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1 Sexually dimorphic murine brain uptake of the 18 kDa 2 translocator protein PET radiotracer [¹⁸F]LW223

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8 Abstract

9 The 18 kDa translocator protein (TSPO) is a well-known biomarker of neuroinflammation, but
10 also plays a role in homeostasis. Positron Emission Tomography (PET) with TSPO radiotracers
11 [¹¹C]PBR28 in humans and [¹⁸F]GE180 in mice have demonstrated sex-dependent uptake
12 patterns in the healthy brain, suggesting sex-dependent TSPO expression, although humans
13 and mice had differing results. This study aimed to assess whether the TSPO PET radiotracer
14 [¹⁸F]LW223 exhibited sexually-dimorphic uptake in healthy murine brain and peripheral
15 organs.

16 Male and female C57Bl6/J mice (13.6±5.4 weeks, 26.8±5.4 g, mean±SD) underwent 2-hour
17 PET scanning post-administration of [¹⁸F]LW223 (6.7±3.6 MBq). Volume of interest and
18 parametric analyses were performed using standard uptake values (90-120 min). Statistical
19 differences were assessed by unpaired t-test or two-way ANOVA with Šidak's test
20 (alpha=0.05).

21 The uptake of [¹⁸F]LW223 was significantly higher across multiple regions of the male mouse
22 brain, with the most pronounced difference detected in hypothalamus ($p<0.0001$). Males also
23 exhibited significantly higher [¹⁸F]LW223 uptake in the heart when compared to females
24 ($p=0.0107$).

25 Data supports previous findings on sexually dimorphic TSPO radiotracer uptake patterns in
26 mice and highlights the need to conduct sex-controlled comparisons in TSPO PET imaging
27 studies.

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13 **Running title:** Sexually dimorphic [¹⁸F]LW223 uptake.

14

15 **Keywords:** TSPO; PET; [¹⁸F]LW223; sex differences.

16 **Abbreviations:** 3 α -HSOR - 3 alpha-hydroxysteroid oxidoreductase; 5 α -R - 5 alpha reductase;
17 CBF – cerebral blood flow; HPLC – high performance liquid chromatography; StAR - acute
18 regulatory protein; *SUV*_{90-120min} - standard uptake values averaged between 90 and 120 minutes;
19 TSPO - 18 kDa translocator protein; VOI – volume of interest.

20

21 **Abbreviated abstract:** Knyzeliene *et al.* report sexually dimorphic murine brain and heart
22 uptake of the novel 18 kDa translocator protein (TSPO) PET radiotracer [¹⁸F]LW223. Results
23 confirm that [¹⁸F]LW223 can detect changes in TSPO *in vivo* and with high sensitivity.

24

25

1

2 **Introduction**

3 The link between sex and susceptibility to a number of neurological diseases and disorders has
4 been well defined and new evidence is constantly emerging.^{1,2} However, prior to studying
5 disease states, and especially when characterising disease biomarkers, it is important to
6 understand whether any sex-dependent differences are present under healthy conditions.
7 Recent positron emission tomography (PET) imaging studies identified such differences for
8 the 18 kDa translocator protein (TSPO), which is usually considered a biomarker for
9 inflammation, but is also known to play a role in steroidogenesis, mitochondrial energy
10 metabolism, cell proliferation, apoptosis and immunomodulation.³⁻⁶ Tuisku *et al.*⁷ has shown
11 that in human subjects, binding of the PET radiotracer [¹¹C]PBR28 to TSPO in the brain was
12 16.3% higher in females than in males. They also found that with age, [¹¹C]PBR28 binding to
13 TSPO in the brain increased in male, but not in female subjects. Conversely, a preclinical study
14 by Biechele *et al.*⁸ showed that male mice had a higher brain uptake of another TSPO PET
15 radiotracer, [¹⁸F]GE180, than females. This study also demonstrated that with age, [¹⁸F]GE180
16 uptake in the murine male brain remained stable, whereas it increased in females. In young
17 adult wild-type mice, [¹⁸F]GE180 brain uptake was approximately 10-15% higher in males
18 compared with females.⁸ These early findings suggested that cross-gender, cross-species and
19 cross-radiotracer binding differences may exist among PET radiotracers targeting TSPO,
20 highlighting the need to carefully investigate these aspects for current and new compounds.

