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The current landscape of the antimicrobial peptide melittin and its therapeutic potential

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Melittin, a main component of bee venom, is a cationic amphiphilic peptide with a linear α -helix structure. It has been reported that melittin can exert pharmacological effects, such as antitumor, antiviral and anti-inflammatory effects in vitro and in vivo. In particular, melittin may be beneficial for the treatment of diseases for which no specific clinical therapeutic agents exist. Melittin can effectively enhance the therapeutic properties of some first-line drugs. Elucidating the mechanism underlying melittin-mediated biological function can provide valuable insights for the application of melittin in disease intervention. However, in melittin, the positively charged amino acids enables it to directly punching holes in cell membranes. The hemolysis in red cells and the cytotoxicity triggered by melittin limit its applications. Melittin-based nanomodification, immuno-conjugation, structural regulation and gene technology strategies have been demonstrated to enhance the specificity, reduce the cytotoxicity and limit the off-target cytolysis of melittin, which suggests the potential of melittin to be used clinically. This article summarizes research progress on antiviral, antitumor and anti-inflammatory properties of melittin, and discusses the strategies of melittin-modification for its future potential clinical applications in preventing drug resistance, enhancing the selectivity to target cells and alleviating cytotoxic effects to normal cells.

KEYWORDS

melittin, pharmacological effect, nanomodification, immuno-conjugation, structural regulation, gene technology strategies

Introduction

Antimicrobial peptides have attracted increasing attention because of their potential benefits in agriculture, the food industry and medicine (1, 2). Antimicrobial peptides, naturally produced bioactive small proteins composed of a few amino acids by all living organisms, can protect against fungi, viruses and bacteria (2, 3). Melittin, the principal component of venom derived from honeybee *Apis mellifera*, is a promising antimicrobial peptide. Melittin is rich in *Apis mellifera* venom and can also be artificially synthesized. Melittin ($C_{131}H_{229}N_{39}O_{31}$) is a cationic short peptide weighting 2840 Da that consists of 26

amino acid residues (GIGAVLKVLTTGLPALISWIKRKRQQ-CONH2) (4) with 4 positive charges in the N-terminal portion and 2 positive charges in the C-terminal region (5). Due to the uneven distribution of polar and the nonpolar amino acid residues, melittin exhibits hydrophilic and lipophilic amphipathic characteristics. The amino terminal region (residues 1-20) plays a hydrophobic role, whereas the carboxyl terminal region performs a hydrophilic role because of a positively charged amino acid (6) in melittin. Each melittin chain is exhibited the structure of a helixhinge-helix motif or bent rod. Amino acid residues 3-10 constitute an α -helix, and residues 12 to 14 form a hinge region, which is joined by an α -helix containing residues 15-24 (7, 8). Four-identical monomers of melittin compose a tetramer, which is the predominance in the abdominal sack of bee (7). When the venom is released, the tetramer is dissociated, yielding the monomer (9, 10). Melittin monomers can attach to the membrane surface, spontaneously bind to natural and artificial membranes (11) by inserting into phospholipid bilayers so that the strict of polar and non-polar moieties across the bilayer are weakened, and the permeability barrier is reduced. Based on the single-molecule dynamics perspective, Xu and colleagues (12) showed that the changes of membranes structure occurred when melittin was adsorbed and accumulated on the surface of cells membrane, which markedly accelerating the lateral diffusion of lipids around the melittin, accordingly, the diffusion facilitated melittin insert in such heterogeneous regions. The inserting process of melittin forms a "U"-bending pathway, takes shapes of toroidal pores with dynamical translocations of lipids and melittin in the lipid bilayer. Dye leakage and quartz crystal microbalance fingerprinting analysis showed that the disrupting action of melittin to membrane were differences response to different types of membrane: melittin-mediated surface-acting mechanism in lipid mixtures (bacterial-mimetic), penetrating the bilayer (mammalianmimetic) and preferring for the domain containing predominantly zwitterionic lipids (domain-forming mixed membranes). It is proposed that melittin-induced membrane permeabilization may be due to toroidal pore or fissure formation in the membrane (13)(Figure 1). Melittin can be applied to monitor lipid-protein interactions in membranes (5, 14). Interestingly, electrochemiluminescence (ECL) imaging efficiently characterize the instant release of vesicle content via melittin-triggered permeabilization (15). Melittin is also applied for the studies related to calmodulin (CaM), a sensor protein of calcium ions, which is regarded as hub protein to bind and modulate more than 200 target proteins in a concentration-dependent manner of calcium ions. Recently, it is reported that the interaction of



melittin and CaM is through melittin anchoring at the hydrophobic pockets of CaM by different sets of residues responding to different conformers (16). The multiple binding modes of melittin-CaM suggest the multiple target potential. Melittin can exert functions through nonmembrane-breaking pathways. A series of studies (17-20) on cardiovascular system diseases and neuropathic pain have reported that melittin exerts antiviral, anti-inflammatory and immune regulatory effects. Melittin could mitigate Staphylococcus aureus-triggered skin infection in CD1 mice and exerted protective effects in a model of methicillin-resistant S. aureus (21), which is significantly difficult to treat with available antibiotics (22). CD1 mice is widely used in the fields of pharmacology, toxicology and etc. because there are less differences in the growth and reproductive performance between different closed colonies raised in different areas of CD1 mice (23), which provides support for the applicable potential of melittin. Melittin inhibits the generation of inflammatory mediators and alleviates the inflammatory reaction in animal models with acute edema and chronic arthritis (24). It is worth noting that melittin can alleviate chemotherapy-induced peripheral neuropathy and ameliorate neuropathic pain in a rodent model (25). The Food and Drug Administration (FDA) approved melittin for relieving pain and swelling triggered by bursitis, tendinitis, rheumatoid arthritis and multiple sclerosis (26, 27). Melittin could hamper the proliferation and growth of human monocytic leukemia cells (28-30). Ceremuga and colleagues observed that melittin could induce the apoptosis of acute lymphoblastic leukemia and chronic myelogenous leukemia cells (31). Melittin suppresses the metastasis of hepatocellular carcinoma tumors (32), inhibits tumor growth in Colon26 NL-17-bearing mice in vivo (33) and exhibits synergistic effects with epirubicin in suppressing colon cancer in vitro (34). The clinically applicable potential of melittin and its functional mechanism are popular issues in the field of antimicrobial peptide research. Given that melittin possesses multi-target mechanisms of biological activity, melittin is considered an attractive candidate for therapeutic diseases in clinic. Therefore, this systematic review aims to summarize some of the important aspects related to melittinmediated antiviral, tumoricidal and anti-inflammatory action. The article also discusses the different modification on melittin such as nanocarrier, immuno-conjugation, peptide-structural regulation and gene technology strategies for preventing the severe hemolytic activity and facilitating therapeutic targeting.

Antiviral properties of melittin

Viral infections can be highly contagious and can cause serious harm to the body, which is reflected in the high fatality rate of many viral infections. Antiviral drugs mainly focus on disrupting the virus replication cycle. However, viruses proliferate quickly and are prone to mutation and drug resistance. The available drugs for targeted treatment are relatively limited. Seeking effective strategies to rapidly inhibit viral replication and prevent or delay drug resistance has attracted increasing attention in the exploration of antiviral agents.

A series of studies have shown the antiviral properties of melittin against herpes simplex virus (HSV) (35-39). HSV mainly attacks the skin, mucous membranes and nerve tissues. Human infection with HSV is relatively common. The sites of viral invasion and lesions vary. Neutralizing antibodies appear in the blood approximately one week after infection, reaching a peak in 3 to 4 weeks and lasting for many years. It is not clear if the protection of neutralization is sufficient although the vaccine elicited neutralizing titers and antibodies to HSV (40). One month after the final administration of HSV-2 gD2 vaccine, mean neutralizing titers against HSV were well-below than those of natural infection (41). Currently, there is no specific drug to cure HSV infections, and treatment strategies mainly include shortening the time of lesion healing or episode duration (42), preventing secondary infections and reducing recurrence. Baghian and colleagues have reported that melittin can impede cell fusion triggered by HSV-1 one of nine herpes viruses that infect humans. The researchers observed that melittin inhibited the attachment and penetration of HSV-1 to the membrane of host's cell, which are major steps of replication cycle in infecting humans of viruses (43). The enveloped viruses attach to the plasma membranes and form close apposition and then release the viral nucleocapsid into cytosol via two-membrane fusion. Membrane microenvironment affects the entry of enveloped viruses (44). Melittin suppressed the activity of the Na⁺ and K⁺ pumps in infected cells (36), which may affect biological processes, such as metabolic reactions, transmembrane transport, adenosine triphosphate (ATP) synthesis and membrane fusion processes, indicating melittin-triggered inhibition of HSV-1 infection. Melittin suppresses HSV-1-triggered fusion of cells by suppressing HSV-1 glycoprotein K, which is necessary for efficiently encapsulating and translocating infectious virions from cytosol to extracellular fluid (45). Melittin exhibited virucidal activity when coincubated with HSV in African green monkey kidney-derived Vero cells (37), which was probably attributed to the direct interaction between melittin and the enveloped virus surface, destabilizing the membrane structure of the virus and inactivating the viral particles (38). Moreover, Albiol Matanic and colleagues showed that melittin impeded HSV 1 and HSV 2 infection but was highly toxic to host cells by osmotic lysis. Melittin can quickly integrate to lipid bilayers, oligomerize and form pores (39). Melittin can induce cell apoptosis (46) and necrosis (47, 48). Discovering strategies that reduce the toxicity of melittin is valuable and necessary for the feasibility of its clinical application.

Human immunodeficiency virus (HIV) leads to the severe impairment of immune function. Patients often suffer from severe opportunistic infections. HIV destroys the human antiviral defense system and promotes the development of cancer. The virus may carry oncogenes that can then be integrated into the DNA of host cells, causing the cells to undergo cancerous transformation, especially when the immune system is suppressed and immune surveillance is lost (49). Anti-HIV management is an important topic in the field of clinical treatment and medical research. It is worth noting that Gag, the structural protein of HIV-1, plays a decisive and specific role in the replication process of HIV-1 apart from being a scaffolding protein (50). Gag coordinates specific

encapsulation of genomic RNA and motivates the formation of viral particles through its automatic assembly. Gag binds to a variety of viral proteins and interacts with intracellular proteins that are needed for its function and then translocates from its translation site to the plasma membrane, allowing the novel virions to be released. Therefore, Gag is considered a target for anti-HIV intervention. Wachinger and colleagues observed that melittin reduced HIV-1 production by affecting the processing of the Gag precursor protein (51). Moreover, melittin significantly reduced HIV-1 replication by damaging HIV proteins in a dose-dependent manner. Wachinger and colleagues obsderved that melittin suppressed viral replication in acutely HIV 1-infected human T lymphoma KE37/1 cells. Under melittin treatment, the levels of intracellular Gag and viral mRNAs were reduced, suggesting that melittin-mediated inhibition of HIV-1 replication occurred through interference with the host cell-directed gene expression of the virus (52). Considering the cytotoxicity of melittin, Hood and colleagues developed lipid film-based nanoparticles loaded with mellitin and investigated the effects of the nanoparticles on HIV-1 infectivity and biosafety. The researcher observed that melittin nanoparticles could capture more HIV-1 and significantly inhibit HIV-1 infection without presenting significant cytotoxicity (53). Interestingly, Jallouk and colleagues enveloped melittin into a lipid shellperfluorocarbon nanoparticle to prevent its cytotoxic effects and enhance its therapeutic efficacy (54). The researchers showed that compared with free melittin, melittin-loaded nanoparticles reduced HIV infection and exhibited less toxicity on sperm and vaginal epithelial cells.

Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) is a crown-like envelope RNA virus with a diameter of 120-160 nm. Patients suffering from SARS-CoV-2 exhibits clinical symptoms, such as ever, cough and fatigue, and showing groundglass lesions in both lungs determined by Computed Tomography imaging. A few patients with severe SARS-CoV-2 infections present cytokine storm syndrome and hyperinflammation. Some cases even develop metabolic acidosis, coagulopathy and multiorgan failure (55, 56). Based on virus biology and pathology, researchers have proposed antiviral agents and immunomodulatory drugs to mitigate the severity of SARS-CoV-2 infection. The SARS-CoV-2 spike protein directly interacts with angiotensin-converting enzyme 2 (ACE-2) receptors in the body, initiating viral replication (57, 58). it is considered a strategy to disrupt the entry of the virus into cells and propagation by preventing the binding of the spike protein with ACE2. Enayathullah and colleagues observed that mellitin treatment could protect Vero cells against SARS-CoV-2 infection (59). The levels of the receptor binding domain (RBD) of the SARS-CoV-2 spike protein were reduced under melatonin treatment in SARS-CoV-2-infected Vero cells. Interestingly, molecular docking assays showed that melittin can interact with the RBD of the SARS-CoV-2 spike protein, suggesting melittin can reduce spike protein-ACE2 interactions. Moreover, the main protease Mpro (3CLpro) performs an important function in SARS-CoV-2 virus replication. Coronaviruses hijack the translational machinery of the host, generating viral proteins. The viral proteins need to be cleaved so that the individual functional proteins can be released for viral replication and transcription. Mpro cleaved viral proteins, facilitating the assembly of viral replicase complex (60). Mpro is considered a viable target for anti-SARS-CoV-2 virus intervention (61). Al-Rabia and colleagues developed a complex using sitagliptin (SIT) and melittin (MEL) to investigate the antiviral effects (62). The researchers demonstrated that the cellular uptake of SIT-MEL was significant in Vero E6 cells infected by SARS-CoV-2 virus. Moreover, Mpro-mediated inhibition was markedly higher with SIT-MEL complex treatment than under SIT or MEL treatment conditions. However, antimicrobial peptides have other limitations in addition to cytotoxicity, such as being easily hydrolyzed and eliminated from the bloodstream or causing allergic responses (63).

