# Cuticular hydrocarbon analysis in forensic entomology: A Review

Hannah Moore[1] and Sue Shemilt[2]

[1]Cranfield Forensic Institute, Cranfield University, Defence Academy of the United Kingdom, Shrivenham, Wiltshire, SN7 8LA, UK
h.e.moore@cranfield.ac.uk

[2]Department of Chemical Ecology, School of Chemical and Physical Science, Keele University, ST5 5BG, England, UK

## Abstract

Blowflies are the first inhabitants of decomposing remains and are therefore of forensic relevance for post mortem interval estimations. Forensic entomology is becoming widely accepted as a branch of forensic science and is being utilised more within forensic casework. This wider use has driven an increase in research being carried out within the field, in particular, in less "classical" techniques such as DNA and chemical analysis in the form of cuticular hydrocarbon analysis. This short review will examine the research currently being studied in the area of cuticular hydrocarbon analysis of forensically important Diptera for species identification and ageing.

**Key words**: Forensic Entomology, PMI estimations, Blowflies, Cuticular Hydrocarbon Analysis

## Introduction

Forensic entomology is the study of insects that utilise decomposing remains to aid legal investigation (Amendt *et al.* 2000; Moore 2013) and its main use has been the estimation of the minimum time since death or minimum post mortem interval estimation (minPMI) (Hart *et al.* 2008). Although the use of insects in criminal investigations has been recorded as early as the 13th Century (Benecke 2001), it is only in the past few decades that this branch of forensic science has gained importance and is being utilised more by police forces for scenes of homicide and abuse (Amendt *et al.* 2000). Due to the abundance of insect activity associated with decomposing remains, they offer a variety of evidence (Moore 2013). Blow flies, among the first species to exploit dead remains (Benecke 2001), can provide information ranging from how long

a person has been dead (Gennard 2007) to determining whether the deceased had ingested drugs prior to death (Hart *et al*. 2008).

The quality of this minPMI evidence, and the forensic entomologist's ability to estimate time since the first fly accessed the body is dependent on accurate insect species identification, their ability to determine the age of the oldest live colonising larvae and their possession of a thorough understanding of entomology, in terms of the ecology and the biology of insects and their environment (Hart *et al*. 2008).

The identification of larvae is usually completed by their collection from the dead host or surrounding area and their subsequent rearing to adult flies. While larvae can be difficult to identify by species morphologically, adult flies are more readily identified via published genital keys (Adams & Hall 2003). This species identification is important because different species have different life cycle timings therefore in order to utilise the correct developmental information, species identification is first required (Ames *et al*. 2006a). The time since colonization is then calculated, which is the estimate based on the age of the most developed insects present when the body is discovered. Current methods for ageing insect specimens involve measuring the length of the larvae or by dissecting any collected pupae, and comparing the measurements to published data in relation to temperature (Hart *et al.* 2008).

In recent years, there has been an increase in developments around DNA-based techniques (Ames *et al*. 2006b; Cainé *et al*. 2009; Harvey *et al*. 2003; Malgorn & Coquoz 1999; Ratcliffe *et al*. 2003; Sperling *et al*. 1994; Stevens & Wall 2001; Vincent *et al*. 2000; Wallman & Donnellan 2001; Wells & Sperling 2001) and chemical analysis research within forensic entomology with the aim of complimenting or enhancing current techniques used within this field of study. This review will concentrate on the analysis of cuticular hydrocarbons (CHC) for the purpose of identification and ageing forensically important Diptera.

Cuticular hydrocarbons are present within the epicuticular wax layer of insect cuticles. This hydrophobic, flexible wax layer also consists of fatty acids and alcohols (Gibbs 1998; Moore 2013). The epicuticular lipids serve a variety of roles in different insects; within social insects,

such as ants, their primary function is to protect against dessication (Blomquist & Bagnères 2010), however research has also suggested that they are used for communication (Martin *et al.* 2008). As well as preventing dessication, the cuticular lipid layer also serves as protection against micro-organisms (Gibbs & Crockett 1998). There are a vast number of different hydrocarbons present on the cuticles of insects, with many different possible combinations. This indicates that potentially each species of insect could have its own unique chemical profile, often referred to as a fingerprint (Everaerts *et al.* 1997; Martin *et al.* 2008; Moore 2013).

