

# Mining the untapped chemical potential of entomopathogenic fungi

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## BACKGROUND

- Given the immense fungal biodiversity (with an estimated 2.2 to 3.8 million species of fungi worldwide [1]) and broad range of fungal habitats, fungi are one of the best sources of natural bioactive compounds, with huge industrial and medicinal potential.
- Entomopathogenic fungi (EPF) are fungi that infect and kill arthropods.
- EPF produce unique secondary metabolites and rapidly adjust their metabolic outputs in response to changes in environmental conditions. [2] These metabolites are usually produced transiently, in low quantities or not at all under laboratory conditions.
- With the market for commercial EPF biopesticides growing considerably in recent years, EPF secondary metabolites present an understudied and biotechnologically valuable opportunity for the pharmaceutical and agricultural industries.

## PROJECT AIM:

Develop liquid culturing techniques of EPF to optimise metabolite production that facilitates structural elucidation and biological activity testing

## METHODS

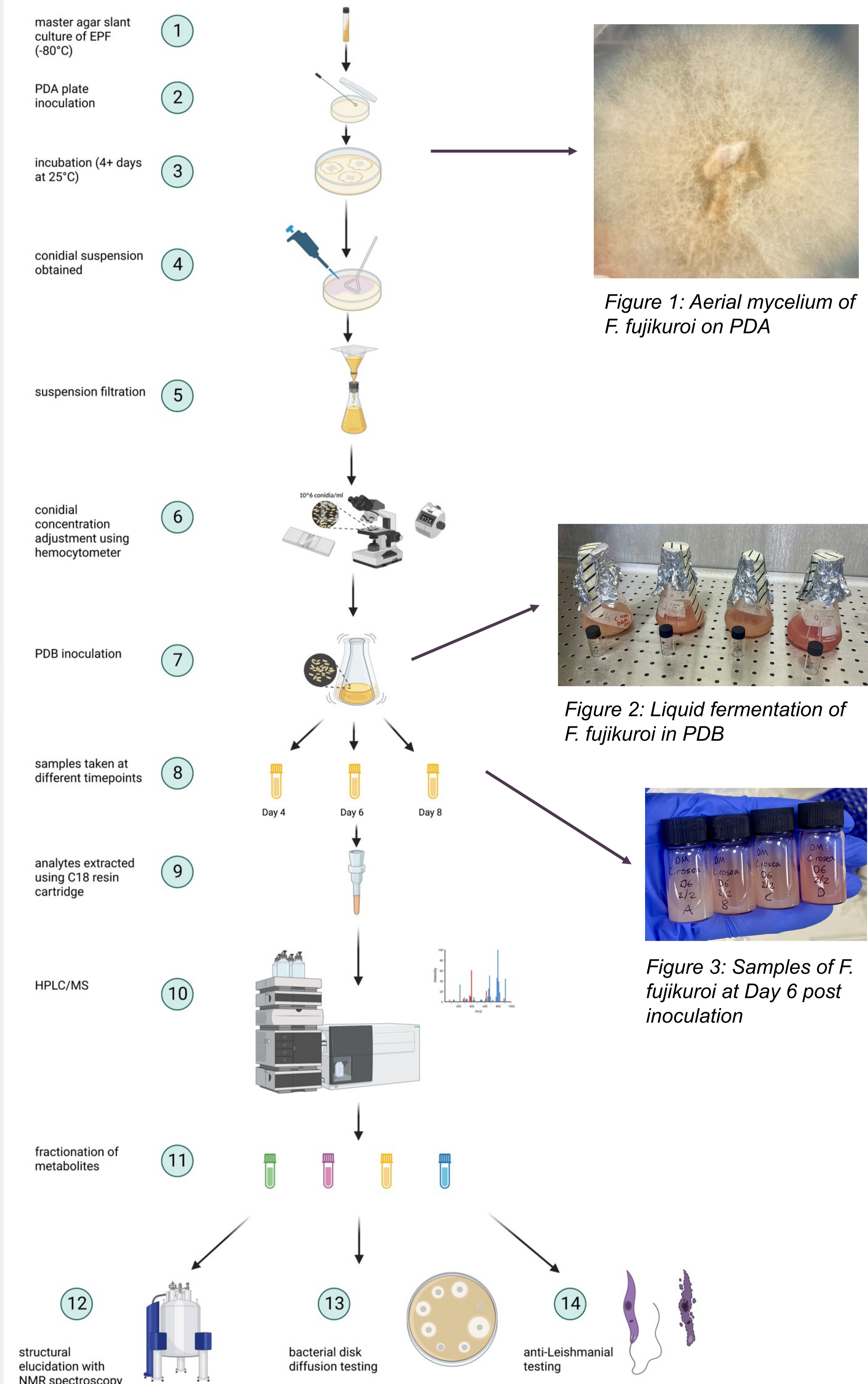


Figure 1: Aerial mycelium of *F. fujikuroi* on PDA

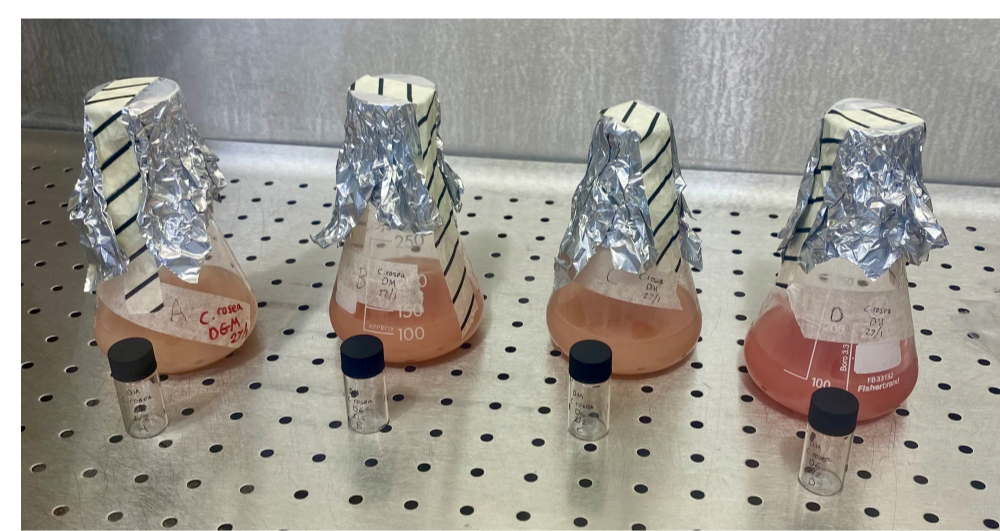


Figure 2: Liquid fermentation of *F. fujikuroi* in PDB

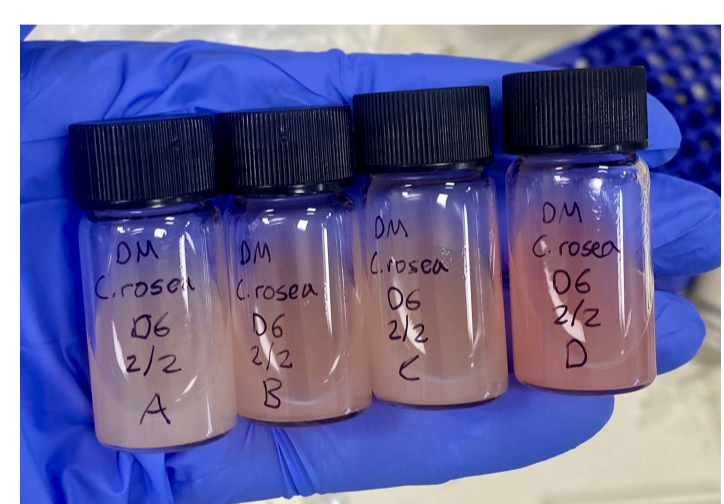


Figure 3: Samples of *F. fujikuroi* at Day 6 post inoculation

## RESULTS

Exploration liquid fermentation experiments in *Fusarium fujikuroi* were performed. *F. fujikuroi* is a fungal plant pathogen that causes bakanae disease in rice seedlings, it has been found to exhibit natural entomopathogenicity. [3]