The anti-influenza A efficacy of melittin have been reported (37). Uddin and colleagues showed that melittin could markdely suppress the replication of influenza A virus in Madin-Daby canine kidney cells in vitro. Additionally, melittin treatment also exhibited protective effects against influenza A virus subtype H1N1-infected C57BL/6 mice in vivo. The mice suffering from H1N1 infection presented clinical signs of respiratory disease, such as ruffled fur, decreased activity, hunched posture and huddling, whereas melittin-treated mice did not show the above signs. Moreover, mice in the melittin treatment group showed lower viral titers in the lung than H1N1-infected mice without melittin treatment. Compared with HSV1 and HSV2, Junin virus (JV) is more sensitive to the inhibitory effects of melittin (64). JV is the etiologic agent of Argentine hemorrhagic fever (AHF). Calomys musculinus is the natural host of JV. JV can be transmitted to humans via the inhalation of aerosolized secretions and excretions of rodents (65). The onset of AHF is slow, and there are no specific symptoms during the early onset of AHF, which may delay the diagnosis. Accompanied by the debilitating hemorrhagic phase, the mortality rate of untreated AHF is 20% to 30% (66). Argentine researchers have been working to identify an effective treatment for AHF. Albiol Matanic and colleagues (64) showed that at a noncytotoxic concentration of 3 µM, melittin could reduce the virus infectivity of HSV-1, HSV-2 and JV by 80%, 80% and 99%, respectively. Melittin also exerts antiviral properties against enterovirus 71 (EV-71) (37). EV-71 is one of the major causative agents that causes hand, foot and mouth disease in young children, which leads to neurological, cardiac and respiratory complications (67). Viral infection with EV-71 is a serious threat to children's health. Melittin treatment could reduce EV-71 replication and ameliorate EV-71 infection-mediated cytopathic effects in HeLa cells in vitro. The antiviral activity of melittin against several kinds of virus associated with human diseases are shown in Supplementary Table 1.

Antitumor potential of melittin

Cancer is a global health problem and is one of the leading causes of death in the world. In recent years, substantial progress has been made in cancer diagnosis and treatment in recent years. However, traditional anticancer treatments still has severe side effects and toxicity and patients still exhibit drug resistance. It is crucial to identify new and effective anticancer treatments with low cytotoxicity. The low toxicity and tumor targeting abilities of antimicrobial peptides suggest their potential as antitumor agents. The antitumor effect of melittin is a popular topic (Supplementary Table 2). In addition to directly punching holes in tumor cell membranes, melittin can modulate the cell cycle, proliferation, angiogenesis metastasis and apoptosis of cancer cells (68).

Colon cancer is the second leading cause of cancer-related deaths worldwide (69). Main strategies for colon cancer treatment are surgery and chemotherapy. The critical problems of chemotherapy are cytotoxicity, nonspecific targeting, and drug resistance. Yaacoub and colleague evaluated the anticancer effects of melittin on a human colon carcinoma HCT116 cells (70). They observed that melittin had inhibitory efficacy on HCT116 cells in a dose-dependent manner, and there were significant synergistic effects between melittin and another main biopeptide of bee venom, phospholipase A2 (PLA2), on the inhibition of HCT116 cells. They speculated that melittin could destroy the glycocalyx of the cell coat on the cell membrane, which facilitated PLA2 directly acting on phospholipids of lipid membrane bilayer, probably causing the necrosis of HCT116 cells. Alfaleh and colleagues enveloped melittin with a fluvastatin nanoparticulate system containing phospholipid and alpha lipoic acid to assess the effects of the nanoparticles against colon cancer. The researchers reported that compared with monotherapy treatments, the hybrid system exhibited more significant inhibition of colon cancer Caco2 cell (71), and the hybrid nanosystem treatment arrested Caco2 cells in G2/M and pre-G1 phases (72). Additionally, melittin-mediated fast action on anticancer effects was reported in the cell lines of colorectal and gastric cancer in vitro. The researchers showed that melittin exhibited anti-tumor effects in a dose-dependent manner in the COLO205 cells (human colon cancer cell lines), HCT-15 cells (human colorectal adenocarcinoma cells). The morphological changes in above cell lines were evident within 30 seconds after melittin addition. Changes in the membrane were gradually observed, such as swelling, breakage or blebbing, and intracellular material was expelled from the cells (73). Melittin inhibited the growth and metastasis in MC38 cells (a murine colon cell line), and increased inflammatory cytokine levels in cancer cells (74). Luo and colleagues observed that melittin-mediated endoplasmic reticulum (ER) stress led to a disruption of calcium homeostasis and facilitated apoptosis in the human colorectal cancer cell line SW480 in vitro. Accordingly, the researchers showed that melittin management impeded tumor growth by triggering ER stress in SW480 tumorbearing mice. No other marked biochemical and hematological toxicity to the mouse bodies were observed (75). The effects of melittin on the bone metastasis in colorectal cancer were assessed (76). A single dose of melittin was given to a mouse model of bone metastasis in human colorectal cancer (bearing HT-29 cells). Melittin treatment could inhibit the growth of metastases. Histological assays present necrosis and inflammatory morphology in melittin-treated metastases, suggesting the potential of melittin to reduce the growth of metastases in colorectal cancer patients.

Melittin treatment also impeded the growth of human gastric cancer (73). Melittin exhibited antitumor effects by inducing apoptosis in SGC-7901 cells (77). Melittin elevated the membrane permeability of mitochondria, reduced mitochondrial membrane

potential and increased reactive oxygen species (ROS) release in SGC-7901 cells. Accordingly, melittin-induced increases in cytochrome C, apoptosis-inducing factor (AIF) and endonuclease G levels were consistent with the downregulation in second mitochondria-derived activator of caspases (Smac)/direct inhibitor of apoptosis gene (IAP) binding protein, activating caspase-3.

Drug resistance is still a critical problem for lung cancer therapy. Cellular mechanisms are related to resistance. Moreover, the tumor microenvironment (TME) is an important factor in the resistance of tumor cells to radiotherapy and chemotherapy (78). The TME is composed of immune cells and mesenchymal cells (79). A critical mechanism in the TME is the activation and infiltration of circulating monocytes, which differentiates into tumor-associated macrophages (TAMs). It is considered that TAMs may be involved in the regulation of tumor growth, infiltration and metastasis. Melittin arrested the cell cycle at G1 phase in a lung cancer cell line and induced apoptosis. Importantly, melittin could suppress the differentiation of circulating monocyte cells into TAMs (80), suggesting the antitumorigenic potential of melittin against lung cancer.

Melittin-mediated inhibition on the tumor invasion of breast cancer have been reported (81, 82). Breast cancer is the second most lethal cancer among women. The invasion and metastasis of cancer cells are the main causes of high mortality in patients suffered from breast cancer (83). The treatment strategies for breast cancer mainly include immunotherapy that targeting breast cancer-related genes, namely, nonspecific expression of human epidermal growth factor receptor (HER2), estrogen, or progesterone receptor and chemotherapy for triple-negative breast cancers (TNBCs). Chemotherapy causes damage to the whole body. Furthermore, once breast cancer resistance to one chemotherapeutic drug develops, it may also promote resistance to other chemotherapeutic drugs, that is, multidrug resistance, thus limiting treatment options. Seeking effective agents to alleviate side effects and overcome acquired resistance to chemotherapeutic drugs in breast cancer is a serious problem that needs to be solved. Considering that approximately fifty percent of TNBC patients overexpress epidermal growth factor receptor (EGFR), the activation of receptor tyrosine kinase (RTK) confers oncogenic regulation via the phosphatidylinositol 3-kinase/protein kinase B (PI3K/Akt) signaling pathway and HER2-enriched breast cancer patients specifically expresses HER2 (84), targeting EGFR may be a viable strategy for anti-breast cancer therapy. The selectivity of melittin to malignant breast cancer cells has attracted much attention (85). The tumoricidal properties of melittin on breast cancer cells have been investigated (86, 87). Melittin could impede primary tumor growth and inhibit the growth of the metastatic sentinel lymph node in breast cancer models in vitro and in vivo (88). Melittin directly acts on MCF7 breast cancer cells (81, 89) and suppresses the migration of breast cancer cell, MDA-MB-231 cell line (90). Melittin can effectively reduce the viability of human SUM159 cells (TNBC cell lines) (85). Scanning electron microscopy showed melittin-mediated membrane disruption of SUM159 cells. The levels of the apoptosis-related proteins, cleaved caspase-3, were markdely

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upregulated under melittin treatment. Moghaddam and colleagues showed that melittin-triggered apoptosis was associated with increases in the levels of mitochondrial fusion protein 2 and dynamin-related protein 1 in 4T1 TNBC cells (91). Interestingly, Duffy and colleagues designed a bifunctional melittin peptide in which an alpha-helical RGD peptide motif containing arginine, glycine and aspartic acid was added to the N-terminus of melittin. RGD-mellitin enhanced the targeting of melittin to the cell membranes in human TNBCs and T11 cells through a charged sequence present in the C-terminus of melittin. Melittin treatment downregulated the protein expression of phosphorylated RTK and the PI3K/Akt signaling pathway (85). Importantly, researchers have demonstrated synergistic effects between melittin and the chemotherapeutic agents docetaxel (85) and epirubicin (34). The resistant of tumor to docetaxel could be effectively treated under melittin administration in T11 xenograft nude mice in vivo. Significant decreases of phosphorylated EGFR and HER2 were also observed in T11-cell-bearing mice treated with melittin. Mir Hassani and colleagues reported that melittin impeded the growth of MDA-MB-231 cells, which is related with melittin-caused suppression on the TME formation (92). Hypoxia-inducible Factor 1 (HIF-1) is one of major drivers for decreasing the pH value of the solid TME by anaerobic respiration, effecting the metabolism of tumor cells (93). The alpha subunit of HIF-1 (HIF-1 α) determines the activity of HIF-1. Interestingly, melittin induced apoptosis of MDA-MB-231 cells. Melittin management reduced the protein expressions and the mRNA levels of HIF-1 α in MDA-MB-231 cells, suggesting melittin-induced suppression of TME formation. Further research is needed to describe the detailed targeting of melittin on breast cancer cells and its mechanism as well as its effects on healthy cells.

Lv and colleagues showed that melittin could inhibit the proliferation, migration and invasion of human hepatocellular carcinoma (HCC) cell lines, MHCC97-H and HepG2 (94). HCC is now the 3rd leading cause of tumor-related death. The increasing mortality rate of HCC is attributed to limited therapeutic agents and the lack of curative treatments (95). Comprehensive treatment is a strategy to improve the prognosis and survival quality of patients suffering from HCC. Sorafenib was an approved agent used for advanced HCC chemotherapy. However, Sorafenib has severe side effects and induces drug resistance, so safe substitutes or adjuvants against advanced HCC are urgently needed (96). Mansour and colleagues reported that melittin synergistically potentiates the virucidal activity of sorafenib in HepG2 cells (97). The combined treatments of melittin with sorafenib (Mel/Sorf) exhibited a synergistic and selective inhibitory effect on HepG2 cells relative to normal liver THLE-2 cells and showed a higher cytotoxic effect on HepG2 cells compared with single treatments. Mel/Sorf treatment could disrupt the cell cycle, arresting HepG2 cells at G2/M phase (97). Computational assays showed that melittin binds to X chromosome-linked inhibitor of apoptosis (XIAP). The elevation of XIAP expression levels was significant in HCC (98). Shi and colleagues observed that a large number of samples from patients with advanced HCC exhibited high expressions of XIAP protein (99). A markedly increased risk of relapse may occur in the patients with XIAP-positive HCC that underwent total resection and orthotopic transplantation of liver. Inhibitors of apoptotic proteins might be a potential anticancer strategy for hepatocellular carcinoma (100). Zhang and colleagues showed that melittin could inhibit cathepsin S-induced angiogenesis and strongly impeded proliferation, invasion and angiogenesis in the HCC cell line MHCC97-H (101). Cathepsin S is a member of the lysosomal cysteine protease family. Cathepsin S degrades many extracellular matrix (ECM) proteins, such as fibronectin, laminin, collagens and elastin (102), promoting TME formation-related immune suppression and tumor growth (103). Cathepsin S is significantly upregulated in the liver of patients with HCC (104). A series of studies (105, 106) showed that downregulating cathepsin S could inhibit the growth, invasion and angiogenesis of tumors. Interestingly, melittin specifically inhibited tumor cell growth and impeded the activity of cathepsin S in HCC cells and Mock/ MHCC97-H cells. Moreover, under melittin treatment, the protein levels of RAS, vascular endothelial growth factor (VEGF)-A, phosphorylated VEGF receptor 2 (VEGFR-2), RAF and mitogen-activated protein kinase kinase 1 (MEK1)/extracellular signal-regulated kinase (ERK)1/2 were strongly reduced in Mock/ MHCC97-H cells (101). Targeting inhibitors to regulate RAS-RAF-MEK-ERK pathway has attracted increasing attention in current research and treatment for cancer (107). Melittin-mediated effects on Mock/MHCC97-H cells show potential for HCC therapy, although more experiments are needed before clinical studies can be carried out. Zhang and colleagues also showed that melittin inhibited the proliferation of HCC and HepG2 cells by downregulating histone deacetylase 2 (HDAC2), leading to the upregulation of a tumor suppressor gene, phosphatase and tensin homolog deleted in chromosome ten (PTEN) (108). HDAC2 is associated with the DNA damage response (109). Shan and colleagues reported that increases in HDAC2 expression levels were significantly related to high-grade tumors, lymph node metastasis and poor prognosis (110). The upregulation of HDAC2 could prevent apoptosis in tumor cells (111). HDAC2 was considered a key target for anticancer treatment (112). The overexpression of HDAC2 was present in patients suffering from HCC (113). Melittin-triggered inhibition of HDAC2 suggested the potential of melittin as a treatment against HCC. Melittin can induce autophagy in HepG2 cells, activating the mitochondrial apoptotic pathway (114). Furthermore, melittin treatment downregulated DNA methyltransferase protein-1 (DNMT1). DNMT1 demethylated and upregulated long noncoding RNA (lncRNA) ADAMTS9-AS2 (94), which is a tumor suppressor (115). Melittin-mediated upregulation of ADAMTS9-AS2 provided new insights into the potential of melittin as a treatment against HCC. The anti-metastatic role of melittin was evaluated. Liu and colleagues demonstrated that melittin impeded the viability, microfilament depolymerization and migration of MHCC97L and MHCC97H HCC cells in vitro. Moreover, under melittin treatment, tumor volume was significantly reduced in nude mice bearing MHCC97H xenografts, whereas the body weight of the mice increased compared with that of tumor xenograft nude mice without melittin treatment. Interestingly, melittin triggered the inhibition of Rac1 (32), a member of the Rho GTPase family. Rac1 is considered to be involved in cancer cell metastasis (116).