The cuticular hydrocarbons of insects are generally long linear molecules, typically ranging in chain lengths of C16 to C37 for Calliphoridae species (Howard & Blomquist 2005). They can be either saturated or unsaturated and can have one or more methyl groups attached to the carbon backbone of the chain. In their saturated form, *n*-alkanes consist of single bonded carbon atoms. In their unsaturated form there can be one (alkene), two (alkadiene) or three (alkatriene) double bonds present along the carbon chain (Moore 2013).

Cuticular hydrocarbon analysis is commonly carried out using Gas Chromatography – Mass Spectrometry (GC-MS) which is a well-established technique that provides separation, identification and characterisation and is widely used within chemistry and forensic science laboratories world-wide (Martin & Drijfhout 2009).

**Cuticular Hydrocarbon analysis for species identification**

There are a number of studies on cuticular chemistry, especially concerning social insects such as ants (Akino *et al.* 2004; Guillem *et al.* 2012; Martin et al. 2008; Tissot et al. 2001), bees (Lavine & Vora 2005), termites (Haverty *et al.* 1997), and wasps (Bernier *et al.* 1998). Other insects have also been well studied such as cockroaches (Everaerts *et al.* 1997), grasshoppers (Chapman & Espelies 1995) and beetles (Lockey 1991, 1992; Page *et al.* 1997). Some of these publications have successfully demonstrated how the analysis of CHCs via the use of GC-MS can provide efficient species identification. A key study was that by Guillem *et al.* (Guillem *et al.* 2012) which proposed the use of chemotaxonomy to address the challenges of morphological identification between ant species. The two chosen ant species, *Myrmica sabuleti* and *M. scabrinodis* are morphologically very similar but by analysing their chemical profile via GC-MS, the presence or absence of two species-specific compounds meant that the two species were

easily distinguishable.

Amongst the aforementioned insects, CHC analysis has also been used for identification purposes of Diptera (Urech *et al*. 2005) as a complimentary technique when the taxonomical identification is not possible.

Ye *et al*. (Ye *et al*. 2007) were the first to publish data from the empty puparial cases of six species of necrophagous flies from China: *Aldrichina graham*, *Chrysomya. megacephala*, *Lucilia sericata*, *Achoetandrus rufifacies*, *Boettcherisca peregrina* and *Parasarcophaga crassipalpis*. A discriminant model yielding eight cuticular hydrocarbon compounds was constructed from the initial extracted GC-MS profiles, allowing for complete separation of the pupal exuviae from the six species. The results were very promising and led to further investigations utilising this fingerprint chemical identification approach within the field of forensic entomology.

Identifying empty puparial cases using taxonomical methods can be challenging as many of the morphological features required are damaged due to the mechanism in which an adult fly emerges from the cases, meaning identification is much more difficult (Braga *et al*. 2013).

Braga *et al*. (Braga *et al*. 2013) examined the puparial cases from four species of Sarcophagidae, *Peckia (Pekia) chrysostoma*, *Peckia (Pattonella) intermutans*, *Sarcophaga (Liopygia) ruficornis* and *Sarcodexia lambens*. They were able to differentiate between the specimens using their chemical profiles and analysing them using GC-MS and Bray-Curtis similarity index for cluster analysis. This shows great potential for analysing entomological evidence when only parts of the insect specimens are available for analysis. The results also highlight the strength of this technique when applied to a family of Diptera that are notoriously difficult to identify using morphological techniques.

Research carried out by Musah *et al*. (Musah *et al*. 2015) also examined the empty puparial cases but looked at four species of Calliphoridae, specifically; *Lucilia cuprina*, *L. sericata*, *Cochliomyia macellaria* and *Chrysomya rufifacies*. *Musca domestica* was also added to the dataset for comparison as this species had been previously sampled by one of the authors. Rather than the typical analytical technique of choice for CHC analysis, GC-MS, Musah *et al*. utilised the rapid ambient ionisation technique of Direct Analysis in Real Time – Mass Spectrometry

(DART-MS). DART analysis does not typically lend itself to hydrocarbon analysis since these compounds can be difficult to detect; especially the *n*-alkanes as they do not readily protonate. This makes the analysis of this type of compound much better suited to a high energy ionisation technique such as Electron Ionisation (in GC-MS) in order for them to become ionised. However, with an adapted sampling technique, *n*-alkanes were able to remain stable and hold their charge, enabling them to be detected with their $O_2^-$ adducts for the first time using DART-MS (Moore 2013). The results showed clear chemical distinctions between the analysed species, yielding a high throughput method for species identification and classification. Work from this initial experiment was continued by looking at the effect of geographical location on the cuticular hydrocarbon profiles of the same species collected from Texas and Ohio. Empty puparial cases from *C. macellaria* were collected from the two regions in North America and analysed using DART-MS. The results showed small chemical differences within the cuticular hydrocarbon profiles and by applying Linear Discriminant Analysis (LDA), their distinct clusters could be visualised enabling differentiation between geographical location (Moore 2013).

