoids may act as signals, advertising that

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Table 3 Proteins differentially expressed during *Cupriavidus* sp. and *Mimosa pudica* co-cultures

Spot	Fold change	*p* value	Mascot score	Sequence coverage (%)	IP	MW (KDa)	Identified protein		COG category	Localization
							Protein name	ID		
Induced proteins										
507	1.5	0.029	147	27	6.1	15.3	Nucleoside-diphosphate kinase Ndk	WP_211957553.1	F	Cytoplasmic/extracellular
629	1.8	0.036	181	40	9.3	32.1	ABC transporter, substrate-binding protein XylF	ODV42702.1	G	Extracellular/cytoplasmic
292	2.4	0.005	109	13	9	42.2	Outer membrane protein Porin	WP_224081450.1	M	Outer membrane
308	2.3	0.041	111	39	5.2	36	Acetoin dehydrogenase E1 component beta-subunit	WP_220630611.1	–	Cytoplasmic
357	1.2	0.026	348	28	5.6	27.5	enoyl-ACP reductase FabI	WP_211945097.1	I	Cytoplasmic/periplasmic
391	1.8	0.034	184	51	6.2	24.1	Peroxiredoxin	WP_211946599.1	O	Periplasmic
499	1.5	0.035	74	12	5.9	16.2	RidA family protein	WP_211958671.1	J	Cytoplasmic
462	1.3	0.041	352	66	5.7	18.1	Cyclophilin/peptidylprolyl isomerase	ODV42271.1	O	Cytoplasmic
199	1.5	0.004	251	37	6.2	43.3	Efflux RND periplasmic adaptor subunit	ODV41844.1	P	Periplasmic
361	1.3	0.007	126	28	7	27.6	Enoyl-CoA hydratase	ODV41316.1	Q	Cytoplasmic
528	1.4	0.036	237	65	6.3	15.2	Universal stress protein UspA	WP_071013637.1	T	Extracellular/cytoplasmic
512	1.3	0.002	156	18	5.2	15.8	Universal stress protein UspA	ODV41046.1	T	Extracellular/cytoplasmic
78	1.7	0.013	216	52	5.5	73.4	Serine protein kinase PrkA	ODV43245.1	T	Cytoplasmic
74	1.7	0.027	130	19	5.5	73.4	Serine protein kinase PrkA	ODV43245.1	T	Cytoplasmic
Repressed proteins										
106	1.3	0.03	163	18	5.5	60.8	Bifunctional aspartate transaminase/aspartate 4-decarboxylase	WP_011298857.1	E	Cytoplasmic
113	2.7	0.01	221	37	8.4	62.5	PPQ-dependent dehydrogenase	WP_224007080.1	G	Periplasmic
625	1.4	0.047	459	42	5.1	57.3	Chaperonin GroEL	WP_035877914.1	O	Cytoplasmic

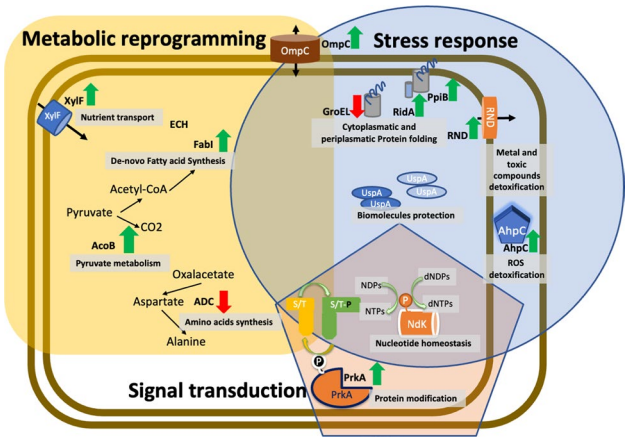

Fig. 10 Model for the proteomics response of *Cupriavidus* sp. UYMMa02A-*Mimosa pudica co-culture* treatment. The illustration shows the main metabolic pathway, biological functions, chemical reactions, enzymes, and other proteins regulated in *Cupriavidus* sp. UYMMa02A grown in a co-culture system with the plant host *M. pudica*. Green and red arrows indicate proteins either upregulated (up direction) or downregulated (down direction) with respect to the control condition. Metabolic pathways affected are shadowed in grey. Color shadows were used to cluster affected proteins according to cellular processes; Metabolic reprogramming (in yellow), Stress response (in blue), and Signal transduction (in orange). Proteins implicated in more than one cellular process have been included at shadow intersections. See the text for a full description

oxidative could occur during the forthcoming interaction with plant roots, as the local production of ROS is a common feature observed during plant-bacteria interactions (Oldroyd et al. 2011; Gourion et al. 2015; Janczarek et al. 2015a, b).