21 The current study aimed to assess sex differences in the uptake of the TSPO PET radiotracer
22 [¹⁸F]LW223 in healthy mice, hypothesising that higher brain uptake would be observed in
23 males compared with females (based on previous results in mice).⁸ Given [¹⁸F]LW223 unique
24 *in vivo* properties, including low non-displaceable volume (V_{ND}) in mouse brain, we also
25 hypothesised that our novel radiotracer had the potential to detect greater differences between
26 males and females with potential to unravel new biological insights by enabling higher
27 sensitivity analysis of regional TSPO changes in the murine brain versus other previously
28 developed TSPO PET radiotracers.⁹ Having rs6971 polymorphism-independent binding in
29 human tissue *ex vivo*, [¹⁸F]LW223 holds great potential to enable TSPO PET imaging across
30 the whole human population.^{10,11} Previously we have also shown that [¹⁸F]LW223 has excellent
31 properties as an imaging biomarker, including low radiometabolism, high free fraction and

1 binding kinetics amenable to the use of simplified outcome measures in both rats and mice.^{9,11}
2 Therefore, prior to its widespread application to image various diseases and disease models, it
3 is important to understand binding patterns of [¹⁸F]LW223 in healthy males and females.
4 Although this study mainly focussed on differences in the brain, it also assessed radiotracer
5 uptake differences at the whole-body level in mice.

6

1 **Materials and methods**

2 **Radiosynthesis of [¹⁸F]LW223**

3 The structure and radiosynthesis of [¹⁸F]LW223 was performed as described previously ¹¹,
4 except the mobile phase flow rate used for purification of the final product using a semi-
5 preparative high performance liquid chromatography (HPLC) system was reduced from 5
6 mL/min to 3 mL/min. [¹⁸F]LW223 was produced with good molar activity, as previously
7 described,¹¹ which is compliant with the radiotracer principle, as demonstrated in our
8 previously published mass effect study in mice.⁹

9 **Animals**

10 The animals used in the study were purchased from Charles River Laboratories (Tranent,
11 United Kingdom). All animal experiments were conducted and authorised by the local
12 University of Edinburgh animal welfare and ethical review committee and in accordance with
13 the Home Office Animals (Scientific Procedures) Act 1986. The animals were housed in IVC
14 cages under standard 12 h light:12 h dark conditions with food and water available *ad libitum*.
15 All animals were scanned during the 12h light cycle conditions.

16 **[¹⁸F]LW223 PET imaging**

17 Young male (n=9, age = 14.69±6.15 weeks, weight = 29.12±3.27 g) and female (n = 5, age =
18 11.56±2.45 weeks, weight = 22.56±5.77 g) C57Bl/6J mice were used in the PET study. The
19 data of seven out of the nine male and five female mice were repurposed from our previous
20 study by MacAskill *et al.*¹¹ Animals were anaesthetised using 1.5-2% isoflurane (Isoflo®
21 APIECE, Zoetis, UK) (50/50 oxygen/nitrous oxide, 1 L/min) and body temperature was
22 maintained using a heated mat. Tail vein cannulations for injection of [¹⁸F]LW223 were
23 performed using butterfly needles (27G 1/2", 12 cm polyurethane tubing, SAI Infusion

1 Technologies, USA), except for two male animals that underwent femoral vein and artery
2 cannulation for radiotracer injection and blood sampling.

3 PET scans were performed immediately post intravenous bolus injection of [¹⁸F]LW223 (n =
4 14, 6.36±3.70 MBq, bolus i.v., mean±SD). Imaging data were acquired using a preclinical
5 PET/CT scanner (nanoPET/CT, Mediso, Hungary). Respiration rate and body temperature
6 were monitored and maintained throughout the imaging session. A 2-hour emission dataset per
7 animal was reconstructed using 3-dimensional 1:5 mode and re-binned as follows: 18 × 10 s,
8 2 × 30 s, 1 × 60 s, 2 × 120 s, 10 × 300 s, 6 × 600 s. All PET studies were reconstructed using
9 Mediso's iterative Tera-Tomo 3D reconstruction algorithm, which includes point spread
10 correction, and the following settings: 4 iterations, 6 subsets, full detector model, normal
11 regularization, spike filter on, voxel size of 0.2 mm and 400-600 keV energy window.
12 Corrections for randoms, scatter and attenuation were applied to all PET data. Immediately
13 post PET imaging session, a 5-minute CT scan (semi-circular full trajectory, maximum field
14 of view, 480 projections, 50 kVp, 300 ms and 1:4 binning) was acquired for attenuation
15 correction and anatomical information. The following parameters were used for CT image
16 reconstruction: matrix size = 121 × 121 × 121 mm, voxel size = 0.25 × 0.25 × 0.25 mm, cosine
17 filter, cut-off at 100%, corrections for offset, gain and pixel.