Previous work from Yew *et al.* (Yew *et al.* 2008) investigated the pheromones in the form of cuticular hydrocarbons, which play a vital role in courtship for the fly species *Drosophila*. This work was carried out on awake, behaving adults using DART and they observed differences in the chemical composition between male and female profiles by detecting unsaturated hydrocarbons. However they did not gain clear, unambiguous mass spectra from the saturated alkanes (Musah *et al*. 2015).

As stated previously, the larval stages are usually difficult to identify to species level due to morphological similarities but they can be done in 3rd instar by experienced entomologists (Szpila 2010) but it can be done as early as the 1st instar stage (Szpila *et al*. 2012). However, it is still usual practice to rear the larvae to adult flies for identification. However this can be a time consuming process when involved in criminal investigations, as identification is the first key step before the correct life cycle timings for aging can be utilised. Chemotaxonomy results from 1st in star larvae of *L. sericata*, *Calliphora vicina* and *Calliphora vomitoria* was published by Moore *et al.* (Moore *et al*. 2014). During this study the authors observed that the species-specific chemical profiles allow for identification to be determined in the immature stages, saving valuable time.

**Cuticular hydrocarbon analysis for ageing**

Cuticular hydrocarbons have also been utilised in entomology for ageing (Brown et al. 2000; Brown, Morton, & Spradbery 1992; Chen *et al.* 1990; Desena et al. 1999; Desena *et al.* 1999; Hugo *et al.* 2006; Tregenza *et al.* 2000) and sex identification (Brown *et al.* 2000, Chen *et al.* 1990; Jackson & Bartelt 1986; Mpuru *et al.* 2001; Steiner *et al.* 2006; Tregenza *et al.* 2000). Trabalon *et al.* (Trabalon *et al.* 1992) examined the cuticular hydrocarbons of *C. vomitoria* with relation to age and sex using GC-MS and Principal Component Analysis (PCA) via liquid extraction from the leg of the adult flies. Usual practice is to extract from the whole insect specimen by submerging them in hexane for approximately 15 minutes. They were able to distinguish between young and old specimens due to the composition of the chemical profiles and they were also able to distinguish between males and females by examining the sex pheromones (alkenes).

Roux *et al.* (Roux *et al.* 2008) were the first to carry out a complete ontogenetical study of three forensically important blowflies (*C. vicina, C. vomitoria* and *Protophormia terraenovae*) using Gas Chromatography with a Flame Ionisation Detector (GC-FID), producing results that have shown the potential to utilise the hydrocarbons for a forensic entomological application. The discrimination analysis they carried out allowed for clear differentiations between the life stages (larvae, post-feeding larvae, puparia and adult fly).

Additional work from Zhu and co-authors (Zhu *et al.* 2006) also showed the potential to use hydrocarbons with a forensic application by being the first to analyse the larvae of *C. rufifacies* using GC-FID and GC-MS. The study presented results which allowed for ageing to be established by adopting the simple statistical analysis of taking the peak area of nonacosane (C29:H) and dividing it by the peak area of eight other selected compounds, giving a peak ratio compared to C29:H. This ratio was seen to significantly increase with age and was followed up by modelling the results using exponential, or power functions. However, the peak ratios can vary considerably between individuals and the method of the analyst selecting the other peaks for use with the ratio can be seen as being a rather subjective means of analysis.

Where a body is found to be highly decomposed and the only surviving entomological evidence remaining are the empty puparial cases, it would be very beneficial to be able to establish the