Several studies have explored the strategy of targeted modification of melittin to enhance its selective and antitumor properties against HCC. Melittin was linked to a homing peptide, SLSLITMLKISR (AM-2), using A(EAAAK)2A to analyze the virucidal activity of this ligation (117). After being incorporated into AM-2, the affinities of melittin to HepG2 cells were elevated, and cell growth was inhibited, suggesting that AM-2 ligand-mediated immunoconjugation may be a valuable strategy for HCC cell targeting. Zhao and colleagues constructed a recombinant immunotoxin by fusing an antiasialoglycoprotein receptor (ASGPR) single-chain variable fragment antibody (C1) to melittin (C1M). They observed that the C1M could bind the surface of HepG2 cells and exhibit cytolytic capacity to HepG2 cells without causing erythrocyte dialysis (118). Qian and colleagues assessed the effects of melittin gene therapy. Melittin gene was carried by an adenovirus vector containing the hypoxia-response element (HRE)-alpha fetoprotein (AFP) promoter to control adenovirus E1a gene expression to target AFP-positive cancer cells under a hypoxic microenvironment. AFP-positive HCC cells were significantly inhibited under the above adenovirus carrying melittin, while under the targeted delivery of the above adenovirus, melittin management suppressed the growth of HCC cells and Hep3B cell xenografts, and increased the overall survival of nude mice bearing Hep3B cells in comparison with those of tumor-bearing mice without melittin treatment (119). Survivin is a member of the inhibitor of apoptosis family. Survivin is transcriptionally urpegulated in most malignant tumors, and its promoter is tumor specific. Using the survivin promoter (pSURV-Mel), Qu and colleagues developed a nonviral vector encoding the melittin gene to assess the anticancer effects of pSURV-Mel in HepG2 cells in vitro and a mouse model bearing human HepG2 cells in vivo (120). The results showed that pSURV-Mel was selectively expressed and induced cytotoxicity in human HepG2 cells but not in the normal liver cell Line L02. pSURV-Mel induced death of HepG2 cells through an apoptosis-dependent pathway. Ling and colleagues developed a recombinant adenovirus carrying AFP promoter and melittin gene (Ad-rAFP-Mel) to analyze the antitumor effects in human HCC cell line BEL-7402 in vitro. They showed that the tumorigenicity rate in BEL-7402 cells transfecting Ad-rAFP-Mel was significantly reduced. In addition, Ad-rAFP-Mel intratumoral injection exhibited marked antineoplastic effects on transplanted tumors in nude mice (121). Anti-HCC effects of AdrAFP-Mel were confirmed by Li and colleagues (122). Ad-rAFP-Mel treatment could inhibit cells proliferation and lead to chromatin condensation and nuclear fragmentation in BEL-7402 cells.

The effects of melittin on prostate cancer were evaluated in PC-3, LNCaP and DU145 cells. Melittin treatment could induce apoptotic cell death by inhibiting NF- κ B and upregulating caspase signaling (123). Chlorotoxin (CTX) is regarded as one of cancer cell-targeting ligands due to its capacity to recognize different tumors (124). CTX could specifically interact with matrix metalloproteinase-2 (MMP-2) and impede MMP-2 activity, which may affect tumor metastasis process (125). Tarokh and colleagues constructed a CTX-targeted nanovector to deliver the melittin gene (126). The CTX-targeted nanovector selectively increased the transfection efficiency of melittin gene in MMP-2 positive human prostate cancer cell line PC3 and inhibited cell viability without affecting the growth of MMP-2 negative NIH3T3 cell line, suggesting the antitumor selectivity of melittin under CTXtargeted nanomodification. In addition, Yan and colleagues showed that melittin could enhance cisplatin sensitivity and inhibit migration of in castration-resistant prostate cancer cells and PC3 cells (127). Interestingly, melittin-triggered tumor impediment and cisplatin sensitivity could be abrogated by the overexpression of lipocalin-2 (LCN2), a gene of the IL-17 signaling pathway. It is nominated the potential of melittin as a candidate for CRPC therapy.

Anti-inflammatory potential of melittin

Inflammation is a defense response of the body against injury. The body maintains homeostasis by limiting infection and repairing damage. However, chronic inflammation can trigger or exacerbate a variety of diseases, such as arthritis, heart disease, arteriosclerosis, inflammatory bowel disease, asthma, central nervous system disease and diabetes. It was reported that melittin could inhibit excessive inflammation, which suggests the melittin can treat inflammationinduced diseases (Supplementary Table 3).

Melittin effectively mitigates the lipopolysaccharide (LPS)mediated pro-inflammatory response in mouse BV2 microglia (128). LPS is thought to be a key molecule in the endotoxic shock associated with gram-negative infection, which is released during cell division, death or an inflammatory response (129). Moon and colleagues showed that melittin inhibited LPS-primed of nitric oxide/inducible nitric oxide synthetase (NO/iNOS) expressions in a dose-dependent manner in BV2 microglia. Melittin treatment reduced the protein expressions of a major mediator of inflammation, prostaglandin E2 (PGE2), and inhibited LPSprimed activation of nuclear factor-kB (NF-kB). Additionally, melittin inhibited the transcription of interleukin-6 (IL-6), interleukin-1 β (IL-1 β) and tumor necrosis factor- α (TNF- α) triggered by LPS in BV2 cells. Han and colleague analyzed the effects of melittin on free radical-triggered neuronal damage (130). H₂O₂ treatment was used to mimic the stimuli of free radicals in the human neuroblastoma SH-SY5Y cell line. Under chronic oxidative stress, cell viability was inhibited, and the protein expressions of Caspase-3 were increased, whereas melittin management significantly reduced DNA fragmentation and increased cell viability. Melittin inhibited H2O2-triggered downregulation of BCL-2 in the protein expressions and mRNA levels. In addition, melittin ameliorated H₂O₂-induced upregulation of pro-apoptotic factors Bax and Caspase-3 in the protein expressions and mRNA levels, indicating melittin-mediated neuroprotective effects against oxidative stress by inhibiting apoptosis in neuronal cells. Considering that oxidative stress is an important inducer of neurodegeneration, these results suggest the potential of melittin for neurodegenerative disease treatment. Amyotrophic lateral sclerosis (ALS) is associated with mutations in superoxide dismutase (SOD1) in motor neurons, excitotoxicity, defective axonal transport, oxidative damage, mitochondrial dysfunction, protein misfolding, abnormal cytoskeleton and neuroinflammation. To analyze the effects of melittin on motor neurons of ALS-related pathological changes, melittin was administered to HSOD1G93A

mice, an ALS model animal. Yan and colleagues showed that melittin treatment improved motor function in mice and alleviated neuronal damage in the spinal cord of HSOD1G93A mice. Melittin administration mitigated α -synuclein misfolding in the brainstem and spinal cord of HSOD1G93A mice, restoring the activity of proteasome in the brainstem and spinal cord (131).

Several researchers have investigated the potential of melittin against rheumatoid arthritis (RA). RA-caused irreversible destruction and functional loss of joints disturb the life of patients. Fibroblast-like synoviocytes (FLS) produce cytokines in the synovial intimal lining, perpetuating inflammation and proteases and then promoting cartilage destruction. Nah evaluated the effects of melittin on FLSs from patients with RA. Under LPS stimuli, the characteristic mediums mediating RA and matrix metalloproteinases (132) were significantly elevated. Under melittin treatment, LPS-induced increases in MMP-3 production and DNA binding activity of NFκB were alleviated in FLS (133). Melittin-mediated inhibition of FLS was confirmed by He and colleagues (134). The researchers observed that melittin markedly inhibited the viability of FLS from patients with RA (RA-FLS), reduced the secretion of IL-1B and promoted apoptosis and autophagy in RA-FLS. FLS survival might depend on the activation of transcription factors, signal transducer and activators of transcription 3 (STAT3), which is regulated by IL-6 (135). It has been reported that inhibiting IL-6 (136) or the IL-6 receptor (137) could improve RA patients' signs and symptoms. The cooperation of IL-6 and soluble IL-6 receptor (sIL6R) can induce FLS proliferation (138). Kim and colleagues showed that IL-6/sIL-6R stimuli activate the STAT3 and nuclear NF-KB p65 in FLS. Melittin treatment ameliorated IL-6/sIL-6R-triggered activation of STAT3 and nuclear translocation of NF-KB p65, increased the protein levels of pro-apoptosis-related proteins, apoptotic protease activating factor 1 (Apaf-1), caspase-9 and caspase-3, and increased the release of cytosolic cytochrome c (139).. Mitogen-activated protein kinase (MAPK) and NF-KB are abnormally activated in the inflammatory pathological process of RA, and the transmission of inflammatory signals among abnormal signaling pathways promotes the release of inflammatory factors (140). Melittin can interact with the P50 subunit (24) and thiol group of IKB kinase (IKK) (141) so that the IKB kinase loses its catalytic effect on the phosphorylation of IκB (inhibitor of NF-κB). NF-κB cannot enter the nucleus. P50 nuclear translocation is also inhibited, which inhibits the production of inflammatory mediators (141, 142). Park and colleagues confirmed that melittin treatment prevented LPS-induced transcription of NFκB and impeded NF-κB DNA binding activity of by suppressing IκB release and p50 translocation (24). Interestingly, melittin treatment could reverse the LPS-primed activation of JNK and NF-KB in Raw 264.7 cells (a murine macrophage line), and the effects above were abolished by a JNK inhibitor (143), suggesting that the properties of melittin on anti-inflammatory and anti-arthritis were involved in regulating JNK signaling pathways.

Ulcerative colitis (UC) is a type of bowel disease with the characteristics of primarily affecting the superficial layers of the rectum and colon and lifelong inflammation (144). Routine therapies cannot achieve satisfactory efficacy, and the side effects cannot be ignored. More effective therapeutic agents are still needed

for UC. Ahmedy and colleagues assessed the antiulcerogenic effect of melittin by oral administration in a colitis mouse model induced by acetic acid (145). Melittin treatment ameliorated acetic acidtriggered decreases of body weight and an increase of colon mass index in colitis model mice. The levels of PGE2, NF-KB, Toll-like receptor 4 (TLR4), tumor necrosis factor (TNF- α), IL-6, secretory phospholipase A2 (sPLA2), p38 MAPK, tumor necrosis factor receptor (TNF-R)-associated factor (TRAF6) and cyclooxygenase 2 (COX-2) were significantly reduced under melittin administration compared with those in colitis model mice without melittin treatment. In addition, melittin attenuated oxidative stress in colitis model mice. Anti-inflammatory property of melittin against UC was assessed in dextran sulfate sodium (DSS)-induced colitis models (146). The synergies between melittin and sulfasalazine were also analyzed to evaluate whether combination therapy could improve the clinical symptoms of UC model mice. The results showed that melittin and the combination of melittin with sulfasalazine significantly alleviated pathological damage in the colon tissue of UC mouse model. Moreover, melittin treatment exerted antioxidant effects on colitis model mice, suggesting that melittin might be a potential alternative agent for UC intervention.

Acute liver failure (ALF) occurs shortly after liver damage in people who have not previously suffered from liver disease. ALF is a severe clinical syndrome with extremely poor prognosis and high mortality. There is no curative treatment for ALF. It is considered that the high mortality rate of ALF may be due to the severe disorder of the systemic inflammatory response (147). Modulating macrophages to suppress inflammation is considered a promising strategy for ALF intervention (148). Park and colleagues assessed the effects of melittin against ALF in a mouse model caused by dgalactosamine (d-GalN) and LPS (d-GalN/LPS) intraperitoneal challenge in vivo (149). They showed that melittin attenuated the d-GalN/LPS-induced increase of IL-1 β and TNF- α in the serum and liver of ALF model mice. Apoptosis was less in the melittin-treated d-GalN/LPS mice than those of d-GalN/LPS-treated group without melittin administration. Moreover, melittin management mitigated the activation of NF-KB in ALF model mice. Melittin-mediated protection against ALF failure by inhibiting inflammatory cytokine release was confirmed by Fan and colleagues (150). The researchers demonstrated the anti-inflammatory activities of melittin in ALF mouse induced by d-GalN/LPS in vivo and LPS-stimulated RAW264.7 macrophages in vitro. Researchers observed that LPS stimulation facilitated aerobic glycolysis in RAW264.7 cells by increasing the levels of Warburg effect-associated enzymes and metabolites, upregulating glycolytic rate, and enhancing Akt/ mTOR/PKM2/HIF-1a signaling. The mortality of ALF mice were significantly reduced under melittin treatment, and the symptoms and signs of ALF were mitigated. Melittin management ameliorated inflammation and exerted antioxidative effects on RAW264.7 cells. Furthermore, melittin treatment attenuated LPS-primed increases in HIF-1 α and PKM2 levels by alleviating LPS-triggered upregulation of Akt/mTOR signals in RAW264.7 cells and ALF model mice, and the melittin-mediated effects above were abolished by shRNA against PKM2. The authors considered that melittin alleviated inflammation in ALF through inhibiting PKM2-mediated Warburg effects.

Acne is a chronic inflammation of the sebaceous glands in the skin follicles, which is closely associated with the inflammatory reaction caused by Propionibacterium acnes (PAC). Lee and colleagues examined the activity of melittin against PAC-triggered inflammatory responses in THP-1 monocytic cells (151). They observed that melittin treatment attenuated heat-killed PACmediated increases of cleavage of caspase-3 and -8 and proinflammatory cytokines, and mitigated PAC-triggered apoptosis in THP-1 cells. In addition, melittin treatment alleviated heat-killed PAC-induced increases in proinflammatory cytokine levels and Toll-like receptor 2 expression levels by inhibiting MAPK and NF-KB signaling pathways in a human keratinocyte HaCaT cells. Melittin also attenuated swelling, cutaneous erythema and the granulomatous response in the ears of living PAC-injected mice than those of mice injected with living PAC without melittin treatment (152).

Melittin could exert anti-inflammatory effects against a kind of inflammatory skin disease, atopic dermatitis (also named atopic eczema). The main clinical manifestations of atopic dermatitis include epidermal thickening, skin dryness and pruritus, a decrease in filaggrin, damage to the epidermal barrier, various allergic reactions mediated by IgE, and disturbance of the immune system (153, 154). Kim and colleagues showed that melittin mitigated ovalbumin-triggered skin symptoms in an atopic dermatitis model mouse *in vivo*. Melittin treatment regulated skin thickness, increased filaggrin levels, reduced inflammatory cytokine and chemokine levels and alleviated the IgE response. Melittin could inhibit the phosphorylation of STAT3, reduce the secretion of IL-4 and IL-13 and increase the protein expressions of filaggrin in HaCaT cells. Further studies may provide more clues regarding melittin in atopic dermatitis treatment (155).

Periodontitis is a type of inflammatory disease affecting gums, periodontal ligament, alveolar bone and cementum, resulting in the destruction of the gingiva. *Porphyromonas gingivalis* (*P. gingivalis*) is a main pathogen of chronic periodontitis and can cause alveolar bone absorption and tooth loss by invading periodontal tissues and releasing toxic substances (156). Kim and colleagues evaluated the effects of melittin on *P. gingivalis* LPS (PgLPS)-infected HaCaT cells (157). They observed that melittin treatment ameliorated PgLPSinduced increases in the protein expressions of IL-6, IL-8, TLR-4, TNF- α and interferon- γ . Melittin incubation mitigated PgLPStriggered activation of extracellular signal-regulated kinase, NF- κ B and PKB/Akt.