HPLC/MS on time course samples demonstrated a change in metabolite production over time, with major peaks noted on Day 10 (Fig 4:2) compared to Day 6 (Fig 4:1), at m/z (Da) of 721 (Fig 4:A), 765 (Fig 4:B) and 707 (Fig 4:C).

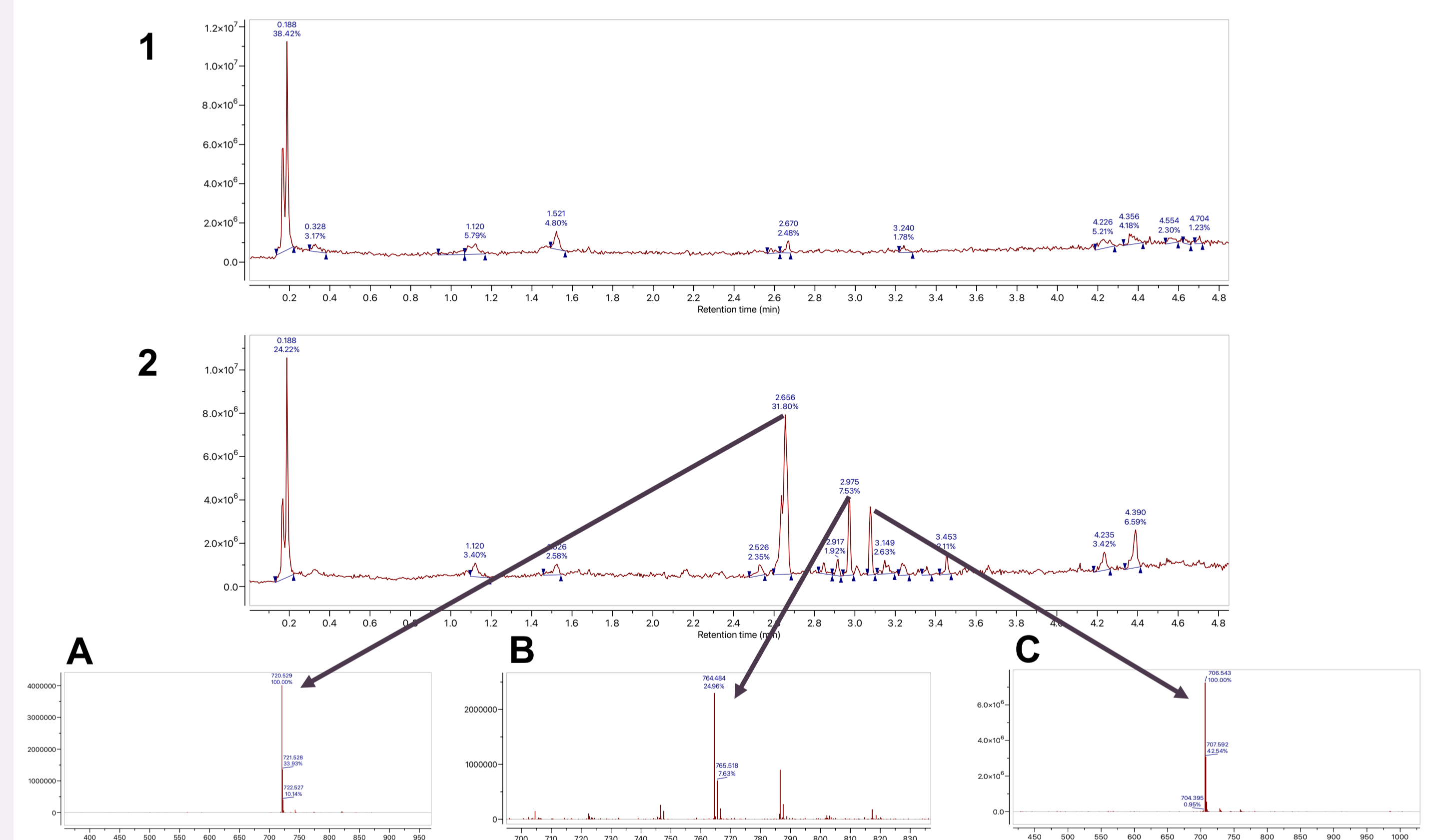


Figure 4: Total ion chromatogram from HPLC/MS of *F. fujikuroi* in acetonitrile at Day 6 (1) and Day 10 (2) post inoculation. Negative-ion mode electrospray ionisation mass spectra major peaks isolated as secondary metabolites (A, B and C).

Metabolite	RT	Height	Area	Total Height %	Total Area %	Width	m/z (Da)	Metabolite	F. oxysporum compound	Molecular weight	Molecular formula	Host	Place
A	2.656	737088	4.4E+07	22.89	31.8	0.021	721	A	Fumonisin B <sub>1</sub>	721	C <sub>21</sub> H <sub>35</sub> NO <sub>15</sub>	Asparagus officinalis	Western Poland
B	2.975	351465	1E+07	10.91	7.53	0.012	765	B	N-Acetylated OH-fumonisin C <sub>1</sub>	765	C <sub>23</sub> H <sub>37</sub> NO <sub>17</sub>	Dianthus caryophyllus	Taejeon, Korea
C	3.076	314854	916245	9.78	6.61	0.014	707	C	Fumonisin C <sub>1</sub>	707	C <sub>23</sub> H <sub>37</sub> NO <sub>15</sub>	Dianthus caryophyllus	Taejeon, Korea

Table 1: Retention times (RT), peak height, area, width and mass (m/z in Da) of metabolites A, B and C

Table 2: Top three predicted molecular formulae for metabolites A, B & C

Literature search revealed *Fusarium oxysporum* secondary metabolites with identical molecular weights [4], suggesting the metabolites could be mycotoxic fumonisins (Table 2).