The increased levels of the bifunctional enzyme Aspartate-Transaminase/Aspartate-4-decarboxylase (AsdA) and the Branched chain amino acids aminotransferase (IlvE) in the presence of luteolin and apigenin suggest that flavonoids impact amino acid metabolism in *Cupriavidus* sp. UYMMa02A. AsdA catalyzes the transamination of oxaloacetate to L-aspartate and the subsequent decarboxylation of aspartate to alanine (Kakimoto et al. 1969; Chen et al. 2000). The exportation of alanine and aspartate, through an amino acid cycle between bacteroids and plants, has been postulated to be important for SNF (Prell and Poole 2006; Prell et al. 2009). Besides this, the increase in the IlvE aminotransferase levels indicates the activation of branched-chain amino acids (BCAA) biosynthesis in response to flavonoids. BCAAs have been shown to have an essential role in the formation of effective symbiosis between the beta-rhizobia *Paraburkholderia phyamtum* STM815, and *Cupriavidus taiwanensis* LMG19424 with *Mimosa pudica* plants (Chen et al. 2012). Furthermore, our recent findings have demonstrated that *C. necator* UYPR2.512 induces the expression of genes related to BCAAs transport in response to luteolin (Rodríguez-Esperón et al. 2022). Nonetheless, it should be considered that the statistical support for ilvE protein is borderline, and further experiments will be needed to clarify its role as well as the role of BCAAs in the *Cupriavidus* sp. UYMMa02A-plant interaction.

A third process influenced by the presence of flavonoids in *Cupriavidus* sp. UYMMa02A is the possible activation of the glyoxylate cycle (Kornberg 1966). In this cycle, isocitrate is converted to succinate and glyoxylate by the enzyme isocitrate lyase (ICL) followed by the synthesis of malate from glyoxylate and acetyl-CoA by malate synthase (MS). The glyoxylate cycle enables growth using C_2 compounds by bypassing the CO_2-generating steps of the TCA cycle while generating the necessary products for gluconeogenesis and other biosynthetic purposes (Dunn et al. 2009). Our hypothesis is supported by the following arguments: (a) The activation of this cycle would explain the observed drop in the levels of the E2 component of the 2-oxoglutarate dehydrogenase complex, as this enzyme is by-passed in the cycle (b) The glyoxylate cycle would fulfill the increased demand for 2-oxoglutarate, the substrate of the induced aspartate transaminase/aspartate 4-decarboxylase. Since each turn of the cycle needs two molecules of acetyl-CoA, the third piece of evidence supporting our proposal is the observed increase in the enoyl-CoA hydratase (ECH) levels. This enzyme catalyzes the second step in the beta-oxidation pathways of fatty acid metabolism (Moskowitz and Merrick 1969) and would supply the needed acetyl-CoA molecules to fuel the glyoxylate cycle. While some of the changes induced by the assayed flavonoids in *Cupriavidus* sp. UYMMa02A can be linked to metabolic processes implicated in the symbiotic interaction between rhizobia and host plants, additional experiments should be performed to determine the significance of these findings in interactions involving *Cupriavidus* rhizobial strains.

To improve the simulation of the initial steps of the symbiotic interaction between *Cupriavidus* sp. UYMMa02A and *Mimosa pudica,* we implement a co-culture device. Under these experimental conditions, we observed a broader bacterial response, as reflected by the number and distribution of proteins with altered expression among COG categories. Many of the proteins with changed expression were predicted to be located at periplasmic or extracellular space, suggesting that *Cupriavidus* sp. UYMMa02A undergoes changes at the membrane and periplasmic levels during the co-culture conditions. Surface remodeling has been well documented in many rhizobia models during their interaction with plant hosts. Variations in exopolysaccharides (EPS), lipopolysaccharides (LPS), and capsular polysaccharides (KPS) composition are required for the proper recognition of rhizobial cells by plant hosts, and for evading the natural plant immune system (Janczarek et al. 2015a, b). Under our experimental conditions, we observed the augmented expression of FabI, an enoyl-ACP reductase involved in