Discussion

Melittin-induced cytotoxicity limits its biological applications

Melittin is characterized by antiviral activity; however, melittininduced destruction of the membrane of red blood cells should not be ignored (4, 158, 159). Gajski and colleagues showed that there are morphological changes in the cell shape, membrane damage, cell granulation and lysis of human peripheral blood lymphocytes (HPBLs), apart from a decline in viability when exposed to melittin (158). Under melittin treatment, the levels of ROS were significantly increased, and oxidation-reduction homeostasis was disrupted. Melittin increased the formation of micronuclei, nuclear buds and nucleoplasmic bridges, suggesting that genome instability was triggered by melittin. Melittin treatment increases the mRNA levels of oxidative stress- and apoptosis-related genes, which suggests that it exerts cytotoxic effects on normal human cells (160). Melittin can cause tonic pain and peripherally persistent pain. Local injection with melittin could cause hyperalgesia, allodynia and inflammatory responses in the injection site (161). Melittin enhances the excitability of spinal nociceptive neurons. Local injection of melittin also leads to hyperresponsiveness of spinal wide dynamic-range neurons under mechanical and thermal stimuli, causing spontaneous nociceptive flinches and pain hypersensitivity (161). Melittin treatment by intradermal injection of triggered mechanical hyperalgesia and heat in the zone of primary injury in humans (162, 163). Subcutaneous injection of melittin triggers heat hypersensitivity by activating tetrodotoxinresistant sodium channels in small primary sensory neurons (164). Melittin-caused toxicity to cells or to the host animal is not uniformly (165). The median hemolytic concentration (HC_{50}) of melittin against human red blood cells (2% suspension) is 16.28 \pm 0.17 μ g/mL and the median lethal concentration (LC₅₀) of melittin against immature human dendritic cells (1 \times 10⁵/mL) is 43.42 \pm 0.86 µg/mL when incubated for 24 h (166). Melittin can trigger morphological changes in mouse peritoneal macrophages at the concentration of 2.5 μ g/mL, and the half-maximal inhibition (IC₅₀) is 5.73 μ g/mL (167). It is necessary to seek strategies to reduce the cytotoxicity and enhance the bioavailability of melittin.

Packaging melittin by nanocarriers is one strategy to minimize the toxicity of melittin

Ligands such as oligonucleotide or peptide molecules can specifically bind to melittin with high affinity. In addition, researchers have shown that the synergistic effects of melittin with some first-line drugs could effectively reduce melittin cytotoxicity and drug resistance. Gui and colleagues developed a nanocomplex system based on polyelectrolytes to allow strong interactions formation between positively charged melittin and negatively charged dextran sulfate using flash nanocomplexation technology (168). They obtained uniform nanocomplexes with high melittin encapsulation efficiency, high payload capacity and tunable release profiles. When treated with free melittin by intraperitoneal (i.p.) injection, there was high acute liver toxicity and a low survival rate in a type II diabetes (T2D) mouse model. In contrast, there was no death in the mice treated with melittin-loaded nanocomplexes. The time of melittin release was prolonged. The blood glucose levels declined after melittin nanocomplex *i.p.* injection. Moreover, mouse body weight was reduced. The author considered that their melittin nanocomplexes reduced acute toxicity and improved pathological indicators, enhancing the applicable potential of melittin. Ye and colleagues (169) constructed lipid-coated polymer nanoparticles

loaded with melittin, which exhibited suitable biocompatibility and stability. The nanosystem contained a PEG-targeting molecular shells and lipid membrane intermediate layer. The formulation was prepared by self-assembly according to intermolecular interactions, comprising hydrophobic effects and electrostatic attraction. Under physiological conditions, the melittin-loaded system exhibited a high encapsulation efficiency and stability. Hemolysis showed that free melittin induced lysis rate of red blood cells was100%. In contrast, neither remarkable hemolysis in the red cells nor specific cytotoxicity was observed under melittinloaded nanosystem treatment, even at high concentrations of melittin. Nanomodification enhanced the selectivity of melittin for tumor cells and markedly inhibited the tumor growth without causing significant normal tissue toxicity or hemolysis in nude mice bearing the human lung adenocarcinoma xenografts.

Li and colleagues (170) designed liposomes with dioleoyl phosphoethanolamine (DOPE)-conjugated hyaluronic acid (HA) inserted by covalent bonds (HA-DOPE). Then, HA-modified liposomes were used to carry melittin (Mel-HA-Lip). HA and its derivatives can interact with specific receptors, such as HA receptor for endocytosis (HARE), receptor of HA-mediated mot and CD44 in the cell membrane (171). CD44 is a widely distributed glycoprotein. Overexpression of CD44 can be observed in the cell membrane of some malignant tumor cells. The specific binding of CD44 with HA can mediate transmembrane transportation (172, 173). The nature of specific binding between HA/HA derivatives and CD44 provides targeted delivery for antitumor drugs to tumor cells (174). HA-modified liposomes enhanced the uptake of melittin by B16F10 cells. Mel-HA-Lip exhibited higher levels of sustainedreleases than Mel-Lip. Furthermore, Mel-HA-Lip performed more obvious antitumor effects compared with Mel-Lip (170). Dai and colleagues developed dual-targeting nanoparticles carrying melittin with HA-conjugated high-density lipoprotein (HDL) as the phospholipid scaffold (MLT-HA-HPPS) to target breast cancer and its sentinel lymph node (SLN) (88). MLT-HA-HPPS exhibited effective cytotoxicity on mouse mammary adenocarcinoma 4T1 cells. MLT-HA-HPPS paracancerous administration significantly suppressed the growth of primary tumor. It is noted that MLT-HA-HPPS could target SLN and impede the growth of metastatic SLN in the lymph node metastasis model bearing 4T1 cells. Tang and colleagues developed TME-responsive MnO2-melittin nanoparticles (M-M NPs) based on an in situ redox reaction between poly(allylamine hydrochloride) and KMnO4 (74). Melittin was carried by carbodiimide method. M-M NPs consumed GSH at a faster rate and facilitated more cytotoxic ROS (•OH) production, inducing tumor cell death without affecting the viability of antigen presenting cells, murine DC cells and bone marrow (BM)-derived macrophages. M-M NPs inhibited tumor growth and metastasis, and increased inflammatory levels in MB49 prostate cancer subcutaneous tumor model mice in vivo and in MC38 colon cancer and B16 melanoma in vitro. Importantly, M-M NPs facilitated MHC-I cross-reactivity via DC, activating tumorspecific CD8 positive T cells, which suggested the antitumor immunotherapy potential of MnO2-melittin.

The peptide-peptide and peptide-drug conjugate strategy may increase the bioavailability and relative safety of melittin

A conjugated peptide-peptide fused with melittin (melittin 8-26) and the pro-apoptotic peptide d(KLAKLAK)2 (dKLA) could inhibit tumor growth and reduce mortality in a mouse model bearing 4T1 TNBC in vivo (82). Melittin-dKLA 8-26 treatment reduces the numbers of M2-like tumor-associated macrophages, which are the main components of TME and involved in the regulation of tumor invasion, angiogenesis, growth, migration and metastasis (71, 175). Additionally, under melittin-dKLA 8-26 administration, lung metastasis was significantly suppressed in 4T1 TNBC mice. Melittin could also increase ROS levels, triggering immunogenic cell death (176). Lee and colleagues (177) demonstrated that melittin-dKLA selectively inhibited M2-like TAMs in cancer mouse model bearing murine Lewis lung carcinoma cells in vivo. Melittin-dKLA could suppress the growth and lower the weights of tumor, and impeded angiogenesis. Jeong and colleagues (178) also showed that the peptide-drug conjugate derived by melittin could inhibit melanoma by targeting M2-like TAMs in a melanoma mouse model bearing B16F10 cells. The research conjugated melittin and mertansine (DM1), an inhibitor targeting the assembly of tubulin into microtubules and cell cycle. Melittin-DM1 significantly increased the survival rates of melanoma mice than those under DM1 management alone. Moreover, a series of studies (86, 87, 179) investigated several strategies, such as innocuous nonspecific toxicity and increased stability of melittin in anti-breast cancer therapy. Daniluk and colleagues showed that compared to melittin alone, using diamond, pristine graphene, graphene oxide and carbon nanoparticles as the carriers for melittin reduced the level of necrosis in MDA-MD-231 and MCF-7 cells (86). Through caveolin-dependent endocytosis, graphene-carried melittin can be transported to MDA-MD-231 cells (87). Melittin-graphene oxide complex reached MDA-MB-231 cells through phagocytosis, microtubule-dependent transport and caveolin-dependent endocytosis. Raveendran and colleagues (179) developed a polyion complex micelle modified by estrone for delivering melittin to hormone-responsive breast cancer cells.

Nabizadeh and colleagues (180) designed a hybrid peptide by fusing truncated-melittin (11 amino acids with substituting arginine for isoleucine in the 5th position) to Lasioglossin (LL-III, 15 amino acids), which can suppress Acinetobacter (A.) baumanni. The hybrid peptide above was named LM1 and was demonstrated to be high stability. The interactions of LM1 with lipid membrane were increased compared with melittin and exhibited antimicrobial potential to halter S. aureus bacteria and A. baumanii. Huang and colleagues designed α -melittin, a hybrid cytolytic peptide, by linking an amphipathic α -helical peptide to the N-terminus of melittin using a GSG linker (181). α -Melittin can bind phospholipids and self-assemble, to form lipid nanoparticles. The lipid nanoparticle-carried α -melittin (α -melittin-NPs) shielded the positive charge of melittin, reducing its cytotoxicity. In addition, α melittin-NPs treatment by intravenous injection suppressed the growth of tumor in melanoma-bearing nude mice. Interestingly, α melittin-NPs could trigger a systemic antitumor response (182). In

comparison with free melittin, *a*-melittin-NPs can facilitate the lymph node uptake of melittin, stimulating activation of antigenpresenting cells activation in C57BL/6 mice. Moreover, α-melittin-NPs could impede primary and distant tumor growth in a model of bilateral flank melanoma by intradermally implanted B16F10 cells. α-Melittin-NPs could also control T-cell-mediated chronic allergic contact dermatitis and atopic dermatitis skin (183). In an allergic contact dermatitis mouse model induced by oxazolone challenge in the ear, both melittin and α -melittin-NPs intradermal injection significantly alleviated ear swelling. In comparison with direct intradermal injection of melittin, α -melittin-NPs treatment alleviated the excess accumulation of leukocytes at the injection position without damaging the epidermal structure integration. α -Melittin-NPs target lymph nodes and suppress allergen-induced maturation of dendritic cells in allergic contact dermatitis mice, whereas α -Melittin-NPs reduce the levels of phosphorylated-RelB (pRelB) and impede the activation and proliferation of CD8⁺ T cells. RelB is a critical signaling molecule of NF-KB signaling. Phosphorylation-activated RelB migrates to the nucleus, priming gene transcription associated with the maturation of dendritic cells (184). Interestingly, α -melittin-NPs controlled the release of Th1type cytokines in allergic contact dermatitis mice and restricted the release of Th2-type cytokines and IgE in mice with atopic dermatitis-like pathology, indicating the immunosuppression against T cells-mediated immune reactions.

Melittin might be used in synergistic therapeutics for medicine in clinic

Codelivery of melittin with antibiotics, chemotherapeutics, small molecule compounds, proteins or gene agents for synergistic therapeutics was explored to develop safer and more efficient strategies against disease. Through hydrogen-bond interactions, melittin (MPI) was wrapped with fluorinated epigallocatechin-3olegate (EGCG) (185). The fluorination of EGCG occurred through the reaction of EGCG and fluorinated compounds by ROS-sensitive linker of oxalyl chloride. EGCG, a potential inhibitor of programmed death-ligand 1 (PD-L1), could regulate the response of tumor cells to immunotherapy (186, 187). The self-assembly of melittin and fluorinated EGCG (FEGCG) was formulated into a fluoronanoparticle system, FEGCG@MPI NPs, which may deliver small molecule drugs but also larger macromolecules (185). Fluorine modification of EGCG ameliorated the hemolysis induced by melittin and promoted the cellular uptake of FEGCG@MPI NPs by Hep3B cells. FEGCG@MPI NPs markedly reduced PD-L1 protein expressions of and disrupted the interactions of PD-L1 and programmed death-1 (PD-1). FEGCG@MPI NP treatment inhibited tumor growth and induced tumor cell apoptosis in HCC mice bearing Hep3B tumor cells. Yazdian-Robati and colleagues developed calcium carbonate nanoparticles (CCNs) with mucin 1-dimer aptamers to carry epirubicin and melittin by using a water-in-oil emulsion method (34). They showed that the codelivery of epirubicin and melittin by using a mixture of mucin 1-dimer aptamer-CCNs-melittin complex and micin 1-dimer aptamer-CCNs-epirubicin complex exerted significant synergistic cytotoxicity in murine colon cancer C26 cell line and MCF-7 cells. Furthermore, under the delivery of mucin-1dimer aptamer-modified CNN, epirubicin and a mixture containing epirubicin and melittin exhibited more effective antitumor effects than free epirubicin treatment in a colon cancer mouse model bearing C26 cells. Wang and colleagues (188) constructed nanolipodisks modified by glycosylated heptapeptide ATWLPPR (9G-A7R) containing 1palmitoyl-2-oleoyl-sn-glycero-3-phosphocholine (POPC) and cholesterol plus 1,2-distearoyl-sn-glycero-3-phosphoethanolamine-N-[methoxy(polyethylene glycol)] (DSPE-PEG) to carry melittin and paclitaxel. 9G-A7R can specifically bind neuropilin-1 and vascular endothelial growth factor receptor 2 overexpressing in glioma cells, enhancing its proteolytic stability in serum and accessibility of A7R to the brain. The nanolipodisks described above could be prominently taken up by glioma. Paclitaxel combined with melittin protected against proteolysis and hemolysis, exhibited synergism against the human glioma cell line U87 and showed enhanced antiglioma properties in glioma-bearing mice. The glycopeptide-enabled paclitaxel/melittin delivery system showed significant anti-glioma effects in an intracranial glioma xenograft model (189). Photodynamic therapy (PDT) is to induce ROS production to induce immunogenic cell death for antitumor therapy. Liu and colleagues developed a serum albumin (SA)-coated boehmite (aluminum hydroxide oxide) organic-inorganic scaffold to envelop melittin and chlorin e6 (Ce6), a photosensitizer. Compared with free melittin treatment, this system significantly reduced hemolysis, enhanced intracellular ROS generation, and exhibited melittinmediated enhancement of PDT effects, inhibiting tumor growth in a mouse model of breast cancer bearing 4T1 cells (176). Motiei and colleagues designed a polyelectrolyte nanocarrier via self-assembly of polyanionic and polycationic chains using folic acid-decorated polyethyleneimine as the shell, dextran sulfate as a complexing agent and polyglutamate grafted chitosan as the core (190). The nanocarrier above was used to deliver LTX-315, a 9-mer cationic amphiphilic peptide from bovine lactoferricin, melittin or miR-34a. LTX-315 can trigger an anticancer immune response, killing human cancer cells (191). miR-34a is a tumor suppressor miRNA. miR-34a downregulation is involved in the proliferation, invasion and metastasis of multiple types of cancers (192). The polyelectrolyte nanosystem carrying miR-34a, melittin or LTX-315 cold controls the release of genes and peptides and exhibits synergistic anti-tumor efficacy of melittin and miR-34a, such as arrest cell cycle and induce apoptosis of MDA-MB-231 cells (190). Asfour and colleagues designed a nanoconjugate of melittin and gabapentin and evaluated the synergistic effects against diabetic wounds (193). It was reported that gabapentin could promote wound healing (194). When conventional treatment fails, gabapentin can be used and compounded to promote dermal healing in the clinic (195). Under gabapentin-melittinconjugated hydroxypropyl-methyl cellulose hydrogel treatment, wound contraction was expedited in streptozotocin-induced diabetic Wistar rats, suggesting potent wound healing activities. Furthermore, the gabapentin-melittin conjugate formulation increased the activity of antioxidant enzymes, reduced lipid peroxidation, inhibited the release of inflammatory factors, and enhanced proliferative and pro-collagen activities, indicating the synergistic effects of melittin with gabapentin on wound healing in diabetes. In addition, it was shown that a hydrogel formulation