## FUTURE WORK

- Repeat fermentation and metabolite profiling with the EPF and rare hyphal parasite *Clonostachys rosea*.
- Develop new liquid fermentation methodologies to replicate the biochemical conditions in insect hosts (eg: culture with insect-derived antimicrobials, identified in Table 3), and analyse differences in secondary metabolite profile.
- Isolate metabolites by fractionation of crude extracts, to enable structural elucidation by NMR spectroscopy and microbiological activity testing.
- Isolate genes responsible for metabolite production using DNA and RNA sequencing, to enlighten our understanding of EPF modes of infection.

Antifungal	Produced by
Drosomycin	fruit fly <i>Drosophila melanogaster</i>
Termiticin	termite <i>Pseudacanthotermes spiniger</i>
Heliomycin	tobacco budworm <i>Heliothis virescens</i>
Gallerimycin	greater wax moth larvae <i>Galleria mellonella</i>
Cecropin A	giant silk moth <i>Hyalopora cecropia</i>
Cecropin B	giant silk moth <i>Hyalopora cecropia</i>
Thanatin	spined soldier bug <i>Podisus maculiferis</i>
Spinigerin	termite <i>Pseudacanthotermes spiniger</i>
Stomoxyn	stable fly <i>Stomoxys calcitrans</i>
Defensin-like antifungal peptide	whitefly <i>Bemisia tabaci</i>

Table 3: Insect-derived antimicrobials

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## References:

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- Hyde, K.D. et al., 2019. The amazing potential of fungi: 50 ways we can exploit fungi industrially. *Fungal Diversity*, 97, pp.1-136.
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