identification as well as the age to allow time since colonisation estimates to be determined. As previously mentioned, empty puparial cases are difficult to identify due to key morpological features being damaged as the adult flies eclose. Since the puparial cases are the hardened cuticle of the post-feeding larvae and are not living, they do not change morphologically over time which makes them impossible to age. However with the discovery of cuticular hydrocarbons analysis, researchers have been publishing some very promising results around ageing empty puparial cases. Zhu *et al.* (Zhu *et al.* 2007) investigated the weathering effects of hydrocarbons extracted from the empty puparial cases of *C. megacephala*. During the study they used GC-MS was used to analyse the hydrocarbons and significant changes of the abundances of the compounds in relation to weathering time was observed. Interestingly, the abundance of the even numbered low molecular weight *n*-alkanes (C22:H, C24:H, C26:H) increased with time which could allow for an approximate age to be established up to 90 days. This was initially carried out under controlled conditions in the laboratory (Zhu et al. 2007) but was then repeated in a field experiment to validate the results (Zhu *et al.* 2013). Once again they reported that the chemical profiles extracted from the empty puparial cases of *C. megacephala* changed significantly during the 90 day weathering process and of the 106 compounds detected, 104 decreased significantly in abundance in correlation to the weathering time. However, in comparison to the lab data, they found the changes for the field study to be much greater, suggesting that cuticular hydrocarbons are significantly influenced by environmental factors. Zhu *et al.* (Zhu *et al.* 2017) continued field validation of empty puparial cases with a different species of forensic importance; *C. rufifacies*. This was another good example where they examined the effect of natural weathering for a characterisation experiment in order to assess the chemical composition changes over time. They presented data for 50 days of weathering time with some in-depth statistical analysis which overall showed a decrease in the hydrocarbon abundances with time, modelled with a modified exponent function. As reported by Zhu in a previous paper (Zhu *et al.* 2007), they once again state that the weathering effects are correlated to the different cuticular hydrocarbon classes, with *n*-alkanes yielding relatively low weathering rates and methyl branched alkanes and alkenes showing relatively high weathering rates (Zhu *et al.* 2017). They also reported that the hydrocarbons weathered more rapidly in comparison to laboratory controlled datasets, again suggesting the importance of the influence of additional environmental factors on the stability of these cuticular compounds.

Further research on the estimation of age from puparial cases was carried out by Moore *et al*. (Moore *et al*. 2017) who published data from the empty puparial cases of *C. vicina* and *L. sericata* over a period of 9 months. They used GC-MS analysis and non-metric multidimensional scaling analysis (NMDS) permutational multivariate analysis of variance (PERMANOVA) and random forest models were applied to the hydrocarbon datasets to examine any chemical changes occurring in relation to time. This study was successful in establishing age differences and is the first of its kind to age empty puparial cases to this time frame. However, although these were preliminary results which showed great potential, it must be repeated in the field to reflect the weathering changes modelled by other researchers.

Two studies have specifically examined chemical changes with time within larvae, enabling the age to be established for *L. sericata*. These studies utilised the techniques of both PCA (Moore *et al*. 2013) and Artificial Neural Networks (ANNs) (Butcher *et al*. 2013). This was not the first time ANNs were used in the field of entomology but it does describe the first use of ANNs for the analysis of hydrocarbons of Diptera for PMI estimations (Butcher *et al*. 2013). Like PCA, ANNs are capable of identifying trends within datasets but they work on a training and learning basis. Once trained, ANNs can reduce the analysis time required by clarifying novel data based on their knowledge of the domain that they have acquired during training (Moore 2013). The results showed significant promise for ageing the larval stages and this was replicated by a study from Moore *et al*. (Moore *et al*. 2016), successfully ageing *C. vomitoria* and *C. vicina* larvae.

Pechal *et al*. (Pechal *et al*. 2014) investigated the use of cuticular hydrocarbons for the chemical ageing of adult flies. The CHCs were examined for up to 30 days post-emergence of *C. macellaria* and *C. rufifacies*. Non-metric multidimensional scaling analysis and permutational multivariate analysis of variance were applied to the hydrocarbon dataset with significant differences detected amongst post-emergence age groups for each species. This was followed by a study carried out by Moore *et al*. (Moore *et al*. 2017) on adult flies up to the same age (30 days) but focussing on *L. sericata*, *C. vicina* and *C. vomitoria* using ANN analysis. This was a proof of concept study which developed a method allowing the age of post-emergence adult blow flies by examining their chemical profiles. However, they found that self-organising map (SOM) were able to classify each of the adult fly ages with very promising results, achieving

100% classification accuracy in the case of the *L. sericata*, and at least 70% classification accuracy for *C. vicina* and *C. vomitoria* (Moore *et al.* 2017).

Cuticular hydrocarbon analysis is in its early stages of research within the field of forensic entomology and it is important that the cuticular chemistry of all forensically important Calliphoridae are studied so they can be used as a base line reference. Braga *et al.* (Braga *et al.* 2016) analysed the CHC from male and female *Chrysomya putoria.* They used GC-MS and cluster analysis with Bray-Curtis measure of the relative abundances, demonstrating that it is possible to differentiate the ages of both sexes using CHC profiles for up to 5-day-old flies.