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fatty-acid biosynthesis, which could reflect an upregulation of lipogenesis for membrane remodeling. Membranes are the primary barriers for cellular interaction with the environment, influencing the uptake and efflux of small molecules. The observed increase in OmpC levels in *Cupriavidus* sp. UYMMa02A during the co-culture could indicate the need to enhance the transport of nutrients, signal molecules, or other metabolites across the bacterial outer membrane. The concomitant levels rise in the periplasmic binding component of an ABC transport system (XylF) putatively involved in ribose/xylose/arabinose/galactose acquisition, suggests an enhanced transport capacity for these sugars. Xylose and ribose have been detected in *M. pudica* root exudate (Klonowska et al. 2018), implying that these sugars may support *Cupriavidus* sp. UYMMa02A metabolism in the rhizosphere of *M. pudica*. In the other hand, we also observed an increase in the levels of a protein predicted to encode the periplasmic adaptor subunit of the resistance-nodule-division (RND) family efflux transporter. In gram-negative bacteria, RND efflux systems work in concert with outer membrane porins, like OmpC, to detoxify deleterious compounds (Nies 2003; Klonowska et al. 2020). The importance of RND systems has been demonstrated in many rhizobial species, where they play a crucial role in survival and competence in the rhizosphere, affecting nodule formation and nitrogen fixation (Klonowska et al. 2012; Santos et al. 2014). Moreover, as observed here, some of these systems were shown to be inducible both by flavonoids and plant host root exudates. Similarly, Klonowska and collaborators found that *Cupriavidus taiwanensis* LMG19424 and *Paraburkholderia phymatum* STM815 RND systems are induced by *M. pudica* root exudates (Klonowska et al. 2018). Our results suggest that this system would also be important for the growth of *Cupriavidus* sp. UYMMa02A in the presence of *M. pudica* root exudates. As mentioned, one role for these efflux systems is the detoxification of harmful compounds. In that sense, the RAST annotation server indicates that the identified protein would be a membrane fusion protein of the CzcB family, which is part of the CzcCBA Heavy metal efflux (HME) RND system (Janssen et al. 2010). CzcCBA systems were first described in *C. metallidurans* CH34 and shown to confer resistance to Co, Zn, and Cd. Our evidence suggests that *Cupriavidus* sp. UYMMa02A activates the extrusion of metals from its cytoplasm, perhaps preventing the formation of reactive oxygen species (ROS). The increase in *Cupriavidus* sp. UYMMa02A peroxiredoxin levels support the hypothesis that ROS are formed during this interaction. Peroxiredoxin is a periplasmic protein implicated in the detoxification of ROS and peroxynitrite, also detected in *Rhizobium leguminosarum*, *C. taiwanensis,* and *P. phymatum* rhizobial strains growing in the presence of root exudates (Ramachandran et al. 2011; Klonowska et al. 2018). Another stress-related induced protein is cyclophilin. Cyclophilins are Peptidyl-prolyl cis/trans isomerases (PPiase, EC: 5.2.1.8) enzymes found in all kingdoms of life. Since these proteins catalyze a rate-limiting step in protein folding, they play a critical role in protein homeostasis. Diverse studies have demonstrated the participation of these proteins in signal transduction, biofilm formation, motility, and adaptation to stress (Skagia et al. 2016; Dimou et al. 2017; Thomloudi et al. 2017). Thomloudi and collaborators showed that the heterologous expression of the two cyclophilins isoforms of *Sinorhizobium meliloti* 1021 in *E. coli* enhance bacterial survival under stress condition, supporting the role of these proteins in stress adaptation in bacteria. In addition, a PPiase mutant in *Azorhizobium caulinodans* ORS571 impairs its symbiosis with *Sesbania rostrata,* reducing nodule size and completely abolishing its nitrogen fixation ability (Suzuki et al. 2007). Along with the increase in the protein levels of the Efflux RND periplasmic adaptor subunit, peroxiredoxin, and PPiase, we also observed an increase in the universal stress protein A (UspA) levels. Universal stress proteins are small cytoplasmic bacterial proteins whose expression is enhanced when the cell is exposed to diverse stress agents and has been implicated in cell survival during prolonged exposure to stress conditions (Nyström and Neidhardt 1994). Induced expression of genes encoding proteins of this family has been observed during *Rhizobium leguminosarum* growing in the rhizosphere of pea, alfalfa, and sugar beet (Ramachandran et al. 2011), as well as in *C. taiwanensis and P. phymatum* growing in the presence of *M. pudica* root exudates (Klonowska et al. 2018). Altogether, the evidence presented here strongly suggests that during the first steps of the symbiosis, *Cupriavidus* sp. UYMMa02A is under considerable stress pressure, a common feature in rhizobia-legumes symbiosis.