embedded tobramycin and melittin could efficiently alleviate biofilmassociated infections in a mouse wound model (196). Recently, Koide and colleagues engineered a temperature-responsive poly(Nisopropylacrylamide) (pNIPAm)-based copolymer. The nanosystem above could trap melittin at 37°C and swell and rapidly release melittin at 25°C. Furthermore, no hemolysis or cytotoxicity at 37°C occurred but hemolysis and cytotoxicity were observed at 25°C (33). Under tumor cooling treatment, melittin was released from the nanosystem only at the cooled site, and growth of tumor was inhibited in Colon26 NL-17-bearing mice *in vivo*, suggesting the applicable potential of melittin for anticancer therapy by local cooling-triggered site-specific release. Melittin modulation that have been reported in the literature are depicted in Figure 2.

Conclusion

Melittin exhibits antiviral, antitumor, and anti-inflammatory functions, although there are practical challenges, such as hemolysis and drug safety. The scientific research on the functional analysis and mechanistic exploration of melittin has progressed, and the understanding of melittin has improved. Importantly, studies using nanomodification, immuno-conjugation, structural regulation and gene technology strategies provide insights into the potential applications of melittin in disease intervention and also for the exploration and application of similar antimicrobial peptides. At present, although there is no trial of melittin on humans that demonstrates the therapeutic properties and investigates the biosafety of melittin management in clinic, it is worth noting that increasing researches undoubtedly indicate the potential of melittin as a candidate for disease intervention. Due to the multi-target efficiency of melittin, further animal experiences should be performed to address the molecular mechanism of melittin action.

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References

1. Van Epps HL. Rene Dubos: unearthing antibiotics. J Exp Med (2006) 203:259. doi: 10.1084/jem.2032fta

2. Huan Y, Kong Q, Mou H, Yi H. Antimicrobial peptides: classification, design, application and research progress in multiple fields. *Front Microbiol* (2020) 11:582779. doi: 10.3389/fmicb.2020.582779

3. Boman HG. Peptide antibiotics and their role in innate immunity. *Annu Rev Immunol* (1995) 13:61–92. doi: 10.1146/annurev.iy.13.040195.000425

4. Habermann E. Bee and wasp venoms. Science (1972) 177:314-22. doi: 10.1126/ science.177.4046.314

5. Raghuraman H, Chattopadhyay A. Melittin: a membrane-active peptide with diverse functions. *Biosci Rep* (2007) 27:189-223. doi: 10.1007/s10540-006-9030-z

 Lee AC, Harris JL, Khanna KK, Hong JH. A comprehensive review on current advances in peptide drug development and design. *Int J Mol Sci* (2019) 20:2383. doi: 10.3390/ijms20102383

7. Terwilliger TC, Eisenberg D. The structure of melittin. II. Interpretation of the structure. J Biol Chem (1982) 257:6016–22. doi: 10.1016/S0021-9258(20)65098-0

8. Bazzo R, Tappin MJ, Pastore A, Harvey TS, Carver JA, Campbell ID. The structure of melittin. A 1H-NMR study in methanol. *Eur J Biochem* (1988) 173:139–46. doi: 10.1111/j.1432-1033.1988.tb13977.x

9. Hider RC. Honeybee venom: a rich source of pharmacologically active peptides. *Endeavour* (1988) 12:60–5. doi: 10.1016/0160-9327(88)90082-8

10. Picoli T, Peter CM, Zani JL, Waller SB, Lopes MG, Boesche KN, et al. Melittin and its potential in the destruction and inhibition of the biofilm formation by Staphylococcus aureus, Escherichia coli and Pseudomonas aeruginosa isolated from bovine milk. *Microb Pathog* (2017) 112:57–62. doi: 10.1016/j.micpath.2017.09.046

11. Lee G, Bae H. Anti-inflammatory applications of melittin, a major component of bee venom: detailed mechanism of action and adverse effects. *Molecules* (2016) 21:616. doi: 10.3390/molecules21050616

12. Xu C, Ma W, Wang K, He K, Chen Z, Liu J, et al. Correlation between singlemolecule dynamics and biological functions of antimicrobial peptide melittin. *J Phys Chem Lett* (2020) 11:4834–41. doi: 10.1021/acs.jpclett.0c01169

13. Pandidan S, Mechler A. Nano-viscosimetry analysis of the membrane disrupting action of the bee venom peptide melittin. *Sci Rep* (2019) 9:10841. doi: 10.1038/s41598-019-47325-y

14. Wang A, Zheng Y, Zhu W, Yang L, Yang Y, Peng J. Melittin-based nano-delivery systems for cancer therapy. *Biomolecules* (2022) 12:118. doi: 10.3390/biom12010118

15. Ben Trad F, Delacotte J, Guille-Collignon M, Lemaître F, Arbault S, Sojic N, et al. Electrochemiluminescence imaging of liposome permeabilization by an antimicrobial peptide: melittin. *Chem BioMed Imaging* (2023) 1:58–65. doi: 10.1021/cbmi.3c00003

16. Durvanger Z, Juhasz T, Liliom K, Harmat V. Structures of calmodulin-melittin complexes show multiple binding modes lacking classical anchoring interactions. *J Biol Chem* (2023) 299:104596. doi: 10.1016/j.jbc.2023.104596

17. Nizet V. Antimicrobial peptide resistance mechanisms of human bacterial pathogens. *Curr Issues Mol Biol* (2006) 8:11–26.

18. Kasozi KI, Niedbala G, Alqarni M, Zirintunda G, Ssempijja F, Musinguzi SP, et al. Bee venom-A potential complementary medicine candidate for SARS-CoV-2 infections. *Front Public Health* (2020) 8:594458. doi: 10.3389/fpubh.2020.594458

19. Erkoc P, von Reumont BM, Luddecke T, Henke M, Ulshofer T, Vilcinskas A, et al. The pharmacological potential of novel melittin variants from the honeybee and solitary bees against inflammation and cancer. *Toxins (Basel)* (2022) 14:818. doi: 10.3390/toxins14120818

20. Pandey P, Khan F, Khan MA, Kumar R, Upadhyay TK. An updated review summarizing the anticancer efficacy of melittin from bee venom in several models of human cancers. *Nutrients* (2023) 15:3111. doi: 10.3390/nu15143111

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fimmu.2024.1326033/ full#supplementary-material

21. Choi JH, Jang AY, Lin S, Lim S, Kim D, Park K, et al. Melittin, a honeybee venom –derived antimicrobial peptide, may target methicillin–resistant Staphylococcus aureus. *Mol Med Rep* (2015) 12:6483–90. doi: 10.3892/mmr.2015.4275

22. Taylor AR. Methicillin-resistant Staphylococcus aureus infections. Prim Care (2013) 40:637–54. doi: 10.1016/j.pop.2013.06.002

23. Kende A, Lai F, Lim PP, Swindale L, Hofstra A, Zhang F, et al. Mode of action hypothesis testing in chemical safety assessments using metabolomics as supporting evidence: Phenobarbital and cyclobutrifluram metabolomics profile comparison. *Toxicol Lett* (2023) 382:13–21. doi: 10.1016/j.toxlet.2023.04.008

24. Park HJ, Lee SH, Son DJ, Oh KW, Kim KH, Song HS, et al. Antiarthritic effect of bee venom: inhibition of inflammation mediator generation by suppression of NF-kappaB through interaction with the p50 subunit. *Arthritis Rheumatol* (2004) 50:3504–15. doi: 10.1002/art.20626

25. Tender T, Rahangdale RR, Balireddy S, Nampoothiri M, Sharma KK, Raghu Chandrashekar H. Melittin, a honeybee venom derived peptide for the treatment of chemotherapy-induced peripheral neuropathy. *Med Oncol* (2021) 38:52. doi: 10.1007/s12032-021-01496-9

26. Silva Oliveira Tde A, Alves do Nascimento AM. [Pharmaceutical Assistance in the Family Healthcare Program: points of affinity and discord in the organization process]. *Cien Saude Cole* (2011) 16:3939–49. doi: 10.1590/s1413-81232011001000031

27. Son DJ, Lee JW, Lee YH, Song HS, Lee CK, Hong JT. Therapeutic application of anti-arthritis, pain-releasing, and anti-cancer effects of bee venom and its constituent compounds. *Pharmacol Ther* (2007) 115:246-70. doi: 10.1016/j.pharmthera.2007.04.004

28. Hait WN, Grais L, Benz C, Cadman EC. Inhibition of growth of leukemic cells by inhibitors of calmodulin: phenothiazines and melittin. *Cancer Chemother Pharmacol* (1985) 14:202–5. doi: 10.1007/BF00258116

29. Saini SS, Chopra AK, Peterson JW. Melittin activates endogenous phospholipase D during cytolysis of human monocytic leukemia cells. *Toxicon* (1999) 37:1605–19. doi: 10.1016/s0041-0101(99)00110-5

30. Moon DO, Park SY, Choi YH, Kim ND, Lee C, Kim GY. Melittin induces Bcl-2 and caspase-3-dependent apoptosis through downregulation of Akt phosphorylation in human leukemic U937 cells. *Toxicon* (2008) 51:112–20. doi: 10.1016/j.toxicon.2007.08.015

31. Ceremuga M, Stela M, Janik E, Gorniak L, Synowiec E, Sliwinski T, et al. Melittin-A natural peptide from bee venom which induces apoptosis in human leukaemia cells. *Biomolecules* (2020) 10:247. doi: 10.3390/biom10020247

32. Liu S, Yu M, He Y, Xiao L, Wang F, Song C, et al. Melittin prevents liver cancer cell metastasis through inhibition of the Rac1-dependent pathway. *Hepatology* (2008) 47:1964–73. doi: 10.1002/hep.22240

33. Koide H, Saito K, Yoshimatsu K, Chou B, Hoshino Y, Yonezawa S, et al. Coolinginduced, localized release of cytotoxic peptides from engineered polymer nanoparticles in living mice for cancer therapy. *J Control Release* (2023) 355:745–59. doi: 10.1016/ j.jconrel.2023.02.020

34. Yazdian-Robati R, Arab A, Ramezani M, Rafatpanah H, Bahreyni A, Nabavinia MS, et al. Smart aptamer-modified calcium carbonate nanoparticles for controlled release and targeted delivery of epirubicin and melittin into cancer cells in *vitro* and in *vivo. Drug Dev Ind Pharm* (2019) 45:603–10. doi: 10.1080/03639045.2019.1569029

35. Baghian A, Jaynes J, Enright F, Kousoulas KG. An amphipathic alpha-helical synthetic peptide analogue of melittin inhibits herpes simplex virus-1 (HSV-1)-induced cell fusion and virus spread. *Peptides* (1997) 18:177–83. doi: 10.1016/s0196-9781(96) 00290-2

36. Baghian A, Kousoulas KG. Role of the Na+,K+ pump in herpes simplex type 1induced cell fusion: melittin causes specific reversion of syncytial mutants with the syn1 mutation to Syn+ (wild-type) phenotype. Virology (1993) 196:548-56. doi: 10.1006/ viro.1993.1510

37. Uddin MB, Lee BH, Nikapitiya C, Kim JH, Kim TH, Lee HC, et al. Inhibitory effects of bee venom and its components against viruses in *vitro* and in *vivo. J Microbiol* (2016) 54:853–66. doi: 10.1007/s12275-016-6376-1

38. Li Q, Zhao Z, Zhou D, Chen Y, Hong W, Cao L, et al. Virucidal activity of a scorpion venom peptide variant mucroporin-M1 against measles, SARS-CoV and influenza H5N1 viruses. *Peptides* (2011) 32:1518–25. doi: 10.1016/j.peptides.2011.05.015

39. Dempsey CE. The actions of melittin on membranes. *Biochim Biophys Acta* (1990) 1031:143-61. doi: 10.1016/0304-4157(90)90006-x

40. Jenks JA, Goodwin ML, Permar SR. The roles of host and viral antibody fc receptors in herpes simplex virus (HSV) and human cytomegalovirus (HCMV) infections and immunity. *Front Immunol* (2019) 10:2110. doi: 10.3389/fimmu.2019.02110

41. Awasthi S, Belshe RB, Friedman HM. Better neutralization of herpes simplex virus type 1 (HSV-1) than HSV-2 by antibody from recipients of GlaxoSmithKline HSV-2 glycoprotein D2 subunit vaccine. *J Infect Dis* (2014) 210:571–5. doi: 10.1093/infdis/jiu177

42. Birkmann A, Zimmermann H. HSV antivirals - current and future treatment options. *Curr Opin Virol* (2016) 18:9–13. doi: 10.1016/j.coviro.2016.01.013

43. Negi G, Sharma A, Dey M, Dhanawat G, Parveen N. Membrane attachment and fusion of HIV-1, influenza A, and SARS-CoV-2: resolving the mechanisms with biophysical methods. *Biophys Rev* (2022) 14:1109–40. doi: 10.1007/s12551-022-00999-7