**Conclusion**

The research carried out by the stated authors in this review above briefly summarises the work currently being carried out using cuticular hydrocarbons within the area of forensic entomology. There is a lot of potential for this work to be utilised for identification, especially in the immature life stages or with Diptera that is more difficult to identify using morphological criteria (e.g. Sarcophagidae). But its real potential comes in focus in relation to ageing. Ageing the post-feeding stage to the day is something that has always been challenging as the larvae do not change in size. Cuticular hydrocarbons shows good accuracy for this life stage as well as adult flies and empty puparial cases which currently cannot be aged using any other technique.

The next step for this field is to replicate the published lab studies and validate this technique by observing the chemical profiles when exposed to environmental factors as this has been reported to alter the ageing models.

**References**

Adams, Z. & Hall, M.J.R. (2003) Methods used for the killing and preservation of blowfly larvae, and their effect on post-mortem larval length. *Forensic Science International*, **138**, 50–61.

Akino, T., Yamamura, K., Wakamura, S. & Yamaoka, R. (2004) Direct behavioral evidence for hydrocarbons as nestmate recognition cues in *Formica japonica* (Hymenoptera: Formicidae). *Applied Entomology and Zoology*, **39**, 381–387.

Amendt, J., Krettek, R., Niess, C., Zehner, R. & Bratzke, H. (2000) Forensic entomology in Germany. *Forensic Science International*, **113**, 309–14.

Ames, C., Turner, B. & Daniel, B. (2006a) Estimating the post-mortem interval (I): The use of genetic markers to aid in identification of Dipteran species and subpopulations. *International Congress Series*, **1288**, 795–797.

Ames, C., Turner, B. & Daniel, B. (2006b) The use of mitochondrial cytochrome oxidase I gene (COI) to differentiate two UK blowfly species - *Calliphora vicina* and *Calliphora vomitoria*. *Forensic science international*, **164**, 179–82.

Benecke, M. (2001) A brief history of forensic entomology. *Forensic Science International*, **120**, 2–14.

Bernier, U.R., Carlson, D.A. & Geden, C.J. (1998) Analysis of the Cuticular Hydrocarbons from Parasitic Wasps of the Genus Muscidifurax. *Journal of the American Society for Mass Spectrometry*, **9**, 320–332.

Blomquist, G.J. & Bagnères, A.G. (Eds.). (2010) *Insect Hydrocarbons: Biology, Biochemistry, and Chemical Ecology*. Cambridge University Press.

Braga, M.V., Bragaa, Pintob, Z.T., de Carvalho Queiroza, M.M., Matsumotod, N. & Blomquist, G.J. (2013) Cuticular hydrocarbons as a tool for the identification of insect species: Puparial cases from Sarcophagidae. *Acta Tropica,* **128**, 479– 485.

Braga, M.V., Bragaa, Pintob, Z.T., de Carvalho Queiroza, M.M., Matsumotod, N. & Blomquist, G.J. (2016) Effect of age on cuticular hydrocarbon profiles in adult *Chrysomya putoria* (Diptera: Calliphoridae). *Forensic Science Internaitonal*, **259**, e37-e47

Brown, W.V., Morton, R. & Spradbery, J.P. (1992) Cuticular hydrocarbons of the Old World screw-worm fly, *Chrysomya bezziana* Villeneuve (Diptera: Calliphoridae). Chemical characterization and quantification by age and sex. *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry*, **101**, 665–671.

Brown, W.V., Rose, H.A., Lacey, M.J. & Wright, K. (2000) The cuticular hydrocarbons of the giant soil-burrowing cockroach *Macropanesthia rhinoceros saussure* (Blattodea: Blaberidae: Geoscapheinae): analysis with respect to age, sex and location. *Comparative Biochemistry and Physiology. Part B, Biochemistry & molecular biology*, **127**, 261–77.

Butcher, J.B., Moore, H.E., Day, C.R., Adam, C.D. & Drijfhout, F.P. (2013) Artificial Meural Network analysis of hydrocarbon profiles for the ageing of *Lucilia sericata* for Post Mortem Interval estimation. *Forensic Science International*, **232**, 25-31.

Cainé, L.M., Real, F.C., Saloña-Bordas, M.I., Pancorbo, M.M. de, Lima, G., Magalhães, T. & Pinheiro, F. (2009) DNA typing of Diptera collected from human corpses in Portugal. *Forensic Science International*, **184**, e21–23.