18 **PET image processing and analysis**

19 Reconstructed images were analysed using PMOD version 3.7 (PMOD Technologies,
20 Switzerland). Volumes of interest (VOIs) were manually drawn around the whole brain, whole
21 heart and whole left and right lungs (with both lungs merged afterwards) using CT images. The
22 whole organ VOIs of spleen, kidneys, liver, adrenals and eyes were drawn using averaged PET
23 images (0-120 min). For regional brain analysis, PET data was co-registered with the mouse
24 brain T2 Magnetic Resonance Imaging (MRI) template and the modified Mirrione mouse brain

1 atlas was used to generate VOIs of the following brain regions: cortex, thalamus, cerebellum,
2 basal forebrain septum, hypothalamus, brain stem, central grey matter, olfactory bulb,
3 amygdala, midbrain, third ventricle, corpus callosum, striatum and hippocampus.¹² The data
4 were extracted as time-activity curves and standard uptake values (SUVs) were calculated as
5 concentration in the VOI divided by the injected dose divided by the animal weight. Average
6 SUV values between 90 and 120 minutes ($SUV_{90-120\ min}$) were used as an outcome measure.
7 This outcome measure was previously validated versus gold-standard invasive kinetic
8 modelling of [¹⁸F]LW223 murine brain PET datasets.⁹

9 **Generation of representative PET images**

10 Representative brain and whole-body PET $SUV_{90-120\ min}$ images for presentation purposes were
11 generated using PMOD version 3.7 (PMOD Technologies, Switzerland). For brain PET
12 images, Gaussian 3D 1.2 mm filter was applied, and they were co-registered with a mouse
13 brain T2 MRI template, whereas Gaussian 3D 1 mm filter was used for whole body PET
14 images.

15 **Parametric brain analysis**

16 To create parametric maps of [¹⁸F]LW223 uptake in the mouse brain, average male (n = 9) and
17 female (n = 5) SUV brain maps were generated. For each animal, the SUV was calculated for
18 each voxel of an average image of 90-120 minutes post-radiotracer injection as in the VOI
19 analysis detailed above. The SUV PET images were normalized to the same space as follows:
20 (1) the CT scans were cropped around the skull and each one was registered to an average MRI
21 atlas by Dorr *et al.*¹³; and (2) the transformation matrices between the PET, CT and MRI atlas
22 were combined to position each SUV image into the MRI space, therefore aligning all SUV
23 images. Each SUV image was smoothed with a Gaussian kernel of $0.2 \times 0.2 \times 0.2$ mm and the
24 brain was masked using the registered MRI atlas. Three separate average SUV brain maps were

1 created and made freely available online (<https://doi.org/10.7488/ds/2988>): (1) for male
2 animals, (2) for female animals and (3) for all animals, where an average over the animals in
3 each group was calculated for each voxel within the brain. Statistical analysis to determine any
4 significant difference between the male and female groups was carried out using R statistical
5 package and the RMINC (R studio package for Medical Imaging NetCDF (Network Common
6 Data Format)) module. The *t*-statistics calculated between groups were overlaid on the MRI
7 atlas.

8 **Statistical analysis**

9 Plotting of graphs and statistical analysis was performed using Prism 9.3.1 (GraphPad, USA).
10 Unpaired t-test or two-way ANOVA with Šidak's post hoc test (alpha=0.05) were used as
11 indicated in the relevant figure legends.