44. Galdiero S, Falanga A, Vitiello G, Vitiello M, Pedone C, D'Errico G, et al. Role of membranotropic sequences from herpes simplex virus type I glycoproteins B and H in the fusion process. *Biochim Biophys Acta* (2010) 1798:579–91. doi: 10.1016/j.bbamem.2010.01.006

45. Jayachandra S, Baghian A, Kousoulas KG. Herpes simplex virus type 1 glycoprotein K is not essential for infectious virus production in actively replicating cells but is required for efficient envelopment and translocation of infectious virions from the cytoplasm to the extracellular space. *J Virol* (1997) 71:5012–24. doi: 10.1128/ JVI.71.7.5012-5024.1997

46. Shaposhnikova VV, Egorova MV, Kudryavtsev AA, Levitman M, Korystov Yu N. The effect of melittin on proliferation and death of thymocytes. *FEBS Lett* (1997) 410:285–8. doi: 10.1016/s0014-5793(97)00578-4

47. Ownby CL, Powell JR, Jiang MS, Fletcher JE. Melittin and phospholipase A2 from bee (Apis mellifera) venom cause necrosis of murine skeletal muscle in *vivo*. *Toxicon* (1997) 35:67–80. doi: 10.1016/s0041-0101(96)00078-5

48. Maher S, McClean S. Melittin exhibits necrotic cytotoxicity in gastrointestinal cells which is attenuated by cholesterol. *Biochem Pharmacol* (2008) 75:1104–14. doi: 10.1016/j.bcp.2007.10.029

49. Masenga SK, Mweene BC, Luwaya E, Muchaili L, Chona M, Kirabo A. HIV-host cell interactions. *Cells* (2023) 12:1351. doi: 10.3390/cells12101351

50. Klingler J, Anton H, Real E, Zeiger M, Moog C, Mely Y, et al. How HIV-1 gag manipulates its host cell proteins: A focus on interactors of the nucleocapsid domain. *Viruses* (2020) 12:888. doi: 10.3390/v12080888

51. Wachinger M, Saermark T, Erfle V. Influence of amphipathic peptides on the HIV-1 production in persistently infected T lymphoma cells. *FEBS Lett* (1992) 309:235–41. doi: 10.1016/0014-5793(92)80780-k

52. Wachinger M, Kleinschmidt A, Winder D, von Pechmann N, Ludvigsen A, Neumann M, et al. Antimicrobial peptides melittin and cecropin inhibit replication of human immunodeficiency virus 1 by suppressing viral gene expression. *J Gen Virol* (1998) 79(Pt 4):731–40. doi: 10.1099/0022-1317-79-4-731

53. Hood JL, Jallouk AP, Campbell N, Ratner L, Wickline SA. Cytolytic nanoparticles attenuate HIV-1 infectivity. *Antivir Ther* (2013) 18:95–103. doi: 10.3851/IMP2346

54. Jallouk AP, Moley KH, Omurtag K, Hu G, Lanza GM, Wickline SA, et al. Nanoparticle incorporation of melittin reduces sperm and vaginal epithelium cytotoxicity. *PloS One* (2014) 9:e95411. doi: 10.1371/journal.pone.0095411

55. Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* (2020) 395:497–506. doi: 10.1016/S0140-6736(20)30183-5

56. Ghosh R, Chatterjee S, Dubey S, Lavie CJ. Famotidine against SARS-CoV2: A hope or hype? *Mayo Clin Proc* (2020) 95:1797–9. doi: 10.1016/j.mayocp.2020.05.027

57. Chilamakuri R, Agarwal S. COVID-19: characteristics and therapeutics. *Cells* (2021) 10:206. doi: 10.3390/cells10020206

58. Hu B, Guo H, Zhou P, Shi ZL. Characteristics of SARS-CoV-2 and COVID-19. Nat Rev Microbiol (2021) 19:141–54. doi: 10.1038/s41579-020-00459-7

59. Enayathullah MG, Parekh Y, Banu S, Ram S, Nagaraj R, Kumar BK, et al. Gramicidin S and melittin: potential anti-viral therapeutic peptides to treat SARS-CoV-2 infection. *Sci Rep* (2022) 12:3446. doi: 10.1038/s41598-022-07341-x

60. Pillaiyar T, Manickam M, Namasivayam V, Hayashi Y, Jung SH. An overview of severe acute respiratory syndrome-coronavirus (SARS-coV) 3CL protease inhibitors: peptidomimetics and small molecule chemotherapy. *J Med Chem* (2016) 59:6595–628. doi: 10.1021/acs.jmedchem.5b01461

61. Vuong W, Khan MB, Fischer C, Arutyunova E, Lamer T, Shields J, et al. Feline coronavirus drug inhibits the main protease of SARS-CoV-2 and blocks virus replication. *Nat Commun* (2020) 11:4282. doi: 10.1038/s41467-020-18096-2

62. Al-Rabia MW, Alhakamy NA, Ahmed OAA, Eljaaly K, Alaofi AL, Mostafa A, et al. Repurposing of sitagliptin- melittin optimized nanoformula against SARS-CoV-2: antiviral screening and molecular docking studies. *Pharmaceutics* (2021) 13:307. doi: 10.3390/pharmaceutics13030307

63. Skalickova S, Heger Z, Krejcova L, Pekarik V, Bastl K, Janda J, et al. Perspective of use of antiviral peptides against influenza virus. *Viruses* (2015) 7:5428-42. doi: 10.3390/v7102883

64. Albiol Matanic VC, Castilla V. Antiviral activity of antimicrobial cationic peptides against Junin virus and herpes simplex virus. Int J Antimicrob Agents (2004) 23:382–9. doi: 10.1016/j.ijantimicag.2003.07.022

65. Kumar S, Yadav D, Singh D, Shakya K, Rathi B, Poonam. Recent developments on Junin virus, a causative agent for Argentine haemorrhagic fever. *Rev Med Virol* (2023) 33:e2419. doi: 10.1002/rmv.2419

66. Enria DA, Briggiler AM, Sanchez Z. Treatment of Argentine hemorrhagic fever. *Antiviral Res* (2008) 78:132–9. doi: 10.1016/j.antiviral.2007.10.010

67. Cox JA, Hiscox JA, Solomon T, Ooi MH, Ng LFP. Immunopathogenesis and virus-host interactions of enterovirus 71 in patients with hand, foot and mouth disease. *Front Microbiol* (2017) 8:2249. doi: 10.3389/fmicb.2017.02249

68. Liu CC, Hao DJ, Zhang Q, An J, Zhao JJ, Chen B, et al. Application of bee venom and its main constituent melittin for cancer treatment. *Cancer Chemother Pharmacol* (2016) 78:1113–30. doi: 10.1007/s00280-016-3160-1

69. Islam MR, Akash S, Rahman MM, Nowrin FT, Akter T, Shohag S, et al. Colon cancer and colorectal cancer: Prevention and treatment by potential natural products. *Chem Biol Interact* (2022) 368:110170. doi: 10.1016/j.cbi.2022.110170

70. Yaacoub C, Rifi M, El-Obeid D, Mawlawi H, Sabatier JM, Coutard B, et al. The cytotoxic effect of apis mellifera venom with a synergistic potential of its two main components-melittin and PLA2-on colon cancer HCT116 cell lines. *Molecules* (2021) 26:2264. doi: 10.3390/molecules26082264

71. Rebelo SP, Pinto C, Martins TR, Harrer N, Estrada MF, Loza-Alvarez P, et al. 3D-3-culture: A tool to unveil macrophage plasticity in the tumour microenvironment. *Biomaterials* (2018) 163:185–97. doi: 10.1016/j.biomaterials.2018.02.030

72. Alfaleh MA, Fahmy O, Al-Rabia MW, Abourehab MAS, Ahmed OAA, Fahmy UA, et al. Hybrid nanoparticulate system of Fluvastatin loaded phospholipid, alpha lipoic acid and melittin for the management of colon cancer. *Sci Rep* (2022) 12:19446. doi: 10.1038/s41598-022-24151-3

73. Soliman C, Eastwood S, Truong VK, Ramsland PA, Elbourne A. The membrane effects of melittin on gastric and colorectal cancer. *PloS One* (2019) 14:e0224028. doi: 10.1371/journal.pone.0224028

74. Tang S, Zhou L, He H, Cui L, Ren Z, Tai Y, et al. MnO(2)-melittin nanoparticles serve as an effective anti-tumor immunotherapy by enhancing systemic immune response. *Biomaterials* (2022) 288:121706. doi: 10.1016/j.biomaterials.2022.121706

75. Luo Y, Xu CM, Luo B, Liang G, Zhang Q. Melittin treatment prevents colorectal cancer from progressing in mice through ER stress-mediated apoptosis. *J Pharm Pharmacol* (2023) 75:645–54. doi: 10.1093/jpp/rgad008

76. Rocha MM, Dariva I, Zornoff GC, De Laurentis GS, Mendes GC, Santana MG, et al. A new therapeutic approach for bone metastasis in colorectal cancer: intratumoral melittin. *J Venom Anim Toxins Incl Trop Dis* (2022) 28:e20210067. doi: 10.1590/1678-9199-JVATITD-2021-0067

77. Kong GM, Tao WH, Diao YL, Fang PH, Wang JJ, Bo P, et al. Melittin induces human gastric cancer cell apoptosis via activation of mitochondrial pathway. *World J Gastroenterol* (2016) 22:3186–95. doi: 10.3748/wjg.v22.i11.3186

78. Cairns R, Papandreou I, Denko N. Overcoming physiologic barriers to cancer treatment by molecularly targeting the tumor microenvironment. *Mol Cancer Res* (2006) 4:61–70. doi: 10.1158/1541-7786.MCR-06-0002

79. Kerkar SP, Restifo NP. Cellular constituents of immune escape within the tumor microenvironment. *Cancer Res* (2012) 72:3125–30. doi: 10.1158/0008-5472.CAN-11-4094

80. Tipgomut C, Wongprommoon A, Takeo E, Ittiudomrak T, Puthong S, Chanchao C. Melittin induced G1 cell cycle arrest and apoptosis in Chago-K1 human bronchogenic carcinoma cells and inhibited the differentiation of THP-1 cells into tumour- associated macrophages. *Asian Pac J Cancer Prev* (2018) 19:3427–34. doi: 10.31557/APJCP.2018.19.12.3427

81. Jeong YJ, Choi Y, Shin JM, Cho HJ, Kang JH, Park KK, et al. Melittin suppresses EGF-induced cell motility and invasion by inhibiting PI3K/Akt/mTOR signaling pathway in breast cancer cells. *Food Chem Toxicol* (2014) 68:218–25. doi: 10.1016/j.jfct.2014.03.022

82. Kim S, Choi I, Han IH, Bae H. Enhanced therapeutic effect of optimized melittindKLA, a peptide agent targeting M2-like tumor-associated macrophages in triplenegative breast cancer. *Int J Mol Sci* (2022) 23:15751. doi: 10.3390/ijms232415751

83. Hu X, Zhang Q, Xing W, Wang W. Role of microRNA/IncRNA intertwined with the Wnt/beta-catenin axis in regulating the pathogenesis of triple-negative breast cancer. *Front Pharmacol* (2022) 13:814971. doi: 10.3389/fphar.2022.814971

84. Shah SP, Roth A, Goya R, Oloumi A, Ha G, Zhao Y, et al. The clonal and mutational evolution spectrum of primary triple-negative breast cancers. *Nature* (2012) 486:395–9. doi: 10.1038/nature10933

85. Duffy C, Sorolla A, Wang E, Golden E, Woodward E, Davern K, et al. Honeybee venom and melittin suppress growth factor receptor activation in HER2-enriched and triple-negative breast cancer. *NPJ Precis Oncol* (2020) 4:24. doi: 10.1038/s41698-020-00129-0

86. Daniluk K, Kutwin M, Grodzik M, Wierzbicki M, Strojny B, Szczepaniak J, et al. Use of selected carbon nanoparticles as melittin carriers for MCF-7 and MDA-MB-231 human breast cancer cells. *Mater (Basel).* (2019) 13:90. doi: 10.3390/ma13010090

87. Daniluk K, Lange A, Pruchniewski M, Malolepszy A, Sawosz E, Jaworski S. Delivery of melittin as a lytic agent via graphene nanoparticles as carriers to breast cancer cells. *J Funct Biomater* (2022) 13:278. doi: 10.3390/jfb13040278

88. Dai Y, Yu X, Leng Y, Peng X, Wang J, Zhao Y, et al. Effective treatment of metastatic sentinel lymph nodes by dual-targeting melittin nanoparticles. *J Nanobiotechnol* (2023) 21:245. doi: 10.1186/s12951-023-02026-7

89. Ip SW, Liao SS, Lin SY, Lin JP, Yang JS, Lin ML, et al. The role of mitochondria in bee venom-induced apoptosis in human breast cancer MCF7 cells. *In Vivo* (2008) 22:237–45.