Chapman, R.F. & Espelies, K.E. (1995) Use of Cuticular Lipids in Grasshopper Taxonomy: A Study of Variation in *Schistocerca shoshone* (Thomas). *Biochemical Systematics and Ecology*, **23**, 383–398.

Chen, C.S., Mulla, M.S., March, R.B. & Chaney, .J.D. (1990) Cuticular hydrocarbon Patterns in Culex-quinquefasciatus as influenced by age and sex and geography. *Bulletin of the Society for Vector Ecology*, **15**, 129–139.

Desena, M.L., Clark, J.M., Edman, J.D., Symington, S.B., Scott, T.W., Clark, G.G. & Peters, T.M. (1999) Potential for ageing Female<i> Aedes aegypti <i/> (Diptera: Culicidae) by Gas Chromatographic Analysis of Cuticular Hydrocarbons, Including a Field Evaluaiton. *Journal of Medical Entomology*, **36**, 811–823.

Desena, M.L., Edman, J.D., Clark, J.M., Symington, S.B. & Scott, T.W. (1999) *Aedes aegypti* (Diptera: Culicidae) Age Determination by Cuticular Hydrocarbon Analysis of Female Legs. *Journal of Medical Entomology*, **36**, 824–830.

Everaerts, C., Farine, J.P. & Brossut, R. (1997) Changes of species specific cuticular hydrocarbon profiles in the cockroaches *Nauphoeta cinerea* and *Leucophaea maderae* reared in heterospecific groups. *Behavioral Ecology and Sociobiology*, **85**, 145–150.

Gennard, D.E. (2007) *Forensic Entomology*. John Wiley & Sons Ltd.

Gibbs, A.G. (1998) Water-Proofing Properties of Cuticular Lipids. *American Zoologist*, **38**, 471–482.

Gibbs, A.G. & Crockett, E.L. (1998) The Biology of Lipids: Integrative and Comparative Perspectives. *Integrative and Comparative Biology*, **38**, 265–267.

Guillem, R.M., Drijfhout, F.P. & Martin, S.J. (2012) Using chemo-taxonomy of host ants to help conserve the large blue butterfly. *Biological Conservation*, **148**, 39–43.

Hart, A.J., Whitaker, A.P. & Hall, M.J.R. (2008) The Use of Forensic Entomology in Criminal Investigations: How it can be of benefit to SIOs. *The Journal of Homicide and Major Incident Investigation*, **4**, 37–48.

Harvey, M.L., Dadour, I.R. & Gaudieri, S. (2003) Mitochondrial DNA cytochrome oxidase I gene: potential for distinction between immature stages of some forensically important fly species (Diptera) in western Australia. *Forensic Science International*, **131**, 134–9.

Haverty, M.I., Collins, M.S., Nelson, L.J. & Thorne, B.L. (1997) Cuticular Hydrocarbons of Termites of the British Virgin Islands. *Journal of Chemical Ecology*, **23**, 927–964.

Howard, R.W. & Blomquist, G.J. (2005) Ecological, behavioral, and biochemical aspects of insect hydrocarbons. *Annual review of entomology*, **50**, 371–93.

Hugo, L.E., Kay, B.H., Eaglesham, G.K., Holling, N. & Ryan, P.A. (2006) Investigation of cuticular hydrocarbons for determining the age and survivorship of australasian mosquitoes. *The American Journal of Tropical Medicine and Hygiene*, **74**, 462–74.

Jackson, L.L. & Bartelt, R.J. (1986) Cuticular Hydrocarbons of Sex. *Insect Biochemistry*, **16**, 433–439.

Lavine, B.K. & Vora, M.N. (2005) Identification of Africanized honeybees. *Journal of Chromatography. A*, **1096**, 69–75.

Lockey, K.H. (1992) Insect hydrocarbon chemotaxonomy: Cuticular hydrocarbons of adult and larval epiphysa species blanchard and adult *Onymacris unguicularis* (HAAG) (tenebrionidae: Coleoptera). *Comparative Biochemistry and Physiology Part B: Comparative Biochemistry*, **102**, 451–470.

Lockey, K.H. (1991) Insect hydrocarbon classes: Implications for chemotaxonomy. *Insect Biochemistry*, **21**, 91–97.

Malgorn, Y. & Coquoz, R. (1999) DNA typing for identification of some species of Calliphoridae. An interest in forensic entomology. *Forensic Science International*, **102**, 111–9.

Martin, S.J. & Drijfhout, F.P. (2009) How reliable is the analysis of complex cuticular hydrocarbon profiles by multivariate statistical methods? *Journal of Chemical Ecology*, **35**, 375–82.