12 **Data availability**

13 Upon manuscript acceptance, all study data will be deposited in the "PET is Wonderful"
14 collection, hosted at the University of Edinburgh DataShare platform:
15 <https://datashare.ed.ac.uk/handle/10283/3219>

16

17 **Results**

18 **[¹⁸F]LW223 uptake was higher in specific regions of the healthy** 19 **male versus the healthy female murine brain**

20 [¹⁸F]LW223 blood uptake and kinetics was comparable in male versus female adult mice (**Fig.**
21 **1**). A 3D TSPO atlas of the female and male mouse brain was generated using *SUV_{90-120min}* PET
22 data collected following injection of [¹⁸F]LW223 (**Fig. 2A**). The VOI analysis showed, that in
23 male mice, [¹⁸F]LW223 PET uptake was up to 50% higher in the basal forebrain septum,

1 hypothalamus, brain stem, olfactory bulb and amygdala when compared to females (**Fig. 2B**).
2 This was confirmed by statistical parametric mapping analysis (**Fig. 2C**), with additional
3 significant differences detected in the striatum, hippocampus, thalamus, midbrain, cerebellum,
4 and in the frontal, entorhinal and parieto-temporal cortices (**Fig. 2C**).

5 **Sex differences of [¹⁸F]LW223 uptake identified in the whole brain** 6 **and heart of healthy mice**

7 In addition to assessing regional uptake of [¹⁸F]LW223 in male and female mouse brain, *SUV*₉₀₋
8 *120min* analysis was performed in peripheral organs that are known to express TSPO (**Fig. 3** and
9 **Fig. 4**). It was found, that in addition to having significantly higher *SUV*_{90-120min} in the brain of
10 healthy males (42% difference), the uptake of [¹⁸F]LW223 was also significantly higher in the
11 heart of male mice (**Fig. 4**). Other organs, such as the lungs, spleen, kidneys and adrenals did
12 not present a sex-dependent *SUV*_{90-120min} of [¹⁸F]LW223.
13

14 **Discussion**

15 Sexual dimorphism in protein expression is becoming increasingly acknowledged in the field
16 of neuroscience. For decades, the majority of neuroscience research has been performed
17 predominately in males, disregarding the fact that the expression of disease markers may
18 significantly differ between males and females even under healthy conditions, leading to
19 knowledge gaps across the field.¹⁴ Recently published data suggested that TSPO may also be
20 one of the proteins within this category.^{7,8}

21 Although the present study did not assess the effect of aging on [¹⁸F]LW223 binding in male
22 and female mouse brain, the results paralleled the outcomes of the equivalently aged C57Bl/6
23 mice from the Biechele *et al.* study, where young males had significantly higher [¹⁸F]LW223

1 *SUV_{90-120min}* across multiple brain regions when compared to females, supporting our
2 hypothesis. Results from this study also showed that global (42%) and regional (up to 50%)
3 changes in the [¹⁸F]LW223 murine male brain uptake versus the female murine brain were
4 more striking than previous reports using other TSPO PET radiotracers, namely [¹⁸F]GE180
5 (10-15% difference). This confirms our secondary hypothesis that [¹⁸F]LW223 has superb
6 sensitivity for detection of TSPO changes in the murine brain and reflects the lower *V_{ND}* of
7 [¹⁸F]LW223. Furthermore, we have previously shown that [¹⁸F]LW223 has good brain
8 penetration followed by slow clearance due to its high affinity to murine TSPO.⁹

9 The reasons behind sexual dimorphism of TSPO are still to be investigated, but a working
10 hypothesis is that it may be involved in sex-dependent regulation of neurosteroid production in
11 the brain. This would resemble expression patterns of other proteins involved in
12 neurosteroidogenesis, such as steroidogenic acute regulatory protein (StAR), 5 alpha reductase
13 (5 α -R) and 3 alpha-hydroxysteroid oxidoreductase (3 α -HSOR), which have been found to
14 differ between male and female rodents.¹⁵ Moreover, levels of neurosteroids, such as
15 dihydroprogesterone, tetrahydroprogesterone, isopregnanolone, dehydroepiandrosterone,
16 testosterone and others, also show regional differences in distribution between male and female
17 brain.¹⁶ However, it is still unclear why rodents and humans exhibit opposite trends when it
18 comes to sexual dimorphism of TSPO.⁷ Our results presented here confirm and expand prior
19 observations with a different TSPO radiotracer [¹⁸F]GE180 in mice.