90. Jung GB, Huh JE, Lee HJ, Kim D, Lee GJ, Park HK, et al. Anti-cancer effect of bee venom on human MDA-MB-231 breast cancer cells using Raman spectroscopy. *BioMed Opt Express* (2018) 9:5703–18. doi: 10.1364/BOE.9.005703

91. Moghaddam FD, Mortazavi P, Hamedi S, Nabiuni M, Roodbari NH. Apoptotic Effects of Melittin on 4T1 Breast Cancer Cell Line is associated with Up Regulation of Mfn1 and Drp1 mRNA Expression. *Anticancer Agents Med Chem* (2020) 20:790–9. doi: 10.2174/1871520620666200211091451

92. Mir Hassani Z, Nabiuni M, Parivar K, Abdirad S, Karimzadeh L. Melittin inhibits the expression of key genes involved in tumor microenvironment formation by suppressing HIF-1alpha signaling in breast cancer cells. *Med Oncol* (2021) 38:77. doi: 10.1007/s12032-021-01526-6

93. Chen C, Pore N, Behrooz A, Ismail-Beigi F, Maity A. Regulation of glut1 mRNA by hypoxia-inducible factor-1. Interaction between H-ras and hypoxia. *J Biol Chem* (2001) 276:9519–25. doi: 10.1074/jbc.M010144200

94. Lv C, Chen J, Huang F, Fang F, Li B. Melittin inhibits the proliferation migration and invasion of HCC cells by regulating ADAMTS9-AS2 demethylation. *Toxicon* (2023) 222:106996. doi: 10.1016/j.toxicon.2022.106996

95. Singal AG, Llovet JM, Yarchoan M, Mehta N, Heimbach JK, Dawson LA, et al. AASLD Practice Guidance on prevention, diagnosis, and treatment of hepatocellular carcinoma. *Hepatology* (2023) 78:1922–65. doi: 10.1097/HEP.000000000000466

96. Abdel-Rahman O, Lamarca A. Development of sorafenib-related side effects in patients diagnosed with advanced hepatocellular carcinoma treated with sorafenib: a systematic-review and meta-analysis of the impact on survival. *Expert Rev Gastroenterol Hepatol* (2017) 11:75–83. doi: 10.1080/17474124.2017.1264874

97. Mansour GH, El-Magd MA, Mahfouz DH, Abdelhamid IA, Mohamed MF, Ibrahim NS, et al. Bee venom and its active component Melittin synergistically potentiate the anticancer effect of Sorafenib against HepG2 cells. *Bioorg Chem* (2021) 116:105329. doi: 10.1016/j.bioorg.2021.105329

98. Notarbartolo M, Cervello M, Giannitrapani L, Meli M, Poma P, Dusonchet L, et al. Expression of IAPs and alternative splice variants in hepatocellular carcinoma tissues and cells. *Ann N Y Acad Sci* (2004) 1028:289–93. doi: 10.1196/annals.1322.033

99. Shi YH, Ding WX, Zhou J, He JY, Xu Y, Gambotto AA, et al. Expression of Xlinked inhibitor-of-apoptosis protein in hepatocellular carcinoma promotes metastasis and tumor recurrence. *Hepatology* (2008) 48:497–507. doi: 10.1002/hep.22393

100. Li G, Chang H, Zhai YP, Xu W. Targeted silencing of inhibitors of apoptosis proteins with siRNAs: a potential anti-cancer strategy for hepatocellular carcinoma. *Asian Pac J Cancer Prev* (2013) 14:4943–52. doi: 10.7314/apjcp.2013.14.9.4943

101. Zhang Z, Zhang H, Peng T, Li D, Xu J. Melittin suppresses cathepsin S-induced invasion and angiogenesis via blocking of the VEGF-A/VEGFR-2/MEK1/ERK1/2 pathway in human hepatocellular carcinoma. *Oncol Lett* (2016) 11:610–8. doi: 10.3892/ol.2015.3957

102. Petanceska S, Canoll P, Devi LA. Expression of rat cathepsin S in phagocytic cells. J Biol Chem (1996) 271:4403–9. doi: 10.1074/jbc.271.8.4403

103. Fuchs N, Meta M, Schuppan D, Nuhn L, Schirmeister T. Novel opportunities for cathepsin S inhibitors in cancer immunotherapy by nanocarrier-mediated delivery. *Cells* (2020) 9:2021. doi: 10.3390/cells9092021

104. Xu J, Li D, Ke Z, Liu R, Maubach G, Zhuo L. Cathepsin S is aberrantly overexpressed in human hepatocellular carcinoma. *Mol Med Rep* (2009) 2:713–8. doi: 10.3892/mmr_00000161

105. Burden RE, Gormley JA, Kuehn D, Ward C, Kwok HF, Gazdoiu M, et al. Inhibition of Cathepsin S by Fsn0503 enhances the efficacy of chemotherapy in colorectal carcinomas. *Biochimie* (2012) 94:487–93. doi: 10.1016/j.biochi.2011.08.017

106. Fan Q, Wang X, Zhang H, Li C, Fan J, Xu J. Silencing cathepsin S gene expression inhibits growth, invasion and angiogenesis of human hepatocellular carcinoma in *vitro. Biochem Biophys Res Commun* (2012) 425:703–10. doi: 10.1016/ j.bbrc.2012.07.013

107. Song Y, Bi Z, Liu Y, Qin F, Wei Y, Wei X. Targeting RAS-RAF-MEK-ERK signaling pathway in human cancer: Current status in clinical trials. *Genes Dis* (2023) 10:76–88. doi: 10.1016/j.gendis.2022.05.006

108. Zhang H, Zhao B, Huang C, Meng XM, Bian EB, Li J. Melittin restores PTEN expression by down-regulating HDAC2 in human hepatocelluar carcinoma HepG2 cells. *PloS One* (2014) 9:e95520. doi: 10.1371/journal.pone.0095520

109. Halley F, Reinshagen J, Ellinger B, Wolf M, Niles AL, Evans NJ, et al. A bioluminogenic HDAC activity assay: validation and screening. *J Biomol Screen* (2011) 16:1227–35. doi: 10.1177/1087057111416004

110. Shan W, Jiang Y, Yu H, Huang Q, Liu L, Guo X, et al. HDAC2 overexpression correlates with aggressive clinicopathological features and DNA-damage response pathway of breast cancer. *Am J Cancer Res* (2017) 7:1213–26.

111. Zhu P, Martin E, Mengwasser J, Schlag P, Janssen KP, Gottlicher M. Induction of HDAC2 expression upon loss of APC in colorectal tumorigenesis. *Cancer Cell* (2004) 5:455–63. doi: 10.1016/s1535-6108(04)00114-x

112. Jo H, Shim K, Kim HU, Jung HS, Jeoung D. HDAC2 as a target for developing anti-cancer drugs. *Comput Struct Biotechnol J* (2023) 21:2048–57. doi: 10.1016/ j.csbj.2023.03.016

113. Noh JH, Bae HJ, Eun JW, Shen Q, Park SJ, Kim HS, et al. HDAC2 provides a critical support to Malignant progression of hepatocellular carcinoma through feedback control of mTORC1 and AKT. *Cancer Res* (2014) 74:1728–38. doi: 10.1158/0008-5472.CAN-13-2109

114. Lv C, Zhang Z, Zhao T, Han MF, Jia DP, Su LZ, et al. The anti-tumour effect of Mel and its role in autophagy in human hepatocellular carcinoma cells. *Am J Transl Res* (2019) 11:931–41.

115. Xie S, Yu X, Li Y, Ma H, Fan S, Chen W, et al. Upregulation of lncRNA ADAMTS9-AS2 Promotes Salivary Adenoid Cystic Carcinoma Metastasis via PI3K/ Akt and MEK/Erk Signaling. *Mol Ther* (2018) 26:2766–78. doi: 10.1016/ j.ymthe.2018.08.018

116. Ma N, Xu E, Luo Q, Song G. Rac1: A regulator of cell migration and a potential target for cancer therapy. *Molecules* (2023) 28:2976. doi: 10.3390/molecules28072976

117. Zhao H, Feng X, Han W, Diao Y, Han D, Tian X, et al. Enhanced binding to and killing of hepatocellular carcinoma cells in *vitro* by melittin when linked with a novel targeting peptide screened from phage display. *J Pept Sci* (2013) 19:639–50. doi: 10.1002/psc.2542

118. Zhao X, Yu Z, Dai W, Yao Z, Zhou W, Zhou W, et al. Construction and characterization of an anti-asialoglycoprotein receptor single-chain variable-fragment-targeted melittin. *Biotechnol Appl Biochem* (2011) 58:405–11. doi: 10.1002/bab.57

119. Qian CY, Wang KL, Fang FF, Gu W, Huang F, Wang FZ, et al. Triplecontrolled oncolytic adenovirus expressing melittin to exert inhibitory efficacy on hepatocellular carcinoma. *Int J Clin Exp Pathol* (2015) 8:10403–11.

120. Qu L, Jiang M, Li Z, Pu F, Gong L, Sun L, et al. Inhibitory effect of biosynthetic nanoscale peptide Melittin on hepatocellular carcinoma, driven by survivin promoter. *J BioMed Nanotechnol* (2014) 10:695–706. doi: 10.1166/jbn.2014.1830

121. Ling CQ, Li B, Zhang C, Zhu DZ, Huang XQ, Gu W, et al. Inhibitory effect of recombinant adenovirus carrying melittin gene on hepatocellular carcinoma. *Ann Oncol* (2005) 16:109–15. doi: 10.1093/annonc/mdi019

122. Li B, Gu W, Zhang C, Huang XQ, Han KQ, Ling CQ. Growth arrest and apoptosis of the human hepatocellular carcinoma cell line BEL-7402 induced by melittin. *Onkologie* (2006) 29:367–71. doi: 10.1159/000094711

123. Park MH, Choi MS, Kwak DH, Oh KW, Yoon DY, Han SB, et al. Anti-cancer effect of bee venom in prostate cancer cells through activation of caspase pathway via inactivation of NF-kappaB. *Prostate* (2011) 71:801–12. doi: 10.1002/pros.21296

124. Fu Y, An N, Li K, Zheng Y, Liang A. Chlorotoxin-conjugated nanoparticles as potential glioma-targeted drugs. *J Neurooncol* (2012) 107:457–62. doi: 10.1007/s11060-011-0763-6

125. Deshane J, Garner CC, Sontheimer H. Chlorotoxin inhibits glioma cell invasion via matrix metalloproteinase-2. *J Biol Chem* (2003) 278:4135–44. doi: 10.1074/ jbc.M205662200

126. Tarokh Z, Naderi-Manesh H, Nazari M. Towards prostate cancer gene therapy: Development of a chlorotoxin-targeted nanovector for toxic (melittin) gene delivery. *Eur J Pharm Sci* (2017) 99:209–18. doi: 10.1016/j.ejps.2016.12.021

127. Yan R, Dai W, Mao Y, Yu G, Li W, Shu M, et al. Melittin inhibits tumor cell migration and enhances cisplatin sensitivity by suppressing IL-17 signaling pathway gene LCN2 in castration-resistant prostate cancer. *Prostate* (2023) 83:1430–45. doi: 10.1002/pros.24605

128. Moon DO, Park SY, Lee KJ, Heo MS, Kim KC, Kim MO, et al. Bee venom and melittin reduce proinflammatory mediators in lipopolysaccharide-stimulated BV2 microglia. *Int Immunopharmacol* (2007) 7:1092–101. doi: 10.1016/j.intimp.2007.04.005

129. Raetz CR, Whitfield C. Lipopolysaccharide endotoxins. Annu Rev Biochem (2002) 71:635–700. doi: 10.1146/annurev.biochem.71.110601.135414

130. Han SM, Kim JM, Park KK, Chang YC, Pak SC. Neuroprotective effects of melittin on hydrogen peroxide-induced apoptotic cell death in neuroblastoma SH-SY5Y cells. BMC Complement Altern Med (2014) 14:286. doi: 10.1186/1472-6882-14-286

131. Yang EJ, Kim SH, Yang SC, Lee SM, Choi SM. Melittin restores proteasome function in an animal model of ALS. *J Neuroinflamm* (2011) 8:69. doi: 10.1186/1742-2094-8-69

132. Cawston T. Matrix metalloproteinases and TIMPs: properties and implications for the rheumatic diseases. *Mol Med Today* (1998) 4:130–7. doi: 10.1016/s1357-4310 (97)01192-1

133. Nah SS, Ha E, Mun SH, Won HJ, Chung JH. Effects of melittin on the production of matrix metalloproteinase-1 and -3 in rheumatoid arthritic fibroblast-like synoviocytes. J Pharmacol Sci (2008) 106:162–6. doi: 10.1254/jphs.sc0070215

134. He SD, Tan N, Sun CX, Liao KH, Zhu HJ, Luo XG, et al. Treatment with melittin induces apoptosis and autophagy of fibroblastlike synoviocytes in patients with rheumatoid arthritis. *Curr Pharm Biotechnol* (2020) 21:734–40. doi: 10.2174/1389201021666191210110826

135. Krause A, Scaletta N, Ji JD, Ivashkiv LB. Rheumatoid arthritis synoviocyte survival is dependent on Stat3. *J Immunol* (2002) 169:6610–6. doi: 10.4049/jimmunol.169.11.6610

136. Nishimoto N, Hashimoto J, Miyasaka N, Yamamoto K, Kawai S, Takeuchi T, et al. Study of active controlled monotherapy used for rheumatoid arthritis, an IL-6 inhibitor (SAMURAI): evidence of clinical and radiographic benefit from an x ray reader-blinded randomised controlled trial of tocilizumab. *Ann Rheum Dis* (2007) 66:1162–7. doi: 10.1136/ard.2006.068064

137. Emery P, Keystone E, Tony HP, Cantagrel A, van Vollenhoven R, Sanchez A, et al. IL-6 receptor inhibition with tocilizumab improves treatment outcomes in patients with rheumatoid arthritis refractory to anti-tumour necrosis factor biologicals: results from a 24-week multicentre randomised placebo-controlled trial. Ann Rheum Dis (2008) 67:1516-23. doi: 10.1136/ard.2008.092932

138. Mihara M, Moriya Y, Kishimoto T, Ohsugi Y. Interleukin-6 (IL-6) induces the proliferation of synovial fibroblastic cells in the presence of soluble IL-6 receptor. *Br J Rheumatol* (1995) 34:321–5. doi: 10.1093/rheumatology/34.4.321

139. Kim SK, Park KY, Yoon WC, Park SH, Park KK, Yoo DH, et al. Melittin enhances apoptosis through suppression of IL-6/sIL-6R complex-induced NF-kappaB and STAT3 activation and Bcl-2 expression for human fibroblast-like synoviocytes in rheumatoid arthritis. *Joint Bone Spine* (2011) 78:471–7. doi: 10.1016/j.jbspin.2011.01.004

140. Feng N, Guo F. Nanoparticle-siRNA: A potential strategy for rheumatoid arthritis therapy? J Control Release (2020) 325:380-93. doi: 10.1016/j.jconrel.2020.07.006

141. Park HJ, Son DJ, Lee CW, Choi MS, Lee US, Song HS, et al. Melittin inhibits inflammatory target gene expression and mediator generation via interaction with IkappaB kinase. *Biochem Pharmacol* (2007) 73:237–47. doi: 10.1016/j.bcp.2006.09.023

142. Maracle CX, Kucharzewska P, Helder B, van der Horst C, Correa de Sampaio P, Noort AR, et al. Targeting non-canonical nuclear factor-kappaB signalling attenuates neovascularization in a novel 3D model of rheumatoid arthritis synovial angiogenesis. *Rheumatol (Oxford)* (2017) 56:294–302. doi: 10.1093/rheumatology/kew393

143. Park HJ, Lee HJ, Choi MS, Son DJ, Song HS, Song MJ, et al. JNK pathway is involved in the inhibition of inflammatory target gene expression and NF-kappaB activation by melittin. *J Inflammation (Lond)* (2008) 5:7. doi: 10.1186/1476-9255-5-7

144. Le Berre C, Honap S, Peyrin-Biroulet L. Ulcerative colitis. Lancet (2023) 402:571-84. doi: 10.1016/S0140-6736(23)00966-2