Finally, we were able to detect the induction of proteins with the potential to transduce host-secreted signals, such as the protein kinase PrkA protein and the Nucleotide dikinase (NdK) protein. PrkA is a member of the conserved serine/threonine protein kinase family widely distributed among Bacteria, Archaea, and Eukarya (Stancik et al. 2018). Serine/Threonine kinases (STKs) act in concert with Serine/Threonine phosphatase (STPs) to introduce or remove phosphate modifications in proteins at these residues. Reversible protein phosphorylation is one of the main mechanisms allowing bacteria to respond to environmental stimuli. Post-translational phosphorylation/dephosphorylation influences the activity of modified proteins by inducing conformational changes and regulating protein–protein interactions. Thus, STKs and STPs influence many different signal transduction pathways in bacteria. In *Rhizobium leguminosarum,* disruption of the *pssZ* gene, encoding an STK, impacts exopolysaccharide synthesis, surface properties, and symbiosis with clover (Lipa et al. 2018). Despite this example, very few studies address the role of STKs in the symbiosis between

rhizobia and legumes. The induction of the *Cupriavidus* sp. UYMMa02A PrkA in the presence of the host plant *Mimosa pudica,* suggests that this STK could have an important role during the initial steps of the symbiotic interaction and demands additional studies to determine its precise role.

On the other hand, Ndks are housekeeping enzymes whose primary role is maintaining the cellular homeostasis of nucleoside triphosphate (NTP) and their deoxy derivatives (dNTPs) pools by catalyzing the reversible γ-phosphate transfer from NTPs (or dNTPs) to NDPs (or dNDPs) (Berg and Joklik 1953). In bacteria, the transfer of high-energy phosphates occurs by a ping-pong mechanism that involves the formation of a phosphor-histidine intermediate at a conserved histidine residue (Lascu and Gonin 2000). Moreover, it has been shown that His-Phosphorylated Ndks can phosphorylate other proteins at histidine residues modulating their activities, and affecting a myriad of bacterial processes (Lu et al. 1996; Attwood and Wieland 2015). For example, the *E. coli* Ndk can transfer its phosphate to the sensor proteins EnvZ and CheA, and then the phosphorylated kinases transfer the high-energy phosphate to their cognate response regulators OmpR and CheY, respectively, which are implicated in osmosis and chemical sensing (Lu et al. 1996). In other bacteria Ndk homologs have been implicated in the regulation of biofilms formation, the function of type 3 secretion systems, the modulation of quorum sensing, and the response to oxidative stress, playing a critical role in bacteria and host interactions (Yu et al. 2017). Interestingly, in some bacteria, Ndk can also be found extracellular and membrane-associated. In *P. aeruginosa,* it has been observed that membrane-associated Ndk correlates with the status of bacterial growth (Shankar et al. 1996). Analysis of the *Cupriavidus* sp. UYMMa02A Ndk sequence suggests that this protein could have both intracellular and periplasmic locations. The results presented here, suggest that this protein would be involved in the interaction between the rhizobial *Cupriavidus* sp. UYMMa02A and the plant host *Mimosa pudica.* Further experiments are needed to determine the exact role of this protein in the interaction, as the role of Ndks in beneficial plant-bacteria interaction has not been studied in detail.

Conclusions

The genome of *Cupriavidus* sp. strain UYMMa02A encodes a conserved symbiotic island that includes a complete *nodDBCIJHASUQ* gene cluster with a nod-box sequence in the promoter region of *nodB*. However, no induction of the *Cupriavidus* sp. UYMMa02A *nod* genes was evidenced in response to pure flavonoids or *Mimosa* spp. root exudates. These results suggest that *Cupriavidus* sp. UYMMA02A may employ Nod-independent mechanisms to stablish symbiosis. Using a quantitative proteomic approach, we detected significant proteomic changes in *Cupriavidus* sp. UYMMa02A when exposed to flavonoids or root exudates. Twenty-four differentially expressed proteins were identified covering diverse bacterial processes ranging from basic metabolism and transport functions to stress response and signal transduction. In the presence of the plant host, the major bacterial responses were related to amino acid metabolism and oxidative stress, which are common features in many rhizobia-legume interactions. Besides this, the increased levels of the proteins PrkA and Ndk proteins during the co-culture conditions indicated the involvement of these versatile proteins in the symbiotic interaction. While further research is necessary to determine the precise roles of the identified proteins, our study presents a new model for interrogating the symbiotic interaction between beta-rhizobia and plant hosts.

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