Martin, S.J., Helanterä, H. & Drijfhout, F.P. (2008) Colony-specific hydrocarbons identify nest mates in two species of Formica ant. *Journal of Chemical Ecology*, **34**, 1072–80.

Martin, S., Vitikainen, E., Helanterä, H. & Drijfhout, F. (2008) Chemical basis of nest-mate discrimination in the ant *Formica exsecta*. *Proceedings of the Royal Soceity B: Biological Sciences*, **275**, 1271–1278.

Moore, H.E. (2013) Analysis of cuticular hydrocarbons in forensically important blowflies using mass spectrometry and its application in Post Mortem Interval estimations. PhD thesis, Keele University, UK.

Moore, H.E., Adam, C.D. & Drijfhout, F.P. (2013) Potential Use of Hydrocarbons for Aging *Lucilia sericata* Blowfly Larvae to Establish the Postmortem Interval. *Journal of Forensic Sciences*, **58**, 404-412.

Moore, H.E., Adam, C.D. & Drijfhout, F.P. (2014) Identifying 1st instar larvae for three forensically important blowfly species using "fingerprint" cuticular hydrocarbon analysis. *Forensic Science International*, **240**, 48-53.

Moore, H.E., Butcher, J.B., Adam, C.D., Day, C.R., Drijfhout, F.P. (2016) Age estimation of Calliphora (Diptera: Calliphoridae) larvae using cuticular hydrocarbon analysis and Artificial Neural Networks. *Forensic Science International*, **268**, 81-91.

Moore, H.E., Butcher, J.B., Adam, C.D., Day, C.R., Drijfhout, F.P. (2016) Age estimation of Calliphora (Diptera: Calliphoridae) larvae using cuticular hydrocarbon analysis and Artificial Neural Networks. *Forensic Science International*, **268**, 81-91.

Moore, H.E., Butcher, J.B., Day, C.R., Drijfhout, F. P. (2017) Adult fly age estimations using cuticular hydrocarbons and Artificial Neural Networks in forensically important Calliphoridae species. *Forensic Science International*, **280**, 233-244.

Moore, H.E., Pechal, J.L., Benbow, M.E. Drijfhout, F.P. (2017) The potential use of cuticular hydrocarbons and multivariate analysis to age empty puparial cases of *Calliphora vicina* and *Lucilia sericata. Scientific Reports*, **7**:1933.

Mpuru, S., Blomquist, G.J., Schal, C., Roux, M., Kuenzli, M., Dusticier, G., Clément, J.L. & Bagnères, A.G. (2001) Effect of age and sex on the production of internal and external hydrocarbons and pheromones in the housefly, *Musca domestica. Insect Biochemistry and Molecular Biology*, **31**, 139–55.

Musah, R.A., Espinoza, E.O., Cody, R.B., Lesiak, A.D., Christensen, E.D., Moore, H.E., Maleknia, S. & Drijfhout, F.P. (2015) A high throughput ambient mass spectrometric approach to species identification and classification from chemical fingerprint signatures. *Scientific Reports*, **5,** 1-16.

Page, M., Nelson, L.J., Blomquist, G.J. & Seybold, S.J. (1997) Cuticular Hydrocarbons as Chemotaxonomic Characters of Pine Engraver Beetles (Ips spp.) in the grandicollis Subgeneric Group. *Journal of Chemical Ecology*, **23**, 1053–1099.

Pechal, J.L., Moore, H.E., Drijfhout, F.P., Benbow, M.E. (2014) Hydrocarbon profiles throughout adult Calliphoridae aging: A promising tool for forensic entomology. *Forensic Science International*, **245**, 65-71.

Ratcliffe, S.T., Webb, D.W., Weinzievr, R.A. & Robertson, H.M. (2003) PCR-RFLP identification of Diptera (Calliphoridae, Muscidae and Sarcophagidae)--a generally applicable method. *Journal of Forensic Sciences*, **48**, 783–5.

Roux, O., Gers, C. & Legal, L. (2008) Ontogenetic study of three Calliphoridae of forensic importance through cuticular hydrocarbon analysis. *Medical and Veterinary Entomology*, **22**, 309–17.

Sperling, F.A., Anderson, G.S. & Hickey, D.A. (1994) A DNA-based approach to the identification of insect species used for postmortem interval estimation. *Journal of Forensic Sciences*, **39**, 418–27.

Steiner, S., Hermann, N. & Ruther, J. (2006) Characterization of a female-produced courtship pheromone in the parasitoid Nasonia vitripennis. *Journal of Chemical Ecology*, **32**, 1687–702.