145. Ahmedy OA, Ibrahim SM, Salem HH, Kandil EA. Antiulcerogenic effect of melittin via mitigating TLR4/TRAF6 mediated NF-kappaB and p38MAPK pathways in acetic acid-induced ulcerative colitis in mice. *Chem Biol Interact* (2020) 331:109276. doi: 10.1016/j.cbi.2020.109276

146. Yaghoubi A, Amel Jamehdar S, Reza Akbari Eidgahi M, Ghazvini K. Evaluation of the therapeutic effect of melittin peptide on the ulcerative colitis mouse model. *Int Immunopharmacol* (2022) 108:108810. doi: 10.1016/j.intimp.2022.108810

147. Rolando N, Wade J, Davalos M, Wendon J, Philpott-Howard J, Williams R. The systemic inflammatory response syndrome in acute liver failure. *Hepatology* (2000) 32:734–9. doi: 10.1053/jhep.2000.17687

148. Tacke F. Targeting hepatic macrophages to treat liver diseases. *J Hepatol* (2017) 66:1300–12. doi: 10.1016/j.jhep.2017.02.026

149. Park JH, Kim KH, Lee WR, Han SM, Park KK. Protective effect of melittin on inflammation and apoptosis in acute liver failure. *Apoptosis* (2012) 17:61–9. doi: 10.1007/s10495-011-0659-0

150. Fan XG, Pei SY, Zhou D, Zhou PC, Huang Y, Hu XW, et al. Melittin ameliorates inflammation in mouse acute liver failure via inhibition of PKM2-mediated Warburg effect. *Acta Pharmacol Sin* (2021) 42:1256–66. doi: 10.1038/s41401-020-00516-0

151. Lee WR, Kim KH, An HJ, Kim JY, Han SM, Lee KG, et al. Protective effect of melittin against inflammation and apoptosis on Propionibacterium acnes-induced human THP-1 monocytic cell. *Eur J Pharmacol* (2014) 740:218–26. doi: 10.1016/ j.ejphar.2014.06.058

152. Lee WR, Kim KH, An HJ, Kim JY, Chang YC, Chung H, et al. The protective effects of melittin on Propionibacterium acnes-induced inflammatory responses in *vitro* and in *vivo. J Invest Dermatol* (2014) 134:1922–30. doi: 10.1038/jid.2014.75

153. Weidinger S, Novak N. Atopic dermatitis. Lancet (2016) 387:1109-22. doi: 10.1016/S0140-6736(15)00149-X

154. Leung DY, Bieber T. Atopic dermatitis. Lancet (2003) 361:151-60. doi: 10.1016/ S0140-6736(03)12193-9

155. Kim WH, An HJ, Kim JY, Gwon MG, Gu H, Jeon M, et al. Beneficial effects of melittin on ovalbumin-induced atopic dermatitis in mouse. *Sci Rep* (2017) 7:17679. doi: 10.1038/s41598-017-17873-2

156. Amano A, Kuboniwa M, Nakagawa I, Akiyama S, Morisaki I, Hamada S. Prevalence of specific genotypes of Porphyromonas gingivalis fimA and periodontal health status. *J Dent Res* (2000) 79:1664–8. doi: 10.1177/00220345000790090501

157. Kim WH, An HJ, Kim JY, Gwon MG, Gu H, Jeon M, et al. Anti-inflammatory effect of melittin on porphyromonas gingivalis LPS-stimulated human keratinocytes. *Molecules* (2018) 23:332. doi: 10.3390/molecules23020332

158. Gajski G, Domijan AM, Zegura B, Stern A, Geric M, Novak Jovanovic I, et al. Melittin induced cytogenetic damage, oxidative stress and changes in gene expression in human peripheral blood lymphocytes. *Toxicon* (2016) 110:56–67. doi: 10.1016/ j.toxicon.2015.12.005

159. Pezzella M, El Hage K, Niesen MJM, Shin S, Willard AP, Meuwly M, et al. Water dynamics around proteins: T- and R-states of hemoglobin and melittin. *J Phys Chem B* (2020) 124:6540–54. doi: 10.1021/acs.jpcb.0c04320

160. Chen J, Guan SM, Sun W, Fu H. Melittin, the major pain-producing substance of bee venom. *Neurosci Bull* (2016) 32:265–72. doi: 10.1007/s12264-016-0024-y

161. Li KC, Chen J. Altered pain-related behaviors and spinal neuronal responses produced by s.c. injection of melittin in rats. *Neuroscience* (2004) 126:753–62. doi: 10.1016/j.neuroscience.2004.03.050

162. Sumikura H, Andersen OK, Drewes AM, Arendt-Nielsen L. A comparison of hyperalgesia and neurogenic inflammation induced by melittin and capsaicin in humans. *Neurosci Lett* (2003) 337:147–50. doi: 10.1016/s0304-3940(02)01325-3

163. Sumikura H, Andersen OK, Drewes AM, Arendt-Nielsen L. Secondary heat hyperalgesia induced by melittin in humans. *Eur J Pain* (2006) 10:121–5. doi: 10.1016/ j.ejpain.2005.01.013

164. Yu YQ, Zhao ZY, Chen XF, Xie F, Yang Y, Chen J. Activation of tetrodotoxinresistant sodium channel NaV1.9 in rat primary sensory neurons contributes to melittin-induced pain behavior. *Neuromol Med* (2013) 15:209–17. doi: 10.1007/ s12017-012-8211-0

165. Guha S, Ferrie RP, Ghimire J, Ventura CR, Wu E, Sun L, et al. Applications and evolution of melittin, the quintessential membrane active peptide. *Biochem Pharmacol* (2021) 193:114769. doi: 10.1016/j.bcp.2021.114769

166. Perez-Cordero JJ, Lozano JM, Cortes J, Delgado G. Leishmanicidal activity of synthetic antimicrobial peptides in an infection model with human dendritic cells. *Peptides* (2011) 32:683–90. doi: 10.1016/j.peptides.2011.01.011

167. Pereira AV, de Barros G, Pinto EG, Tempone AG, Orsi Rde O, Dos Santos LD, et al. Melittin induces in *vitro* death of Leishmania (Leishmania) infantum by triggering the cellular innate immune response. *J Venom Anim Toxins Incl Trop Dis* (2016) 22:1. doi: 10.1186/s40409-016-0055-x

168. Gui Z, Zhu J, Ye S, Ye J, Chen J, Ling Y, et al. Prolonged melittin release from polyelectrolyte-based nanocomplexes decreases acute toxicity and improves blood glycemic control in a mouse model of type II diabetes. *Int J Pharm* (2020) 577:119071. doi: 10.1016/j.ijpharm.2020.119071

169. Ye R, Zheng Y, Chen Y, Wei X, Shi S, Chen Y, et al. Stable loading and delivery of melittin with lipid-coated polymeric nanoparticles for effective tumor therapy with negligible systemic toxicity. *ACS Appl Mater Interf* (2021) 13:55902–12. doi: 10.1021/acsami.1c17618

170. Li Y, Ruan S, Wang Z, Feng N, Zhang Y. Hyaluronic acid coating reduces the leakage of melittin encapsulated in liposomes and increases targeted delivery to melanoma cells. *Pharmaceutics* (2021) 13:1235. doi: 10.3390/pharmaceutics13081235

171. Park K, Lee MY, Kim KS, Hahn SK. Target specific tumor treatment by VEGF siRNA complexed with reducible polyethyleneimine-hyaluronic acid conjugate. *Biomaterials* (2010) 31:5258–65. doi: 10.1016/j.biomaterials.2010.03.018

172. Naor D, Wallach-Dayan SB, Zahalka MA, Sionov RV. Involvement of CD44, a molecule with a thousand faces, in cancer dissemination. *Semin Cancer Biol* (2008) 18:260–7. doi: 10.1016/j.semcancer.2008.03.015

173. Koltai T, Reshkin SJ, Carvalho TMA, Cardone RA. Targeting the stromal protumoral hyaluronan-CD44 pathway in pancreatic cancer. *Int J Mol Sci* (2021) 22:3953. doi: 10.3390/ijms22083953

174. Surace C, Arpicco S, Dufay-Wojcicki A, Marsaud V, Bouclier C, Clay D, et al. Lipoplexes targeting the CD44 hyaluronic acid receptor for efficient transfection of breast cancer cells. *Mol Pharm* (2009) 6:1062–73. doi: 10.1021/mp800215d

175. Schmieder A, Michel J, Schonhaar K, Goerdt S, Schledzewski K. Differentiation and gene expression profile of tumor-associated macrophages. *Semin Cancer Biol* (2012) 22:289–97. doi: 10.1016/j.semcancer.2012.02.002

176. Liu H, Hu Y, Sun Y, Wan C, Zhang Z, Dai X, et al. Co-delivery of bee venom melittin and a photosensitizer with an organic-inorganic hybrid nanocarrier for photodynamic therapy and immunotherapy. *ACS Nano* (2019) 13:12638–52. doi: 10.1021/acsnano.9b04181

177. Lee C, Jeong H, Bae Y, Shin K, Kang S, Kim H, et al. Targeting of M2-like tumor-associated macrophages with a melittin-based pro-apoptotic peptide. *J Immunother Cancer* (2019) 7:147. doi: 10.1186/s40425-019-0610-4

178. Jeong C, Kim J, Han IH, Kim S, Choi I, Kim H, et al. Melittin derived peptidedrug conjugate, M-DM1, inhibits tumor progression and induces effector cell infiltration in melanoma by targeting M2 tumor-associated macrophages. *Front Immunol* (2023) 14:1178776. doi: 10.3389/fimmu.2023.1178776

179. Raveendran R, Chen F, Kent B, Stenzel MH. Estrone-decorated polyion complex micelles for targeted melittin delivery to hormone-responsive breast cancer cells. *Biomacromolecules* (2020) 21:1222–33. doi: 10.1021/acs.biomac.9b01681

180. Nabizadeh S, Rahbarnia L, Nowrozi J, Farajnia S, Hosseini F. Rational design of hybrid peptide with high antimicrobial property derived from Melittin and Lasioglossin. J Biomol Struct Dyn (2023), 1–9. doi: 10.1080/07391102.2023.2274971

181. Huang C, Jin H, Qian Y, Qi S, Luo H, Luo Q, et al. Hybrid melittin cytolytic Peptide-driven ultrasmall lipid nanoparticles block melanoma growth in *vivo. ACS Nano* (2013) 7:5791–800. doi: 10.1021/nn400683s

182. Yu X, Dai Y, Zhao Y, Qi S, Liu L, Lu L, et al. Melittin-lipid nanoparticles target to lymph nodes and elicit a systemic anti-tumor immune response. *Nat Commun* (2020) 11:1110. doi: 10.1038/s41467-020-14906-9

183. Liu Z, Fan Z, Liu J, Wang J, Xu M, Li X, et al. Melittin-carrying nanoparticle suppress T cell-driven immunity in a murine allergic dermatitis model. *Adv Sci* (*Weinh*) (2023) 10:e2204184. doi: 10.1002/advs.202204184

184. Li M, Zhang X, Zheng X, Lian D, Zhang ZX, Ge W, et al. Immune modulation and tolerance induction by RelB-silenced dendritic cells through RNA interference. *J Immunol* (2007) 178:5480–7. doi: 10.4049/jimmunol.178.9.5480

185. Sun M, Wu Y, Zhou Z, Liu S, Mao S, Li G. Co-delivery of EGCG and melittin with self-assembled fluoro-nanoparticles for enhanced cancer therapy. *Aging (Albany NY)* (2023) 15:4875–88. doi: 10.18632/aging.204769

186. Ravindran Menon D, Li Y, Yamauchi T, Osborne DG, Vaddi PK, Wempe MF, et al. EGCG inhibits tumor growth in melanoma by targeting JAK-STAT signaling and its downstream PD-L1/PD-L2-PD1 axis in tumors and enhancing cytotoxic T-cell responses. *Pharm (Basel).* (2021) 14:1081. doi: 10.3390/ph14111081

187. Rawangkan A, Wongsirisin P, Namiki K, Iida K, Kobayashi Y, Shimizu Y, et al. Green tea catechin is an alternative immune checkpoint inhibitor that inhibits PD-L1 expression and lung tumor growth. *Molecules* (2018) 23:2071. doi: 10.3390/molecules23082071

188. Wang H, Wang X, Xie C, Zhang M, Ruan H, Wang S, et al. Nanodisk-based glioma-targeted drug delivery enabled by a stable glycopeptide. *J Control Release* (2018) 284:26–38. doi: 10.1016/j.jconrel.2018.06.006

189. Wang H, Wang S, Wang R, Wang X, Jiang K, Xie C, et al. Co-delivery of paclitaxel and melittin by glycopeptide-modified lipodisks for synergistic anti-glioma therapy. *Nanoscale* (2019) 11:13069–77. doi: 10.1039/c9nr01820d

190. Motiei M, Aboutalebi F, Forouzanfar M, Dormiani K, Nasr-Esfahani MH, Mirahmadi-Zare SZ. Smart co-delivery of miR-34a and cytotoxic peptides (LTX-315 and melittin) by chitosan based polyelectrolyte nanocarriers for specific cancer cell death induction. *Mater Sci Eng C Mater Biol Appl* (2021) 128:112258. doi: 10.1016/j.msec.2021.112258

191. Sveinbjornsson B, Camilio KA, Haug BE, Rekdal O. LTX-315: a first-in-class oncolytic peptide that reprograms the tumor microenvironment. *Future Med Chem* (2017) 9:1339–44. doi: 10.4155/fmc-2017-0088

192. Lin X, Chen W, Wei F, Zhou BP, Hung MC, Xie X. Nanoparticle Delivery of miR-34a Eradicates Long-term-cultured Breast Cancer Stem Cells via Targeting C22ORF28 Directly. *Theranostics* (2017) 7:4805–24. doi: 10.7150/thno.20771

193. Asfour HZ, Alhakamy NA, Ahmed OAA, Fahmy UA, Md S, El-Moselhy MA, et al. Enhanced healing efficacy of an optimized gabapentin-melittin nanoconjugate gelloaded formulation in excised wounds of diabetic rats. *Drug Deliv* (2022) 29:1892–902. doi: 10.1080/10717544.2022.2086943

194. Gajski G, Garaj-Vrhovac V. Melittin: a lytic peptide with anticancer properties. *Environ Toxicol Pharmacol* (2013) 36:697–705. doi: 10.1016/j.etap.2013.06.009

195. Riepl M. Compounding pearls – wound care: drugs and formulations for dermal healing, part 2. Int J Pharm Compd (2021) 25:282–7.

196. Maiden MM, Zachos MP, Waters CM. Hydrogels embedded with melittin and tobramycin are effective against Pseudomonas aeruginosa biofilms in an animal wound model. *Front Microbiol* (2019) 10:1348. doi: 10.3389/fmicb.2019.01348