20 The uptake of [¹⁸F]LW223 in peripheral mouse organs with known TSPO expression was also
21 investigated. It is important to gain this understanding as TSPO PET imaging is increasingly
22 applied for imaging inflammation beyond the brain, as well as for assessing functional heart-
23 brain TSPO axis.^{11,17,18} Opposed to previous findings on relative TSPO gene expression in
24 Balb/cJ mice, in the current study, C57Bl/6J males exhibited significantly higher uptake of
25 [¹⁸F]LW223 in the heart when compared to females, indicating higher levels of TSPO.¹⁹

1 Although this contradictory finding could be influenced by gene expression differences
2 between Balb/c and C57Bl/6 mouse strains, these results highlight the importance of
3 conducting sex-controlled TSPO PET imaging studies when assessing functional TSPO axes
4 or inflammatory conditions across the body.²⁰

5 Since *SUV_{90-120min}* was performed to quantify [¹⁸F]LW223 PET data in this study, it is possible
6 that the results might be impacted by possible confounding factors, such as sex-dependent
7 cerebral blood flow (CBF) differences. However, no data is currently available to support this
8 hypothesis, as no sex-dependent differences in CBF were previously found in rodents.²¹

9 As eluded previously, [¹⁸F]LW223 has a number of important advantages compared with
10 previously developed TSPO radiotracers, which can resolve the current bottleneck in clinical
11 TSPO PET imaging. Here we show that [¹⁸F]LW223 uptake across mouse brain regions and
12 heart is sex-dependent, with male mice expressing higher levels of TSPO when compared to
13 females. Gaining this information during the preclinical development of [¹⁸F]LW223 is
14 beneficial, as it aids better design of future preclinical and clinical studies with this lead TSPO
15 radiotracer, for which a translational package is already available, including dosimetry
16 analysis.¹¹ Our findings also highlighted the need for studies investigating sexually dimorphic
17 target expression patterns across the field of preclinical neuroscience, which would expand the
18 current understanding on brain functions and allow for the use of patient-tailored diagnostic
19 measures.

20

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8 Foundation. CA-C and CL are supported by Edinburgh Imaging.

9 **Competing interests**

10 A patent for TSPO binders has been submitted (application GB1810312.7 and
11 PCT/EP2019/066546). No other potential conflicts of interest relevant to this article exist.

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26

27

1 **Figure legends**

2 **Figure 1. [¹⁸F]LW223 whole blood time-activity curves.** (A) Male and female mouse image-
3 derived whole-blood time-activity curves from 0 to 120 minutes post-radiotracer
4 administration. (B) Insert showing whole-blood time-activity curves from 0 to 3 minutes post-
5 radiotracer administration. Data presented as mean±SD, n=9 males and n=5 females.

6

7 **Figure 2. Sex-dependent [¹⁸F]LW223 uptake differences detected during regional**
8 **analysis of a mouse brain.** (A) Representative images of female (n=5) and male (n=9) average
9 [¹⁸F]LW223 *SUV*_{90-120min} brain atlases. (B) A comparison of regional [¹⁸F]LW223 uptake in
10 female (n=5) and male (n=9) mouse brains (mean±SD; two-way ANOVA, Šidák's multiple
11 comparison test, alpha=0.05, * p=0.0112, ** p≤0.0079, **** p<0.0001). (C) Statistical
12 parametric map showing t-statistics for differences between the female (n=5) and male (n=9)
13 brain [¹⁸F]LW223 *SUV*_{90-120min}.

14

15 **Figure 3. Representative [¹⁸F]LW223 *SUV*_{90-120min} total-body images of female and male**
16 **mice.** Legend: B – brain, L- lungs, H – heart, K – kidneys, A – adrenals, S – spleen.

17

18 **Figure 4. Young male mice have higher [¹⁸F]LW223 uptake in brain and heart compared**
19 **with young female mice.** The data represents uptake of [¹⁸F]LW223 in brain and peripheral
20 organs of female (n=5) and male (n=9) mice (mean ± SD; * p=0.0107, ** p=0.0095, unpaired
21 t-test, alpha=0.05).

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