Stevens, J. & Wall, R. (2001) Genetic relationships between blowflies (Calliphoridae) of forensic importance. *Forensic Science International*, **120**, 116–123.

Szpila, K. (2010) Key for the Identification of Third Instars of European Blowflies (Diptera: Calliphoridae) of Forensic Importance. *Current Concepts in Forensic Entomology*. (eds J. Amendt, C.P. Campobasso, M.L. Goff & M. Grassberger), p. 47. Springer.

Szpila, K., Hall, M.J.R., Pape, T. & Grzywacz, a. (2012) Morphology and identification of first instars of the European and Mediterranean blowflies of forensic importance. Part II. Luciliinae. *Medical and veterinary entomology*, 87–100.

Tissot, M., Nelson, D.R. & Gordon, D.M. (2001) Qualitative and quantitative differences in cuticular hydrocarbons between laboratory and field colonies of *Pogonomyrmex barbatus*. *Comparative Biochemistry and Physiology. Part B, Biochemistry & Molecular Biology*, **130**, 349–58.

Trabalon, M., Campan, M., Clement, J.L., Lange, C. & Miquel, M.T. (1992) Cuticular Hydrocarbons of *Calliphora vomitoria* (Diptera): Relation to Age and Sex. *General and Comparative Endocrinology*, **85**, 208–216.

Tregenza, T., Buckley, S.H., Pritchard, V.L. & Butlin, R.K. (2000) Inter- and Intra-Population Effects of Sex and Age on Epicticular Composition of Meadow Grasshopper,*Chorthippus parallelus*. *Journal of Chemical Ecology*, **26**, 257–278.

Urech, R., Brown, G.W., Moore, C.J. & Green, P.E. (2005) Cuticular hydrocarbons of buffalo fly, *Haematobia exigua*, and chemotaxonomic differentiation from horn fly, *H. irritans*. *Journal of Chemical Ecology*, **31**, 2451–61.

Vincent, S., Vian, J.M. & Carlotti, M.P. (2000) Partial sequencing of the cytochrome oxydase b subunit gene I: a tool for the identification of European species of blow flies for postmortem interval estimation. *Journal of Forensic Sciences*, **45**, 820–3.

Wallman, J.F. & Donnellan, S.C. (2001) The utility of mitochondrial DNA sequences for the identification of forensically important blowflies (Diptera : Calliphoridae) in southeastern Australia. *Forensic Science International*, **120**, 60–67.

Wells, J.D. & Sperling, F.A.H. (2001) DNA-based identification of forensically important Chrysomyinae (Diptera : Calliphoridae). *Forensic Science International*, **120**, 110–115.

Ye, G., Li, K., Zhu, J., Zhu, G. & Hu, C. (2007) Cuticular hydrocarbon composition in pupal exuviae for taxonomic differentiation of six necrophagous flies. *Journal of Medical Entomology*, **44**, 450–6.

Yew, J.Y., Cody, R.B. & Kravitz, E. a. (2008) Cuticular hydrocarbon analysis of an awake behaving fly using direct analysis in real-time time-of-flight mass spectrometry. *Proceedings of the National Academy of Sciences of the United States of America*, **105**, 7135–40.

Zhu, G.H., Xu, X.H., Yu, X.J., Zhang, Y. & Wang, J.F. (2007) Puparial case hydrocarbons of *Chrysomya megacephala* as an indicator of the postmortem interval. *Forensic Science International*, **169**, 1–5.

Zhu, G.H., Ye, G.Y., Hu, C., Xu, X.H. & Li, K. (2006) Development changes of cuticular hydrocarbons in *Chrysomya rufifacies* larvae: potential for determining larval age. *Medical and Veterinary Entomology*, **20**, 438–44.

Zhu, G.H., Yu, X.J., Xie, L.X., Luo, H.,Wang, D., Lv, J.Y. & Xu, X.H (2013) Time of Death Revealed by Hydrocarbons of Empty Puparia of *Chrysomya megacephala* (Fabricius) (Diptera: Calliphoridae): A Field Experiment. *PLoS ONE*, **8**, 1-7.

Zhu, G.H., Jia, Z.J., Yu, X.J., Wu, K.S., Chen, L.S., Lv, J.Y. & Benbow, M.E. (2017) Predictable weathering of puparial hydrocarbons of necrophagous flies for determining the postmortem interval: a field experiment using Chrysomya rufifacies. *International Journal of Legal Medicine*, **131**, 885–894.